



## The Role of Natural Antibodies to CC Chemokine Receptor 5 in HIV Infection

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The CC chemokine receptor 5 (CCR5) is responsible for immune and inflammatory responses by mediation of chemotactic activity in leukocytes, although it is expressed on different cell types. It has been shown to act as co-receptor for the human and simian immunodeficiency viruses (HIV-1, HIV-2, and SIV). Natural reactive antibodies (Abs) recognizing first loop (ECL1) of CCR5 have been detected in several pools of immunoglobulins from healthy donors and from several cohorts of either HIV-exposed but uninfected subjects (ESN) or HIV-infected individuals who control disease progression (LTNP) as well. The reason of development of anti-CCR5 Abs in the absence of autoimmune disease is still unknown; however, the presence of these Abs specific for CCR5 or for other immune receptors and mediators probably is related to homeostasis maintenance. The majority of anti-CCR5 Abs is directed to HIV binding site (N-terminus and ECL2) of the receptor. Conversely, it is well known that ECL1 of CCR5 does not bind HIV; thus, the anti-CCR5 Abs directed to ECL1 elicit a long-lasting internalization of CCR5 but not interfere with HIV binding directly; these Abs block HIV infection in either epithelial cells or CD4+ T lymphocytes and the mechanism differs from those ones described for all other CCR5-specific ligands. The Ab-mediated CCR5 internalization allows the formation of a stable signalosome by interaction of CCR5, β-arrestin2 and ERK1 proteins. The signalosome degradation and the subsequent de novo proteins synthesis determine the CCR5 reappearance on the cell membrane with a very long-lasting kinetics (8 days). The use of monoclonal Abs to CCR5 with particular characteristics and mode of action may represent a novel mode to fight viral infection in either vaccinal or therapeutic strategies.

Keywords: CC chemokine receptor 5, anti-CC chemokine receptor 5 antibodies, CC chemokine receptor 5 signalosome, HIV infection, HIV protection, CC chemokine receptor 5-based vaccine, CC chemokine receptor 5-based therapy

## INTRODUCTION

The CC chemokine receptor 5 (CCR5) belongs to G protein-coupled receptors (GPCRs), which represent the largest known superfamily of signal transducers and play functional roles in the response to exposure to light and odor as well as in cellular response to different types of signaling molecules (1). They consist approximately 4% of coded human genome (2) and represent one of the most important and largest groups of targets for therapeutics (3). Among them, the chemokine receptors

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are responsible for immune and inflammatory responses by mediation of chemotactic activity in leukocytes, even though they are expressed on a wide range of cell types, such as T and B cells, monocytes–macrophages, granulocytes, NK, DC, astrocytes, and neurons, and also on epithelium, endothelium, vascular smooth muscle, and fibroblasts (4–8).

CCR5 has also been implicated in hematopoiesis and it has been demonstrated that it act as co-receptor for the human and simian immunodeficiency viruses (HIV-1, HIV-2, and SIV) either independently of, or together with, the receptor CD4 (9–12). In particular, binding of viral gp120 of HIV-1 to CD4 triggers a conformational change in gp120 itself, which permits its binding to CCR5 and finally the viral entry into the cells (13, 14).

CCR5 is undoubtedly the main HIV-1 and HIV-2 co-receptor, involved in virus entry and cell-to-cell spread (15); interestingly, these R5-tropic viruses (CCR5 dependent strains) are associated with the initial infection (16), while HIV strains using the CXCR4 co-receptor are detected rarely in the early infection (11, 15, 17).

It is well known that chemokine receptor agonists, such as the  $\beta$ -chemokines RANTES (CCL5), MIP-1 $\alpha$  (CCL3), and MIP-1 $\beta$  (CCL4), inhibit HIV infection of susceptible cells *in vitro* (18–21).

Interestingly, the number of CCR5 molecules expressed on cell surface is correlated with the levels of viral infection (13) and it has been described a variation of the level of CCR5 molecules among individuals (15), which is due to both environmental and genetic aspects. Indeed, it has been shown that high levels of CCR5, in some developing countries such as Africa, is environmentally driven and it has been hypothesized that it is due to parasitic infections (22). Whereas a CCR5-negative phenotype has been described in either some subjects, which are resistant to HIV infection (exposed to HIV but seronegative subjects, so called ESN) or in Caucasians and in other ethnic groups worldwide; the reduced or absent expression of CCR5 in these populations has been attributed to a genetic mutation, named  $\Delta 32$ , a deletion of 32 base-pair in CCR5 gene that produces a truncated form of the receptor, which is not expressed on the cell membrane (23). Several clinical studies underlined that homozygous mutation affecting the expression of CCR5 confers a total resistance against HIV infection (24–28); whereas heterozygotes for CCR5 $\Delta$ 32 are not associated with complete HIV protection (15) but progress slowly in the infection, most likely due to the reduction of CCR5 levels on the cell surface (29). CCR5 $\Delta$ 32 is spontaneous in 4-18% of Askenazi Jews and European people but it has not been found in Pacific and Asian indigenes (21, 24, 25, 28); this mutation occurs mostly like a heterozygous defect on CCR5 gene (10-20%), with the highest frequencies in Nordic European countries, and only less than 1% is a homozygous mutation, as reported in several study population (24, 25, 30-33). In addition, it has been shown that the frequency of CCR5 $\Delta$ 32 genotype is higher also among ESN and HIV-infected individuals who control disease progression without treatment (so called LTNP) compared to HIV-1 treated seropositive subjects and people from the general population (34, 35). Of note, different levels of CCR5 expression among different individuals do not affect immune functions (36), in fact its absence is not associated with medical dysfunction (37). Nevertheless the prevalence of homozygosity for CCR5 $\Delta$ 32 mutation, which results in the absence of CCR5

expression, has been found increased in either West Nile infected subjects or in tick-borne encephalitis (38, 39) and reviewed in Venuti et al. (21).

More interestingly, anti-CCR5 natural Abs have been discovered and they also showed HIV-blocking properties (40–43).

Overall, several numbers of strategies aimed to the prevention of CCR5 function in the HIV entry has been developed and tested.

## MECHANISM OF GENERATION OF ANTI-SELF ANTIBODIES (Abs)

Natural Abs represent the first line of defense against pathogens; they are usually present in human serum as IgG3, IgM, and IgA and are generated in the absence of previous immune activation (44, 45).

The identification of B-1 cells (a subset of B cells), able to produce different self-reactive Abs, has demonstrated the capability of the immune system to interact with self-repertoire (45–47). It has been established that, in human, B cells are able to proliferate and to secrete Abs after exposure to lipopolysaccharide (LPS) from the Gram-negative bacteria membrane independent to the specific B-cell receptor (BCR) (48). Nevertheless, natural human Abs can also cross-react with microbial antigens, thus allowing host protection to pathogen independent of the previous microbial invasion (44).

B-1 cells are detected in the pleural and peritoneal cavity and represent the first line of defense, but they are present in the spleen and bone marrow as well, in which they secrete a higher proportion of circulating natural Abs (40, 45). The activation status of B-1 cells is BCR independent (49) and after their fast redistribution from the body cavities, B-1 cells are able to differentiate and to secrete abundant amounts of IgM and/or IgA (50).

The partial differentiation of B-1 cells and their ability to respond rapidly are fundamental for the Abs production to elucidate host protection to pathogens infection *via* mucosal surfaces and blood. In fact, the production of natural IgM at a steady state by B-1 cells represents a relevant protection against pathogen replication before the development of the antigenspecific response (40, 44, 45, 51, 52).

Many functions have been proposed for natural Abs such as a first line role in host defense and also a regulative part in homeostasis maintenance (40, 45, 53, 54). In addition, B-1 cells produce IgM that stimulate B-2 cells to elicit IgG (45, 55, 56), but they can also lead to induce the IgA production in response to antigen stimulation especially in the serum or in the intestinal lamina propria (40, 57).

Since the Eighties, when the AIDS was first described, several signals of autoimmune dysfunction were reported in subjects infected with HIV, such as B cell altered pathway, with production of high quantity of Abs and also of anti-cell Abs (58–60). These abnormalities, at the beginning, were related to HIV-vs-host activity but other pieces of evidence suggested that some anti-cell Abs may be considered like a host-vs-HIV reactions. Actually, it was shown that some broadly neutralizing human Abs produced during the HIV infection were autoreactive (61).

The finding led to suppose that immunotolerance mechanisms represent a disadvantage for these types of Abs (62, 63). Notably, the studies regarding the follow-up of HIV patients treated with three broadly neutralizing Abs, established that only one of them exhibited a low level of *in vivo* autoreactivity, while autoimmune-related adverse events were not detected in the study (64).

Many healthy donors displayed the presence of natural reactive Abs specific for CCR5 in several pools of immunoglobulins (41). Interestingly, different types of HIV-blocking Abs have been isolated from several cohorts of either ESN or HIV-infected individuals (40). The reason of development of anti-CCR5 Abs in the absence of autoimmune disease is still unknown; however, the presence of these Abs specific for CCR5 or for other immune receptors and mediators probably is related to homeostasis maintenance (40). Virus-induced alterations of self antigens can provide an increase of either auto-immunogenic proteins and the corresponding auto-Abs. Host factors itself, or other concomitant or latent viral infections, could activate these perturbations in the host cells, leading to conformational changes in host receptors and to remodeling from a self protein to a non-self antigenic epitope, as reviewed by Lopalco (40).

#### CCR5 AND ITS RELATED Abs

CCR5 shows a classic structure composed of seven transmembrane domains with N-terminus and three extracellular loops (ECL1, 2, and 3), which have immunogenic properties. The two longer domains (N-terminus and ECL2) are recruited for HIV binding (65–67). Its preferential ligands are MIP-1 $\alpha$ , MIP-1 $\beta$ , and RANTES and the binding of these molecules could interfere sterically with the viral envelope protein (Env) gp120 of HIV binding resulting in an inhibition of viral infection (15). An alternative model of protection is that ligand-induced chemokine receptor internalization eliminates the co-receptor from the cell surface (68); obviously, these two mechanisms are not mutually exclusive.

Anti-CCR5 natural Abs were found also in individuals with  $\Delta$ 32 mutation, sexual partners of subjects who were wild type for CCR5 gene, thus suggesting that CCR5 can be considered as an alloantigen (40, 42, 69, 70). Moreover, hemophilic patients subjected to continuous blood transfusions, ESN and LTNP show Abs to CCR5 directed specifically to the first external loop (ECL1) (21, 29, 41–43, 69, 71–75); these natural Abs have been identified in serum and also in other biological fluids, such as semen, cervicovaginal secretion and saliva in subject with different genetic background (75).

