



NK Cell Exhaustion

Jiacheng Bi^{1*} and Zhigang Tian^{2,3*}

¹ Shenzhen Laboratory of Antibody Engineering, Institute of Biomedicine and Biotechnology, Shenzhen Institutes of Advanced Technology, Chinese Academy of Sciences, Shenzhen, China, ² School of Life Sciences and Medical Center, Institute of Immunology, Key Laboratory of Innate Immunity and Chronic Disease of Chinese Academy of Science, University of Science and Technology of China, Hefei, China, ³ Collaborative Innovation Center for Diagnosis and Treatment of Infectious Diseases, State Key Laboratory for Diagnosis and Treatment of Infectious Diseases, First Affiliated Hospital, College of Medicine, Zhejiang University, Hangzhou, China

Natural killer cells are important effector lymphocytes of the innate immune system, playing critical roles in antitumor and anti-infection host defense. Tumor progression or chronic infections, however, usually leads to exhaustion of NK cells, thus limiting the antitumor/infection potential of NK cells. In many tumors or chronic infections, multiple mechanisms might contribute to the exhaustion of NK cells, such as dysregulated NK cell receptors signaling, as well as suppressive effects by regulatory cells or soluble factors within the microenvironment. Better understanding of the characteristics, as well as the underlying mechanisms of NK cell shut also could reveal novel NK cell-based antitumor/infection targets. Here, we provide an overview of our current knowledge on NK cell exhaustion in tumors, and in chronic infections.

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*Correspondence:

Jiacheng Bi jc.bi@siat.ac.cn; Zhigang Tian tzg@ustc.edu.cn

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INTRODUCTION

Immune cell exhaustion describes the status of dysfunction of immune cells, usually under the settings of tumors or chronic infections (1, 2). Such status, usually associated with poor control of malignancies or infections, is characterized by decreased effector functions (1). For T cells, exhaustion is accompanied by phenotypic changes (1), epigenetic modifications (3), and alterations in transcriptional profiles (4). Multiple negative regulatory pathways (e.g., immunoregulatory cytokines and PD-1) have been shown to be involved in the exhaustion of T cells (1). The in-depth descriptions of the molecular characteristics of T cell exhaustion have not only provided a framework for better understanding T cell biology in these contexts but have also given rise to T cell-based antitumor or anti-infection immunotherapy, which includes immune checkpoint blockade (5) and adoptive T cell therapy (6, 7).

NK cells, as a critical part of the innate immune system, are an important effector lymphocyte population in antitumor and anti-infection immunity (8–10). Evidence supporting its essential roles includes the correlation of poor cytotoxicity of NK cells in the peripheral blood with higher risk of cancer (11). Also, the expression of NKp30 and NKG2D on NK cells from melanoma metastatic lymph node (M-LN) negatively correlated with percentages of tumor cells in M-LN (12). Not only the potentials of NK cells in controlling blood cancers and tumor metastasis have already been widely appreciated but the tumor infiltration of NK cells was also associated with good prognosis in multiple solid tumors (13–17).

However, under the settings of tumors and chronic infections, NK cells exhibit an exhausted status similar with exhausted T cells, displaying poor effector function and altered phenotype. Although the exact mechanisms leading to NK cell exhaustion in tumors and chronic infections are poorly

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defined, emerging studies to be discussed below have shown that multiple negative regulatory pathways in these contexts might contribute to such exhausted status of NK cells, such as dysregulated NK cell receptors signaling, as well as suppressive effects by regulatory cells or soluble factors within the microenvironment. Here, we reviewed current understanding of the characteristics and the mechanisms of NK cell exhaustion, as well as ongoing efforts trying to reverse such state of NK cells.

NK CELL EFFECTOR FUNCTIONS

NK cells mediate antitumor or anti-infection immunity by production of effector cytokines, or by direct cytotoxic activity (8). NK cells are an early and essential source of IFN- γ *in vivo* (18). IFN- γ either directly enhances target cell immunogenicity (19) or facilitates adaptive immunity (20, 21). Besides rapid production of IFN- γ , NK cells also directly eliminate transformed cells or infected cells through cytotoxic activity dependent on perforin and granzyme (22–24), or inducing target cell apoptosis by TNF- α (25), FasL (26), and TRAIL (27). In addition to the effector functions, NK cells also potentiate adaptive immune response through DC editing and maturation (28, 29).

Unlike cytotoxic T cells, NK cells are recombinase independent, and do not need to be primed before effector functions, which makes NK cells a rapid responder in host immunity. Activation of NK cells depends on the integration of activating signals and inhibitory signals from cell surface receptors (30), upon recognition of target cells (31) or interaction with accessory cells (32). Activating receptors include NKG2D, CD16, NCRs, CD226 (DNAM-1), and 2B4, among which, CD16 plays a key role in antibody-dependent cell-mediated cytotoxicity as the $Fc\gamma$ receptor. Inhibitory receptors include self-MHC I-recognizing KIRs in human or Ly49s in mice, NKG2A, TIM-3, TIGIT, and CD96.

CHARACTERISTICS OF NK CELL EXHAUSTION

Exhausted Effector Functions

Despite the potential cytolytic activity of NK cells against tumor cells or infected cells, NK cells exhibited impaired effector functions in hosts with tumors or chronic infections (Figure 1). For example, progression of multiple myeloma in mice was associated with decreased percentages of NK cells (33). At single cell levels, tumor-infiltrating NK cells produced decreased effector cytokines IFN-y and GM-CSF in mouse models (34). NK cells in cancer patients showed diminished cytolytic activity, as evidenced by lower expression of cytolytic molecules, such as granzymes, perforin, FasL, and TRAIL (35). Intratumoral NK cells from patients with various cancers produced decreased IFN-γ (36, 37), CD107a (36, 37), granzyme B (36), and perforin (36) and exhibited impaired cytolytic activity (38), compared with NK cells from peritumor regions or from the peripheral blood. Such exhaustion of NK cell functions seems to be the result of an active process in tumors or chronic infections, since adoptively transferred murine NK cells into mice with leukemia rapidly lost IFN-y production, followed by loss of cytotoxicity after homeostatic proliferation in the presence of tumor (39).

Exhausted Phenotypes

The functional exhaustion of NK cells in tumors and chronic infections is sometimes accompanied with the downregulated expression of certain surface activating receptors on NK cells (Figure 1). NKG2D was frequently downregulated on NK cells in patients with various kinds of malignancies, e.g., pancreatic cancer, gastric cancer, colorectal cancer (35), breast cancer (38), and chronic lymphocytic leukemia (40), as well as in patients with chronic virus infection, such as HBV (41). Compromised NKG2D signaling in this context was also evidenced by downregulation of DAP10, the signaling adaptor of NKG2D (41). Besides NKG2D, CD16 (38), NCRs (NKp30, NKp44, and NKp46) (35, 38, 40-42), CD226 (33, 38, 40, 42, 43), and 2B4 (41) expression on NK cells also usually decreased under settings of tumors or chronic infections. Dysregulated expression of these receptors in patients could be restored in remission (38). Given that NK cell activation result from an integration of activating and inhibitory signals (30), weakened signals from activating receptors might result in the lost of integrated signaling balance toward domination by inhibitory signals, thus gradually inducing NK cell exhaustion.

