



Novel Insights Into Triple-Negative Breast Cancer Prognosis by Comprehensive Characterization of Aberrant Alternative Splicing

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Background: Alternative splicing (AS) is important in the regulation of gene expression and aberrant AS is emerging as a major factor in the pathogenesis of human conditions, including cancer. Triple-negative breast cancer (TNBC) is the most challenging subtype of breast cancer with strong invasion, high rate of metastasis, and poor prognosis. Here we report a systematic profiling of aberrant AS in TNBC.

Methods: The percent spliced in (PSI) values for AS events in 151 TNBC patients were obtained from The Cancer Genome Atlas (TCGA) SpliceSeq database. Univariate Cox and stepwise Multivariate Cox regression analyses were conducted to find the best prognostic AS model. Splicing regulatory networks were constructed by prognosis-related spliceosome and aberrant AS events. Additionally, pathway enrichment and gene set enrichment analysis (GSEA) were further employed to reveal the significant pathways for prognosis-related AS genes. Finally, splicing regulatory networks were constructed via Spearman's rank correlation coefficients between prognosis-related AS events and splicing factor expressions.

Results: A total of 1,397 prognosis-associated AS events were identified in TNBC. The majority of the parent genes of prognostic AS events exhibited direct interactions to each other in the STRING gene network. Pathways of focal adhesion ($p < 0.001$), RNA splicing ($p = 0.007$), homologous recombination ($p = 0.042$) and ECM-receptor interaction ($p = 0.046$) were found to be significantly enriched for prognosis-related AS. Additionally, the area under curve (AUC) of the best AS prognostic predictor model reached 0.949, showing a powerful capability to predict outcomes. The Exon Skip (ES) type of AS events displayed more robust and efficient capacity in predicting performance than any other specific AS events type in terms of prognosis. The ES AS signature might confer a strong oncogenic phenotype in the high-risk group with elevated activities in cell cycle and SUMOylating pathways of tumorigenesis, while programmed cell death and metabolism pathways were found to be enriched in the low-risk group of TNBC.

The splicing correlation network also revealed a regulatory mode of prognostic splicing factors (SFs) in TNBC.

Conclusion: Our analysis of AS events in TNBC could not only contribute to elucidating the tumorigenesis mechanism of AS but also provide clues to uncovering underlying prognostic biomarkers and therapeutic targets for further study.

Keywords: alternative splicing, triple-negative breast cancer, The Cancer Genome Atlas, prognostic biomarker, splicing correlation network

INTRODUCTION

Breast cancer is one of the most lethal malignancies in women worldwide with an estimated 268,600 new cases diagnosed per year in the United States (DeSantis et al., 2019). From a pathological viewpoint, breast cancer can be characterized into four basic subtypes with the presence or absence of two hormone receptors including estrogen receptor (ER), progesterone receptor (PR), and human epidermal growth factor receptor 2 (HER2) (Onitilo et al., 2009; Dai et al., 2016). TNBC is defined as the lack of ER, PR and the absence of protein overexpression/gene amplification of HER2, which accounts for approximately 15 to 20% of all breast cancers (Iorio et al., 2016). Compared with other subtypes, TNBC is the most clinically challenging subtype due to its strong invasion, high rate of metastasis, and poor OS. To date, chemotherapy remains the only standard treatment, while other therapies did not exhibit any significantly improvement of survival for TNBC (Bianchini et al., 2016). Furthermore, TNBC is a highly heterogeneous disease whose molecular mechanism of progression and aggressiveness is still unclear. Therefore, exploration of the mechanism that promotes TNBC progression and development of novel predictive prognostic biomarkers are crucial.

Over the last few decades, major efforts have been made to reveal the underlying mechanisms of TNBC that could assist in the prediction of prognostic biomarkers and the guidance of anti-cancer treatments. In addition, extensive studies have been performed on genomic, transcriptomic, and even DNA methylation features to classify TNBC into intrinsic molecular subtypes, which have widely expanded our knowledge on TNBC (Bianchini et al., 2016; Stirzaker et al., 2016; Shi et al., 2018). However, these studies only focus on mutation and abnormal gene expression while ignoring other important oncogenesis mechanisms such as aberrant alternative splicing.

Alternative splicing is a post-transcriptional modification mechanism to ensure high transcript and protein diversity during the lifespan of eukaryotic organisms (Wang et al., 2015). It has been estimated that up to 95% of multi-exon genes undergo

the AS process, and the vast majority of them vary across different human cells and tissues (Pan et al., 2008; Wang et al., 2008; Ke and Chasin, 2011). AS events are categorized into seven common types as follows: Alternate Acceptor site (AA); Alternate Donor site (AD); Alternate Promoter (AP); Alternate Terminator (AT); Exon Skip (ES); Mutually Exclusive Exons (ME), and RI (Figure 1A) (Ryan et al., 2012, 2016). Growing evidence indicated that the aberrant AS patterns were implicated in many human cancers (Oltean and Bates, 2014). Deep analyses of AS events in non-small cell lung cancer, gastrointestinal cancer and breast cancer have provided abundant clues linking aberrant AS events and the tumorigenesis of cancer (Li et al., 2017; Lin et al., 2018; Zhang et al., 2019). Particularly, global dysregulation of AS was regarded as a driver event causing breast cancer progression and metastasis (Read and Natrajan, 2018). Despite this, the underlying regulatory mechanisms of AS and their clinical implications were not yet fully understood in TNBC. Hence, identification of aberrant AS events as a potential prognostic biomarker and therapeutic target for TNBC is both promising and imperative.

In the present study, we conducted a comprehensive analysis of genome-wide AS events in the TNBC from TCGA. Subsequently, we determined prognosis-associated AS events and uncovered reliable and distinct prognostic signatures for TNBC. In addition, we constructed a prognosis-associated splicing factor-alternative splicing (SF-AS) network to explore the splicing factors and their potential targets. Our findings represent an initial attempt to uncover TNBC related AS events. Moreover, further exploration of splicing regulatory factors on their AS targets may aid in fully understanding the underlying mechanism of splicing factors, which could eventually help to elucidate the role that AS plays in the tumorigenesis of TNBC.

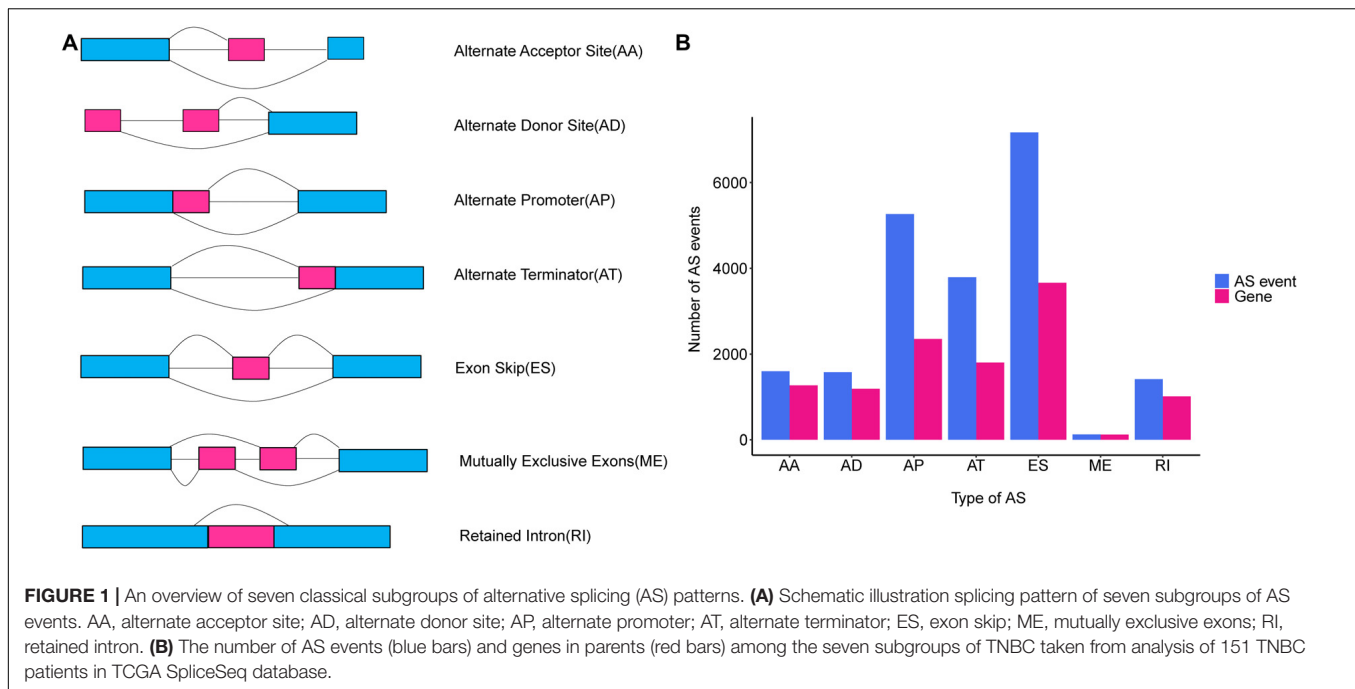
MATERIALS AND METHODS

TCGA RNAseq and Alternative Splicing Event Data Process

RNAseq-HTSeq data and clinical data of patients who were diagnosed with primary breast cancer were downloaded from the TCGA¹. The inclusive criteria for categorizing TNBC are as follows: (1) loss of expression for ER and PR, and HER2 by immunohistochemistry (IHC), and the absence of the amplification of HER2 by fluorescent *in situ* hybridization (FISH)