The majority of anti-CCR5 Abs is directed to HIV binding site (N-terminus and ECL2) of the receptor. Conversely, Abs to ECL1–CCR5, which induce a long-lasting internalization of the receptor (29), are capable to block HIV infection in either CD4+ T lymphocytes or epithelial cells, this latter one through transcytosis, which mimics mucosal transmission (76) and this mechanism differs from that induced by all the other ligands directed to CCR5 (40). First of all, the natural Abs recognize ECL1 whereas CCR5 agonists specifically bind to the ECL2 of CCR5. Second and more important, the long-lasting internalization of CCR5 with natural anti-CCR5 Abs seems to be a unique mechanism not demonstrated for other CCR5 modulating molecules so far. Indeed, by using monoclonal antibodies (mAbs) that recognize the N-terminus and the second loop of CCR5, it has been shown a differentially modulation of receptor activity; thus suggesting that each CCR5 extramembrane region can display different properties (65, 77, 78).

A clinical study, related to the presence and the activity of Abs to ECL1 in the sera of some LTNP, clearly demonstrated that the loss of these Abs observed during the follow-up of these subjects was significantly associated with the clinical progression of the disease (29). Moreover, in another studies, a total of 206 Asian and Caucasian ESN subjects have been tested for the presence of anti-CCR5 Abs directed to ECL1 and 9% resulted positive (43, 75, 79), similar percentage (9.8%) have been found in different cohorts of HIV seropositive subjects (total subjects 336) (29, 80), although only in LTNP anti-CCR5 Abs have been associated with resistance and showed anti HIV property in vitro (29, 81). Strikingly, anti-CCR5-ECL1 Abs resulted HIV protective only when they were directed to a conformational epitope within ECL1 loop (43, 75). A total of 325 healthy controls have even analyzed as well but none resulted positive for anti-CCR5 Abs, thus suggesting that these Abs could be elicited by low levels of viral antigenic stimulation; that could explain why these Abs have been found in ESN and LTNP people but not in subjects who were not exposed to HIV or progressed and developed AIDS. Another hypothesis could be that anti-CCR5 Abs are elicited during other antigenic stimulations (different from HIV), which induce alterations of self-repertoire, thus eliciting anti-self responses. Finally, the priming due to endogenous retroviral proteins, which share homology with HIV env protein, could elicit in some HIV-exposed subjects a specific immune response.

Of note, these ECL1 specific Abs do not induce alteration in immune functions, as demonstrated by healthy subjects with anti-CCR5 Abs (45) or by elicited anti-CCR5 Abs in animal models such as mice and macaques (82–84) as further specified in the section of CCR5 immunization as vaccination strategy.

The ECL2 domain represents the binding site for both HIV and chemokines, so the Abs that recognize this site can prevent chemokine binding and/or signaling (66), although N-terminus is specific for viral binding only. For example, 2D7 is one of the most potent mAb directed to ECL2 that blocks HIV-1 entry into CD4 T cells, but not the transcytosis carried out with epithelial cells (66, 76, 85). An anti-CCR5 mAb named PRO140 is a humanized mAb that targets a conformational epitope between N-terminus and ECL2 and it deeply blocks viral entry (86). Another fully human IgG4 mAb with a strong activity against various HIV-1 isolates is CCR5mAb004 (87).

A recent study has demonstrated for the first time that the region designated as the membrane-proximal region (MPR), between the N-terminus and the ECL1, is important for HIV-1 infections (16). In fact, the Abs directed to this epitope block the infection of R5-tropic HIV-1 without affecting X4-tropic strain; furthermore, the substitution of MPR with the equivalent region of CCR2b, CXCR4, or CCR3 significantly abrogates viral infection (16). Both these findings provide an argument against the possible use of a target therapy with CCR5-specific Abs.

## ENDOCYTOSIS AND *DE NOVO* SYNTHESIS OF CCR5 WITH NATURAL ANTI-CCR5 Abs

Ligands binding to CCR5 leads to conformational changes, which include desensitization and internalization (88). Two major mechanisms of rapid receptor regulation have been distinguished, specifically homologous (agonist-specific) and heterologous (agonist-nonspecific) desensitization, and both mechanisms are really important in fine tuning leukocyte responses (89, 90). Homologous desensitization requires phosphorylation of the receptor binding mediated by members of the GPCR kinases (GRK) family (91). This in turn leads to the association of  $\beta$ -arrestin1/2 with the receptor and to desensitization via uncoupling of the receptor and G protein (77, 92); in particular,  $\beta$ -arrestins bound physically with the receptors and initiate endocytosis through clathrin-coated vescicles and also act as scaffold proteins in crosstalk with other signaling pathways (93). Conversely, heterologous desensitization is traditionally defined as a state of cellular refractoriness to different agonists after receptor phosphorylation sites different from GRK mediated by second messenger-activated protein kinases, such as PKC (90).

CCR5 internalization can also induce a different second pathway, which recruits caveolae and it is independent of clathrincoated pits. Caveolae are microdomains able to be internalized under precise conditions or in a controlled manner (13, 94).

It is well known that, after endocytosis, the GPCR proteins are also classified in receptors that are recycled, slowly or rapidly, to the cell membrane after their resensitization and those that should be degraded (77, 95–97). CCR5 is usually recycled after desensitization (4): after stimulation with natural ligands, CCR5 is internalized into the trans-Golgi network (TGN) *via* the endosome recycling compartment (ERC) (98) and, when the resensitization process is complete, it can return to the cell surface (4, 98). However, rare examples of post-endocytic sorting for GPCRs mediated by ligands have been reported (77, 99–101).

Bönsch and colleagues have recently shown that different ligands of the same GPGR are able to induce different phosphorylation pathways, which may be a relevant factor for the interaction with  $\beta$ -arrestins (77, 102). In addition, ligands trigger a characteristic short-term kinetics of CCR5 internalization, which transiently involves  $\beta$ -arrestins with consequent rapid recycling or degradation on the cell membrane; conversely, natural anti ECL1-CCR5 Abs induce a specific long-lasting kinetics of CCR5 internalization (29) with the recruitment of an ERK1-mediated pathway (70, 77). Of note, a hitherto unrecognized mechanism of CCR5 modulation mediated by G-protein-dependent ERK1 was comprehensively reported; in particular, natural anti-CCR5 Abs led to activation of ERK1 which is localized predominantly in the cytosol and it interacts directly with the CCR5 protein, thus inducing the degradation of CCR5 with a consequent de novo synthesis (70); the re-expression of CCR5 on the cell surface needs several days (70). This finding is actually important for the design of suitable microbicide or therapeutic tool that could inhibit HIV infection for several days after application by using a

Furthermore, it is largely reported that GPCRs, considering the stability of interaction with  $\beta$ -arrestins after agonist stimulation, can be functionally divided into two general classes: (i) "Class A" receptors, such as  $\beta 2$  adrenergic receptor ( $\beta 2AR$ ), develop transient complexes with  $\beta$ -arrestins transiently ubiquinated and with weak activation of ERK1/2; by contrast, (ii) "Class B" receptors, such as vasopressin V2 receptor ( $V_2R$ ), develop tight receptor- $\beta$ arrestins complexes, regulated by its constant ubiquitination and a durable activation of ERK1/2 which is located mainly into the endosomes. Endosomes complexes containing activated GPCRs, activated and ubiquitinated β-arrestins and phosphorylated ERK are called "signalosome" (77, 102, 103). In fact, it is well understood that the ubiquitination status of  $\beta$ -arrestin has a relevant role for its interaction with proteins responsible for endocytosis (e.g., clathrin) and for signaling (e.g., ERK1/2), and influences temporal and spatial dissociation of the complex (104-108). Overall, CCR5 is classified as a "Class A" receptor, but stimulation with anti-CCR5 Abs lead to the translation into a very long-lasting Class B type (77, 102, 106).

Very recently, it has been published the different ability of two RANTES analogous (5P14 and PSC) to induce the development of stable complexes between CCR5 and  $\beta$ -Arrestin1. Briefly, PSC-RANTES is able to induce a long-duration of recruitment of  $\beta$ -Arrestin1 to CCR5 compared to 5P14-RANTES, which elicits a temporary recruitment. Notably, the experiments have been carried out and the results assessed at short time only (50 min) (1). Therefore, it is possible to determine the fate of the internalized receptor by the aid of specific CCR5-ligands, suggesting that the stability of ligand-induced receptor-arrestin complexes has a crucial role in the sorting mechanism (1, 77).

In a very relevant way, these published data underline that the binding of natural Abs induces modifications in CCR5 signaling, which leads ligand-induced post-endocytic sorting in a very long-lasting Class B trafficking (77). Furthermore, in T cell, anti-CCR5 Abs that recognize ECL1 are able to induce a CCR5-negative phenotype, ERK1-mediated, by the strong support of  $\beta$ -arrestin2 (as shown in **Figure 1**); otherwise, it is possible that this mechanism could be specific for T cells only (77, 109).

