



Survival and Clinicopathological Significance of SIRT1 Expression in Cancers: A Meta-Analysis

Min Sun^{1,2†}, Mengyu Du^{2†}, Wenhua Zhang³, Sisi Xiong⁴, Xingrui Gong², Peijie Lei⁵, Jin Zha², Hongrui Zhu², Heng Li¹, Dong Huang^{1*} and Xinsheng Gu^{6*}

¹ Department of General Surgery, Taihe Hospital, Hubei University of Medicine, Shiyan, China, ² Department of Anesthesiology, Institute of Anesthesiology, Taihe Hospital, Hubei University of Medicine, Shiyan, China, ³ Department of Pharmaceutical Sciences, College of Pharmacy, Hubei University of Medicine, Shiyan, China, ⁴ School of Nursing, Hubei University of Medicine, Shiyan, China, ⁵ The First Clinical School, Hubei University of Medicine, Shiyan, China, ⁶ Department of Pharmacology, College of Basic Medical Sciences, Hubei University of Medicine, Shiyan, China

OPEN ACCESS

Edited by:

Yang Yang, Northwest University, China

Reviewed by:

Fan Peng, Fourth Military Medical University, China Zu-Bing Mei, Shuguang Hospital, Shanghai University of Traditional Chinese Medicine, China

*Correspondence:

Dong Huang hd_814@sohu.com Xinsheng Gu gu.xinsheng@gmail.com

[†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Cellular Endocrinology, a section of the journal Frontiers in Endocrinology

Received: 10 December 2018 Accepted: 11 February 2019 Published: 13 March 2019

Citation:

Sun M, Du M, Zhang W, Xiong S, Gong X, Lei P, Zha J, Zhu H, Li H, Huang D and Gu X (2019) Survival and Clinicopathological Significance of SIRT1 Expression in Cancers: A Meta-Analysis. Front. Endocrinol. 10:121. doi: 10.3389/fendo.2019.00121 **Background:** Silent information regulator 2 homolog 1 (SIRT1) is an evolutionarily conserved enzymes with nicotinamide adenine dinucleotide (NAD)⁺-dependent deacetylase activity. SIRT1 is involved in a large variety of cellular processes, such as genomic stability, energy metabolism, senescence, gene transcription, and oxidative stress. SIRT1 has long been recognized as both a tumor promoter and tumor suppressor. Its prognostic role in cancers remains controversial.

Methods: A meta-analysis of 13,138 subjects in 63 articles from PubMed, EMBASE, and Cochrane Library was performed to evaluate survival and clinicopathological significance of SIRT1 expression in various cancers.

Results: The pooled results of meta-analysis showed that elevated expression of SIRT1 implies a poor overall survival (OS) of cancer patients [Hazard Ratio (HR) = 1.566, 95% CI: 1.293–1.895, P < 0.0001], disease free survival (DFS) (HR = 1.631, 95% CI: 1.250–2.130, P = 0.0003), event free survival (EFS) (HR = 2.534, 95% CI: 1.602–4.009, P = 0.0001), and progress-free survival (PFS) (HR = 3.325 95% CI: 2.762–4.003, P < 0.0001). Elevated SIRT1 level was associated with tumor stage [Relative Risk (RR) = 1.299, 95% CI: 1.114–1.514, P = 0.0008], lymph node metastasis (RR = 1.172, 95% CI: 1.010–1.360, P = 0.0363), and distant metastasis (RR = 1.562, 95% CI: 1.022–2.387, P = 0.0392). Meta-regression and subgroup analysis revealed that ethnic background has influence on the role of SIRT1 expression in predicting survival and clinicopathological characteristics of cancers. Overexpression of SIRT1 predicted a worse OS and higher TNM stage and lymphatic metastasis in Asian population especially in China.

Conclusion: Our data suggested that elevated expression of SIRT1 predicted a poor OS, DFS, EFS, PFS, but not for recurrence-free survival (RFS) and cancer-specific survival (CCS). SIRT1 overexpression was associated with higher tumor stage, lymph node metastasis, and distant metastasis. SIRT1-mediated molecular events and biological processes could be an underlying mechanism for metastasis and SIRT1 is a therapeutic target for inhibiting metastasis, leading to good prognosis.

Keywords: SIRT1, cancer, prognosis, clinicopathological significance, meta-analysis

1

INTRODUCTION

Silent information regulator 2 homolog 1 (SIRT1) is an evolutionarily conserved enzymes with nicotinamide adenine dinucleotide $(NAD)^+$ -dependent deacetylase activity and a member of the mammalian sirtuin family. It is expressed in almost all human tissues and localized in both nuclei and cytoplasm (1). Its substrates include histones and non-histone proteins such as transcription factors (2–4). SIRT1 is involved in a large variety of cellular processes, such as genomic stability, energy metabolism, senescence, gene transcription, and oxidative stress (5). It has been shown to be involved in a spectrum of diseases, including cancer, diabetes, obesity, and neurodegenerative diseases (6–8). SIRT1 plays an important role in regulating glucose and lipid metabolism and regulates malignancy in tumors (9).

SIRT1 has long been recognized as both a tumor promoter and tumor suppressor (10-12). This is also shown in recent studies. SIRT1 promotes proliferation, migration, and invasion of breast cancer cell line MCF-7 (13). SIRT1 promotes proliferation and paclitaxel-resistance of human cervical cancer cells (14). Yang et al. found that SIRT1 levels are lower in non-small cell lung cancer (NSCLC) than the normal control group (15), but Gharabaghi et al. found that SIRT1 are over expressed in NSCLC (16). The role of SIRT1 in prognosis of cancer was also investigated in several studies. Over expression of SIRT1 suggests poor prognosis in luminal breast cancer (17) and serous epithelial ovarian cancer (EOC) (18), gastric cancer (19), high pathological stage and worse overall survival in the lung adenocarcinoma patients (20), decreased survival and increased relapse in breast cancer patients (3, 21), colorectal carcinoma patients (22), lymphangiogenesis, lymphovascular invasion, and prognosis in pN0 esophageal squamous cell carcinoma (23), soft tissue sarcomas (24), both operable triple-negative and non-triple-negative breast cancer (25), hepatocellular carcinoma (26), gastric carcinoma (27), diffuse large B-cell lymphoma (28). On the other hand, SIRT1 expression is found to be associated with good prognosis for head and neck squamous cell carcinoma patients (29), and colorectal cancer (30). Therefore, the prognostic and clinicopathological significance of SIRT1 abnormal expression in cancers remain to be elucidated.

Prognostic value and clinicopathological association of SIRT1 with cancers have been analyzed in previous meta-analysis (31–36). However, the studies included in these meta-analysis were limited to mostly Asian population, single or several cancer types, or they were published several years ago (31–36). In the present study, we conducted an updated and more comprehensive meta-analysis and subgroup analysis to reveal the prognostic value and

clinicopathological association of SIRT1 abnormal expression in cancers.

METHODS

Search Strategy

We retrieved literature published in between 1966 and April 1st, 2018 by searching PubMed, EMBASE, and Cochrane Library with the keywords (1) "SIRT1" OR "sirtuin 1" OR "SIR2" OR "SIR2L1" OR "SIR2alpha" OR "silent mating type information regulation 2 homolog-1" AND (2) "tumor OR cancer OR carcinoma OR neoplasm" and using the search strategies as illustrated in **Supplementary Tables 1A–C**. We selected and evaluated all relevant studies and review articles about SIRT1 and inquired the authors for unpublished raw data. The search and selection of articles for the study were separately conducted based on a common set of criteria. The divergence in opinion were settled through discussion among ourselves.

Inclusion and Exclusion Criteria

This meta-analysis was conducted according to Meta-analysis of Observational Studies in Epidemiology (MOOSE) Checklist. Studies enrolled in this analysis satisfied the following requirements: (i) patients must be diagnosed with cancer via pathology; (ii) The expression of SIRT1 must be determined by quantitative real-time polymerase chain reaction (q-PCR), immunohistochemistry (IHC), or in situ hybridization (ISH); (iii) The correlation between SIRT1 expression and prognosis or clinicopathological features was investigated; (iv) The Hazard Ratio (HR) and its 95% confidence interval (CI) for survival indicator on the basis of SIRT1 expression level were readily available or could be calculated indirectly; (v) The most representative and most accurate study was adopted when a single sample source was used in multiple studies to avoid unnecessary cohort overlapping. Studies that have satisfied the abovementioned inclusion requirements were further ruled out if they had any of the following flaws: (i) duplicated articles or data; (ii) not human studies; (iii) review articles or letters; (iv) lack of sufficient data or information to get HR; (v) articles not written in English.

Quality Assessment of Included Studies

We used the Newcastle-Ottawa Scale (NOS) to assess the quality of each included study. Scores \geq 7 were considered high quality. We used a "star system" for case-control studies (**Supplementary Table 2**).

Data Extraction

We extracted the following data from the full texts of eligible studies: (i) the first author; (ii) publication year; (iii) characteristics of the studies, which comprised of the patients' nationality, sample size, tumor type, and clinicopathological characteristics; (iv) the assay method and cut-off value of SIRT1; (v) HRs of SIRT1 expression for OS, disease-free survival (DFS), event-free survival (EFS), recurrence-free survival (RFS), cancer-specific survival (CCS), progression-free survival (PFS); (vi) if the HR for OS, DFS, EFS, RFS, CCS and PFS were

Abbreviations: OS, overall survival; SIRT1, silent information regulator 1, sirtuin-1; HR, hazard ratio; RR, relative risk; CI, confidence interval; DFS, disease free survival; EFS, event free survival; PFS, progress-free survival; RFS, recurrence-free survival; CCS, cancer-specific survival; NAD, nicotinamide adenine dinucleotide; q-PCR, quantitative real-time polymerase chain reaction; IHC, immunohistochemistry; ISH, *in situ* hybridization; NA, not available; NOS, newcastle-ottawa scale; NSCLC, non-small cell lung cancer; EOC, epithelial ovarian cancer; TNM, tumor, node, metastasis; MOOSE, meta-analysis of observational Studies in Epidemiology.



calculated by both univariate and multivariate analyses, the latter was our first choice, given that these results were adjusted for confounding factors. If a study did not report the HR, we estimated HR and their corresponding 95% CI using the method described by Parmar et al. (37) and Tierney et al. (38). We recovered the data of Kaplan-Meier curves via the Engauge Digitizer version 9.8 (http://markummitchell.github.io/ engauge-digitizer) and calculated the HR and its 95% CI. We repeated this process three times to reduce variability. Any divergence regarding the extraction and interpretation of all data was resolved by discussion among ourselves until consensus was reached.

Statistical Analysis

All the HRs and their 95% CIs were combined to evaluate the effect of SIRT1 high expression on prognosis. If the pooled HR < 1 and their 95% CI did not overlap the invalid line in the forest plot, the high expression of SIRT1 predicted a good OS. If the 95% CI overlapped the invalid line, the combined HR was considered insignificant. Otherwise, the combined HR predicted a poor OS. The heterogeneity of the pooled results was examined via Cochrane's Q test and Higgins' I-squared, and P < 0.1 or $I^2 \ge 25\%$ was considered high heterogeneity. If P >0.1 and $I^2 < 25\%$, we ignored the influence of heterogeneity and pooled the overall result using a fixed effects model, otherwise employing the random effects model. The potential publication bias was assessed by a funnel plot, and Egger's test (39). P < 0.05 was considered significant. Statistical analysis was carried out using the "metafor" and "meta" packages of the R/BioConductor (version 3.5.1).

RESULTS

Search Results

We found 2,397 articles in PubMed, 2,460 articles in EMBASE, 20 articles in Cochrane library, and one articles through the references. We had a total of 3,733 articles after removing 1,145 duplicated articles. We then ruled out 2,953 articles which were review, letters, laboratory studies, or articles irrelevant to present research. We further excluded 717 full-text articles according to the inclusion and exclusion criteria of this study. The remaining 63 articles were finally eligible and included in this meta-analysis (**Figure 1**).

Study Demographics

The 63 eligible articles were published in between 2008 and 2017 with 63 studies that included a total of 13,138 participants from 9 countries who represented 16 cancer types and Asian and Caucasian ethnic groups (**Table 1**). The mean and median value were selected as the cut-off value in most articles. All studies measured SIRT1 expression in tumor tissue or serum via q-PCR, IHC, or ISH.

Correlation Between SIRT1 Expression and Prognosis

We performed meta-analysis of correlation between SIRT1 expression and OS, DFS, EFS, RFS, CCS, and PFS. The results and analysis of publication bias are presented in **Table 2**. The results showed that higher SIRT1 expression indicated an unfavorable OS (n = 48, HR: 1.566, 95% CI: [1.293, 1.895], P < 0.0001, $I^2 = 81.3\%$) (**Figure 2**), poor patient DFS (n = 14, HR: 1.631, 95% CI: [1.250–2.130], P = 0.0003, $I^2 = 72.6\%$, **Figure 3A**), poor

References	Study design	Country	Case (N)	Type of Cancer	Disease stage	Follow-up (Mo.) median(range)	Race	Sample type	Survival end points	Adjusted variables	NOS score
Stenzinger et al. (40)	Retrospective	Germany	129	Pancreatic ductal adenocarcinoma	2	54	Caucasian	Tissue	SO	Stage, grade	2
Li et al. (41)	Retrospective	China	91	Pancreatic ductal adenocarcinoma		NA	Asian	Tissue	NA	NA	Ø
Teramae et al. (42)	Retrospective	Japan	62	Uterine cervical cancer	=	NA	Asian	Tissue	SO	NA	7
Asaka et al. (43)	Retrospective	Japan	108	Endometrial carcinoma	\geq	NA	Asian	Tissue	OS, DFS	NA	œ
Jang et al. (44)	Retrospective	Korea	64	Ovarian cancer	\geq	29 (1–137)	Asian	Tissue	SO	Age, P53	9
Shuang et al. (18)	Retrospective	China	63	Ovarian cancer	2	NA	Asian	Serum	SO	Age, grade, FIGO stage, Iymphatic metastasis	9
Mvunta et al. (45)	Retrospective	Japan	68	Ovarian cancer	2	74	Asian	Tissue	SO	Stage, lymphovascular space invasion	2
Zhang et al. (46)	Retrospective	China	22	Osteosarcoma	NA	NA	Asian	Tissue	SO	NA	7
Feng et al. (47)	Retrospective	China	89	Osteosarcoma	≡⊥	11 (3–83)	Asian	Tissue	SO	Stage, neoadjuvant chemotherapy	œ
Kim et al. (24)	Retrospective	Korea	104	Soft tissue sarcoma	2	NA	Asian	Tissue	OS, EFS	P53, β-catenin, stage, depth of tumor, tumor necrosis, distant metastasis	ω
Noguchi et al. (29)	Retrospective	Japan	437	Head and neck squamous cell carcinoma	2	46.6 (1–174)	Asian	Tissue	RFS, CSS	Age, gender, tumor site, tumor status, lymph node status, distant metastasis, pathologic grade	ω
Yu et al. (48)	Retrospective	China	120	Laryngeal and hypopharyngeal carcinomas	≥	AN	Asian	Tissue	SO	NA	2
Batra et al. (49)	Retrospective	India	94	Retinoblastoma	\geq	59	Asian	Tissue	NA	NA	7
Chen et al. (50)	Retrospective	China	206	Esophageal squamous cell carcinoma	Ī	55.9 (5–86)	Asian	Tissue	OS, DFS	Differentiation, T status, stage, VEGF-C, peritumoral lymphatic microvessel density	2
He et al. (51)	Retrospective	China	86	Esophageal squamous cell carcinoma	=	NA	Asian	Tissue	SO	Gender, clinical stage, histological grade, lymph node metastasis	2
Feng et al. (52)	Retrospective	China	34	Pelvis chondrosarcoma	≡⊥	37.7 vs. 53.8*	Asian	Tissue	SO	NA	œ
Noh et al. (53)	Retrospective	Korea	200	Renal cell carcinoma	≡ ⊥	NA	Asian	Tissue	OS, RFS, CSS	NA	9
Jeh et al. (54)	Retrospective	Korea	102	Renal cell carcinoma	≥⊥	73	Asian	Tissue	CSS	Age, T stage, grade, metastasis, karnofsky performance status	2
Jang et al. (28)	Retrospective	Korea	104	Diffuse large B cell lymphoma	≥⊥	17.3 (1–135)	Asian	Tissue	OS, EFS	International prognostic index	ω

(Continued)