¹<https://tcga-data.nci.nih.gov/tcga/>

Abbreviations: AA, alternate acceptor site; AD, alternate donor site; AP, alternate promoter; AS, alternative splicing; AT, alternate terminator; AUC, area under the curve; BP, biological process; CC, cellular component; DAVID, Database for Annotation, Visualization and Integrated Discovery; ES, exon skip; GO, gene ontology; GSEA, gene set enrichment analysis; HR, hazard ratios; KEGG, Kyoto Encyclopedia of Genes and Genomes; ME, mutually exclusive exons; MF, molecular function; MSigDB, Molecular Signatures Database; OS, overall survival; PSI, percent spliced in; RI, retained intron; ROC, receiver operator characteristic; SF, splicing factor; TCGA, The Cancer Genome Atlas; TNBC, triple-negative breast cancer.



being needed if the IHC result was equivocal (Penault-Llorca and Viale, 2012); (2) patients with clinicopathological information and who had an OS of at least 30 days; (3) patients did not suffer from other types of tumors. Based on these screening criteria, a total of 151 TNBC samples were enrolled in this study (Penault-Llorca and Viale, 2012).

AS profilings of TNBC cohorts were downloaded from TCGA SpliceSeq² (Ryan et al., 2016), a web-based resource for generating splicing patterns of TCGA tumors. Percent-splice-in (PSI) value is used to quantify splicing events, it represents the ratio of inclusive reads over inclusive and exclusive reads normalized by read length in each splicing event, and always ranges from 0 to 1. The inclusion criteria for a reliable set of AS events were required the percentage of Samples with PSI Value $\geq 75\%$ and a standard deviation of > 0.05 . In addition, missing splicing data were imputed by the K-nearest neighbor (KNN) algorithm from Pamr package in R.

UpSet Plot Functional and Pathway Enrichment and PPI Networks Analysis

Given that one gene harbors more than one type of AS events, intersections between protein-coding genes in seven types of AS events in TNBC were visualized by UpsetR package in R. The GO (Ashburner et al., 2000) of BP, CC, MF, and KEGG pathway (Kanehisa et al., 2017) were analyzed by the DAVID (version 6.8) (Huang da et al., 2009) web tool with default settings, and was visualized by the gglpot2 package in R. Subsequently, the parent genes of the most prognostically significant AS events ($p < 0.001$) were imported into STRING database (version 11) (Szklarczyk et al., 2019) with a confident

score > 0.4 to construct a gene-interaction network. Cytoscape (version 3.7.1) (Shannon et al., 2003) was further applied to display their relationships in the gene-interaction network.

Survival Analysis and the Construction of Prognostic Model

For each type of AS events, TNBC cohorts were divided into two groups by a median cut of PSI value. Univariate Cox proportional hazard regression analysis was employed to detect the survival-related AS events with $p < 0.05$ in all seven AS subgroups. The most significant top 10 genes in each AS event model were chosen as candidates for the forest plots. The top 10 prognostically aberrant AS gene were then merged into a Multivariate Cox regression model to build candidate prognostic models. To make our model more practical and robust, a forward stepwise method was used to search a minimal set of candidate AS events. In addition, a combination of AS PSI levels weighted by regression coefficient (β) from the Multivariate Cox regression analysis was used to construct a risk score model (Scosyrev and Glimm, 2019). The risk score for each patient was as follows: Risk score = $\beta_1 \times AS_1 + \beta_2 \times AS_2 + \dots + \beta_n \times AS_n$. Kaplan-Meier Curves (Logrank Tests) were used to verify whether the predictive models could differentiate the OS between high-risk and low-risk groups in TNBC. The AUC of the Receiver Operating Characteristic (ROC) curve was calculated by survival ROC package for each AS events based on the prognostic model. All statistical analyses were performed using R (version 3.5.3).

Gene Set Enrichment Analysis (GSEA) for the AS Event-Based Classifier

To further explore the biological pathways in the AS Event-Based predictive model, GSEA analysis (Subramanian et al., 2005)

²<http://bioinformatics.mdanderson.org/TCGASpliceSeq>

was performed to verify the differences in BP and pathways between high-risk and low-risk groups with the JAVA program using the annotated gene set of “C2: Canonical pathways” which was downloaded from the MSigDB³. After performing 1000 permutation steps, the gene sets with nominal $p < 0.05$ were considered to be significantly enriched.

Construction of the Splicing Factors (SFs) Correlation Networks

The associations between prognostic AS events and their regulated SFs were further investigated. A list of 71 known SFs was extracted from the SpliceAid2 database⁴ (Piva et al., 2012). The curated level 3 mRNA-seq data of SFs were downloaded from TCGA portal¹ and normalized by TMM method in edgeR packages (Robinson et al., 2010). To find a potential regulatory network between the prognosis-associated SFs and AS events, Univariate Cox regression analysis was first conducted to screen survival-associated SFs, and then Spearman test was implemented to analyze the correlation between the mRNA expression level of prognostic SFs and PSI values of AS events. The significantly correlated pairs for SFs with PSI values of AS events ($|r| > 0.4$, $p < 0.001$) were selected as the candidates for the splicing correlation network. Lastly, Cytoscape (version 3.7.1) was further used to plot the potential SF-AS regulatory network of the significant correlation between SFs and AS events.

RESULTS

Integrated Overview of AS Events Profiling in TNBC Cohort

The comprehensive genome-wide AS events profiling of 151 TNBC patients with clinical implications was generated by TCGA SpliceSeq, in among which the median follow-up OS was 30.5 months (range 1 to 117 months). To give a precise description for each AS event, taking “HNRNPA1_ES_212638” for example, “HNRNPA1” represented the parent gene of the AS event, “ES” was the AS type, and the number “212638” stood for the AS event ID from the TCGA SpliceSeq database. Totally, we detected 20,931 AS events from 7,358 genes, including 7,166 ESs in 3,661 genes, 1,413 RIs in 1012 genes, 5,263 APs in 2,353 genes, 3,789 ATs in 1,802 genes, 1,575 ADs in 1,189 genes, 1,600 AAs in 1,269 genes and 125 MEs in 122 genes (Figure 1B). Additionally, it should be noted that one gene harbored up to three AS events on average. Among these genes, Collagen Type I Alpha 2 Chain (COL1A2) had the max number of AS events ($n = 48$), followed by Mitochondrial Ribosomal Protein L55 (MRPL55) ($n = 45$) and Arylformamidase (AFMID) ($n = 36$), which suggested that one gene with multiple AS events could result in combinatorial arrangements and greatly contribute to transcriptome and proteome diversity. Among these splicing patterns, ES events were observed as the most frequent type, which accounted for 34.2% of the AS types, followed by AP (25.1%) and AT (18.1%) events.

