



Dual Face of Vγ9Vδ2-T Cells in Tumor Immunology: Anti- versus Pro-Tumoral Activities

Zheng Xiang and Wenwei Tu*

Li Ka Shing Faculty of Medicine, Department of Paediatrics and Adolescent Medicine, Laboratory for Translational Immunology, University of Hong Kong, Hong Kong, Hong Kong

 $V\gamma 9V\delta 2$ -T cells are considered as potent effector cells for tumor immunotherapy through directly killing tumor cells and indirectly regulating other innate and adaptive immune cells to establish antitumoral immunity. The antitumoral activity of $V\gamma 9V\delta 2$ -T cells is governed by a complicated set of activating and inhibitory cell receptors. In addition, cytokine milieu in tumor microenvironment can also induce the pro-tumoral activities and functional plasticity of $V\gamma 9V\delta 2$ -T cells. Here, we review the anti- versus pro-tumoral activities of $V\gamma 9V\delta 2$ -T cells and discuss the mechanisms underlying the recognition, activation, differentiation and regulation of $V\gamma 9V\delta 2$ -T cells in tumor immunosurveillance. The comprehensive understanding of the dual face of $V\gamma 9V\delta 2$ -T cells in tumor immunology may improve the therapeutic efficacy and clinical outcomes of $V\gamma 9V\delta 2$ -T cell-based tumor immunotherapy.

OPEN ACCESS

Edited by:

Michael Sitkovsky, Northeastern University, United States

Reviewed by:

Viktor Umansky, Deutsches Krebsforschungszentrum (DKFZ), Germany Silvia Deaglio, University of Turin, Italy

> ***Correspondence:** Wenwei Tu wwtu@hku.hk

Specialty section:

This article was submitted to T Cell Biology, a section of the journal Frontiers in Immunology

Received: 11 April 2017 Accepted: 11 August 2017 Published: 28 August 2017

Citation:

Xiang Z and Tu W (2017) Dual Face of V_Y9V82-T Cells in Tumor Immunology: Anti- versus Pro-Tumoral Activities. Front. Immunol. 8:1041. doi: 10.3389/fimmu.2017.01041 Keywords: $V\gamma 9V\delta 2$ -T cells, antitumor activity, pro-tumor activity, tumor immunity, tumor immunotherapy

INTRODUCTION

Human $\gamma\delta$ -T cells can be classified into two main subsets depending on the expression of T cell receptor (TCR) δ chain (1). V δ 1 $\gamma\delta$ -T cells with different V γ elements account for the majority of mucosal-associated lymphoid tissue $\gamma\delta$ -T cells, and they mediate the immune responses to *Listeria monocytogenes*, Cytomegalovirus, and certain hematological malignancies (2, 3). In contrast, $\gamma\delta$ -T cells bearing the V δ 2 gene with the co-expression of the V γ 9 chain (V γ 9V δ 2-T cells) are abundant in the peripheral blood and lymphoid organs of most healthy individuals, and they are involved in the first line of the immune responses to mycobacteria, Epstein–Barr virus (EBV), and some solid tumors (1, 2, 4–10).

The V γ 9V δ 2-T cell is an important component of immune effector cells that contribute to tumor immunosurveillance against many types of tumors, such as lymphoma, myeloma (11, 12), hepatocellular, and colorectal carcinoma (13), and prostate (14), lung (15), colon (16), breast (17), and ovary cancers (18). Recently, our group discovered a novel strategy to treat EBV-induced B-cell lymphoma by boosting V γ 9V δ 2-T cell immunity (19, 20). V γ 9V δ 2-T cells can directly kill tumor cells through the secretion of cytolytic molecules or indirectly prime and modulate immunological functions of other innate and adaptive immune cells to develop and establish profound antitumor immunity (21, 22). Unlike the conventional $\alpha\beta$ -T cells, the V γ 9V δ 2-T cell is a member of the non-conventional lymphocyte family (23), and the antigen recognition of V γ 9V δ 2-T cells is major histocompatibility complex (MHC)-unrestricted (24, 25). Recent growing evidence has suggested that the V γ 9V δ 2-T cell is one of the most attractive candidates for antitumor immunotherapy. In this review, we will discuss recent advances in the basic V γ 9V δ 2-T cell research and evidence from clinical applications of V γ 9V δ 2-T cells. Most importantly, this review will provide an overview of the current knowledge about mechanisms of V γ 9V δ 2-T cell mediated antitumor immunity and the potential limitations of V γ 9V δ 2-T cell-based immunotherapy.

PLASTICITY OF V γ 9V δ 2-T CELLS IN TUMOR IMMUNITY

The antitumor activity of V γ 9V δ 2-T cells is influenced by their functional plasticity that is driven by environmental factors (26). Similar to $\alpha\beta$ -T cells, V γ 9V δ 2-T cells also display the plasticity that contributes to their functional specialization (27). Accumulating evidence indicates that V γ 9V δ 2-T cells can differentiate into the cells with various characteristics associated with Th1-like, Th2-like, Th17-like, follicular T helper cells (Tfh)-like, or regulatory T cells (Treg)-like characteristics (26).

Upon phosphoantigen stimulation, $V\gamma 9V\delta 2$ -T cells preferentially differentiate into Th1-like cells with profound IFN- γ and TNF- α responses (28, 29). Th1-like $V\gamma 9V\delta 2$ -T cells can be induced through isopentenyl pyrophosphate (IPP) activation with IL-12 and anti-IL-4 antibody, and even the addition of IL-21 (30–32). Phosphoantigens and IL-2 can promote their cytolytic activity by upregulating CD56 expression and increasing granule secretion (32, 33). Interestingly, $V\gamma 9V\delta 2$ -T cells can also be polarized into Th2-like cells, which are characterized by increased secretion of IL-4 upon stimulation with IPP, IL-4, and anti-IL-12 antibody (30).

V γ 9V δ 2-T cells with Tfh-like functions can be induced by IL-21 and phosphoantigens stimulation (34, 35). These Tfh-like V γ 9V δ 2-T cells also have the capability to migrate into the lymph node germinal center (35). Similar to Tfh CD4⁺ T-cells, cell-to-cell contact is necessary for the B cell helper activity of the Tfh-like V γ 9V δ 2-T cells.

IL-17-producing γδ-T cells have been extensively discussed in the murine model (36). Recent findings also suggested that human γδ-T cells can produce IL-17 (37, 38). Some groups reported that naïve Vγ9Vδ2-T cells can be induced into the Th17-like phenotype or mixed Th1/Th17-like phenotype (39–41). Vγ9Vδ2-T cells require IL-1β, IL-23, and TGF-β, but not IL-6, for differentiation into Th17-like cells (41). In human colorectal cancer (CRC), activated inflammatory dendritic cells (DCs) polarize Vγ9Vδ2-T cells into IL-17-producing γδ-T cells, which can secrete high levels of IL-17 in an IL-23-dependent manner (42).

Upon stimulation of IPP in the presence of IL-15 and TGF- β , the V γ 9V δ 2-T cells can be induced into transcription factor forkhead box P3 (Foxp3)-expressing Treg-like $\gamma\delta$ -T cells with regulatory/immunosuppressive function (43). When combined with IL-15, IL-2, TGF- β , and phosphoantigen stimulation, decitabine can also induce the immunoregulatory activity of V γ 9V δ 2-T cells (44).

Therefore, $V\gamma 9V\delta 2$ -T cells can be induced into different functional subsets depending on the cytokine milieu in the tumor microenvironment.

ANTITUMORAL RESPONSE OF $V\gamma 9V\delta 2$ -T CELLS

The antitumoral activity of V γ 9V δ 2-T cells has been well studied (45, 46). Their antitumoral activity mostly relies on the recognition of phosphoantigens and stressed molecules by TCR (47) and

other cellular receptors (48), like NKG2D (49). These receptors can respond to perturbations in the endogenous isoprenoid biosynthesis and the presence of "danger signals" that occur during cell stress and malignant transformation. Other molecules can act as co-stimulatory signals to regulate the antitumoral activity of V γ 9V δ 2-T cells (48). Apart from the direct cytotoxicity of V γ 9V δ 2-T cells, these cells can also stimulate and regulate other immune components to establish the antitumoral activity (7, 50). Last, but not least, the homing receptors expressed on V γ 9V δ 2-T cells lead cell migration to tumor sites where they display broad and potent antitumoral activity (51).

Mechanisms of Tumor Cell Recognition TCRγδ Recognition

Non-Peptide Ligands

In tumor immunosurveillance, although Vγ9Vδ2-T cells share some features with other immune cells, such as NK cells and $\alpha\beta$ -T cells (21, 52), a distinctive characteristic of V γ 9V δ 2-T cells is the TCR-dependent recognition of non-peptidic phosphorylated antigens (also called phosphoantigens). Vy9V82-TCR recognizes the molecules that are normally expressed in specific conditions, such as when cells undergo stressful conditions. For example, (E)-4-Hydroxy-3-methyl-but-2-enyl pyrophosphate (HMBPP) and IPP are the products of the prokaryotic isoprenoid pathway and the mevalonate isoprenoid pathway, respectively (53-56). All of these products can be recognized by V γ 9V δ 2-T cells and lead to subsequent activation of $V\gamma 9V\delta 2$ -T cells. The levels of these naturally occurring metabolites are too low to be detected as a dangerous signal by $V\gamma 9V\delta 2$ -T cells in normal cells. The dysfunctional metabolism of malignant tumor cells can result in the accumulation of endogenous phosphoantigens that are recognized by V γ 9V δ 2-T cells. Furthermore, bisphosphonates (such as pamidronate and zoledronate) can cause tumor cells to be more sensitive to the cytotoxicity of Vy9V82-T cells via inhibition of the farnesyl pyrophosphate synthase enzyme in the isoprenoid pathway, which leads to IPP accumulation (57). Meanwhile, current findings have indicated that several molecules, such as F1-ATPase (combined with apolipoprotein A-I, called Apo A-I) (58, 59) and butyrophilin 3A1 (BTN3A1, CD277), might be involved with phosphoantigens to mediate Vy9V82-T cells activation (60, 61) (Figure 1).