Survival Significance of SIRT1 in Various Cancers

Continued
Ξ
3LE
Ā

Frontiers in Endocrinology | www.frontiersin.org

References	Study design	Country	Case (N)	Type of Cancer	Disease stage	Follow-up (Mo.) median(range)	Race	Sample type	Survival end points	Adjusted variables	NOS score
Ren et al. (55)	Retrospective	China	45	Angioimmunoblastic T cell lymphoma	2	AA	Asian	Tissue	PFS	P53, LDH, hemoglobin, γ-Globulin, sex, age, international prognostic index score, stage	ω
Nosho et al. (56)	Retrospective	NSA	485	Colorectal cancer	2	A	Caucasian	Tissue	SS CSS	Age, year of diagnosis, sex, body Mass index, tumor location, stage, grade, microsatellite instability, the CpG island methylator phenotype	Q
Jang et al. (2)	Retrospective	Korea	497	Colorectal cancer	\geq	70.8	Asian	Tissue	OS, DFS	Histological grade, AJCC stage	Ø
Jung et al. (30)	Retrospective	Korea	349	Colorectal cancer	≥⊥	55.3	Asian	Tissue	OS	Age, location, TNM stage, Histologic grade, β-catenin	ω
Benard et al. (57)	Retrospective	Netherlands	254	Colorectal cancer		103.2	Caucasian	Tissue	OS	NA	7
Chen et al. (22)	Retrospective	China	102	Colorectal cancer	N-II	NA	Asian	Tissue	OS	Gender, age, metastasis, TNM stage	ω
Lv et al. (58)	Retrospective	China	120	Colorectal cancer		53.3 (1-78)	Asian	Tissue	SO	NA	9
Lee et al. (59)	Retrospective	China	351	Colorectal cancer		NA	Asian	Tissue	DFS, CSS	NA	7
Cheng et al. (60)	Retrospective	China	06	Colorectal cancer		NA	Asian	Tissue	OS	NA	00
Chen et al. (61)	Retrospective	China	172	Hepatocellular Carcinoma		125 (45–236)	Asian	Tissue	OS	NA	Q
Jang et al. (62)	Retrospective	Korea	154	Hepatocellular Carcinoma	21-1	NA	Asian	Tissue	OS, DFS	Stage, albumin, AFP, c-Myc, P53	α
Hao et al. (63)	Retrospective	China	66	Hepatocellular Carcinoma	\geq	NA	Asian	Tissue	SO	NA	7
Cheng et al. (64)	Retrospective	China	148	Hepatocellular Carcinoma		NA	Asian	Tissue	SO	NA	α
Li et al. (65)	Retrospective	China	72	Hepatocellular Carcinoma		NA	Asian	Tissue	OS, DFS	NA	œ
Liu et al. (66)	Retrospective	China	148	Hepatocellular Carcinoma	=	NA	Asian	Tissue	NA	NA	7
Cha et al. (27)	Retrospective	Korea	177	Gastric Cancer		NA	Asian	Tissue	OS, RFS	TNM stage	7
Feng et al. (67)	Retrospective	China	176	Gastric Cancer		NA	Asian	Tissue	OS	NA	9
Kang et al. (68)	Retrospective	Korea	452	Gastric Cancer	2	53.3 (3-83)	Asian	Tissue	SO	Lymph node metastasis, depth of invasion, lymphatic invasion, histologic grade, DBC1, cytoplasmic β-catenin	~
Noguchi et al. (19)	Retrospective	Japan	557	Gastric Cancer		69 (6–142)	Asian	Tissue	CSS	NA	7
											(Continued)

Continued
-
щ
В
₽

Frontiers in Endocrinology | www.frontiersin.org

References	Study design	Country	Case (N)	Type of Cancer	Disease stage	Follow-up (Mo.) median(range)	Race	Sample type	Survival end points	Adjusted variables	NOS score
Qiu et al. (69)	Retrospective	China	96	Gastric Cancer		31.6 (6–78)	Asian	Tissue	OS, RFS	Lymph node metastasis, Beclin1 expression	ω
Szász et al. (70)	Retrospective	Hungary	1065	Gastric Cancer		NA	Caucasian	Tissue	SO	NA	œ
Zhang et al. (71)	Retrospective	China	128	Gastric Cancer		NA	Asian	Tissue	SO	NA	80
Zhang et al. (72)	Retrospective	China	176	Gastric Cancer		NA	Asian	Tissue	SO	NA	9
Ren et al. (73)	Retrospective	NSA	348	Colorectal cancer		NA	Caucasian	Tissue	SO	NA	00
Shin et al. (74)	Retrospective	Korea	45	Ovarian cancer	AN	NA	Asian	Tissue	SO	NA	9
Zhang et al. (75)	Retrospective	China	50	Colorectal cancer	NA	NA	Asian	Tissue	NA	NA	7
Cao et al. (76)	Retrospective	China	150	Breast carcinoma		161	Asian	Tissue	OS, DFS	Lymph node metastasis,	œ
										I NM stage, EK status, PK status, Snail expression	
Jin et al. (77)	Retrospective	Korea	319	Breast carcinoma	≡⊥	NA	Asian	Tissue	OS, DFS	AJCC stage, lymphatic invasion. DCIS	9
Kim et al. (78)	Retrospective	Korea	278	Breast carcinoma	NA	63.78 (2–74)	Asian	Tissue	DFS	T stage, caspase3,	7
										lymphovascular invasion	
Lee et al. (21)	Retrospective	NSA	142	Breast carcinoma	\geq	NA	Caucasian	Tissue	OS, EFS	Stage, HER2 status, P53 expression	ω
Chung et al. (79)	Retrospective	Korea	344	Breast carcinoma		NA	Asian	Tissue	OS, DFS	T stage, lymphatic	9
										invasion, DCIS	
Derr et al. (3)	Retrospective	Netherlands	822	Breast carcinoma		11.8 (0.16–27.55)	Caucasian	Tissue	OS, DFS	NA	Q
Wu et al. (25)	Retrospective	China	134	Breast carcinoma	≡_	154	Asian	Tissue	OS, DFS	Stages, P53, Lymph nodes status	ω
Lee et al. (80)	Retrospective	Korea	688	Breast carcinoma		190.8	Asian	Tissue	NA	NA	00
Chung et al. (81)	Retrospective	Korea	427	Breast carcinoma		NA	Asian	Tissue	DFS	NA	9
Zhang et al. (82)	Retrospective	China	149	Breast carcinoma	AN	101.03 vs. 88.38*	Asian	Tissue	SO	NA	Q
Sung et al. (83)	Retrospective	Korea	28	Breast carcinoma	N-I	NA	Asian	Tissue	NA	NA	9
Gharabaghi et al.	Retrospective	Iran	40	NSCLC	NA	NA	Caucasian	Tissue	SO	Gender, age, histologic	9
(01)										graue, i stage, iyrripri node metastasis, BIRC6	
										expression	
Li et al. (20)	Retrospective	China	75	NSCLC		NA	Asian	Tissue	SO	Age, TNM stage	7
Lin et al. (84)	Retrospective	China	260	NSCLC	NA	37.1 (0–128)	Asian	Tissue	SO	NA	00
Noh et al. (85)	Retrospective	Korea	144	NSCLC	NA	NA	Asian	Tissue	NA	NA	7
Zhang et al. (86)	Retrospective	China	295	NSCLC	≥l–III	AA	Asian	Tissue	OS	Tumor stage, tumor differentiation	9
Chen et al. (23)	Retrospective	China	125	NSCLC	\geq	NA	Asian	Tissue	NA	NA	œ
Grbesa et al. (87)	Retrospective	Spain	179	NSCLC		45	Asian	Tissue	OS, RFS	Stage	9
Mo., month; NSCLC CSS, cancer-specifi growth factor recept), non-small cell lu c survival; RFS, re tor 2; BIRC6, Bac	Ing cancer; NA, no scurrence-free surv uloviral IAP repeat-	ot available; rival; LDH, lt -containing	NOS, newcastle-ottawa s actate dehydrogenase; Dt 6.	cale. FIGO, the in 3C1, deleted in b.	temational federation reast cancer 1; ER, ∈	n of gynecology . sstrogen receptc	and obstetrics; Ov nr; PR, progestero	S, overall survival; EFS ne receptor; DCIS, d	S, event-free survival; DFS, diseas uctal carcinoma in situ; HER2, huu	e-free survival; man epidermal
*The median surviva	I time of high exp	ression group vs. k	ow express	ion group.							

6

TABLE 2	Survival eff	ects of SIRT	l overexpression	and the proc	nosis of patients
	Our vivar on	0013 01 01111	1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0		pationito.

Outcome	No. of trials (patients)	HR (95%Cl) Fixed-effect estimate	<i>P</i> -value of Fixed-effect Model	HR (95%Cl) Random-effect estimate	<i>P</i> value of Random-effect Model	Heterogeneity <i>I</i> ² (%), <i>P</i> -value	<i>P</i> -value of Egger's test, Begg's test
OS	48 (9573)	1.259 (1.170–1.355)	<0.0001	1.566 (1.293–1.895)	<0.0001	81.3%, <0.0001	0.0043, 0.1884
DFS	14 (3982)	1.482 (1.308–1.679)	< 0.0001	1.631 (1.250–2.130)	0.0003	72.6%, <0.0001	0.2234, 0.2503
EFS	3 (350)	2.534 (1.602–4.009)	0.0001	2.534 (1.602-4.009)	0.0001	0.0%, 0.8557	0.1174, 0.1172
RFS	5 (1089)	1.253 (0.996–1.575)	0.0542	1.936 (0.903–4.151)	0.0898	88.90%, <0.0001	0.0037, 0.3272
CCS	6 (2132)	1.097 (0.900–1.338)	0.3591	1.229 (0.757–1.994)	0.4037	77.3%, 0.0005	0.6331, 0.3476
PFS	2 (340)	3.325 (2.762–4.003)	<0.0001	3.325 (2.762-4.003)	<0.0001	0.0%, 0.9089	NA, NA

HR, hazard ratio; *CI*, confidence interval; *OS*, overall survival; *DFS*, disease free survival; *EFS*, event free survival; *RFS*, recurrence-free survival; *CCS*, cancer-specific survival; *PFS*, progress-free survival; *NA*, not available. l^2 , index for assessing heterogeneity; value $\geq 25\%$ indicates a moderate to high heterogeneity; Egger's test, *P*-value of Egger's regression for asymmetry assessment; Begg's test, *P*-value of Begg and Mazumdar rank correlation test for asymmetry assessment. Bold italics indicate statistically significant values (P < 0.05).

EFS (n = 3, HR: 2.534, 95% CI: [1.602, 4.009], P = 0.0001, $I^2 = 0\%$, **Figure 3B**), and poor PFS (n = 2, HR: 3.325, 95% CI: [2.762, 4.003], P < 0.0001, $I^2 = 0\%$, **Figure 3C**), but not correlated with RFS of the Asian or tissue (n = 5, HR: 1.936, 95% CI: [0.903 - 4.151], P = 0.0898, $I^2 = 88.9\%$) (**Figure 3D**) or CCS (n = 6, HR: 1.229, 95% CI: [0.757–1.994], P = 0.4037, $I^2 = 77.3\%$) (**Figure 3E**).

Correlation Between SIRT1 Expression and Prognosis of Cancer Types

Cancer type subgroup analysis showed that SIRT1 overexpression was associated with a worse OS in osteosarcoma (n = 2, HR: 1.661, 95% CI: [1.162, 2.372], P = 0.0053, $I^2 = 0\%$), esophageal squamous cell carcinoma (n = 2, HR: 1.781, 95% CI: [1.197, 2.652], P = 0.0044, $I^2 = 0\%$), hepatocellular carcinoma (n = 5, HR: 1.969, 95% CI: [1.539, 2.520], P < 0.0001, $I^2 = 0\%$), breast carcinoma (n = 7, HR: 1.744, 95% CI: [1.022, 2.978], P < 0.0416, $I^2 = 70.18\%$), NSCLC (n = 5, HR: 1.929, 95% CI: [1.259, 2.957], P < 0.0025, $I^2 = 59.40\%$), whereas SIRT1 overexpression was not correlated with the OS in ovarian cancer (n = 4, HR: 1.971, 95% CI: [0.899, 4.323], P = 0.0903, $I^2 = 55.18\%$), colorectal carcinoma (n = 8, HR: 0.932, 95% CI: [0.636, 1.366], P = 0.7198, $I^2 = 82.96\%$), gastric carcinoma (n = 7, HR: 1.535, 95% CI: [0.864, 2.726], P = 0.1436, $I^2 = 87.90\%$) (**Supplementary Figure 1A**).

SIRT1 overexpression was associated with a worse DFS in colorectal cancer (n = 3, HR: 1.544, 95% CI: [1.061, 2.247], P = 0.0233, $I^2 = 68.69\%$), and breast carcinoma (n = 7, HR: 1.819, 95% CI: [1.026, 3.223], P = 0.0404, $I^2 = 84.59\%$), whereas it was not correlated with the DFS in hepatocellular carcinoma (n = 2, HR: 1.357, 95% CI: [0.872, 2.113], P = 0.1758, $I^2 = 9.41\%$) (**Supplementary Figure 2A**).

SIRT1 overexpression was correlated with RFS of Gastric Cancer (n = 2, HR: 2.734, 95% CI: [1.694, 4.413], P < 0.0001, $I^2 = 0\%$), Renal cell carcinoma (n = 1, HR: 2.233, 95% CI: [1.088, 4.583]), and NSCLC (HR: 2.698, 95% CI: [1.457, 4.996]), whereas SIRT1 overexpression was negatively correlated with RFS of head and neck squamous cell carcinoma (HR: 0.655, 95% CI: [0.478, 0.897], P = 0.0084) (**Supplementary Table 3**).

SIRT1 overexpression was correlated with CCS in head and neck squamous cell carcinoma (n = 1, HR: 0.640, 95% CI: [0.453, 0.905], P = 0.0116), gastric cancer (n = 1, HR: 1.450,

95% CI: [1.018, 2.066], P = 0.0396), and renal cell carcinoma (n = 2, HR: 1.478, 95% CI: [0.124, 17.621]), but not with CCS in colorectal cancer (HR: 1.344, 95% CI: [0.716, 2.521], P = 0.3577) (**Supplementary Table 3**).

Correlation Between SIRT1 Expression and Prognosis of Cancer in Different Countries

Analysis of country subgroups showed that high expression of SIRT1 was correlated with poor OS in China (n = 24, HR: 1.661, 95% CI: [1.339, 2.060], P < 0.0001, $I^2 = 63.03\%$), Korea (n = 12, HR: 1.902, 95% CI: [1.187, 3.047], P = 0.0075, $I^2 = 80.65\%$), Japan (n = 3, HR: 1.940, 95% CI: [1.029, 3.655], P = 0.0405, $I^2 = 0\%$), but not in USA (n = 3, HR: 1.043, 95% CI: [0.465, 2.338], P = 0.9193, $I^2 = 84.36\%$), or Netherlands (n = 2, HR: 1.003, 95% CI: [0.671, 1.498], P = 0.9893, $I^2 = 73.79\%$) (Supplementary Figure 1B).

SIRT1 overexpression was also correlated with poor DFS in China (n = 6, HR: 2.021, 95% CI: [1.612, 2.534], P < 0.0001, $I^2 = 0\%$), but not in Korea (n = 6, HR: 1.321, 95% CI: [0.773, 2.259]) (**Supplementary Figure 2B**).

SIRT1 overexpression was correlated with poor EFS in Korea (n = 2, HR: 2.714, 95% CI: [1.506, 4.894], P = 0.0009, $I^2 = 0\%$) and USA (n = 1, HR: 2.280, 95% CI: [1.098, 4.734]).

Correlation Between SIRT1 Expression and Prognosis of Cancer in Asian and Caucasian

Elevated SIRT1 expression predicted a significantly worse OS in Asian population with cancers (HR: 1.708, 95% CI: [1.406, 2.076], P < 0.0001, $I^2 = 69.59\%$) rather than in Caucasian population (HR: 1.04, 95% CI: [0.75, 1.45], P < 0.01, $I^2 = 81\%$) (**Supplementary Figure 1C**).

SIRT1 expression predicted a significantly worse DFS in Asian population with cancers (n = 13, HR: 1.683, 95% CI: [1.235, 2.294], P < 0.0010, $I^2 = 74.27\%$), whereas one article suggested that increased expression of SIRT1 is correlated with Caucasian patient DFS (HR: 1.344, 95% CI: [1.040; 1.736], P = 0.0237) (**Supplementary Figure 2C**).