³<http://software.broadinstitute.org/gsea/index.jsp>

⁴www.introni.it/spliceaid.html

Identification of Prognosis-Associated AS Events in TNBC

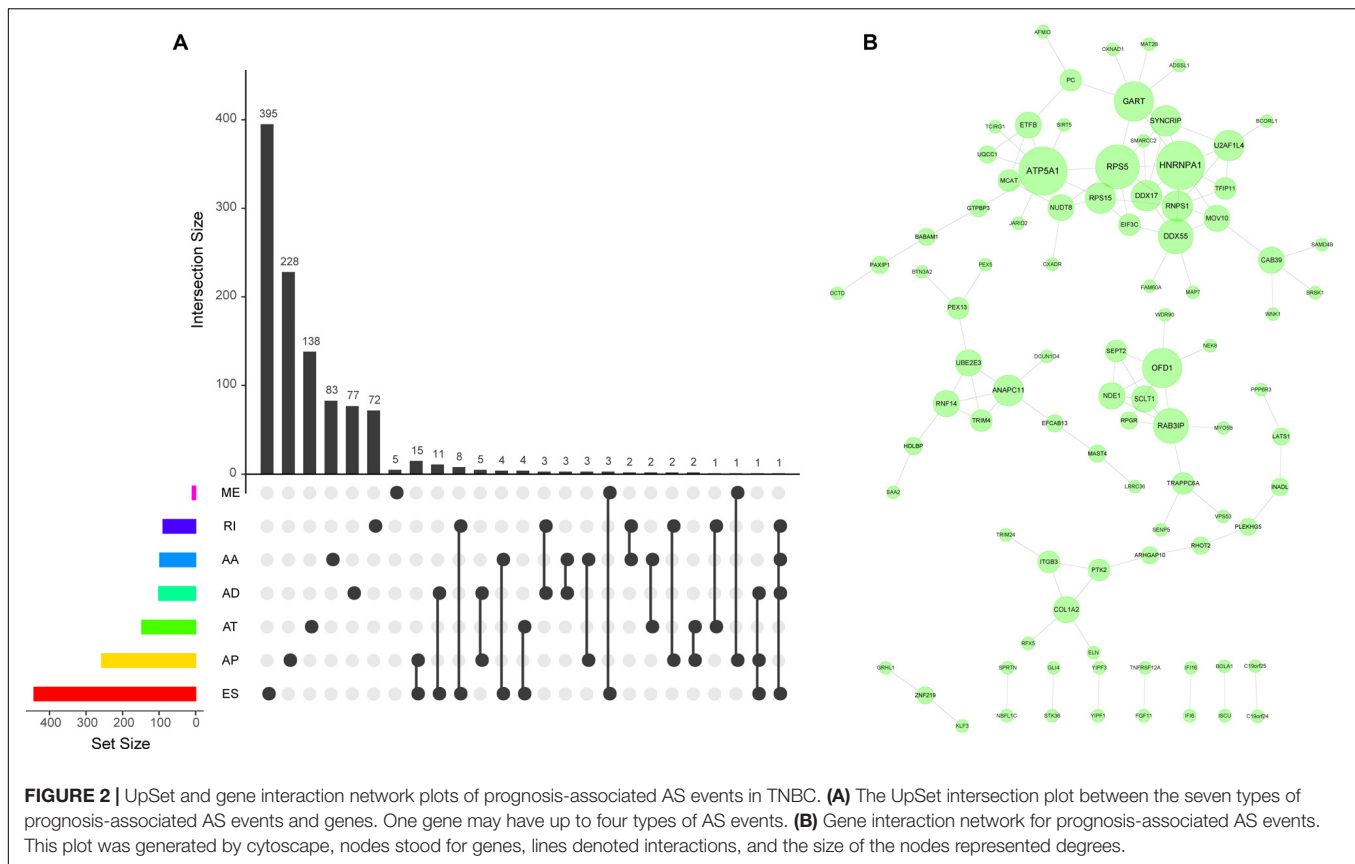
To explore the association between AS events and prognosis in patients with TNBC, we conducted cox univariate analyses to evaluate the prognostic factor in each AS event type. TNBC patients were categorized into low- and high-PSI groups according to the median PSI value. Consequently, a total of 1,397 AS events were found to be significantly associated with OS (log-rank $p < 0.05$), which accounted for 6.7% of all AS events and 14.5% of all the patent genes in TNBC (Supplementary Table 1). It was noteworthy that most of these significant prognosis-associated AS events (800 ASs out of 1,397 ASs) were favorable prognostic factors ($HR < 1$). To quantitatively analyze the overlap between genes and AS events amongst the seven AS types, the UpSet plot was created to visualize the intersecting sets in Figure 2A, indicating that most of the prognosis-associated AS genes might have one AS events type, and some of them may have up to four AS event types. For example, AA, AD, ES, and RI events of TMEM205 were found to be significantly associated with TNBC patients' survival.

Protein-Protein Network Analyses of Survival-Associated AS Events

To investigate the interactions among these genes of prognosis-associated AS events, we conducted gene interaction network analyses in TNBC. Parent genes of prognosis-associated AS events ($p < 0.001$) were imported to the STRING PPI database (version 11) with a score > 0.4 . A PPI-network generated by cytoscape illustrated the hub genes (HNRNPA1, RPS5, DDX55, OFD1, et al.) and their interactions in the prognosis related network (Figure 2B). Our results suggested that the majority of the parent genes of prognostic AS events exhibited direct gene-gene interaction to each other in the STRING gene network, indicating AS dysregulation genes played a crucial role in the process of TNBC tumorigenesis.

Functional Enrichment Analysis of Survival-Associated AS Events

Next, in order to evaluate the potential impact of prognosis-associated AS events on corresponding gene biological functions in TNBC, GO, and KEGG pathways enrichment analyses for their parent genes were performed. The results revealed that a total of 49 significant terms in GO BP categories were closely related to TNBC, including regulation of transcription, DNA-templated ($p < 0.001$), and RNA splicing ($p = 0.007$) (Figure 3A). Besides, 29 pathways in GO CC indicated significant differences in terms such as focal adhesion ($p < 0.001$) and Cul4-RING E3 ubiquitin ligase complex ($p = 0.0015$) (Figure 3B). We also observed that 24 pathways in GO MF (Figure 3C) were highlighted, including significant difference in terms of poly(A) RNA binding ($p < 0.001$), RNA polymerase II core promoter proximal region sequence-specific DNA binding ($p = 0.0016$), as well as SUMO-specific protease activity ($p = 0.005$). Additionally, 5 KEGG pathways associated with TNBC carcinogenesis were significantly affected in TNBC ($p < 0.05$, Figure 3D), including Regulation of actin cytoskeleton ($p = 0.0015$), Homologous



recombination ($p = 0.042$) and ECM-receptor interaction ($p = 0.046$). Enriched KEGG pathways were depicted in **Figure 3D**, and the detailed significant pathway enrichment results were listed in **Supplementary Table 2**. Collectively, these findings suggested that the prognostic associated AS events not only played a vital role in tumor proliferation pathways such as Homologous recombination and RNA splicing, but also participated in invasion and metastasis related processes such as focal adhesion and ECM-receptor interaction, which could contribute to uncovering the underlying mechanisms linking OS-associated AS events and protein biological function during the carcinogenesis in TNBC.

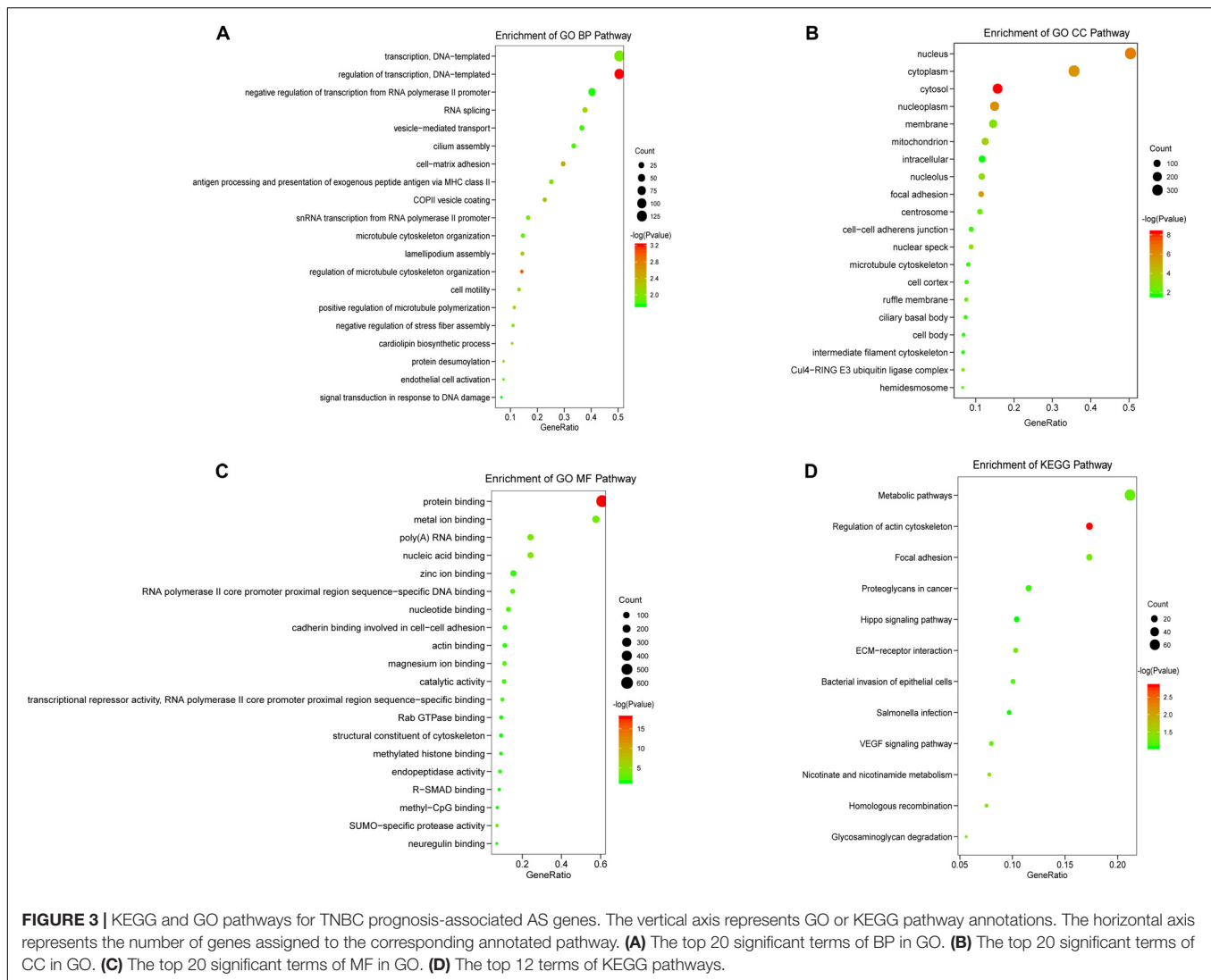
Construction of the Prognostic Predictors in TNBC Cohort