Another phenotypic signature of NK cell exhaustion is the upregulation of inhibitory receptors (Figure 1). For example, PD-1, as a well-known target in immunotherapy, is a proven checkpoint on T cells. PD-1 overexpression in NK cell line resulted in decreased degranulation, indicating that PD-1 signaling is suppressive not only on T cells but also on NK cells (44). PD-1 was found to be upregulated on NK cells from tumor patients, such as those with Kaposi sarcoma (44), renal cell carcinoma (45), multiple myeloma (46), and EBV-associated posttransplant lymphoproliferative disorders (47). Such upregulation of PD-1 on NK cells was found confined to a subset of CD56dimNKG2A-K IR+CD57+ cells, as reported in HCMV, and ovarian carcinoma patients (48), where such subset was frequently detected (48). Such high PD-1 expression was found associated with reduced proliferative capability in response to cytokines (48), impaired degranulation (44, 48), and poor cytokine production (44, 47, 48) by NK cells. In addition, in renal cell carcinoma, increased PD-1 expression on NK cells in the peripheral blood was correlated to disease stage; the expression was significantly reduced soon after surgical resection of the primary tumor (45). Together, these data indicate that PD-1 is not only associated with the exhausted status of NK cells but PD-1 signaling also might contribute to NK cell exhaustion.

Inhibitory receptor NKG2A was increased on CD56^{dim} NK cells from hepatocellular carcinoma (HCC) patients (36). Such high expression of NKG2A, suggested to be induced by IL-10, contributed to NK cell dysfunction in these patients, correlated with plasma IL-10 levels, and predicted a poor prognosis (36). NKG2A was also expressed on higher percentages of NK cells in active chronic hepatitis B (CHB) patients, than in inactive CHB patients or healthy controls (49). In consistent with this, NK cells from HBV-carrier mice also had higher percentages of NKG2A⁺ NK cells compared with control mice (49). NKG2A was inhibitory on active-CHB patients-derived peripheral NK cell cytotoxicity (49). Importantly, NKG2A-Qa-1 interaction in HBV-carrier mice contributed to HBV persistence (49). These data suggest



NK cell exhaustion to boost antitumor or anti-infection immunity.

that inhibitory receptor NKG2A also might contribute to NK cell exhaustion in tumors and chronic infections.

Tim-3, a novel checkpoint on T cells, was shown to be an inhibitory receptor on NK cells (50) and was upregulated on NK cells from PBMCs of patients with CHB infection (51). Tim-3 suppressed the cytotoxicity of NK cells from these patients (51), suggesting that Tim-3 might be involved in the exhausted cytolytic function of NK cells in these patients.

Exhausted Signaling/Transcriptional Programs

The maintenance of NK cell function relies on an active signaling/ transcriptional program, which seems to be perturbed in tumors or chronic infections, thus contributing to NK cell exhaustion. IL-15 signaling was shown to be defective in tumor-bearing mice, which impeded NK cell maturation and IFN- γ production (52). Overexpression of IL-15 reversed the maturation defects in NK cells (52). Along with the exhaustion of NK cells, the transcriptional factor Eomes and T-bet were downregulated in adoptive transferred murine NK cells into mice with leukemia (39). Enforced expression of Eomes in NK cells partially reversed NK cell dysfunction (39), indicating that the repressed expression of Eomes in NK cells in leukemia is not only an exhaustion-associated transcriptional signature but also at least partially accounted for the exhaustion of NK cells.

IMMUNOREGULATORY PATHWAYS INVOLVED IN NK CELL EXHAUSTION

Detrimental Modulations of Checkpoint Ligands

Together with the dysregulation of NK cell surface receptors, detrimental modulations of ligand expression on transformed

or infected cell surface for NK cell receptors lead to aggravated inhibitory signals, and to dampened activating signals, which might lean the integrated signals balance toward inhibition, and promote the exhaustion of NK cells in tumors or chronic infections. For example, HLA-E, the ligand for inhibitory receptor NKG2A, was more frequently expressed by tumor cells than by normal cells, which aggravated NKG2A-mediated inhibition of NK cell activity (53). CD200, a ligand of NK cell inhibitory receptor CD200R, is upregulated in acute myeloid leukemia and is associated with poor patient outcome (54). Galactin-9, the ligand for human NK cell inhibitory receptor Tim-3 (55), was expressed at high levels on acute myeloid leukemia cells (56). In these examples, upregulated ligand for inhibitory receptor aggravated the inhibitory signaling. Whereas CD48, the ligand for activating receptor 2B4, was reported to be downregulated by oncogenic proteins in acute myeloid leukemia, thus weakening activating signaling from 2B4 (57). Presence of soluble ligand of activating receptors could also weaken activating signaling. Activating receptor GITR ligand (sGITRL) was detected in the sera of patients with various tumors (58). sGITRL or patients' sera that contained sGITRL might block activating receptor GITR interaction with its ligand and were suppressive on NK cell cytotoxicity by negatively regulating NF-KB activity (58). Fag2 protein of Fusobacterium nucleatum, bacteria present in the tumor microenvironment, is a ligand for inhibitory receptor TIGIT (59-63). Faq2 was found able to bind to tumor cells and to mediate inhibition of NK cell activity by triggering TIGIT signaling (59). The abovementioned examples, either aggravating inhibitory signaling or dampening activating signaling, might promote NK cell exhaustion within tumor environment.

Upregulated ligands for NK cell activating receptors by tumor cells, in some cases, could be paradoxically inhibitory on NK cell function, possibly contributing to NK cell exhaustion as well. MICA/B, ligands for human NKG2D, was frequently expressed in both solid tumors (64) and leukemia (65). NK cells from patients with acute myeloid leukemia patients showed downregulated expression of activating receptors, NKp46, NKp30, CD226, 2B4, and CD94/NKG2C (65). Soluble MICA/B also downregulated NKG2D expression on an NK cell line in vitro (64), suggesting that downregulation of activating receptors on NK cells might be a result of chronic exposure of NK cells to tumor cells (65). Similarly, CD112, the ligand for both activating receptor CD226 and inhibitory receptor TIGIT in humans, is frequently expressed at high levels on tumor cells (65). Coculture with CD112expressing leukemic cells in vitro was shown to downregulate CD226 on NK cells (43). In line with this, CD112 expression on leukemic blasts was negatively correlated with CD226 expression on NK cells (43). On the other hand, Rae-1, the murine ligand for NKG2D, when expressed by RMA cells, was reported to be acquired by NK cells, which elicited NK cell fratricide by neighboring NK cells through the NKG2D-induced perforin pathway both *in vitro* and *in vivo* (66). In the cases demonstrated above, upregulated activating ligands, either downregulated expression of activating receptor after NK cell chronic exposure to tumor cells or induced NK cell fratricide, ultimately leading to weakened signaling through activating receptors, which, together with other negative regulatory pathways in tumor microenvironment, might lead to NK cell exhaustion.

Posttranslational alterations of ligands for NK cell surface receptors might also be involved in NK cell exhaustion. For example, posttranslational modifications reduced the affinity of MICA for NKG2D, impairing activating signals from NKG2D (67). Diminished activating signaling might then gradually induce the exhausted status of NK cells in tumor microenvironment.

Regulatory Cells

In addition to modulations of NK cell surface receptor signaling, suppressive immune cells and non-immune cells exist in the microenvironment of tumors or chronic infections, which rendered exhaustion of NK cells (**Figure 1**).

CD4⁺CD25⁺Foxp3⁺ regulatory T cells are a critical subset of T cells that maintain immune tolerance (68–70). Tumor growth or chronic infections promotes the expansion of Tregs (71–73). Treg cells are inhibitory on NKG2D expression on NK cells, and on NK cell cytotoxicity (74). Evidence that Treg cells are related to NK cell exhaustion is that absolute numbers of Treg cells in patients with gastrointestinal stromal tumors inversely correlated with NK cell induction (74).

Tumor growth also promotes the expansion of another suppressive cell type, CD11b⁺Gr1⁺ myeloid-derived suppressor cells (MDSCs) (75). Expansion of MDSCs inversely correlated with activation of NK cells in tumor patients and mice (76), linking MDSCs with NK cell exhaustion. MDSCs suppressed IL-2-mediated NK cell cytotoxicity by affecting the activity of STAT5 in a contact-dependent manner (77). HCV-induced MDSCs suppressed NK cell IFN- γ production *via* an arginase-1-dependent inhibition of mechanistic target of rapamycin (mTOR) activation (78). MDSCs inhibited NKG2D expression on NK cells and suppressed both cytotoxicity and IFN- γ production, both *in vitro* and *in vivo via* membrane-bound TGF- β in mice (76). These data indicate that MDSCs could mediate NK cell exhaustion in tumors and chronic infections through their suppressive functions.