Study	TE	seTE	Hazard	Ratio	HR	95%-CI	Weight (fixed)	Weight (random)
Stenzinger, 2013	0.50 (0.2499		+	1.65	[1.01; 2.69]	2.3%	2.6%
Teramae, 2014	0.41 (0.4894		<u> </u>	1.51	[0.58; 3.94]	0.6%	1.7%
Asaka, 2015	0.08 (0.8845		<u> </u>	1.08	[0.19; 6.11]	0.2%	0.9%
Jang, 2009	-0.25 (0.5397	+		0.78	[0.27; 2.25]	0.5%	1.6%
Shuang, 2015	0.36 (0.4447	_		1.43	[0.60; 3.42]	0.7%	1.9%
Mvunta, 2016	1.10 (0.4930		<u>ii</u> 1	- 3.00	[1.14; 7.88]	0.6%	1.7%
Zhang, 2016	0.46 (0.3127	-		1.59	10.86: 2.931	1.4%	2.3%
Feng. 2016	0.53 (0.2238		<u></u>	1.70	[1.10: 2.63]	2.8%	2.6%
Kim 2013	2 31 (0 6434			→ 10.06	[2 85 35 51]	0.3%	1.3%
Yu 2013	-0.58 (0 4828		1	0.56	$[0.22 \cdot 1.44]$	0.6%	1.8%
Chen 2014	0.54 (0.1020			1 71	[1 01 2 89]	2.0%	2.5%
He 2015	0.63 (0.2074			1.71	[1.01, 2.00] $[1.02 \cdot 3.47]$	1.0%	2.3%
Fong 2017	0.00 (0.0717		15	- 1.00	[1.02, 5.47]	0 10/	2.370
Nob 2012	1 22 (0.3717		1	- 2.70	[0.10, 0.19]	0.170	1 70/
long 2008	1.55 (0.4917			- 3.79	[1.45, 9.94]	0.0%	1.770
Jang, 2000	1.00 (0.0073		1		[1.45, 15.72]	0.4%	1.470
Nosho, 2009	0.07 0	0.1700	_	-	1.07		4.5%	2.8%
Jang, 2012	0.26 0	0.1602			1.30	[0.95; 1.78]	5.5%	2.8%
Jung, 2013	-0.62 (0.2865			0.54	[0.31; 0.95]	1.7%	2.4%
Benard, 2014	-0.22 (0.1746			0.80	[0.57; 1.13]	4.6%	2.8%
Chen, 2014	-0.90 (0.3117			0.41	[0.22; 0.75]	1.4%	2.3%
Lv, 2014	0.76 (0.2375			2.13	[1.34; 3.39]	2.5%	2.6%
Cheng, 2016	0.84 (0.3785		•	2.31	[1.10; 4.85]	1.0%	2.1%
Chen, 2011	0.62 (0.1954		H -	1.85	[1.26; 2.71]	3.7%	2.7%
Jang, 2012	0.76 (0.2693			2.14	[1.26; 3.63]	1.9%	2.5%
Hao, 2014	0.34 (0.3671	-		1.41	[0.69; 2.90]	1.0%	2.1%
Cheng, 2016	0.82 (0.2874		<u> </u>	2.27	[1.29; 3.99]	1.7%	2.4%
Li, 2016	1.02 (0.5288	-	•	- 2.77	[0.98; 7.81]	0.5%	1.6%
Cha, 2009	1.46 (0.4289			- 4.32	[1.86; 10.01]	0.8%	1.9%
Feng, 2011	0.51 (0.4346	-	+	1.66	[0.71; 3.89]	0.7%	1.9%
Kang, 2012	-0.40 (0.2156	-		0.67	[0.44: 1.02]	3.0%	2.7%
Qiu. 2016	0.87 (0.3221			2.39	[1.27: 4.50]	1.4%	2.3%
Szász, 2016	-0.45 (0.0872			0.64	[0.54: 0.76]	18.5%	3.0%
Zhang 2017	0.89 (0 3585	_		2 43	[1 21 4 91]	1 1%	2.2%
Zhang 2013	0.54 (0 4462	_		1 72	[0 72 4 12]	0.7%	1.9%
Ren 2017	-0.73 (0 2458			0.48	[0.30: 0.78]	2.3%	2.6%
Shin 2016	1.82 (0.7170		1	—— 6.19	[1 52: 25 23]	0.3%	1.0%
C_{20} 2014	1.52 (0.7170			4.55	[1.62; 20.20]	0.5%	1.2%
lin 2015	0.82 (0.0220			2.00	[1.00, 12.00]	0.0%	1.0%
	1 16 (0.4243		1	_ 2.20	[0.30, 5.13]	0.070	1.970
Chung 2016	0.01 (0.0421			1.01	[1.10, 9.20]	0.0 %	1.0 /0
Dorr 2014	0.010	0.0397			1.01	[U.29, 3.34]	10.0%	1.3%
	1.19 (1		1.21		10.2%	2.9%
VVU, 2012	1.28 (0.5005		1	- 3.59	[1.34; 9.57]	0.6%	1.7%
Zhang, 2016	-0.63 (0.38/1	-•		0.53	[0.25; 1.13]	0.9%	2.1%
Gnarabaghi, 2016	0.92 (0.5012	1	•	2.51	[U.94; 6.70]	0.6%	1.7%
LI, 2015	-1.06 (0.6193			0.35	[U.10; 1.17]	0.4%	1.4%
Lin, 2015	0.73 (0.2568			2.07	[1.25; 3.42]	2.1%	2.5%
Zhang, 2013	0.91 (0.1276		1 🛨	2.48	[1.93; 3.18]	8.6%	2.9%
Grbesa, 2015	0.79 (0.3436		1 1 1 1	2.19	[1.12; 4.30]	1.2%	2.2%
Fixed effect model				•	1.26	[1.17; 1.35]	100.0%	
Random effects model				•	1.57	[1.29; 1.90]		100.0%
	10.04		1 1 1	1	1			

FIGURE 2 | Forest plot of SIRT1 expression and overall survival in various cancers.

A	тг	OCTE		Hazard Patia		цр	95%-01	Weight	Weight (random)
Study	IE	SELE				пК	95%-CI	(fixea)	(random)
Asaka 2015	0.39	0 5519				1 48	[0 50 · 4 37]	1.3%	4 0%
Chen 2014	0.58	0 2652				1.78	[1 06: 3 00]	5.8%	8.0%
lang 2012	0.00	0.1390			,	1 14	[0.87:150]	21.0%	10.2%
Ly 2014	0.10	0.1000				7.1 7 7.13	[1 37 3 31]	8 1%	8 7%
	0.70	0.2240				1 66	[1.07, 0.01]	10.2%	0.7%
Lee, 2013	0.31	0.1997				1.00	[1.12, 2.40]	7 10/2	9.2 /0
	0.17	0.2304				1.19	[0.74, 1.09]	7.170	0.0% E 40/
LI, 2016	0.00	0.4255				1.90		2.2%	5.4% 4.00/
	1.57	0.5258			- '	4.83	[1.72; 13.54]	1.5%	4.2%
Jin, 2015	0.48	0.3077				1.61	[0.88; 2.94]	4.3%	7.2%
Kim, 2015	1.16	0.7453				3.19 [0.74; 13.75]	0.7%	2.6%
Chung, 2016	1.12	0.2417				3.07	[1.91; 4.93]	6.9%	8.4%
Derr, 2014	0.30	0.1307				1.34	[1.04; 1.74]	23.7%	10.4%
Wu, 2012	1.09	0.3852			:	2.96	[1.39; 6.30]	2.7%	6.0%
Chung, 2015	-0.96	0.3038	_		(0.38	[0.21; 0.69]	4.4%	7.3%
Fixed effect model				•		1.48	[1.31; 1.68]	100.0%	
Random effects mode				-		1.63	[1.25: 2.13]		100.0%
Heterogeneity: $I^2 = 73\%$, μ	0 < 0.01								
			0.1	0.5 1 2	10				
В			•	0.0 . 1				Weight	Weight
Study	TE	seTE		Hazard Ratio		HR	95%-CI	(fixed)	(random)
Kim 2013	0 90	0 3807			_	2 46	[1 17 5 10]	37.8%	37.8%
lang 2008	1 16	0.0007				3 20	[1.17, 0.10]	22.8%	22.8%
Jang, 2000	0.92	0.4304				2.20	[1.22, 0.30]	22.070	22.070
Lee, 2010	0.02	0.3720				2.20	[1.10, 4.73]	39.4%	39.4%
Fixed offerst model						0 50	14 60. 4 041	400.00/	
Fixed effect model						2.53	[1.60; 4.01]	100.0%	400.00/
Random effects mode))				1	2.53	[1.60; 4.01]		100.0%
Heterogeneity: $I^2 = 0\%$, p	0 = 0.86		1		1				
С			0.2	0.5 1 2 5	5			Weight	Weight
Study	TE	seTE		Hazard Ratio		HR	95%-CI	(fixed)	(random)
								` '	· · ·
Ren, 2016	1.28	0.6522		•		3.58	[1.00; 12.85]	2.1%	2.1%
Zhang, 2013	1.20	0.0956				3.32	[2.75; 4.00]	97.9%	97.9%
0,				T			. , .		
Fixed effect model				↓ ↓		3.33	[2.76: 4.00]	100.0%	
Random effects mode	el					2 22	2 76 4 001		100.0%
						3.33			
Heterogeneity: $l^2 = 0\%$	0 = 0.91					3.33	[2.10, 4.00]		
Heterogeneity: $I^2 = 0\%$, μ	0 = 0.91		0 1	05 1 2		3.33	[2.70, 4.00]	Woight	Waight
Heterogeneity: $I^2 = 0\%$, μ D	o = 0.91	TE	0.1	0.5 1 2	 10	э.ээ up	05%-01	Weight	Weight
Heterogeneity: / ² = 0%, µ D Study	o = 0.91 TE	seTE	0.1	0.5 1 2 Hazard Ratio	 10	HR	95%-CI	Weight (fixed)	Weight (random)
Heterogeneity: / ² = 0%, / D Study Noguchi, 2013	0 = 0.91 TE -0 42	seTE	0.1	0.5 1 2 Hazard Ratio	 10	3.33 HR 0.66	95%-CI	Weight (fixed)	Weight (random) 22.1%
Heterogeneity: / ² = 0%, / D Study Noguchi, 2013 Noh. 2013	0 = 0.91 TE -0.42	seTE 2 0.1606	0.1	0.5 1 2 Hazard Ratio	 10	0.66 2 23	95%-Cl [0.48; 0.90] [1.09: 4.58]	Weight (fixed) 53.1% 10.2%	Weight (random) 22.1% 19.1%
Heterogeneity: / ² = 0%, / D Study Noguchi, 2013 Noh, 2013 Cha, 2009	0 = 0.91 TE −0.42 0.80	seTE 2 0.1606 0 0.3668 3 0 3752	0.1	0.5 1 2 Hazard Ratio		0.66 2.23	95%-Cl [0.48; 0.90] [1.09; 4.58] [1 73: 7 53]	Weight (fixed) 53.1% 10.2% 9.7%	Weight (random) 22.1% 19.1% 18.9%
Heterogeneity: / ² = 0%, <i>µ</i> D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Oiu, 2016	0 = 0.91 TE −0.42 0.80 1.28	seTE 2 0.1606 0 0.3668 3 0.3752	0.1	0.5 1 2 Hazard Ratio	 	0.66 2.23 3.61	95%-CI [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19: 4.10]	Weight (fixed) 53.1% 10.2% 9.7% 13.2%	Weight (random) 22.1% 19.1% 18.9% 19.9%
Heterogeneity: / ² = 0%, <i>µ</i> D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbeez, 2015	0 = 0.91 TE −0.42 0.80 1.28 0.80	seTE 2 0.1606 0 0.3668 3 0.3752 0 0.3218	0.1	0.5 1 2 Hazard Ratio	 10 	0.66 2.23 3.61 2.23	95%-Cl [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00]	Weight (fixed) 53.1% 10.2% 9.7% 13.2%	Weight (random) 22.1% 19.1% 18.9% 19.9% 20.0%
Heterogeneity: / ² = 0%, <i>p</i> D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015	0 = 0.91 TE -0.42 0.80 1.28 0.80 0.99	 seTE 0.1606 0.3668 0.3752 0.3218 0.3144 	0.1	0.5 1 2 Hazard Ratio	 	0.66 2.23 3.61 2.23 2.70	95%-Cl [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00]	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.8%	Weight (random) 22.1% 19.1% 18.9% 19.9% 20.0%
Heterogeneity: / ² = 0%, / D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015	0 = 0.91 TE -0.42 0.80 1.28 0.80 0.99	 seTE 0.1606 0.3668 0.3752 0.3218 0.3144 	0.1	0.5 1 2 Hazard Ratio		0.66 2.23 3.61 2.23 2.70	95%-CI [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00]	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.8%	Weight (random) 22.1% 19.1% 18.9% 19.9% 20.0%
Heterogeneity: / ² = 0%, / D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015 Fixed effect model	= 0.91 TE −0.42 0.80 1.28 0.80 0.99	 seTE 0.1606 0.3668 0.3752 0.3218 0.3144 	0.1	0.5 1 2 Hazard Ratio		0.66 2.23 3.61 2.23 2.70 1.25	95%-Cl [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00]	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.8% 100.0%	Weight (random) 22.1% 19.1% 18.9% 19.9% 20.0%
Heterogeneity: $l^2 = 0\%$, p D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015 Fixed effect model Random effects model Hoterogeneity: $l^2 = 000'$	⇒ = 0.91 TE -0.42 0.80 1.28 0.80 0.99 	 seTE 0.1606 0.3668 0.3752 0.3218 0.3144 	0.1	0.5 1 2 Hazard Ratio	 	0.66 2.23 3.61 2.23 2.70 1.25 1.94	95%-Cl [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00] [1.00; 1.58] [0.90; 4.15]	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.8% 100.0%	Weight (random) 22.1% 19.1% 18.9% 19.9% 20.0%
Heterogeneity: / ² = 0%, <i>p</i> D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015 Fixed effect model Random effects model Heterogeneity: / ² = 89%,	p = 0.91 TE -0.42 0.80 1.28 0.80 0.99 9 9 <i>p</i> < 0.01	seTE 2 0.1606 0 0.3668 3 0.3752 0 0.3218 0 0.3144	0.1	0.5 1 2 Hazard Ratio	 	0.66 2.23 3.61 2.23 2.70 1.25 1.94	95%-Cl [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00] [1.00; 1.58] [0.90; 4.15]	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.8% 100.0% 	Weight (random) 22.1% 19.1% 18.9% 20.0% 100.0%
Heterogeneity: / ² = 0%, // D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015 Fixed effect model Random effects model Heterogeneity: / ² = 89%, E Study	p = 0.91 TE -0.42 0.80 1.28 0.80 0.99 P p < 0.01 TE	seTE	0.1	0.5 1 2 Hazard Ratio	 5	0.66 2.23 3.61 2.23 2.70 1.25 1.94	95%-Cl [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00] [1.00; 1.58] [0.90; 4.15]	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.8% 100.0% Weight (fixed)	Weight (random) 22.1% 19.1% 18.9% 20.0% 100.0% Weight (random)
Heterogeneity: $l^2 = 0\%$, p D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015 Fixed effect model Random effects model Heterogeneity: $l^2 = 89\%$, E Study	p = 0.91 TE -0.42 0.80 1.28 0.80 0.99 P p < 0.01 TE	 seTE 0.1606 0.3668 0.3752 0.3218 0.3144 seTE 	0.1	0.5 1 2 Hazard Ratio	 5	0.66 2.23 3.61 2.23 2.70 1.25 1.94 HR	95%-Cl [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00] [1.46; 5.00] [1.00; 1.58] [0.90; 4.15] 95%-Cl	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.8% 100.0% Weight (fixed)	Weight (random) 22.1% 19.1% 18.9% 20.0% 100.0% Weight (random)
Heterogeneity: $l^2 = 0\%$, p D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015 Fixed effect model Random effects model Heterogeneity: $l^2 = 89\%$, E Study Noguchi, 2013	p = 0.91 TE -0.42 0.80 1.28 0.80 0.99 P p < 0.01 TE -0.45	 seTE 0.1606 0.3668 0.3752 0.3218 0.3144 seTE 0.1768 	0.1	0.5 1 2 Hazard Ratio	 5	 3.33 HR 0.66 2.23 3.61 2.23 2.70 1.25 1.94 HR 0.64 	95%-Cl [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00] [1.00; 1.58] [0.90; 4.15] 95%-Cl [0.45; 0.91]	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.8% 100.0% Weight (fixed) 32.8%	Weight (random) 22.1% 19.1% 18.9% 20.0% 100.0% Weight (random) 22.8%
Heterogeneity: $l^2 = 0\%$, p D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015 Fixed effect model Random effects model Heterogeneity: $l^2 = 89\%$, E Study Noguchi, 2013 Noh, 2013	p = 0.91 TE -0.42 0.80 1.28 0.80 0.99 P p < 0.01 TE -0.45 1.49	 seTE 0.1606 0.3668 0.3752 0.3218 0.3144 seTE 0.1768 0.6264 	0.1	0.5 1 2 Hazard Ratio	 	 HR 0.66 2.23 3.61 2.23 2.70 1.25 1.94 HR 0.64 4.42 	95%-Cl [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00] [1.00; 1.58] [0.90; 4.15] 95%-Cl [0.45; 0.91] [1.29; 15.08]	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.8% 100.0% Weight (fixed) 32.8% 2.6%	Weight (random) 22.1% 19.1% 18.9% 20.0% 100.0% Weight (random) 22.8% 9.7%
Heterogeneity: $l^2 = 0\%$, p D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015 Fixed effect model Random effects model Heterogeneity: $l^2 = 89\%$, E Study Noguchi, 2013 Noh, 2013 Jeh, 2017	p = 0.91 TE -0.42 0.80 1.28 0.80 0.99 P p < 0.01 TE -0.45 1.49 -1.07	 seTE 0.1606 0.3668 0.3752 0.3218 0.3144 seTE 0.1768 0.6264 1.1518 	0.1	0.5 1 2 Hazard Ratio	 	 3.33 HR 0.66 2.23 3.61 2.23 2.70 1.25 1.94 HR 0.64 4.42 0.34 	95%-CI [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00] [1.00; 1.58] [0.90; 4.15] 95%-CI [0.45; 0.91] [1.29; 15.08] [0.04; 3.28]	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.8% 100.0% Weight (fixed) 32.8% 2.6% 0.8%	Weight (random) 22.1% 19.1% 18.9% 20.0% 100.0% Weight (random) 22.8% 9.7% 3.9%
Heterogeneity: $l^2 = 0\%$, p D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015 Fixed effect model Random effects model Heterogeneity: $l^2 = 89\%$, E Study Noguchi, 2013 Noh, 2013 Jeh, 2017 Nosbo, 2009	p = 0.91 TE -0.42 0.80 0.99 p < 0.01 TE -0.45 1.49 -1.07 -0.02	 seTE 0.1606 0.3668 0.3752 0.3218 0.3144 seTE 0.1768 0.6264 1.1518 0.2444 	0.1	0.5 1 2 Hazard Ratio	- - - 5	HR 0.66 2.23 3.61 2.23 2.70 1.25 1.94 HR 0.64 4.42 0.34 0.98	95%-Cl [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00] [1.00; 1.58] [0.90; 4.15] 95%-Cl [0.45; 0.91] [1.29; 15.08] [0.04; 3.28] [0.61; 1.58]	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.8% 100.0% Weight (fixed) 32.8% 2.6% 0.8% 0.8%	Weight (random) 22.1% 19.1% 18.9% 20.0% 100.0% Weight (random) 22.8% 9.7% 3.9% 20.6%
Heterogeneity: $l^2 = 0\%$, p D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015 Fixed effect model Random effects model Heterogeneity: $l^2 = 89\%$, E Study Noguchi, 2013 Noh, 2013 Jeh, 2017 Nosho, 2009 Lee, 2015	p = 0.91 TE -0.42 0.80 1.28 0.99 p < 0.01 TE -0.45 1.49 -1.07 -0.02 0.62	 seTE 0.1606 0.3668 0.3752 0.3218 0.3144 seTE 0.1768 0.6264 1.1518 0.2444 0.2547 	0.1	0.5 1 2 Hazard Ratio	 	HR 0.66 2.23 3.61 2.23 2.70 1.25 1.94 HR 0.64 4.42 0.34 0.98 1.86	95%-Cl [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00] [1.00; 1.58] [0.90; 4.15] 95%-Cl [0.95; 0.91] [1.29; 15.08] [0.04; 3.28] [0.04; 3.28] [0.04; 3.28]	Weight (fixed) 53.1% 10.2% 9.7% 13.8% 100.0% Weight (fixed) 32.8% 2.6% 0.8% 17.1% 15.3%	Weight (random) 22.1% 19.1% 18.9% 20.0% 100.0% Weight (random) 22.8% 9.7% 3.9% 20.6% 20.2%
Heterogeneity: $l^2 = 0\%$, p D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015 Fixed effect model Random effects model Random effects model Heterogeneity: $l^2 = 89\%$, E Study Noguchi, 2013 Noh, 2013 Jeh, 2017 Nosho, 2009 Lee, 2015 Noguchi 2014	p = 0.91 TE -0.42 0.80 1.28 0.80 0.99 p < 0.01 TE -0.45 1.49 -1.07 -0.20 0.62 0.62	 seTE 0.1606 0.3668 0.3752 0.3218 0.3144 seTE 0.1768 0.6264 1.1518 0.2444 0.2484 0.1805 	0.1	0.5 1 2 Hazard Ratio	 	HR 0.66 2.23 3.61 2.23 2.70 1.25 1.94 HR 0.64 4.42 0.34 0.98 1.86 1.45	95%-Cl [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00] [1.00; 1.58] [0.90; 4.15] 95%-Cl [0.45; 0.91] [1.29; 15.08] [0.04; 3.28] [0.4; 3.28] [0.4; 3.28] [1.02; 3.09]	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.2% 13.8% 100.0% Weight (fixed) 32.8% 2.6% 0.8% 17.1% 15.3%	Weight (random) 222.1% 19.1% 18.9% 20.0% 20.0% 100.0% Weight (random) 22.8% 9.7% 3.9% 20.6% 20.2%
Heterogeneity: $l^2 = 0\%$, p D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015 Fixed effect model Random effects model Random effects model Heterogeneity: $l^2 = 89\%$, E Study Noguchi, 2013 Noh, 2013 Jeh, 2017 Nosho, 2009 Lee, 2015 Noguchi, 2014	p = 0.91 TE -0.42 0.80 1.28 0.80 0.99 p < 0.01 TE -0.45 1.49 -1.07 -0.22 0.62 0.37	 seTE 0.1606 0.3668 0.3752 0.3218 0.3144 seTE 0.1768 0.6264 1.1518 0.2444 0.2444 0.2587 0.1805 	0.1	0.5 1 2 Hazard Ratio	 5	HR 0.66 2.23 3.61 2.23 2.70 1.25 1.94 HR 0.64 4.42 0.34 0.98 1.86 1.45	95%-Cl [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00] [1.00; 1.58] [0.90; 4.15] 95%-Cl [0.45; 0.91] [1.29; 15.08] [0.04; 3.28] [0.4; 3.28] [0.4; 3.28] [1.12; 3.09] [1.02; 2.07]	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.8% 100.0% Weight (fixed) 32.8% 2.6% 0.8% 17.1% 15.3% 31.4%	Weight (random) 222.1% 19.1% 18.9% 20.0% 20.0% 100.0% Weight (random) 22.8% 9.7% 3.9% 20.6% 20.2% 22.7%
Heterogeneity: $l^2 = 0\%$, p D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015 Fixed effect model Random effects model Random effects model Heterogeneity: $l^2 = 89\%$, E Study Noguchi, 2013 Noh, 2013 Jeh, 2017 Nosho, 2009 Lee, 2015 Noguchi, 2014 Fixed effect model	p = 0.91 TE -0.42 0.80 1.28 0.80 0.99 p < 0.01 TE -0.45 1.49 -1.07 -0.22 0.62 0.37	 seTE 0.1606 0.3668 0.3752 0.3218 0.3144 seTE 0.1768 0.6264 1.1518 0.2444 0.2444 0.2587 0.1805 	0.1	0.5 1 2 Hazard Ratio	 	HR 0.66 2.23 3.61 2.23 2.70 1.25 1.94 HR 0.64 4.42 0.34 0.98 1.86 1.45 1.10	95%-CI [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00] [1.00; 1.58] [0.90; 4.15] 95%-CI [0.45; 0.91] [1.29; 15.08] [0.04; 3.28] [0.4; 3.28] [0.4; 3.28] [0.4; 3.28] [1.12; 3.09] [1.02; 2.07] [0.90; 1.341]	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.2% 13.8% 100.0% Weight (fixed) 32.8% 2.6% 0.8% 17.1% 15.3% 31.4%	Weight (random) 222.1% 19.1% 18.9% 20.0% 20.0% 100.0% Weight (random) 22.8% 9.7% 3.9% 20.6% 20.2% 22.7%
Heterogeneity: $l^2 = 0\%$, p D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015 Fixed effect model Random effects model Heterogeneity: $l^2 = 89\%$, E Study Noguchi, 2013 Noh, 2013 Jeh, 2017 Nosho, 2009 Lee, 2015 Noguchi, 2014 Fixed effect model Random effects model Random effects model	p = 0.91 TE -0.42 0.80 1.28 0.80 0.99 p < 0.01 TE -0.45 1.49 -1.07 -0.02 0.62 0.37	 seTE 0.1606 0.3668 0.3752 0.3218 0.3144 seTE 0.1768 0.6264 1.1518 0.2444 0.2587 0.1805 	0.1	0.5 1 2 Hazard Ratio		HR 0.66 2.23 3.61 2.23 2.70 1.25 1.94 HR 0.64 4.42 0.34 0.98 1.86 1.45 1.10 1.23	95%-Cl [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00] [1.00; 1.58] [0.90; 4.15] 95%-Cl [0.45; 0.91] [1.29; 15.08] [0.04; 3.28] [0.04; 3.28] [0.04; 3.28] [1.12; 3.09] [1.12; 3.09] [1.02; 2.07] [0.90; 1.34] [0.76; 1.99]	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.8% 100.0% Weight (fixed) 32.8% 2.6% 0.8% 17.1% 15.3% 31.4%	Weight (random) 222.1% 19.1% 18.9% 20.0% 100.0% Weight (random) 222.8% 9.7% 3.9% 20.6% 20.2% 22.7%
Heterogeneity: $l^2 = 0\%$, p D Study Noguchi, 2013 Noh, 2013 Cha, 2009 Qiu, 2016 Grbesa, 2015 Fixed effect model Random effects mode Heterogeneity: $l^2 = 89\%$, E Study Noguchi, 2013 Noh, 2013 Jeh, 2017 Nosho, 2009 Lee, 2015 Noguchi, 2014 Fixed effect model Random effects mode Heterogeneity : $l^2 = 77\%$	p = 0.91 TE -0.42 0.80 1.28 0.80 0.99 p < 0.01 TE -0.45 1.49 -1.07 -0.02 0.62 0.37 I p < 0.01	seTE 0.1606 0.3668 0.3752 0.3218 0.3144 seTE 0.1768 0.6264 1.1518 0.2444 0.2587 0.1805	0.1	0.5 1 2 Hazard Ratio	 	HR 0.66 2.23 3.61 2.23 2.70 1.25 1.94 HR 0.64 4.42 0.98 1.86 1.45 1.10 1.23	95%-Cl [0.48; 0.90] [1.09; 4.58] [1.73; 7.53] [1.19; 4.19] [1.46; 5.00] [1.00; 1.58] [0.90; 4.15] 95%-Cl [0.45; 0.91] [1.29; 15.08] [0.04; 3.28] [0.04; 3.28] [0.04; 3.28] [0.12; 3.09] [1.12; 3.09] [1.02; 2.07] [0.90; 1.34] [0.76; 1.99]	Weight (fixed) 53.1% 10.2% 9.7% 13.2% 13.8% 100.0% Weight (fixed) 32.8% 2.6% 0.8% 17.1% 15.3% 31.4%	Weight (random) 22.1% 19.1% 18.9% 20.0% 100.0% Weight (random) 22.8% 9.7% 3.9% 20.6% 20.2% 22.7%