To evaluate the independent prognostic factors of AS events in the TNBC cohort, the top 10 significant prognostic related AS events were chosen as candidates, which could be displayed in forest plots (**Figure 4**). Multivariate Cox regression analysis for independent prognostic factors was subsequently employed to construct prognostic predictor models in seven AS events type, respectively. Risk score was first calculated in each AS events type, and the TNBC patients were stratified into high- and low-risk subgroups by the median value of risk scores. In our analysis of each splicing prognostic signature, the prognostic models built with seven different AS types 8 AA events, 5

AD events, 4 AP events, 4 AT events, 6 ES events, 2 MEs or 5 RI events, showed great powers ($p < 0.0001$) to predict the prognostic outcome respectively (**Figure 5**). Particularly the prognostic predictor of single ES events type exhibited the most powerful capability to predict the outcome amongst the seven prognostic models. Moreover, time dependent ROC curves from 3 years survival were also employed to compare the efficiency among AS signatures. As shown in **Figure 5**, an AUC value of > 0.7 was observed in each type of AS in TNBC, and the ES prognostic model exhibited the best predictor efficiency at distinguishing favorable or adverse outcomes (AUC = 0.949), followed by the AP models and RI models with AUCs of 0.944 and 0.926 respectively. The detailed information of these prognosis-associated AS events was summarized in **Supplementary Table 3**. Additionally, the distribution of survival status, risk score, and the heatmaps for the seven AS splicing patterns in TNBC patients were shown in **Figure 6**. It could be confirmed that the ES events type displayed a more robust and efficient capacity in predicting performance than any other specific AS events type in aspects of prognosis.

Gene Set Enrichment Analysis (GSEA) of the ES AS Signature

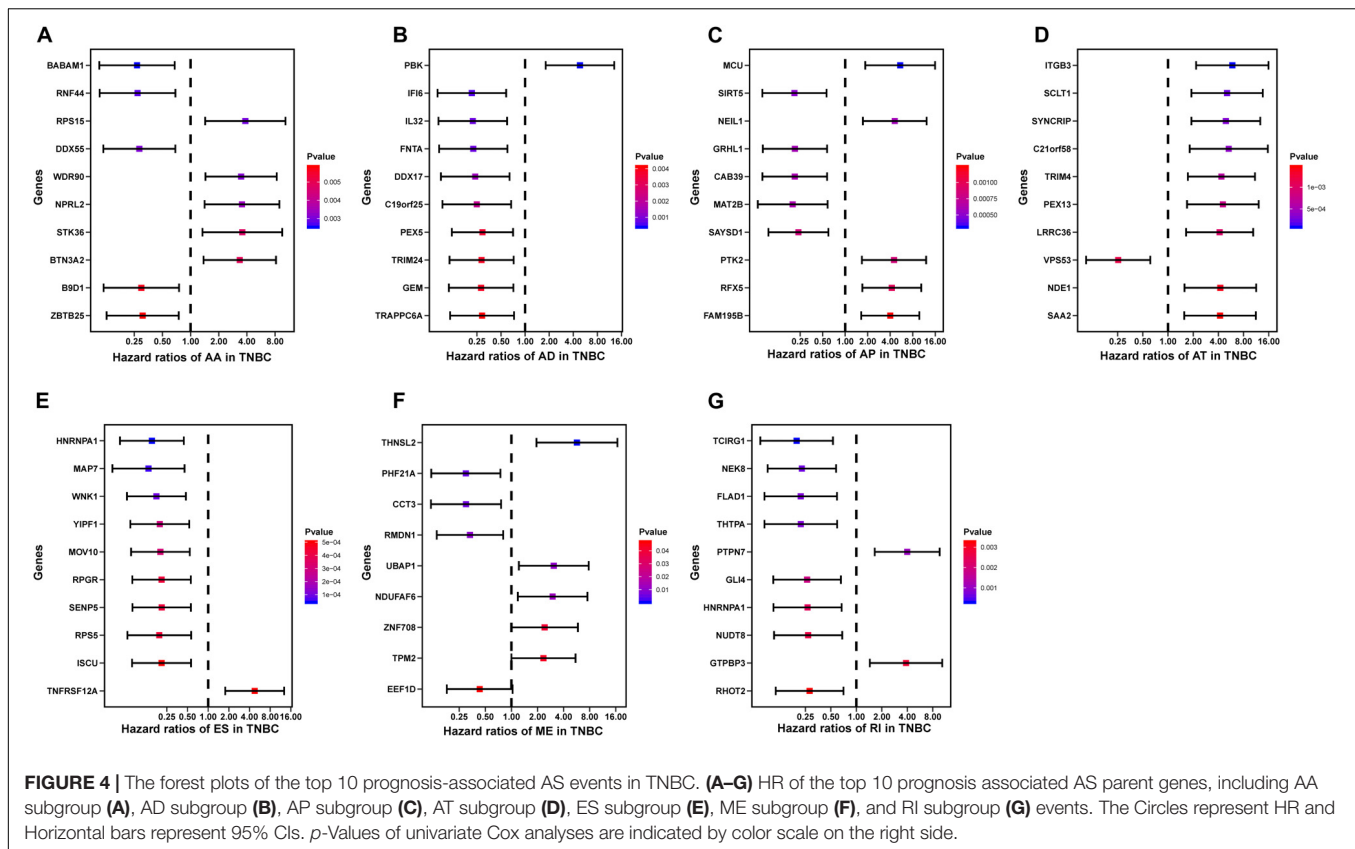
To explore the specific pathways involved in the ES prognosis-associated AS signature, GSEA analyses were further implemented. By using GSEA with expression of mRNA



between low- and high-risk groups from the ES AS signature, several oncogenic pathways were significantly enriched in the high-risk group, including Sumoylation ($p = 0.01$), Cell cycle ($p = 0.018$), Homologous DNA pairing and strand exchange ($p = 0.022$), POLO like kinase mediated events ($p = 0.035$) (Figure 7), which exhibited that they were involved in the tumorigenesis and proliferation of TNBC. On the contrary, specific genes overexpressed in the low-risk group showed significant enrichment in Regulated necrosis ($p = 0.002$), TRIF mediated programmed cell death ($p = 0.006$), Amino sugar and nucleotide sugar metabolism ($p = 0.02$), Coenzyme A biosynthesis ($p = 0.021$), and Vitamin B2 riboflavin metabolism ($p = 0.025$). Detailed GSEA results were listed in **Supplementary Table 4**. Overall, the ES AS signature might confer a strong oncogenic phenotype in the high-risk group with elevated activities in cell cycle and SUMOylating pathways of tumorigenesis while programmed cell death and metabolism pathways were found to be enriched in the low-risk group of TNBC.

Construction of a Potential SF-AS Regulatory Network

It is widely acknowledged that several cancer spliceosomes are involved in the regulation of AS events during tumor progress (El Marabti and Younis, 2018), therefore construction of the SF-AS regulatory network based on the significant correlations between prognosis-associated SFs and AS events was necessary. Herein, a total of 71 experimentally validated SFs from the SpliceAid2 database were selected to identify prognosis-related splicing factors. We detected 5 key SFs whose expression levels were strongly associated with prognosis ($p < 0.01$) in TNBC, these are Heterogeneous Nuclear Ribonucleoprotein U (HNRNPU), Splicing Factor 3b Subunit 1 (SF3B1), FMRP Translational Regulator 1 (FMR1), Polypyrimidine Tract Binding Protein 2 (PTBP2), and Splicing Factor Proline and Glutamine Rich (SFPQ). Furthermore, spearman correlation coefficient was used to assess the association between prognosis-associated SFs and AS events. After that, we characterized the SF-AS regulatory network by five prognostic splicing factors and the