Besides CD11b⁺Gr1⁺ cells, the tumor promoting roles of other myeloid cells (79, 80) have been well accepted, which are potentially suppressive on NK cell functions in tumors as well. For example, in human gastric cancers, tumor-infiltrating monocytes/macrophages were physically close to NK cells, and their percentages were negatively correlated with the percentages of IFN- γ^+ and TNF- α^+ cells among NK cells (16), indicating that these tumor-infiltrating monocytes/macrophages are associated with NK cell exhaustion. Such monocytes/macrophages could impair expression of IFN- γ , TNF- α , and Ki-67 in NK cells in a TGF- β -dependent manner *in vitro* (16), suggesting that these monocytes/macrophages might contribute to the exhaustion of NK cells in tumors.

On the other hand, increasing evidence indicated the NK-suppressive roles of non-immune cells in tumors. Fibroblasts derived from metastatic melanomas mediated inhibition of IL-2-induced upregulation of CD226 on NK cells in a contact-dependent manner and counteracted IL-2-induced upregulation of NKp44 and NKp30 through releasing PGE(2) (81). Fibroblasts from HCC were significantly superior to foreskin-derived fibroblasts at impairing NK cell activation, which was mediated by PGE(2)

and IDO (82). However, the contribution of tumor-derived fibroblasts to NK cell exhaustion in physiological conditions still needs further investigations.

Exosomes

Tumor-associated exosomes are immunoregulatory microvesicles secreted from tumor cells (Figure 1). Breast tumor cellderived exosomes inhibited perforin expression and cytolytic activity ex vivo and in vitro and reduced NK cell percentages in both the lung and spleen of naive mice (83). Tumor cell-derived exosomes suppressed NK cell function by expressing TGF-β, and by expressing ligands for NKG2D to downregulating NKG2D expressed by NK cells (84). Exosomes from the sera of acute myeloid leukemia patients also showed NK-suppressive effects by similar mechanisms (85). Moreover, exosomes from hypoxic tumors showed TGF-β1 and miR-210- and miR-23a-dependent suppression on NK cell function, as compared with exosomes from normoxic tumor cells, in multiple tumor models (86). These studies demonstrated the inhibitory effects of tumorassociated exosomes on NK cells and suggest that they might be involved in the exhausted status of NK cells in these contexts.

Suppressive Cytokines

Suppressive cytokines are important factors that might promote NK cell exhaustion in tumors and in chronic infections (Figure 1). TGF- β was usually detected at high levels in the settings of tumors (87) or chronic infections (41), indicating that TGF- β is highly associated with tumors and chronic infections. Besides, inhibitory effects of TGF-β on NK cells are well documented. Expression of NKG2D was shown to be downregulated by TGF- β (88). TGF- β also induced miR-183 to repress DAP12 transcription, thus suppressing signaling of activating NK cell receptors (89). TGF-\beta-treated human NK cells exhibited decreased cytolytic activity, with abrogated perforin polarization to the immune synapse (89). Breast tumor cell-secreted TGF-β suppressed NK cell expression of activation marker CD69, degranulation marker CD107a, effector cytokines IFN-y and TNF- α , and cytotoxicity against target cells (38). Despite these in vitro studies and correlation analyses, the exact contributions of TGF- β to NK cell exhaustion in physiological conditions have yet to be revealed.

Hypoxia

Other characteristics of tumor microenvironment might contribute to NK cell exhaustion, e.g., hypoxia (**Figure 1**). Within tumor microenvironment, oxygen is usually less available than in normal tissues, which has been long associated with the immune suppressive characteristics of tumors (90). Hypoxia has been shown to induce NK cell suppression. Hypoxia induced hypoxia-inducible factor 1 α in NK cells, decreased expression of NKG2D expression (91), and abrogated the upregulation of NKp46, NKp30, NKp44, and NKG2D in response to activating cytokines (92), thus impairing the capacity of killing target cells (92). Future studies are required to elucidate the role of hypoxia in NK cell exhaustion *in vivo*.

REVERSION OF EXHAUSTION

Blockade of immune checkpoints, as an important part of immunotherapy, proved effective in reversing the exhaustion of T cells to boost antitumor immunity. However, in immunotherapy for tumors and chronic infections, treatment with anti-PD-1 monoclonal antibody alone or with an additional anti-CTLA-4 monoclonal antibody has exhibited clinical benefits only for some patients, suggesting the need for combining extra therapeutics that further counteract the immuno-suppresive mechanisms by tumors or infected cells. In this context, mechanisms that mediate NK cell exhaustion in chronic disorders, as well as pathways for maintaining NK cell self-tolerance, may be targeted to reinvigorate NK cells and provide additionally enhanced immunity (**Figure 1**).

Checkpoint Blockades/Agonisms

First of all, emerging checkpoint blockade strategies are being tested for the potential in reversing NK cell exhaustion in tumors and chronic infections. While blocking KIR, in combination with CTLA-4 blockade, is still under clinical trials for treatment of advanced tumors (NCT01750580), preclinical studies have shown that KIR (or Ly49I/C in mice) blockade boosted NK cell activity in tumor models. Treatment of leukemia-bearing mice with F(ab')₂ of a blocking antibody against mouse Ly49I/C or adoptive transfer of NK cells treated ex vivo with F(ab')2 increased the survival rate (93). F(ab')₂ of anti-Ly49I/C also enhanced the antileukemia activity of anti-huCD20 monoclonal antibody in an EL4-huCD20 leukemia mouse model in an NK-dependent manner (94). A fully human monoclonal antibody, lirilumab, reacts with KIR2DL1/2/3, preventing their binding to HLA-C (95). Administration of lirilumab enhanced the beneficial effect of anti-huCD20 monoclonal antibody, rituximab, on mouse survival in the EL4-huCD20 leukemia model in Rag1KO-KIR Tg mice, whose effect was abrogated when NK cells were depleted (94). Anti-KIR monoclonal antibody and lenalidomide also combined to enhance NK cell versus multiple myeloma effect (96).

Besides KIR, preclinical studies have been revealing more potential NK cell checkpoints. Therapeutic CD94/NKG2A blockade with an anti-NKG2A monoclonal blocking antibody, monalizumab, restored direct cytotoxicity of NK cells against chronic lymphoid leukemia cells in vitro (97). Blocking NKG2A-Qa-1 interaction in vivo in HBV-carrier mice promoted viral clearance in an NK cell-dependent manner (49). CD96, together with activating receptor CD226 and inhibitory receptor TIGIT, constitutes a receptor family that bind nectins and nectin-like family proteins (e.g., CD155 and CD112) and regulates NK cell functions (98). CD96, previously shown to promote human NK cell-target cell adhesion (99), was later revealed to compete with CD226 for CD155 binding and directly inhibits IFN- γ production by NK cells in mice (100). CD96^{-/-} mice displayed resistance to carcinogenesis and experimental lung metastasis (100). Blocking CD96 with a monoclonal antibody inhibited experimental metastases in multiple mouse models (101).

Agonist antibody to augment activating signaling for NK cells is another potent strategy to boost NK cell activity. Administration of anti-4-1BB agonist monoclonal antibody enhanced the expression of CD69 activation marker on NK cells and improved disease control in an NK cell-dependent manner in mice with established multiple myeloma (33).

