



Molecular signatures and phylogenomic analysis of the genus *Burkholderia*: proposal for division of this genus into the emended genus *Burkholderia* containing pathogenic organisms and a new genus *Paraburkholderia* gen. nov. harboring environmental species

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The genus *Burkholderia* contains large number of diverse species which include many clinically important organisms, phytopathogens, as well as environmental species. However, currently, there is a paucity of biochemical or molecular characteristics which can reliably distinguish different groups of *Burkholderia* species. We report here the results of detailed phylogenetic and comparative genomic analyses of 45 sequenced species of the genus *Burkholderia*. In phylogenetic trees based upon concatenated sequences for 21 conserved proteins as well as 16S rRNA gene sequence based trees, members of the genus *Burkholderia* grouped into two major clades. Within these main clades a number of smaller clades including those corresponding to the clinically important *Burkholderia cepacia* complex (BCC) and the *Burkholderia pseudomallei* groups were also clearly distinguished. Our comparative analysis of protein sequences from *Burkholderia* spp. has identified 42 highly specific molecular markers in the form of conserved sequence indels (CSIs) that are uniquely found in a number of well-defined groups of *Burkholderia* spp. Six of these CSIs are specific for a group of *Burkholderia* spp. (referred to as Clade I in this work) which contains all clinically relevant members of the genus (viz. the BCC and the *B. pseudomallei* group) as well as the phytopathogenic *Burkholderia* spp. The second main clade (Clade II), which is composed of environmental *Burkholderia* species, is also distinguished by 2 identified CSIs that are specific for this group. Additionally, our work has also identified multiple CSIs that serve to clearly demarcate a number of smaller groups of *Burkholderia* spp. including 3 CSIs that are specific for the *B. cepacia* complex, 4 CSIs that are uniquely found in the *B. pseudomallei* group, 5 CSIs that are specific for the phytopathogenic *Burkholderia* spp. and 22 other CSI that distinguish two groups within Clade II. The described molecular markers provide highly specific means for the demarcation of different groups of *Burkholderia* spp. and they also offer novel and useful targets for the development of diagnostic assays for the clinically important members of the BCC or the *pseudomallei* groups. Based upon the results of phylogenetic analyses, the identified CSIs and the pathogenicity profile of *Burkholderia* species, we are proposing a division of the genus *Burkholderia* into two genera. In this new proposal, the emended genus *Burkholderia* will correspond to the Clade I and it will contain only the clinically relevant and phytopathogenic *Burkholderia* species. All other *Burkholderia* spp., which are primarily environmental, will be transferred to a new genus *Paraburkholderia* gen. nov.

Keywords: *Burkholderia*, *Burkholderia cepacia* complex, conserved signature indels, phylogenetic trees, molecular signatures

INTRODUCTION

The genus *Burkholderia* is a morphologically, metabolically, and ecologically diverse group of gram-negative bacteria (Yabuuchi et al., 1992; Coenye and Vandamme, 2003; Mahenthiralingam et al., 2005; Palleroni, 2005; Compant et al., 2008). *Burkholderia* species are ubiquitous in the environment (Coenye and

Vandamme, 2003). They inhabit a wide range of ecological niches, ranging from soil to the human respiratory tract (Coenye and Vandamme, 2003). A group of 17 closely related *Burkholderia* species, the *Burkholderia cepacia* complex (BCC), are responsible for prevalent and potentially lethal pulmonary infections in immunocompromised individuals, such as individuals with cystic

fibrosis (Mahenthiralingam et al., 2002, 2005; Biddick et al., 2003; Hauser et al., 2011). *Burkholderia pseudomallei*, a *Burkholderia* species related to the BCC, is the causative agent for the disease melioidosis, a potentially lethal septic infection which accounts for up to 20% of all community-acquired septicemias in some regions (White, 2003; Limmathurotsakul and Peacock, 2011). Other species related to the BCC are the causative agents of major infections in both animals (*Burkholderia mallei*) and plants (*Burkholderia glumae* and *Burkholderia gladioli*) (Whitlock et al., 2007; Nandakumar et al., 2009).

In spite of the large diversity and varied pathogenicity among the >70 members of the group, all *Burkholderia* species are currently placed within one genus (Coenye and Vandamme, 2003; Palleroni, 2005). The phylogeny and taxonomy of the genus *Burkholderia* is primarily defined on the basis of 16S rRNA sequence analysis (Yabuuchi et al., 1992; Palleroni, 2005; Yarza et al., 2008). The inferences obtained from 16S rRNA analysis have been further substantiated by other phylogenetic methods, including *recA* gene based analysis (Payne et al., 2005), *acdS* gene based analysis (Onofre-Lemus et al., 2009), DNA–DNA hybridization (Gillis et al., 1995), whole cell fatty acid analysis (Stead, 1992), multilocus sequence analysis (Tayeb et al., 2008; Spilker et al., 2009; Estrada-de los Santos et al., 2013), gene gain/loss analysis (Zhu et al., 2011), and whole genome phylogenetic analysis (Ussery et al., 2009; Segata et al., 2013). In many of these phylogenetic studies, the members of the genus *Burkholderia* can be divided into two or more distinct phylogenetic groups, with one group consisting of members of the BCC and related species (Payne et al., 2005; Tayeb et al., 2008; Yarza et al., 2008; Spilker et al., 2009; Ussery et al., 2009; Gyaneshwar et al., 2011; Vandamme and Dawyndt, 2011; Zhu et al., 2011; Estrada-de los Santos et al., 2013; Segata et al., 2013). Although there are some commonly shared features among closely related groups of *Burkholderia* species, there is no known morphological, biochemical, or molecular characteristic specific to the larger phylogenetic groups within the genus (ex. the BCC and related species).

The advent of next generation sequencing methods has led to a rapid increase in the number of genome sequences available for bacterial species (Mardis, 2008). The availability of these sequences for members of the genus *Burkholderia* provides us better means to evaluate the phylogenetic relationships among different species (Ciccarelli et al., 2006; Wu et al., 2009). Importantly, the large data sets of sequences allows for the use of comparative genomic techniques to discover novel molecular markers that can provide independent evidence for different phylogenetic groups within the genus *Burkholderia* (Gupta, 1998, 2014; Gao and Gupta, 2012). In this work, we describe one type of molecular marker, conserved sequence insertions or deletions (CSIs), which are uniquely present in protein sequences from a defined group of organisms, that can be used to delineate different phylogenetic groups of *Burkholderia* species independently of traditional phylogenetic methods (Gupta, 1998, 2001; Gao and Gupta, 2012). Our comparative analysis of *Burkholderia* genomes has led to the identification of 42 unique CSIs that delineate different phylogenetic groups within the genus in clear molecular terms. A clade of *Burkholderia* containing the BCC and related organisms (Clade

I) was supported by both phylogenetic evidence and 6 identified CSIs. We have also identified 3 CSIs specific for the BCC, 4 CSIs specific for the *B. pseudomallei* group, and 5 CSIs specific for the plant pathogenic *Burkholderia* spp. The remaining members of the genus *Burkholderia* formed another monophyletic clade (Clade II) in our phylogenetic trees which was supported by 2 CSIs. Within Clade II, we identified two smaller clades of *Burkholderia* that were supported by 16 and 6 CSIs. The grouping of members of the genus *Burkholderia* into at least two large, monophyletic groups has also been observed in a large body of prior phylogenetic research (Payne et al., 2005; Tayeb et al., 2008; Yarza et al., 2008; Spilker et al., 2009; Ussery et al., 2009; Gyaneshwar et al., 2011; Zhu et al., 2011; Estrada-de los Santos et al., 2013; Segata et al., 2013). Based on the phylogenetic evidence and our identified CSIs, we propose division of the genus *Burkholderia* into two genera: an emended genus *Burkholderia* containing clinically important and phytopathogenic members of the genus and a new genus *Paraburkholderia* gen. nov. harboring the environmental species.

MATERIALS AND METHODS

PHYLOGENETIC ANALYSIS

A concatenated sequence alignment of 21 highly conserved proteins (viz. ArgRS, EF-G, GyrA, GyrB, Hsp60, Hsp70, IleRS, RecA, RpoB, RpoC, SecY, ThrRS, TrpS, UvrD, ValRS, 50S ribosomal proteins L1, L5 and L6, and 30S ribosomal proteins S2, S8 and S11) was used to perform phylogenetic analysis. Due to their presence in most bacteria, these proteins have been extensively utilized for phylogenetic studies (Gupta, 1998, 2009; Kyripides et al., 1999; Harris et al., 2003; Charlebois and Doolittle, 2004; Ciccarelli et al., 2006). The amino acid sequences for these conserved proteins were obtained from NCBI database for all of the species/strains listed in **Table 1**, which includes 45 sequenced species of the genus *Burkholderia*. Furthermore, three genomes from other members of class *Betaproteobacteria* (viz. *Cupriavidus necator* N-1, *Bordetella pertussis* Tohama I, and *Neisseria meningitidis* MC58), serving as outgroups in our analysis, were also retrieved from NCBI database. Depending on genome availability, type strains were selected for most of the species. Multiple sequence alignments for these proteins were created using Clustal_X 1.83 and concatenated into a single alignment file (Jeanmougin et al., 1998). Poorly aligned regions from the alignment file were removed using Gblocks 0.91b and the resulting alignment, which contained 7688 aligned characters, was ultimately utilized for phylogenetic analysis (Castresana, 2000). A maximum likelihood (ML) tree based on 100 bootstrap replicates of this alignment was constructed using MEGA 6.0 while employing Jones–Taylor–Thornton substitution model (Jones et al., 1992; Tamura et al., 2013).

A maximum likelihood 16S rRNA gene sequence consensus tree was also created for 101 sequences, which included 97 representative strains from the genus *Burkholderia* and four outgroup sequences from the genera *Cupriavidus* and *Ralstonia*. The sequences utilized in the study were obtained from the Ribosomal Database Project (RDP III) (Cole et al., 2009) and NCBI. All the sequences were aligned using MAAFT 7 (Kato and Standley, 2013) and a ML tree based upon 1000 bootstrap replicates of

Table 1 | Genome characteristics of the sequenced members of the genus *Burkholderia*.

Organism	BioProject	Size (Mb)	GC%	Chromosomes	Proteins	References
<i>Burkholderia cenocepacia</i> J2315	PRJNA57953	8.06	66.9	3	7116	Holden et al., 2009
<i>Burkholderia pseudomallei</i> K96243	PRJNA57733	7.25	68.1	2	5727	Holden et al., 2004
<i>Burkholderia mallei</i> ATCC 23344	PRJNA57725	5.84	68.5	2	5022	Nierman et al., 2004
<i>Burkholderia thailandensis</i> E264	PRJNA58081	6.72	67.6	2	5632	Kim et al., 2005
<i>Burkholderia oklahomensis</i> C6786	PRJNA54789	6.99	67.0	–	6954	NMRC ^b
<i>Burkholderia multivorans</i> ATCC 17616	PRJNA58909	7.01	66.7	3	6111	DOE ^d
<i>Burkholderia ambifaria</i> AMMD	PRJNA58303	7.53	66.8	3	6610	Coenye et al., 2001b
<i>Burkholderia glumae</i> BGR1	PRJNA59397	7.28	67.9	2	5773	Lim et al., 2009
<i>Burkholderia xenovorans</i> LB400	PRJNA57823	9.73	62.6	3	8702	Chain et al., 2006
<i>Burkholderia</i> sp. CCGE1002	PRJNA42523	7.88	63.3	3	6889	Ormeno-Orrillo et al., 2012
<i>Burkholderia</i> sp. CCGE1001	PRJNA42975	6.83	63.6	2	5965	DOE ^d
<i>Burkholderia</i> sp. CCGE1003	PRJNA46253	7.04	63.2	2	5988	DOE ^d
<i>Burkholderia</i> sp. Ch1-1	PRJNA48975	8.74	62.4	–	7742	DOE ^d
<i>Burkholderia</i> sp. H160	PRJNA55101	7.89	62.9	–	7460	Ormeno-Orrillo et al., 2012
<i>Burkholderia</i> sp. 383	PRJNA58073	8.68	66.3	3	7716	DOE ^d
<i>Burkholderia spreintiae</i> WSM5005	PRJNA66661	7.76	63.2	–	–	DOE ^d
<i>Burkholderia</i> sp. Y123	PRJNA81081	8.90	63.3	3	7804	Lim et al., 2012
<i>Burkholderia</i> sp. SJ98	PRJNA160003	7.88	61.4	–	7268	Kumar et al., 2012
<i>Burkholderia</i> sp. WSM2230	PRJNA165309	6.31	63.1	–	–	DOE ^d
<i>Burkholderia</i> sp. KJ006	PRJNA165871	6.63	67.2	3	6024	Kwak et al., 2012
<i>Burkholderia</i> sp. TJJ49	PRJNA179699	7.38	66.9	–	8940	Khan et al., 2013
<i>Burkholderia</i> sp. BT03	PRJNA180532	10.64	61.9	–	10126	Oak Ridge ^c
<i>Burkholderia</i> sp. WSM2232	PRJNA182741	7.21	63.1	–	–	DOE ^d
<i>Burkholderia</i> sp. WSM3556	PRJNA182743	7.68	61.8	–	–	DOE ^d
<i>Burkholderia</i> sp. URHA0054	PRJNA190816	7.24	62.8	–	–	DOE ^d
<i>Burkholderia</i> sp. WSM4176	PRJNA199219	9.07	62.9	–	8336	DOE ^d
<i>Burkholderia</i> sp. JPY251	PRJNA199221	8.61	63.1	–	7873	DOE ^d
<i>Burkholderia</i> sp. JPY347	PRJNA199222	6.39	63.1	–	5963	DOE ^d
<i>Burkholderia</i> sp. RPE64	PRJNA205541	6.96	63.1	3	6498	Shibata et al., 2013
<i>Burkholderia vietnamiensis</i> G4	PRJNA58075	8.39	65.7	3	7617	DOE ^d
<i>Burkholderia dolosa</i> AUO158	PRJNA54351	6.42	66.8	–	4795	Broad Institute ^a
<i>Burkholderia phymatum</i> STM815	PRJNA58699	8.68	62.3	2	7496	Vandamme et al., 2002b
<i>Burkholderia phytofirmans</i> PsJN	PRJNA58729	8.21	62.3	2	7241	Weilharter et al., 2011
<i>Burkholderia ubonensis</i> Bu	PRJNA54793	6.93	67.3	–	7181	NMRC ^b
<i>Burkholderia graminis</i> C4D1M	PRJNA54887	7.48	62.9	–	6747	DOE ^d
<i>Burkholderia rhizoxinica</i> HKI 454	PRJNA60487	3.75	60.7	1	3870	Lackner et al., 2011
<i>Burkholderia gladioli</i> BSR3	PRJNA66301	9.05	67.4	2	7411	Seo et al., 2011
<i>Burkholderia cepacia</i> GG4	PRJNA173858	6.47	66.7	2	5825	Hong et al., 2012
<i>Candidatus Burkholderia kirkii</i> UZHbot1	PRJNA74017	4.01	62.9	–	2069	Van Oevelen et al., 2002b
<i>Burkholderia mimosarum</i> LMG 23256	PRJNA163559	8.41	63.9	–	–	DOE ^d
<i>Burkholderia terrae</i> BS001	PRJNA168186	11.29	61.8	–	10234	Nazir et al., 2012
<i>Burkholderia pyrrocinia</i> CH-67	PRJNA199595	8.05	67.4	–	7324	Song et al., 2012
<i>Burkholderia kururiensis</i> M130	PRJNA199910	7.13	65.0	–	6311	Coutinho et al., 2013
<i>Burkholderia phenoliruptrix</i> BR3459a	PRJNA176370	7.65	63.1	2	6496	Oliveira Cunha et al., 2012
<i>Burkholderia bryophila</i> 376MFSHa3.1	PRJNA201182	7.38	61.9	–	6722	DOE ^d

^a The Broad Institute Genome Sequencing Platform (Broad Institute).^b Naval Medical Research Center/ Biological Defense Research Directorate (NMRC).^c Oak Ridge National Lab (Oak Ridge).^d DOE Joint Genome Institute (DOE).

this alignment was constructed using the General Time Reversible Model (Tavaré, 1986) in MEGA 6.0 (Tamura et al., 2013).