# INDUCTION OF ANTI-CCR5 Abs AS VACCINATION STRATEGY

Published data, obtained in mice and macaques, demonstrate the capability of either anti-CCR5 Abs to display HIV-blocking properties or vaccines against CCR5 to prevent the problem of virus variability and viral escape (82, 110–113). Accordingly, the development of Abs as functional inhibitors of CCR5 is the big goal that could be reached, since Abs can provide protection by causing very low toxicity (113). Several groups have investigated the possibility to use *in vivo* Abs specific to CCR5 (82, 83, 111, 112, 114–116). Interestingly, when a long-term intranasal immunization was performed, it has elicited specific IgA and IgG in both mucosal secretions and sera of the immunized mice. Such systemic and mucosal Abs induce a CCR5-negative



phenotype on both peripheral and mucosal cells, thus blocking HIV replication in vitro (111). In accordance with this result, the use of ECL1-CCR5 peptide, chimeric-generated in the context of the capsid protein of Flock House Virus, elicits Abs able to induce CCR5 internalization and re-expression with a very slow kinetics which needs 4 weeks after immunization to be recovered (82). Furthermore, in a subsequent study, it has been published that the substitution of amino acids within ECL1 in position 95 and 96 elicited Abs, which induced stronger long-lasting internalization of CCR5, whereas amino acid substitutions in position 92, 98 and 99 abrogated biological activity of such Abs (112), thus highlighting the importance of the epitope in driving different trafficking pathway. Moreover, in a recent study performed in mice, several aspects of anti-CCR5 immunization, including the use of all the extramembrane domains of CCR5 have been tested, to better understand the ideal schedule to reach long-lasting and strong immune responses. Interestingly, ECL1 and ECL2 showed stronger responses compared to the N-terminus; they achieved nearly complete CCR5 downregulation, and they blocked HIV infection (82). In addition, in this study was not observed any immune dysfunction in T cell responses or histopathological alterations in organs and tissues in relation to the presence or

the induction of Abs specific for CCR5. The possibility of longterm toxicity and any functional impact of anti-CCR5 Abs needs additional studies; however, the findings showed in this latter study are supported by other published studies, where no adverse events were reported in CCR5-immunized macaques after 3 years of follow-up (84). In addition, it has recently published that the prophylactic immunization of macaques with virus-like particle specific for two CCR5 regions is safe and immunogenic and is capable to reduce highly virus replication in a subset of the animals (83). On the other hand, Bogers and colleagues used an immunization approach to target both virus and CCR5 (three extracellular peptides of CCR5, an N-terminal HIV gp120 fragment generated in transgenic plants and recombinant SIV p27) (117); this strategy of vaccination showed a significant block of the virus infection by eliciting good serum and vaginal quantity of Abs (117). More recently, Peabody et al. demonstrated that the immunization with recombinant vectors, which enable the CCR5-ECL2 region to recreate its native conformation, overcomes the issue of tolerance and induces the appropriate immune response (118).

Although several strategies aimed at inducing a CCR5-negative phenotype to prevent HIV-1 entry, the earlier immunization

studies in macaques observed little or no protection against SIV challenge (116, 118), probably due to poor selection of CCR5 antigen or to the correct peptide sequence in the wrong conformation. Indeed, it has previously demonstrated that immunization with ECL1 domain, in a linear conformation, does not elicit serological Abs responses that bind to the native molecule (111) and, moreover, in macaques, the immunization with ECL2 in its native conformation induces immune responses with expected properties (84). Nevertheless, Chain and colleagues have recently defined a new linear epitope of CCR5 within the N-terminus domain recognized by two independently produced mAbs; in particular, they found that RoAb13 Ab is capable to bind to both linear peptide and native form of the epitope and the sulfation of tyrosines at CCR5 N-terminus enhanced its binding to the peptide (119). RoAb13 has been previously reported to block HIV infection (120) but also blocks migration of monocytes after the chemokine binding to CCR5 or in the presence of inflammatory macrophage conditioned medium (119).

A significant challenge in the design of anti-CCR5 Abs is that they must be purely "blocking Abs" that either bind to the epitope in such a way to occlude the viral receptor or Abs binding results in receptor internalization. The most effective anti-pathogen Abs are able to engage host defense mechanisms, such as Complement or ADCC (Antibody-Dependent Cell-mediated Cytotoxicity), thus resulting protective against HIV infection (121) although these functions could result in inhibition of the effectiveness of immune responses. Moreover, as reported by Pastori et al., it is possible to elicit the production of murine serum anti-ECL1-CCR5 Abs at levels 300-fold greater than those found in humans and that the quantity of murine CCR5-specific immunoglobulins reached 50% of total Igs (82). It is noteworthy that such HIV-1 blocking Abs are present in serum and mucosal fluids from subjects with different genetic backgrounds (75), thus suggesting that it is possible to elicit these Abs in subjects coming from both developing as well as developed countries. In addition, an individual who received a stem cell transplant from a CCR5-negative donor, for acute myeloid leukemia treatment, is believed to be the only patient to have been cured of HIV (119, 122).

## ANTI-CCR5 Abs IN THE IMMUNE-PROPHYLAXIS AGAINST HIV INFECTION

The Abs can prevent viral infection by several mechanisms of action: (1) can directly block virus attachment to the cell by leading the Abs to bind either virus or receptor and/or co-receptor on host cells; (2) can block fusion at cell surface at the post-binding/ pre-fusion state as well (87). For reducing the development of viral escape variant, it has been highly considered to target the conserved cellular receptors, such as CCR5, for treatment of HIV infection. In particular, as HIV needs the presence of one co-receptor in dependence of the strain (CCR5 and/or CXCR4) in association with the receptor CD4, mAbs against cellular proteins have been developed and are being tested in clinical trials. A humanized mAb directed to CD4, named ibalizumab, exert an antiviral property not inhibiting the binding of gp120 but by a post-binding conformational effects, which prevents the interaction between CD4-gp120 and CXCR4 or CCR5 (123,

124). Three clinical trials have been reported, which underlined its efficacy (87). For sure, one emerging therapy is based on the use of CCR5-specific Abs; in particular, CCR5mAb004 appears safe and effective in the reduction of viral load when tested in clinical trials (87). Interestingly, another study involving the mAb PRO140 showed virologic suppression without blocking the response of the receptor to chemokines; however, the highest tolerated dose of this mAb has not been determined, proposing a substantial margin of safety for PRO140 in dependence of the site of administration (87, 125). In all these clinical trials, the use of anti-CCR5 Abs did not induce any alterations in other lymphocyte functions, thus confirming their safety.

Of note, the use of Abs instead of chemokines or classical antiretroviral therapy could reduce the complication related to drugs resistance and also the unwanted interactions with redundant CCR receptors. For example, ST6 is a Fab fragment obtained from a mAb specific for a unique sequence of N-terminus CCR5 and it was engineered in a single-chain antibody (scFv) fused with an ER retention peptide; the usage of such scFv by intracellular immunization was able to downregulate the receptor from cell membrane both in macaques and in human cells, whereas the expression of CXCR4 was not affected. Moreover, the modified cells were not infected with R5-HIV (126). In a subsequent study, it has been demonstrated that transformed primary T cells, with a CCR5 intrabody (an Ab that binds its receptor at intracellular level), were resistant to HIV infection (21). Finally, scFvs directed to CCR5 were utilized, as well, to lead viral pseudotyped lentiviral vectors to cells that express CCR5 (127).

Very recently emerged the evidence that combinations of HIV-blocking Abs will likely be more effective that single one as reviewed by Margolis (128). Alternatively, the bio-engineering, which generates Abs either with different specificities (129) or anchored to target cells (130), has given a proof of concept to generate more potent HIV-blocking Abs.

## OTHER STRATEGIES AIMED AT BLOCKING HIV INFECTION THROUGH CCR5

Anti-CCR5 strategies include also the utilization of small molecule drugs, such as Maraviroc, which binds in the transmembrane regions of CCR5 and it is a functional antagonist that prevents CCR5 signaling from cell surface and even if it is currently in clinical trials (131), it has been approved for use in many jurisdictions.<sup>1</sup> Nevertheless, there is low enthusiasm to utilize it as front-line therapy in HIV-infected patients (23), thus it is currently in use in HIV treatment-multiexperienced patients only (132). Moreover, HIV-1 escape mutants to Maraviroc have been described and reviewed by Harada and Yoshimura (133).

Since the discovery that natural ligands of CCR5 (RANTES, MIP-1 $\alpha$ , and MIP-1 $\beta$ ) show anti-HIV activity (1, 19, 86, 134, 135), a large numbers of modified analogs have been tested due to their short half-lives (<10 min) (134, 136) but no one has been tested in human clinical trial due to low antiviral activity *in vivo*. The most

<sup>&</sup>lt;sup>1</sup>https://aidsinfo.nih.gov/guidelines/search/1/CELSENTRI/0.



promising described so far was PSC-RANTES that shows several non-natural, non-coded structures in the N-terminal region (137, 138). It displays an important inhibition of HIV entry, CCR5 dependent, *in vitro* (137) and also a full protection against R5-tropic SHIV infection in a macaque vaginal challenge model (139); although this high potency *in vitro*, it requires high concentration to give protection in macaques (138, 139). Considering that it is capable to induce an intracellular sequestration of CCR5 longer than RANTES, it could be helpful for topical HIV prevention (140). Using a strategy based on phage display, Gaertner and collaborators obtained three different modified PSC-RANTES, which exhibit only natural amino acids: 6P4-RANTES, which prolongs the intracellular sequestration of CCR5; 5P12-RANTES has no detectable G protein signaling and does not bring about receptor sequestration; and 5P14-RANTES, which induces the internalization of CCR5 with no detectable G protein-linked signaling activity (138). Another relevant RANTES derivative is named AOP-RANTES and it was obtained by first generating an aldehyde-like group at the NH2-terminus of RANTES and then reacting with aminooxypentane; it is able to block R5-tropic strain infection on macrophages *in vitro* (141). AOP-RANTES induces >90% downregulation of cell membrane expression of

CCR5 on monocytes/macrophages, lymphocytes and inhibits CCR5 recycling on cell surface whereas RANTES does not (142).

As HIV entry process requires expression of both CCR5 and CD4 on cell membrane, receptor- and co-receptor-mimetic peptides (143, 144) have been proposed as an alternative strategy to block HIV entry but, as for chemokines, no one has been already tested in human clinical trial.

A summary of the immunologic approaches that use CCR5 as target to block HIV transmission/infection is showed in **Figure 2**.

Hematopoietic stem cell transplant using a CCR5 $\Delta$ 32 donor led to the only known cure of HIV-1 infection (122, 145) and T cells treated with engineered nucleases that introduce mutations at the CCR5 locus are resistant to HIV (146–150), accelerating ongoing efforts to develop gene editing- and cell-based therapeutic agents for HIV (15, 151, 152).

Another promising method of gene editing is the use of CRISPR/Cas9 system (Clustered Regularly interspaced palindromic repeats sequences) to target human cells for the disruption of CCR5 gene, otherwise the off-targeting is still a major limit to be overcome (153–155). Furthermore, DNA binding proteins, for example, the transcription activator-like effectors (TALEs), which are vegetal proteins, have been used *in vitro* and showed effects similar to those obtained with engineered nuclease (156).

Zinc finger nucleases (ZFNs) are other common and versatile DNA binding proteins utilized in several cell types. In addition, CCR5–ZFN-modified autologous CD4+ T lymphocytes have been used in a phase I clinical trial and this approach resulted safe (149).

To shutdown CCR5 expression, several RNA-based technologies have been used also with good results, such as RNA silencing (siRNA), antisense RNAs targeting different cellular and viral genes or ribozymes with catalytic activity (157–159); in particular, pseudotyped lentivirus and adenoviruses vectors have been used with good results for transducing siRNA-coding sequence into the cells. In the same way to that described for gene editing, off-targeting activity and over-expression of antisense RNA could cause a toxic effect (160) and could activate innate immune response as well (161).