Checkpoint Modulations

Besides checkpoint inhibitors and agonists, emerging agents potentially reverse NK cell exhaustion by counteracting the detrimental regulation of NK cell receptors or their ligands in the microenvironment of tumors or chronic infections. For example, tumor cell surface expression of NKG2D ligands, either in human or in mice, could be therapeutically upregulated to enhance NK cell activation, by treating tumor cells with alkylating agent (102), proteasome inhibitors (103), hyperploidy-inducing agents (104), histone deacetylation inhibitors (105), or inhibitors for glycogen synthase kinase-3 (106). On the other hand, tumor surface expression of MHC class I might also be downmodulated to increase miss-self recognition-mediated activation of NK cells, e.g., by proteasome inhibitors (107) or PI3K inhibitors (108).

Cytokines

In addition to NK cell surface receptor signaling-targeting/modulating agents, treatment with activating cytokines or blocking the signaling of suppressive cytokines might reverse the NK-disfavoring cytokine milieu that might promote NK cell exhaustion in tumors and chronic infections. Treatment with NK-activating cytokines, IL-12 and IL-18, or with an IL-2 mutant (H9 "superkine") increased survival of MHC-I-deficient tumorbearing mice, accompanied by restoration of effector functions of MHC-I-deficient tumor-infiltrating NK cells (109). Alternatively, another NK-activating cytokine, IL-15 was under clinical trials for treatment of various tumors. In addition, IL-15 fused with the extracellular domain of NKG2D was shown to exhibit enhanced NK cell tumor infiltration and increased suppression of xenografted tumors growth in nude mice infused with human PBMC (110). On the other hand, blockade of TGFBR1 with a small molecule Galunisertib restored TGF-β-induced downregulation of NK cell activating receptors, CD226, NKp30, and NKG2D, and cytolytic molecules, TRAIL, perforin, and granzyme A, increasing direct cytotoxicity and ADCC of ex vivo activated NK cells against neuroblastoma cells in vitro (111).

REMAINING QUESTIONS AND FUTURE DIRECTIONS

In many tumors and chronic infections, the potential of NK cells to produce cytotoxic molecules and effector cytokines is restricted, accompanied with detrimentally modulated expression of surface receptors/ligands, indicating that, similar with T cells, NK cells are also exhausted in these settings. Based on our limited knowledge about NK cell exhaustion, there have been attempts trying to reverse NK cell exhaustion to boost antitumor or anti-infection immunity. However, current barriers in reversing NK cell exhaustion lie beyond the lack of in-depth understanding of such status.

Recent studies suggest that exhausted CD8⁺ T cells in many tumors and chronic infections may represent a distinct lineage,

in that exhausted T cells display multiple signatures distinct with effector T cells and memory T cells (1, 3). On the other hand, the molecular basis of exhausted NK cell in tumors and chronic infections remains largely unexplored.

One feature of exhausted CD8⁺ T cells in tumors and chronic infections is that transcriptional pathways are used differently by exhausted CD8⁺ T cells than by effector and memory CD8⁺ T cells. For example, Blimp-1 is expressed at aberrantly high levels by exhausted CD8⁺ T cells during chronic infections, which promotes the expression of inhibitory receptors and exhaustion (112). Although the transcriptional factors specific for exhausted NK cells are yet to be defined, Eomes and T-bet, essential for the effector function of NK cells (113), were shown to be downregulated in exhausted NK cells in leukemia (39). Overexpression of Eomes restored the exhausted function of NK cells, indicating that downregulated Eomes accounted for the exhausted phenotype of NK cells in this context (39).

Besides transcriptional regulations, cellular metabolism is also important for lymphocyte development and effector function (114–116). Through comparing CD8⁺ T cells in either chronic or acute infections, the exhaustion of CD8⁺ T cells in chronic infections was linked to suppressed bioenergetics, despite persistent signaling of mechanistic target of rapamycin (mTOR) pathway (117). Since activation of mTOR pathway is critical for the effector function of NK cells (118), the exhausted function of NK cells suggests that this pathway in NK cells might be altered in tumors and chronic infections, which remains to be determined.

Emerging evidence appreciates the role of epigenetic regulation in immune cell differentiation and function (119). Exhausted CD8⁺ T cells exhibited stable and state-specific epigenetic modifications distinct from functional memory CD8⁺ T cells (120, 121). The histone-lysine N-methyltransferase, enhancer of zeste homolog 2 (EZH2), was shown to regulate NK cell differentiation and function (122), indicating that epigenetic regulation plays a critical role in shaping NK cell response. However, the epigenetic signature of exhausted NK cells in tumors and chronic infections has not been revealed.

Taken together, the molecular basis, such as transcriptional profiles, epigenetic state, and metabolic regulation, of exhausted NK cells in tumors and chronic infections, as well as the clinical relevance, is still elusive. Future studies are required to reveal novel mechanisms/pathways, e.g., in the aspects mentioned above, hopefully leading to novel targets for NK-based immunotherapy.

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Both JB and ZT conceived and wrote the manuscript.

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REFERENCES

- 1. Wherry EJ. T cell exhaustion. Nat Immunol (2011) 12:492-9. doi:10.1038/ ni.2035
- 2. Joyce JA, Fearon DT. T cell exclusion, immune privilege, and the tumor microenvironment. Science (2015) 348:74-80. doi:10.1126/science.aaa6204
- 3. Wherry EJ, Kurachi M. Molecular and cellular insights into T cell exhaustion. Nat Rev Immunol (2015) 15:486-99. doi:10.1038/nri3862
- 4. Angelosanto JM, Wherry EJ. Transcription factor regulation of CD8+ T-cell memory and exhaustion. Immunol Rev (2010) 236:167-75. doi:10.1111/j. 1600-065X.2010.00927.x
- 5. Sharma P, Allison JP. The future of immune checkpoint therapy. Science (2015) 348:56-61. doi:10.1126/science.aaa8172
- 6. Lim WA, June CH. The principles of engineering immune cells to treat cancer. Cell (2017) 168:724-40. doi:10.1016/j.cell.2017.01.016
- 7. Rosenberg SA, Restifo NP. Adoptive cell transfer as personalized immunotherapy for human cancer. Science (2015) 348:62-8. doi:10.1126/science. aaa4967
- 8. Vivier E, Tomasello E, Baratin M, Walzer T, Ugolini S. Functions of natural killer cells. Nat Immunol (2008) 9:503-10. doi:10.1038/ni1582
- 9. Herberman RB, Nunn ME, Holden HT, Lavrin DH. Natural cytotoxic reactivity of mouse lymphoid cells against syngeneic and allogeneic tumors. II. Characterization of effector cells. Int J Cancer (1975) 16:230-9. doi:10.1002/ iic 2910160204
- 10. Kiessling R, Klein E, Wigzell H. "Natural" killer cells in the mouse. I. Cytotoxic cells with specificity for mouse Moloney leukemia cells. Specificity and distribution according to genotype. Eur J Immunol (1975) 5:112-7. doi:10.1002/eji.1830050208
- 11. Vivier E, Ugolini S, Blaise D, Chabannon C, Brossay L. Targeting natural killer cells and natural killer T cells in cancer. Nat Rev Immunol (2012) 12:239-52. doi:10.1038/nri3174
- 12. Messaoudene M, Fregni G, Fourmentraux-Neves E, Chanal J, Maubec E, Mazouz-Dorval S, et al. Mature cytotoxic CD56(bright)/CD16(+) natural killer cells can infiltrate lymph nodes adjacent to metastatic melanoma. Cancer Res (2014) 74:81-92. doi:10.1158/0008-5472.CAN-13-1303
- 13. Coca S, Perez-Piqueras J, Martinez D, Colmenarejo A, Saez MA, Vallejo C, et al. The prognostic significance of intratumoral natural killer cells in patients with colorectal carcinoma. Cancer (1997) 79:2320-8. doi:10.1002/ (SICI)1097-0142(19970615)79:12<2320::AID-CNCR5>3.0.CO;2-P
- 14. Ishigami S, Natsugoe S, Tokuda K, Nakajo A, Xiangming C, Iwashige H, et al. Clinical impact of intratumoral natural killer cell and dendritic cell infiltration in gastric cancer. Cancer Lett (2000) 159:103-8. doi:10.1016/ \$0304-3835(00)00542-5
- 15. Villegas FR, Coca S, Villarrubia VG, Jimenez R, Chillon MJ, Jareno J, et al. Prognostic significance of tumor infiltrating natural killer cells subset CD57 in patients with squamous cell lung cancer. Lung Cancer (2002) 35:23-8. doi:10.1016/S0169-5002(01)00292-6
- 16. Peng LS, Zhang JY, Teng YS, Zhao YL, Wang TT, Mao FY, et al. Tumorassociated monocytes/macrophages impair NK-cell function via TGFbeta1 in human gastric cancer. Cancer Immunol Res (2017) 5:248-56. doi:10.1158/ 2326-6066.CIR-16-0152
- 17. Jin S, Deng Y, Hao JW, Li Y, Liu B, Yu Y, et al. NK cell phenotypic modulation in lung cancer environment. PLoS One (2014) 9:e109976. doi:10.1371/ journal.pone.0109976
- 18. Stetson DB, Mohrs M, Reinhardt RL, Baron JL, Wang ZE, Gapin L, et al. Constitutive cytokine mRNAs mark natural killer (NK) and NKT cells poised for rapid effector function. J Exp Med (2003) 198:1069-76. doi:10.1084/ jem.20030630
- Kaplan DH, Shankaran V, Dighe AS, Stockert E, Aguet M, Old LJ, et al. 19. Demonstration of an interferon gamma-dependent tumor surveillance system in immunocompetent mice. Proc Natl Acad Sci USA (1998) 95:7556-61. doi:10.1073/pnas.95.13.7556
- 20. Mocikat R, Braumuller H, Gumy A, Egeter O, Ziegler H, Reusch U, et al. Natural killer cells activated by MHC class I(low) targets prime dendritic cells to induce protective CD8 T cell responses. Immunity (2003) 19:561-9. doi:10.1016/S1074-7613(03)00264-4
- 21. Martin-Fontecha A, Thomsen LL, Brett S, Gerard C, Lipp M, Lanzavecchia A, et al. Induced recruitment of NK cells to lymph nodes provides IFN-gamma for T(H)1 priming. Nat Immunol (2004) 5:1260-5. doi:10.1038/ni1138