IDENTIFICATION OF MOLECULAR MARKERS (CSIs)

BLASTp searches were conducted for all proteins from chromosomes 2 and 3 (accession numbers NC_008061 and NC_008061) of *Burkholderia cenocepacia* J2315 (Holden et al., 2009) to identify CSIs that are shared by different members of the genus *Burkholderia*. Species that appeared as top hits with high scoring homologs (E values $< 1e^{-20}$) from the genus *Burkholderia* and other outgroups were selected. Multiple sequence alignments were created using the Clustal_X 1.83 (Jeanmougin et al., 1998). These alignments were visually inspected for the presence of insertions or deletions (indels) restricted to either some or all members of the genus *Burkholderia* and flanked by at least 5–6 conserved amino acid residues on both sides in the neighboring 30–40 amino acids. Indel queries that were not flanked by conserved regions were not further evaluated. The species specificity of the indel queries meeting the above criterion was further evaluated by performing BLASTp searches on short sequence segments containing the insertions or deletions, and their flanking conserved regions (60–100 amino acids long). The searches were conducted against the NCBI non-redundant (nr) database and a minimum of 250 BLAST hits were examined for the presence or absence of CSIs. The results of these analyses were evaluated as described in detail in our recent work (Gupta, 2014). Signature files for the CSIs that were specific for members of the genus *Burkholderia* were created and formatted using the programs SIG_CREATE and SIG_STYLE (accessible from Gleans.net) as described by Gupta (2014). The sequence alignment files presented here contain information for all detected insertions or deletions from the *Burkholderia* group of interest, but only a limited number from species that are serving as outgroups. Sequence information for different strains of various species is not shown, but they all exhibited similar pattern. Lastly, unless otherwise indicated, the CSIs shown here are specifically found in the indicated groups and similar CSIs were not detected in the 250 Blast hits with the query sequences.

RESULTS

BRANCHING PATTERN OF *BURKHOLDERIA* SPECIES IN CONCATENATED PROTEIN AND 16S rRNA TREES

Genome sequences of 45 species of *Burkholderia* were available from the NCBI genome database at the time of this work (NCBI, 2014). Some characteristics of these genomes are listed in **Table 1**. The genome sizes of the sequenced *Burkholderia* species show large variation (from 3.75–11.29 Mb) and the numbers of proteins in them also varied in a similar proportion. In this work we have produced a ML phylogenetic tree based on the concatenated amino acid sequences of 21 conserved housekeeping and ribosomal proteins obtained from 45 sequenced *Burkholderia* species (**Figure 1**). The *Burkholderia* species formed two large clades in the protein based ML tree: One consisting of the BCC and related organisms (Clade I) and another comprised mainly of environmental or poorly characterized *Burkholderia* species (Clade II). Within Clade I, three smaller, distinct clades are also observed. The first of these clades (Clade Ia) is wholly comprised

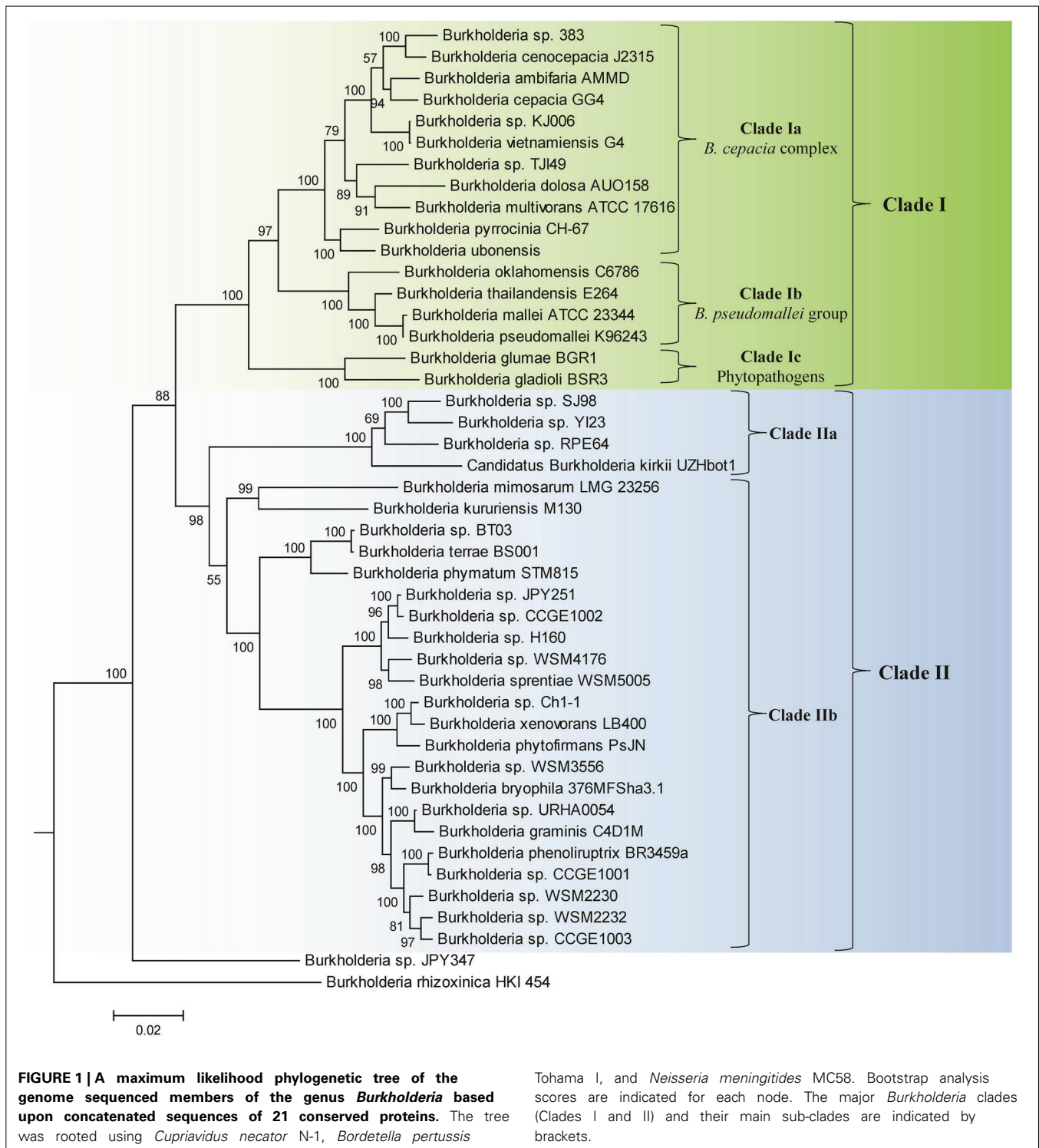
of the sequenced BCC species, the second clade (Clade Ib) groups *B. pseudomallei* and closely related species, and the third clade (Clade Ic) consists of the plant pathogenic species, *B. glumae* and *B. gladioli*. Clade II could also be divided into two smaller clades, Clade IIa and Clade IIb. Clade IIa is separated from Clade IIb by a long branch, suggesting that a large amount of genetic divergence has occurred between the two groups. In addition to the two main clades of *Burkholderia*, two species, *Burkholderia* sp. JPY347 and *Burkholderia rhizoxinica*, branched early in the tree and did not associate with either Clade I or II.

We have also constructed a 16S rRNA based ML phylogenetic tree for 97 *Burkholderia* strains and candidate species (**Figure 2**). In this 16S rRNA based phylogenetic tree we observed broadly similar patterns to our protein based phylogeny. A clade consisting of the BCC and related organisms (Clade I) was clearly resolved. The three subclades within Clade I, the BCC (Clade Ia), the *B. pseudomallei* group (Clade Ib), and the plant pathogenic species (Clade Ic) were well resolved, though some species exhibited aberrant branching (ex. *B. oklahomensis* and *B. pseudomultivorans*). A large assemblage of the remaining *Burkholderia* species, roughly corresponding to Clade II in our concatenated protein based phylogenetic tree, was also observed in the 16S rRNA tree. However, due to significant number of unsequenced *Burkholderia* species which are present in the 16S rRNA database it is difficult to accurately identify the groups within Clade II of the 16S rRNA tree which correspond to Clades IIa and IIb in our concatenated protein based phylogenetic tree. Bootstrap support for branches in the 16S rRNA based tree were also significantly lower than they were in the concatenated protein tree indicating that some of the observed branching patterns may not be reliable. However, the clade consisting of the BCC and related organisms (Clade I) has strong bootstrap support and has been identified in a large number of previous 16S rRNA based phylogenetic studies (Yabuuchi et al., 1992; Palleroni, 2005; Yarza et al., 2008; Suarez-Moreno et al., 2012).

MOLECULAR SIGNATURES DISTINGUISHING THE CLADE I AND CLADE II *BURKHOLDERIA*

Rare genetic changes, such as insertions and deletions in essential genes/proteins, which occur in a common ancestor can be inherited by the various decedent species related to this common ancestor (Gupta, 1998; Rokas and Holland, 2000; Gogarten et al., 2002; Gupta and Griffiths, 2002). Due to the rarity and the specific presence of these rare genetic changes to a related group of organisms, they can serve as important molecular markers and provide a novel means to understand the evolutionary interrelationships between different closely related species (Gupta, 1998; Gupta and Griffiths, 2002; Gao and Gupta, 2012).

The comparative analysis of protein sequences from *Burkholderia* species that was carried out in the present work has identified a number of CSIs that serve to clearly distinguish a number of different clades within the genus *Burkholderia*. These studies have led to identification of 6 CSIs that are specific for the Clade I *Burkholderia*, consisting of the BCC and related organisms, enabling clear distinction of this group from all other *Burkholderia*. This clade, which contains all well characterized pathogens within the genus, represents the most clinically



relevant group within the *Burkholderia*. All species within this clade are potentially pathogenic to human, animals, or plants and most have been isolated from clinical human samples (Simpson et al., 1994; Mahenthiralingam et al., 2002, 2005; Biddick et al., 2003; O'Carroll et al., 2003). One example of a CSI that is specific to the Clade I *Burkholderia* is shown in **Figure 3A**. In this case, a one amino acid deletion is present in a highly conserved region

of a periplasmic amino acid-binding protein. The indel is flanked on both sides by highly conserved regions indicating that it is not the result of alignment artifacts and that it is a reliable genetic characteristic. This CSI is present in all of the sequenced members of the Clade I *Burkholderia*, but absent in all other bacterial homologs of this protein. Our work has identified 5 additional CSIs in other widely distributed proteins that are

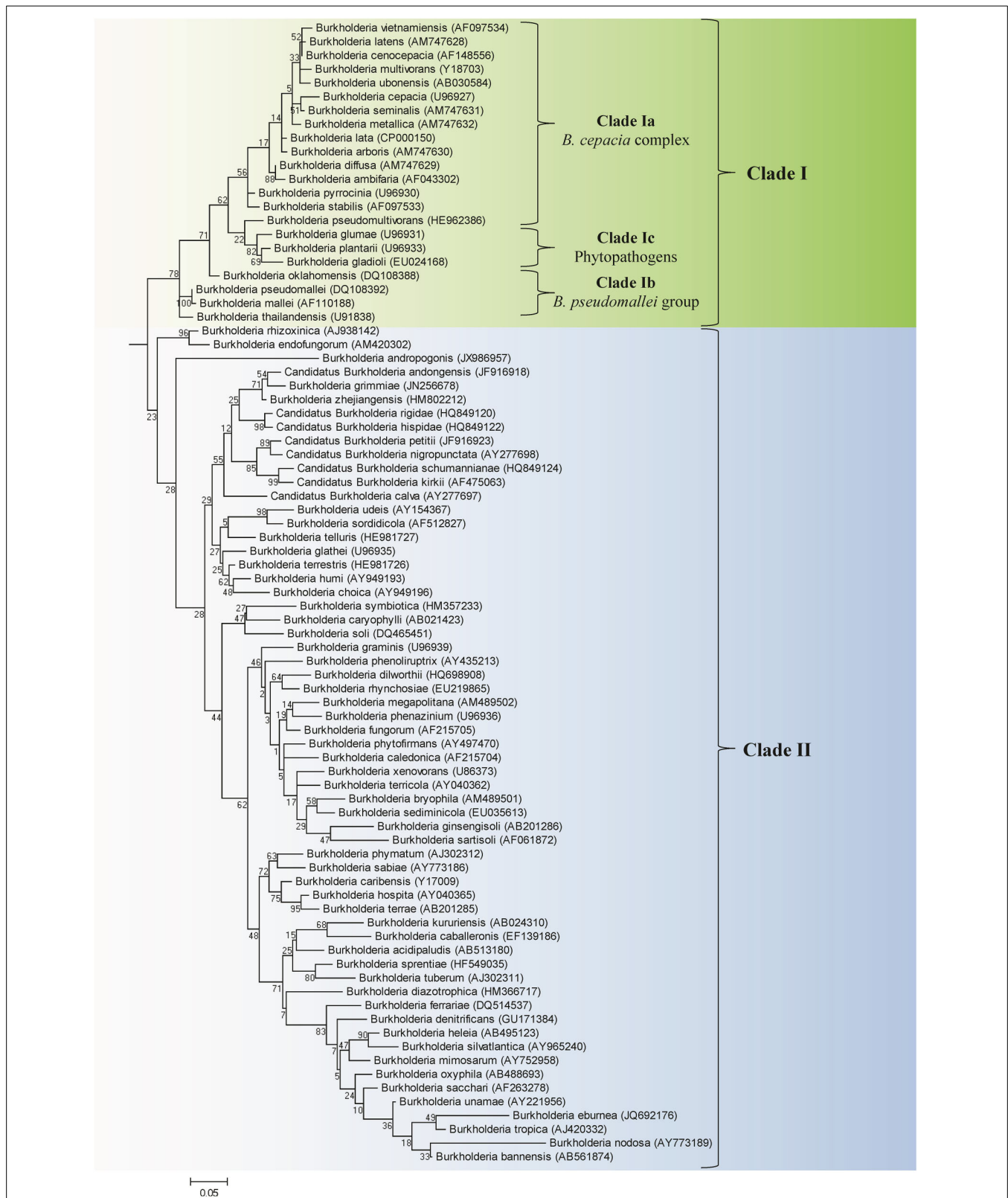


FIGURE 2 | A maximum likelihood tree based on the 16S rRNA gene sequences of 97 members of the genus *Burkholderia*. Accession numbers for the 16S rRNA sequenced used for each organism are provided in the brackets following the name of the organism. The tree

was rooted using four species from the genera *Cupriavidus* and *Ralstonia*. Bootstrap analysis scores are indicated for each node. The major *Burkholderia* clades (Clades I and II) and the subclades within Clade I are indicated by brackets.