FIGURE 3 | Forest plots of main survival outcomes compared SIRT1 overexpression with underexpression. (A) DFS. (B) EFS. (C) PFS. (D) RFS. (E) CCS.

TABLE 3 The associations of SIRT1 overexpression with the clinicopathological characteristics of the study patients.	
---	--

Clinicopathological parameters	No. of trials (patients)	RR (95%Cl) Fixed-effect estimate	P-value of Fixed-effect Model	RR (95%Cl) Random-effect estimate	<i>P</i> -value of Random-effect Model	Heterogeneity / ² (%), <i>P</i> -value	<i>P</i> -value of Egger's test, Begg's test
Tumor stage	33 (5857)	1.133 (1.062–1.209)	0.0002	1.299 (1.114–1.514)	0.0008	77.4%, <0.0001	0.0070, 0.1827
Lymphatic metastasis	29 (6354)	1.046 (0.995–1.100)	0.0763	1.172 (1.010–1.360)	0.0363	86.3%, <0.0001	0.0637, 0.4308
Distant metastasis	14 (2632)	1.607 (1.312–1.968)	<0.0001	1.562 (1.022–2.387)	0.0392	71.0%, <0.0001	0.6780, 0.3520
Tumor size	21 (2469)	1.143 (1.050–1.245)	0.0021	1.101 (0.984–1.232)	0.0924	41.7%, 0.0241	0.1660, 0.2047
Depth of tumor invasion	19 (4689)	1.036 (0.982–1.093)	0.1912	1.113 (0.985–1.258)	0.0852	81.70%, <0.0001	0.0903, 0.1955
Differentiation	28 (5740)	1.010 (0.940–1.085)	0.7841	1.055 (0.931–1.196)	0.3996	63.10%, <0.0001	0.1170, 0.3847
Age	38 (7223)	1.052 (1.004–1.102)	0.0345	1.043 (0.973–1.118)	0.2373	43.50%, 0.0027	0.5651, 0.8308
Gender	34 (6129)	1.003 (0.967–1.040)	0.8739	0.991 (0.950–1.035)	0.6858	35.00%, 0.0247	0.1727, 0.3353

RR, relative risk; CI, confidence interval; l^2 , index for assessing heterogeneity; value $\geq 25\%$ indicates a moderate to high heterogeneity; Egger's test, P-value of Egger's regression for asymmetry assessment; Begg's test: P-value of Begg and Mazumdar rank correlation test for asymmetry assessment. Bold italics indicate statistically significant values (P < 0.05).

Correlation Between SIRT1 Expression and Clinicopathological Characteristics

We performed an analysis of the association of SIRT1 expression with clinicopathological characteristics (Table 3). The results showed that overexpression of SIRT1 was significantly correlated with TNM stage. Higher SIRT1 expression indicated high TNM stage for various malignancies (n = 33, RR: 1.299; 95% CI: [1.114, 1.514], P = 0.0008, $I^2 = 77.4\%$, Figure 4). SIRT1 expression was significantly correlated with lymphatic metastasis (n = 29, RR: 1.172, 95% CI: [1.010, 1.360], P = 0.0363, $I^2 = 86.3\%$, Figure 5), distant metastasis (*n* = 14, RR: 1.562, 95% CI: [1.022, 2.387], P = 0.0392, $I^2 = 71.0\%$, Figure 6), but not correlated with tumor size (RR:1.101, 95% CI [0.984-1.232], $I^2 = 41.7\%$), depth of tumor invasion (RR: 1.113, 95% CI [0.985–1.258], $I^2 =$ 81.7%), differentiation (RR: 1.055, 95% CI [0.931–1.196], $I^2 =$ 63.1%), gender (RR: 0.991, 95% CI [0.950–1.035], $I^2 = 35.0\%$), or age (RR: 1.043, 95% CI [0.973–1.118], $I^2 = 43.5\%$) (Table 3, Supplementary Figure 3).

Correlation Between SIRT1 Expression and Clinicopathological Characteristics of Cancers Types

We performed analysis of correlation between SIRT1 expression and clinicopathological characteristics of cancers types (**Supplementary Table 4**). The results showed that SIRT1 overexpression was associated with a higher TNM stage in hepatocellular carcinoma (n = 6, RR: 1.611, 95% CI: [1.185, 2.188], P = 0.0023, $I^2 = 55.30\%$), but not correlated with the TNM stage in pancreatic ductal adenocarcinoma (n = 2, RR: 2.275, 95% CI: [0.928, 5.579], P = 0.0725, $I^2 = 0\%$), ovarian cancer (n = 2, RR: 0.820, 95% CI: [0.561, 1.201], P = 0.3082, $I^2 = 3.18\%$), colorectal cancer (n = 4, RR: 1.146, 95% CI: [0.817, 1.608], P = 0.4290, $I^2 = 70.90\%$), gastric cancer (n = 6, RR: 1.264, 95% CI: [0.823, 1.942], P = 0.2842, $I^2 = 92.48\%$), breast carcinoma (n = 5, RR: 1.411, 95% CI: [0.846, 2.356], P = 0.1873, $I^2 = 65.10\%$), or NSCLC (n = 2, RR: 1.389, 95\% CI: [0.661, 2.917], P = 0.3853, $I^2 = 16.51\%$) (**Supplementary Figure 4A**). SIRT1 overexpression was associated with distant metastasis in pancreatic ductal adenocarcinoma (n = 2, RR: 2.046, 95% CI: [1.153, 3.631], P = 0.0144, $I^2 = 0\%$) and breast carcinoma (n = 2, RR: 3.257, 95% CI: [1.777, 5.970], P = 0.0001, $I^2 = 0\%$), but not in colorectal cancer (n = 3, RR: 1.140, 95% CI: [0.444, 2.923], P =0.7857, $I^2 = 80.57\%$) or gastric cancer (n = 2, RR: 1.316, 95% CI: [0.679, 2.551], P = 0.4160, $I^2 = 0\%$) (**Supplementary Figure 5A**).

Correlation Between SIRT1 Expression and Clinicopathological Characteristics of Cancers in Different Countries

We performed analysis of correlation between SIRT1 expression and clinicopathological characteristics of cancers in different countries (**Supplementary Table 4**). The results showed that SIRT1 overexpression was associated with a higher TNM stage $(n = 17, \text{ RR: } 1.638, 95\% \text{ CI: } [1.404, 1.911], P < 0.0001, I^2 =$ 41.16%) (**Supplementary Figure 4B**) and lymphatic metastasis in China (n = 11, RR: 1.411, 95% CI: [1.155, 1.724], P = 0.0007, $I^2 = 68.48\%$), and not with lymphatic metastasis in Japan (n = 3, RR: 0.964, 95% CI: [0.657, 1.415]), or Korea (n = 12, RR: 1.166, 95% CI: [0.898, 1.516]) (**Supplementary Figure 6A**).

Correlation Between SIRT1 Expression and Clinicopathological Characteristics of Cancers in Asian and Caucasian

We performed analysis of correlation between SIRT1 expression and clinicopathological characteristics of cancers in Asian and Caucasian (**Supplementary Table 4**). The results showed that SIRT1 overexpression predicted a significantly higher TNM stage in Asian population with cancers (n = 30, RR: 1.323, 95% CI: [1.124, 1.559], P = 0.0008, $I^2 = 78.76\%$) rather than that in Caucasian population (n = 3, RR: 0.919, 95% CI: [0.744, 1.136], P = 0.4352, $I^2 = 0\%$) (**Supplementary Figure 4C**). However, publication bias was suspected based on the Egger's test (P =0.0070) rather than Begg's test (P = 0.1827).