- 22. Trapani JA, Smyth MJ. Functional significance of the perforin/granzyme cell death pathway. Nat Rev Immunol (2002) 2:735-47. doi:10.1038/ nri911
- 23. van den Broek MF, Kagi D, Zinkernagel RM, Hengartner H. Perforin dependence of natural killer cell-mediated tumor control in vivo. Eur I Immunol (1995) 25:3514-6. doi:10.1002/eii.1830251246
- 24. Smyth MJ, Thia KY, Cretney E, Kelly JM, Snook MB, Forbes CA, et al. Perforin is a major contributor to NK cell control of tumor metastasis. J Immunol (1999) 162:6658-62.
- 25. Wajant H, Pfizenmaier K, Scheurich P. Tumor necrosis factor signaling. Cell Death Differ (2003) 10:45-65. doi:10.1038/sj.cdd.4401189
- 26. Arase H, Arase N, Saito T. Fas-mediated cytotoxicity by freshly isolated natural killer cells. J Exp Med (1995) 181:1235-8. doi:10.1084/jem.181.3.1235
- 27. Kayagaki N, Yamaguchi N, Nakayama M, Takeda K, Akiba H, Tsutsui H, et al. Expression and function of TNF-related apoptosis-inducing ligand on murine activated NK cells. J Immunol (1999) 163:1906-13.
- 28. Moretta L, Ferlazzo G, Bottino C, Vitale M, Pende D, Mingari MC, et al. Effector and regulatory events during natural killer-dendritic cell interactions. Immunol Rev (2006) 214:219-28. doi:10.1111/j.1600-065X.2006.00450.x
- 29. Ferlazzo G, Morandi B. Cross-talks between natural killer cells and distinct subsets of dendritic cells. Front Immunol (2014) 5:159. doi:10.3389/ fimmu.2014.00159
- 30. Long EO, Kim HS, Liu D, Peterson ME, Rajagopalan S. Controlling natural killer cell responses: integration of signals for activation and inhibition. Annu Rev Immunol (2013) 31:227-58. doi:10.1146/annurevimmunol-020711-075005
- 31. Lanier LL. NK cell recognition. Annu Rev Immunol (2005) 23:225-74. doi:10.1146/annurev.immunol.23.021704.115526
- 32. Newman KC, Riley EM. Whatever turns you on: accessory-cell-dependent activation of NK cells by pathogens. Nat Rev Immunol (2007) 7:279-91. doi:10.1038/nri2057
- 33. Guillerey C, Ferrari de Andrade L, Vuckovic S, Miles K, Ngiow SF, Yong MC, et al. Immunosurveillance and therapy of multiple myeloma are CD226 dependent. J Clin Invest (2015) 125:2077-89. doi:10.1172/JCI77181
- 34. Paul S, Kulkarni N, Shilpi, Lal G. Intratumoral natural killer cells show reduced effector and cytolytic properties and control the differentiation of effector Th1 cells. Oncoimmunology (2016) 5:e1235106. doi:10.1080/2162402X. 2016.1235106
- 35. Peng YP, Zhu Y, Zhang JJ, Xu ZK, Qian ZY, Dai CC, et al. Comprehensive analysis of the percentage of surface receptors and cytotoxic granules positive natural killer cells in patients with pancreatic cancer, gastric cancer, and colorectal cancer. J Transl Med (2013) 11:262. doi:10.1186/1479-5876-11-262
- 36. Sun C, Xu J, Huang Q, Huang M, Wen H, Zhang C, et al. High NKG2A expression contributes to NK cell exhaustion and predicts a poor prognosis of patients with liver cancer. Oncoimmunology (2017) 6:e1264562. doi:10.1080/ 2162402X.2016.1264562
- 37. Platonova S, Cherfils-Vicini J, Damotte D, Crozet L, Vieillard V, Validire P, et al. Profound coordinated alterations of intratumoral NK cell phenotype and function in lung carcinoma. Cancer Res (2011) 71:5412-22. doi:10.1158/0008-5472.CAN-10-4179
- 38. Mamessier E, Sylvain A, Thibult ML, Houvenaeghel G, Jacquemier J, Castellano R, et al. Human breast cancer cells enhance self tolerance by promoting evasion from NK cell antitumor immunity. J Clin Invest (2011) 121:3609-22. doi:10.1172/JCI45816
- 39. Gill S, Vasey AE, De Souza A, Baker J, Smith AT, Kohrt HE, et al. Rapid development of exhaustion and down-regulation of eomesodermin limit the antitumor activity of adoptively transferred murine natural killer cells. Blood (2012) 119:5758-68. doi:10.1182/blood-2012-03-415364
- 40. Parry HM, Stevens T, Oldreive C, Zadran B, McSkeane T, Rudzki Z, et al. NK cell function is markedly impaired in patients with chronic lymphocytic leukaemia but is preserved in patients with small lymphocytic lymphoma. Oncotarget (2016) 7:68513-26. doi:10.18632/oncotarget.12097
- 41. Sun C, Fu B, Gao Y, Liao X, Sun R, Tian Z, et al. TGF-beta1 down-regulation of NKG2D/DAP10 and 2B4/SAP expression on human NK cells contributes to HBV persistence. PLoS Pathog (2012) 8:e1002594. doi:10.1371/journal. ppat.1002594
- 42. Bozzano F, Picciotto A, Costa P, Marras F, Fazio V, Hirsch I, et al. Activating NK cell receptor expression/function (NKp30, NKp46, DNAM-1)

during chronic viraemic HCV infection is associated with the outcome of combined treatment. *Eur J Immunol* (2011) 41:2905–14. doi:10.1002/eji.201041361