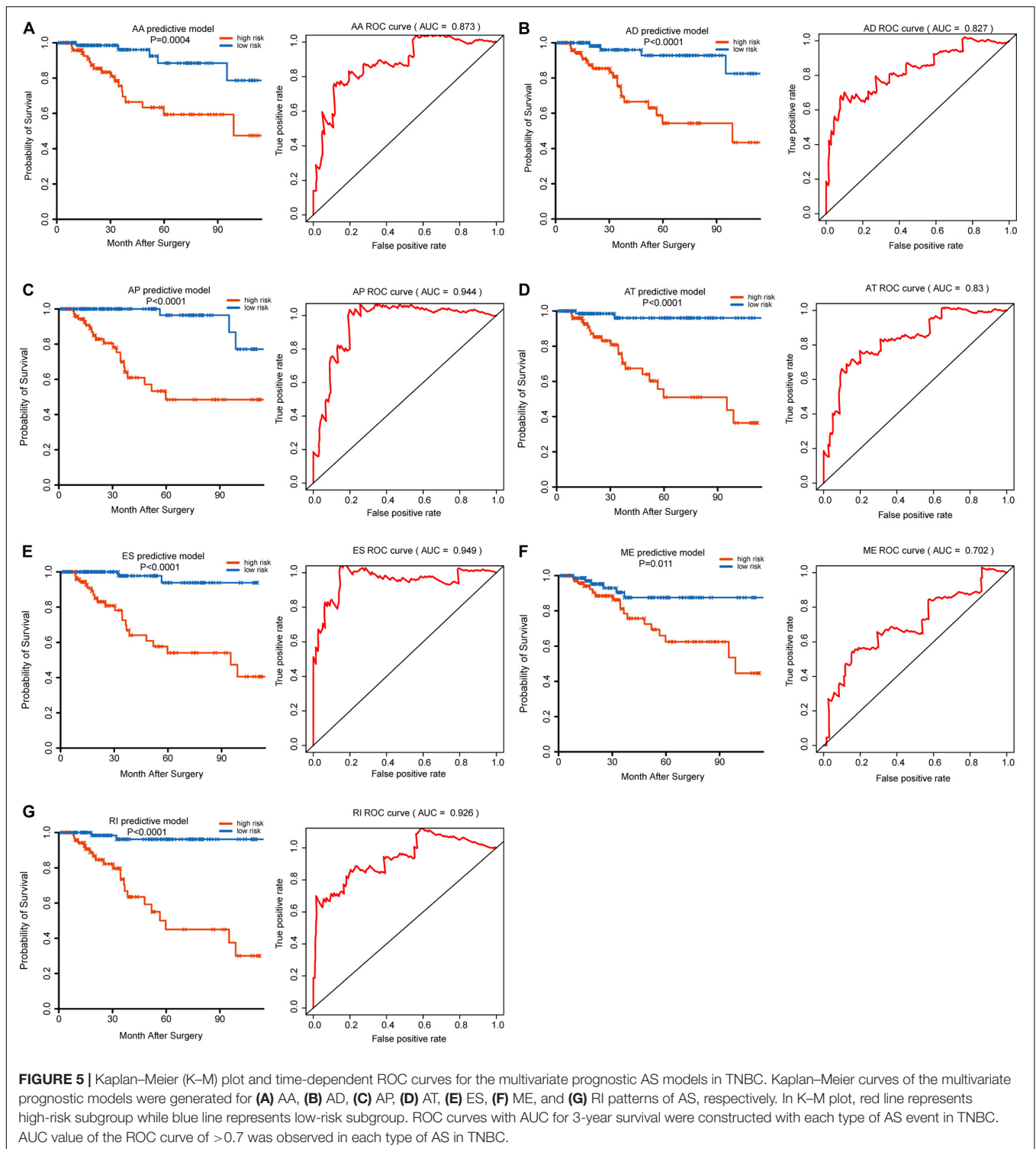
most significant survival-associated AS events ($p < 0.001$) in TNBC (Supplementary Table 5). As depicted in the SF-AS regulatory network (Figure 8A), it was observed that most favorable survival-related AS events (pink dots) were positively regulated (pink edges) with the expression of SFs while the majority of adverse prognostic AS events (green dots) were negatively (green edges) correlated by SFs, which was consistent with results of survival-associated SFs. Notably, the majority of key AS factors (orange dots) were observed to be related to more than one AS event, and some of them even played opposite roles in regulation of different AS events. Moreover, we also observed that different SFs could affect the same AS event simultaneously, for example, splicing factors SFPQ and HNRNPA1 were both significantly associated with AA of QKI, which implied that splicing process of the same gene might be co-regulated by different prognostic SFs. Representative correlations between SFs and their survival-related AS events were displayed in the scatter plots (Figures 7B–E). For example, expression of SF3B1 was positively correlated with AP of SMAD4 with $r = 0.55$, $p < 0.001$ (Figure 8B), whereas FMR1 negatively correlated with ES of TP53BP2 with $r = -0.53$, $p < 0.001$ (Figure 8C).

DISCUSSION

Alternative splicing is regarded as one of the important gene regulatory processes that could enable one single gene to

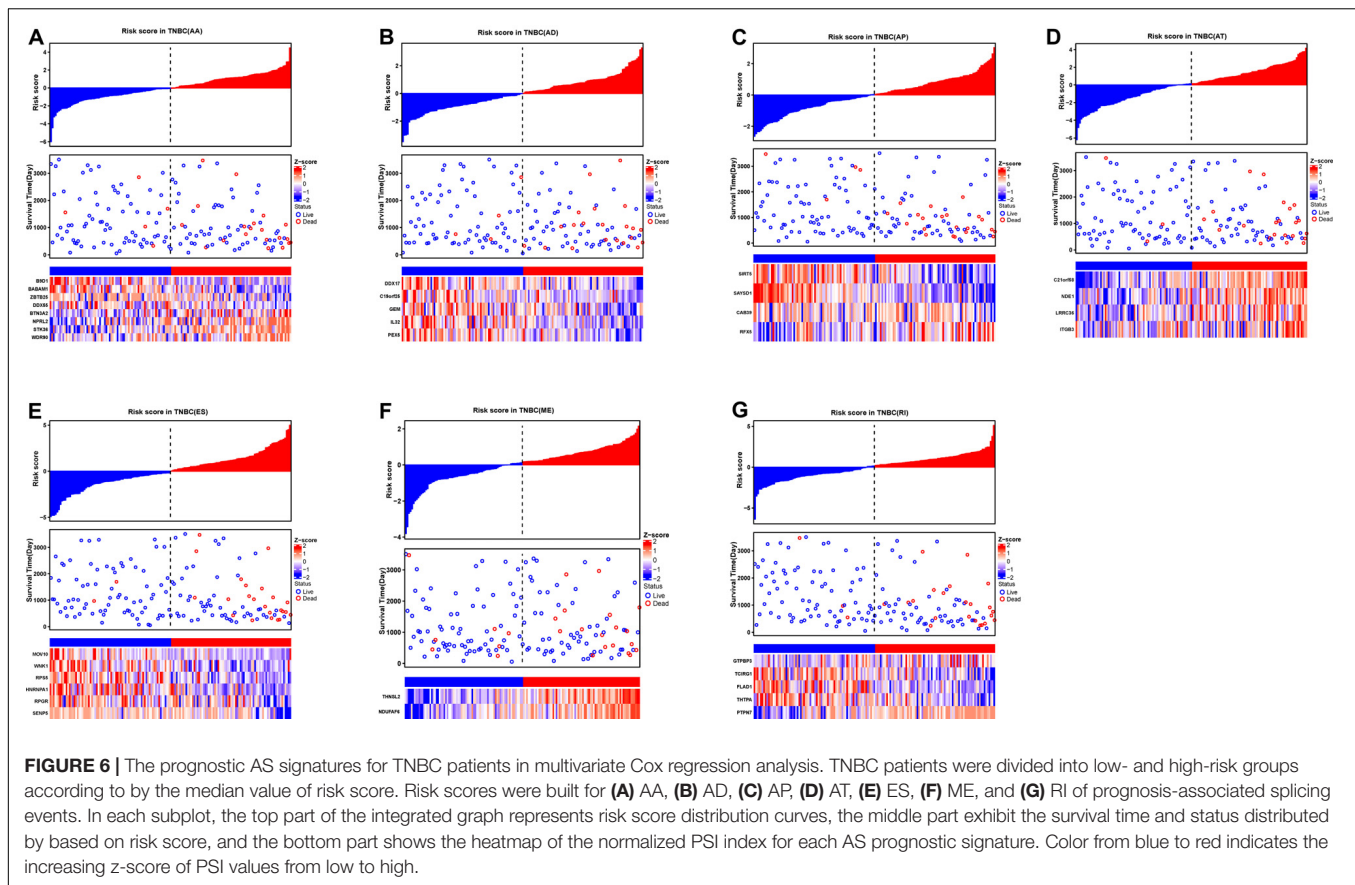
generate multiple mRNAs and translated various proteome in human genome. AS has a tremendous impact on processes of cell growth and development, and accumulating investigations suggest that AS perturbation engaged in multiple tumorigenesis processes such as cancer onset, progression, metastasis, hypoxia, as well as angiogenesis, particularly dysregulated in colorectal adenocarcinoma and lung adenocarcinoma (Misquitta-Ali et al., 2011; Lokody, 2014). Carcinogenesis in TNBC is a complex process that involves aberrant mRNA AS and the production of protein diversity. Previous studies demonstrated that the AS events or regulator factors in breast cancer were aberrantly expressed, thus contributing to the pathogenesis mechanisms of tumor formation (Read and Natrajan, 2018). For example, Han et al. (2017) discovered that intron retention was the most dominant type of hypoxia-induced AS in breast cancer. Ke et al. (2018) identified TDP43 as a crucial splicing regulator that interacted with SRSF3, which was responsible for the unique AS profile in TNBC. Nevertheless, these studies were based solely at the level of single genes, while a systematic analysis at the level of the whole genome that could better characterize aberrant AS events as potential prognostic and predictive biomarkers in TNBC is still lacking.