Peptide Ligands

(1) Self ligands: in addition to non-peptide ligands, $V\gamma9V\delta2$ -T cells can also recognize some molecules of cellular origin, which could be capable of indicating cellular stress or malignant transformation (49, 62). Several self-antigens have been confirmed to bind to $V\gamma9V\delta2$ -TCR, including heat shock protein-60 (HSP 60) (63), U16-binding protein 4 (ULBP-4) (64), human MutS homolog 2 (hMSH2) (63, 65), and F1-ATP synthase (F1-ATPase) (59, 66). The expressions of these proteins are shown to be upregulated on the surface of different tumor cells and they can promote recognition by $V\gamma9V\delta2$ -T cells. It is intriguing that ULBP-4 and hMSH2 can also bind to NKG2D to induce the cytotoxicity



FIGURE 1 | Underlying mechanisms implicated in regulating antitumoral activity of $V\gamma9V82$ -T cells. $V\gamma9V82$ -T cells can distinguish between tumorous and normal cells using T cell receptor (TCR) and other innate receptors to sense isopentenyl pyrophosphate (IPP) levels and stress signals (such as MICA/B, ULBP4, and MSH2) displayed on target cells. Most importantly, TCR $\gamma\delta$ is the predominant factor that can trigger cell activation without any contribution of other co-stimulators, such as NKG2D. Following TCR-dependent activation, $V\gamma9V82$ -T cells recognize and kill tumor cells by releasing effector molecules, such as granzymes and perforin, and Th-1 cytokines, inducing target cell apoptosis *via* Fas/FasL, TNF-related apoptosis-inducing ligand (TRAIL) and TNF- α pathways, and antibody-dependent cell-mediated cytotoxicity through CD16 expression. The activation threshold is finely regulated by inhibitory receptors, such as NKG2A/CD94. Moreover, adhesion patterns, such as lymphocyte function-associated antigen 1 (LFA-1)/intercellular adhesion molecule-1 (ICAM-1), are also involved in regulating the antitumoral activity of V γ 9V δ 2-T cells. The chemokine receptors, including CCR5, control the ability of V γ 9V δ 2-T cell to migrate to the tumor site. The survival and proliferation of V γ 9V δ 2-T cells are mostly modulated by different cytokines, such as IL-2 and IL-15.

of V γ 9V δ 2-T cells against tumor cells through TCR and NKG2D engagement (63–65) (**Figure 1**).

(2) Non-self ligands: tetanus toxoid (67), Ig λ light chain (68), and viral proteins, such as glycoprotein I from *Herpes simplex* (69) and staphylococcal enterotoxin A (70), are antigens that were reported to be capable of stimulating V γ 9V δ 2-T cell responses.

Cell Receptor Engagement

Besides the $V\gamma 9V\delta 2$ -TCR engagement, some other cellular receptors, especially the NK receptors (NKRs), are involved

in the effective triggering of antitumoral responses of V γ 9V δ 2-T cells (49) (**Figure 1**). Together with previous studies, we reported that NKG2D can bind to its ligands (71), such as MICA, MICB, and ULBP-1, -2, -3, and -4, which are expressed in different tumors, including leukemia, lymphoma, ovarian, and colon carcinoma (72–74). In particular, the high expression level of ULBP1 indicates the susceptibility of lymphoma to V γ 9V δ 2-T cell-mediated cytolysis (74). Furthermore, ULBP-4 expression is detected on the cell surface of EBV-transformed lymphoid cells lines as well as on colon, ovarian, and liver cancer cells (64).

Another NKR implicated in tumor recognition by V γ 9V δ 2-T cells is the DNAX accessory molecule-1 (DNAM-1) (75, 76). Nectin-like-5 and Nectin-2, ligands of DNAM-1, are expressed on most hepatocellular carcinoma (HCC) cell lines (75). In addition, some V γ 9V δ 2-T cells also express NKp44, which can mediate their cytotoxic activity against multiple myeloma (MM) cell lines (77, 78).

Similar to NK cells, $V\gamma 9V\delta 2$ -T cells also express high levels of CD16 (Fc γ R III) upon phosphoantigen stimulation (79), and thus leading to antibody-dependent cell-mediated cytotoxicity (ADCC) against tumor cells (80–83).

$\gamma\delta\text{-}T$ CELLS ACT AS EFFECTOR CELLS

$V\gamma 9V\delta 2$ -T Cells with Killer Functions

Interaction of TCR and/or NKG2D with their respective ligands can stimulate the activation of V γ 9V δ 2-T cells. Once activated, V γ 9V δ 2-T cells secrete IFN- γ and TNF- α , and increase the release of antitumor effector molecules, such as perforin and granzymes. The DNAM-1 signaling pathway can positively regulate the cytotoxic activity and IFN- γ secretion of V γ 9V δ 2-T cells against a broad range of tumors.

Antibody-dependent cell-mediated cytotoxicity mediated by V γ 9V δ 2-T cells can be activated *via* the binding of CD16 to antibodies, such as rituximab, trastuzumab, atumumab, and alemtuzumab, coated on the certain tumor cells (80–83).

In addition, activated V γ 9V δ 2-T cells can also induce tumor cell apoptosis *via* TNF-related apoptosis-inducing ligand and Fas/FasL pathways (84–86).

$V\gamma 9V\delta 2$ -T Cells with Helper Functions

Activated Vy9V82-T cells may secrete chemokines, such as C-C motif chemokine ligand 3 (CCL3), CCL4, C-X-C motif chemokine 10 (CXCL10), and CXCL13, to recruit αβ-T cells, B cells, NK cells, and macrophages/DCs to the tumor site (22, 31, 87). Activated V γ 9V δ 2-T cells not only stimulate DC maturation and macrophage activation (88, 89), but also induce CD4⁺ and CD8⁺ $\alpha\beta$ -T cell differentiation for enhancing antitumoral activity through secretion of IFN- γ and TNF- α and upregulation of CD40L expression (88, 90, 91). Moreover, activated Vγ9Vδ2-T cells can mimic antigen presentation cell (APC) functions by stimulating the antitumoral activity of $\alpha\beta$ -T cells through the upregulation of several surface molecules, such as MHC I and II, CD40, CD83, and CD86 (92, 93). Activated Vy9V82-T cells can also present glycolipid antigens to iNKT cells through the uptake of CD-1d by trogocytosis. Subsequently, activated iNKT cells can trigger downstream reactions to boost antitumoral immunity (94). Furthermore, activated CD137L+ Vy9V82-T cells can stimulate the antitumoral activity of NK cells by acting as a co-stimulator through interacting with CD137 expressed on NK cells. Thus, the costimulatory signals can upregulate the cytotoxic activity of NK cells to kill the solid tumor cells, which usually show resistance to NK cells (95). Although the contribution of IL-17 in tumor surveillance is still controversial (96-99), some in vitro and in vivo evidence have indicated that IL-17 secreted by Vy9V82-T cells might be involved in antitumoral immune responses via indirect mechanisms (96) (Figure 2).

$V\gamma 9V\delta 2$ -T Cells with Homing Functions

CC and CXC chemokines are produced by most tumor cells, such as breast, cervix, pancreatic, and ovarian tumor cells (57). To provide optimal protection against tumor cells, the cytotoxic V γ 9V δ 2-T cells must migrate from the blood-stream to tumor site (33, 100, 101) (**Figure 2**). Unlike V δ 1 $\gamma\delta$ -T cells which express lymph node homing chemokine receptor C-C chemokine receptor 7 (CCR7), the circulating V γ 9V δ 2-T cells preferentially express inflammatory homing chemokine receptor CCR5, which can mediate the migration of V γ 9V δ 2-T cells to CCR5 ligands that are expressed in tumor cells (20, 102). Apart from the predominantly expressed CCR5, V γ 9V δ 2-T cells also express other chemokine receptor 3, which are involved in regulating the homing ability of V γ 9V δ 2-T cells (13, 87, 103).

In addition, some of the adhesion molecules, such as lymphocyte function-associated antigen 1 (LFA-1), L-selectin, and CD44v6, are also involved in the migration of V γ 9V δ 2-T cells to the tumor site (104) (**Figure 1**).

REGULATION MECHANISMS OF $\gamma\delta\text{-}T$ CELL ACTIVATION

$V\gamma 9V\delta 2$ -T Cell Activation

In tumor immunosurveillance, the antitumoral activities of V γ 9V δ 2-T cells is composed of several steps (23): (1) detecting and sensing any types of stress signals in a non-MHC-restricted manner; (2) producing a huge quantity of effector molecules that can kill tumor cells through direct and indirect mechanisms; and (3) exhibiting potent cytotoxic and cytolytic activities against a broad panel of tumors. In order to tightly monitor the initiation and development of tumorigenesis, the activation of V γ 9V δ 2-T cells is triggered by TCR-mediated recognition and precisely regulated by various innate immune cells and cytokine receptors. Furthermore, the activation and differentiation of V γ 9V δ 2-T cells are molecularly controlled by surface receptors that capture key extracellular cues and convey downstream intracellular signals (105). More detailed information can be found in **Figure 1**.

Regulation of V γ 9V δ 2-T Cell Activation Regulation by TCR

Vγ9Vδ2-T cells mainly recognize the non-peptide ligands, phosphoantigens, which are shown to act as a trigger signal to activate Vγ9Vδ2-T cells (105). Cipriani et al. concluded that IPP induces rapid and persistent PKC-dependent phosphorylation of ERK 1/2, p38 MAPK, and JNK, which can lead to the activation of NF- κ B and AP-1 and the secretion of IFN- γ and TNF- α (106). Some studies have shown that crosslinking of CD3 and phosphoantigens stimulation can induce highly sustained calcium signaling in V γ 9V δ 2-T cells *via* phosphorylation of Zap70, Pi3K, LAT, ERK 1/2, and p38 MAPK (105, 107, 108).

Due to upregulated self-antigen expressions in the transformed tumor cells, $V\gamma 9V\delta 2$ -T cells discriminate these malignant cells by directly binding to these self-antigens (49), such as HSP 60,



also suppress innate and adaptive immune responses. Besides the "EFFECTOR" function, Vy9V82-T cells can also exhibit the "HELPER" functions in antitumor immune immunity. The activated Vy9V82-T cells can promote the maturation of DCs and the activation of macrophages. These Vy9V82-T cells can act as APC and initiate the differentiation and stimulation of tumor antigen-specific CD4+ T and CD8+ T cells. Moreover, Vy9V82-T cells can present antigens to iNKT and B cells. Vy9V82-T cells can promote NK cell antitumor response by delivering a costimulatory signal. APC, antigen presentation cell; DCs, dendritic cells; iNKT, invariant NKT cells; IPP, isopentenyl pyrophosphate; MDSCs, myeloid-derived suppressor cells; NK, natural killer cell; N-BPs, nitrogen-containing bisphosphonates; and TAMs, tumor-associated macrophages.

ULBP-4, hMSH2, and F1-ATPase, which can enhance the activation and cytotoxicity of $V\gamma 9V\delta 2$ -T cells.

Regulation by NK-Associated Receptors

The expression of NK-associated receptors is a distinguishing feature of V γ 9V δ 2-T cells that mediates the recognition of stress ligands expressed by normal cells during infection and cell transformation by different pathogens. NKG2D and its ligands, such as MICA/B and ULBPs (64, 74, 109), are well-defined NK-associated receptors and ligands for V γ 9V δ 2-T cells. For example, expressions of these ligands are normally induced during cellular stress, such as DNA damage that occurs in tumor cells.