### CONCLUSION

The incidence of natural allo- or auto-responses in healthy people, without symptoms or signals of autoimmune disease, and also the capability of eliciting and maintaining strong and longlasting HIV-blocking Abs in animal models, suggests that some autoimmune mechanisms could be positively utilized to give a better protection or a higher response to HIV in HIV-exposed individuals and in HIV-positive subjects. Allo- and auto-immune responses could allow a new key to analyze HIV tricks in immune escape and offer unexploited strategies to fight HIV with its own arms. CCR5 is the most important co-receptor in the early stages of infection, and half or more of all infected individuals move to AIDS harboring only CCR5 (R5)-tropic viruses. Epidemiology studies clearly established that CCR5 plays a crucial role in the transmission and pathogenesis of HIV *in vivo*.

As in CCR5-defective individuals were not found inflammatory and immune alterations or disfunctions, CCR5 has been defined as a redundant molecule in humans (12, 141, 162, 163), and as the variability of HIV *env*, CCR5 has become a relevant target to generate drugs and immune modulatory molecules to block HIV transmission and subsequent infection.

Overall, these findings together with the data reported for *in vivo* (clinical trials) and *in vitro* (laboratory findings) studies support the view that CCR5 could represent an excellent target to fight HIV and a good alternative to classical antiviral approaches, although it should be taken into account the concomitant geographical location of  $CCR5\Delta32$  and other pathologies, such as West Nile infection or tick-borne encephalitis.

The development of a sterilizing vaccine capable to prevent HIV infection totally is the highest and the most expected effort, still far from being reached. Over the past 30 years, there has been a huge global effort to develop an effective prophylactic vaccine against HIV/AIDS. This is a significant challenge since no previously licensed vaccine in current use has been designed without the presence of a significant "convalescent population," i.e., patients who have been patently infected and demonstrated subsequent clearance of the pathogen. Such a patient population usually supplies critical information for characterizing adaptive immunological responses associated with "protection." One of the main reasons of failure in developing an effective AIDS vaccine could be the mainstream concept that the most relevant information derive from studying the immune responses in patients who have not cleared the virus. Thus, the design of a CCR5-based vaccine, which takes advantage of data generated in a small but significant clinical cohorts of individuals such as ESN or LTNP could represent an excellent target to generate new vaccination strategy, as these subjects represent a sort of vaccinated/cured subjects and this protective status can be induced and reproduced in all subject. It is relevant underline that natural anti-CCR5 Abs reproduce a protective status similar to that one observed for  $\Delta 32$  mutation, although an approach based on CCR5 vaccine in individuals who can contract HIV infection may be a more possible and safe goal compared to gene therapy, taking into account the HIV epidemiology and the trouble of implementing CCR5 gene therapy in people living in developing countries.

Nowadays, there are many antiviral drugs used in therapy but the most related problem is the development of drug-resistant strain of virus that invalidates the positive effects obtained with the therapy utilized. Conversely, the possibility of using monoclonal Abs as therapy, with particular characteristics and mode of action, may represent a novel mode to fight viral infection disease. Overall, Abs show low toxicity together with high specificity and versatility.

It is well known that the first effective treatment of infectious disease was the "serum therapy" (administration of hyperimmune sera from immunized animals or human donors) and only after the discovery of antibiotic therapy in association with the development in vaccine design, this treatment was abandoned for mostly of infections (87, 164, 165).

The possibility of usage of Abs in clinical practice was opened from the opportunity of generate and manipulate Abs with different specific epitope recognition, such as the mAbs (87). In fact, in the last years, mAbs have begun a new class of clinical drug utilized in inflammatory diseases, immunology, and oncology; only their development for infection treatment is going slowly.

Strategies aimed to prevent infection, such as usage of condoms, represent another effective line of defense to fight the HIV epidemic. However, social and ethnic "barriers" impede effective protection of many people. Therapeutic Abs to CCR5 could offer an alternative for primary prevention of HIV and their availability would greatly empower women/men to protect themselves and their partners. Indeed, Abs formulated as a topical product could control the disease without affecting social and procreation aspects. In addition, proceeding directly at the HIV transmission level, the passive immunotherapy approach will help to prevent and reduce both further infection and disease incidence, respectively.

Other strategies involve ART (Anti Retroviral Therapy), which is a strong treatment program utilized to suppress HIV viral replication and the progression of HIV disease. The typical regimen combines three or more different drugs, such as nucleosidic or non-nucleosidic inhibitors of reverse transcriptase, protease, and integrase inhibitors. ART is the only current available treatment for HIV patients and it is being used in many developing countries with the help of WHO.<sup>2</sup> Nevertheless, it has limitations in terms of high cost, intolerance, bad compliance, and insurgence of resistance (166, 167).

For this reason, a new strategy has emerged to identify blocking Abs against the HIV receptors or co-receptors, either as active-immunizations such as a vaccine or passive-immunizations such as the use of CCR5-based immuno-prophylaxis.

<sup>2</sup>WHO | Ten years in public health 2007-2017. WHO Available at: http://www.who. int/publications/10-year-review/dg-letter/en/.

#### REFERENCES

- Bönsch C, Munteanu M, Rossitto-Borlat I, Fürstenberg A, Hartley O. Potent anti-HIV chemokine analogs direct post-endocytic sorting of CCR5. *PLoS One* (2015) 10:e0125396. doi:10.1371/journal.pone.0125396
- Fredriksson R, Lagerström MC, Lundin L-G, Schiöth HB. The G-proteincoupled receptors in the human genome form five main families. Phylogenetic analysis, paralogon groups, and fingerprints. *Mol Pharmacol* (2003) 63:1256–72. doi:10.1124/mol.63.6.1256
- Jacoby E, Bouhelal R, Gerspacher M, Seuwen K. The 7 TM G-protein-coupled receptor target family. *ChemMedChem* (2006) 1:761–82. doi:10.1002/ cmdc.200600134
- Signoret N, Pelchen-Matthews A, Mack M, Proudfoot AE, Marsh M. Endocytosis and recycling of the HIV coreceptor CCR5. *J Cell Biol* (2000) 151:1281–94. doi:10.1083/jcb.151.6.1281
- Pelchen-Matthews A, Signoret N, Klasse PJ, Fraile-Ramos A, Marsh M. Chemokine receptor trafficking and viral replication. *Immunol Rev* (1999) 168:33–49. doi:10.1111/j.1600-065X.1999.tb01281.x
- Murphy PM, Baggiolini M, Charo IF, Hébert CA, Horuk R, Matsushima K, et al. International union of pharmacology. XXII. Nomenclature for chemokine receptors. *Pharmacol Rev* (2000) 52:145–76.
- Rottman JB, Ganley KP, Williams K, Wu L, Mackay CR, Ringler DJ. Cellular localization of the chemokine receptor CCR5. Correlation to cellular targets of HIV-1 infection. *Am J Pathol* (1997) 151:1341–51.
- Barmania F, Pepper MS. C-C chemokine receptor type five (CCR5): an emerging target for the control of HIV infection. *Appl Transl Genom* (2013) 2:3–16. doi:10.1016/j.atg.2013.05.004

Interestingly, natural human Abs that recognize the ECL1 of the receptor induce a long-lasting internalization of CCR5 by triggering the recruitment of  $\beta$ -arrestin2; this event induces the accumulation of the two proteins (CCR5 and  $\beta$ -arrestin2) into the cytoplasm and leads to the activation of ERK1, which is retained into the cytosol as well. This stable CCR5 signalosome persists into the cells at least 48 h; after that, it may be targeted for degradation with consequent *de novo* synthesis of the proteins complex and, consequently, CCR5 reappears on the cell membrane with long-lasting kinetics (8 days) (70, 77). This particular mechanism could be used for designing molecules that work synergistically for stable maintenance of the signalosome into the cells and for driving the complex to degradation; thus permits to reach a longlasting CCR5 disappearance from cell membrane which could inhibit HIV infection for a long time.

These findings may support the discovery of innovative therapeutic tools where CCR5 is an important player for microbial control and/or elimination (168) and as well as for the regulation T cell function in autoimmune diseases, such as rheumatoid arthritis, type 1 diabetes, multiple sclerosis (169), and in tumorigenesis (170, 171).

### **AUTHOR CONTRIBUTIONS**

AV wrote the review, CP performed the figures and revised the whole manuscript. LL wrote the review and supervised the figures and the whole text.

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- Signoret N, Rosenkilde MM, Klasse PJ, Schwartz TW, Malim MH, Hoxie JA, et al. Differential regulation of CXCR4 and CCR5 endocytosis. *J Cell Sci* (1998) 111(Pt 18):2819–30.
- Endres MJ, Clapham PR, Marsh M, Ahuja M, Turner JD, McKnight A, et al. CD4-independent infection by HIV-2 is mediated by fusin/CXCR4. *Cell* (1996) 87:745–56. doi:10.1016/S0092-8674(00)81393-8
- 11. Berger EA. HIV entry and tropism: the chemokine receptor connection. *AIDS* (1997) 11 Suppl A:S3–16.
- Moore JP, Trkola A, Dragic T. Co-receptors for HIV-1 entry. Curr Opin Immunol (1997) 9:551–62. doi:10.1016/S0952-7915(97)80110-0
- Mueller A, Kelly E, Strange PG. Pathways for internalization and recycling of the chemokine receptor CCR5. *Blood* (2002) 99:785–91. doi:10.1182/blood.V99. 3.785
- Littman DR. Chemokine receptors: keys to AIDS pathogenesis? Cell (1998) 93:677–80. doi:10.1016/S0092-8674(00)81429-4
- Lopalco L. CCR5: from natural resistance to a new anti-HIV strategy. Viruses (2010) 2:574–600. doi:10.3390/v2020574
- Tan Y, Tong P, Wang J, Zhao L, Li J, Yu Y, et al. The membrane-proximal region of C–C chemokine receptor type 5 participates in the infection of HIV-1. *Front Immunol* (2017) 8:478. doi:10.3389/fimmu.2017.00478
- Connor RI, Sheridan KE, Ceradini D, Choe S, Landau NR. Change in coreceptor use correlates with disease progression in HIV-1-infected individuals. *J Exp Med* (1997) 185:621–8. doi:10.1084/jem.185.4.621
- Signoret N, Hewlett L, Wavre S, Pelchen-Matthews A, Oppermann M, Marsh M. Agonist-induced endocytosis of CC chemokine receptor 5 is clathrin dependent. *Mol Biol Cell* (2005) 16:902–17. doi:10.1091/mbc. E04-08-0687