In our study, we downloaded RNA splicing data of 151 TNBC samples from TCGA SpliceSeq database and performed a genome-wide profiling analysis to elucidate the oncogenic roles of AS in TNBC. Our analysis preliminarily detected 1,397 AS



events from 1,069 parent genes that were significantly associated with OS in TNBC. According to our results, ES events were observed as the most frequent type, accounting for nearly one third of the AS types, followed by AP and AT events, which was concordant with other studies (Xiong et al., 2018; Li et al., 2019; Zhang et al., 2019). Notably, among the genes with multiple

AS events, Collagen Type I Alpha 2 Chain (COL1A2), which is a subunit of type I collagen, has the maximum number of AS events ($n = 48$). In addition, further analysis also identified COL1A2 AS events as being significantly associated with survival. Evidence showed that COL1A2 could activate $\beta 1$ -integrin and the activation, along with the epithelial-mesenchymal transition,

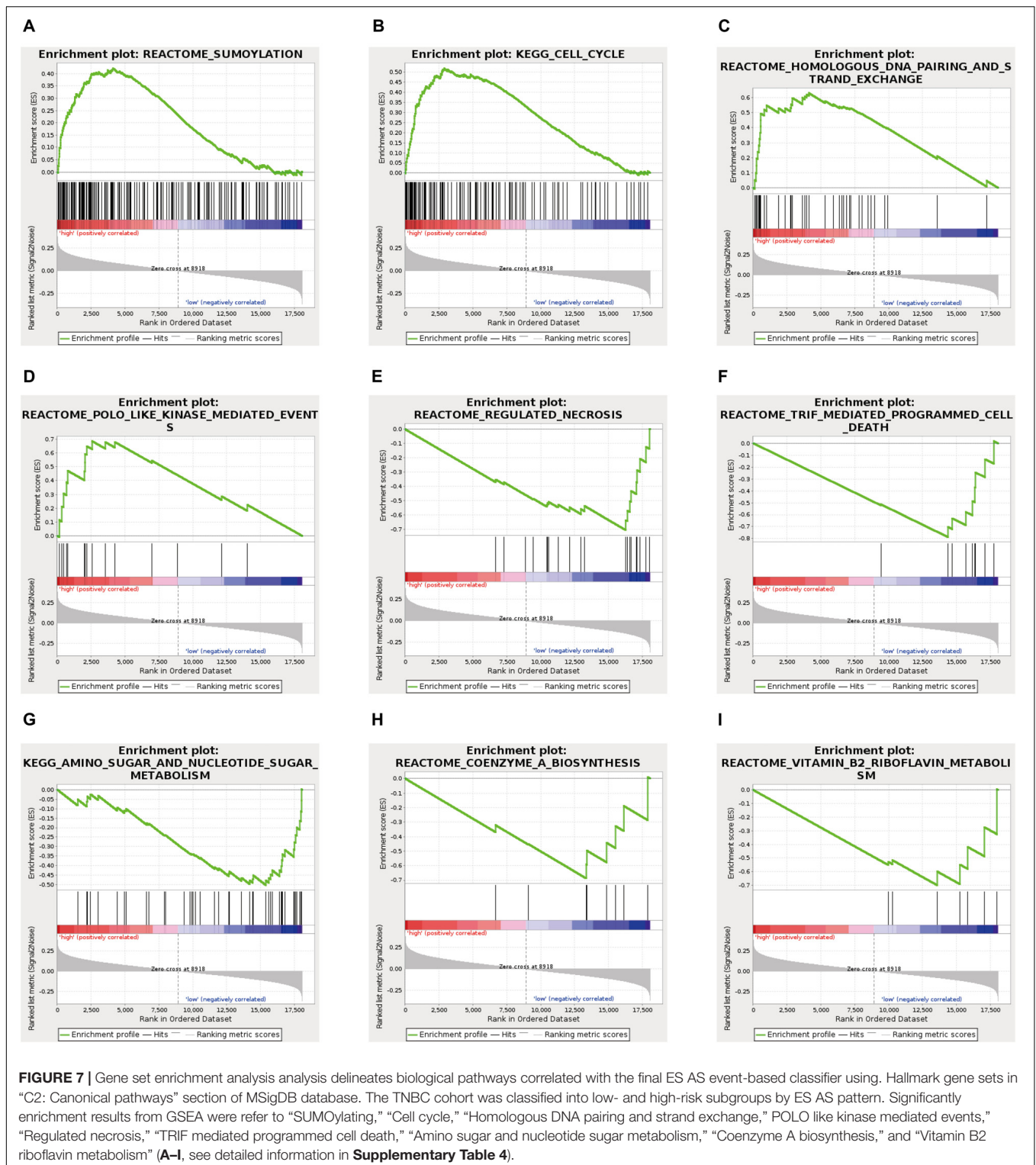


participated in the development of various human malignancies (Yu et al., 2018).

From the gene-interaction network analysis, we found that the genes with survival-associated AS events including HNRNPA1, RPS5, DDX55, and OFD1 were hub nodes of the network which might play vital roles in cancer progress. Recent studies have suggested that HNRNPA1 is involved in the progression and metastasis of several cancers (Chen et al., 2018), including lung, stomach, prostate, and breast cancers (Nadiminty et al., 2015). HNRNPA1 is also a member of the hnRNP family of RNA-binding proteins, which are involved in RNA maturation and translation, as well as pre-mRNA AS (Yang et al., 2019). However, the exact role of AS events in Heterogeneous nuclear ribonucleoprotein A1(HNRNPA1) and its molecular mechanism remains unknown. The hub genes (including HNRNPA1) identified in our gene-interaction network could help to find new clues for the tumorigenesis mechanism of TNBC. GO and KEGG pathway analysis using DAVID were performed to investigate the underlying biological functions for prognosis-associated AS events in TNBC. GO pathway analysis revealed that the genes of prognosis-associated AS events were enriched in focal adhesion, Cul4-RING E3 ubiquitin ligase complex, RNA splicing, and poly(A) RNA binding pathways, and some of which were referred to as the regulator of RNA splicing process (Zhang and Manley, 2013; El Marabti and Younis, 2018). Furthermore, the KEGG

enrichment analysis brought us new insights for elucidating hallmark cancer pathways of TNBC, including Regulation of actin cytoskeleton, Homologous recombination and ECM-receptor interaction which play crucial roles in oncogenesis processes (Wallace et al., 2012). For example, lack of focal adhesion and corruption of ECM-receptor interactions has been regarded as a vital characteristic in most cancers. Additional research concerning how these AS events induced aberrant activation of tumor signaling pathways could be explored in the future.