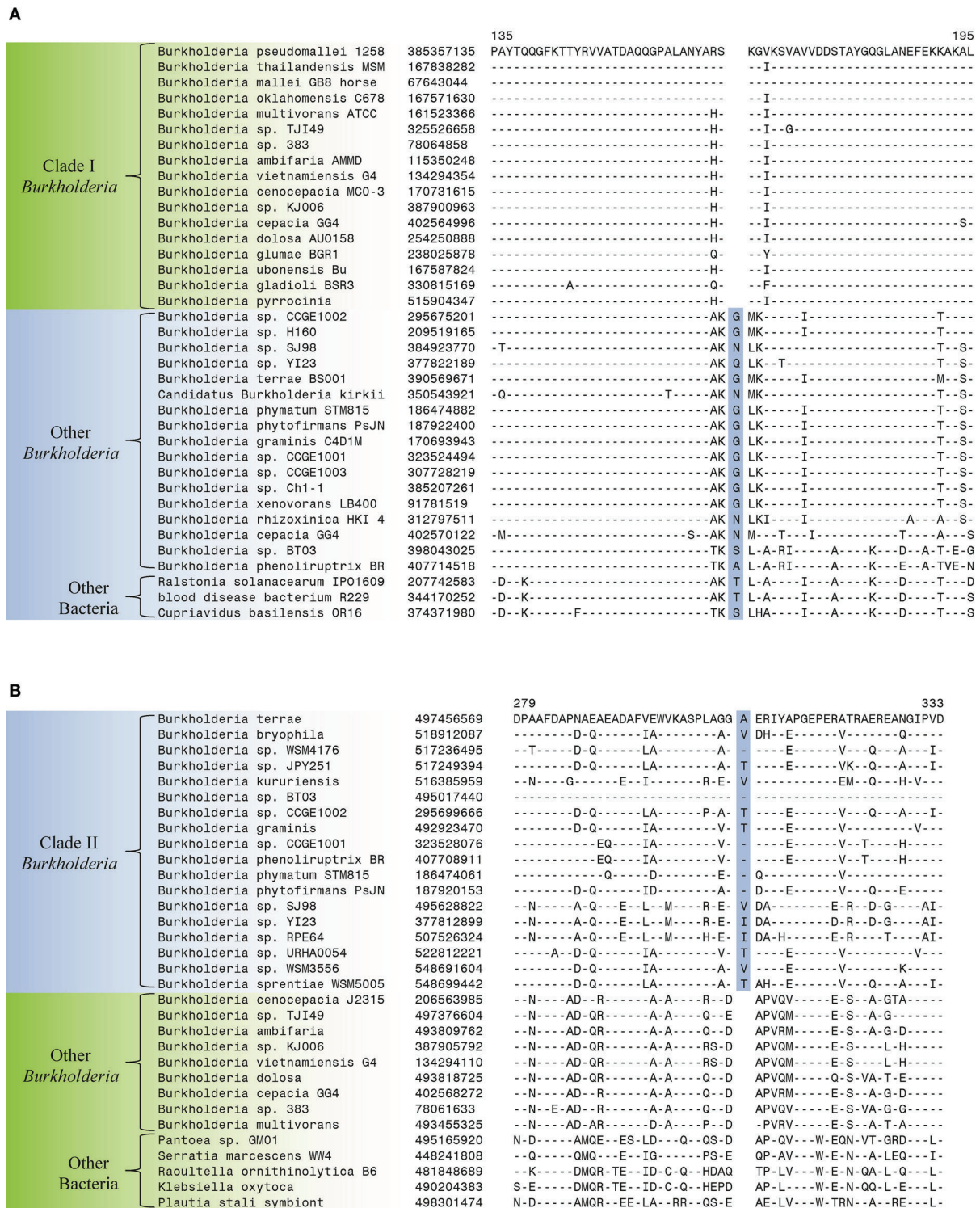


FIGURE 3 | Partial sequence alignments of (A) a periplasmic amino acid-binding protein showing a 1 amino acid deletion identified in all members of Clade I of the genus *Burkholderia* (B) a dehydrogenase showing a 1 amino acid insertion (boxed) identified only in members of Clade II of the genus *Burkholderia*. These CSIs were not found in the sequence homologs of these proteins from any other sequenced bacteria. In each case, sequence information for a *Burkholderia* species and a limited number other bacteria are shown, but unless otherwise indicated, similar

CSIs were detected in all members of the indicated group and not detected in any other bacterial species in the top 250 BLAST hits. The dashes (-) in the alignments indicate identity with the residue in the top sequence. GenBank identification (GI) numbers for each sequence are indicated in the second column. Sequence information for other CSIs specific to the members of Clade I and Clade II of the genus *Burkholderia* are presented in Supplemental Figures 1–5 and Supplemental Figure 6, respectively, and their characteristics are summarized in **Table 2**.

specific for the Clade I *Burkholderia* and sequence alignments for these CSIs are shown in Supplemental Figures 1–5 and a summary of their characteristics is provided in **Table 2**.

Two additional CSIs identified in this work are specific for the Clade II *Burkholderia* species which is made up of mainly environmental organisms. One of these CSIs, shown in **Figure 3B**, consists of a one amino acid insertion in a dehydrogenase protein that is uniquely found in members of the Clade II *Burkholderia* and absent in all other *Burkholderia* species as well all other bacterial groups. A sequence alignment for another CSI that is specific for the Clade II *Burkholderia* (a 2 aa deletion in a LysR family of transcription regulator protein) is shown in Supplemental Figure 6 and its characteristics are summarized in **Table 2**.

CSIs DISTINGUISHING DIFFERENT MAIN GROUPS WITHIN THE CLADE I BURKHOLDERIA

The species within Clade I of the genus *Burkholderia* are responsible for a range of human, animal, and plant diseases (Biddick et al., 2003; Mahenthiralingam et al., 2005). The members of Clade I (i.e., the BCC and related *Burkholderia*) are commonly separated into 3 main groups which correspond to clades identified in our phylogenetic trees. The first group, the members of the BCC (Clade 1a), are prevalent pathogens in cystic fibrosis patients, the second group, the *B. pseudomallei* group (Clade 1b), contains the causative agents of melioidosis and glanders, while the third group contains the plant pathogenic *Burkholderia* species (Clade 1c) (White, 2003; Mahenthiralingam et al., 2005; Whitlock et al., 2007; Nandakumar et al., 2009). Our analysis has identified 3 CSIs that are specific for all members of the BCC clade (Clade 1a). One example of a BCC clade specific CSI is shown in **Figure 4A**. This CSI consists of a 2 amino acid insertion in a conserved region of a histidine utilization repressor which is only found in members of the BCC. Sequence alignments for two other BCC clade specific CSIs are shown in Supplemental Figures 7, 8 and their characteristics are summarized in **Table 3**.

Our work has also identified 4 CSIs that are specific for the *B. pseudomallei* group (Clade 1b) which contains the most prevalent human pathogen within the genus, *B. pseudomallei* (Wiersinga et al., 2006). One example of a CSI specific to the *B. pseudomallei* group, which consists of a 1 amino acid insertion in a conserved region of a periplasmic oligopeptide-binding

protein, is shown in **Figure 4B**. Sequence alignments for three other CSIs in three different proteins that are specific for the *B. pseudomallei* group are shown in Supplemental Figures 9–11 and their characteristics are summarized in **Table 3**.

We have also identified 5 CSIs that are specific for the major plant pathogenic group within the genus *Burkholderia* (Clade 1c) which contains the species *B. glumae* and *B. gladioli*. An example of a CSI representing this group is shown in **Figure 4C**. This CSI consists of a 1 amino acid insertion in a conserved region of a SMP-30/gluconolactonase/LRE-like region-containing protein that is found in the members of Clade 1c of the genus *Burkholderia* but absent in all other *Burkholderia* and all other bacterial groups. Sequence alignments for the other 4 CSIs are shown in Supplemental Figures 12–15 and their key features are highlighted in **Table 3**.

CSIs THAT ARE SPECIFIC FOR TWO GROUPS WITHIN THE CLADE II BURKHOLDERIA

The species within Clade II of the genus *Burkholderia* inhabit a variety of environmental niches, but there is little evidence of their colonization of healthy or immunocompromised human patients (Coenye and Vandamme, 2003). The branching of different groups within Clade II is not well resolved in 16S rRNA trees and there is currently a lack of sequence data that can be used to generate trees based on concatenated gene sets that reliably resolve the interrelationships of the clade while sufficiently reflecting the total diversity of species within the clade (**Figures 1, 2**) (Cole et al., 2009; NCBI, 2014). Despite the limited sequence data, we have been able to identify two robust groups within Clade II that are supported by a number of CSIs. The first Clade, Clade IIa, primarily consists of unclassified members of the genus and candidatus *Burkholderia* species (**Figure 1**). Clade IIa is supported by 16 CSIs identified in this work. One example of a CSI specific for Clade IIa, consisting of a 1 amino acid insertion in 3-phosphoglycerate dehydrogenase, is shown in **Figure 5A**. This insertion is present in a highly conserved region of this protein in all sequenced members of Clade IIa and absent in all other *Burkholderia* and all other bacterial groups. Sequence alignments for the other 15 CSIs that are specific for Clade IIa *Burkholderia* spp. are shown in Supplemental Figures 16–30 and their characteristics are summarized in **Table 3**.

Table 2 | Conserved signature indels specific for the two major clades within the genus *Burkholderia*.

Protein Name	GI Number	Figures	Indel size	Indel position ^a	Specificity
Periplasmic amino acid-binding protein	385357135	Figure 3A	1 aa del	135–195	Clade I
Putative lyase	167724527	Supplemental Figure 1	1 aa del	70–121	Clade I
4-hydroxybenzoate 3-monooxygenase	238023559	Supplemental Figure 2	1 aa ins	101–171	Clade I
6-phosphogluconate dehydrogenase, decarboxylating	330820932	Supplemental Figure 3	1 aa ins	137–202	Clade I
Putative lipoprotein	121598811	Supplemental Figure 4	1 aa del	363–393	Clade I
Sarcosine oxidase subunit alpha	493818877	Supplemental Figure 5	3 aa ins	904–965	Clade I
Dehydrogenase	497456569	Figure 3B	1 aa ins	279–333	Clade II
LysR family transcriptional regulator	187919777	Supplemental Figure 6	2 aa del	260–294	Clade II

^aThe region of the specified protein that contains the indel.

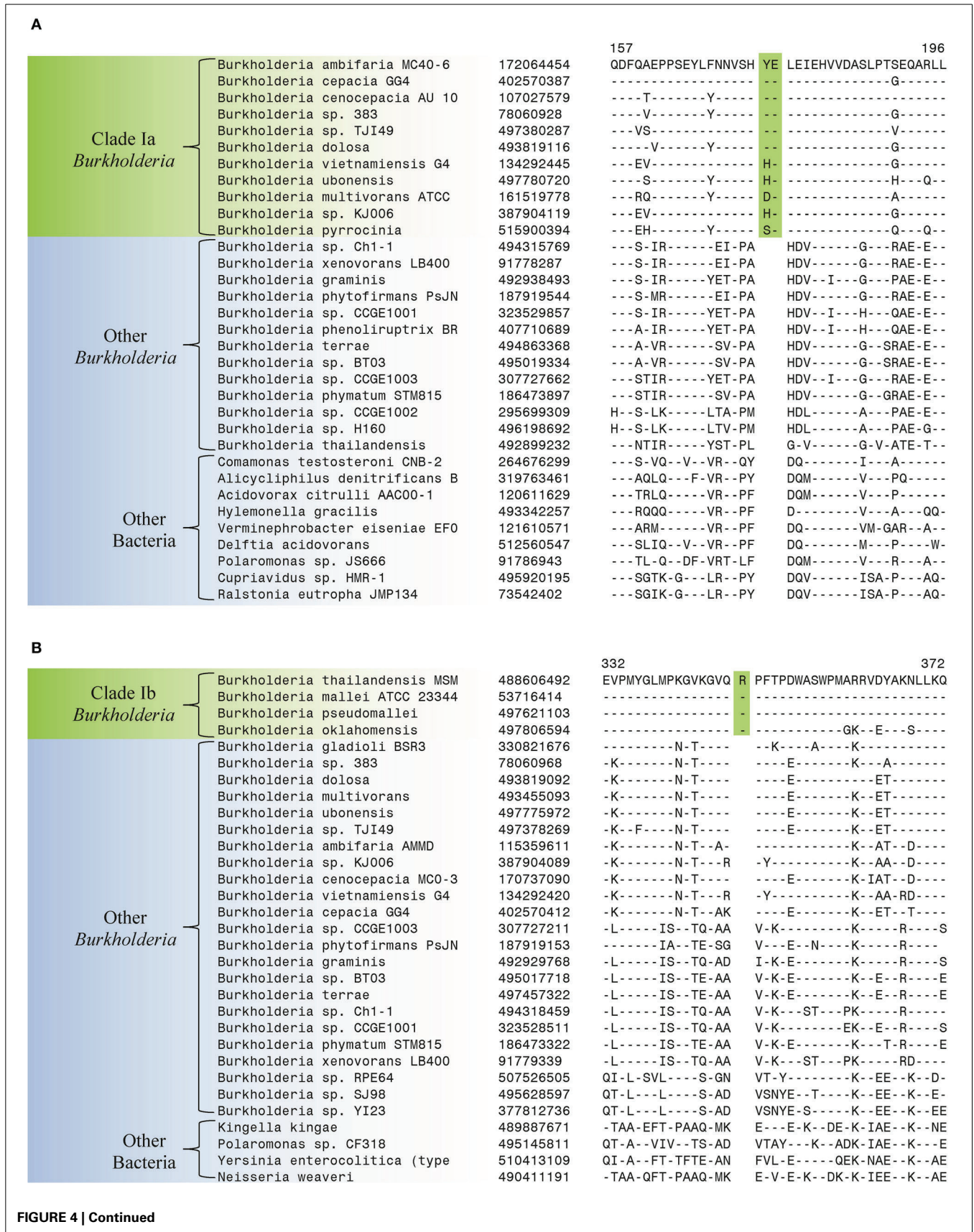
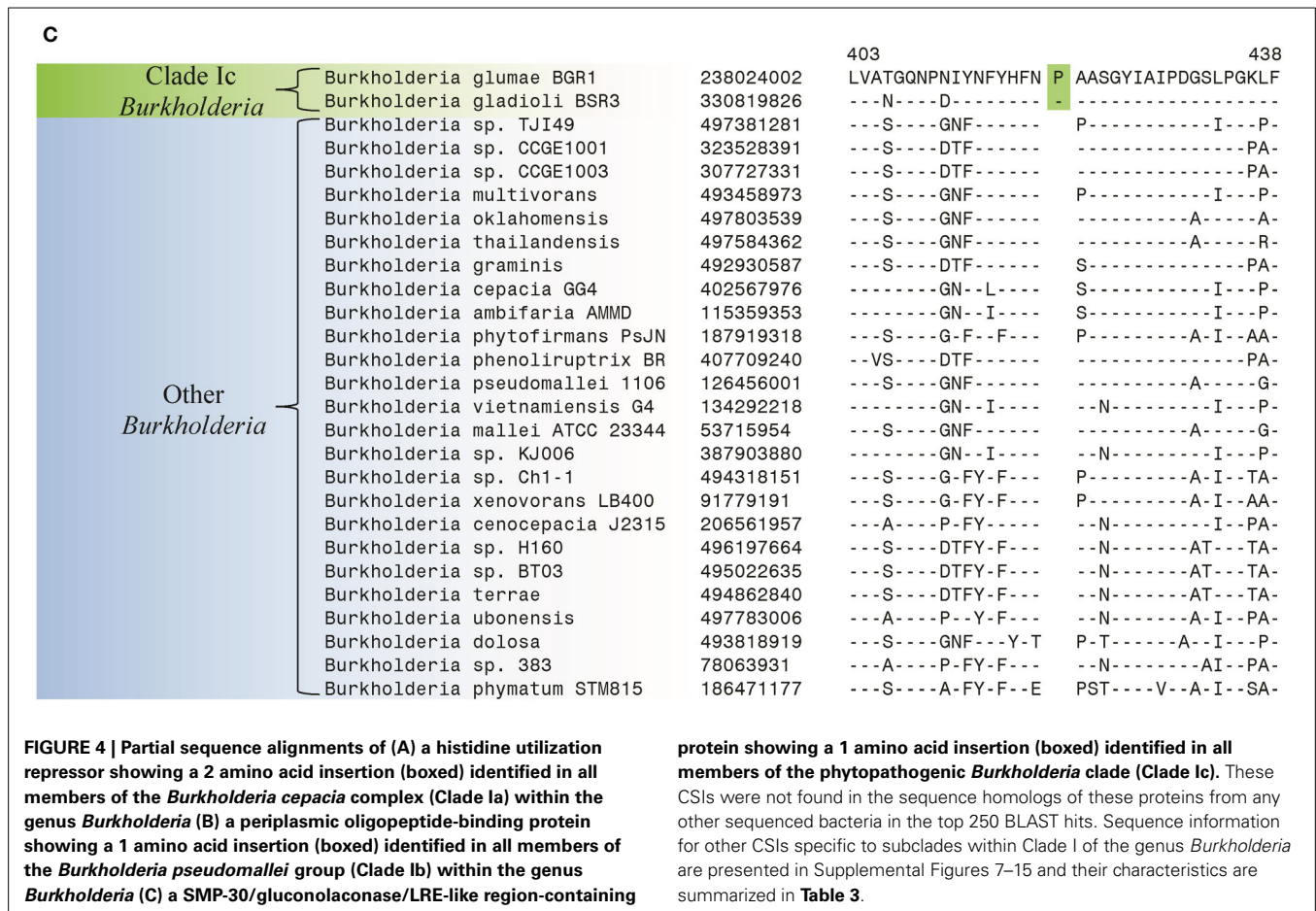


FIGURE 4 | Continued



The second group within Clade II of the *Burkholderia* (Clade IIb), is comprised of a large variety of environmental *Burkholderia* species (Coenye and Vandamme, 2003; Suarez-Moreno et al., 2012). Our analysis has identified 6 CSIs that are specific to this large group of *Burkholderia* species. One example of a CSI specific to the members of Clade IIb of the genus *Burkholderia* is shown in **Figure 5B**. The CSI consists of a one amino acid insertion in 4-hydroxyacetophenone monooxygenase, which is only present in members of Clade IIb of the genus *Burkholderia* and not in protein homologs from any other sequenced bacterial group. Information for other 5 CSIs which are specific to members of Clade IIb of the genus *Burkholderia* are shown in Supplemental Figures 31–35 and their characteristics are summarized in **Table 3**.