Whether NKG2D plays a primary stimulatory or a co-stimulatory role in V γ 9V δ 2-T cells is still being debated (49). Some evidence have indicated that an additive effect of the NKG2D pathway on TCR-mediated activation through increasing cytokine production (110), and upregulating intracellular calcium mobilization and enhancing cytotoxic activity (105). On the other hand, other studies have reported that the NKG2D pathway alone can activate V γ 9V δ 2-T cells without TCR engagement (111), and the blockade of the NKG2D pathway, but not the blockade of γ \delta-TCR, can inhibit the cytotoxicity of V γ 9V δ 2-T cells against some hematological tumors (74).

DNAX accessory molecule-1 is another NK-associated receptor that regulates V γ 9V δ 2-T cells, whereby blocking the interaction between DNAM-1 and its ligands, such as Nectin-like-5 and Nectin-2, could impair the cytotoxic capacity and IFN- γ production of V γ 9V δ 2-T cells against HCC cells (75).

A recent study demonstrated that NKp44, a natural cytotoxicity receptor, is involved in the cytotoxicity of V γ 9V δ 2-T cells against MM cell lines that lack the expression of NKG2D ligands (77). Because V γ 9V δ 2-T cells are potentially highly self-reactive, TCR-mediated activity needs to be tightly controlled by a close interplay between activating and inhibitory NKRs (105). Like NK cells, most human circulating V γ 9V δ 2-T cells express several inhibitory NKRs belonging to the lectin-like receptor family (such as the NKG2A/CD94 heterodimer), or the immunoglobulin (Ig) family [such as Ig-like transcript 2 (ILT-2)]. Upon interaction with the classical and/or non-classical MHC I molecules, the inhibitory signals are delivered by these receptors to V γ 9V δ 2-T cells. This mechanism allows TCR-activated V γ 9V δ 2-T cells to target the tumor cells rather than the normal cells depending on the expression level of the MHC I molecules, and thus preventing self-reactivity while enhancing antitumoral activity (105). In addition, NKG2C/CD94 also can regulate V γ 9V δ 2-T cell effector functions, including cytokine secretion, cell proliferation, and cytotoxic activity (112).

Regulation by Co-Stimulatory Receptors

There are two main types of costimulatory receptors that are expressed on Vy9V82-T cells: the Ig and tumor necrosis factor receptor (TNFR) superfamilies. Ribot et al. suggested that CD28, which is an important member of the Ig superfamily, is constitutively expressed on lymphoid Vy9V82-T cells. Co-stimulation with CD28 promotes the survival and proliferation of Vy9V82-T cells by enhancing IL-2 production, whereas blocking antibodies target its B7 ligands (CD80 and CD86) can inhibit cell survival and proliferation (113). CD27 is a member of the TNFR superfamily that is expressed in 80% of V γ 9V δ 2-T cells (114). CD27 pathway has been confirmed to play critical roles in co-stimulating Vγ9Vδ2-T cell activation. Furthermore, CD27 is also involved in promoting cell proliferation, upregulating the antiapoptotic gene Bcl2a1 to maintain cell survival and enhancing IFN-y production and cytotoxicity of Vy9V82-T cells (105, 114). Upon activation, CD30, another member of the TNFR superfamily, is also expressed on Vy9V82-T cells and it can increase pro-inflammatory cytokine production and promote TCR-induced calcium fluxes (115). A high level of CD137L that is expressed on activated Vy9V82-T cells can transmit a reversal signal to regulate $V\gamma 9V\delta 2$ -T cell activation (95, 116).

Regulation by Adhesion Molecules

In addition to TCR and NKR, other surface receptors, such as adhesion molecules, are also important for regulating Vγ9V82-T cell function. To date, several adhesion molecules, including LFA-1/intercellular adhesion molecule-1 (ICAM-1) (117), CD2/ LFA-1/3 (118), and CD6/CD166 (119), have been identified as regulators in modulating Vγ9V82-T cell activation.

High expressions of LFA-1 and its ligand, ICAM-1, have been detected on the surface of V γ 9V δ 2-T cells and most of the tumor cell lines, respectively. LFA-1/ICAM-1 adhesive interaction is necessary but not sufficient for IFN- γ production and the cytotoxicity of V γ 9V δ 2-T cells (117). The interaction between LFA-3 on V γ 9V δ 2-T cells and CD2 on lymphoid cells can stimulate TNF- α secretion by V γ 9V δ 2-T cells (118). Additionally, CD6 is a member of the scavenger receptor family, which is also expressed on the surface of V γ 9V δ 2-T cells. CD6 binds to its ligand, CD166,

which is associated with the capability of tumor cells to activate $V\gamma 9V\delta 2$ -T cells upon phosphoantigen induction (119).

Most importantly, the interactions between LFA-1/ICAM-1 and CD2/LFA-3 can stabilize the immunological synapses after V γ 9V δ 2-TCR/phosphoantigen triggers the formation of immunological synapses (120, 121).

Regulation by Toll-Like Receptors (TLRs)

The roles of TLRs in regulating the activation of V γ 9V δ 2-T cells are not completely understood. After ligation of TLR2, TLR3, and TLR5, the V γ 9V δ 2-T cells can produce IFN- γ , TNF- α , granulocyte-macrophage colony-stimulating factor, CCL3, and CCL5 (122). The triggering of TLR3, 4, 5, and 9 can also induce the early activation of V γ 9V δ 2-T cell and the production of IFN- γ (105). In addition, TLR3 and TLR7 agonists can enhance the antitumoral activity of V γ 9V δ 2-T cells against adenocarcinoma cells (123). Nevertheless, these responses require simultaneous stimulation of V γ 9V δ 2-TCR (105).

Regulation by Cytokine Receptors

The interleukin receptors are essential for the development and homeostasis of V γ 9V δ 2-T cells due to the pivotal effect of interleukins on cell proliferation, differentiation, and survival, and the regulation of the immunological functions of the V γ 9V δ 2-T cells. IL-2 and IL-15 can be used to stimulate V γ 9V δ 2-T cell expansion, but they cannot fully induce effector functions without $\gamma\delta$ -TCR-mediated activation and downstream signals, such as the ERK and AKT pathway (124, 125). IL-12 and IL-18 are reported to be beneficial for differentiation into IFN- γ^+ V γ 9V δ 2-T cells (126). Recent studies also indicated that other interleukins, such as IL-23, IL-1 β (39, 40, 127), and IL-21 (34, 35), are also involved in the induction of V γ 9V δ 2-T cell plasticity.

Regulation by Inhibitory Receptors

Like inhibitory NKRs, some other inhibitory molecules, such as programmed cell death protein-1 (PD-1) and B- and T-lymphocyte attenuator (BTLA), act in a similar manner in Vy9V82-T cells. PD-1 and BTLA can negatively regulate Vy9V82-T cell responses. Vy9V82-T cells do not normally express detectable PD-1; however, once activated, the expression of PD-1 is upregulated in Vy9V82-T cells (128). Iwasaki et al. found that the interaction of PD-1 and PD-L1 can attenuate the antitumoral activity of Vy9V82-T cells by downregulating IFN-y production and lowering cytotoxicity (128). BTLA engagement can downregulate phosphoantigen/ $\gamma\delta$ -TCR-mediated signaling and inhibit Vy9V82-T cell proliferation, including the response to lymphoma cells (129). Moreover, the HLA-G expressed on the surface of tumor cells can bind to the ILT-2 on V γ 9V δ 2-T cells. This interaction could impair cytokine production and the cytotoxicity of V γ 9V δ 2-T cells (130).

$\begin{array}{l} \mbox{PRO-TUMORAL RESPONSE} \\ \mbox{OF } V\gamma 9V\delta 2\mbox{-T CELLS} \end{array}$

Recently, more evidence indicates that $V\gamma9V\delta2\text{-}T$ cells can display the functions for promoting tumor development

through direct or indirect strategies (27). As mentioned above, the differentiation of unique subpopulations of $V\gamma 9V\delta 2$ -T cells with immunosuppressive features can be induced in the presence of specific stimuli, such as in the tumor-established microenvironment (Figure 2). For example, $V\gamma 9V\delta 2$ -T cells may display Th2-, Th17-, or Treg-like profile and produce IL-4, IL-17, IL-10, and TGF-β, respectively. IL-10 and TGF-β are the cytokines with immunosuppressive activity, while IL-10 can impair APC function of DCs, and induce the pro-tumoral functions of Vy9V82-T cells (131, 132). Some evidence indicates that IL-17 produced by Th17-like Vy9V82-T cells can directly promote the proliferation and dissemination of tumor cells in breast cancer (133). Most importantly, in the tumor microenvironment, IL-17 plays a critical regulatory role to other immune components, such as MDSCs and macrophage, and thus indirectly influences tumor immunosurveillance (42). Moreover, similar to classical Treg cells, the suppressive effect of regulatory-like Vy9V82-T cells also depends on cell-to-cell contact via CD86/CTLA-4 and PD-L1/PD-1 interactions between activated Vy9V82-T cells and their responder cells (134) (Figure 3).

More recently, Peters et al. demonstrated that $V\gamma 9V\delta 2$ -T cell is a major source of IL-9 *in vitro* (135). According to this study,



FIGURE 3 | Antitumoral versus pro-tumoral activities of human Vy9V82-T cells. Vy9V82-T cells display dual effects in the immune surveillance against tumors, including antitumoral and pro-tumoral functions. On one hand, $V_{\gamma}9V\delta2$ -T cells can control and suppress tumor development directly through potent cytotoxic capability and indirectly by regulating other innate and adaptive immune cells to establish antitumoral immune responses. First, $V\gamma 9V\delta 2$ -T cells can recognize and eliminate tumor cells with their potent killing ability. Second, they can promote dendritic cell maturation and macrophage activation. Third, $V_{\gamma}9V\delta2$ -T cells can present antigens to $\alpha\beta\text{-T}$ cells and iNKT cells. Fourth, they can stimulate and enhance NK cell antitumoral response. Finally, $V_{\gamma}9V\delta2$ -T cells can help B cells produce antibodies. On the other hand, in the tumor microenvironment, the $V\gamma 9V\delta 2$ -T cells can be induced into Th-17-like cells and Treg-like cells by specific stimuli and they can secret immunosuppressive cytokines, such as IL-17, IL-10, and TGF-β, to suppress and modulate innate and adaptive antitumoral immunity. These strategies will be beneficial for tumor evasion/ growth, metastasis, and angiogenesis.