- Cocchi F, DeVico AL, Garzino-Demo A, Arya SK, Gallo RC, Lusso P. Identification of RANTES, MIP-1 alpha, and MIP-1 beta as the major HIVsuppressive factors produced by CD8+ T cells. *Science* (1995) 270:1811–5. doi:10.1126/science.270.5243.1811
- Bleul CC, Farzan M, Choe H, Parolin C, Clark-Lewis I, Sodroski J, et al. The lymphocyte chemoattractant SDF-1 is a ligand for LESTR/fusin and blocks HIV-1 entry. *Nature* (1996) 382:829–33. doi:10.1038/382829a0
- 21. Venuti A, Lopalco L. Tackling HIV: genetic vs. immune CCR5 targeting. *J AIDS Clin Res* (2014) 5:344–52. doi:10.4172/2155-6113.1000344
- Clerici M, Butto S, Lukwiya M, Saresella M, Declich S, Trabattoni D, et al. Immune activation in Africa is environmentally-driven and is associated with upregulation of CCR5. Italian-Ugandan AIDS Project. *AIDS* (2000) 14:2083–92. doi:10.1097/00002030-200009290-00003
- Kim MB, Giesler KE, Tahirovic YA, Truax VM, Liotta DC, Wilson LJ. CCR5 receptor antagonists in preclinical to phase II clinical development for treatment of HIV. *Expert Opin Investig Drugs* (2016) 25:1377–92. doi:10.108 0/13543784.2016.1254615
- 24. Huang Y, Paxton WA, Wolinsky SM, Neumann AU, Zhang L, He T, et al. The role of a mutant CCR5 allele in HIV-1 transmission and disease progression. *Nat Med* (1996) 2:1240–3. doi:10.1038/nm1196-1240
- Liu R, Paxton WA, Choe S, Ceradini D, Martin SR, Horuk R, et al. Homozygous defect in HIV-1 coreceptor accounts for resistance of some multiply-exposed individuals to HIV-1 infection. *Cell* (1996) 86:367–77. doi:10.1016/S0092-8674(00)80110-5
- Paxton WA, Liu R, Kang S, Wu L, Gingeras TR, Landau NR, et al. Reduced HIV-1 infectability of CD4+ lymphocytes from exposed-uninfected individuals: association with low expression of CCR5 and high production of beta-chemokines. *Virology* (1998) 244:66–73. doi:10.1006/viro.1998.9082
- Paxton WA, Martin SR, Tse D, O'Brien TR, Skurnick J, VanDevanter NL, et al. Relative resistance to HIV-1 infection of CD4 lymphocytes from persons who remain uninfected despite multiple high-risk sexual exposure. *Nat Med* (1996) 2:412–7. doi:10.1038/nm0496-412
- Samson M, Libert F, Doranz BJ, Rucker J, Liesnard C, Farber C-M, et al. Resistance to HIV-1 infection in Caucasian individuals bearing mutant alleles of the CCR-5 chemokine receptor gene. *Nature* (1996) 382:722–5. doi:10.1038/382722a0
- Pastori C, Weiser B, Barassi C, Uberti-Foppa C, Ghezzi S, Longhi R, et al. Long-lasting CCR5 internalization by antibodies in a subset of long-term nonprogressors: a possible protective effect against disease progression. *Blood* (2006) 107:4825–33. doi:10.1182/blood-2005-06-2463
- Rahimi H, Farajollahi MM, Hosseini A. Distribution of the mutated delta 32 allele of CCR5 co-receptor gene in Iranian population. *Med J Islam Repub Iran* (2014) 28:140–5.
- Karam W, Jurjus R, Khoury N, Khansa H, Assad C, Zalloua P, et al. Frequency of the CCR5-delta 32 chemokine receptor gene mutation in the Lebanese population. *East Mediterr Health J* (2004) 10:671–5.
- 32. Dean M, Carrington M, Winkler C, Huttley GA, Smith MW, Allikmets R, et al. Genetic restriction of HIV-1 infection and progression to AIDS by a deletion allele of the CKR5 structural gene. Hemophilia Growth and Development Study, Multicenter AIDS Cohort Study, Multicenter Hemophilia Cohort Study, San Francisco City Cohort, ALIVE Study. Science (1996) 273:1856–62.
- 33. Adojaan M, Mölder T, Männik A, Kivisild T, Villems R, Krispin T, et al. High prevalence of the CCR5Delta32 HIV-resistance mutation among Estonian HIV type 1-infected individuals. *AIDS Res Hum Retroviruses* (2007) 23:193–7. doi:10.1089/aid.2006.0113
- 34. Trecarichi EM, Tumbarello M, de Gaetano Donati K, Tamburrini E, Cauda R, Brahe C, et al. Partial protective effect of CCR5-Delta 32 heterozygosity in a cohort of heterosexual Italian HIV-1 exposed uninfected individuals. *AIDS Res Ther* (2006) 3:22. doi:10.1186/1742-6405-3-22
- 35. Stewart GJ, Ashton LJ, Biti RA, Ffrench RA, Bennetts BH, Newcombe NR, et al. Increased frequency of CCR-5 delta 32 heterozygotes among long-term non-progressors with HIV-1 infection. The Australian Long-Term Non-Progressor Study Group. *AIDS* (1997) 11:1833–8. doi:10.1097/00002030-199715000-00007
- Hogan CM, Hammer SM. Host determinants in HIV infection and disease. Part 2: genetic factors and implications for antiretroviral therapeutics. *Ann Intern Med* (2001) 134:978–96. doi:10.7326/ 0003-4819-134-9\_Part\_1-200105010-00013
- 37. Ghorban K, Dadmanesh M, Hassanshahi G, Momeni M, Zare-Bidaki M, Arababadi MK, et al. Is the CCR5  $\Delta$  32 mutation associated with immune

system-related diseases? Inflammation (2013) 36:633-42. doi:10.1007/s10753-012-9585-8

- Glass WG, McDermott DH, Lim JK, Lekhong S, Yu SF, Frank WA, et al. CCR5 deficiency increases risk of symptomatic West Nile virus infection. J Exp Med (2006) 203:35–40. doi:10.1084/jem.20051970
- Kindberg E, Mickiene A, Ax C, Akerlind B, Vene S, Lindquist L, et al. A deletion in the chemokine receptor 5 (CCR5) gene is associated with tickborne encephalitis. J Infect Dis (2008) 197:266–9. doi:10.1086/524709
- 40. Lopalco L. Natural anti-CCR5 antibodies in HIV-infection and -exposure. *J Transl Med* (2011) 9(Suppl 1):S4. doi:10.1186/1479-5876-9-S1-S4
- Bouhlal H, Hocini H, Quillent-Grégoire C, Donkova V, Rose S, Amara A, et al. Antibodies to C-C chemokine receptor 5 in normal human IgG block infection of macrophages and lymphocytes with primary R5-tropic strains of HIV-1. *J Immunol* (2001) 166:7606–11.
- 42. Ditzel HJ, Rosenkilde MM, Garred P, Wang M, Koefoed K, Pedersen C, et al. The CCR5 receptor acts as an alloantigen in CCR5Delta32 homozygous individuals: identification of chemokineand HIV-1-blocking human antibodies. *Proc Natl Acad Sci U S A* (1998) 95:5241–5. doi:10.1073/pnas.95.9.5241
- 43. Lopalco L, Barassi C, Pastori C, Longhi R, Burastero SE, Tambussi G, et al. CCR5-reactive antibodies in seronegative partners of HIV-seropositive individuals down-modulate surface CCR5 in vivo and neutralize the infectivity of R5 strains of HIV-1 In vitro. *J Immunol* (2000) 164:3426–33.
- Xu X, Ng SM, Hassouna E, Warrington A, Oh S-H, Rodriguez M. Humanderived natural antibodies: biomarkers and potential therapeutics. *Future Neurol* (2015) 10:25–39. doi:10.2217/fnl.14.62
- Panda S, Ding JL. Natural antibodies bridge innate and adaptive immunity. J Immunol (2015) 194:13–20. doi:10.4049/jimmunol.1400844
- Banchereau J, Steinman RM. Dendritic cells and the control of immunity. Nature (1998) 392:245–52. doi:10.1038/32588
- Zitvogel L. Dendritic and natural killer cells cooperate in the control/switch of innate immunity. J Exp Med (2002) 195:F9–14. doi:10.1084/jem.20012040
- Muramatsu M, Kinoshita K, Fagarasan S, Yamada S, Shinkai Y, Honjo T. Class switch recombination and hypermutation require activation-induced cytidine deaminase (AID), a potential RNA editing enzyme. *Cell* (2000) 102:553–63. doi:10.1016/S0092-8674(00)00078-7
- Coutinho A, Kazatchkine MD, Avrameas S. Natural autoantibodies. Curr Opin Immunol (1995) 7:812–8. doi:10.1016/0952-7915(95)80053-0
- Conrad K, Bachmann MP, Matsuura E, Shoenfeld Y. From animal models to human genetics: research on the induction and pathogenicity of autoantibodies. *Autoimmun Rev* (2005) 4:178–87. doi:10.1016/j.autrev.2004.10.001
- Harindranath N, Ikematsu H, Notkins AL, Casali P. Structure of the VH and VL segments of polyreactive and monoreactive human natural antibodies to HIV-1 and *Escherichia coli* beta-galactosidase. *Int Immunol* (1993) 5:1523–33. doi:10.1093/intimm/5.12.1523
- 52. Quan CP, Berneman A, Pires R, Avrameas S, Bouvet JP. Natural polyreactive secretory immunoglobulin A autoantibodies as a possible barrier to infection in humans. *Infect Immun* (1997) 65:3997–4004.
- Duan B, Morel L. Role of B-1a cells in autoimmunity. *Autoimmun Rev* (2006) 5:403–8. doi:10.1016/j.autrev.2005.10.007
- Elkon K, Casali P. Nature and functions of autoantibodies. Nat Clin Pract Rheumatol (2008) 4:491–8. doi:10.1038/ncprheum0895
- Sutterwala FS, Noel GJ, Salgame P, Mosser DM. Reversal of proinflammatory responses by ligating the macrophage Fcgamma receptor type I. *J Exp Med* (1998) 188:217–22. doi:10.1084/jem.188.1.217
- Berland R, Wortis HH. Origins and functions of B-1 cells with notes on the role of CD5. *Annu Rev Immunol* (2002) 20:253–300. doi:10.1146/annurev. immunol.20.100301.064833
- Kaminski DA, Stavnezer J. Enhanced IgA class switching in marginal zone and B1 B cells relative to follicular/B2 B cells. *J Immunol* (2006) 177:6025–9.
- De Milito A, Nilsson A, Titanji K, Thorstensson R, Reizenstein E, Narita M, et al. Mechanisms of hypergammaglobulinemia and impaired antigen-specific humoral immunity in HIV-1 infection. *Blood* (2004) 103:2180–6. doi:10.1182/blood-2003-07-2375
- Friedli F, Rieben R, Wegmüller E, Moerenhout M, Nydegger UE. Normal levels of allo- but increased levels of potentially autoreactive antibodies against ABO histo-blood group antigens in AIDS patients. *Clin Immunol Immunopathol* (1996) 80:96–100. doi:10.1006/clin.1996.0099
- Hunziker L, Recher M, Macpherson AJ, Ciurea A, Freigang S, Hengartner H, et al. Hypergammaglobulinemia and autoantibody induction