Elevated SIRT1 expression predicted a significantly distant metastasis in Caucasian population with cancers (n = 2, RR:

Higi Study	h expres Event	ssion Size	Low ex Event	<pression Size</pression 	n Risk	Ratio	RR	95%-CI	Weight (fixed)	Weight (random)
									· ·	, ,
Stenzinger, 2013	4	36	5	93		10 •	2.07	[0.59; 7.27]	0.3%	1.2%
Li, 2016	7	39	3	42	_	•	2.51	[0.70; 9.04]	0.3%	1.1%
Jang, 2009	18	33	16	22		ł.	0.75	[0.50; 1.12]	2.0%	3.8%
Mvunta, 2016	4	11	17	57		<u> </u>	1.22	[0.51; 2.93]	0.6%	1.9%
Kim, 2013	42	75	9	29			1.80	[1.01; 3.22]	1.3%	3.0%
Noguchi, 2013	196	348	61	89	+	8	0.82	[0.69; 0.97]	10.0%	4.9%
Yu, 2013	30	37	48	83			1.40	[1.10; 1.78]	3.0%	4.6%
Feng, 2017	18	25	1	9		•	6.48	[1.00; 41.79]	0.2%	0.6%
Noh, 2013	22	119	16	81	-	<u>1</u>	0.94	[0.52; 1.67]	2.0%	3.0%
Jang, 2008	41	77	14	27	-		1.03	[0.68: 1.56]	2.1%	3.8%
Nosho, 2009	69	165	132	280			0.89	[0.71; 1.10]	10.0%	4.7%
Juna. 2013	105	235	56	114	-	1	0.91	[0.72: 1.15]	7.7%	4.7%
Lee. 2015	37	68	12	38			1.72	[1.03: 2.89]	1.6%	3.3%
Chen. 2011	64	95	41	77			1.27	[0.98: 1.63]	4.6%	4.6%
Jang 2012	12	55	24	99		<u>E</u>	0.90	[0 49 1 66]	1.8%	2.9%
Hao 2014	30	76	4	23	-	<u>.</u>	2 27	[0 89 5 77]	0.6%	1.8%
Cheng 2016	31	48	. 12	39			2 10	[1 25: 3 51]	1.4%	3.3%
Li 2016	23	41	5	31			3 48	[1 49 8 11]	0.6%	2.0%
Liu 2016	45	76	26	72			1 64	[1 14 2 35]	2.7%	4 1%
Cha 2009	79	130	14	47		1	2.04	[1 29 3 23]	2.1%	3.6%
Eeng 2011	66	81	62	95		1	1 25	[1.20, 0.20]	5.9%	4 9%
Kang 2012	58	255	100	197		8	0.45	[0.34: 0.58]	11.6%	4.5%
Noguchi 2014	161	345	82	212		16	1 21	[0.04, 0.00]	10.4%	4.8%
	49	53	24	43		<u>19</u>	1.66	[1 26 2 18]	2.7%	4.5%
Zhang 2017	30	68	27	15			2.87	[1.20, 2.10]	0.5%	1.6%
Zhang 2015	16	30	1	20		<u>}</u>	2.07	[1.02, 0.00]	0.5%	1.0%
Cao 2014	18	116		20			1 32	[1.04, 0.01]	0.0%	1.0%
	10	82		40		ij	1.02	[0.40, 0.00]	0.070	1.0%
Mu 2012	10	72	17	40 62		8	2 1 2	[0.41, 3.03]	1 0%	3.6%
	40	207	111	471			2.10	[1.39, 3.41]	0.00/	3.0 /0
Lee, 2010 Sung. 2010	12	207	144	4/1		d .	_ 202	[0.72, 1.20]	9.0%	4.0%
Sung, 2010	13	23	1	0 16			- 2.03 2.55		0.2%	0.0%
LI, 2015	17	50	2	15			2.00		0.3%	1.0%
Cnen, 2014	1	26	24	99			1.11	[0.54; 2.29]	1.0%	2.4%
Fixed effect model		3197		2660		•	1.13	[1.06; 1.21]	100.0%	
Random effects model	l					•	1.30	[1.11; 1.51]		100.0%
Heterogeneity: $I^2 = 77\%$, μ	0.01 > c				I I					

FIGURE 4 | Forest plots of tumor stage compared SIRT1 overexpression with underexpression.

3.830, 95% CI: [1.445, 10.154], P = 0.0069, $I^2 = 0\%$), but not in Asian population (n = 12, RR: 1.422, 95% CI: [0.913, 2.217], P = 0.1198, $I^2 = 72.85\%$) (Supplementary Figure 5B).

Elevated SIRT1 expression predicted a significantly higher lymphatic metastasis in Asian population with cancers (n = 29, RR: 1.239, 95% CI: [1.056, 1.453], P = 0.0086, $I^2 = 86.81\%$), but not correlated with lymphatic metastasis in Caucasian population (n = 3, RR: 0.777, 95% CI: [0.526, 1.147], P = 0.2040, $I^2 = 76.11\%$) (**Supplementary Figure 6B**).

Meta-Regression Analysis of Heterogeneity for Overall Survival and Publication Bias

We performed a meta-regression to explore the source of high heterogeneity for OS (**Table 4**). All potential factors could not significantly explain heterogeneity in the meta-analyses of the SIRT1 expression with survival outcomes in the *post-hoc* analysis, with the exception of ethnicity (**Supplementary Table 5**). Meta-regression analysis demonstrated a statistically significant correlation between ethnicity and OS (P = 0.022). From the meta-regression result, we conducted a subgroup analysis with groups of patients Asian or Caucasian (**Supplementary Figure 1C**). This subgroup analysis demonstrated a significantly lower heterogeneity value in Asian group (n = 40, RR: 1.708, 95% CI: [1.406, 2.076], $I^2 = 69.59\%$), which suggests that the high SIRT1 expression has stronger efficacy in the Asian population than the Caucasian population.

Meta-regression also used to explore the source of high heterogeneity for clinicopathological outcomes (Further details are provided in **Supplementary Table 6**). As to tumor stage, meta-regression analysis demonstrated a statistically significant correlation between tumor stage and country (P < 0.05), published year (P = 0.0169), and sample size (P = 0.0004). This subgroup analysis demonstrated a significantly lower heterogeneity value in China (n = 17, RR: 1.638, 95% CI: [1.404, 1.911], $I^2 = 41.16\%$), which suggests that the high SIRT1

Н	igh expres	ssion	Low ex	xpressio	n			Weight	Weight
Study	Event	Size	Event	Size	Risk Ratio	RR	95%-CI	(fixed)	(random)
Stenzinger, 2013	27	36	71	93	-	0.98	[0.79; 1.22]	2.9%	4.4%
Li, 2016	22	39	16	52		1.83	[1.12; 3.00]	1.0%	3.2%
Mvunta, 2016	2	11	10	57		1.04	[0.26; 4.10]	0.2%	0.9%
Kim, 2013	12	74	3	30		1.62	[0.49; 5.34]	0.3%	1.2%
Noguchi, 2013	156	348	52	89		0.77	[0.62; 0.95]	6.1%	4.5%
Yu, 2013	23	37	29	83		1.78	[1.21; 2.62]	1.3%	3.7%
He, 2015	34	54	13	32	1: .	1.55	[0.97; 2.47]	1.2%	3.3%
Noh, 2013	1	119	1	81 —		0.68	[0.04; 10.73]	0.1%	0.3%
Jang, 2012	103	208	178	289		0.80	[0.68; 0.95]	10.9%	4.6%
Jung, 2013	98	235	47	114	÷	1.01	[0.78; 1.32]	4.6%	4.2%
Lv, 2014	23	32	41	88		1.54	[1.13; 2.11]	1.6%	4.0%
Cha, 2009	95	130	18	47		1.91	[1.31; 2.78]	1.9%	3.7%
Feng, 2011	67	81	61	95		1.29	[1.08; 1.54]	4.1%	4.6%
Kang, 2012	96	255	213	293		0.52	[0.44; 0.62]	14.5%	4.6%
Noguchi, 2014	172	255	173	302		1.18	[1.03; 1.34]	11.6%	4.7%
Qiu, 2016	50	53	22	43		1.84	[1.37; 2.49]	1.8%	4.1%
Zhang, 2017	46	68	5	15	1: 1:	2.03	[0.97; 4.23]	0.6%	2.2%
Zhang, 2015	13	30	6	20		1.44	[0.66; 3.17]	0.5%	2.1%
Cao, 2014	46	116	21	34	- - -	0.64	[0.45; 0.91]	2.4%	3.9%
Jin, 2015	39	87	63	232		1.65	[1.21; 2.26]	2.5%	4.0%
Kim, 2015	78	117	133	161	.	0.81	[0.70; 0.93]	8.2%	4.7%
Lee, 2010	36	82	16	20	- - -	0.55	[0.40; 0.76]	1.9%	4.0%
Chung, 2016	58	146	52	198		1.51	[1.11; 2.06]	3.2%	4.1%
Lee, 2016	120	206	237	470	•	1.16	[1.00; 1.34]	10.6%	4.7%
Sung, 2010	14	23	1	5		3.04	[0.51; 18.11]	0.1%	0.6%
Gharabaghi, 2016	16	27	9	13		0.86	[0.53; 1.38]	0.9%	3.3%
Lin, 2015	23	36	133	224	- -	1.08	[0.82; 1.41]	2.7%	4.2%
Noh, 2013	58	96	12	48	·	2.42	[1.44; 4.05]	1.2%	3.1%
Chen, 2014	12	26	26	99		1.76	[1.03; 2.99]	0.8%	3.0%
Fixed effect model		3027		3327		1.05	[1.00; 1.10]	100.0%	
Random effects mod	lel				•	1.17	[1.01; 1.36]		100.0%
Heterogeneity: I ² = 86%	b, <i>p</i> < 0.01						_		
					0.1 0.5 1 2 10				

FIGURE 5 | Forest plots of lymphatic metastasis compared SIRT1 overexpression with underexpression.

	High expres	ssion	Low ex	pression				Weight	Weight
Study	Event	Size	Event	Size	Risk Ratio	RR	95%-CI	(fixed)	(random)
Stenzinger, 2013	2	36	2	93		2.58 [0.	38; 17.65]	0.9%	3.5%
Li, 2016	18	39	12	52	- <u>}</u> -	2.00 [1	.10; 3.65]	8.8%	9.6%
Feng, 2016	39	49	17	40		1.87 [1	.27; 2.76]	15.9%	10.8%
Kim, 2013	29	74	2	30	1 	- 5.88 [1.	50; 23.10]	2.4%	5.3%
Feng, 2017	8	25	7	9		0.41 [0	.21; 0.80]	8.8%	9.2%
Jeh, 2017	2	16	22	86	∎ <u> </u>	0.49 [0	.13; 1.88]	5.9%	5.4%
Jang, 2012	7	208	12	289		0.81 [0	.32; 2.02]	8.5%	7.7%
Jung, 2013	23	235	17	114		0.66 [0	.37; 1.18]	19.5%	9.7%
Lv, 2014	12	32	12	88	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	2.75 [1	.38; 5.48]	5.4%	9.1%
Cha, 2009	3	130	1	47		1.08 [0.	12; 10.17]	1.2%	2.8%
Noguchi, 2014	24	345	11	212		1.34 [0	.67; 2.68]	11.6%	9.0%
Lee, 2010	27	82	3	40	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	4.39 [1.	42; 13.61]	3.4%	6.5%
Wu, 2012	26	72	8	64	+	2.89 [1	.41; 5.92]	7.2%	8.9%
Chen, 2014	2	26	1	99	++++++	—— 7.62 [0.	72; 80.76]	0.4%	2.5%
Fixed effect model		1369		1263	•	1.61 [1.	31; 1.97]	100.0%	
Random effects m	odel					1.56 [1.	02; 2.39]		100.0%
Heterogeneity: $I^2 = 7$	1%, <i>p</i> < 0.01					-			
. ,					0.1 0.51 2 10				

FIGURE 6 | Forest plots of distant metastasis compared SIRT1 overexpression with underexpression.

TABLE 4 | Meta-regression analysis of heterogeneity for overall survival.

Moderators	Variables of regression	HR _{interaction} (95% CI)	P-value of regression	l ²	Cochrane Q (P-value)
Year	Year	1.001(0.990-1.012)	0.840	81.69%	<0.001
Sample size	Sample size	2.578(0.674-9.860)	0.166	43.18%	0.152
Follow up	Follow up	0.741(0.076-7.236)	0.796	72.89%	< 0.001
Country	Intercept	1.648(1.291-2.104)	<0.001	71.56%	< 0.001
	Germany	1.001(0.334-2.997)	0.999	71.56%	< 0.001
	Hungary	0.388(0.143-1.051)	0.063	71.56%	< 0.001
	Iran	1.523(0.380–6.105)	0.553	71.56%	< 0.001
	Japan	1.134(0.463–2.775)	0.783	71.56%	< 0.001
	Korea	1.079(0.695–1.675)	0.736	71.56%	< 0.001
	Netherlands	0.600(0.285-1.263)	0.178	71.56%	< 0.001
	Spain	1.330(0.405–4.374)	0.638	71.56%	< 0.001
	USA	0.605(0.302-1.215)	0.158	71.56%	< 0.001
Tumor type	Intercept	1.716(1.055–2.792)	0.030	75.22%	< 0.001
	Colorectal cancer	0.543(0.293-1.008)	0.053	75.22%	< 0.001
	Diffuse large B cell lymphoma	2.786(0.559–13.892)	0.211	75.22%	< 0.001
	Endometrial carcinoma	0.629(0.082-4.849)	0.657	75.22%	< 0.001
	Esophageal squamous cell carcinoma	1.044(0.412-2.640)	0.928	75.22%	< 0.001
	Gastric Cancer	0.851(0.442-1.638)	0.630	75.22%	< 0.001
	Hepatocellular Carcinoma	1.168(0.574–2.373)	0.668	75.22%	< 0.001
	Laryngeal and hypopharyngeal carcinomas	0.326(0.078–1.370)	0.126	75.22%	<0.001
	NSCLC	1.071(0.521-2.204)	0.852	75.22%	< 0.001
	Osteosarcoma	0.960(0.384–2.399)	0.930	75.22%	< 0.001
	Ovarian cancer	1.135(0.478–2.692)	0.774	75.22%	< 0.001
	Pancreatic ductal adenocarcinoma	0.961(0.294-3.144)	0.948	75.22%	< 0.001
	Pelvis chondrosarcoma	0.711(0.080–6.345)	0.760	75.22%	< 0.001
	Renal cell carcinoma	2.208(0.520-9.383)	0.283	75.22%	< 0.001
	Soft tissue sarcoma	5.863(1.115–30.823)	0.037	75.22%	< 0.001
	Uterine cervical cancer	0.880(0.208–3.727)	0.862	75.22%	< 0.001
Race	Intercept	1.705(1.414–2.056)	< 0.001	73.32%	< 0.001
	Caucasian	0.619(0.411–0.932)	0.022	73.32%	< 0.001
Sample type	Intercept	1.430(0.353–5.799)	0.617	81.72%	< 0.001
	Tissue	1.097(0.267–4.510)	0.898	81.72%	<0.001

 $HR_{interaction}$, interaction effect calculated by meta-regression; Positive direction indicates that possible moderators might strengthen OS in the SIRT1 overexpression relative to underexpression. Bold italics indicate statistically significant values (P < 0.05).

expression has stronger efficacy in the China population than the other countries. As to tumor size, meta-regression analysis demonstrated a statistically significant correlation between tumor size and published year (P = 0.0260). As to depth of tumor invasion, meta-regression analysis demonstrated a statistically significant correlation between depth of tumor invasion and sample size (P = 0.0044).

We used funnel plots and Egger's regression models to assess potential publication bias (**Tables 2**, **3**). The association between HRs (**Supplementary Figure 7**) or RRs (**Supplementary Figure 8**) and standard error for the SIRT1 expression was demonstrated in funnel plots, with each plot point representing a study. In regards to the OS, RFS, and TNM stage, we found that Egger's regression yielded potential publication bias (**Figure 7**).

DISCUSSION

In the current study, we conducted a meta-analysis of 13,138 subjects in 63 articles from PubMed, EMBASE and Cochrane library to evaluate prognostic and clinicopathological significance of SIRT1 expression in cancers. We found that elevated expression of SIRT1 was correlated with a poor OS of cancer patients, DFS, EFS, and PFS, but couldn't predict RFS or CCS. Elevated SIRT1 expression was associated with TNM stage, lymph node metastasis, and distant metastasis, but not with tumor size, depth of tumor invasion, differentiation, gender, or age. Our findings provide a clue to understanding prognostic and clinicopathological significance of SIRT1 expression in cancers.