V γ 9V δ 2-T cells can secret IL-9 when culturing with TGF- β and IL-15 and in the absence of IL-4. IL-9-producing CD8+ T cells and Th9 cells are always considered as new players in antitumoral immune responses (136-138); however, in some studies, IL-9 also displays potential pro-tumoral activity. Qiu et al. found that in ALK⁺ anaplastic large-cell lymphoma, IL-9 can promote lymphoma cell proliferation and colony formation by regulating Jak3 activation in an autocrine manner (139). Furthermore, IL-9 can abolish the establishment of immunological memory to tumor re-challenge and exhibits an immunomodulatory role in pro-tumoral immunity (140). Indeed, elevated serum IL-9 was found in the patients with metastatic breast cancer or diffuse large B-cell lymphoma (DLBCL) (141, 142). Meantime, Lv et al. found that IL-9 can support the survival of DLBCL cells and enhance the resistance of these tumor cells to chemotherapeutic drugs by upregulating p21CIP1 genes (141). Thus, in tumor immunity, the exact immunological roles of IL-9-producing Vy9V82-T cells are still unknown. So far, the evidence for pro-tumoral activity of Vγ9Vδ2-T cells mostly comes from ex vivo experiments. More clinical evidence is needed to evaluate the potential pro-tumor ability of Vy9V82-T cells in vivo.

THERAPEUTIC EFFECTS OF $V\gamma9V\delta2$ -T CELLS IN CLINICAL TRIALS

To date, two main therapeutic strategies based on $V\gamma 9V\delta 2$ -T cells have been proposed for tumor immunotherapy: the in vivo expansion of Vy9V82-T cells by aminobisphosphonates, and the adoptive transfer of *ex vivo*-expanded V γ 9V δ 2-T cells (143). Several clinical trials have been initiated to evaluate the safety and the rapeutic efficacy of V γ 9V δ 2-T cell immunotherapy based on in vivo and ex vivo expansion. The clinical outcomes of these strategies have been confirmed in patients with different types of tumors, including leukemia/lymphoma, melanoma, RCC, hormone-refractory prostate cancer, breast cancer, NSCLC, CRC, MM, gastrointestinal tumors, ovarian cancer, cervical cancer, and bone cancer (45, 57). For instance, Wilhelm et al. first reported that administration of pamidronate and low-dose of IL-2 to expand Vy9V82-T cells in vivo was well tolerated and could induce objective tumor response in patients with low-grade non-Hodgkin lymphoma or MM (18). Lang and coworkers conducted a pilot trial to determine the therapeutic effects of zoledronate with low-dose of IL-2 in patients with RCC (144). Furthermore, zoledronate with low dose of IL-2 have also been tested in treating prostate cancer and advanced breast cancer where partial remissions haves been reported (14, 17). Bennouna et al. conducted a phase I study that, after treatment of bromohydrin pyrophosphate (BrHPP) and IL-2, in total of 28 patients, 12 patients had stable disease and 16 had progressive disease after three cycles of administration (145).

Adoptive transfer of zoledronate-expanded V γ 9V δ 2-T cells was reported to induce a complete remission of RCC patient with lung metastasis. (146). Kobayashi et al. also reported that adoptive transfer of V γ 9V δ 2-T cells in combination with zoledronate and IL-2 was well tolerated and the objective clinical responses could

be achieved in some patients with advanced RCC (147). More detailed information about clinical studies of V γ 9V δ 2-T cells had been systematically reviewed in recent articles summarized by Deniger et al. and Kobayashi and Tanaka (45, 148).

Although these two strategies have yielded clinical success, there are still some limitations. For the in vivo expansion and activation of $V\gamma 9V\delta 2$ -T cells, one of the limitations is that the sustained proliferative activity is impaired, probably due to an energy or exhaustion of Vy9V82-T cells induced by the successive infusions of BrHPP and IL-2 (45, 149). Additionally, Kalvan et al. demonstrated that neutrophils in human peripheral blood could uptake of zoledronate and cause the suppression of Vy9V82-T cells (150). Fowler et al. suggested that systemic use of zoledronate reduced the tumor homing ability of Vy9V82-T cells (151). Indeed, repeated administration of zoledronate and IL-2 in the patients with breast cancer and RCC could inhibit the proliferative capacity of V γ 9V δ 2-T cells and reduced the responsiveness of V γ 9V δ 2-T cells to re-stimulation (144, 152). For the adoptive cell transfer therapy, the main problem is difficulty to expand Vy9V82-T cells ex vivo from the advanced cancer patients with limited initial number of Vy9V82-T cells, especially after radiotherapy and chemotherapy (45). Moreover, the effect of immunosuppressive tumor microenvironment on adoptively transferred V γ 9V δ 2-T cells is still not clear.

Up to now, aminobisphosphonates have only been found to target and stimulate Vy9V82-T cell subsets, but they do not target other subpopulations of $\gamma\delta$ -T cells, such as V δ 1-T cells. Some synthetic phosphoantigens, such as BrHPP, HMBPP, and 2-methyl-3-butenyl-1-pyrophosphate (2M3B1PP), are also found to activate and expand V γ 9V δ 2-T cells by mimicking the effects of aminobisphosphonates (153). Upon stimulation by these compounds, $V\gamma 9V\delta 2$ -T cells can enhance their antitumoral activities by release of IFN- γ and TNF- α (57). The administration of BrHPP was also tested in a Phase I study; the data showed that the BrHPP treatment was well tolerated and expansion of Vy9V82-T cells was successfully induced in patients with solid tumors (145). In addition to synthetic phosphoantigens, some monoclonal antibodies targeting γδ-TCR are also good candidates for tumor immunotherapy. Starick et al. demonstrated that BTN3A (CD277)-specific monoclonal antibody 20.1 induced

REFERENCES

- Chien YH, Meyer C, Bonneville M. Gammadelta Tcells: first line of defense and beyond. AnnuRevImmunol(2014)32:121–55.doi:10.1146/annurev-immunol-032713-120216
- Poggi A, Zocchi MR. gammadelta T Lymphocytes as a first line of immune defense: old and new ways of antigen recognition and implications for cancer immunotherapy. *Front Immunol* (2014) 5:575. doi:10.3389/fimmu. 2014.00575
- 3. Hovav AH. Human gammadelta T cells: rapid, stable and clonally reactive. *Cell Mol Immunol* (2017) 14(8):646–8. doi:10.1038/cmi.2017.33
- Zheng J, Liu Y, Lau YL, Tu W. gammadelta-T cells: an unpolished sword in human anti-infection immunity. *Cell Mol Immunol* (2013) 10(1):50–7. doi:10.1038/cmi.2012.43
- O'Brien RL, Born WK. Dermal gammadelta T cells what have we learned? Cell Immunol (2015) 296(1):62–9. doi:10.1016/j.cellimm.2015.01.011

TCR-mediated activation of V γ 9V δ 2-T cells (154). This study not only provided the novel mechanisms involved in V γ 9V δ 2-T cell activation but also showed the potential for using BTN3A-specific antibodies to manipulate V γ 9V δ 2-T cell immunity in tumor immunotherapy (154, 155).

Some recent studies have focused on the development of novel protocols to expand V γ 9V δ 2-T cells using immobilized antigens (156), agonistic monoclonal antibodies (15, 101), and tumordriven artificial antigen-presenting cells (157, 158). Meanwhile, V γ 9V δ 2-T cells are also amenable to genetic modifications *via* the introduction of $\alpha\beta$ TCRs (159), and chimeric antigen receptors (160). Furthermore, V γ 9V δ 2-T cells are suggested to act as a novel cellular vaccine to treat cancer patients (161).

CONCLUSION

Recent advances in V γ 9V δ 2-T cell research have paved a way for developing innovative therapeutic strategies for tumor immunotherapy. As discussed in this review, V γ 9V δ 2-T cells can recognize tumor cells through TCR and other cell surface receptors, and their antitumoral activity is strictly regulated by activating and inhibitory receptors and their ligands. In addition, stimuli/cytokine milieu in tumor microenvironment can also induce pro-tumoral activity and functional plasticity of V γ 9V δ 2-T cells. Further study on the dual face of V γ 9V δ 2-T cells in tumor immunology could optimize current therapeutic protocols and improve the therapeutic efficacy and clinical outcomes of V γ 9V δ 2-T cell-based tumor immunotherapy.

AUTHOR CONTRIBUTIONS

ZX and WT wrote the manuscript. WT revised the manuscript.

FUNDING

This work was supported in part by Theme-based Research Scheme (Project No. T11-705/14N), the General Research Fund (HKU780113M, 17121214, and 17115015), the Area of Excellence Program supported by the University Grants Committee of the Hong Kong SAR, China (AoE/M-06/08).

- Bonneville M, Chen ZW, Dechanet-Merville J, Eberl M, Fournie JJ, Jameson JM, et al. Chicago 2014 – 30 years of gammadelta T cells. *Cell Immunol* (2014) 296(1):3–9. doi:10.1016/j.cellimm.2014.11.001
- Ferreira LM. Gammadelta T cells: innately adaptive immune cells? Int Rev Immunol (2013) 32(3):223–48. doi:10.3109/08830185.2013.783831
- Li H, Xiang Z, Feng T, Li J, Liu Y, Fan Y, et al. Human Vgamma9Vdelta2-T cells efficiently kill influenza virus-infected lung alveolar epithelial cells. *Cell Mol Immunol* (2013) 10(2):159–64. doi:10.1038/cmi.2012.70
- Tu W, Zheng J, Liu Y, Sia SF, Liu M, Qin G, et al. The aminobisphosphonate pamidronate controls influenza pathogenesis by expanding a gammadelta T cell population in humanized mice. *J Exp Med* (2011) 208(7):1511–22. doi:10.1084/jem.20110226
- Qin G, Mao H, Zheng J, Sia SF, Liu Y, Chan PL, et al. Phosphoantigenexpanded human gammadelta T cells display potent cytotoxicity against monocyte-derived macrophages infected with human and avian influenza viruses. J Infect Dis (2009) 200(6):858–65. doi:10.1086/605413