mechanisms in viral infections. Nat Immunol (2003) 4:343–9. doi:10.1038/ ni911

- Haynes BF, Fleming J, St Clair EW, Katinger H, Stiegler G, Kunert R, et al. Cardiolipin polyspecific autoreactivity in two broadly neutralizing HIV-1 antibodies. *Science* (2005) 308:1906–8. doi:10.1126/science.1111781
- Haynes BF, Mascola JR. The quest for an antibody-based HIV vaccine. Immunol Rev (2017) 275:5–10. doi:10.1111/imr.12517
- Kelsoe G, Haynes BF. Host controls of HIV broadly neutralizing antibody development. *Immunol Rev* (2017) 275:79–88. doi:10.1111/imr.12508
- 64. Vcelar B, Stiegler G, Wolf HM, Muntean W, Leschnik B, Mehandru S, et al. Reassessment of autoreactivity of the broadly neutralizing HIV antibodies 4E10 and 2F5 and retrospective analysis of clinical safety data. *AIDS* (2007) 21:2161–70. doi:10.1097/QAD.0b013e328285da15
- Blanpain C, Vanderwinden J-M, Cihak J, Wittamer V, Le Poul E, Issafras H, et al. Multiple active states and oligomerization of CCR5 revealed by functional properties of monoclonal antibodies. *Mol Biol Cell* (2002) 13:723–37. doi:10.1091/mbc.01-03-0129
- 66. Lee B, Sharron M, Blanpain C, Doranz BJ, Vakili J, Setoh P, et al. Epitope mapping of CCR5 reveals multiple conformational states and distinct but overlapping structures involved in chemokine and coreceptor function. *J Biol Chem* (1999) 274:9617–26. doi:10.1074/jbc.274.14.9617
- Olson WC, Jacobson JM. CCR5 monoclonal antibodies for HIV-1 therapy. *Curr Opin HIV AIDS* (2009) 4:104–11. doi:10.1097/COH.0b013e3283224015
- Wells TN, Proudfoot AE, Power CA, Marsh M. Chemokine receptors the new frontier for AIDS research. *Chem Biol* (1996) 3:603–9. doi:10.1016/ S1074-5521(96)90126-X
- Grene E, Pinto LA, Kwak-Kim JY, Giorgi JV, Landay AL, Kessler HA, et al. Increased levels of anti-CCR5 antibodies in sera from individuals immunized with allogeneic lymphocytes. *AIDS* (2000) 14:2627–8. doi:10.1097/00002030-200011100-00035
- Venuti A, Pastori C, Siracusano G, Riva A, Sciortino MT, Lopalco L. ERK1based pathway as a new selective mechanism to modulate CCR5 with natural antibodies. *J Immunol* (2015) 195:3045–57. doi:10.4049/jimmunol.1500708
- Eslahpazir J, Jenabian M-A, Bouhlal H, Hocini H, Carbonneil C, Grésenguet G, et al. Infection of macrophages and dendritic cells with primary R5-tropic human immunodeficiency virus type 1 inhibited by natural polyreactive anti-CCR5 antibodies purified from cervicovaginal secretions. *Clin Vaccine Immunol* (2008) 15:872–84. doi:10.1128/CVI.00463-07
- Barassi C, Marenzi C, Pastori C, Longhi R, Lazzarin A, Lopalco L. A new prospective against HIV infection: induction of murin CCR5-downregulating antibodies. *New Microbiol* (2004) 27:85–94.
- Devito C, Broliden K, Kaul R, Svensson L, Johansen K, Kiama P, et al. Mucosal and plasma IgA from HIV-1-exposed uninfected individuals inhibit HIV-1 transcytosis across human epithelial cells. *J Immunol* (2000) 165:5170–6.
- Devito C, Hinkula J, Kaul R, Lopalco L, Bwayo JJ, Plummer F, et al. Mucosal and plasma IgA from HIV-exposed seronegative individuals neutralize a primary HIV-1 isolate. *AIDS* (2000) 14:1917–20. doi:10.1097/00002030-200009080-00006
- 75. Lopalco L, Barassi C, Paolucci C, Breda D, Brunelli D, Nguyen M, et al. Predictive value of anti-cell and anti-human immunodeficiency virus (HIV) humoral responses in HIV-1-exposed seronegative cohorts of European and Asian origin. J Gen Virol (2005) 86:339–48. doi:10.1099/vir.0.80585-0
- 76. Bomsel M, Pastori C, Tudor D, Alberti C, Garcia S, Ferrari D, et al. Natural mucosal antibodies reactive with first extracellular loop of CCR5 inhibit HIV-1 transport across human epithelial cells. *AIDS* (2007) 21:13–22. doi:10.1097/QAD.0b013e328011049b
- 77. Venuti A, Pastori C, Pennisi R, Riva A, Sciortino MT, Lopalco L. Class B  $\beta$ -arrestin2-dependent CCR5 signalosome retention with natural antibodies to CCR5. *Sci Rep* (2016) 6:39382. doi:10.1038/srep39382
- Olson WC, Rabut GE, Nagashima KA, Tran DN, Anselma DJ, Monard SP, et al. Differential inhibition of human immunodeficiency virus type 1 fusion, gp120 binding, and CC-chemokine activity by monoclonal antibodies to CCR5. J Virol (1999) 73:4145–55.
- Barassi C, Lazzarin A, Lopalco L. CCR5-specific mucosal IgA in saliva and genital fluids of HIV-exposed seronegative subjects. *Blood* (2004) 104:2205–6. doi:10.1182/blood-2004-06-2134
- Grene E, Pinto LA, Landay AL, Kessler HA, Anderson SA, Dolan MJ, et al. Anti-CCR5 antibodies in sera of HIV-positive individuals. *Hum Immunol* (2001) 62:143–5. doi:10.1016/S0198-8859(00)00243-3

- Brombin C, Diomede L, Tudor D, Drillet AS, Pastori C, Poli E, et al. A nonparametric procedure for defining a new humoral immunologic profile in a pilot study on HIV infected patients. *PLoS One* (2013) 8:e58768. doi:10.1371/ journal.pone.0058768
- Pastori C, Diomede L, Venuti A, Fisher G, Jarvik J, Bomsel M, et al. Induction of HIV-blocking anti-CCR5 IgA in Peyers's patches without histopathological alterations. *J Virol* (2014) 88:3623–35. doi:10.1128/ JVI.03663-13
- Van Rompay KKA, Hunter Z, Jayashankar K, Peabody J, Montefiori D, LaBranche CC, et al. A vaccine against CCR5 protects a subset of macaques upon intravaginal challenge with simian immunodeficiency virus SIVmac251. *J Virol* (2014) 88:2011–24. doi:10.1128/JVI.02447-13
- Chackerian B, Briglio L, Albert PS, Lowy DR, Schiller JT. Induction of autoantibodies to CCR5 in macaques and subsequent effects upon challenge with an R5-tropic simian/human immunodeficiency virus. *J Virol* (2004) 78:4037–47. doi:10.1128/JVI.78.8.4037-4047.2004
- Wu L, LaRosa G, Kassam N, Gordon CJ, Heath H, Ruffing N, et al. Interaction of chemokine receptor CCR5 with its ligands: multiple domains for HIV-1 gp120 binding and a single domain for chemokine binding. *J Exp Med* (1997) 186:1373–81. doi:10.1084/jem.186.8.1373
- Trkola A, Ketas TJ, Nagashima KA, Zhao L, Cilliers T, Morris L, et al. Potent, broad-spectrum inhibition of human immunodeficiency virus type 1 by the CCR5 monoclonal antibody PRO 140. J Virol (2001) 75:579–88. doi:10.1128/ JVI.75.2.579-588.2001
- Flego M, Ascione A, Cianfriglia M, Vella S. Clinical development of monoclonal antibody-based drugs in HIV and HCV diseases. *BMC Med* (2013) 11:4. doi:10.1186/1741-7015-11-4
- Oppermann M. Chemokine receptor CCR5: insights into structure, function, and regulation. *Cell Signal* (2004) 16:1201–10. doi:10.1016/j. cellsig.2004.04.007
- Hüttenrauch F, Pollok-Kopp B, Oppermann M. G protein-coupled receptor kinases promote phosphorylation and beta-arrestin-mediated internalization of CCR5 homo- and hetero-oligomers. *J Biol Chem* (2005) 280:37503–15. doi:10.1074/jbc.M500535200
- Ali H, Richardson RM, Haribabu B, Snyderman R. Chemoattractant receptor cross-desensitization. *J Biol Chem* (1999) 274:6027–30. doi:10.1074/jbc.274.10.6027
- Pitcher JA, Freedman NJ, Lefkowitz RJ. G protein-coupled receptor kinases. Annu Rev Biochem (1998) 67:653–92. doi:10.1146/annurev. biochem.67.1.653
- Luttrell LM, Lefkowitz RJ. The role of beta-arrestins in the termination and transduction of G-protein-coupled receptor signals. J Cell Sci (2002) 115:455–65.
- Perry SJ, Lefkowitz RJ. Arresting developments in heptahelical receptor signaling and regulation. *Trends Cell Biol* (2002) 12:130–8. doi:10.1016/ S0962-8924(01)02239-5
- Okamoto Y, Ninomiya H, Miwa S, Masaki T. Cholesterol oxidation switches the internalization pathway of endothelin receptor type A from caveolae to clathrin-coated pits in Chinese hamster ovary cells. *J Biol Chem* (2000) 275:6439–46. doi:10.1074/jbc.275.9.6439
- Moore CAC, Milano SK, Benovic JL. Regulation of receptor trafficking by GRKs and arrestins. *Annu Rev Physiol* (2007) 69:451–82. doi:10.1146/ annurev.physiol.69.022405.154712
- Hanyaloglu AC, von Zastrow M. Regulation of GPCRs by endocytic membrane trafficking and its potential implications. *Annu Rev Pharmacol Toxicol* (2008) 48:537–68. doi:10.1146/annurev.pharmtox.48.113006.094830
- Marchese A, Paing MM, Temple BRS, Trejo J. G protein-coupled receptor sorting to endosomes and lysosomes. *Annu Rev Pharmacol Toxicol* (2008) 48:601–29. doi:10.1146/annurev.pharmtox.48.113006.094646
- Escola J-M, Kuenzi G, Gaertner H, Foti M, Hartley O. CC chemokine receptor 5 (CCR5) desensitization: cycling receptors accumulate in the trans-Golgi network. J Biol Chem (2010) 285:41772–80. doi:10.1074/jbc.M110.153460
- Marie N, Lecoq I, Jauzac P, Allouche S. Differential sorting of human delta-opioid receptors after internalization by peptide and alkaloid agonists. *J Biol Chem* (2003) 278:22795–804. doi:10.1074/jbc.M300084200
- 100. Han S, Xiao K, Kim J, Wu J-H, Wisler JW, Nakamura N, et al. MARCH2 promotes endocytosis and lysosomal sorting of carvedilol-bound  $\beta(2)$ -adrenergic receptors. *J Cell Biol* (2012) 199:817–30. doi:10.1083/jcb.201208192