- Sanchez-Correa B, Gayoso I, Bergua JM, Casado JG, Morgado S, Solana R, et al. Decreased expression of DNAM-1 on NK cells from acute myeloid leukemia patients. *Immunol Cell Biol* (2012) 90:109–15. doi:10.1038/ icb.2011.15
- Beldi-Ferchiou A, Lambert M, Dogniaux S, Vely F, Vivier E, Olive D, et al. PD-1 mediates functional exhaustion of activated NK cells in patients with Kaposi sarcoma. *Oncotarget* (2016) 7:72961–77. doi:10.18632/ oncotarget.12150
- 45. MacFarlane AW, Jillab M, Plimack ER, Hudes GR, Uzzo RG, Litwin S, et al. PD-1 expression on peripheral blood cells increases with stage in renal cell carcinoma patients and is rapidly reduced after surgical tumor resection. *Cancer Immunol Res* (2014) 2:320–31. doi:10.1158/2326-6066. CIR-13-0133
- 46. Benson DM Jr, Bakan CE, Mishra A, Hofmeister CC, Efebera Y, Becknell B, et al. The PD-1/PD-L1 axis modulates the natural killer cell versus multiple myeloma effect: a therapeutic target for CT-011, a novel monoclonal anti-PD-1 antibody. *Blood* (2010) 116:2286–94. doi:10.1182/blood-2010-02-271874
- Wiesmayr S, Webber SA, Macedo C, Popescu I, Smith L, Luce J, et al. Decreased NKp46 and NKG2D and elevated PD-1 are associated with altered NK-cell function in pediatric transplant patients with PTLD. *Eur J Immunol* (2012) 42:541–50. doi:10.1002/eji.201141832
- Pesce S, Greppi M, Tabellini G, Rampinelli F, Parolini S, Olive D, et al. Identification of a subset of human natural killer cells expressing high levels of programmed death 1: a phenotypic and functional characterization. J Allergy Clin Immunol (2017) 139:335–46.e3. doi:10.1016/j.jaci. 2016.04.025
- Li F, Wei H, Wei H, Gao Y, Xu L, Yin W, et al. Blocking the natural killer cell inhibitory receptor NKG2A increases activity of human natural killer cells and clears hepatitis B virus infection in mice. *Gastroenterology* (2013) 144:392–401. doi:10.1053/j.gastro.2012.10.039
- Li YH, Zhou WH, Tao Y, Wang SC, Jiang YL, Zhang D, et al. The Galectin-9/ Tim-3 pathway is involved in the regulation of NK cell function at the maternal-fetal interface in early pregnancy. *Cell Mol Immunol* (2016) 13:73–81. doi:10.1038/cmi.2014.126
- Ju Y, Hou N, Meng J, Wang X, Zhang X, Zhao D, et al. T cell immunoglobulinand mucin-domain-containing molecule-3 (Tim-3) mediates natural killer cell suppression in chronic hepatitis B. *J Hepatol* (2010) 52:322–9. doi:10.1016/j.jhep.2009.12.005
- Richards JO, Chang X, Blaser BW, Caligiuri MA, Zheng P, Liu Y. Tumor growth impedes natural-killer-cell maturation in the bone marrow. *Blood* (2006) 108:246–52. doi:10.1182/blood-2005-11-4535
- Lo Monaco E, Tremante E, Cerboni C, Melucci E, Sibilio L, Zingoni A, et al. Human leukocyte antigen E contributes to protect tumor cells from lysis by natural killer cells. *Neoplasia* (2011) 13:822–30. doi:10.1593/neo.101684
- Coles SJ, Wang EC, Man S, Hills RK, Burnett AK, Tonks A, et al. CD200 expression suppresses natural killer cell function and directly inhibits patient anti-tumor response in acute myeloid leukemia. *Leukemia* (2011) 25:792–9. doi:10.1038/leu.2011.1
- Ndhlovu LC, Lopez-Verges S, Barbour JD, Jones RB, Jha AR, Long BR, et al. Tim-3 marks human natural killer cell maturation and suppresses cell-mediated cytotoxicity. *Blood* (2012) 119:3734–43. doi:10.1182/ blood-2011-11-392951
- Gleason MK, Lenvik TR, McCullar V, Felices M, O'Brien MS, Cooley SA, et al. Tim-3 is an inducible human natural killer cell receptor that enhances interferon gamma production in response to galectin-9. *Blood* (2012) 119:3064–72. doi:10.1182/blood-2011-06-360321
- Elias S, Yamin R, Golomb L, Tsukerman P, Stanietsky-Kaynan N, Ben-Yehuda D, et al. Immune evasion by oncogenic proteins of acute myeloid leukemia. *Blood* (2014) 123:1535–43. doi:10.1182/blood-2013-09-526590
- Baltz KM, Krusch M, Baessler T, Schmiedel BJ, Bringmann A, Brossart P, et al. Neutralization of tumor-derived soluble glucocorticoid-induced TNFRrelated protein ligand increases NK cell anti-tumor reactivity. *Blood* (2008) 112:3735–43. doi:10.1182/blood-2008-03-143016
- 59. Gur C, Ibrahim Y, Isaacson B, Yamin R, Abed J, Gamliel M, et al. Binding of the Fap2 protein of *Fusobacterium nucleatum* to human inhibitory receptor

TIGIT protects tumors from immune cell attack. *Immunity* (2015) 42:344–55. doi:10.1016/j.immuni.2015.01.010