- Cordova A, Toia F, La Mendola C, Orlando V, Meraviglia S, Rinaldi G, et al. Characterization of human gammadelta T lymphocytes infiltrating primary malignant melanomas. *PLoS One* (2012) 7(11):e49878. doi:10.1371/ journal.pone.0049878
- Dudley ME, Wunderlich JR, Yang JC, Sherry RM, Topalian SL, Restifo NP, et al. Adoptive cell transfer therapy following non-myeloablative but lymphodepleting chemotherapy for the treatment of patients with refractory metastatic melanoma. *J Clin Oncol* (2005) 23(10):2346–57. doi:10.1200/JCO.2005.00.240
- Bouet-Toussaint F, Cabillic F, Toutirais O, Le Gallo M, Thomas de la Pintiere C, Daniel P, et al. Vgamma9Vdelta2 T cell-mediated recognition of human solid tumors. Potential for immunotherapy of hepatocellular and colorectal carcinomas. *Cancer Immunol Immunother* (2008) 57(4):531–9. doi:10.1007/ s00262-007-0391-3
- 14. Dieli F, Vermijlen D, Fulfaro F, Caccamo N, Meraviglia S, Cicero G, et al. Targeting human $\gamma\delta$ T cells with zoledronate and interleukin-2 for immunotherapy of hormone-refractory prostate cancer. *Cancer Res* (2007) 67(15):7450–7. doi:10.1158/0008-5472.CAN-07-0199
- Kang N, Zhou J, Zhang T, Wang L, Lu F, Cui Y, et al. Adoptive immunotherapy of lung cancer with immobilized anti-TCRgammadelta antibody-expanded human gammadelta T-cells in peripheral blood. *Cancer Biol Ther* (2009) 8(16):1540–9. doi:10.4161/cbt.8.16.8950
- Corvaisier M, Moreau-Aubry A, Diez E, Bennouna J, Mosnier JF, Scotet E, et al. V gamma 9V delta 2 T cell response to colon carcinoma cells. *J Immunol* (2005) 175(8):5481–8. doi:10.4049/jimmunol.175.8.5481
- Meraviglia S, Eberl M, Vermijlen D, Todaro M, Buccheri S, Cicero G, et al. In vivo manipulation of Vgamma9Vdelta2 T cells with zoledronate and lowdose interleukin-2 for immunotherapy of advanced breast cancer patients. *Clin Exp Immunol* (2010) 161(2):290–7. doi:10.1111/j.1365-2249.2010. 04167.x
- Wilhelm M, Kunzmann V, Eckstein S, Reimer P, Weissinger F, Ruediger T, et al. Gammadelta T cells for immune therapy of patients with lymphoid malignancies. *Blood* (2003) 102(1):200–6. doi:10.1182/blood-2002-12-3665
- Dharnidharka VR, Mohanakumar T. New approaches to treating B-cell cancers induced by Epstein-Barr virus. N Engl J Med (2015) 372(6):569–71. doi:10.1056/NEJMcibr1415117
- Xiang Z, Liu Y, Zheng J, Liu M, Lv A, Gao Y, et al. Targeted activation of human Vgamma9Vdelta2-T cells controls Epstein-Barr virus-induced B cell lymphoproliferative disease. *Cancer Cell* (2014) 26(4):565–76. doi:10.1016/j. ccr.2014.07.026
- Woo SR, Corrales L, Gajewski TF. Innate immune recognition of cancer. *Annu Rev Immunol* (2015) 33:445–74. doi:10.1146/annurev-immunol-032414-112043
- 22. Tyler CJ, Doherty DG, Moser B, Eberl M. Human Vgamma9/Vdelta2 T cells: innate adaptors of the immune system. *Cell Immunol* (2015) 296(1):10–21. doi:10.1016/j.cellimm.2015.01.008
- Lafont V, Sanchez F, Laprevotte E, Michaud HA, Gros L, Eliaou JF, et al. Plasticity of gammadelta T cells: impact on the anti-tumor response. *Front Immunol* (2014) 5:622. doi:10.3389/fimmu.2014.00622
- Morita CT, Beckman EM, Bukowski JF, Tanaka Y, Band H, Bloom BR, et al. Direct presentation of nonpeptide prenyl pyrophosphate antigens to human gamma delta T cells. *Immunity* (1995) 3(4):495–507. doi:10.1016/1074-7613(95)90178-7
- Morita CT, Tanaka Y, Bloom BR, Brenner MB. Direct presentation of non-peptide prenyl pyrophosphate antigens to human gamma delta T cells. *Res Immunol* (1996) 147(5):347–53. doi:10.1016/0923-2494(96)89649-0
- Wesch D, Peters C, Siegers GM. Human gamma delta T regulatory cells in cancer: fact or fiction? *Front Immunol* (2014) 5:598. doi:10.3389/ fimmu.2014.00598
- Caccamo N, Todaro M, Sireci G, Meraviglia S, Stassi G, Dieli F. Mechanisms underlying lineage commitment and plasticity of human gammadelta T cells. *Cell Mol Immunol* (2013) 10(1):30–4. doi:10.1038/cmi.2012.42
- Dunne MR, Mangan BA, Madrigal-Estebas L, Doherty DG. Preferential Th1 cytokine profile of phosphoantigen-stimulated human Vgamma9-Vdelta2 T cells. *Mediators Inflamm* (2010) 2010:704941. doi:10.1155/2010/ 704941
- 29. Thedrez A, Sabourin C, Gertner J, Devilder MC, Allain-Maillet S, Fournie JJ, et al. Self/non-self discrimination by human gammadelta T cells:

simple solutions for a complex issue? *Immunol Rev* (2007) 215:123-35. doi:10.1111/j.1600-065X.2006.00468.x

- Wesch D, Glatzel A, Kabelitz D. Differentiation of resting human peripheral blood gamma delta T cells toward Th1- or Th2-phenotype. *Cell Immunol* (2001) 212(2):110–7. doi:10.1006/cimm.2001.1850
- Vermijlen D, Ellis P, Langford C, Klein A, Engel R, Willimann K, et al. Distinct cytokine-driven responses of activated blood gammadelta T cells: insights into unconventional T cell pleiotropy. *J Immunol* (2007) 178(7):4304–14. doi:10.4049/jimmunol.178.7.4304
- Thedrez A, Harly C, Morice A, Salot S, Bonneville M, Scotet E. IL-21-mediated potentiation of antitumor cytolytic and proinflammatory responses of human V gamma 9V delta 2 T cells for adoptive immunotherapy. *J Immunol* (2009) 182(6):3423–31. doi:10.4049/jimmunol.0803068
- Qin G, Liu Y, Zheng J, Ng IH, Xiang Z, Lam KT, et al. Type 1 responses of human Vgamma9Vdelta2 T cells to influenza A viruses. *J Virol* (2011) 85(19):10109–16. doi:10.1128/JVI.05341-11
- Bansal RR, Mackay CR, Moser B, Eberl M. IL-21 enhances the potential of human gammadelta T cells to provide B-cell help. *Eur J Immunol* (2012) 42(1):110–9. doi:10.1002/eji.201142017
- Caccamo N, Todaro M, La Manna MP, Sireci G, Stassi G, Dieli F. IL-21 regulates the differentiation of a human gammadelta T cell subset equipped with B cell helper activity. *PLoS One* (2012) 7(7):e41940. doi:10.1371/journal. pone.0041940
- Sutton CE, Lalor SJ, Sweeney CM, Brereton CF, Lavelle EC, Mills KH. Interleukin-1 and IL-23 induce innate IL-17 production from gammadelta T cells, amplifying Th17 responses and autoimmunity. *Immunity* (2009) 31(2):331–41. doi:10.1016/j.immuni.2009.08.001
- Deknuydt F, Scotet E, Bonneville M. Modulation of inflammation through IL-17 production by gammadelta T cells: mandatory in the mouse, dispensable in humans? *Immunol Lett* (2009) 127(1):8–12. doi:10.1016/j.imlet.2009.08.003
- Wu X, Zhang JY, Huang A, Li YY, Zhang S, Wei J, et al. Decreased Vdelta2 gammadelta T cells associated with liver damage by regulation of Th17 response in patients with chronic hepatitis B. *J Infect Dis* (2013) 208(8):1294–304. doi:10.1093/infdis/jit312
- Caccamo N, La Mendola C, Orlando V, Meraviglia S, Todaro M, Stassi G, et al. Differentiation, phenotype, and function of interleukin-17-producing human Vgamma9Vdelta2 T cells. *Blood* (2011) 118(1):129–38. doi:10.1182/ blood-2011-01-331298
- Ness-Schwickerath KJ, Jin C, Morita CT. Cytokine requirements for the differentiation and expansion of IL-17A- and IL-22-producing human Vgamma2Vdelta2 T cells. *J Immunol* (2010) 184(12):7268–80. doi:10.4049/ jimmunol.1000600
- Ness-Schwickerath KJ, Morita CT. Regulation and function of IL-17A- and IL-22-producing gammadelta T cells. *Cell Mol Life Sci* (2011) 68(14):2371–90. doi:10.1007/s00018-011-0700-z
- Wu P, Wu D, Ni C, Ye J, Chen W, Hu G, et al. gammadeltaT17 cells promote the accumulation and expansion of myeloid-derived suppressor cells in human colorectal cancer. *Immunity* (2014) 40(5):785–800. doi:10.1016/j. immuni.2014.03.013
- 43. Casetti R, Agrati C, Wallace M, Sacchi A, Martini F, Martino A, et al. Cutting edge: TGF-beta1 and IL-15 Induce FOXP3+ gammadelta regulatory T cells in the presence of antigen stimulation. *J Immunol* (2009) 183(6):3574–7. doi:10.4049/jimmunol.0901334
- 44. Hu Y, Cui Q, Gu Y, Sheng L, Wu K, Shi J, et al. Decitabine facilitates the generation and immunosuppressive function of regulatory gammadeltaT cells derived from human peripheral blood mononuclear cells. *Leukemia* (2013) 27(7):1580–5. doi:10.1038/leu.2012.345
- Kobayashi H, Tanaka Y. gammadelta T cell immunotherapy-A review. *Pharmaceuticals* (2015) 8(1):40–61. doi:10.3390/ph8010040
- Rei M, Pennington DJ, Silva-Santos B. The emerging protumor role of gammadelta T lymphocytes: implications for cancer immunotherapy. *Cancer Res* (2015) 75(5):798–802. doi:10.1158/0008-5472.CAN-14-3228
- Wiemer DF, Wiemer AJ. Opportunities and challenges in development of phosphoantigens as Vgamma9Vdelta2 T cell agonists. *Biochem Pharmacol* (2014) 89(3):301–12. doi:10.1016/j.bcp.2014.03.009
- Ribeiro ST, Ribot JC, Silva-Santos B. Five layers of receptor signaling in gammadelta T-cell differentiation and activation. *Front Immunol* (2015) 6:15. doi:10.3389/fimmu.2015.00015