- 101. Groer CE, Schmid CL, Jaeger AM, Bohn LM. Agonist-directed interactions with specific beta-arrestins determine mu-opioid receptor trafficking, ubiquitination, and dephosphorylation. *J Biol Chem* (2011) 286:31731–41. doi:10.1074/jbc.M111.248310
- Jean-Charles P-Y, Rajiv V, Shenoy SK. Ubiquitin-related roles of β-arrestins in endocytic trafficking and signal transduction. *J Cell Physiol* (2016) 231:2071–80. doi:10.1002/jcp.25317
- 103. Shenoy SK, Drake MT, Nelson CD, Houtz DA, Xiao K, Madabushi S, et al. beta-arrestin-dependent, G protein-independent ERK1/2 activation by the beta2 adrenergic receptor. J Biol Chem (2006) 281:1261–73. doi:10.1074/jbc. M506576200
- Lefkowitz RJ, Shenoy SK. Transduction of receptor signals by beta-arrestins. Science (2005) 308:512–7. doi:10.1126/science.1109237
- Reiter E, Lefkowitz RJ. GRKs and beta-arrestins: roles in receptor silencing, trafficking and signaling. *Trends Endocrinol Metab* (2006) 17:159–65. doi:10.1016/j.tem.2006.03.008
- 106. Shenoy SK. Deubiquitinases and their emerging roles in β-arrestin-mediated signaling. *Methods Enzymol* (2014) 535:351–70. doi:10.1016/ B978-0-12-397925-4.00020-1
- 107. Luttrell LM, Roudabush FL, Choy EW, Miller WE, Field ME, Pierce KL, et al. Activation and targeting of extracellular signal-regulated kinases by beta-arrestin scaffolds. *Proc Natl Acad Sci U S A* (2001) 98:2449–54. doi:10.1073/ pnas.041604898
- 108. Tohgo A, Choy EW, Gesty-Palmer D, Pierce KL, Laporte S, Oakley RH, et al. The stability of the G protein-coupled receptor-beta-arrestin interaction determines the mechanism and functional consequence of ERK activation. *J Biol Chem* (2003) 278:6258–67. doi:10.1074/jbc.M212231200
- 109. Fox JM, Kasprowicz R, Hartley O, Signoret N. CCR5 susceptibility to ligand-mediated down-modulation differs between human T lymphocytes and myeloid cells. *J Leukoc Biol* (2015) 98:59–71. doi:10.1189/ jlb.2A0414-193RR
- 110. Devito C, Zuber B, Schröder U, Benthin R, Okuda K, Broliden K, et al. Intranasal HIV-1-gp160-DNA/gp41 peptide prime-boost immunization regimen in mice results in long-term HIV-1 neutralizing humoral mucosal and systemic immunity. *J Immunol* (2004) 173:7078–89.
- 111. Barassi C, Soprana E, Pastori C, Longhi R, Buratti E, Lillo F, et al. Induction of murine mucosal CCR5-reactive antibodies as an anti-human immunodeficiency virus strategy. *J Virol* (2005) 79:6848–58. doi:10.1128/ JVI.79.11.6848-6858.2005
- 112. Pastori C, Clivio A, Diomede L, Consonni R, De Mori GMS, Longhi R, et al. Two amino acid substitutions within the first external loop of CCR5 induce human immunodeficiency virus-blocking antibodies in mice and chickens. *J Virol* (2008) 82:4125–34. doi:10.1128/JVI.02232-07
- 113. Li L, Tian JH, Yang K, Zhang P, Jia WQ. Humanized PA14 (a monoclonal CCR5 antibody) for treatment of people with HIV infection. *Cochrane Database Syst Rev* (2014) 7:CD008439. doi:10.1002/14651858.CD008439. pub3
- 114. Wu K, Xue X, Wang Z, Yan Z, Shi J, Han W, et al. Construction, purification, and immunogenicity of recombinant cystein-cystein type chemokine receptor 5 vaccine. *Protein Expr Purif* (2006) 49:108–13. doi:10.1016/j. pep.2006.02.020
- 115. Misumi S, Nakayama D, Kusaba M, Iiboshi T, Mukai R, Tachibana K, et al. Effects of immunization with CCR5-based cycloimmunogen on simian/ HIVSF162P3 challenge. *J Immunol* (2006) 176:463–71.
- Bogers WMJM, Bergmeier LA, Oostermeijer H, ten Haaft P, Wang Y, Kelly CG, et al. CCR5 targeted SIV vaccination strategy preventing or inhibiting SIV infection. *Vaccine* (2004) 22:2974–84. doi:10.1016/j.vaccine.2004.02.050
- Bogers WM, Bergmeier LA, Ma J, Oostermeijer H, Wang Y, Kelly CG, et al. A novel HIV-CCR5 receptor vaccine strategy in the control of mucosal SIV/HIV infection. AIDS (2004) 18:25–36. doi:10.1097/00002030-200401020-00003
- Peabody DS, Manifold-Wheeler B, Medford A, Jordan SK, do Carmo Caldeira J, Chackerian B. Immunogenic display of diverse peptides on viruslike particles of RNA phage MS2. *J Mol Biol* (2008) 380:252–63. doi:10.1016/j. jmb.2008.04.049
- Chain B, Arnold J, Akthar S, Brandt M, Davis D, Noursadeghi M, et al. A linear epitope in the N-terminal domain of CCR5 and its interaction with antibody. *PLoS One* (2015) 10:e0128381. doi:10.1371/journal.pone.0128381

- 120. Ji C, Brandt M, Dioszegi M, Jekle A, Schwoerer S, Challand S, et al. Novel CCR5 monoclonal antibodies with potent and broad-spectrum anti-HIV activities. *Antiviral Res* (2007) 74:125–37. doi:10.1016/j.antiviral.2006.11.003
- Lewis GK, Pazgier M, DeVico AL. Survivors remorse: antibody-mediated protection against HIV-1. *Immunol Rev* (2017) 275:271–84. doi:10.1111/ imr.12510
- 122. Allers K, Hütter G, Hofmann J, Loddenkemper C, Rieger K, Thiel E, et al. Evidence for the cure of HIV infection by CCR5Δ32/Δ32 stem cell transplantation. *Blood* (2011) 117:2791–9. doi:10.1182/blood-2010-09-309591
- Burlone ME, Budkowska A. Hepatitis C virus cell entry: role of lipoproteins and cellular receptors. *J Gen Virol* (2009) 90:1055–70. doi:10.1099/ vir.0.008300-0
- 124. Song R, Franco D, Kao C-Y, Yu F, Huang Y, Ho DD. Epitope mapping of ibalizumab, a humanized anti-CD4 monoclonal antibody with anti-HIV-1 activity in infected patients. *J Virol* (2010) 84:6935–42. doi:10.1128/JVI. 00453-10
- 125. Jacobson JM, Lalezari JP, Thompson MA, Fichtenbaum CJ, Saag MS, Zingman BS, et al. Phase 2a study of the CCR5 monoclonal antibody PRO 140 administered intravenously to HIV-infected adults. *Antimicrob Agents Chemother* (2010) 54:4137–42. doi:10.1128/AAC.00086-10
- 126. Steinberger P, Andris-Widhopf J, Bühler B, Torbett BE, Barbas CF. Functional deletion of the CCR5 receptor by intracellular immunization produces cells that are refractory to CCR5-dependent HIV-1 infection and cell fusion. *Proc Natl Acad Sci U S A* (2000) 97:805–10. doi:10.1073/pnas.97.2.805
- 127. Aires da Silva F, Costa MJL, Corte-Real S, Goncalves J. Cell type-specific targeting with sindbis pseudotyped lentiviral vectors displaying anti-CCR5 single-chain antibodies. *Hum Gene Ther* (2005) 16:223–34. doi:10.1089/ hum.2005.16.223
- Margolis DM, Koup RA, Ferrari G. HIV antibodies for treatment of HIV infection. *Immunol Rev* (2017) 275:313–23. doi:10.1111/imr.12506
- 129. Gardner MR, Kattenhorn LM, Kondur HR, von Schaewen M, Dorfman T, Chiang JJ, et al. AAV-expressed eCD4-Ig provides durable protection from multiple SHIV challenges. *Nature* (2015) 519:87–91. doi:10.1038/nature14264
- 130. Xie J, Sok D, Wu NC, Zheng T, Zhang W, Burton DR, et al. Immunochemical engineering of cell surfaces to generate virus resistance. *Proc Natl Acad Sci U* S A (2017) 114:4655–60. doi:10.1073/pnas.1702764114
- 131. Pett SL, Amin J, Horban A, Andrade-Villanueva J, Losso M, Porteiro N, et al. Week 96 results of the randomized, multicentre Maraviroc Switch (MARCH) study. *HIV Med* (2017). doi:10.1111/hiv.12532
- 132. Van Der Ryst E. Maraviroc a CCR5 antagonist for the treatment of HIV-1 infection. *Front Immunol* (2015) 6:277. doi:10.3389/fimmu.2015.00277
- Harada S, Yoshimura K. Driving HIV-1 into a vulnerable corner by taking advantage of viral adaptation and evolution. *Front Microbiol* (2017) 8:390. doi:10.3389/fmicb.2017.00390
- Hartley O, Offord RE. Engineering chemokines to develop optimized HIV inhibitors. *Curr Protein Pept Sci* (2005) 6:207–19. doi:10.2174/ 1389203054065400
- Cerini F, Offord R, McGowan I, Hartley O. Stability of 5P12-RANTES, a candidate rectal microbicide, in human rectal lavage. *AIDS Res Hum Retroviruses* (2017) 33:768–77. doi:10.1089/AID.2016.0199
- Corbeau P, Reynes J. CCR5 antagonism in HIV infection: ways, effects, and side effects. AIDS (2009) 23:1931–43. doi:10.1097/QAD.0b013e32832e71cd
- 137. Hartley O, Gaertner H, Wilken J, Thompson D, Fish R, Ramos A, et al. Medicinal chemistry applied to a synthetic protein: development of highly potent HIV entry inhibitors. *Proc Natl Acad Sci U S A* (2004) 101:16460–5. doi:10.1073/pnas.0404802101
- Gaertner H, Cerini F, Escola J-M, Kuenzi G, Melotti A, Offord R, et al. Highly potent, fully recombinant anti-HIV chemokines: reengineering a low-cost microbicide. *Proc Natl Acad Sci U S A* (2008) 105:17706–11. doi:10.1073/ pnas.0805098105
- Lederman MM, Veazey RS, Offord R, Mosier DE, Dufour J, Mefford M, et al. Prevention of vaginal SHIV transmission in rhesus macaques through inhibition of CCR5. *Science* (2004) 306:485–7. doi:10.1126/science.1099288
- 140. Kuhmann SE, Hartley O. Targeting chemokine receptors in HIV: a status report. Annu Rev Pharmacol Toxicol (2008) 48:425–61. doi:10.1146/annurev. pharmtox.48.113006.094847