DISCUSSION

The genus *Burkholderia* is one of the largest groups of species within the class *Betaproteobacteria* (Palleroni, 2005; Parte, 2013). The genus contains a variety of bacteria that inhabit a wide range of ecological niches including a number of bacteria that have pathogenic potential (Yabuuchi et al., 1992; Coenye and Vandamme, 2003; Mahenthiralingam et al., 2005; Palleroni, 2005; Compant et al., 2008). The phylogeny of the genus *Burkholderia* has been studied using a wide array of methodologies based on phenotypic, biochemical, genetic, and genomic characteristics (Stead, 1992; Gillis et al., 1995; Payne et al., 2005; Tayeb et al.,

2008; Onofre-Lemus et al., 2009; Spilker et al., 2009; Ussery et al., 2009; Gyaneshwar et al., 2011; Vandamme and Dawyndt, 2011; Zhu et al., 2011; Estrada-de los Santos et al., 2013). These studies have provided novel insights into the evolutionary relationship of the species within the genus *Burkholderia*. However, no taxonomic changes have been made to date due to a lack of discrete, distinguishing characteristics identified for the different phylogenetic lineages within the genus (Estrada-de los Santos et al., 2013).

In the present work, we have outlined two major groups of species within the genus *Burkholderia*: Clade I, which contains all pathogenic members of the genus, and Clade II, which contains a large variety of environmental species. These two groups were found to branch distinctly in a highly resolved phylogenetic tree based on a large number of concatenated protein sequences produced in this work (**Figure 1**). Evidence for the distinctness of Clade I organisms from other *Burkholderia* species has been observed in a wide range of previous phylogenetic studies (Payne et al., 2005; Tayeb et al., 2008; Yarza et al., 2008; Spilker et al., 2009; Ussery et al., 2009; Gyaneshwar et al., 2011; Vandamme and Dawyndt, 2011; Zhu et al., 2011; Suarez-Moreno et al., 2012; Estrada-de los Santos et al., 2013; Segata et al., 2013). Importantly, we have also identified 6 and 2 CSIs that serve as discrete molecular characteristics of Clade I and Clade II, respectively (**Figure 6** and **Table 2**). These CSIs are the

Table 3 | Conserved signature indels specific for groups within Clades I and II.

Protein Name	GI Number	Figures	Indel size	Indel position ^a	Specificity
Histidine utilization repressor	172064454	Figure 4A	2 aa ins	157–196	Clade Ia
Molybdate ABC transporter substrate-binding protein	189352411	Supplemental Figure 7	1 aa ins	110–158	Clade Ia
Acid phosphatase	221203041	Supplemental Figure 8	1 aa ins	305–338	Clade Ia
Periplasmic oligopeptide-binding protein	488606492	Figure 4B	1 aa ins	332–372	Clade Ib
OpgC protein	53716883	Supplemental Figure 9	1 aa ins	137–204	Clade Ib
Polysaccharide deacetylase family protein	167725414	Supplemental Figure 10	1 aa ins	29–63	Clade Ib
Thioredoxin domain protein	497613277	Supplemental Figure 11	1 aa ins	247–294	Clade Ib
SMP-30/gluconolactonase/LRE-like region-containing protein	238024002	Figure 4C	1 aa ins	403–438	Clade Ic
Cation efflux protein	330820376	Supplemental Figure 12	1 aa ins	129–160	Clade Ic
putative peptidoglycan-binding LysM/M23B peptidase	238024763	Supplemental Figure 13	1 aa ins	155–198	Clade Ic
SMP-30/gluconolactonase/LRE-like region-containing protein	238024002	Supplemental Figure 14	2 aa del	80–130	Clade Ic
hypothetical protein bgla_2g22890	330821370	Supplemental Figure 15	1 aa ins	322–358	Clade Ic
3-phosphoglycerate dehydrogenase	494056927	Figure 5A	1 aa ins	61–100	Clade IIa
Hypothetical protein BY123_A021470	377821591	Supplemental Figure 16	1 aa del	16–76	Clade IIa
Prepilin peptidase	377821714	Supplemental Figure 17	1 aa ins	179–230	Clade IIa
Uracil-DNA glycosylase	495619839	Supplemental Figure 18	2 aa ins	191–230	Clade IIa
Hypothetical protein BY123_A015260	377820970	Supplemental Figure 19	2 aa ins	221–270	Clade IIa
Carboxylate-amine ligase	377822128	Supplemental Figure 20	1 aa del	321–362	Clade IIa
NADH:ubiquinone oxidoreductase subunit M	494056355	Supplemental Figure 21	3 aa ins	303–348	Clade IIa
NADH:ubiquinone oxidoreductase subunit L	494056354	Supplemental Figure 22	1 aa ins	538–585	Clade IIa
ABC transporter	377821271	Supplemental Figure 23	1 aa del	59–99	Clade IIa
Hypothetical protein BY123_A002220	377819666	Supplemental Figure 24	2 aa ins	133–172	Clade IIa
16S rRNA-processing protein RimM	494056031	Supplemental Figure 25	1 aa ins	147–201	Clade IIa
FAD linked oxidase domain-containing protein	377819737	Supplemental Figure 26	1 aa ins	106–144	Clade IIa
Preprotein translocase subunit SecD	495626933	Supplemental Figure 27	1 aa del	306–341	Clade IIa
Mechanosensitive ion channel protein MscS	494057445	Supplemental Figure 28	3 aa ins	101–143	Clade IIa
Hypothetical protein BY123_A006130	377820057	Supplemental Figure 29	1 aa ins	199–253	Clade IIa
Uroporphyrinogen-III synthase	494056428	Supplemental Figure 30	7 aa ins	37–79	Clade IIa
4-hydroxyacetophenone monooxygenase	496202984	Figure 5B	1 aa ins	380–449	Clade IIb
Transposase A-like protein	187923943	Supplemental Figure 31	1 aa ins	5–50	Clade IIb
Group 1 glycosyl transferase	186475830	Supplemental Figure 32	1 aa ins	153–194	Clade IIb
4-hydroxyacetophenone monooxygenase	496202984	Supplemental Figure 33	3 aa ins	145–219	Clade IIb
Undecaprenyl-phosphate glucose phosphotransferase	209521823	Supplemental Figure 34	1 aa ins	208–275	Clade IIb
putative flavin-binding monooxygenase-like protein	186476032	Supplemental Figure 35	3 aa ins	102–148	Clade IIb

^a The region of the specified protein that contains the indel.

first discrete features that have been identified that are unique to either Clade I or Clade II of the genus *Burkholderia*. These CSIs act as independent verification of the phylogenetic trends identified in this and other studies and provide clear evidence that the species from the Clade I are distinct from all other *Burkholderia* and that they are derived from a common ancestor exclusive of all other *Burkholderia*. Although sequence information for Clade II members is at present somewhat limited, based upon the shared presence of two CSIs by them, it is likely that they are also derived from a common ancestor exclusive of other bacteria.

Additionally, we have identified molecular evidence, in the form of large numbers of CSIs, which support the distinctiveness of several smaller groups within the genus *Burkholderia*. The most important of these groups, the *B. cepacia* complex (BCC; Clade Ia) and the *B. pseudomallei* group (Clade Ib), are

supported by the 3 and 4 of the identified CSIs, respectively. The BCC are a group of opportunistic pathogens which colonize immunodeficient human hosts and are among the most prevalent and lethal infections in cystic fibrosis patients (Mahenthalingam et al., 2002, 2005; Biddick et al., 2003; Hauser et al., 2011). The 17 species that make up the BCC are closely related and form a tight monophyletic cluster within the genus *Burkholderia* (Vandamme and Dawyndt, 2011). The *B. pseudomallei* group consists of 4 closely related species: *B. pseudomallei*, the causative agent of the highly lethal septicemia melioidosis (White, 2003; Limmathurotsakul and Peacock, 2011), *B. mallei*, the causative agent of the equine disease glanders and occasional human infections (Whitlock et al., 2007), and the largely non-pathogenic organisms, *Burkholderia thailandensis* and *Burkholderia oklahomensis* (Deshazer, 2007). The identified CSIs are highly specific characteristics of these two important pathogenic groups and they

A

		61	100
Clade IIa <i>Burkholderia</i>	Candidatus <i>Burkholderia</i> kirkii	494056927	QLLDKCPKLRMISQTGRAGG G HIDIDACTERGIAVLEGTG
	<i>Burkholderia</i> sp. SJ98	495627620	-----
Other <i>Burkholderia</i>	<i>Burkholderia</i> sp. YI23	377819458	-----V-----
	<i>Burkholderia</i> sp. RPE64	507514550	-----F-----
	<i>Burkholderia</i> sp. JPY347	517252484	-----H-----KV-A
	<i>Burkholderia</i> pyrrocinia	515904055	---S-L-H-----I-S
	<i>Burkholderia</i> xenovorans LB400	91781423	-----L-R-----KVSS
	<i>Burkholderia</i> cenocepacia	493541519	---S-L-N-----VSS
	<i>Burkholderia</i> oklahomensis	497786533	---G-L-H-----VST
	<i>Burkholderia</i> phytotfirmans PsJN	187922310	-----L-R-----KVSS
	<i>Burkholderia</i> multivorans ATCC	161526298	---A-L-N-----ISS
	<i>Burkholderia</i> graminis	492932713	-----L-R-----KVSS
	<i>Burkholderia</i> phymatum STM815	186474800	-----L-H-----K-S
	<i>Burkholderia</i> glumae BGR1	238028950	---A-L-H-----KVSS
	<i>Burkholderia</i> dolosa	493815470	---A-L-N-----ISS
	<i>Burkholderia</i> ubonensis	497776121	---A-L-H-----I-T
	<i>Burkholderia</i> thailandensis MSM	488604417	---G-L-H-----VST
	<i>Burkholderia</i> mallei ATCC 23344	53724668	---G-L-H-----VST
	<i>Burkholderia</i> pseudomallei K962	53717757	---G-L-H-----VST
	<i>Burkholderia</i> bryophila	518911408	-----L-R-----KVS
	<i>Burkholderia</i> ambifaria	493802193	---A-L-N-----ISS
	<i>Burkholderia</i> lata	78067952	---A-L-N-----ISS
	<i>Burkholderia</i> cepacia GG4	402565104	---A-L-N-----ISS
	<i>Burkholderia</i> gladioli BSR3	330818693	---A-L-H-----K-SS
	<i>Burkholderia</i> kururiensis	516382324	-----L-H-----KVSS
	<i>Burkholderia</i> rhizoxinica HKI 4	312797609	-----L-R-NV-----VSS
	Cupriavidus sp. WS	519050400	---E-L-----KI-----
	Cupriavidus basilensis	493151034	---E-L-----KIV-----V
	Variovorax paradoxus S110	239815615	---IE-L-----KL-----V
	Pandoraea sp. SD6-2	498504991	---I-L-R-I-----T
	Ramlibacter tataouinensis TT3	337279845	---VE-L-R-L-----P
	Comamonas testosteroni	489166115	-----L-R-KL-A-----K-A
	Variovorax sp. CF313	495112131	---IE-L-----KL-----VA
	Polaromonas sp. CF318	495147651	---VIE-L-----KL-V-----V
	Curvibacter lanceolatus	518403185	---IE-L-----L-----V-S
	Caldimonas manganoxidans	518390521	---E-L-R-KL-----V-P
Hydrogenophaga sp. PBC	497204968	---IE-L-R-KL-A-----I-S	
Acidovorax sp. NO-1	496179791	---VE-L-R-KL-A-----KV-S	
Delftia sp. Cs1-4	333914892	---E-L-R-KL-A-----K-A	

B

		380	449	
Clade IIb <i>Burkholderia</i>	<i>Burkholderia</i> sp. H160	496202984	GTGFLVNDVRAPFEVTLGGTDLSTLW L RDGPEAYLGTSIANFNFFMIVGPNLGLGH	
	<i>Burkholderia</i> sp. CCGE1002	295676766	---Q---P-----D-----	
	<i>Burkholderia</i> sp. Ch1-1	494327703	---Q---G---D---D-A---G-M---	
	<i>Burkholderia</i> phytotfirmans PsJN	187924282	---Q---G---D---D-A---Q---	
	<i>Burkholderia</i> sp. CCGE1001	323525825	---Q---G---D---V-A---GA---	
	<i>Burkholderia</i> xenovorans LB400	91783855	---Q---G---D---D-A---GA---	
	<i>Burkholderia</i> graminis	492936566	---Q---G---D---V-A---GA---	
	<i>Burkholderia</i> terrae	494862209	---Q---G---D---A---GA---	
	<i>Burkholderia</i> sp. BT03	495008604	---Q---G---D---A---GA---	
	<i>Burkholderia</i> phymatum STM815	186476032	---Q---G---D-A-I-A-GA---	
	<i>Burkholderia</i> sp. CCGE1003	307729928	---Q---GP---D---I-A-G---	
	<i>Burkholderia</i> sp. WSM2232	548691109	---GPA---I-A-G---	
	<i>Burkholderia</i> sp. WSM2230	548606735	---QF---GP---D---V-A---GA---	
	<i>Burkholderia</i> sp. WSM3556	548691174	---QF-G-G---IS-AN-RA---	
	<i>Burkholderia</i> sp. URHA0054	522812216	---Q---G---D---V-A---GA---	
	<i>Burkholderia</i> sprengiae WSM5005	548699263	---Q---P-----D-E---	
	<i>Burkholderia</i> mimosarum LMG 23256	548694927	---Q---D---Q---I-Q-G-M---	
	<i>Burkholderia</i> bryophila	518914327	---QF-GFG---I-A-RA---	
	<i>Burkholderia</i> phenoliruptrix BR3459a	407713185	---Q---G---D---V-A---GA---	
	<i>Burkholderia</i> sp. WSM4176	517228895	---Q---P-----D-----	
	<i>Burkholderia</i> sp. JPY251	517246749	---Q---P-----D-----	
	Other <i>Burkholderia</i>	<i>Burkholderia</i> kururiensis	516386272	---H---G---V-A---GA---
		<i>Burkholderia</i> thailandensis	497584847	---AATEFL-MHIV-RA-VT-NDA-
<i>Burkholderia</i> cenocepacia		493525653	---AATEFLS-MRI-R-L-NDA-	
<i>Burkholderia</i> vietnamiensis G4		134291654	---AATEFLS-MRI-RD-L-NDA-	
<i>Burkholderia</i> cepacia GG4		402569879	---AATSFLS-MRI-RN-L-NDA-	
<i>Burkholderia</i> oklahomensis		497808405	---AATEFL-MRIV-RA-VT-NDA-	
<i>Burkholderia</i> ambifaria MC40-6		172062537	---AATAFLS-MRI-RD-R-NDA-	
<i>Burkholderia</i> gladioli BSR3		330821758	---AATEFLS-MRI-R-LE-NEV-	
<i>Burkholderia</i> multivorans		493454498	---Q-A-PFPRGAI-V-R-L-IVDA-	
Cupriavidus basilensis		493139021	---QAT-FL-MR---D-I-NQA-	
Other Bacteria	Acidovorax avenae subsp. avena	326316554	---AATEFLS-MKI-R-LE-NAA-	
	Acinetobacter sp. NIPH 1867	491354526	---AATEFM-MOI---N-KR-KDQ-	
	Pseudomonas resinovorans NBRC	512617894	---AAT-FL-MSIR---RE-NDA-	
	Hahella chejuensis KCTC 2396	83647045	A---QAAEAVI---DLI---VS-NQA-	

FIGURE 5 | Partial sequence alignments of (A) 3-phosphoglycerate dehydrogenase showing a 1 amino acid insertion (boxed) identified in all members of Clade IIa of the genus *Burkholderia* (B) 4-hydroxyacetophenone monooxygenase showing a 1 amino acid insertion (boxed) identified only in members of Clade IIb of the genus

Burkholderia. These CSIs were not found in the sequence homologs of these proteins from any other sequenced bacteria in the top 250 BLAST hits. Sequence information for other CSIs specific to subclades within Clade II of the genus *Burkholderia* are presented in Supplemental Figures 16–35 and their characteristics are summarized in Table 3.