- Stanietsky N, Simic H, Arapovic J, Toporik A, Levy O, Novik A, et al. The interaction of TIGIT with PVR and PVRL2 inhibits human NK cell cytotoxicity. *Proc Natl Acad Sci U S A* (2009) 106:17858–63. doi:10.1073/ pnas.0903474106
- Stanietsky N, Rovis TL, Glasner A, Seidel E, Tsukerman P, Yamin R, et al. Mouse TIGIT inhibits NK-cell cytotoxicity upon interaction with PVR. *Eur J Immunol* (2013) 43:2138–50. doi:10.1002/eji.201243072
- 62. Li M, Xia P, Du Y, Liu S, Huang G, Chen J, et al. T-cell immunoglobulin and ITIM domain (TIGIT) receptor/poliovirus receptor (PVR) ligand engagement suppresses interferon-gamma production of natural killer cells via beta-arrestin 2-mediated negative signaling. *J Biol Chem* (2014) 289:17647–57. doi:10.1074/jbc.M114.572420
- Bi J, Zhang Q, Liang D, Xiong L, Wei H, Sun R, et al. T-cell Ig and ITIM domain regulates natural killer cell activation in murine acute viral hepatitis. *Hepatology* (2014) 59:1715–25. doi:10.1002/hep.26968
- Ren J, Nie Y, Lv M, Shen S, Tang R, Xu Y, et al. Estrogen upregulates MICA/B expression in human non-small cell lung cancer through the regulation of ADAM17. *Cell Mol Immunol* (2015) 12:768–76. doi:10.1038/cmi.2014.101
- 65. Sanchez-Correa B, Morgado S, Gayoso I, Bergua JM, Casado JG, Arcos MJ, et al. Human NK cells in acute myeloid leukaemia patients: analysis of NK cell-activating receptors and their ligands. *Cancer Immunol Immunother* (2011) 60:1195–205. doi:10.1007/s00262-011-1050-2
- Nakamura K, Nakayama M, Kawano M, Amagai R, Ishii T, Harigae H, et al. Fratricide of natural killer cells dressed with tumor-derived NKG2D ligand. *Proc Natl Acad Sci U S A* (2013) 110:9421–6. doi:10.1073/pnas.1300140110
- Tsuboi S, Sutoh M, Hatakeyama S, Hiraoka N, Habuchi T, Horikawa Y, et al. A novel strategy for evasion of NK cell immunity by tumours expressing core2 O-glycans. *EMBO J* (2011) 30:3173–85. doi:10.1038/emboj. 2011.215
- Ohkura N, Kitagawa Y, Sakaguchi S. Development and maintenance of regulatory T cells. *Immunity* (2013) 38:414–23. doi:10.1016/j.immuni.2013.03.002
- Li Z, Li D, Tsun A, Li B. FOXP3+ regulatory T cells and their functional regulation. *Cell Mol Immunol* (2015) 12:558–65. doi:10.1038/cmi.2015.10
- Liu ZM, Wang KP, Ma J, Guo Zheng S. The role of all-trans retinoic acid in the biology of Foxp3+ regulatory T cells. *Cell Mol Immunol* (2015) 12:553–7. doi:10.1038/cmi.2014.133
- Hori S, Nomura T, Sakaguchi S. Control of regulatory T cell development by the transcription factor Foxp3. *Science* (2003) 299:1057–61. doi:10.1126/ science.1079490
- 72. Beyer M, Kochanek M, Darabi K, Popov A, Jensen M, Endl E, et al. Reduced frequencies and suppressive function of CD4+CD25hi regulatory T cells in patients with chronic lymphocytic leukemia after therapy with fludarabine. *Blood* (2005) 106:2018–25. doi:10.1182/blood-2005-02-0642
- Zhai N, Chi X, Li T, Song H, Li H, Jin X, et al. Hepatitis C virus core protein triggers expansion and activation of CD4(+)CD25(+) regulatory T cells in chronic hepatitis C patients. *Cell Mol Immunol* (2015) 12:743–9. doi:10.1038/ cmi.2014.119
- Ghiringhelli F, Menard C, Terme M, Flament C, Taieb J, Chaput N, et al. CD4+CD25+ regulatory T cells inhibit natural killer cell functions in a transforming growth factor-beta-dependent manner. *J Exp Med* (2005) 202:1075–85. doi:10.1084/jem.20051511
- Gabrilovich DI. Myeloid-derived suppressor cells. Cancer Immunol Res (2017) 5:3–8. doi:10.1158/2326-6066.CIR-16-0297
- Li H, Han Y, Guo Q, Zhang M, Cao X. Cancer-expanded myeloid-derived suppressor cells induce anergy of NK cells through membrane-bound TGFbeta 1. *J Immunol* (2009) 182:240–9. doi:10.4049/jimmunol.182.1.240
- Liu C, Yu S, Kappes J, Wang J, Grizzle WE, Zinn KR, et al. Expansion of spleen myeloid suppressor cells represses NK cell cytotoxicity in tumor-bearing host. *Blood* (2007) 109:4336–42. doi:10.1182/blood-2006-09-046201
- Goh CC, Roggerson KM, Lee HC, Golden-Mason L, Rosen HR, Hahn YS. Hepatitis C virus-induced myeloid-derived suppressor cells suppress NK cell IFN-gamma production by altering cellular metabolism via arginase-1. *J Immunol* (2016) 196:2283–92. doi:10.4049/jimmunol.1501881
- Liu Y, Cao X. Intratumoral dendritic cells in the anti-tumor immune response. Cell Mol Immunol (2015) 12:387–90. doi:10.1038/cmi.2014.130
- Liu Y, Cao X. The origin and function of tumor-associated macrophages. *Cell Mol Immunol* (2015) 12:1–4. doi:10.1038/cmi.2014.83

- Balsamo M, Scordamaglia F, Pietra G, Manzini C, Cantoni C, Boitano M, et al. Melanoma-associated fibroblasts modulate NK cell phenotype and antitumor cytotoxicity. *Proc Natl Acad Sci U S A* (2009) 106:20847–52. doi:10.1073/pnas.0906481106
- Li T, Yang Y, Hua X, Wang G, Liu W, Jia C, et al. Hepatocellular carcinoma-associated fibroblasts trigger NK cell dysfunction via PGE2 and IDO. *Cancer Lett* (2012) 318:154–61. doi:10.1016/j.canlet.2011.12.020
- Liu C, Yu S, Zinn K, Wang J, Zhang L, Jia Y, et al. Murine mammary carcinoma exosomes promote tumor growth by suppression of NK cell function. *J Immunol* (2006) 176:1375–85. doi:10.4049/jimmunol.176.3.1375
- Clayton A, Mitchell JP, Court J, Linnane S, Mason MD, Tabi Z. Human tumor-derived exosomes down-modulate NKG2D expression. *J Immunol* (2008) 180:7249–58. doi:10.4049/jimmunol.180.11.7249
- Szczepanski MJ, Szajnik M, Welsh A, Whiteside TL, Boyiadzis M. Blastderived microvesicles in sera from patients with acute myeloid leukemia suppress natural killer cell function via membrane-associated transforming growth factor-beta1. *Haematologica* (2011) 96:1302–9. doi:10.3324/haematol. 2010.039743
- Berchem G, Noman MZ, Bosseler M, Paggetti J, Baconnais S, Le Cam E, et al. Hypoxic tumor-derived microvesicles negatively regulate NK cell function by a mechanism involving TGF-beta and miR23a transfer. *Oncoimmunology* (2016) 5:e1062968. doi:10.1080/2162402X.2015.1062968
- Dong M, Blobe GC. Role of transforming growth factor-beta in hematologic malignancies. *Blood* (2006) 107:4589–96. doi:10.1182/blood-2005-10-4169
- Castriconi R, Cantoni C, Della Chiesa M, Vitale M, Marcenaro E, Conte R, et al. Transforming growth factor beta 1 inhibits expression of NKp30 and NKG2D receptors: consequences for the NK-mediated killing of dendritic cells. *Proc Natl Acad Sci U S A* (2003) 100:4120–5. doi:10.1073/ pnas.0730640100
- Donatelli SS, Zhou JM, Gilvary DL, Eksioglu EA, Chen X, Cress WD, et al. TGF-beta-inducible microRNA-183 silences tumor-associated natural killer cells. *Proc Natl Acad Sci U S A* (2014) 111:4203–8. doi:10.1073/pnas. 1319269111
- Palazon A, Goldrath AW, Nizet V, Johnson RS. HIF transcription factors, inflammation, and immunity. *Immunity* (2014) 41:518–28. doi:10.1016/j. immuni.2014.09.008
- Sarkar S, Germeraad WT, Rouschop KM, Steeghs EM, van Gelder M, Bos GM, et al. Hypoxia induced impairment of NK cell cytotoxicity against multiple myeloma can be overcome by IL-2 activation of the NK cells. *PLoS* One (2013) 8:e64835. doi:10.1371/journal.pone.0064835
- Balsamo M, Manzini C, Pietra G, Raggi F, Blengio F, Mingari MC, et al. Hypoxia downregulates the expression of activating receptors involved in NK-cell-mediated target cell killing without affecting ADCC. *Eur J Immunol* (2013) 43:2756–64. doi:10.1002/eji.201343448
- Koh CY, Blazar BR, George T, Welniak LA, Capitini CM, Raziuddin A, et al. Augmentation of antitumor effects by NK cell inhibitory receptor blockade in vitro and in vivo. *Blood* (2001) 97:3132–7. doi:10.1182/blood. V97.10.3132
- 94. Kohrt HE, Thielens A, Marabelle A, Sagiv-Barfi I, Sola C, Chanuc F, et al. Anti-KIR antibody enhancement of anti-lymphoma activity of natural killer cells as monotherapy and in combination with anti-CD20 antibodies. *Blood* (2014) 123:678–86. doi:10.1182/blood-2013-08-519199
- 95. Romagne F, Andre P, Spee P, Zahn S, Anfossi N, Gauthier L, et al. Preclinical characterization of 1-7F9, a novel human anti-KIR receptor therapeutic antibody that augments natural killer-mediated killing of tumor cells. *Blood* (2009) 114:2667–77. doi:10.1182/blood-2009-02-206532
- Benson DM Jr, Bakan CE, Zhang S, Collins SM, Liang J, Srivastava S, et al. IPH2101, a novel anti-inhibitory KIR antibody, and lenalidomide combine to enhance the natural killer cell versus multiple myeloma effect. *Blood* (2011) 118:6387–91. doi:10.1182/blood-2011-06-360255
- McWilliams EM, Mele JM, Cheney C, Timmerman EA, Fiazuddin F, Strattan EJ, et al. Therapeutic CD94/NKG2A blockade improves natural killer cell dysfunction in chronic lymphocytic leukemia. *Oncoimmunology* (2016) 5:e1226720. doi:10.1080/2162402X.2016.1226720
- Martinet L, Smyth MJ. Balancing natural killer cell activation through paired receptors. *Nat Rev Immunol* (2015) 15:243–54. doi:10.1038/nri3799
- Fuchs A, Cella M, Giurisato E, Shaw AS, Colonna M. Cutting edge: CD96 (tactile) promotes NK cell-target cell adhesion by interacting with the

poliovirus receptor (CD155). J Immunol (2004) 172:3994-8. doi:10.4049/ jimmunol.172.7.3994