Our current study indicates that overexpression of SIRT1 is correlated with poor OS, DFS, EFS, and PFS, but not with RFS or



CCS, suggesting that SIRT1 expression is significantly correlated with poor prognosis as a global factor but not a restricted factor to tumor itself. It has been shown that SIRT1 is not a protein only found to a specific tissues or organs, instead, its expression can be found in almost all human tissues (1) and involved in a large variety of cellular processes, such as genomic stability, energy metabolism, senescence, gene transcription, and oxidative stress (5) by acting on a wide spectrum of proteins, including histones and transcription factors (2-4). This render SIRT1 plays multiple roles to regulate biological process in multi-systems. Melatonin is a pleiotropic molecule synthesized by pineal gland and many other organs and has important cytoprotective effects in many tissues including aging, neurodegenerative diseases, immunomodulation, and cancer and modulates DNA damage response (88, 89). Melatonin counteracts tumor metastases by modulating cell-cell and cell-matrix interaction, extracellular matrix remodeling, cytoskeleton reorganization, epithelialmesenchymal transition, and angiogenesis (90). Recent studies showed that the upregulated SIRT1 signaling pathway is involved in protective effects of melatonin on vascular endothelium against aging-, oxidative stress-, lipopolysaccharide-, and ischemia-induced damage (91) and delays ovarian aging (92). SIRT1 is induced in normal cells and inhibited in tumor cells by melatonin (88, 89). SIRT1 may mediate the pleiotropic function of melatonin in cancer progression and metastasis. SIRT1 is an endocrine regulator of thyroid and parathyroid hormone function (93-95), and steroid hormone receptor activity (96, 97). SIRT1 is a regulator in immunity and autoimmunity, such as dendritic cell activation, T-regulatory cells (98-102). SIRT1 is also a regulator of lipid and carbohydrate metabolism (9, 103-105). In addition, SIRT1 regulates nervous system by inhibiting neuronal apoptosis and damage as well module nerve regeneration (103, 106, 107). Therefore, SIRT1 is a global factor for endocrine, immunity, metabolism, and nervous system, and affect poor OS, DFS, EFS, and PFS, but not with RFS or CCS in cancer patients.

In the current study, we found that SIRT1 overexpression was associated with TNM stage, lymph node metastasis, and distant metastasis, but not with tumor size, depth of tumor invasion, differentiation, gender, or age, suggesting that SIRT1 promotes metastasis but not growth, proliferation, and invasion of cancer tissues. Tumor is locally initiated and proliferated and may invade near tissues. Tumor size, depth of tumor invasion, and differentiation are terms used to characterize tumors which are locally confined in the early stage of malignancy (108, 109). Metastasis is the characteristics of advanced malignancy of cancer progression (110–113). Our data indicate that SIRT1 overexpression is associated with metastasis but not with tumor characteristics of early stage, suggesting that SIRT1 expression can predict advanced malignancy and is a potential therapeutic target for inhibiting metastasis of advanced cancer.

We performed subgroup analysis because of high heterogeneity in included studies. Correlation analysis between SIRT1 expression and prognosis of cancer types showed that SIRT1 overexpression predicted worse OS of osteosarcoma, esophageal squamous cell carcinoma, OS but not DFS of hepatocellular carcinoma, OS and DFS of breast carcinoma, OS and RFS of NSCLC, DFS but not OS or CCS in colorectal cancer, RFS and CCS but not OS of gastric cancer, RFS and CCS of renal cell carcinoma, CCS but not RFS in head and neck squamous cell carcinoma. SIRT1 overexpression cannot predict OS in ovarian cancer. Correlation analysis between SIRT1 expression and clinicopathological characteristics of cancers types showed that SIRT1 overexpression was associated with a higher TNM stage in hepatocellular carcinoma, but not in pancreatic ductal adenocarcinoma, ovarian cancer, colorectal cancer, gastric cancer, breast carcinoma, or NSCLC. SIRT1 overexpression was associated with distant metastasis in pancreatic ductal adenocarcinoma and breast carcinoma, but not in colorectal cancer or gastric cancer. From these results, we are unable to draw a unanimous conclusion, probably because there is a deficiency of studies that employ all prognostic indexes OS, DFS, EFS, RFS, CCS, and PFS or a full range of clinicopathological characteristics to study the role of SIRT1 expression in survival of patients with a specific cancer type. More thorough studies are warranted.

Our subgroup correlation analysis between SIRT1 expression and prognosis of cancer in different countries and ethnic groups showed that high expression of SIRT1 predicted poor OS and DFS in China, poor OS and EFS but not DFS in Korea, poor OS in Japan, EFS in USA, but not OS in USA or Netherlands. Elevated SIRT1 expression predicted worse OS and DFS in Asian population with cancers, poor DFS but not OS in Caucasian population. Our subgroup analysis between SIRT1 expression and clinicopathological characteristics of cancers in different countries and ethnic groups showed that SIRT1 overexpression was associated with a higher TNM stage and lymphatic metastasis in China and Asian population except lymphatic metastasis in Japan or Korea, and not higher TNM stage and lymphatic metastasis in Caucasian population, We also found that SIRT1 overexpression predicted distant metastasis in Caucasian population, but not in Asians. These results indicate that ethnic background has influence on the role of SIRT1 expression in predicting the OS and clinicopathological characteristics of cancers. This is consistent with recent studies that showed SIRT1 expression is lower in NSCLC than the normal control group in a group of Chinese patients (15), and overexpressed in NSCLC in an Iran population (16). Our study showed that overexpression of SIRT1 predicted a worse OS in the Asian but not in the Caucasian, a higher TNM stage and lymphatic metastasis in Asian population especially in China but not in the Caucasian. This is consistent with the results of our meta-regression analysis. The effects of ethnic background on the role of SIRT1 expression in predicting the OS and clinicopathological characteristics of cancers need further collaborative investigation.

It has been established that there are significant differences between Asian and Caucasian populations in genetic and epigenetic background, dietary, environmental factors (114, 115). These factors are essential for not only initiation and progression, but also metastasis of cancers (116, 117). Mutations and extensive polymorphisms of SIRT1 were found in Chinese and Japanese (118–121) and 41 cancer lines (122). Although the data on mutations and polymorphisms of SIRT1 are very limited, we speculate that difference in SIRT1 mutations and polymorphisms may be one of accounts for difference in predicting OS and TNM stage and lymphatic metastasis of cancer by SIRT1 expression. This deserves further investigation (123).

It is known that metastasis is an independent predictor for poor prognosis of many cancer types (124–126). We find that elevated expression of SIRT1 was correlated with OS, DFS, EFS, and PFS. SIRT1 overexpression is also correlated with TNM stage, lymph node metastasis, and distant metastasis, but not with tumor size, depth of tumor invasion, differentiation, gender, or age. Overexpression of SIRT1 predicted a worse OS and higher TNM stage and lymphatic metastasis in Asian population especially in China. Therefore, overexpression of SIRT1 may promote lymphatic metastasis of cancers that lead to poor OS, DFS, EFS, and PFS. It is likely that SIRT1-mediated molecular events and biological processes could be an underlying mechanism for metastasis.

Our study is consistent with the most recent study by Wang et al. in that SIRT1 overexpression was significantly correlated with the OS in solid cancers, especially in liver cancer and lung cancer based on 7,369 cases from 37 studies and most of them are Asians (34). Consistently, the study by Hong et al showed that high SIRT1 expression correlated with vascular invasion and was not significantly correlated with overall survival rates in colon cancer (36). Study with 3024 patients by Wu et al showed that high SIRT1 expression predicts poor survival in noncolorectal gastrointestinal cancer, but not in colorectal cancer (35). SIRT1 expression was correlated with depth of invasion, lymph node metastasis and TNM stage and predicted a poor OS in colorectal cancer patients based on an analysis with seven studies (33). In an analysis of 1,650 patients in seven studies, high SIRT1 expression predicts a poor prognosis of gastric cancer patients and linked with patients' age, T stage, N stage, and tumor differentiation (32). Analysis by Cao et al. based on six studies involving 604 patients showed that SIRT1 expression was correlated with poor DFS and OS and high TNM stage and lymph node metastasis (31). However, we have performed study on survival and clinicopathological significance of SIRT1 expression in cancers more comprehensively. First, we included 63 eligible articles and a total of 13,138 participants in our study. These patients represented 9 countries and 16 cancer types as well as Asian and Caucasian ethnic groups. Second, we investigated both clinicopathological and prognostic significance of SIRT1 expression based on comprehensive clinical data and performed a series of subgroup analysis based on prognostic types, clinicopathological characteristics, cancer types, ethnic groups, countries. These stratifications provide more vehicles in understanding the survival and clinicopathological significance of SIRT1 expression in cancers.

There are also limitations in our study. Firstly, we found that heterogeneity existed in the meta-analysis as indicated by the I^2 values. It is predictable because of presence of interstudy differences in study design (prospective and retrospective), enrolled populations, treatment regimen, duration of follow-up, outcome measures, and other study and clinical characteristics (127). The heterogeneity among the studies remained, despite the usage of a random-effects model and subgroup analyses (128). Secondly, there is publication bias for SIRT1 expression and prognosis or clinicopathological characteristics as indicated by asymmetry of funnel plots for OS, DFS, EFS, RFS, CCS, PFS, and clinicopathological characteristics. Thirdly, we barely explored the correlation between SIRT1 overexpression and patient survival in terms of clinical parameters. Other elements that may contribute to the heterogeneity, such as therapeutic regimen, pathological grade, body mass index, and mean age, were not analyzed due to the lack of sufficient data (129). Fourthly, we performed a quantitative meta-analysis based mostly on secondary data, which could lead to inaccurate results because of a shortage of original individual patient data (130). Finally, we conducted our study based on the mRNA expression of SIRT1 or the protein levels, although the changes in the mRNA and protein levels of SIRT1 are consistent in several cancer types

(15, 16, 131, 132). The study by Hong et al who determined SIRT1 expression using immunohistochemistry showed similar results to ours study in relation with vascular invasion and overall survival rates in colon cancer (36). We should extensively investigate the prognostic and clinicopathological significance of SIRT1 expression at protein level in the future.

In conclusion, we have found that elevated expression of SIRT1 can predict poor OS, DFS, EFS, and PFS, but not with RFS or CCS, TNM stage, lymph node metastasis, and distant metastasis, but not tumor size, depth of tumor invasion, differentiation of cancers. Ethnic background has influence on the role of SIRT1 expression in predicting survival and clinicopathological characteristics of cancers. Overexpression of SIRT1 predicted a worse OS and higher TNM stage and lymphatic metastasis in Asian population especially in China. SIRT1-mediated molecular events and biological processes could be an underlying mechanism for metastasis and SIRT1 is a potential therapeutic target for inhibiting cancer metastasis. More studies that employ all prognostic indexes OS, DFS, EFS, RFS, CCS, and PFS or a full range of clinicopathological characteristics to study the role of SIRT1 expression in survival of patients with a specific cancer type, and mutations and polymorphisms of SIRT1 in cancers of different ethnic groups need to be further investigated in the future.

DATA AVAILABILITY

All datasets generated for this study are included in the manuscript and the supplementary files.

AUTHOR CONTRIBUTIONS

MS, DH, XGu, and HZ participated in research design. MS, WZ, MD, SX, XGo, PL, HL, and JZ performed data analysis. MS and XGu wrote or contributed to the writing of the manuscript.

FUNDING

This research was supported by the Natural Science Foundation of Hubei Provincial Department of Education (Q20182105), Chen Xiao-ping Foundation for the development of science and technology of Hubei Provincial (CXPJJH11800001-2018333), Natural Science Foundation of Hubei Province of China (2016CFB530) and Faculty Development Foundation of Hubei University of Medicine (2014QDJZR01), and National Students' platform for innovation and entrepreneurship

REFERENCES

- Fagerberg L, Hallstrom BM, Oksvold P, Kampf C, Djureinovic D, Odeberg J, et al. Analysis of the human tissue-specific expression by genome-wide integration of transcriptomics and antibody-based proteomics. *Mol Cell Proteomics*. (2014) 13:397–406. doi: 10.1074/mcp.M113.035600
- Jang SH, Min KW, Paik SS, Jang KS. Loss of SIRT1 histone deacetylase expression associates with tumour progression in colorectal adenocarcinoma. *J Clin Pathol.* (2012) 65:735–9. doi: 10.1136/jclinpath-2012-200685

training program (201810929005, 201810929009, 201810929068, and 201813249010).

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fendo. 2019.00121/full#supplementary-material

Supplementary Table 1 Databases searching terms. **(a)** Search criterion of Medline (via PubMed, from inception to April 1st, 2018) (n = 2397). **(b)** Search criterion of Embase (from 1966 to July 24, 2018) (n = 2460). **(c)** Search criterion of Cochrane Library (April 1st, 2018) (n = 20).

Supplementary Table 2 | Results of quality assessment using the Newcastle–Ottawa Scale for the included studies.

Supplementary Table 3 | Subgroup analysis for SIRT1 overexpression and the prognosis in cancers.

Supplementary Table 4 | Subgroup analysis for SIRT1 overexpression and clinicopathological parameters in cancers.

Supplementary Table 5 | Meta-regression analysis of heterogeneity for survival outcomes.

Supplementary Table 6 | Meta-regression analysis of heterogeneity for clinicopathological parameters in cancers.

Supplementary Figure 1 | Forest plot of subgroup analysis for SIRT1 overexpression and OS in cancers. (A) Cancer subgroup, (B) Country subgroup, (C) Ethnicity subgroup.

Supplementary Figure 2 | Forest plot of subgroup analysis for SIRT1 overexpression and DFS in cancers. (A) Cancer subgroup, (B) Country subgroup, (C) Ethnicity subgroup.

Supplementary Figure 3 | Forest plots of non-essential clinicopathological outcomes compared SIRT1 overexpression with underexpression. (A) Tumor size,
 (B) Depth of tumor invasion, (C) Differentiation, (D) Gender, (E) Age.

Supplementary Figure 4 | Forest plot of subgroup analysis for SIRT1 overexpression and TNM stage in cancers. (A) Cancer subgroup, (B) Country subgroup, (C) Ethnicity subgroup.

Supplementary Figure 5 | Forest plot of subgroup analysis for SIRT1 overexpression and distant metastasis in cancers. (A) Cancer subgroup, (B) Ethnicity subgroup.

Supplementary Figure 6 | Forest plot of subgroup analysis for SIRT1 overexpression and lymphatic metastasis in cancers. (A) Country subgroup, (B) Ethnicity subgroup.

Supplementary Figure 7 | Funnel plot for publication bias for SIRT1 expression and prognosis. (A) OS, (B) DFS, (C) EFS, (D) RFS, (E) CCS.

Supplementary Figure 8 | Funnel plot for publication bias for SIRT1 expression and clinicopathological characteristics. (A) Age, (B) Gender, (C) Tumor stage,
 (D) Distant metastasis, (E) Lymphatic invasion, (F) Tumor size, (G) Depth of tumor invasion, (H) Differentiation.