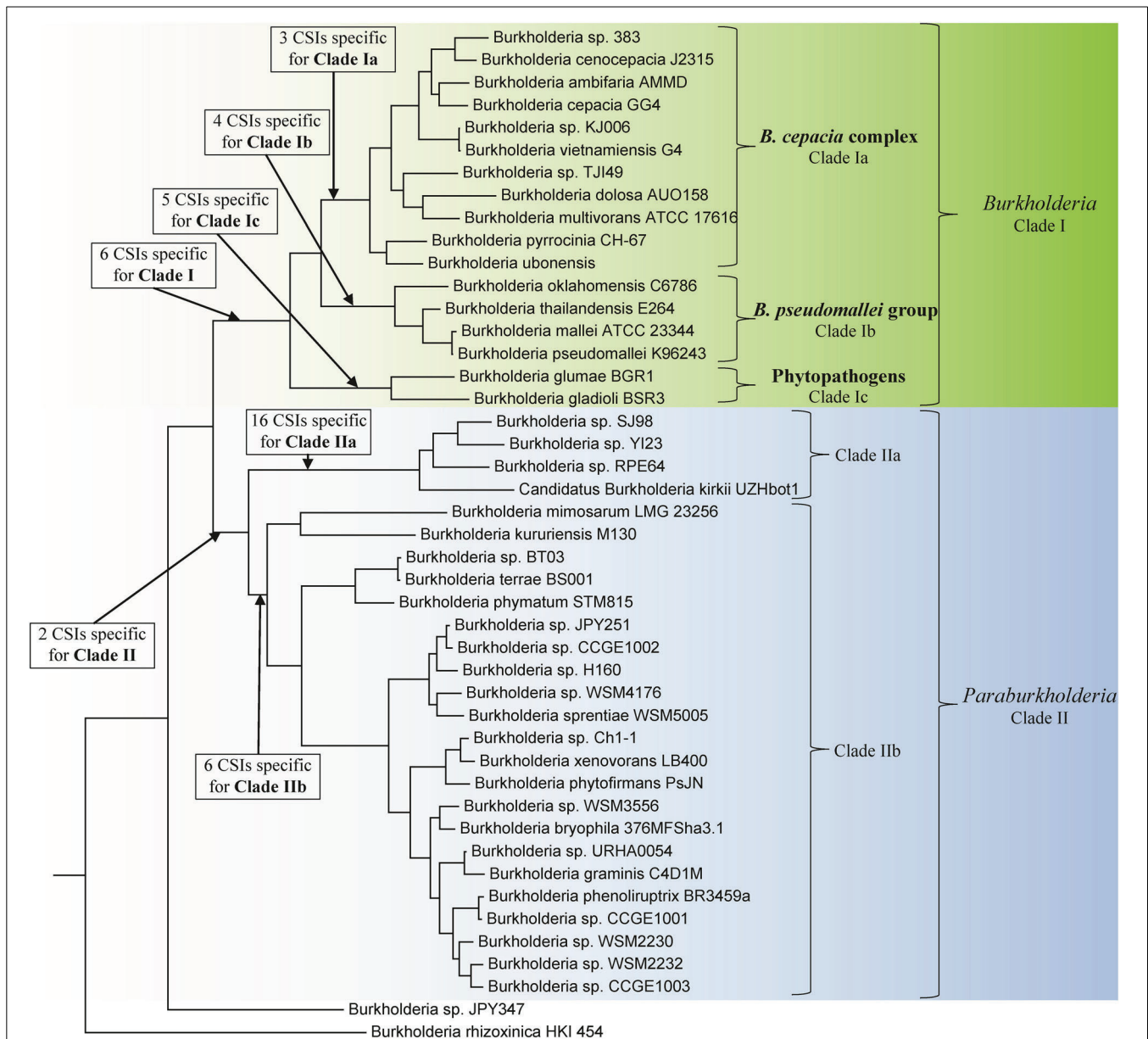


FIGURE 6 | A summary diagram depicting the distribution of identified CSIs and the proposed names of the two major groups (Clade I and II) within *Burkholderia*. The major *Burkholderia* clades are indicated by brackets and highlighting.

provide novel and useful targets for the development of diagnostic assays for either the BCC or the *B. pseudomallei* group (Ahmad et al., 2011; Wong et al., 2014). We have identified CSIs for three other groups within the genus *Burkholderia*: A group of plant pathogenic *Burkholderia* related to the BCC and *B. pseudomallei* group (Clade Ic), a group containing unnamed and candidate *Burkholderia* species (Clade IIa), and a group consisting of environmental *Burkholderia* (Clade IIb). We have identified 6, 16, and 6 CSIs for these three groups, respectively. These CSIs provide important differentiating characteristics for these groups, particularly for Clades IIa and IIb which are related groups that have no other identified differentiating characteristics (Suarez-Moreno et al., 2012).

The phylogenetic analyses, identified CSIs, and the pathogenic characteristics of the different *Burkholderia* species presented in this work strongly suggest that the genus *Burkholderia* is made up of at least two distinct lineages. One lineage consisting of the BCC and related organisms (Clade I) and another consisting of a wide range of environmental organisms (Clade II). This latter clade is phylogenetically highly diverse and there is a paucity of sequence information available for its members. Thus, it is possible that in future this latter clade may be found to consist of more than one distinct bacterial lineage, however, it is currently clear that Clade I and Clade II represent distinct lineages. Evidence for the distinctness of the Clade I members from other *Burkholderia* species has been identified in

Table 4 | Descriptions of the new combinations in the genus *Paraburkholderia* gen. nov.

New Combination	Basonym	Type Strain	References
<i>Paraburkholderia acidipaludis</i> comb. nov.	<i>Burkholderia acidipaludis</i>	SA33 NBRC 101816 VTCC-D6-6	Aizawa et al., 2010b
<i>Candidatus Paraburkholderia andongensis</i> comb. nov.	<i>Candidatus Burkholderia andongensis</i>	—	Lemaire et al., 2011
<i>Paraburkholderia andropogonis</i> comb. nov.	<i>Burkholderia andropogonis</i>	ATCC 23061 CCUG 32772 CFBP 2421 CIP 105771 DSM 9511 ICMP 2807 JCM 10487 LMG 2129 NCPFB 934 NRRL B-14296	Gillis et al., 1995
<i>Paraburkholderia aspalathi</i> comb. nov.	<i>Burkholderia aspalathi</i>	VG1C DSM 27239 LMG 27731	Mavengere et al., 2014
<i>Paraburkholderia bannensis</i> comb. nov.	<i>Burkholderia bannensis</i>	E25 BCC 36998 NBRC 103871	Aizawa et al., 2011
<i>Paraburkholderia bryophila</i> comb. nov.	<i>Burkholderia bryophila</i>	1S18 CCUG 52993 LMG 23644	Vandamme et al., 2007
<i>Paraburkholderia caballeronis</i> comb. nov.	<i>Burkholderia caballeronis</i>	TNe-841 CIP 110324 LMG 26416	Martínez-Aguilar et al., 2013
<i>Paraburkholderia caledonica</i> comb. nov.	<i>Burkholderia caledonica</i>	W50D CCUG 42236 CIP 107098 JCM 21561 LMG 19076 NBRC 102488	Coenye et al., 2001a
<i>Candidatus Paraburkholderia calva</i> comb. nov.	<i>Candidatus Burkholderia calva</i>	—	Van Oevelen et al., 2004
<i>Paraburkholderia caribensis</i> comb. nov.	<i>Burkholderia caribensis</i>	MWAP64 CCUG 42847 CIP 106784 DSM 13236 LMG 18531	Achouak et al., 1999
<i>Paraburkholderia caryophylli</i> comb. nov.	<i>Burkholderia caryophylli</i>	ATCC 25418 CCUG 20834 CFBP 2429 CFBP 3818 CIP 105770 DSM 50341 HAMB1 2159 ICMP 512	Yabuuchi et al., 1992

(Continued)

Table 4 | Continued

New Combination	Basonym	Type Strain	References
		JCM 9310 JCM 10488 LMG 2155 NCPFB 2151	
<i>Paraburkholderia choica</i> comb. nov.	<i>Burkholderia choica</i>	LMG 22940 CCUG 63063	Vandamme et al., 2013
<i>Paraburkholderia denitrificans</i> comb. nov.	<i>Burkholderia denitrificans</i>	KIS30-44 DSM 24336 KACC 12733	Lee et al., 2012
<i>Paraburkholderia diazotrophica</i> comb. nov.	<i>Burkholderia diazotrophica</i>	JPY461 NKMU-JPY461 BCRC 80259 KCTC 23308 LMG 26031	Sheu et al., 2013
<i>Paraburkholderia dilworthii</i> comb. nov.	<i>Burkholderia dilworthii</i>	WSM3556 LMG 27173 HAMBI 3353	De Meyer et al., 2014
<i>Paraburkholderia eburne</i> comb. nov.	<i>Burkholderia eburne</i>	RR11 KEMC 7302-065 JCM 18070	Kang et al., 2014
<i>Paraburkholderia endofungorum</i> comb. nov.	<i>Burkholderia endofungorum</i>	HKI 456 CIP 109454 DSM 19003	Partida-Martinez et al., 2007
<i>Paraburkholderia ferrariae</i> comb. nov.	<i>Burkholderia ferrariae</i>	FeGI01 CECT 7171 DSM 18251 LMG 23612	Valverde et al., 2006
<i>Paraburkholderia fungorum</i> comb. nov.	<i>Burkholderia fungorum</i>	Croize P763-2 CCUG 31961 CIP 107096 JCM 21562 LMG 16225 NBRC 102489	Coenye et al., 2001a
<i>Paraburkholderia ginsengisoli</i> comb. nov.	<i>Burkholderia ginsengisoli</i>	KMY03 KCTC 12389 NBRC 100965	Kim et al., 2006
<i>Paraburkholderia glathei</i> comb. nov.	<i>Burkholderia glathei</i>	ATCC 29195 CFBP 4791 CIP 105421 DSM 50014 JCM 10563 LMG 14190	Vandamme et al., 1997
<i>Paraburkholderia graminis</i> comb. nov.	<i>Burkholderia graminis</i>	C4D1M ATCC 700544 CCUG 42231 CIP 106649 LMG 18924	Viallard et al., 1998

(Continued)

Table 4 | Continued

New Combination	Basonym	Type Strain	References
<i>Paraburkholderia grimmiae</i> comb. nov.	<i>Burkholderia grimmiae</i>	R27 CGMCC 1.11013 DSM 25160	Tian et al., 2013
<i>Paraburkholderia heleia</i> comb. nov.	<i>Burkholderia heleia</i>	SA41 NBRC 101817 VTCC-D6-7	Aizawa et al., 2010a
<i>Candidatus Paraburkholderia hispidae</i> comb. nov.	<i>Candidatus Burkholderia hispidae</i>	—	Lemaire et al., 2012
<i>Paraburkholderia hospita</i> comb. nov.	<i>Burkholderia hospita</i>	LMG 20598 CCUG 43658	Goris et al., 2002
<i>Paraburkholderia humi</i> comb. nov.	<i>Burkholderia humi</i>	LMG 22934 CCUG 63059	Vandamme et al., 2013
<i>Candidatus Paraburkholderia kirkii</i> comb. nov.	<i>Candidatus Burkholderia kirkii</i>	—	Van Oevelen et al., 2002a
<i>Paraburkholderia kururiensis</i> comb. nov.	<i>Burkholderia kururiensis</i>	KP23 ATCC 700977 CCUG 43663 CIP 106643 DSM 13646 JCM 10599 LMG 19447	Zhang et al., 2000
<i>Paraburkholderia megapolitana</i> comb. nov.	<i>Burkholderia megapolitana</i>	A3 CCUG 53006 LMG 23650	Vandamme et al., 2007
<i>Paraburkholderia mimosarum</i> comb. nov.	<i>Burkholderia mimosarum</i>	PAS44 BCRC 17516 LMG 23256	Chen et al., 2006
<i>Candidatus Paraburkholderia nigropunctata</i> comb. nov.	<i>Candidatus Burkholderia nigropunctata</i>	—	Van Oevelen et al., 2004
<i>Paraburkholderia nodosa</i> comb. nov.	<i>Burkholderia nodosa</i>	Br3437 BCRC 17575 LMG 23741	Chen et al., 2007
<i>Paraburkholderia oxyphila</i> comb. nov.	<i>Burkholderia oxyphila</i>	OX-01 DSM 22550 NBRC 105797	Otsuka et al., 2011
<i>Candidatus Paraurkholderia petiti</i> comb. nov.	<i>Candidatus Burkholderia petiti</i>	—	Lemaire et al., 2011
<i>Paraburkholderia phenazinium</i> comb. nov.	<i>Burkholderia phenazinium</i>	ATCC 33666 CCUG 20836 CFBP 4793 CIP 106502 DSM 10684 JCM 10564 LMG 2247 NCIMB 11027	Viallard et al., 1998

(Continued)

Table 4 | Continued

New Combination	Basonym	Type Strain	References
<i>Paraburkholderia phenoliruptrix</i> comb. nov.	<i>Burkholderia phenoliruptrix</i>	AC1100 CCUG 48558 LMG 22037	Coenye et al., 2004
<i>Paraburkholderia phymatum</i> comb. nov.	<i>Burkholderia phymatum</i>	STM815 LMG 21445 CCUG 47179	Vandamme et al., 2002a
<i>Paraburkholderia phytofirmans</i> comb. nov.	<i>Burkholderia phytofirmans</i>	PsJN CCUG 49060 LMG 22146	Sessitsch et al., 2005
<i>Paraburkholderia rhizoxinica</i> comb. nov.	<i>Burkholderia rhizoxinica</i>	HKI 454 CIP 109453 DSM 19002	Partida-Martinez et al., 2007
<i>Paraburkholderia rhynchosiae</i> comb. nov.	<i>Burkholderia rhynchosiae</i>	WSM3937 LMG 27174 HAMBI 3354	De Meyer et al., 2013b
<i>Candidatus Paraburkholderia rigidae</i> comb. nov.	<i>Candidatus Burkholderia rigidae</i>	—	Lemaire et al., 2012
<i>Paraburkholderia sabiae</i> comb. nov.	<i>Burkholderia sabiae</i>	Br3407 BCRC 17587 LMG 24235	Chen et al., 2008
<i>Paraburkholderia sacchari</i> comb. nov.	<i>Burkholderia sacchari</i>	CCT 6771 CCUG 46043 CIP 107211 IPT 101 LMG 19450	Brämer et al., 2001
<i>Paraburkholderia sartisoli</i> comb. nov.	<i>Burkholderia sartisoli</i>	RP007 CCUG 53604 ICMP 13529 LMG 24000	Vanlaere et al., 2008
<i>Candidatus Paraburkholderia schumanniana</i> comb. nov.	<i>Candidatus Burkholderia schumanniana</i>	—	Lemaire et al., 2012
<i>Paraburkholderia sedimicola</i> comb. nov.	<i>Burkholderia sedimicola</i>	HU2-65W KCTC 22086 LMG 24238	Lim et al., 2008
<i>Paraburkholderia silvatlantica</i> comb. nov.	<i>Burkholderia silvatlantica</i>	SRMrh-20 ATCC BAA-1244 LMG 23149	Perin et al., 2006
<i>Paraburkholderia soli</i> comb. nov.	<i>Burkholderia soli</i>	GP25-8 DSM 18235 KACC 11589	Yoo et al., 2007
<i>Paraburkholderia sordidicola</i> comb. nov.	<i>Burkholderia sordidicola</i>	CCUG 49583 JCM 11778 KCTC 12081	Lim et al., 2003
<i>Paraburkholderia spreintiae</i> comb. nov.	<i>Burkholderia spreintiae</i>	WSM5005 LMG 27175 HAMBI 3357	De Meyer et al., 2013a