- Correia DV, Lopes A, Silva-Santos B. Tumor cell recognition by gammadelta T lymphocytes: T-cell receptor vs. NK-cell receptors. *Oncoimmunology* (2013) 2(1):e22892. doi:10.4161/onci.22892
- Lo Presti E, Dieli F, Meraviglia S. Tumor-infiltrating gammadelta T lymphocytes: pathogenic role, clinical significance, and differential programing in the tumor microenvironment. *Front Immunol* (2014) 5:607. doi:10.3389/ fimmu.2014.00607
- Fowler DW, Bodman-Smith MD. Harnessing the power of Vdelta2 cells in cancer immunotherapy. *Clin Exp Immunol* (2015) 180(1):1–10. doi:10.1111/ cei.12564
- Bonneville M, O'Brien RL, Born WK. Gammadelta T cell effector functions: a blend of innate programming and acquired plasticity. *Nat Rev Immunol* (2010) 10(7):467–78. doi:10.1038/nri2781
- 53. Hintz M, Reichenberg A, Altincicek B, Bahr U, Gschwind RM, Kollas AK, et al. Identification of (E)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate as a major activator for human gammadelta T cells in *Escherichia coli*. *FEBS Lett* (2001) 509(2):317–22. doi:10.1016/S0014-5793(01)03191-X
- Altincicek B, Moll J, Campos N, Foerster G, Beck E, Hoeffler JF, et al. Cutting edge: human gamma delta T cells are activated by intermediates of the 2-C-methyl-D-erythritol 4-phosphate pathway of isoprenoid biosynthesis. *J Immunol* (2001) 166(6):3655–8. doi:10.4049/jimmunol.166.6.3655
- Gober HJ, Kistowska M, Angman L, Jeno P, Mori L, De Libero G. Human T cell receptor gammadelta cells recognize endogenous mevalonate metabolites in tumor cells. J Exp Med (2003) 197(2):163–8. doi:10.1084/jem.20021500
- Chen ZW. Multifunctional immune responses of HMBPP-specific Vgamma2Vdelta2 T cells in *M. tuberculosis* and other infections. *Cell Mol Immunol* (2013) 10(1):58–64. doi:10.1038/cmi.2012.46
- 57. Gogoi D, Chiplunkar SV. Targeting gamma delta T cells for cancer immunotherapy: bench to bedside. *Indian J Med Res* (2013) 138(5):755–61.
- Mookerjee-Basu J, Vantourout P, Martinez LO, Perret B, Collet X, Perigaud C, et al. F1-adenosine triphosphatase displays properties characteristic of an antigen presentation molecule for Vgamma9Vdelta2 T cells. *J Immunol* (2010) 184(12):6920–8. doi:10.4049/jimmunol.0904024
- Scotet E, Martinez LO, Grant E, Barbaras R, Jeno P, Guiraud M, et al. Tumor recognition following Vgamma9Vdelta2 T cell receptor interactions with a surface F1-ATPase-related structure and apolipoprotein A-I. *Immunity* (2005) 22(1):71–80. doi:10.1016/j.immuni.2004.11.012
- Harly C, Guillaume Y, Nedellec S, Peigne CM, Monkkonen H, Monkkonen J, et al. Key implication of CD277/butyrophilin-3 (BTN3A) in cellular stress sensing by a major human gammadelta T-cell subset. *Blood* (2012) 120(11):2269–79. doi:10.1182/blood-2012-05-430470
- Sandstrom A, Peigne CM, Leger A, Crooks JE, Konczak F, Gesnel MC, et al. The intracellular B30.2 domain of butyrophilin 3A1 binds phosphoantigens to mediate activation of human Vgamma9Vdelta2 T cells. *Immunity* (2014) 40(4):490–500. doi:10.1016/j.immuni.2014.03.003
- Chien YH, Konigshofer Y. Antigen recognition by gammadelta T cells. *Immunol Rev* (2007) 215:46–58. doi:10.1111/j.1600-065X.2006.00470.x
- Chen H, He X, Wang Z, Wu D, Zhang H, Xu C, et al. Identification of human T cell receptor gammadelta-recognized epitopes/proteins via CDR3delta peptide-based immunobiochemical strategy. *J Biol Chem* (2008) 283(18):12528–37. doi:10.1074/jbc.M708067200
- Kong Y, Cao W, Xi X, Ma C, Cui L, He W. The NKG2D ligand ULBP4 binds to TCRgamma9/delta2 and induces cytotoxicity to tumor cells through both TCRgammadelta and NKG2D. *Blood* (2009) 114(2):310–7. doi:10.1182/ blood-2008-12-196287
- 65. Dai Y, Chen H, Mo C, Cui L, He W. Ectopically expressed human tumor biomarker MutS homologue 2 is a novel endogenous ligand that is recognized by human gammadelta T cells to induce innate anti-tumor/virus immunity. *J Biol Chem* (2012) 287(20):16812–9. doi:10.1074/jbc.M111.327650
- Vantourout P, Mookerjee-Basu J, Rolland C, Pont F, Martin H, Davrinche C, et al. Specific requirements for Vgamma9Vdelta2 T cell stimulation by a natural adenylated phosphoantigen. *J Immunol* (2009) 183(6):3848–57. doi:10.4049/jimmunol.0901085
- Kozbor D, Trinchieri G, Monos DS, Isobe M, Russo G, Haney JA, et al. Human TCR-gamma+/delta+, CD8+ T lymphocytes recognize tetanus toxoid in an MHC-restricted fashion. *J Exp Med* (1989) 169(5):1847–51. doi:10.1084/jem.169.5.1847
- 68. Wright A, Lee JE, Link MP, Smith SD, Carroll W, Levy R, et al. Cytotoxic T lymphocytes specific for self tumor immunoglobulin express T cell

receptor delta chain. J Exp Med (1989) 169(5):1557–64. doi:10.1084/jem.169. 5.1557

- 69. Sciammas R, Bluestone JA. HSV-1 glycoprotein I-reactive TCR gamma delta cells directly recognize the peptide backbone in a conformationally dependent manner. *J Immunol* (1998) 161(10):5187–92.
- Rust CJ, Verreck F, Vietor H, Koning F. Specific recognition of staphylococcal enterotoxin A by human T cells bearing receptors with the V gamma 9 region. *Nature* (1990) 346(6284):572–4. doi:10.1038/346572a0
- Gasser S, Orsulic S, Brown EJ, Raulet DH. The DNA damage pathway regulates innate immune system ligands of the NKG2D receptor. *Nature* (2005) 436(7054):1186–90. doi:10.1038/nature03884
- Champsaur M, Lanier LL. Effect of NKG2D ligand expression on host immune responses. *Immunol Rev* (2010) 235(1):267–85. doi:10.1111/j.0105-2896.2010.00893.x
- Girlanda S, Fortis C, Belloni D, Ferrero E, Ticozzi P, Sciorati C, et al. MICA expressed by multiple myeloma and monoclonal gammopathy of undetermined significance plasma cells costimulates pamidronate-activated gammadelta lymphocytes. *Cancer Res* (2005) 65(16):7502–8. doi:10.1158/ 0008-5472.CAN-05-0731
- 74. Lanca T, Correia DV, Moita CF, Raquel H, Neves-Costa A, Ferreira C, et al. The MHC class Ib protein ULBP1 is a nonredundant determinant of leukemia/lymphoma susceptibility to gammadelta T-cell cytotoxicity. *Blood* (2010) 115(12):2407–11. doi:10.1182/blood-2009-08-237123
- Toutirais O, Cabillic F, Le Friec G, Salot S, Loyer P, Le Gallo M, et al. DNAX accessory molecule-1 (CD226) promotes human hepatocellular carcinoma cell lysis by Vgamma9Vdelta2 T cells. *Eur J Immunol* (2009) 39(5):1361–8. doi:10.1002/eji.200838409
- 76. Wrobel P, Shojaei H, Schittek B, Gieseler F, Wollenberg B, Kalthoff H, et al. Lysis of a broad range of epithelial tumour cells by human gamma delta T cells: involvement of NKG2D ligands and T-cell receptor- versus NKG2D-dependent recognition. *Scand J Immunol* (2007) 66(2–3):320–8. doi:10.1111/j.1365-3083.2007.01963.x
- 77. von Lilienfeld-Toal M, Nattermann J, Feldmann G, Sievers E, Frank S, Strehl J, et al. Activated gammadelta T cells express the natural cytotoxicity receptor natural killer p 44 and show cytotoxic activity against myeloma cells. *Clin Exp Immunol* (2006) 144(3):528–33. doi:10.1111/j.1365-2249. 2006.03078.x
- Baychelier F, Sennepin A, Ermonval M, Dorgham K, Debre P, Vieillard V. Identification of a cellular ligand for the natural cytotoxicity receptor NKp44. *Blood* (2013) 122(17):2935–42. doi:10.1182/blood-2013-03-489054
- 79. Lafont V, Liautard J, Liautard JP, Favero J. Production of TNF-alpha by human V gamma 9V delta 2 T cells via engagement of Fc gamma RIIIA, the low affinity type 3 receptor for the Fc portion of IgG, expressed upon TCR activation by nonpeptidic antigen. *J Immunol* (2001) 166(12):7190–9. doi:10.4049/jimmunol.166.12.7190
- Braza MS, Klein B, Fiol G, Rossi JF. gammadelta T-cell killing of primary follicular lymphoma cells is dramatically potentiated by GA101, a type II glycoengineered anti-CD20 monoclonal antibody. *Haematologica* (2011) 96(3):400–7. doi:10.3324/haematol.2010.029520
- Gertner-Dardenne J, Bonnafous C, Bezombes C, Capietto AH, Scaglione V, Ingoure S, et al. Bromohydrin pyrophosphate enhances antibody-dependent cell-mediated cytotoxicity induced by therapeutic antibodies. *Blood* (2009) 113(20):4875–84. doi:10.1182/blood-2008-08-172296
- Tokuyama H, Hagi T, Mattarollo SR, Morley J, Wang Q, So HF, et al. V gamma 9 V delta 2 T cell cytotoxicity against tumor cells is enhanced by monoclonal antibody drugs – rituximab and trastuzumab. *Int J Cancer* (2008) 122(11):2526–34. doi:10.1002/ijc.23365
- Capietto AH, Martinet L, Fournie JJ. Stimulated gammadelta T cells increase the in vivo efficacy of trastuzumab in HER-2+ breast cancer. *J Immunol* (2011) 187(2):1031–8. doi:10.4049/jimmunol.1100681
- Todaro M, D'Asaro M, Caccamo N, Iovino F, Francipane MG, Meraviglia S, et al. Efficient killing of human colon cancer stem cells by gammadelta T lymphocytes. *J Immunol* (2009) 182(11):7287–96. doi:10.4049/jimmunol.0804288
- Li Z, Xu Q, Peng H, Cheng R, Sun Z, Ye Z. IFN-gamma enhances HOS and U2OS cell lines susceptibility to gammadelta T cell-mediated killing through the Fas/Fas ligand pathway. *Int Immunopharmacol* (2011) 11(4):496–503. doi:10.1016/j.intimp.2011.01.001
- 86. Dokouhaki P, Schuh NW, Joe B, Allen CA, Der SD, Tsao MS, et al. NKG2D regulates production of soluble TRAIL by ex vivo expanded human

gammadelta T cells. *Eur J Immunol* (2013) 43(12):3175–82. doi:10.1002/ eji.201243150