- Derr RS, van Hoesel AQ, Benard A, Goossens-Beumer IJ, Sajet A, Dekker-Ensink NG, et al. High nuclear expression levels of histone-modifying enzymes LSD1, HDAC2 and SIRT1 in tumor cells correlate with decreased survival and increased relapse in breast cancer patients. *BMC Cancer*. (2014) 14:604. doi: 10.1186/1471-2407-14-604
- Xu Z, Yang Y, Li B, Li Y, Xia K, Li X, et al. Checkpoint suppressor 1 suppresses transcriptional activity of ERalpha and breast cancer cell proliferation via deacetylase SIRT1. *Cell Death Dis.* (2018) 9:559. doi: 10.1038/s41419-018-0629-3

- Tran D, Bergholz J, Zhang H, He H, Wang Y, Zhang Y, et al. Insulin-like growth factor-1 regulates the SIRT1-p53 pathway in cellular senescence. *Aging Cell.* (2014) 13:669–78. doi: 10.1111/acel. 12219
- Imai S, Guarente L. Ten years of NAD-dependent SIR2 family deacetylases: implications for metabolic diseases. *Trends Pharmacol Sci.* (2010) 31:212–20. doi: 10.1016/j.tips.2010.02.003
- Sebastian C, Satterstrom FK, Haigis MC, Mostoslavsky R. From sirtuin biology to human diseases: an update. J Biol Chem. (2012) 287:42444–52. doi: 10.1074/jbc.R112.402768
- Carafa V, Rotili D, Forgione M, Cuomo F, Serretiello E, Hailu GS, et al. Sirtuin functions and modulation: from chemistry to the clinic. *Clin Epigenet*. (2016) 8:61. doi: 10.1186/s13148-016-0224-3
- Zhu S, Dong Z, Ke X, Hou J, Zhao E, Zhang K, et al. The roles of sirtuins family in cell metabolism during tumor development. *Semin Cancer Biol.* (2018). doi: 10.1016/j.semcancer.2018.11.003. [Epub ahead of print].
- Liu T, Liu PY, Marshall GM. The critical role of the class III histone deacetylase SIRT1 in cancer. *Cancer Res.* (2009) 69:1702–5. doi: 10.1158/0008-5472.CAN-08-3365
- Fang Y, Nicholl MB. Sirtuin 1 in malignant transformation: friend or foe? Cancer Lett. (2011) 306:10–4. doi: 10.1016/j.canlet.2011.02.019
- 12. Fang Y, Nicholl MB. A dual role for sirtuin 1 in tumorigenesis. *Curr Pharm Des.* (2014) 20:2634–6.
- Xu Y, Qin Q, Chen R, Wei C, Mo Q. SIRT1 promotes proliferation, migration, and invasion of breast cancer cell line MCF-7 by upregulating DNA polymerase delta1 (POLD1). *Biochem Biophys Res Commun.* (2018) 502:351–7. doi: 10.1016/j.bbrc.2018.05.164
- Xia X, Zhou X. Knockdown of SIRT1 inhibits proliferation and promotes apoptosis of paclitaxel-resistant human cervical cancer cells. *Cell Mol Biol.* (2018) 64:36–41.
- Yang F. The expression and mechanism of Sirt1 and AMPK in nonsmall cell lung cancer. J BUON. (2018) 23:106–10.
- Gharabaghi MA. Diagnostic investigation of BIRC6 and SIRT1 protein expression level as potential prognostic biomarkers in patients with nonsmall cell lung cancer. *Clin Respir J.* (2018) 12:633–8. doi: 10.1111/crj.12572
- Tan J, Liu Y, Maimaiti Y, Wang C, Yan Y, Zhou J, et al. Combination of SIRT1 and Src overexpression suggests poor prognosis in luminal breast cancer. *Onco Targets Ther.* (2018) 11:2051–61. doi: 10.2147/OTT.S162503
- Shuang T, Wang M, Zhou Y, Shi C. Over-expression of Sirt1 contributes to chemoresistance and indicates poor prognosis in serous epithelial ovarian cancer (EOC). *Med Oncol.* (2015) 32:260. doi: 10.1007/s12032-015-0706-8
- Noguchi A, Kikuchi K, Zheng H, Takahashi H, Miyagi Y, Aoki I, et al. SIRT1 expression is associated with a poor prognosis, whereas DBC1 is associated with favorable outcomes in gastric cancer. *Cancer Med.* (2014) 3:1553–61. doi: 10.1002/cam4.310
- Li C, Wang L, Zheng L, Zhan X, Xu B, Jiang J, et al. SIRT1 expression is associated with poor prognosis of lung adenocarcinoma. *Onco Targets Ther.* (2015) 8:977–84. doi: 10.2147/ott.s82378
- Lee H, Kim KR, Noh SJ, Park HS, Kwon KS, Park BH, et al. Expression of DBC1 and SIRT1 is associated with poor prognosis for breast carcinoma. *Hum Pathol.* (2011) 42:204–13. doi: 10.1016/j.humpath.2010.05.023
- Chen X, Sun K, Jiao S, Cai N, Zhao X, Zou H, et al. High levels of SIRT1 expression enhance tumorigenesis and associate with a poor prognosis of colorectal carcinoma patients. *Sci Rep.* (2014) 4:7481. doi: 10.1038/srep07481
- Chen X, Hokka D, Maniwa Y, Ohbayashi C, Itoh T, Hayashi Y. Sirt1 is a tumor promoter in lung adenocarcinoma. Oncol Lett. (2014) 8:387–93. doi: 10.3892/ol.2014.2057
- 24. Kim JR, Moon YJ, Kwon KS, Bae JS, Wagle S, Yu TK, et al. Expression of SIRT1 and DBC1 is associated with poor prognosis of soft tissue sarcomas. *PLoS ONE.* (2013) 8:e74738. doi: 10.1371/journal.pone.0074738
- Wu M, Wei W, Xiao X, Guo J, Xie X, Li L, et al. Expression of SIRT1 is associated with lymph node metastasis and poor prognosis in both operable triple-negative and non-triple-negative breast cancer. *Med Oncol.* (2012) 29:3240–9. doi: 10.1007/s12032-012-0260-6
- Chen HC, Jeng YM, Yuan RH, Hsu HC, Chen YL. SIRT1 promotes tumorigenesis and resistance to chemotherapy in hepatocellular carcinoma and its expression predicts poor prognosis. *Ann Surg Oncol.* (2012) 19:2011– 9. doi: 10.1245/s10434-011-2159-4

- Cha EJ, Noh SJ, Kwon KS, Kim CY, Park BH, Park HS, et al. Expression of DBC1 and SIRT1 is associated with poor prognosis of gastric carcinoma. *Clin Cancer Res.* (2009) 15:4453–9. doi: 10.1158/1078-0432.ccr-08-3329
- Jang KY, Hwang SH, Kwon KS, Kim KR, Choi HN, Lee NR, et al. SIRT1 expression is associated with poor prognosis of diffuse large B-cell lymphoma. *Am J Surg Pathol.* (2008) 32:1523–31. doi: 10.1097/PAS.0b013e31816b6478
- Noguchi A, Li X, Kubota A, Kikuchi K, Kameda Y, Zheng H, et al. SIRT1 expression is associated with good prognosis for head and neck squamous cell carcinoma patients. Oral Surg Oral Med Oral Pathol Oral Radiol. (2013) 115:385–92. doi: 10.1016/j.oooo.2012.12.013
- Jung W, Hong KD, Jung WY, Lee E, Shin BK, Kim HK, et al. SIRT1 expression is associated with good prognosis in colorectal cancer. *Korean J Pathol.* (2013) 47:332–9. doi: 10.4132/KoreanJPathol.2013.47.4.332
- Cao YW, Li YC, Wan GX, Du XM, Li F. Clinicopathological and prognostic role of SIRT1 in breast cancer patients: a meta-analysis. *Int J Clin Exp Med.* (2015) 8:616–24.
- 32. Jiang B, Chen JH, Yuan WZ, Ji JT, Liu ZY, Wu L, et al. Prognostic and clinical value of Sirt1 expression in gastric cancer: A systematic meta-analysis. J Huazhong Univ Sci Technolog Med Sci. (2016) 36:278–84. doi: 10.1007/s11596-016-1580-0
- 33. Zu G, Ji A, Zhou T, Che N. Clinicopathological significance of SIRT1 expression in colorectal cancer: a systematic review and meta analysis. *Int J Surg.* (2016) 26:32–7. doi: 10.1016/j.ijsu.2016.01.002
- Wang C, Yang W, Dong F, Guo Y, Tan J, Ruan S, et al. The prognostic role of Sirt1 expression in solid malignancies: a meta-analysis. *Oncotarget*. (2017) 8:66343–51. doi: 10.18632/oncotarget.18494
- Wu S, Jiang J, Liu J, Wang X, Gan Y, Tang Y. Meta-analysis of SIRT1 expression as a prognostic marker for overall survival in gastrointestinal cancer. *Oncotarget*. (2017) 8:62589–99. doi: 10.18632/oncotarget.19880
- Hong WG, Pyo JS. The clinicopathological significance of SIRT1 expression in colon cancer: an immunohistochemical study and meta-analysis. *Pathol Res Pract.* (2018) 214:1550–5. doi: 10.1016/j.prp.2018.07.022
- Parmar MK, Torri V, Stewart L. Extracting summary statistics to perform meta-analyses of the published literature for survival endpoints. *Stat Med.* (1998) 17:2815–34.
- Tierney JF, Stewart LA, Ghersi D, Burdett S, Sydes MR. Practical methods for incorporating summary time-to-event data into meta-analysis. *Trials*. (2007) 8:16. doi: 10.1186/1745-6215-8-16
- Stuck AE, Rubenstein LZ, Wieland D. Bias in meta-analysis detected by a simple, graphical test. Asymmetry detected in funnel plot was probably due to true heterogeneity. *BMJ*. (1998) 316: 469; author reply 470–461.
- Stenzinger A, Endris V, Klauschen F, Sinn B, Lorenz K, Warth A, et al. High SIRT1 expression is a negative prognosticator in pancreatic ductal adenocarcinoma. *BMC Cancer.* (2013) 13:450. doi: 10.1186/1471-2407-13-450
- Li S, Hong H, Lv H, Wu G, Wang Z. SIRT 1 overexpression is associated with metastasis of Pancreatic Ductal Adenocarcinoma (PDAC) and promotes migration and growth of PDAC cells. *Med Sci Monit.* (2016) 22:1593–600. doi: 10.12659/MSM.896697
- Teramae M, Fukuda T, Wada T, Kawanishi M, Imai K, Yamauchi M, et al. Sirtuin1 expression predicts the efficacy of neoadjuvant chemotherapy for locally advanced uterine cervical cancer. *Mol Clin Oncol.* (2015) 3:73–8. doi: 10.3892/mco.2014.427
- Asaka R, Miyamoto T. Sirtuin 1 promotes the growth and cisplatin resistance of endometrial carcinoma cells: a novel therapeutic target. *Lab Invest.* (2015) 95:1363–73. doi: 10.1038/labinvest.2015.119
- 44. Jang KY, Kim KS, Hwang SH, Kwon KS, Kim KR, Park HS, et al. Expression and prognostic significance of SIRT1 in ovarian epithelial tumours. *Pathology*. (2009) 41:366–71. doi: 10.1080/00313020902 884451
- 45. Mvunta DH, Miyamoto T, Asaka R, Yamada Y, Ando H, Higuchi S, et al. Overexpression of SIRT1 is associated with poor outcomes in patients with ovarian carcinoma. *Appl Immunohistochem Mol Morphol.* (2017) 25:415–21. doi: 10.1097/pai.00000000000316
- Zhang N, Xie T, Xian M, Wang YJ, Li HY, Ying MD, et al. SIRT1 promotes metastasis of human osteosarcoma cells. *Oncotarget*. (2016) 7:79654–69. doi: 10.18632/oncotarget.12916

- Feng H, Guo P, Wang J, Xu J, Xie C, Gao F. Expression of Leptin and Sirtuin-1 is associated with poor prognosis in patients with osteosarcoma. *Pathol Res*
- Pract. (2016) 212:319–24. doi: 10.1016/j.prp.2016.02.002
 48. Yu XM, Liu Y, Jin T, Liu J, Wang J, Ma C, et al. The expression of SIRT1 and DBC1 in laryngeal and hypopharyngeal carcinomas. *PLoS ONE*. (2013) 8:e66975. doi: 10.1371/journal.pone.0066975
- Batra A, Kashyap S, Singh L, Bakhshi S. Sirtuin1 expression and correlation with histopathological features in retinoblastoma. *Ocul Oncol Pathol.* (2015) 2:86–90. doi: 10.1159/000439594
- Chen GQ, Tian H, Yue WM, Li L, Li SH, Qi L, et al. SIRT1 expression is associated with lymphangiogenesis, lymphovascular invasion and prognosis in pN0 esophageal squamous cell carcinoma. *Cell Biosci.* (2014) 4:48. doi: 10.1186/2045-3701-4-48
- He Z, Yi J, Jin L, Pan B, Chen L, Song H. Overexpression of Sirtuin-1 is associated with poor clinical outcome in esophageal squamous cell carcinoma. *Tumour Biol.* (2016) 37:7139–48. doi: 10.1007/s13277-015-4459-y
- Feng H, Wang J, Xu J, Xie C, Gao F, Li Z. The expression of SIRT1 regulates the metastaticplasticity of chondrosarcoma cells by inducing epithelialmesenchymal transition. *Sci Rep.* (2017) 7:41203. doi: 10.1038/srep41203
- Noh SJ, Kang MJ, Kim KM, Bae JS, Park HS, Moon WS, et al. Acetylation status of P53 and the expression of DBC1, SIRT1, and androgen receptor are associated with survival in clear cell renal cell carcinoma patients. *Pathology*. (2013) 45:574–80. doi: 10.1097/PAT.0b013e3283652c7a
- Jeh SU, Park JJ, Lee JS, Kim DC, Do J, Lee SW, et al. Differential expression of the sirtuin family in renal cell carcinoma: aspects of carcinogenesis and prognostic significance. Urol Oncol. (2017) 35:675.e9– 675.e15. doi: 10.1016/j.urolonc.2017.08.016
- 55. Ren Y, Gu Y, Wang W, Wang J, Li X, Wang Y, et al. Expression of the significance of silent information regulator type-1 in Angioimmunoblastic T-cell lymphoma is greater association with tumorigenesis and has strong implications for adverse prognosis. *Cell Cycle.* (2016) 15:1545–51. doi: 10.1080/15384101.2016.1172157
- Nosho K, Shima K, Irahara N, Kure S, Firestein R, Baba Y, et al. SIRT1 histone deacetylase expression is associated with microsatellite instability and CpG island methylator phenotype in colorectal cancer. *Mod Pathol.* (2009) 22:922–32. doi: 10.1038/modpathol.2009.49
- Benard A, Goossens-Beumer IJ, van Hoesel AQ, Horati H, de Graaf W, Putter H, et al. Nuclear expression of histone deacetylases and their histone modifications predicts clinical outcome in colorectal cancer. *Histopathology*. (2015) 66:270–82. doi: 10.1111/his.12534
- Lv L, Shen Z, Zhang J, Zhang H, Dong J, Yan Y, et al. Clinicopathological significance of SIRT1 expression in colorectal adenocarcinoma. *Med Oncol.* (2014) 31:965. doi: 10.1007/s12032-014-0965-9
- Lee WY, Lee WT, Cheng CH, Chen KC, Chou CM, Chung CH, et al. Repositioning antipsychotic chlorpromazine for treating colorectal cancer by inhibiting sirtuin 1. Oncotarget. (2015) 6:27580–95. doi: 10.18632/oncotarget.4768
- Cheng F, Su L, Yao C, Liu L, Shen J, Liu C, et al. SIRT1 promotes epithelial-mesenchymal transition and metastasis in colorectal cancer by regulating Fra-1 expression. *Cancer Lett.* (2016) 375:274–83. doi: 10.1016/j.canlet.2016.03.010
- Chen J, Zhang B, Wong N, Lo AW, To KF, Chan AW, et al. Sirtuin 1 is upregulated in a subset of hepatocellular carcinomas where it is essential for telomere maintenance and tumor cell growth. *Cancer Res.* (2011) 71:4138– 49. doi: 10.1158/0008-5472.can-10-4274
- Jang KY, Noh SJ, Lehwald N, Tao GZ, Bellovin DI, Park HS, et al. SIRT1 and c-Myc promote liver tumor cell survival and predict poor survival of human hepatocellular carcinomas. *PLoS ONE*. (2012) 7:e45119. doi: 10.1371/journal.pone.0045119
- Hao C, Zhu PX, Yang X, Han ZP, Jiang JH, Zong C, et al. Overexpression of SIRT1 promotes metastasis through epithelial-mesenchymal transition in hepatocellular carcinoma. *BMC Cancer*. (2014) 14:978. doi: 10.1186/1471-2407-14-978
- 64. Cheng J, Liu C, Liu L, Chen X, Shan J, Shen J, et al. MEK1 signaling promotes self-renewal and tumorigenicity of liver cancer stem cells via maintaining SIRT1 protein stabilization. *Oncotarget.* (2016) 7:20597–611. doi: 10.18632/oncotarget.7972