(Continued)

Table 4 | Continued

New Combination	Basonym	Type Strain	References
<i>Paraburkholderia symbiotica</i> comb. nov.	<i>Burkholderia symbiotica</i>	JPY-345 NKMU-JPY-345 BCRC 80258 KCTC 23309 LMG 26032	Sheu et al., 2012
<i>Paraburkholderia telluris</i> comb. nov.	<i>Burkholderia telluris</i>	LMG 22936 CCUG 63060	Vandamme et al., 2013
<i>Paraburkholderia terrae</i> comb. nov.	<i>Burkholderia terrae</i>	KMY02 KCTC 12388 NBRC 100964	Yang et al., 2006
<i>Paraburkholderia terrestris</i> comb. nov.	<i>Burkholderia terrestris</i>	LMG 22937 CCUG 63062	Vandamme et al., 2013
<i>Paraburkholderia terricola</i> comb. nov.	<i>Burkholderia terricola</i>	CCUG 44527 LMG 20594	Goris et al., 2002
<i>Paraburkholderia tropica</i> comb. nov.	<i>Burkholderia tropica</i>	Ppe8 ATCC BAA-831 DSM 15359 LMG 22274	Reis et al., 2004
<i>Paraburkholderia tuberum</i> comb. nov.	<i>Burkholderia tuberum</i>	STM678 CCUG 47178 LMG 21444	Vandamme et al., 2002a
<i>Paraburkholderia udeis</i> comb. nov.	<i>Burkholderia udeis</i>	LMG 27134 CCUG 63061	Vandamme et al., 2013
<i>Paraburkholderia unamae</i> comb. nov.	<i>Burkholderia unamae</i>	MTI-641 ATCC BAA-744 CIP 107921	Caballero-Mellado et al., 2004
<i>Paraburkholderia xenovorans</i> comb. nov.	<i>Burkholderia xenovorans</i>	LB400 CCUG 46959 LMG 21463 NRRL B-18064	Goris et al., 2004
<i>Paraburkholderia zhejiangensis</i> comb. nov.	<i>Burkholderia zhejiangensis</i>	OP-1 KCTC 23300	Lu et al., 2012

a number of previous phylogenetic studies (Payne et al., 2005; Tayeb et al., 2008; Yarza et al., 2008; Spilker et al., 2009; Ussery et al., 2009; Gyaneshwar et al., 2011; Vandamme and Dawyndt, 2011; Zhu et al., 2011; Suarez-Moreno et al., 2012; Estrada-de los Santos et al., 2013; Segata et al., 2013). Estrada-de los Santos et al. (2013) recently completed a phylogenetic analysis of the genus *Burkholderia* utilizing the multilocus sequence analysis of *atpD*, *gltB*, *lepA*, and *recA* genes in combination with the 16S rRNA gene, which provides compelling evidence for the presence of two distinct evolutionary lineages within the genus *Burkholderia*. However, these authors have refrained from formally proposing a division of the genus into two genera due to a paucity of differentiating characteristics for the two groups. Our comparative analysis of *Burkholderia* genomes has identified a set of distinctive molecular characteristics that

clearly differentiate the two evolutionary lineages within the genus *Burkholderia* in addition the phylogenetic evidence. In light of the abundance of phylogenetic and molecular evidence for the presence of two distinct evolutionary lineages within the genus *Burkholderia*, and the distinct pathogenicity profiles of the members of these two groups, we are proposing that genus *Burkholderia* should be divided into two separate genera. The first of these monophyletic genera, which comprises of all the clinically relevant species and clearly distinguished from all other *Burkholderia* species, will retain the name *Burkholderia* (Clade I). For the remainder of the *Burkholderia* species (Clade II), which include a wide range of environmental species, we propose the name *Paraburkholderia* gen. nov. An emended description of the genus *Burkholderia* and a description of *Paraburkholderia* gen. nov. are provided below. Brief descriptions of the new species

combinations within *Paraburkholderia* gen. nov. are presented in Table 4.

EMENDED DESCRIPTION OF THE GENUS *BURKHOLDERIA* (Yabuuchi et al., 1993 EMEND. Gillis et al., 1995)

The genus contains the type species *B. cepacia* (Yabuuchi et al., 1993). The species from this genus are gram-negative, straight or slightly curved rods, which exhibit motility mediated by one or more polar flagella. Only, *B. mallei* lacks flagella and is non-motile. The species do not produce sheaths or prosthecae and do not go through any resting stages. Most species are able to accumulate and utilize poly- β -hydroxybutyrate (PHB) for growth. The species are mostly aerobic chemoorganotrophs, but some species are capable of anaerobic respiration using nitrate as the terminal electron acceptor. The G+C content for the members of the genus ranges from 65.7 to 68.5%. The members of the genus form a distinct monophyletic clade in phylogenetic trees, and they are distinguished from all other bacteria by the conserved sequence indels reported in this work in the following proteins: Periplasmic amino acid-binding protein, 4-hydroxybenzoate 3-monooxygenase, 6-phosphogluconate dehydrogenase, Sarcosine oxidase subunit alpha, a putative lipoprotein, and a putative lyase (Table 2).

DESCRIPTION OF THE GENUS *PARABURKHOLDERIA* GEN. NOV.

The genus contains the type species *Paraburkholderia graminis* comb. nov. (Basonym: *Burkholderia graminis*, Viillard et al., 1998) The species from this genus are gram-negative straight or slightly curved rods with one or more polar flagella. Other morphological and metabolic characteristics are similar to genus *Burkholderia*. The G+C content for the members of the genus ranges from 61.4 to 65.0%. The species are not associated with humans. The members of this genus generally form a distinct clade in the neighborhood of genus *Burkholderia* in phylogenetic trees, and they lack the molecular signatures which are specific for *Burkholderia*. Most of the sequenced members from this genus contain the conserved sequence indels reported in this work in the protein sequences of an unnamed dehydrogenase and a LysR family transcriptional regulator (Table 2).

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <http://www.frontiersin.org/journal/10.3389/fgene.2014.00429/abstract>

REFERENCES

- Achouak, W., Christen, R., Barakat, M., Martel, M.-H., and Heulin, T. (1999). *Burkholderia caribensis* sp. nov., an exopolysaccharide-producing bacterium isolated from vertisol microaggregates in Martinique. *Int. J. Syst. Bacteriol.* 49, 787–794. doi: 10.1099/00207713-49-2-787
- Ahmod, N. Z., Gupta, R. S., and Shah, H. N. (2011). Identification of a *Bacillus anthracis* specific indel in the *yeaC* gene and development of a rapid pyrosequencing assay for distinguishing *B. anthracis* from the *B. cereus* group. *J. Microbiol. Methods* 87, 278–285. doi: 10.1016/j.mimet.2011.08.015
- Aizawa, T., Ve, N. B., Nakajima, M., and Sunairi, M. (2010a). *Burkholderia heleaia* sp. nov., a nitrogen-fixing bacterium isolated from an aquatic plant, *Eleocharis dulcis*, that grows in highly acidic swamps in actual acid sulfate soil areas of Vietnam. *Int. J. Syst. Evol. Microbiol.* 60, 1152–1157. doi: 10.1099/ijs.0.1015198-0
- Aizawa, T., Ve, N. B., Vijarnsorn, P., Nakajima, M., and Sunairi, M. (2010b). *Burkholderia acidipaludis* sp. nov., aluminium-tolerant bacteria isolated from Chinese water chestnut (*Eleocharis dulcis*) growing in highly acidic swamps in South-East Asia. *Int. J. Syst. Evol. Microbiol.* 60, 2036–2041. doi: 10.1099/ijs.0.018283-0
- Aizawa, T., Vijarnsorn, P., Nakajima, M., and Sunairi, M. (2011). *Burkholderia bannensis* sp. nov., an acid-neutralizing bacterium isolated from torped grass (*Panicum repens*) growing in highly acidic swamps. *Int. J. Syst. Evol. Microbiol.* 61, 1645–1650. doi: 10.1099/ijs.0.026278-0
- Biddick, R., Spilker, T., Martin, A., and LiPuma, J. J. (2003). Evidence of transmission of *Burkholderia cepacia*, *Burkholderia multivorans* and *Burkholderia dolosa* among persons with cystic fibrosis. *FEMS Microbiol. Lett.* 228, 57–62. doi: 10.1016/S0378-1097(03)00724-9
- Brämer, C. O., Vandamme, P., da Silva, L. F., Gomez, J., and Steinbüchel, A. (2001). Polyhydroxyalkanoate-accumulating bacterium isolated from soil of a sugar-cane plantation in Brazil. *Int. J. Syst. Evol. Microbiol.* 51, 1709–1713. doi: 10.1099/00207713-51-5-1709
- Caballero-Mellado, J., Martínez-Aguilar, L., Paredes-Valdez, G., and Estrada-de los Santos, P. (2004). *Burkholderia unamae* sp. nov., an N₂-fixing rhizospheric and endophytic species. *Int. J. Syst. Evol. Microbiol.* 54, 1165–1172. doi: 10.1099/ijs.0.02951-0
- Castresana, J. (2000). Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. *Mol. Biol. Evol.* 17, 540–552. doi: 10.1093/oxfordjournals.molbev.a026334
- Chain, P. S., Deneff, V. J., Konstantinidis, K. T., Vergez, L. M., Agullo, L., Reyes, V. L., et al. (2006). *Burkholderia xenovorans* LB400 harbors a multi-replicon, 9.73-Mbp genome shaped for versatility. *Proc. Natl. Acad. Sci. U.S.A.* 103, 15280–15287. doi: 10.1073/pnas.0606924103
- Charlebois, R. L., and Doolittle, W. F. (2004). Computing prokaryotic gene ubiquity: rescuing the core from extinction. *Genome Res.* 14, 2469–2477. doi: 10.1101/gr.3024704
- Chen, W.-M., de Faria, S. M., Chou, J.-H., James, E. K., Elliott, G. N., Sprent, J. I., et al. (2008). *Burkholderia sabiae* sp. nov., isolated from root nodules of *Mimosa caesalpiniiifolia*. *Int. J. Syst. Evol. Microbiol.* 58, 2174–2179. doi: 10.1099/ijs.0.65816-0
- Chen, W.-M., De Faria, S. M., James, E. K., Elliott, G. N., Lin, K.-Y., Chou, J.-H., et al. (2007). *Burkholderia nodosa* sp. nov., isolated from root nodules of the woody Brazilian legumes *Mimosa bimucronata* and *Mimosa scabrella*. *Int. J. Syst. Evol. Microbiol.* 57, 1055–1059. doi: 10.1099/ijs.0.64873-0
- Chen, W.-M., James, E. K., Coenye, T., Chou, J.-H., Barrios, E., De Faria, S. M., et al. (2006). *Burkholderia mimosarum* sp. nov., isolated from root nodules of *Mimosa* spp. from Taiwan and South America. *Int. J. Syst. Evol. Microbiol.* 56, 1847–1851. doi: 10.1099/ijs.0.64325-0
- Ciccarelli, F. D., Doerks, T., Von Mering, C., Creevey, C. J., Snel, B., and Bork, P. (2006). Toward automatic reconstruction of a highly resolved tree of life. *Science* 311, 1283–1287. doi: 10.1126/science.1123061
- Coenye, T., Henry, D., Speert, D. P., and Vandamme, P. (2004). *Burkholderia phenoliruptrix* sp. nov., to accommodate the 2, 4, 5-trichlorophenoxyacetic acid and halophenol-degrading strain AC1100. *Syst. Appl. Microbiol.* 27, 623–627. doi: 10.1078/0723202042369992
- Coenye, T., Laevens, S., Willems, A., Ohlén, M., Hannant, W., Govan, J., et al. (2001a). *Burkholderia fungorum* sp. nov. and *Burkholderia caledonica* sp. nov., two new species isolated from the environment, animals and human clinical samples. *Int. J. Syst. Evol. Microbiol.* 51, 1099–1107. doi: 10.1099/00207713-51-3-1099
- Coenye, T., Mahenthalingam, E., Henry, D., LiPuma, J. J., Laevens, S., Gillis, M., et al. (2001b). *Burkholderia ambifaria* sp. nov., a novel member of the *Burkholderia cepacia* complex including biocontrol and cystic fibrosis-related isolates. *Int. J. Syst. Evol. Microbiol.* 51, 1481–1490. doi: 10.1099/00207713-51-4-1481
- Coenye, T., and Vandamme, P. (2003). Diversity and significance of *Burkholderia* species occupying diverse ecological niches. *Environ. Microbiol.* 5, 719–729. doi: 10.1046/j.1462-2920.2003.00471.x
- Cole, J. R., Wang, Q., Cardenas, E., Fish, J., Chai, B., Farris, R. J., et al. (2009). The ribosomal database project: improved alignments and new tools for rRNA analysis. *Nucleic Acids Res.* 37, D141–D145. doi: 10.1093/nar/gkn879
- Compant, S., Nowak, J., Coenye, T., Clement, C., and Ait Barka, E. (2008). Diversity and occurrence of *Burkholderia* spp. in the natural environment. *FEMS Microbiol. Rev.* 32, 607–626. doi: 10.1111/j.1574-6976.2008.00113.x