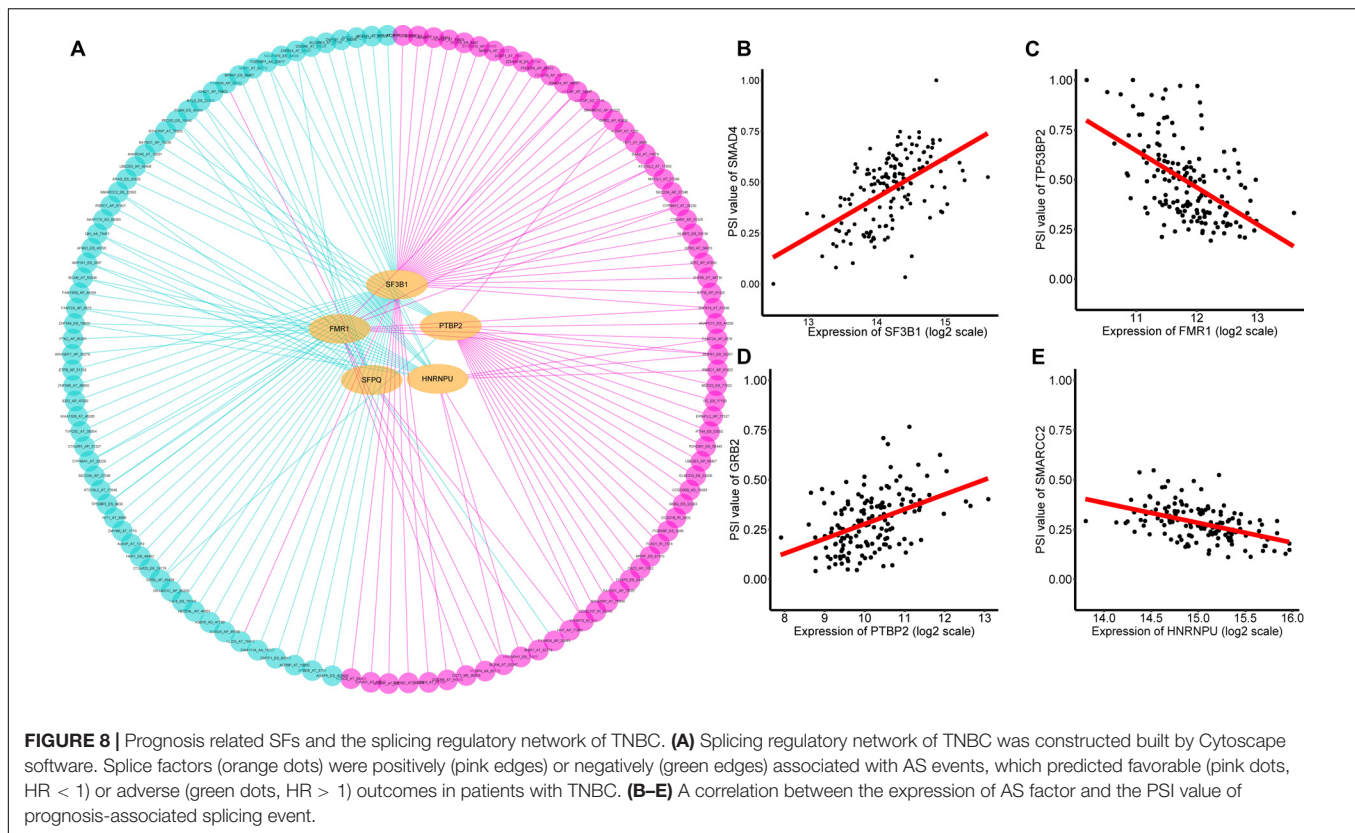
Recently, a prognostic model based on mRNA-lncRNA signature including three mRNA (FCGR1A, RSAD2, CHRDL1), and two lncRNAs (HIF1A-AS2 and AK124454), was constructed to assess the survival and prognosis of TNBC patients (Jiang et al., 2016). However, a prognostic model based on AS events has not yet been reported. Therefore, further analysis on the prognostic models built by single types of seven AS patterns (AA, AD, AP, AT, ES, ME, and RI) was conducted to evaluate the prognosis. The results showed that the prognostic predictor of the ES type exhibited better efficiency to predict the outcome than the other six types of AS prognostic models. In addition, The AUC of ROC from 3 years survival for this high-efficiency prognostic model reach 0.949, providing a more accurate prediction in risk stratification for TNBC patients. GSEA analyses were further employed to reveal specific pathways involved in the ES AS signature between high- and



low-risk groups. Notably, we identified genes in the high-risk group that displayed significant enrichment in cell cycle and Sumoylation pathways while the genes in low-risk group exhibited significant enrichment in programmed cell death, and metabolism pathways. Taken together, this research helps us

to investigate prognostic AS patterns of the TNBC cohort, which could potentially lead to the effective use of prognostic biomarkers in TNBC.

Another notable finding was the crucial roles of the correlation network between prognostic splicing factors and



the significant prognostic-related AS events. In this study, we obtained five splicing factors that were closely associated with the OS of TNBC patients ($p < 0.01$), including Heterogeneous Nuclear Ribonucleoprotein U (HNRNPU), Splicing Factor 3b Subunit 1 (SF3B1), FMRP Translational Regulator 1 (FMR1), Polypyrimidine Tract Binding Protein 2 (PTBP2), and Splicing Factor Proline and Glutamine Rich (SFPQ). Moreover, we observed that expression of PTBP2 had a high correlation with GRB2 AP ($r = 0.44$, $p < 0.001$, **Figure 8D**). Recent studies reported that GRB2 acts as an adaptor protein which plays a central role in the regulation of ARF1 and ARF6 activation in invasive breast cancer (Haines et al., 2014). Additionally, we also found that HNRNPU had significant correlation with SMARCC2 ES ($r = -0.49$, $p < 0.001$, **Figure 8E**). SMARCC2, encoding subunits of Mammalian SWI/SNF chromatin remodeling complexes, is thought to be a core factor in chromosomal rearrangements of chromatin regulators (Kadoch and Crabtree, 2015). Recent studies suggested that abnormalities in SWI/SNF complex subunits play a crucial role in breast cancer (Ribeiro-Silva et al., 2019). Our SF-AS network would further strengthen the association of aberrant AS of ES in SMARCC2 with HNRNPU, which could provide invaluable clues to uncover potential AS prognostic biomarker for TNBC. Collectively, our findings present a better understanding of the underlying mechanism of splicing factors, which could eventually help to elucidate the pathogenic role of AS in TNBC.

CONCLUSION

In summary, we have provided a comprehensive analysis of aberrant AS events in TNBC. Our study constructed an ES AS signature to evaluate the prognostic outcomes, which were found to show a more accurate prediction in risk stratification for TNBC patients. Moreover, the splicing correlation network between prognostic splicing factors and aberrant AS events revealed a strong regulatory mode, which could provide a better understanding of the underlying mechanisms in the TNBC spliceosome. The work achieved in our study could not only contribute to elucidating the tumorigenic mechanism of AS, it could also provide new clues to uncover potential prognostic biomarkers and therapeutic targets for further study.

DATA AVAILABILITY STATEMENT

RNAseq-HTSeq data and clinical data of patients who were diagnosed with primary breast cancer were downloaded from the TCGA (<https://tcga-data.nci.nih.gov/tcga/>). AS profiling of TNBC cohorts were downloaded from TCGA SpliceSeq (<http://bioinformatics.mdanderson.org/TCGASpliceSeq>).

AUTHOR CONTRIBUTIONS

SG and ZS designed the study and wrote the original draft. All authors analyzed and reviewed the data, read and agreed to

the published version of the manuscript. SG, ZS, and DS-L edited the manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fgene.2020.00534/full#supplementary-material>