- 100. Chan CJ, Martinet L, Gilfillan S, Souza-Fonseca-Guimaraes F, Chow MT, Town L, et al. The receptors CD96 and CD226 oppose each other in the regulation of natural killer cell functions. *Nat Immunol* (2014) 15:431–8. doi:10.1038/ni.2850
- 101. Blake SJ, Stannard K, Liu J, Allen S, Yong MC, Mittal D, et al. Suppression of metastases using a new lymphocyte checkpoint target for cancer immunotherapy. *Cancer Discov* (2016) 6:446–59. doi:10.1158/2159-8290. CD-15-0944
- 102. Antonangeli F, Soriani A, Ricci B, Ponzetta A, Benigni G, Morrone S, et al. Natural killer cell recognition of in vivo drug-induced senescent multiple myeloma cells. *Oncoimmunology* (2016) 5:e1218105. doi:10.1080/21624 02X.2016.1218105
- 103. Niu C, Jin H, Li M, Zhu S, Zhou L, Jin F, et al. Low-dose bortezomib increases the expression of NKG2D and DNAM-1 ligands and enhances induced NK and gammadelta T cell-mediated lysis in multiple myeloma. *Oncotarget* (2017) 8:5954–64. doi:10.18632/oncotarget.13979
- 104. Acebes-Huerta A, Lorenzo-Herrero S, Folgueras AR, Huergo-Zapico L, Lopez-Larrea C, Lopez-Soto A, et al. Drug-induced hyperploidy stimulates an antitumor NK cell response mediated by NKG2D and DNAM-1 receptors. *Oncoimmunology* (2016) 5:e1074378. doi:10.1080/2162402X.2015. 1074378
- 105. Schmudde M, Braun A, Pende D, Sonnemann J, Klier U, Beck JF, et al. Histone deacetylase inhibitors sensitize tumour cells for cytotoxic effects of natural killer cells. *Cancer Lett* (2008) 272:110–21. doi:10.1016/j.canlet. 2008.06.027
- 106. Fionda C, Malgarini G, Soriani A, Zingoni A, Cecere F, Iannitto ML, et al. Inhibition of glycogen synthase kinase-3 increases NKG2D ligand MICA expression and sensitivity to NK cell-mediated cytotoxicity in multiple myeloma cells: role of STAT3. *J Immunol* (2013) 190:6662–72. doi:10.4049/ jimmunol.1201426
- 107. Shi J, Tricot GJ, Garg TK, Malaviarachchi PA, Szmania SM, Kellum RE, et al. Bortezomib down-regulates the cell-surface expression of HLA class I and enhances natural killer cell-mediated lysis of myeloma. *Blood* (2008) 111:1309–17. doi:10.1182/blood-2007-03-078535
- 108. Bommarito D, Martin A, Forcade E, Nastke MD, Ritz J, Bellucci R. Enhancement of tumor cell susceptibility to natural killer cell activity through inhibition of the PI3K signaling pathway. *Cancer Immunol Immunother* (2016) 65:355–66. doi:10.1007/s00262-016-1804-y
- 109. Ardolino M, Azimi CS, Iannello A, Trevino TN, Horan L, Zhang L, et al. Cytokine therapy reverses NK cell anergy in MHC-deficient tumors. *J Clin Invest* (2014) 124:4781–94. doi:10.1172/JCI74337
- 110. Chen Y, Chen B, Yang T, Xiao W, Qian L, Ding Y, et al. Human fused NKG2D-IL-15 protein controls xenografted human gastric cancer through the recruitment and activation of NK cells. *Cell Mol Immunol* (2017) 14:293–307. doi:10.1038/cmi.2015.81
- 111. Tran HC, Wan Z, Sheard MA, Sun J, Jackson JR, Malvar J, et al. TGFbetaR1 blockade with galunisertib (LY2157299) enhances anti-neuroblastoma activity of the anti-GD2 antibody dinutuximab (ch14.18) with natural killer cells. *Clin Cancer Res* (2017) 23:804–13. doi:10.1158/1078-0432.CCR-16-1743
- 112. Shin H, Blackburn SD, Intlekofer AM, Kao C, Angelosanto JM, Reiner SL, et al. A role for the transcriptional repressor Blimp-1 in CD8(+) T cell exhaustion during chronic viral infection. *Immunity* (2009) 31:309–20. doi:10.1016/j.immuni.2009.06.019
- 113. Simonetta F, Pradier A, Roosnek E. T-bet and eomesodermin in NK cell development, maturation, and function. *Front Immunol* (2016) 7:241. doi:10.3389/fimmu.2016.00241
- Buck MD, Sowell RT, Kaech SM, Pearce EL. Metabolic instruction of immunity. Cell (2017) 169:570–86. doi:10.1016/j.cell.2017.04.004
- Kouidhi S, Elgaaied AB, Chouaib S. Impact of metabolism on T-cell differentiation and function and cross talk with tumor microenvironment. *Front Immunol* (2017) 8:270. doi:10.3389/fimmu.2017.00270
- Gardiner CM, Finlay DK. What fuels natural killers? Metabolism and NK cell responses. Front Immunol (2017) 8:367. doi:10.3389/fimmu.2017. 00367
- 117. Bengsch B, Johnson AL, Kurachi M, Odorizzi PM, Pauken KE, Attanasio J, et al. Bioenergetic insufficiencies due to metabolic alterations regulated by

the inhibitory receptor PD-1 are an early driver of CD8(+) T cell exhaustion. *Immunity* (2016) 45:358–73. doi:10.1016/j.immuni.2016.07.008

- 118. Marcais A, Cherfils-Vicini J, Viant C, Degouve S, Viel S, Fenis A, et al. The metabolic checkpoint kinase mTOR is essential for IL-15 signaling during the development and activation of NK cells. *Nat Immunol* (2014) 15:749–57. doi:10.1038/ni.2936
- 119. Shih HY, Sciume G, Poholek AC, Vahedi G, Hirahara K, Villarino AV, et al. Transcriptional and epigenetic networks of helper T and innate lymphoid cells. *Immunol Rev* (2014) 261:23–49. doi:10.1111/imr.12208
- Pauken KE, Sammons MA, Odorizzi PM, Manne S, Godec J, Khan O, et al. Epigenetic stability of exhausted T cells limits durability of reinvigoration by PD-1 blockade. *Science* (2016) 354:1160–5. doi:10.1126/science. aaf2807
- 121. Sen DR, Kaminski J, Barnitz RA, Kurachi M, Gerdemann U, Yates KB, et al. The epigenetic landscape of T cell exhaustion. *Science* (2016) 354:1165–9. doi:10.1126/science.aae0491

122. Yin J, Leavenworth JW, Li Y, Luo Q, Xie H, Liu X, et al. Ezh2 regulates differentiation and function of natural killer cells through histone methyltransferase activity. *Proc Natl Acad Sci U S A* (2015) 112:15988–93. doi:10.1073/pnas.1521740112

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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