- Coutinho, B. G., Passos da Silva, D., Previato, J. O., Mendonca-Previato, L., and Venturi, V. (2013). Draft genome sequence of the rice endophyte *Burkholderia kururiensis* M130. *Genome Announc.* 1, e0022512–e0022512. doi: 10.1128/genomeA.00225-12
- De Meyer, S. E., Cnockaert, M., Ardley, J. K., Maker, G., Yates, R., Howieson, J. G., et al. (2013a). *Burkholderia sprentiae* sp. nov., isolated from *Lebeckia ambigua* root nodules. *Int. J. Syst. Evol. Microbiol.* 63(Pt 11), 3950–3957. doi: 10.1099/ijs.0.048777-0
- De Meyer, S. E., Cnockaert, M., Ardley, J. K., Trengove, R. D., Garau, G., Howieson, J. G., et al. (2013b). *Burkholderia rhynchosiae* sp. nov., isolated from *Rhynchosia ferulifolia* root nodules. *Int. J. Syst. Evol. Microbiol.* 63(Pt 11), 3944–3949. doi: 10.1099/ijs.0.048751-0
- De Meyer, S. E., Cnockaert, M., Ardley, J. K., Van Wyk, B.-E., Vandamme, P. A., and Howieson, J. G. (2014). *Burkholderia dilworthii* sp. nov., isolated from *Lebeckia ambigua* root nodules. *Int. J. Syst. Evol. Microbiol.* 64(Pt 4), 1090–1095. doi: 10.1099/ijs.0.058602-0
- Deshazer, D. (2007). Virulence of clinical and environmental isolates of *Burkholderia oklahomensis* and *Burkholderia thailandensis* in hamsters and mice. *FEMS Microbiol. Lett.* 277, 64–69. doi: 10.1111/j.1574-6968.2007.00946.x
- Estrada-de los Santos, P., Vinuesa, P., Martínez-Aguilar, L., Hirsch, A. M., and Caballero-Mellado, J. (2013). Phylogenetic analysis of *Burkholderia* species by multilocus sequence analysis. *Curr. Microbiol.* 67, 51–60. doi: 10.1007/s00284-013-0330-9
- Gao, B., and Gupta, R. S. (2012). Microbial systematics in the post-genomics era. *Antonie Van Leeuwenhoek* 101, 45–54. doi: 10.1007/s10482-011-9663-1
- Gillis, M., Van Van, T., Bardin, R., Goor, M., Hebbbar, P., Willems, A., et al. (1995). Polyphasic taxonomy in the genus *Burkholderia* leading to an emended description of the genus and proposition of *Burkholderia vietnamiensis* sp. nov. for N₂-fixing isolates from rice in Vietnam. *Int. J. Syst. Bacteriol.* 45, 274–289. doi: 10.1099/00207713-45-2-274
- Gogarten, J. P., Doolittle, W. F., and Lawrence, J. G. (2002). Prokaryotic evolution in light of gene transfer. *Mol. Biol. Evol.* 19, 2226–2238. doi: 10.1093/oxfordjournals.molbev.a004046
- Goris, J., Dejonghe, W., Falsen, E., De Clerck, E., Geeraerts, B., Willems, A., et al. (2002). Diversity of transconjugants that acquired plasmid pJP4 or pEMT1 after inoculation of a donor strain in the A- and B-horizon of an agricultural soil and description of *Burkholderia hospita* sp. nov. and *Burkholderia terricola* sp. nov. *Syst. Appl. Microbiol.* 25, 340–352. doi: 10.1078/0723-2020-00134
- Goris, J., De Vos, P., Caballero-Mellado, J., Park, J., Falsen, E., Quensen, J. F., et al. (2004). Classification of the biphenyl- and polychlorinated biphenyl-degrading strain LB400T and relatives as *Burkholderia xenovorans* sp. nov. *Int. J. Syst. Evol. Microbiol.* 54, 1677–1681. doi: 10.1099/ijs.0.63101-0
- Gupta, R. S. (1998). Protein phylogenies and signature sequences: a reappraisal of evolutionary relationships among archaeobacteria, eubacteria, and eukaryotes. *Microbiol. Mol. Biol. Rev.* 62, 1435.
- Gupta, R. S. (2001). The branching order and phylogenetic placement of species from completed bacterial genomes, based on conserved indels found in various proteins. *Int. Microbiol.* 4, 187–202. doi: 10.1007/s10123-001-0037-9
- Gupta, R. S. (2009). Protein signatures (molecular synapomorphies) that are distinctive characteristics of the major cyanobacterial clades. *Int. J. Syst. Evol. Microbiol.* 59, 2510. doi: 10.1099/ijs.0.005678-0
- Gupta, R. S. (2014). *Identification of Conserved Indels that are Useful for Classification and Evolutionary Studies Methods in Microbiology*, Vol. 41. Oxford: Academic Press.
- Gupta, R. S., and Griffiths, E. (2002). Critical issues in bacterial phylogeny. *Theor. Popul. Biol.* 61, 423–434. doi: 10.1006/tpbi.2002.1589
- Gyaneshwar, P., Hirsch, A. M., Moulin, L., Chen, W.-M., Elliott, G. N., Bontemps, C., et al. (2011). Legume-nodulating *betaproteobacteria*: diversity, host range, and future prospects. *Mol. Plant Microbe Interact.* 24, 1276–1288. doi: 10.1094/MPMI-06-11-0172
- Harris, J. K., Kelley, S. T., Spiegelman, G. B., and Pace, N. R. (2003). The genetic core of the universal ancestor. *Genome Res.* 13, 407–412. doi: 10.1101/gr.652803
- Hauser, A. R., Jain, M., Bar-Meir, M., and McColley, S. A. (2011). Clinical significance of microbial infection and adaptation in cystic fibrosis. *Clin. Microbiol. Rev.* 24, 29–70. doi: 10.1128/CMR.00036-10
- Holden, M. T., Seth-Smith, H. M., Crossman, L. C., Sebaihia, M., Bentley, S. D., Cerdano-Tarraga, A. M., et al. (2009). The genome of *Burkholderia cenocepacia* J2315, an epidemic pathogen of cystic fibrosis patients. *J. Bacteriol.* 191, 261–277. doi: 10.1128/JB.01230-08
- Holden, M. T., Titball, R. W., Peacock, S. J., Cerdano-Tarraga, A. M., Atkins, T., Crossman, L. C., et al. (2004). Genomic plasticity of the causative agent of melioidosis, *Burkholderia pseudomallei*. *Proc. Natl. Acad. Sci. U.S.A.* 101, 14240–14245. doi: 10.1073/pnas.0403302101
- Hong, K. W., Koh, C. L., Sam, C. K., Yin, W. F., and Chan, K. G. (2012). Complete genome sequence of *Burkholderia* sp. Strain GG4, a betaproteobacterium that reduces 3-oxo-N-acylhomoserine lactones and produces different N-acylhomoserine lactones. *J. Bacteriol.* 194, 6317–6312. doi: 10.1128/JB.01578-12
- Jeanmougin, F., Thompson, J. D., Gouy, M., Higgins, D. G., and Gibson, T. J. (1998). Multiple sequence alignment with Clustal X. *Trends Biochem. Sci.* 23, 403. doi: 10.1016/S0968-0004(98)01285-7
- Jones, D. T., Taylor, W. R., and Thornton, J. M. (1992). The rapid generation of mutation data matrices from protein sequences. *Comput. Appl. Biosci.* CABIOS, 8, 275–282.
- Kang, S. R., Srinivasan, S., and Lee, S. S. (2014). *Burkholderia eburnea* sp. nov., isolated from peat soil. *Int. J. Syst. Evol. Microbiol.* 64(Pt 4), 1108–1115. doi: 10.1099/ijs.0.051078-0
- Katoh, K., and Standley, D. M. (2013). MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Mol. Biol. Evol.* 30, 772–780. doi: 10.1093/molbev/mst010
- Khan, A., Asif, H., Studholme, D. J., Khan, I. A., and Azim, M. K. (2013). Genome characterization of a novel *Burkholderia cepacia* complex genomovar isolated from dieback affected mango orchards. *World J. Microbiol. Biotechnol.* 29, 2033–2044. doi: 10.1007/s11274-013-1366-5
- Kim, H.-B., Park, M.-J., Yang, H.-C., An, D.-S., Jin, H.-Z., and Yang, D.-C. (2006). *Burkholderia ginsengisoli* sp. nov., a β -glucosidase-producing bacterium isolated from soil of a ginseng field. *Int. J. Syst. Evol. Microbiol.* 56, 2529–2533. doi: 10.1099/ijs.0.64387-0
- Kim, H. S., Schell, M. A., Yu, Y., Ulrich, R. L., Sarria, S. H., Nierman, W. C., et al. (2005). Bacterial genome adaptation to niches: divergence of the potential virulence genes in three *Burkholderia* species of different survival strategies. *BMC Genomics* 6:174. doi: 10.1186/1471-2164-6-174
- Kumar, S., Vikram, S., and Raghava, G. P. (2012). Genome sequence of the nitroaromatic compound-degrading bacterium *Burkholderia* sp. strain SJ98. *J. Bacteriol.* 194, 3286–3212. doi: 10.1128/JB.00497-12
- Kwak, M. J., Song, J. Y., Kim, S. Y., Jeong, H., Kang, S. G., Kim, B. K., et al. (2012). Complete genome sequence of the endophytic bacterium *Burkholderia* sp. strain KJ006. *J. Bacteriol.* 194, 4432–4433. doi: 10.1128/JB.00821-12
- Kyrpides, N., Overbeek, R., and Ouzounis, C. (1999). Universal protein families and the functional content of the last universal common ancestor. *J. Mol. Evol.* 49, 413–423. doi: 10.1007/PL00006564
- Lackner, G., Moebius, N., Partida-Martinez, L., and Hertweck, C. (2011). Complete genome sequence of *Burkholderia rhizoxinica*, an Endosymbiont of *Rhizopus microsporus*. *J. Bacteriol.* 193, 783–784. doi: 10.1128/JB.01318-10
- Lee, C.-M., Weon, H.-Y., Yoon, S.-H., Kim, S.-J., Koo, B.-S., and Kwon, S.-W. (2012). *Burkholderia denitrificans* sp. nov., isolated from the soil of Dokdo Island, Korea. *J. Microbiol.* 50, 855–859. doi: 10.1007/s12275-012-1554-2
- Lemaire, B., Robbrecht, E., van Wyk, B., Van Oevelen, S., Verstraete, B., Prinsen, E., et al. (2011). Identification, origin, and evolution of leaf nodulating symbionts of *Sericanthe (Rubiaceae)*. *J. Microbiol.* 49, 935–941. doi: 10.1007/s12275-011-1163-5
- Lemaire, B., Van Oevelen, S., De Block, P., Verstraete, B., Smets, E., Prinsen, E., et al. (2012). Identification of the bacterial endosymbionts in leaf nodules of *Pavetta (Rubiaceae)*. *Int. J. Syst. Evol. Microbiol.* 62, 202–209. doi: 10.1099/ijs.0.028019-0
- Lim, J. H., Baek, S.-H., and Lee, S.-T. (2008). *Burkholderia sediminicola* sp. nov., isolated from freshwater sediment. *Int. J. Syst. Evol. Microbiol.* 58, 565–569. doi: 10.1099/ijs.0.65502-0
- Lim, J., Lee, T. H., Nahm, B. H., Choi, Y. D., Kim, M., and Hwang, I. (2009). Complete genome sequence of *Burkholderia glumae* BGR1. *J. Bacteriol.* 191, 3758–3759. doi: 10.1128/JB.00349-09
- Lim, J. S., Choi, B. S., Choi, A. Y., Kim, K. D., Kim, D. I., Choi, I. Y., et al. (2012). Complete genome sequence of the fenitrothion-degrading *Burkholderia* sp. strain YI23. *J. Bacteriol.* 194, 896–811. doi: 10.1128/JB.06479-11
- Lim, Y. W., Baik, K. S., Han, S. K., Kim, S. B., and Bae, K. S. (2003). *Burkholderia sordidicola* sp. nov., isolated from the white-rot fungus *Phanerochaete sordida*. *Int. J. Syst. Evol. Microbiol.* 53, 1631–1636. doi: 10.1099/ijs.0.02456-0