- 65. Li Y, Xu S, Li J, Zheng L, Feng M, Wang X, et al. SIRT1 facilitates hepatocellular carcinoma metastasis by promoting PGC-1alpha-mediated mitochondrial biogenesis. Oncotarget. (2016) 7:29255-74. doi: 10.18632/oncotarget.8711
- 66. Liu L, Liu C, Zhang Q, Shen J, Zhang H, Shan J, et al. SIRT1mediated transcriptional regulation of SOX2 is important for self-renewal of liver cancer stem cells. *Hepatology*. (2016) 64:814–27. doi: 10.1002/hep. 28690
- Feng AN, Zhang LH, Fan XS, Huang Q, Ye Q, Wu HY, et al. Expression of SIRT1 in gastric cardiac cancer and its clinicopathologic significance. *Int J Surg Pathol.* (2011) 19:743–50. doi: 10.1177/1066896911412181
- Kang Y, Jung WY, Lee H, Lee E, Kim A, Kim BH. Expression of SIRT1 and DBC1 in gastric adenocarcinoma. *Korean J Pathol.* (2012) 46:523–31. doi: 10.4132/KoreanJPathol.2012.46.6.523
- Qiu G, Li X. The prognostic role of SIRT1-autophagy axis in gastric cancer. Evid Based Complement Alternat Med. (2016) 2016;6869415. doi: 10.1155/2016/5989681
- Szasz AM, Lanczky A, Nagy A, Forster S, Hark K, Green JE, et al. Cross-validation of survival associated biomarkers in gastric cancer using transcriptomic data of 1,065 patients. *Oncotarget*. (2016) 7:49322–33. doi: 10.18632/oncotarget.10337
- Zhang S, Huang S, Deng C, Cao Y, Yang J, Chen G, et al. Coordinated overexpression of SIRT1 and STAT3 is associated with poor survival outcome in gastric cancer patients. *Oncotarget*. (2017) 8:18848–60. doi: 10.1016/j.nut.2016.09.008
- Zhang LH, Huang Q, Fan XS, Wu HY, Yang J, Feng AN. Clinicopathological significance of SIRT1 and p300/CBP expression in gastroesophageal junction (GEJ) cancer and the correlation with E-cadherin and MLH1. *Pathol Res Pract.* (2013) 209:611–7. doi: 10.1016/j.prp.2013.03.012
- Ren NS, Ji M, Tokar EJ, Busch EL, Xu X, Lewis D, et al. Haploinsufficiency of SIRT1 enhances glutamine metabolism and promotes cancer development. *Curr Biol.* (2017) 27:483–94. doi: 10.1016/j.cub.2016.12.047
- Shin DH, Choi YJ, Jin P, Yoon H, Chun YS, Shin HW, et al. Distinct effects of SIRT1 in cancer and stromal cells on tumor promotion. *Oncotarget*. (2016) 7:23975–87. doi: 10.18632/oncotarget.8073
- Zhang XZ, Chen SQ, Cheng ML, Cao FL, Cheng YF. The expression and correlation of SIRT1 and Phospho-SIRT1 in colorectal cancer. *Int J Clin Exp Med.* (2015) 8:809–17.
- Cao YW, Li WQ, Wan GX, Li YX, Du XM, Li YC, et al. Correlation and prognostic value of SIRT1 and Notch1 signaling in breast cancer. J Exp Clin Cancer Res. (2014) 33:97. doi: 10.1186/s13046-014-0097-2
- 77. Jin MS, Hyun CL, Park IA, Kim JY, Chung YR, Im SA, et al. SIRT1 induces tumor invasion by targeting epithelial mesenchymal transition-related pathway and is a prognostic marker in triple negative breast cancer. *Tumour Biol.* (2016) 37:4743–53. doi: 10.1007/s13277-015-4231-3
- Kim H, Lee KH, Park IA, Chung YR, Im SA, Noh DY, et al. Expression of SIRT1 and apoptosis-related proteins is predictive for lymph node metastasis and disease-free survival in luminal A breast cancer. *Virchows Arch.* (2015) 467:563–70. doi: 10.1007/s00428-015-1815-7
- 79. Chung SY, Jung YY, Park IA, Kim H, Chung YR, Kim JY, et al. Oncogenic role of SIRT1 associated with tumor invasion, lymph node metastasis, and poor disease-free survival in triple negative breast cancer. *Clin Exp Metastasis*. (2016) 33:179–85. doi: 10.1007/s10585-015-9767-5
- Lee JJ, Lee HJ, Son BH, Kim SB, Ahn JH, Ahn SD, et al. Expression of FOXM1 and related proteins in breast cancer molecular subtypes. *Int J Exp Pathol.* (2016) 97:170–7. doi: 10.1111/iep.12187
- Chung YR, Kim H, Park SY, Park IA, Jang JJ, Choe JY, et al. Distinctive role of SIRT1 expression on tumor invasion and metastasis in breast cancer by molecular subtype. *Hum Pathol.* (2015) 46:1027–35. doi: 10.1016/j.humpath.2015.03.015
- Zhang W, Luo J, Yang F, Wang Y, Yin Y, Strom A, et al. BRCA1 inhibits ARmediated proliferation of breast cancer cells through the activation of SIRT1. *Sci Rep.* (2016) 6:22034. doi: 10.1038/srep22034
- Sung JY, Kim R, Kim JE, Lee J. Balance between SIRT1 and DBC1 expression is lost in breast cancer. *Cancer Sci.* (2010) 101:1738–44. doi: 10.1111/j.1349-7006.2010.01573.x

- Lin SY, Peng F. Association of SIRT1 and HMGA1 expression in non-small cell lung cancer. Oncol Lett. (2016) 11:782–8. doi: 10.3892/ol.2015.3914
- Noh SJ, Baek HA, Park HS, Jang KY, Moon WS, Kang MJ, et al. Expression of SIRT1 and cortactin is associated with progression of non-small cell lung cancer. *Pathol Res Pract.* (2013) 209:365–70. doi: 10.1016/j.prp.2013.03.011
- Zhang T, Rong N, Chen J, Zou C, Jing H, Zhu X, et al. SIRT1 expression is associated with the chemotherapy response and prognosis of patients with advanced NSCLC. *PLoS ONE.* (2013) 8:e79162. doi: 10.1371/journal.pone.0079162
- Grbesa I, Pajares MJ, Martinez-Terroba E, Agorreta J, Mikecin AM, Larrayoz M, et al. Expression of sirtuin 1 and 2 is associated with poor prognosis in non-small cell lung cancer patients. *PLoS ONE.* (2015) 10:e0124670. doi: 10.1371/journal.pone.0124670
- Majidinia M, Sadeghpour A, Mehrzadi S, Reiter RJ, Khatami N, Yousefi B. Melatonin: a pleiotropic molecule that modulates DNA damage response and repair pathways. J Pineal Res. (2017) 63:e12416. doi: 10.1111/jpi.12416
- Mayo JC, Sainz RM, Gonzalez Menendez P, Cepas V, Tan DX, Reiter RJ. Melatonin and sirtuins: a "not-so unexpected" relationship. J Pineal Res. (2017) 62:e12391. doi: 10.1111/jpi.12391
- Su SC, Hsieh MJ, Yang WE, Chung WH, Reiter RJ, Yang SF. Cancer metastasis: mechanisms of inhibition by melatonin. *J Pineal Res.* (2017) 62:e12370. doi: 10.1111/jpi.12370
- 91. Lee FY, Sun CK, Sung PH, Chen KH, Chua S, Sheu JJ, et al. Daily melatonin protects the endothelial lineage and functional integrity against the aging process, oxidative stress, and toxic environment and restores blood flow in critical limb ischemia area in mice. *J Pineal Res.* (2018) 65:e12489. doi: 10.1111/jpi.12489
- Tamura H, Kawamoto M, Sato S, Tamura I, Maekawa R, Taketani T, et al. Long-term melatonin treatment delays ovarian aging. *J Pineal Res.* (2017) 62:e12381. doi: 10.1111/jpi.12381
- Cordeiro A, de Souza LL, Oliveira LS, Faustino LC, Santiago LA, Bloise FF, et al. Thyroid hormone regulation of Sirtuin 1 expression and implications to integrated responses in fasted mice. *J Endocrinol.* (2013) 216:181–93. doi: 10.1530/JOE-12-0420
- Thakran S, Sharma P, Attia RR, Hori RT, Deng X, Elam MB, et al. Role of sirtuin 1 in the regulation of hepatic gene expression by thyroid hormone. J Biol Chem. (2013) 288:807–18. doi: 10.1074/jbc.M112.437970
- 95. Fei Y, Shimizu E, McBurney MW, Partridge NC. Sirtuin 1 is a negative regulator of parathyroid hormone stimulation of matrix metalloproteinase 13 expression in osteoblastic cells: role of sirtuin 1 in the action of PTH on osteoblasts. J Biol Chem. (2015) 290:8373–82. doi: 10.1074/jbc.M114.602763
- Dai Y, Ngo D, Forman LW, Qin DC, Jacob J, Faller DV. Sirtuin 1 is required for antagonist-induced transcriptional repression of androgenresponsive genes by the androgen receptor. *Mol Endocrinol.* (2007) 21:1807– 21. doi: 10.1210/me.2006-0467
- Moore RL, Dai Y, Faller DV. Sirtuin 1 (SIRT1) and steroid hormone receptor activity in cancer. J Endocrinol. (2012) 213:37–48. doi: 10.1530/JOE-11-0217
- Kong S, McBurney MW, Fang D. Sirtuin 1 in immune regulation and autoimmunity. *Immunol Cell Biol.* (2012) 90:6–13. doi: 10.1038/icb.2011.102
- 99. Yang H, Lee SM, Gao B, Zhang J, Fang D. Histone deacetylase sirtuin 1 deacetylates IRF1 protein and programs dendritic cells to control Th17 protein differentiation during autoimmune inflammation. *J Biol Chem.* (2013) 288:37256–66. doi: 10.1074/jbc.M113.527531
- 100. Akimova T, Xiao H, Liu Y, Bhatti TR, Jiao J, Eruslanov E, et al. Targeting sirtuin-1 alleviates experimental autoimmune colitis by induction of Foxp3+ T-regulatory cells. *Mucosal Immunol.* (2014) 7:1209–20. doi: 10.1038/mi.2014.10
- 101. Luciano JA, Kautza B, Darwiche S, Martinez S, Stratimirovic S, Waltz P, et al. Sirtuin 1 agonist minimizes injury and improves the immune response following traumatic shock. *Shock.* (2015) 44(Suppl. 1):149–55. doi: 10.1097/SHK.00000000000412
- 102. Owczarczyk AB, Schaller MA, Reed M, Rasky AJ, Lombard DB, Lukacs NW. Sirtuin 1 regulates dendritic cell activation and autophagy during respiratory syncytial virus-induced immune responses. *J Immunol.* (2015) 195:1637–46. doi: 10.4049/jimmunol.1500326
- 103. Zhang Y, Li H, Cao Y, Zhang M, Wei S. Sirtuin 1 regulates lipid metabolism associated with optic nerve regeneration. *Mol Med Rep.* (2015) 12:6962–8. doi: 10.3892/mmr.2015.4286

- 104. Alayon AN, Ortega Avila JG, Echeverri Jimenez I. Carbohydrate metabolism and gene expression of sirtuin 1 in healthy subjects after Sacha inchi oil supplementation: a randomized trial. *Food Funct.* (2018) 9:1570–7. doi: 10.1039/c7fo01956d
- 105. Cao MM, Lu X, Liu GD, Su Y, Li YB, Zhou J. Resveratrol attenuates type 2 diabetes mellitus by mediating mitochondrial biogenesis and lipid metabolism via Sirtuin type 1. *Exp Ther Med.* (2018) 15:576–84. doi: 10.3892/etm.2017.5400
- 106. Yan P, Bai L, Lu W, Gao Y, Bi Y, Lv G. Regulation of autophagy by AMP-activated protein kinase/sirtuin 1 pathway reduces spinal cord neurons damage. *Iran J Basic Med Sci.* (2017) 20:1029–36. doi: 10.22038/JJBMS.2017.9272
- 107. Shi X, Pi L, Zhou S, Li X, Min F, Wang S, et al. Activation of Sirtuin 1 Attenuates High Glucose-Induced Neuronal Apoptosis by Deacetylating p53. Front Endocrinol. (2018) 9:274. doi: 10.3389/fendo.2018.00274
- 108. Knudsen BS, Vasioukhin V. Mechanisms of prostate cancer initiation and progression. Adv Cancer Res. (2010) 109:1–50. doi: 10.1016/B978-0-12-380890-5.00001-6
- 109. Leong SP, Aktipis A, Maley C. Cancer initiation and progression within the cancer microenvironment. *Clin Exp Metastasis*. (2018). doi: 10.1007/s10585-018-9921-y. [Epub ahead of print].
- Patel LR, Camacho DF, Shiozawa Y, Pienta KJ, Taichman RS. Mechanisms of cancer cell metastasis to the bone: a multistep process. *Future Oncol.* (2011) 7:1285–97. doi: 10.2217/fon.11.112
- 111. Farnsworth RH, Achen MG, Stacker SA. The evolving role of lymphatics in cancer metastasis. *Curr Opin Immunol.* (2018) 53:64–73. doi: 10.1016/j.coi.2018.04.008
- 112. Franchino F, Ruda R, Soffietti R. Mechanisms and Therapy for Cancer Metastasis to the Brain. Front Oncol. (2018) 8:161. doi: 10.3389/fonc.2018.00161
- 113. Hamidi H, Ivaska J. Every step of the way: integrins in cancer progression and metastasis. Nat Rev Cancer. (2018) 18:533–48. doi: 10.1038/s41568-018-0038-z
- 114. Hur YM, Kaprio J, Iacono WG, Boomsma DI, McGue M, Silventoinen K, et al. Genetic influences on the difference in variability of height, weight and body mass index between Caucasian and East Asian adolescent twins. *Int J Obes.* (2008) 32:1455–67. doi: 10.1038/ijo.2008.144
- 115. Tarabay M, Elshazli R, Settin A. African vs. Caucasian and Asian difference for the association of interleukin-10 promotor polymorphisms with type 2 diabetes mellitus (a meta-analysis study). *Meta Gene.* (2016) 9:10–7. doi: 10.1016/j.mgene.2016.02.006
- Chatterjee A, Rodger EJ, Eccles MR. Epigenetic drivers of tumourigenesis and cancer metastasis. *Semin Cancer Biol.* (2018) 51:149–59. S1044-579X(17)30053-6
- 117. Pavlidis ET, Pavlidis TE. Current molecular and genetic aspects of pancreatic cancer, the role of Metastasis Associated Proteins (MTA): a review. J Invest Surg. (2018) 31:54–66. doi: 10.1080/08941939.2016. 1269854
- 118. Shimoyama Y, Suzuki K, Hamajima N, Niwa T. Sirtuin 1 gene polymorphisms are associated with body fat and blood pressure in Japanese. *Transl Res.* (2011) 157:339–47. doi: 10.1016/j.trsl.2011.02.004
- 119. Shimoyama Y, Mitsuda Y, Tsuruta Y, Suzuki K, Hamajima N, Niwa T. SIRTUIN 1 gene polymorphisms are associated with cholesterol metabolism and coronary artery calcification in Japanese hemodialysis patients. *J Ren Nutr.* (2012) 22:114–9. doi: 10.1053/j.jrn.2011.10.025
- 120. Chen Z, Zhai Y, Zhang W, Teng Y, Yao K. Single Nucleotide Polymorphisms of the Sirtuin 1 (SIRT1) gene are associated with age-related macular degeneration in chinese han individuals: a case-control pilot study. *Medicine*. (2015) 94:e2238. doi: 10.1097/MD.00000000002238
- 121. Lv Y, Lin S, Peng F. SIRT1 gene polymorphisms and risk of lung cancer. Cancer Manag Res. (2017) 9:381-6. doi: 10.1016/j.canlet.2017. 09.001
- 122. Han J, Hubbard BP, Lee J, Montagna C, Lee HW, Sinclair DA, et al. Analysis of 41 cancer cell lines reveals excessive allelic loss and novel mutations in the SIRT1 gene. *Cell Cycle*. (2013) 12:263–70. doi: 10.4161/cc.23056
- 123. Mei ZB, Duan CY, Li CB, Cui L, Ogino S. Prognostic role of tumor PIK3CA mutation in colorectal cancer: a systematic review and meta-analysis. *Ann Oncol.* (2016) 27:1836–48. doi: 10.1093/annonc/mdw264

- 124. Tsutsumi S, Watanabe R, Tabe Y, Fujii T, Morita H, Kigure W, et al. Extranodal metastasis predicts poor survival in advanced colorectal cancer. *Hepatogastroenterology*. (2012) 59:372-4. doi: 10.5754/hge 10139
- 125. Funazo T, Nomizo T, Kim YH. Liver metastasis is associated with poor progression-free survival in patients with non-small cell lung cancer treated with nivolumab. J Thorac Oncol. (2017) 12:e140–1. doi: 10.1016/j.jtho.2017.04.027
- 126. Ambe PC, Godde D, Storkel S, Zirngibl H, Bonicke L. Extra nodular metastasis is a poor prognostic factor for overall survival in node-positive patients with colorectal cancer. *Int J Colorectal Dis.* (2018) 33:403–9. doi: 10.1007/s00384-018-2991-0
- 127. Mei Z, Shi L, Wang B, Yang J, Xiao Z, Du P, et al. Prognostic role of pretreatment blood neutrophil-to-lymphocyte ratio in advanced cancer survivors: a systematic review and meta-analysis of 66 cohort studies. *Cancer Treat Rev.* (2017) 58:1–13. doi: 10.1016/j.ctrv.2017. 05.005
- 128. Mei Z, Liang M, Li L, Zhang Y, Wang Q, Yang W. Effects of statins on cancer mortality and progression: a systematic review and meta-analysis of 95 cohorts including 1,111,407 individuals. *Int J Cancer*. (2017) 140:1068–81. doi: 10.1002/ijc.30526
- 129. Xiao Y, Zheng L, Mei Z, Xu C, Liu C, Chu X, et al. The impact of metformin use on survival in prostate cancer: a systematic review and meta-analysis. Oncotarget. (2017) 8:100449–58. doi: 10.18632/oncotarget. 22117

- 130. Mei ZB, Zhang ZJ, Liu CY, Liu Y, Cui A, Liang ZL, et al. Survival benefits of metformin for colorectal cancer patients with diabetes: a systematic review and meta-analysis. *PLoS ONE.* (2014) 9:e91818. doi: 10.1371/journal.pone.0091818
- 131. Rifai K, Judes G, Idrissou M, Daures M, Bignon YJ, Penault-Llorca F, et al. Dual SIRT1 expression patterns strongly suggests its bivalent role in human breast cancer. *Oncotarget.* (2017) 8:110922–30. doi: 10.18632/oncotarget.23006
- 132. Subramaniyan B, Kumar V, Mathan G. Effect of sodium salt of Butrin, a novel compound isolated from Butea monosperma flowers on suppressing the expression of SIRT1 and Aurora B kinase-mediated apoptosis in colorectal cancer cells. *Biomed Pharmacother*. (2017) 90:402–13. doi: 10.1016/j.biopha.2017.03.086

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Sun, Du, Zhang, Xiong, Gong, Lei, Zha, Zhu, Li, Huang and Gu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.