- 87. Cipriani B, Borsellino G, Poccia F, Placido R, Tramonti D, Bach S, et al. Activation of C-C beta-chemokines in human peripheral blood gammadelta T cells by isopentenyl pyrophosphate and regulation by cytokines. *Blood* (2000) 95(1):39–47.
- Conti L, Casetti R, Cardone M, Varano B, Martino A, Belardelli F, et al. Reciprocal activating interaction between dendritic cells and pamidronate-stimulated gammadelta T cells: role of CD86 and inflammatory cytokines. *J Immunol* (2005) 174(1):252–60. doi:10.4049/jimmunol.174.1.252
- Dunne MR, Madrigal-Estebas L, Tobin LM, Doherty DG. (E)-4-hydroxy-3-methyl-but-2 enyl pyrophosphate-stimulated Vgamma9Vdelta2 T cells possess T helper type 1-promoting adjuvant activity for human monocytederived dendritic cells. *Cancer Immunol Immunother* (2010) 59(7):1109–20. doi:10.1007/s00262-010-0839-8
- Ismaili J, Olislagers V, Poupot R, Fournie JJ, Goldman M. Human gamma delta T cells induce dendritic cell maturation. *Clin Immunol* (2002) 103(3 Pt 1):296–302. doi:10.1006/clim.2002.5218
- Devilder MC, Maillet S, Bouyge-Moreau I, Donnadieu E, Bonneville M, Scotet E. Potentiation of antigen-stimulated V gamma 9V delta 2 T cell cytokine production by immature dendritic cells (DC) and reciprocal effect on DC maturation. *J Immunol* (2006) 176(3):1386–93. doi:10.4049/ jimmunol.176.3.1386
- Brandes M, Willimann K, Bioley G, Levy N, Eberl M, Luo M, et al. Crosspresenting human gammadelta T cells induce robust CD8+ alphabeta T cell responses. Proc Natl Acad Sci U S A (2009) 106(7):2307–12. doi:10.1073/ pnas.0810059106
- Brandes M, Willimann K, Moser B. Professional antigen-presentation function by human gammadelta T cells. *Science* (2005) 309(5732):264–8. doi:10.1126/science.1110267
- Schneiders FL, Prodohl J, Ruben JM, O'Toole T, Scheper RJ, Bonneville M, et al. CD1d-restricted antigen presentation by Vgamma9Vdelta2-T cells requires trogocytosis. *Cancer Immunol Res* (2014) 2(8):732–40. doi:10.1158/2326-6066.CIR-13-0167
- Maniar A, Zhang X, Lin W, Gastman BR, Pauza CD, Strome SE, et al. Human gammadelta T lymphocytes induce robust NK cell-mediated antitumor cytotoxicity through CD137 engagement. *Blood* (2010) 116(10):1726–33. doi:10.1182/blood-2009-07-234211
- Kryczek I, Wei S, Szeliga W, Vatan L, Zou W. Endogenous IL-17 contributes to reduced tumor growth and metastasis. *Blood* (2009) 114(2):357–9. doi:10.1182/blood-2008-09-177360
- Ngiow SF, Smyth MJ, Teng MW. Does IL-17 suppress tumor growth? Blood (2010) 115(12):2554–5; author reply 6–7. doi:10.1182/blood-2009-11-254607
- Muranski P, Restifo NP. Does IL-17 promote tumor growth? *Blood* (2009) 114(2):231–2. doi:10.1182/blood-2009-04-215541
- Du JW, Xu KY, Fang LY, Qi XL. Interleukin-17, produced by lymphocytes, promotes tumor growth and angiogenesis in a mouse model of breast cancer. *Mol Med Rep* (2012) 6(5):1099–102. doi:10.3892/mmr.2012.1036
- Futagbi G, Welbeck JE, Tetteh JK, Hviid L, Akanmori BD. Selective activation of TCR-gammadelta+ cells in endemic Burkitt's lymphoma. *Malar J* (2007) 6:69. doi:10.1186/1475-2875-6-69
- 101. Zhou J, Kang N, Cui L, Ba D, He W. Anti-gammadelta TCR antibodyexpanded gammadelta T cells: a better choice for the adoptive immunotherapy of lymphoid malignancies. *Cell Mol Immunol* (2012) 9(1):34–44. doi:10.1038/cmi.2011.16
- 102. Hannani D, Ma Y, Yamazaki T, Dechanet-Merville J, Kroemer G, Zitvogel L. Harnessing gammadelta T cells in anticancer immunotherapy. *Trends Immunol* (2012) 33(5):199–206. doi:10.1016/j.it.2012.01.006
- Glatzel A, Wesch D, Schiemann F, Brandt E, Janssen O, Kabelitz D. Patterns of chemokine receptor expression on peripheral blood gamma delta T lymphocytes: strong expression of CCR5 is a selective feature of V delta 2/V gamma 9 gamma delta T cells. *J Immunol* (2002) 168(10):4920–9. doi:10.4049/ jimmunol.168.10.4920
- 104. Thomas ML, Badwe RA, Deshpande RK, Samant UC, Chiplunkar SV. Role of adhesion molecules in recruitment of Vdelta1 T cells from the peripheral blood to the tumor tissue of esophageal cancer patients. *Cancer Immunol Immunother* (2001) 50(4):218–25. doi:10.1007/s002620100190

- Nedellec S, Bonneville M, Scotet E. Human Vgamma9Vdelta2 T cells: from signals to functions. *Semin Immunol* (2010) 22(4):199–206. doi:10.1016/j. smim.2010.04.004
- 106. Cipriani B, Knowles H, Chen L, Battistini L, Brosnan CF. Involvement of classical and novel protein kinase C isoforms in the response of human V gamma 9V delta 2 T cells to phosphate antigens. *J Immunol* (2002) 169(10):5761–70. doi:10.4049/jimmunol.169.10.5761
- 107. Correia DV, d'Orey F, Cardoso BA, Lanca T, Grosso AR, deBarros A, et al. Highly active microbial phosphoantigen induces rapid yet sustained MEK/Erk- and PI-3K/Akt-mediated signal transduction in anti-tumor human gammadelta T-cells. *PLoS One* (2009) 4(5):e5657. doi:10.1371/journal.pone.0005657
- Gomes AQ, Martins DS, Silva-Santos B. Targeting gammadelta T lymphocytes for cancer immunotherapy: from novel mechanistic insight to clinical application. *Cancer Res* (2010) 70(24):10024–7. doi:10.1158/0008-5472. CAN-10-3236
- 109. Groh V, Rhinehart R, Secrist H, Bauer S, Grabstein KH, Spies T. Broad tumor-associated expression and recognition by tumor-derived gamma delta T cells of MICA and MICB. *Proc Natl Acad Sci U S A* (1999) 96(12):6879–84. doi:10.1073/pnas.96.12.6879
- 110. Das H, Groh V, Kuijl C, Sugita M, Morita CT, Spies T, et al. MICA engagement by human Vgamma2Vdelta2 T cells enhances their antigen-dependent effector function. *Immunity* (2001) 15(1):83–93. doi:10.1016/ S1074-7613(01)00168-6
- 111. Rincon-Orozco B, Kunzmann V, Wrobel P, Kabelitz D, Steinle A, Herrmann T. Activation of V gamma 9V delta 2 T cells by NKG2D. *J Immunol* (2005) 175(4):2144–51. doi:10.4049/jimmunol.175.4.2144
- 112. Angelini DF, Zambello R, Galandrini R, Diamantini A, Placido R, Micucci F, et al. NKG2A inhibits NKG2C effector functions of gammadelta T cells: implications in health and disease. *J Leukoc Biol* (2011) 89(1):75–84. doi:10.1189/jlb.0710413
- 113. Ribot JC, Debarros A, Mancio-Silva L, Pamplona A, Silva-Santos B. B7-CD28 costimulatory signals control the survival and proliferation of murine and human gammadelta T cells via IL-2 production. *J Immunol* (2012) 189(3):1202–8. doi:10.4049/jimmunol.1200268
- 114. Ribot JC, debarros A, Silva-Santos B. Searching for "signal 2": costimulation requirements of gammadelta T cells. *Cell Mol Life Sci* (2011) 68(14):2345–55. doi:10.1007/s00018-011-0698-2
- 115. Ferrarini M, Delfanti F, Gianolini M, Rizzi C, Alfano M, Lazzarin A, et al. NF-kappa B modulates sensitivity to apoptosis, proinflammatory and migratory potential in short- versus long-term cultured human gamma delta lymphocytes. *J Immunol* (2008) 181(9):5857–64. doi:10.4049/jimmunol. 181.9.5857
- 116. Shao Z, Schwarz H. CD137 ligand, a member of the tumor necrosis factor family, regulates immune responses via reverse signal transduction. *J Leukoc Biol* (2011) 89(1):21–9. doi:10.1189/jlb.0510315
- 117. Kato Y, Tanaka Y, Tanaka H, Yamashita S, Minato N. Requirement of species-specific interactions for the activation of human gamma delta T cells by pamidronate. *J Immunol* (2003) 170(7):3608–13. doi:10.4049/ jimmunol.170.7.3608
- 118. Wang P, Malkovsky M. Different roles of the CD2 and LFA-1 T-cell co-receptors for regulating cytotoxic, proliferative, and cytokine responses of human V gamma 9/V delta 2 T cells. *Mol Med* (2000) 6(3):196–207.
- 119. Kato Y, Tanaka Y, Hayashi M, Okawa K, Minato N. Involvement of CD166 in the activation of human gamma delta T cells by tumor cells sensitized with nonpeptide antigens. *J Immunol* (2006) 177(2):877–84. doi:10.4049/ jimmunol.177.2.877
- 120. Castro MA, Nunes RJ, Oliveira MI, Tavares PA, Simoes C, Parnes JR, et al. OX52 is the rat homologue of CD6: evidence for an effector function in the regulation of CD5 phosphorylation. *J Leukoc Biol* (2003) 73(1):183–90. doi:10.1189/jlb.0902437
- 121. Hassan NJ, Simmonds SJ, Clarkson NG, Hanrahan S, Puklavec MJ, Bomb M, et al. CD6 regulates T-cell responses through activation-dependent recruitment of the positive regulator SLP-76. *Mol Cell Biol* (2006) 26(17):6727–38. doi:10.1128/MCB.00688-06
- Beetz S, Wesch D, Marischen L, Welte S, Oberg HH, Kabelitz D. Innate immune functions of human gammadelta T cells. *Immunobiology* (2008) 213(3–4):173–82. doi:10.1016/j.imbio.2007.10.006