REFERENCES

- Ashburner, M., Ball, C. A., Blake, J. A., Botstein, D., Butler, H., Cherry, J. M., et al. (2000). Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. *Nat. Genet.* 25, 25–29. doi: 10.1038/75556
- Bianchini, G., Balko, J. M., Mayer, I. A., Sanders, M. E., and Gianni, L. (2016). Triple-negative breast cancer: challenges and opportunities of a heterogeneous disease. *Nat. Rev. Clin. Oncol.* 13, 674–690. doi: 10.1038/nrclinonc.2016.66
- Chen, Y., Liu, J., Wang, W., Xiang, L., Wang, J., Liu, S., et al. (2018). High expression of hnRNP1 promotes cell invasion by inducing EMT in gastric cancer. *Oncol. Rep.* 39, 1693–1701. doi: 10.3892/or.2018.6273
- Dai, X., Xiang, L., Li, T., and Bai, Z. (2016). Cancer hallmarks, biomarkers and breast cancer molecular subtypes. *J. Cancer* 7, 1281–1294. doi: 10.7150/jca.13141
- DeSantis, C. E., Ma, J., Gaudet, M. M., Newman, L. A., Miller, K. D., Goding Sauer, A., et al. (2019). Breast cancer statistics, 2019. *CA Cancer J. Clin.* 69, 438–451. doi: 10.3322/caac.21583
- El Marabti, E., and Younis, I. (2018). The cancer spliceome: reprogramming of alternative splicing in cancer. *Front. Mol. Biosci.* 5:80. doi: 10.3389/fmolb.2018.00080
- Haines, E., Saucier, C., and Claing, A. (2014). The adaptor proteins p66Shc and Grb2 regulate the activation of the GTPases ARF1 and ARF6 in invasive breast cancer cells. *J. Biol. Chem.* 289, 5687–5703. doi: 10.1074/jbc.M113.516047
- Han, J., Li, J., Ho, J. C., Chia, G. S., Kato, H., Jha, S., et al. (2017). Hypoxia is a key driver of alternative splicing in human breast cancer cells. *Sci. Rep.* 7:4108. doi: 10.1038/s41598-017-04333-0
- Huang da, W., Sherman, B. T., and Lempicki, R. A. (2009). Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat. Protoc.* 4, 44–57. doi: 10.1038/nprot.2008.211
- Iorio, E., Caramujo, M. J., Cecchetti, S., Spadaro, F., Carpinelli, G., Canese, R., et al. (2016). Key players in choline metabolic reprogramming in triple-negative breast cancer. *Front. Oncol.* 6:205. doi: 10.3389/fonc.2016.00205
- Jiang, Y. Z., Liu, Y. R., Xu, X. E., Jin, X., Hu, X., Yu, K. D., et al. (2016). Transcriptome analysis of triple-negative breast cancer reveals an integrated mRNA-lncRNA signature with predictive and prognostic value. *Cancer Res.* 76, 2105–2114. doi: 10.1158/0008-5472.CAN-15-3284
- Kadoch, C., and Crabtree, G. R. (2015). Mammalian SWI/SNF chromatin remodeling complexes and cancer: mechanistic insights gained from human genomics. *Sci. Adv.* 1:e1500447. doi: 10.1126/sciadv.1500447
- Kanehisa, M., Furumichi, M., Tanabe, M., Sato, Y., and Morishima, K. (2017). KEGG: new perspectives on genomes, pathways, diseases and drugs. *Nucleic Acids Res.* 45, D353–D361. doi: 10.1093/nar/gkx1092
- Ke, H., Zhao, L., Zhang, H., Feng, X., Xu, H., Hao, J., et al. (2018). Loss of TDP43 inhibits progression of triple-negative breast cancer in coordination with SRSF3. *Proc. Natl. Acad. Sci. U.S.A.* 115, E3426–E3435. doi: 10.1073/pnas.1714573115
- Ke, S., and Chasin, L. A. (2011). Context-dependent splicing regulation: exon definition, co-occurring motif pairs and tissue specificity. *RNA Biol.* 8, 384–388. doi: 10.4161/rna.8.3.14458
- Li, Y., Sun, N., Lu, Z., Sun, S., Huang, J., Chen, Z., et al. (2017). Prognostic alternative mRNA splicing signature in non-small cell lung cancer. *Cancer Lett.* 393, 40–51. doi: 10.1016/j.canlet.2017.02.016
- Li, Z. X., Zheng, Z. Q., Wei, Z. H., Zhang, L. L., Li, F., Lin, L., et al. (2019). Comprehensive characterization of the alternative splicing landscape in head and neck squamous cell carcinoma reveals novel events associated with tumorigenesis and the immune microenvironment. *Theranostics* 9, 7648–7665. doi: 10.7150/tno.36585
- Lin, P., He, R. Q., Ma, F. C., Liang, L., He, Y., Yang, H., et al. (2018). Systematic Analysis of Survival-Associated Alternative Splicing Signatures in Gastrointestinal Pan-Adenocarcinomas. *EBioMedicine* 34, 46–60. doi: 10.1016/j.ebiom.2018.07.040
- Lokody, I. (2014). Alternative splicing: aberrant splicing promotes colon tumour growth. *Nat. Rev. Cancer* 14, 382–383. doi: 10.1038/nrc3753
- Misquitta-Ali, C. M., Cheng, E., O'Hanlon, D., Liu, N., McClade, C. J., Tsao, M. S., et al. (2011). Global profiling and molecular characterization of alternative splicing events misregulated in lung cancer. *Mol. Cell. Biol.* 31, 138–150. doi: 10.1128/MCB.00709-10
- Nadiminty, N., Tummala, R., Liu, C., Lou, W., Evans, C. P., and Gao, A. C. (2015). NF-kappaB2/p52:c-Myc:hnRNP1 pathway regulates expression of androgen receptor splice variants and enzalutamide sensitivity in prostate cancer. *Mol. Cancer Ther.* 14, 1884–1895. doi: 10.1158/1535-7163.MCT-14-1057
- Oltean, S., and Bates, D. O. (2014). Hallmarks of alternative splicing in cancer. *Oncogene* 33, 5311–5318. doi: 10.1038/onc.2013.533
- Onitilo, A. A., Engel, J. M., Greenlee, R. T., and Mukesh, B. N. (2009). Breast cancer subtypes based on ER/PR and Her2 expression: comparison of clinicopathologic features and survival. *Clin. Med. Res.* 7, 4–13. doi: 10.3121/cm.2009.825
- Pan, Q., Shai, O., Lee, L. J., Frey, B. J., and Blencowe, B. J. (2008). Deep surveying of alternative splicing complexity in the human transcriptome by high-throughput sequencing. *Nat. Genet.* 40, 1413–1415. doi: 10.1038/ng.259
- Penault-Llorca, F., and Viale, G. (2012). Pathological and molecular diagnosis of triple-negative breast cancer: a clinical perspective. *Ann. Oncol.* 23(Suppl. 6), vi19–vi22. doi: 10.1093/annonc/mds190
- Piva, F., Giulietti, M., Burini, A. B., and Principato, G. (2012). SpliceAid 2: a database of human splicing factors expression data and RNA target motifs. *Hum. Mutat.* 33, 81–85. doi: 10.1002/humu.21609
- Read, A., and Natrajan, R. (2018). Splicing dysregulation as a driver of breast cancer. *Endocr. Relat. Cancer* 25, R467–R478. doi: 10.1530/erc-18-0068
- Ribeiro-Silva, C., Vermeulen, W., and Lans, H. (2019). SWI/SNF: complex complexes in genome stability and cancer. *DNA Repair* 77, 87–95. doi: 10.1016/j.dnarep.2019.03.007
- Robinson, M. D., McCarthy, D. J., and Smyth, G. K. (2010). edgeR: a bioconductor package for differential expression analysis of digital gene expression data. *Bioinformatics* 26, 139–140. doi: 10.1093/bioinformatics/btp616
- Ryan, M., Wong, W. C., Brown, R., Akbani, R., Su, X., Broom, B., et al. (2016). TCGASpliceSeq a compendium of alternative mRNA splicing in cancer. *Nucleic Acids Res.* 44, D1018–D1022. doi: 10.1093/nar/gkv1288
- Ryan, M. C., Cleland, J., Kim, R., Wong, W. C., and Weinstein, J. N. (2012). SpliceSeq: a resource for analysis and visualization of RNA-Seq data on alternative splicing and its functional impacts. *Bioinformatics* 28, 2385–2387. doi: 10.1093/bioinformatics/bts452
- Scosyrev, E., and Glimm, E. (2019). Power analysis for multivariable Cox regression models. *Stat. Med.* 38, 88–99. doi: 10.1002/sim.7964
- Shannon, P., Markiel, A., Ozier, O., Baliga, N. S., Wang, J. T., Ramage, D., et al. (2003). Cytoscape: a software environment for integrated models of biomolecular interaction networks. *Genome Res.* 13, 2498–2504. doi: 10.1101/gr.1239303

- Shi, Y., Jin, J., Ji, W., and Guan, X. (2018). Therapeutic landscape in mutational triple negative breast cancer. *Mol. Cancer* 17:99. doi: 10.1186/s12943-018-0850-9
- Stirzaker, C., Zotenko, E., and Clark, S. J. (2016). Genome-wide DNA methylation profiling in triple-negative breast cancer reveals epigenetic signatures with important clinical value. *Mol. Cell Oncol.* 3:e1038424. doi: 10.1080/23723556.2015.1038424
- Subramanian, A., Tamayo, P., Mootha, V. K., Mukherjee, S., Ebert, B. L., Gillette, M. A., et al. (2005). Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc. Natl. Acad. Sci. U.S.A.* 102, 15545–15550. doi: 10.1073/pnas.0506580102
- Szklarczyk, D., Gable, A. L., Lyon, D., Junge, A., Wyder, S., Huerta-Cepas, J., et al. (2019). STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. *Nucleic Acids Res.* 47, D607–D613. doi: 10.1093/nar/gky1131
- Wallace, S. S., Murphy, D. L., and Sweasy, J. B. (2012). Base excision repair and cancer. *Cancer Lett.* 327, 73–89. doi: 10.1016/j.canlet.2011.12.038
- Wang, E. T., Sandberg, R., Luo, S., Khrebtkova, I., Zhang, L., Mayr, C., et al. (2008). Alternative isoform regulation in human tissue transcriptomes. *Nature* 456, 470–476. doi: 10.1038/nature07509
- Wang, Y., Liu, J., Huang, B. O., Xu, Y. M., Li, J., Huang, L. F., et al. (2015). Mechanism of alternative splicing and its regulation. *Biomed. Rep.* 3, 152–158. doi: 10.3892/br.2014.407
- Xiong, Y., Deng, Y., Wang, K., Zhou, H., Zheng, X., Si, L., et al. (2018). Profiles of alternative splicing in colorectal cancer and their clinical significance: a study based on large-scale sequencing data. *EBioMedicine* 36, 183–195. doi: 10.1016/j.ebiom.2018.09.021
- Yang, Q., Zhao, J., Zhang, W., Chen, D., and Wang, Y. (2019). Aberrant alternative splicing in breast cancer. *J. Mol. Cell Biol.* 11, 920–929. doi: 10.1093/jmcb/mjz033
- Yu, Y., Liu, D., Liu, Z., Li, S., Ge, Y., Sun, W., et al. (2018). The inhibitory effects of COL1A2 on colorectal cancer cell proliferation, migration, and invasion. *J. Cancer* 9, 2953–2962. doi: 10.7150/jca.25542
- Zhang, D., Duan, Y., Cun, J., and Yang, Q. (2019). Identification of prognostic alternative splicing signature in breast carcinoma. *Front. Genet.* 10:278. doi: 10.3389/fgene.2019.00278
- Zhang, J., and Manley, J. L. (2013). Misregulation of pre-mRNA alternative splicing in cancer. *Cancer Discov.* 3, 1228–1237. doi: 10.1158/2159-8290.CD-13-0253

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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