- Limmathurotsakul, D., and Peacock, S. J. (2011). Melioidosis: a clinical overview. *Br. Med. Bull.* 99, 125–139. doi: 10.1093/bmb/ldr007
- Lu, P., Zheng, L.-Q., Sun, J.-J., Liu, H.-M., Li, S.-P., Hong, Q., et al. (2012). *Burkholderia zhejiangensis* sp. nov., a methyl-parathion-degrading bacterium isolated from a wastewater-treatment system. *Int. J. Syst. Evol. Microbiol.* 62(Pt 6), 1337–1341. doi: 10.1099/ijs.0.035428-0
- Mahenthiralingam, E., Baldwin, A., and Vandamme, P. (2002). *Burkholderia cepacia* complex infection in patients with cystic fibrosis. *J. Med. Microbiol.* 51, 533–538.
- Mahenthiralingam, E., Urban, T. A., and Goldberg, J. B. (2005). The multifarious, multireplicon *Burkholderia cepacia* complex. *Nat. Rev. Microbiol.* 3, 144–156. doi: 10.1038/nrmicro1085
- Mardis, E. R. (2008). The impact of next-generation sequencing technology on genetics. *Trends Genet.* 24, 133–141. doi: 10.1016/j.tig.2007.12.007
- Martínez-Aguilar, L., Salazar-Salazar, C., Méndez, R. D., Caballero-Mellado, J., Hirsch, A. M., Vásquez-Murrieta, M. S., et al. (2013). *Burkholderia caballeronis* sp. nov., a nitrogen fixing species isolated from tomato (*Lycopersicon esculentum*) with the ability to effectively nodulate *Phaseolus vulgaris*. *Antonie Van Leeuwenhoek* 104, 1063–1071. doi: 10.1007/s10482-013-0028-9
- Mavengere, N. R., Ellis, A. G., and Le Roux, J. J. (2014). *Burkholderia aspalathi* sp. nov., isolated from root nodules of the South African legume *Aspalathus abietina* Thunb. *Int. J. Syst. Evol. Microbiol.* 64, 1906–1912. doi: 10.1099/ijs.0.057067-0
- Nandakumar, R., Shahjahan, A., Yuan, X., Dickstein, E., Groth, D., Clark, C., et al. (2009). *Burkholderia glumae* and *B. gladioli* cause bacterial panicle blight in rice in the southern United States. *Plant Dis.* 93, 896–905. doi: 10.1094/PDIS-93-9-0896
- Nazir, R., Hansen, M. A., Sorensen, S., and van Elsland, J. D. (2012). Draft genome sequence of the soil bacterium *Burkholderia terrae* strain BS001, which interacts with fungal surface structures. *J. Bacteriol.* 194, 4480–4481. doi: 10.1128/JB.00725-12
- NCBI. (2014). *NCBI Genome Database*. <http://www.ncbi.nlm.nih.gov/genome/>
- Nierman, W. C., DeShazer, D., Kim, H. S., Tettelin, H., Nelson, K. E., Feldblyum, T., et al. (2004). Structural flexibility in the *Burkholderia mallei* genome. *Proc. Natl. Acad. Sci. U.S.A.* 101, 14246–14251. doi: 10.1073/pnas.0403306101
- O'Carroll, M. R., Kidd, T. J., Coulter, C., Smith, H. V., Rose, B. R., Harbour, C., et al. (2003). *Burkholderia pseudomallei*: another emerging pathogen in cystic fibrosis. *Thorax* 58, 1087–1091. doi: 10.1136/thorax.58.12.1087
- Oliveira Cunha, C., Goda Zuleta, L. F., Paula de Almeida, L. G., Prioli Ciapina, L., Lustrino Borges, W., Pitard, R. M., et al. (2012). Complete genome sequence of *Burkholderia phenoliruptrix* BR3459a (CLA1), a heat-tolerant, nitrogen-fixing symbiont of *Mimosa flocculosa*. *J. Bacteriol.* 194, 6675–6676. doi: 10.1128/JB.01821-12
- Onofre-Lemus, J., Hernández-Lucas, I., Girard, L., and Caballero-Mellado, J. (2009). ACC (1-aminocyclopropane-1-carboxylate) deaminase activity, a widespread trait in *Burkholderia* species, and its growth-promoting effect on tomato plants. *Appl. Environ. Microbiol.* 75, 6581–6590. doi: 10.1128/AEM.01240-09
- Ormeno-Orrillo, E., Rogel, M. A., Chueire, L. M., Tiedje, J. M., Martínez-Romero, E., and Hungria, M. (2012). Genome sequences of *Burkholderia* sp. strains CCGE1002 and H160, isolated from legume nodules in Mexico and Brazil. *J. Bacteriol.* 194, 6927–6912. doi: 10.1128/JB.01756-12
- Otsuka, Y., Muramatsu, Y., Nakagawa, Y., Matsuda, M., Nakamura, M., and Murata, H. (2011). *Burkholderia oxyphila* sp. nov., a bacterium isolated from acidic forest soil that catabolizes (+)-catechin and its putative aromatic derivatives. *Int. J. Syst. Evol. Microbiol.* 61, 249–254. doi: 10.1099/ijs.0.017368-0
- Palleroni, N. J. (2005). "Genus I. *Burkholderia* Yabuuchi et al. 1993, 398^{VP} (Effective publication: Yabuuchi et al. 1992, 1268) emend. Gillis et al. 1995, 286*," in *Bergey's Manual of Systematic Bacteriology*, 2^{Edn}, Vol. 2, eds D. J. Brenner, N. R. Krieg, G. M. Garrity, and J. T. Staley (New York, NY: Springer), 575–600.
- Parte, A. C. (2013). LPSN—list of prokaryotic names with standing in nomenclature. *Nucleic Acids Res.* 42, D613–D616. doi: 10.1093/nar/gkt1111
- Partida-Martínez, L. P., Groth, I., Schmitt, I., Richter, W., Roth, M., and Hertweck, C. (2007). *Burkholderia rhizoxinica* sp. nov. and *Burkholderia endofungorum* sp. nov., bacterial endosymbionts of the plant-pathogenic fungus *Rhizopus microsporus*. *Int. J. Syst. Evol. Microbiol.* 57, 2583–2590. doi: 10.1099/ijs.0.64660-0
- Payne, G. W., Vandamme, P., Morgan, S. H., LiPuma, J. J., Coenye, T., Weightman, A. J., et al. (2005). Development of a recA gene-based identification approach for the entire *Burkholderia* genus. *Appl. Environ. Microbiol.* 71, 3917–3927. doi: 10.1128/AEM.71.7.3917-3927.2005
- Perin, L., Martínez-Aguilar, L., Paredes-Valdez, G., Baldani, J., Estrada-de Los Santos, P., Reis, V., et al. (2006). *Burkholderia silvatlantica* sp. nov., a diazotrophic bacterium associated with sugar cane and maize. *Int. J. Syst. Evol. Microbiol.* 56, 1931–1937. doi: 10.1099/ijs.0.64362-0
- Reis, V., Estrada-De los Santos, P., Tenorio-Salgado, S., Vogel, J., Stoffels, M., Guyon, S., et al. (2004). *Burkholderia tropica* sp. nov., a novel nitrogen-fixing, plant-associated bacterium. *Int. J. Syst. Evol. Microbiol.* 54, 2155–2162. doi: 10.1099/ijs.0.02879-0
- Rokas, A., and Holland, P. W. H. (2000). Rare genomic changes as a tool for phylogenetics. *Trends Ecol. Evol.* 15, 454–459. doi: 10.1016/S0169-5347(00)01967-4
- Segata, N., Bornigen, D., Morgan, X. C., and Huttenhower, C. (2013). PhyloPhlAn is a new method for improved phylogenetic and taxonomic placement of microbes. *Nat. Commun.* 4, 2304. doi: 10.1038/ncomms3304
- Seo, Y. S., Lim, J., Choi, B. S., Kim, H., Goo, E., Lee, B., et al. (2011). Complete genome sequence of *Burkholderia gladioli* BSR3. *J. Bacteriol.* 193, 3149–3111. doi: 10.1128/JB.00420-11
- Sessitsch, A., Coenye, T., Sturz, A., Vandamme, P., Barka, E. A., Salles, J., et al. (2005). *Burkholderia phytofirmans* sp. nov., a novel plant-associated bacterium with plant-beneficial properties. *Int. J. Syst. Evol. Microbiol.* 55, 1187–1192. doi: 10.1099/ijs.0.63149-0
- Sheu, S.-Y., Chou, J.-H., Bontemps, C., Elliott, G. N., Gross, E., James, E. K., et al. (2012). *Burkholderia symbiotica* sp. nov., isolated from root nodules of *Mimosa* spp. native to north-east Brazil. *Int. J. Syst. Evol. Microbiol.* 62(Pt 9), 2272–2278. doi: 10.1099/ijs.0.037408-0
- Sheu, S.-Y., Chou, J.-H., Bontemps, C., Elliott, G. N., Gross, E., dos Reis Junior, F. B., et al. (2013). *Burkholderia diazotrophica* sp. nov., isolated from root nodules of *Mimosa* spp. *Int. J. Syst. Evol. Microbiol.* 63(Pt 2), 435–441. doi: 10.1099/ijs.0.039859-0
- Shibata, T. F., Maeda, T., Nikoh, N., Yamaguchi, K., Oshima, K., Hattori, M., et al. (2013). Complete genome sequence of *Burkholderia* sp. Strain RPE64, bacterial symbiont of the bean bug *Riptortus pedestris*. *Genome Announc.* 1, 10–13. doi: 10.1128/genomeA.00441-13
- Simpson, I. N., Finlay, J., Winstanley, D. J., Dewhurst, N., Nelson, J. W., Butler, S. L., et al. (1994). Multi-resistance isolates possessing characteristics of both *Burkholderia (Pseudomonas) cepacia* and *Burkholderia gladioli* from patients with cystic fibrosis. *J. Antimicrob. Chemother.* 34, 353–361. doi: 10.1093/jac/34.3.353
- Song, J. Y., Kwak, M. J., Lee, K. Y., Kong, H. G., Kim, B. K., Kwon, S. K., et al. (2012). Draft genome sequence of the antifungal-producing plant-benefiting bacterium *Burkholderia pyrrocinia* CH-67. *J. Bacteriol.* 194, 6649–6650. doi: 10.1128/JB.01779-12
- Spilker, T., Baldwin, A., Bumford, A., Dowson, C. G., Mahenthiralingam, E., and LiPuma, J. J. (2009). Expanded multilocus sequence typing for *Burkholderia* species. *J. Clin. Microbiol.* 47, 2607–2610. doi: 10.1128/JCM.00770-09
- Stead, D. (1992). Grouping of plant-pathogenic and some other *Pseudomonas* spp. by using cellular fatty acid profiles. *Int. J. Syst. Bacteriol.* 42, 281–295. doi: 10.1099/00207713-42-2-281
- Suarez-Moreno, Z. R., Caballero-Mellado, J., Coutinho, B. G., Mendonca-Previato, L., James, E. K., and Venturi, V. (2012). Common features of environmental and potentially beneficial plant-associated *Burkholderia*. *Microb. Ecol.* 63, 249–266. doi: 10.1007/s00248-011-9929-1
- Tamura, K., Stecher, G., Peterson, D., Filipski, A., and Kumar, S. (2013). MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Mol. Biol. Evol.* 30, 2725–2729. doi: 10.1093/molbev/mst197
- Tavaré, S. (1986). "Some probabilistic and statistical problems in the analysis of DNA sequences," in *Lectures on Mathematics in the Life Sciences*, 17^{Edn}, ed R. M. Miura (Providence, RI: American Mathematical Society), 57–86.
- Tayeb, L. A., Lefevre, M., Passet, V., Diancourt, L., Brisse, S., and Grimont, P. A. (2008). Comparative phylogenies of *Burkholderia*, *Ralstonia*, *Comamonas*, *Brevundimonas* and related organisms derived from *rpoB*, *gyrB* and *rrs* gene sequences. *Res. Microbiol.* 159, 169–177. doi: 10.1016/j.resmic.2007.12.005
- Tian, Y., Kong, B. H., Liu, S. L., Li, C. L., Yu, R., Liu, L., et al. (2013). *Burkholderia grimmiae* sp. nov., isolated from a xerophilous moss (*Grimmia montana*). *Int. J. Syst. Evol. Microbiol.* 63(Pt 6), 2108–2113. doi: 10.1099/ijs.0.045492-0
- Ussery, D. W., Kiil, K., Lagesen, K., Sicheritz-Ponten, T., Bohlin, J., and Wassenaar, T. M. (2009). The genus *Burkholderia*: analysis of 56 genomic sequences. *Genome Dyn.* 6, 140–147. doi: 10.1159/000235768

- Valverde, A., Delvasto, P., Peix, A., Velázquez, E., Santa-Regina, I., Ballester, A., et al. (2006). *Burkholderia ferrariae* sp. nov., isolated from an iron ore in Brazil. *Int. J. Syst. Evol. Microbiol.* 56, 2421–2425. doi: 10.1099/ijs.0.64498-0
- Vandamme, P., and Dawyndt, P. (2011). Classification and identification of the *Burkholderia cepacia* complex: past, present and future. *Syst. Appl. Microbiol.* 34, 87–95. doi: 10.1016/j.syapm.2010.10.002
- Vandamme, P., De Brandt, E., Houf, K., Salles, J. F., van Elsas, J. D., Spilker, T., et al. (2013). *Burkholderia humi* sp. nov., *Burkholderia choica* sp. nov., *Burkholderia telluris* sp. nov., *Burkholderia terrestris* sp. nov. and *Burkholderia udeis* sp. nov.: *Burkholderia glathei*-like bacteria from soil and rhizosphere soil. *Int. J. Syst. Evol. Microbiol.* 63(Pt 12), 4707–4718. doi: 10.1099/ijs.0.048900-0
- Vandamme, P., Goris, J., Chen, W. M., de Vos, P., and Willems, A. (2002b). *Burkholderia tuberum* sp. nov. and *Burkholderia phymatum* sp. nov., nodulate the roots of tropical legumes. *Syst. Appl. Microbiol.* 25, 507–512. doi: 10.1078/07232020260517634
- Vandamme, P., Goris, J., Chen, W.-M., De Vos, P., and Willems, A. (2002a). *Burkholderia tuberum* sp. nov. and *Burkholderia phymatum* sp. nov., nodulate the roots of tropical legumes. *Syst. Appl. Microbiol.* 25, 507–512. doi: 10.1078/07232020260517634
- Vandamme, P., Holmes, B., Vancanneyt, M., Coenye, T., Hoste, B., Coopman, R., et al. (1997). Occurrence of multiple genomovars of *Burkholderia cepacia* in cystic fibrosis patients and proposal of *Burkholderia multivorans* sp. nov. *Int. J. Syst. Bacteriol.* 47, 1188–1200. doi: 10.1099/00207713-47-4-1188
- Vandamme, P., Opelt, K., Knöchel, N., Berg, C., Schönmann, S., De Brandt, E., et al. (2007). *Burkholderia bryophila* sp. nov. and *Burkholderia megapolitana* sp. nov., moss-associated species with antifungal and plant-growth-promoting properties. *Int. J. Syst. Evol. Microbiol.* 57, 2228–2235. doi: 10.1099/ijs.0.65142-0
- Vanlaere, E., van der Meer, J. R., Falsen, E., Salles, J. F., De Brandt, E., and Vandamme, P. (2008). *Burkholderia sartisoli* sp. nov., isolated from a polycyclic aromatic hydrocarbon-contaminated soil. *Int. J. Syst. Evol. Microbiol.* 58, 420–423. doi: 10.1099/ijs.0.65451-0
- Van Oevelen, S., De Wachter, R., Vandamme, P., Robbrecht, E., and Prinsen, E. (2002a). Identification of the bacterial endosymbionts in leaf galls of Psychotria (Rubiaceae, angiosperms) and proposal of *Candidatus Burkholderia kirkii* sp. nov. *Int. J. Syst. Evol. Microbiol.* 52, 2023–2027. doi: 10.1099/ijs.0.02103-0
- Van Oevelen, S., De Wachter, R., Vandamme, P., Robbrecht, E., and Prinsen, E. (2002b). Identification of the bacterial endosymbionts in leaf galls of Psychotria (Rubiaceae, angiosperms) and proposal of *Candidatus Burkholderia kirkii* sp. nov. *Int. J. Syst. Evol. Microbiol.* 52(Pt 6), 2023–2027. doi: 10.1099/ijs.0.02103-0
- Van Oevelen, S., De Wachter, R., Vandamme, P., Robbrecht, E., and Prinsen, E. (2004). *Candidatus Burkholderia calva* and *Candidatus Burkholderia nigropunctata* as leaf gall endosymbionts of African Psychotria. *Int. J. Syst. Evol. Microbiol.* 54, 2237–2239. doi: 10.1099/ijs.0.63188-0
- Viallard, V., Poirier, I., Cournoyer, B., Haurat, J., Wiebkin, S., Ophel-Keller, K., et al. (1998). *Burkholderia graminis* sp. nov., a rhizospheric *Burkholderia* species, and reassessment of [*Pseudomonas*] phenazinium, [*Pseudomonas*] pyrrocinia and [*Pseudomonas*] *glathei* as *Burkholderia*. *Int. J. Syst. Bacteriol.* 48, 549–563. doi: 10.1099/00207713-48-2-549
- Weilharter, A., Mitter, B., Shin, M. V., Chain, P. S., Nowak, J., and Sessitsch, A. (2011). Complete genome sequence of the plant growth-promoting endophyte *Burkholderia phytofirmans* strain PsJN. *J. Bacteriol.* 193, 3383–3384. doi: 10.1128/JB.05055-11
- White, N. J. (2003). Melioidosis. *Lancet* 361, 1715. doi: 10.1016/S0140-6736(03)13374-0
- Whitlock, G. C., Estes, D. M., and Torres, A. G. (2007). Glanders: off to the races with *Burkholderia mallei*. *FEMS Microbiol. Lett.* 277, 115–122. doi: 10.1111/j.1574-6968.2007.00949.x
- Wiersinga, W. J., van der Poll, T., White, N. J., Day, N. P., and Peacock, S. J. (2006). Melioidosis: insights into the pathogenicity of *Burkholderia pseudomallei*. *Nat. Rev. Microbiol.* 4, 272–282. doi: 10.1038/nrmicro1385
- Wong, S. Y., Paschos, A., Gupta, R. S., and Schellhorn, H. E. (2014). Insertion/deletion-based approach for the detection of *Escherichia coli* O157:H7 in freshwater environments. *Environ. Sci. Technol.* 48, 11462–11470. doi: 10.1021/es502794h
- Wu, D., Hugenholtz, P., Mavromatis, K., Pukall, R., Dalin, E., Ivanova, N. N., et al. (2009). A phylogeny-driven genomic encyclopaedia of Bacteria and Archaea. *Nature* 462, 1056–1060. doi: 10.1038/nature08656
- Yabuuchi, E., Kosako, Y., Oyaizu, H., Yano, I., Hotta, H., Hashimoto, Y., et al. (1992). Proposal of *Burkholderia* gen. nov. and transfer of seven species of the genus *Pseudomonas* homology group II to the new genus, with the type species *Burkholderia cepacia* (Palleroni and Holmes 1981) comb. nov. *Microbiol. Immunol.* 36, 1251–1275. doi: 10.1111/j.1348-0421.1992.tb02129.x
- Yabuuchi, E., Kosako, Y., Oyaizu, H., Yano, I., Hotta, H., Hashimoto, Y., et al. (1993). *Burkholderia* gen. nov. validation of the publication of new names and new combinations previously effectively published outside the IJSB, List no 43. *Int. J. Syst. Bacteriol.* 43, 398–399. doi: 10.1099/00207713-43-2-398
- Yang, H.-C., Im, W.-T., Kim, K. K., An, D.-S., and Lee, S.-T. (2006). *Burkholderia terrae* sp. nov., isolated from a forest soil. *Int. J. Syst. Evol. Microbiol.* 56, 453–457. doi: 10.1099/ijs.0.63968-0
- Yarza, P., Richter, M., Peplies, J., Euzéby, J., Amann, R., Schleifer, K. H., et al. (2008). The All-Species Living Tree project: A 16S rRNA-based phylogenetic tree of all sequenced type strains. *Syst. Appl. Microbiol.* 31, 241–250. doi: 10.1016/j.syapm.2008.07.001
- Yoo, S.-H., Kim, B.-Y., Weon, H.-Y., Kwon, S.-W., Go, S.-J., and Stackebrandt, E. (2007). *Burkholderia soli* sp. nov., isolated from soil cultivated with Korean ginseng. *Int. J. Syst. Evol. Microbiol.* 57, 122–125. doi: 10.1099/ijs.0.64471-0
- Zhang, H., Hanada, S., Shigematsu, T., Shibuya, K., Kamagata, Y., Kanagawa, T., et al. (2000). *Burkholderia kururiensis* sp. nov., a trichloroethylene (TCE)-degrading bacterium isolated from an aquifer polluted with TCE. *Int. J. Syst. Evol. Microbiol.* 50, 743–749. doi: 10.1099/00207713-50-2-743
- Zhu, B., Zhou, S., Lou, M., Zhu, J., Li, B., Xie, G., et al. (2011). Characterization and inference of gene gain/loss along *Burkholderia* evolutionary history. *Evol. Bioinform. Online* 7, 191. doi: 10.4137/EBO.S7510

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