- 123. Shojaei H, Oberg HH, Juricke M, Marischen L, Kunz M, Mundhenke C, et al. Toll-like receptors 3 and 7 agonists enhance tumor cell lysis by human gammadelta T cells. *Cancer Res* (2009) 69(22):8710–7. doi:10.1158/0008-5472.CAN-09-1602
- 124. Hayashi K, Altman A. Protein kinase C theta (PKCtheta): a key player in T cell life and death. *Pharmacol Res* (2007) 55(6):537–44. doi:10.1016/j. phrs.2007.04.009
- Garcia VE, Jullien D, Song M, Uyemura K, Shuai K, Morita CT, et al. IL-15 enhances the response of human gamma delta T cells to nonpeptide [correction of nonpetide] microbial antigens. *J Immunol* (1998) 160(9):4322–9.
- 126. Tsuda J, Li W, Yamanishi H, Yamamoto H, Okuda A, Kubo S, et al. Involvement of CD56brightCD11c+ cells in IL-18-mediated expansion of human gammadelta T cells. J Immunol (2011) 186(4):2003–12. doi:10.4049/ jimmunol.1001919
- 127. Moens E, Brouwer M, Dimova T, Goldman M, Willems F, Vermijlen D. IL-23R and TCR signaling drives the generation of neonatal Vgamma9Vdelta2 T cells expressing high levels of cytotoxic mediators and producing IFN-gamma and IL-17. *J Leukoc Biol* (2011) 89(5):743–52. doi:10.1189/jlb.0910501
- 128. Iwasaki M, Tanaka Y, Kobayashi H, Murata-Hirai K, Miyabe H, Sugie T, et al. Expression and function of PD-1 in human gammadelta T cells that recognize phosphoantigens. *Eur J Immunol* (2011) 41(2):345–55. doi:10.1002/ eji.201040959
- 129. Gertner-Dardenne J, Fauriat C, Orlanducci F, Thibult ML, Pastor S, Fitzgibbon J, et al. The co-receptor BTLA negatively regulates human Vgamma9Vdelta2 T-cell proliferation: a potential way of immune escape for lymphoma cells. *Blood* (2013) 122(6):922–31. doi:10.1182/blood-2012-11-464685
- 130. Lesport E, Baudhuin J, Sousa S, LeMaoult J, Zamborlini A, Rouas-Freiss N, et al. Inhibition of human gamma delta [corrected] T-cell antitumoral activity through HLA-G: implications for immunotherapy of cancer. *Cell Mol Life Sci* (2011) 68(20):3385–99. doi:10.1007/s00018-011-0632-7
- 131. Kuhl AA, Pawlowski NN, Grollich K, Blessenohl M, Westermann J, Zeitz M, et al. Human peripheral gammadelta T cells possess regulatory potential. *Immunology* (2009) 128(4):580–8. doi:10.1111/j.1365-2567.2009. 03162.x
- 132. Mao Y, Yin S, Zhang J, Hu Y, Huang B, Cui L, et al. A new effect of IL-4 on human gammadelta T cells: promoting regulatory Vdelta1 T cells via IL-10 production and inhibiting function of Vdelta2 T cells. *Cell Mol Immunol* (2016) 13(2):217–28. doi:10.1038/cmi.2015.07
- Murugaiyan G, Saha B. Protumor vs antitumor functions of IL-17. J Immunol (2009) 183(7):4169–75. doi:10.4049/jimmunol.0901017
- Peters C, Oberg HH, Kabelitz D, Wesch D. Phenotype and regulation of immunosuppressive Vdelta2-expressing gammadelta T cells. *Cell Mol Life Sci* (2014) 71(10):1943–60. doi:10.1007/s00018-013-1467-1
- Peters C, Hasler R, Wesch D, Kabelitz D. Human Vdelta2 T cells are a major source of interleukin-9. *Proc Natl Acad Sci U S A* (2016) 113(44):12520–5. doi:10.1073/pnas.1607136113
- 136. Lu Y, Hong B, Li H, Zheng Y, Zhang M, Wang S, et al. Tumor-specific IL-9producing CD8+ Tc9 cells are superior effector than type-I cytotoxic Tc1 cells for adoptive immunotherapy of cancers. *Proc Natl Acad Sci U S A* (2014) 111(6):2265–70. doi:10.1073/pnas.1317431111
- Quezada SA, Peggs KS. An antitumor boost to TH9 cells. Nat Immunol (2014) 15(8):703–5. doi:10.1038/ni.2945
- Vegran F, Apetoh L, Ghiringhelli F. Th9 cells: a novel CD4 T-cell subset in the immune war against cancer. *Cancer Res* (2015) 75(3):475–9. doi:10.1158/0008-5472.CAN-14-2748
- Qiu L, Lai R, Lin Q, Lau E, Thomazy DM, Calame D, et al. Autocrine release of interleukin-9 promotes Jak3-dependent survival of ALK+ anaplastic large-cell lymphoma cells. *Blood* (2006) 108(7):2407–15. doi:10.1182/ blood-2006-04-020305
- 140. Hoelzinger DB, Dominguez AL, Cohen PA, Gendler SJ. Inhibition of adaptive immunity by IL9 can be disrupted to achieve rapid T-cell sensitization and rejection of progressive tumor challenges. *Cancer Res* (2014) 74(23):6845–55. doi:10.1158/0008-5472.CAN-14-0836
- 141. Lv X, Feng L, Ge X, Lu K, Wang X. Interleukin-9 promotes cell survival and drug resistance in diffuse large B-cell lymphoma. J Exp Clin Cancer Res (2016) 35(1):106. doi:10.1186/s13046-016-0374-3

- 142. Carlsson A, Wingren C, Kristensson M, Rose C, Ferno M, Olsson H, et al. Molecular serum portraits in patients with primary breast cancer predict the development of distant metastases. *Proc Natl Acad Sci U S A* (2011) 108(34):14252–7. doi:10.1073/pnas.1103125108
- 143. Fournie JJ, Sicard H, Poupot M, Bezombes C, Blanc A, Romagne F, et al. What lessons can be learned from gammadelta T cell-based cancer immunotherapy trials? *Cell Mol Immunol* (2013) 10(1):35–41. doi:10.1038/cmi. 2012.39
- 144. Lang JM, Kaikobad MR, Wallace M, Staab MJ, Horvath DL, Wilding G, et al. Pilot trial of interleukin-2 and zoledronic acid to augment gammadelta T cells as treatment for patients with refractory renal cell carcinoma. *Cancer Immunol Immunother* (2011) 60(10):1447–60. doi:10.1007/s00262-011-1049-8
- 145. Bennouna J, Levy V, Sicard H, Senellart H, Audrain M, Hiret S, et al. Phase I study of bromohydrin pyrophosphate (BrHPP, IPH 1101), a Vgamma9Vdelta2 T lymphocyte agonist in patients with solid tumors. *Cancer Immunol Immunother* (2010) 59(10):1521–30. doi:10.1007/s00262-010-0879-0
- 146. Kobayashi H, Tanaka Y, Shimmura H, Minato N, Tanabe K. Complete remission of lung metastasis following adoptive immunotherapy using activated autologous gammadelta T-cells in a patient with renal cell carcinoma. *Anticancer Res* (2010) 30(2):575–9.
- 147. Kobayashi H, Tanaka Y, Yagi J, Minato N, Tanabe K. Phase I/II study of adoptive transfer of gammadelta T cells in combination with zoledronic acid and IL-2 to patients with advanced renal cell carcinoma. *Cancer Immunol Immunother* (2011) 60(8):1075–84. doi:10.1007/s00262-011-1021-7
- Deniger DC, Moyes JS, Cooper LJ. Clinical applications of gamma delta T cells with multivalent immunity. *Front Immunol* (2014) 5:636. doi:10.3389/ fimmu.2014.00636
- 149. Sicard H, Ingoure S, Luciani B, Serraz C, Fournie JJ, Bonneville M, et al. In vivo immunomanipulation of V gamma 9V delta 2 T cells with a synthetic phosphoantigen in a preclinical nonhuman primate model. *J Immunol* (2005) 175(8):5471–80. doi:10.4049/jimmunol.175.8.5471
- 150. Kalyan S, Chandrasekaran V, Quabius ES, Lindhorst TK, Kabelitz D. Neutrophil uptake of nitrogen-bisphosphonates leads to the suppression of human peripheral blood gammadelta T cells. *Cell Mol Life Sci* (2014) 71(12):2335–46. doi:10.1007/s00018-013-1495-x
- 151. Fowler DW, Copier J, Dalgleish AG, Bodman-Smith MD. Zoledronic acid causes gammadelta T cells to target monocytes and down-modulate inflammatory homing. *Immunology* (2014) 143(4):539–49. doi:10.1111/ imm.12331
- 152. Sugie T, Murata-Hirai K, Iwasaki M, Morita CT, Li W, Okamura H, et al. Zoledronic acid-induced expansion of gammadelta T cells from early-stage breast cancer patients: effect of IL-18 on helper NK cells. *Cancer Immunol Immunother* (2013) 62(4):677–87. doi:10.1007/s00262-012-1368-4
- 153. Kondo M, Izumi T, Fujieda N, Kondo A, Morishita T, Matsushita H, et al. Expansion of human peripheral blood gammadelta T cells using zoledronate. *J Visualized Exp* (2011) (55). doi:10.3791/3182
- 154. Starick L, Riano F, Karunakaran MM, Kunzmann V, Li J, Kreiss M, et al. Butyrophilin 3A (BTN3A, CD277)-specific antibody 20.1 differentially activates Vgamma9Vdelta2 TCR clonotypes and interferes with phosphoantigen activation. *Eur J Immunol* (2017) 47(6):982–92. doi:10.1002/eji. 201646818
- Franchini DM, Michelas M, Lanvin O, Poupot M, Fournie JJ. BTN3A1antibodies and phosphoantigens: TCRVgamma9Vdelta2 "see" the difference. *Eur J Immunol* (2017) 47(6):954–7. doi:10.1002/eji.201747058
- 156. Qi J, Zhang J, Zhang S, Cui L, He W. Immobilized MICA could expand human Vdelta1 gammadelta T cells in vitro that displayed major histocompatibility complex class I chain-related A-dependent cytotoxicity to human epithelial carcinomas. *Scand J Immunol* (2003) 58(2):211–20. doi:10.1046/j.1365-3083.2003.01288.x
- 157. Deniger DC, Maiti SN, Mi T, Switzer KC, Ramachandran V, Hurton LV, et al. Activating and propagating polyclonal gamma delta T cells with broad specificity for malignancies. *Clin Cancer Res* (2014) 20(22):5708–19. doi:10.1158/1078-0432.CCR-13-3451
- 158. Fisher JP, Yan M, Heuijerjans J, Carter L, Abolhassani A, Frosch J, et al. Neuroblastoma killing properties of Vdelta2 and Vdelta2-negative gammadeltaT cells following expansion by artificial antigen-presenting cells. *Clin Cancer Res* (2014) 20(22):5720–32. doi:10.1158/1078-0432.CCR-13-3464

- 159. Hiasa A, Nishikawa H, Hirayama M, Kitano S, Okamoto S, Chono H, et al. Rapid alphabeta TCR-mediated responses in gammadelta T cells transduced with cancer-specific TCR genes. *Gene Ther* (2009) 16(5):620–8. doi:10.1038/ gt.2009.6
- 160. Rischer M, Pscherer S, Duwe S, Vormoor J, Jurgens H, Rossig C. Human gammadelta T cells as mediators of chimaeric-receptor redirected anti-tumour immunity. *Br J Haematol* (2004) 126(4):583–92. doi:10.1111/j.1365-2141. 2004.05077.x
- Khan MW, Eberl M, Moser B. Potential use of gammadelta T cell-based vaccines in cancer immunotherapy. *Front Immunol* (2014) 5:512. doi:10.3389/ fimmu.2014.00512

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Xiang and Tu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.