- 141. Simmons G, Clapham PR, Picard L, Offord RE, Rosenkilde MM, Schwartz TW, et al. Potent inhibition of HIV-1 infectivity in macrophages and lymphocytes by a novel CCR5 antagonist. *Science* (1997) 276:276–9. doi:10.1126/ science.276.5310.276
- 142. Mack M, Luckow B, Nelson PJ, Cihak J, Simmons G, Clapham PR, et al. Aminooxypentane-RANTES induces CCR5 internalization but inhibits recycling: a novel inhibitory mechanism of HIV infectivity. J Exp Med (1998) 187:1215–24. doi:10.1084/jem.187.8.1215
- 143. Dogo-Isonagie C, Lam S, Gustchina E, Acharya P, Yang Y, Shahzad-ul-Hussan S, et al. Peptides from second extracellular loop of C-C chemokine receptor type 5 (CCR5) inhibit diverse strains of HIV-1. *J Biol Chem* (2012) 287:15076–86. doi:10.1074/jbc.M111.332361
- 144. Bobyk KD, Mandadapu SR, Lohith K, Guzzo C, Bhargava A, Lusso P, et al. Design of HIV coreceptor derived peptides that inhibit viral entry at submicromolar concentrations. *Mol Pharm* (2017) 14:2681–9. doi:10.1021/acs. molpharmaceut.7b00155
- 145. Hütter G, Nowak D, Mossner M, Ganepola S, Müssig A, Allers K, et al. Longterm control of HIV by CCR5 Delta32/Delta32 stem-cell transplantation. N Engl J Med (2009) 360:692–8. doi:10.1056/NEJMoa0802905
- 146. Didigu CA, Wilen CB, Wang J, Duong J, Secreto AJ, Danet-Desnoyers GA, et al. Simultaneous zinc-finger nuclease editing of the HIV coreceptors ccr5 and cxcr4 protects CD4+ T cells from HIV-1 infection. *Blood* (2014) 123:61–9. doi:10.1182/blood-2013-08-521229
- 147. Mock U, Machowicz R, Hauber I, Horn S, Abramowski P, Berdien B, et al. mRNA transfection of a novel TAL effector nuclease (TALEN) facilitates efficient knockout of HIV co-receptor CCR5. *Nucleic Acids Res* (2015) 43:5560–71. doi:10.1093/nar/gkv469
- 148. Romano Ibarra GS, Paul B, Sather BD, Younan PM, Sommer K, Kowalski JP, et al. Efficient modification of the CCR5 locus in primary human T cells with megaTAL nuclease establishes HIV-1 resistance. *Mol Ther Nucleic Acids* (2016) 5:e352. doi:10.1038/mtna.2016.56
- 149. Tebas P, Stein D, Tang WW, Frank I, Wang SQ, Lee G, et al. Gene editing of CCR5 in autologous CD4 T cells of persons infected with HIV. N Engl J Med (2014) 370:901–10. doi:10.1056/NEJMoa1300662
- 150. Ye L, Wang J, Beyer AI, Teque F, Cradick TJ, Qi Z, et al. Seamless modification of wild-type induced pluripotent stem cells to the natural CCR5∆32 mutation confers resistance to HIV infection. *Proc Natl Acad Sci U S A* (2014) 111:9591–6. doi:10.1073/pnas.1407473111
- Bhoj VG, Thibodeaux SR, Levine BL. Novel gene and cellular therapy approaches for treating HIV. *Discov Med* (2016) 21:283–92.
- Hale M, Mesojednik T, Romano Ibarra GS, Sahni J, Bernard A, Sommer K, et al. Engineering HIV-resistant, anti-HIV chimeric antigen receptor T cells. *Mol Ther* (2017) 25:570–9. doi:10.1016/j.ymthe.2016.12.023
- Manjunath N, Yi G, Dang Y, Shankar P. Newer gene editing technologies toward HIV gene therapy. *Viruses* (2013) 5:2748–66. doi:10.3390/v5112748
- Cho SW, Kim S, Kim JM, Kim J-S. Targeted genome engineering in human cells with the Cas9 RNA-guided endonuclease. *Nat Biotechnol* (2013) 31:230–2. doi:10.1038/nbt.2507
- 155. Cradick TJ, Fine EJ, Antico CJ, Bao G. CRISPR/Cas9 systems targeting β-globin and CCR5 genes have substantial off-target activity. *Nucleic Acids Res* (2013) 41:9584–92. doi:10.1093/nar/gkt714
- 156. Mussolino C, Alzubi J, Fine EJ, Morbitzer R, Cradick TJ, Lahaye T, et al. TALENs facilitate targeted genome editing in human cells with high specificity and low cytotoxicity. *Nucleic Acids Res* (2014) 42:6762–73. doi:10.1093/ nar/gku305
- 157. Shimizu S, Kamata M, Kittipongdaja P, Chen KN, Kim S, Pang S, et al. Characterization of a potent non-cytotoxic shRNA directed to the HIV-1

co-receptor CCR5. Genet Vaccines Ther (2009) 7:8. doi:10.1186/1479-0556-7-8

- 158. Cordelier P, Morse B, Strayer DS. Targeting CCR5 with siRNAs: using recombinant SV40-derived vectors to protect macrophages and microglia from R5-tropic HIV. *Oligonucleotides* (2003) 13:281–94. doi:10.1089/154545703322616961
- 159. Bai J, Gorantla S, Banda N, Cagnon L, Rossi J, Akkina R. Characterization of anti-CCR5 ribozyme-transduced CD34+ hematopoietic progenitor cells in vitro and in a SCID-hu mouse model in vivo. *Mol Ther* (2000) 1:244–54. doi:10.1006/mthe.2000.0038
- 160. An DS, Qin FX-F, Auyeung VC, Mao SH, Kung SKP, Baltimore D, et al. Optimization and functional effects of stable short hairpin RNA expression in primary human lymphocytes via lentiviral vectors. *Mol Ther* (2006) 14:494–504. doi:10.1016/j.ymthe.2006.05.015
- 161. Whitehead KA, Dahlman JE, Langer RS, Anderson DG. Silencing or stimulation? siRNA delivery and the immune system. *Annu Rev Chem Biomol Eng* (2011) 2:77–96. doi:10.1146/annurev-chembioeng-061010-114133
- Stewart G. Chemokine genes beating the odds. Nat Med (1998) 4:275–7. doi:10.1038/nm0398-275
- Cohen J. Exploiting the HIV-chemokine nexus. *Science* (1997) 275:1261–4. doi:10.1126/science.275.5304.1261
- 164. Keller MA, Stiehm ER. Passive immunity in prevention and treatment of infectious diseases. *Clin Microbiol Rev* (2000) 13:602–14. doi:10.1128/ CMR.13.4.602-614.2000
- Saylor C, Dadachova E, Casadevall A. Monoclonal antibody-based therapies for microbial diseases. *Vaccine* (2009) 27(Suppl 6):G38–46. doi:10.1016/j. vaccine.2009.09.105
- 166. Asiimwe S, Ross JM, Arinaitwe A, Tumusiime O, Turyamureeba B, Roberts DA, et al. Expanding HIV testing and linkage to care in southwestern Uganda with community health extension workers. *J Int AIDS Soc* (2017) 20:2163. doi:10.7448/ias.20.5.21633
- 167. Hickey MD, Odeny TA, Petersen M, Neilands TB, Padian N, Ford N, et al. Specification of implementation interventions to address the cascade of HIV care and treatment in resource-limited settings: a systematic review. *Implement Sci* (2017) 12:102. doi:10.1186/s13012-017-0630-8
- Lederman MM, Penn-Nicholson A, Cho M, Mosier D. Biology of CCR5 and its role in HIV infection and treatment. *JAMA* (2006) 296:815–26. doi:10.1001/jama.296.7.815
- Ajuebor MN, Carey JA, Swain MG. CCR5 in T cell-mediated liver diseases: what's going on? J Immunol (2006) 177:2039–45.
- Velasco-Velázquez M, Xolalpa W, Pestell RG. The potential to target CCL5/ CCR5 in breast cancer. *Expert Opin Ther Targets* (2014) 18:1265–75. doi:10. 1517/14728222.2014.949238
- Chang W-J, Du Y, Zhao X, Ma L-Y, Cao G-W. Inflammation-related factors predicting prognosis of gastric cancer. World J Gastroenterol (2014) 20:4586–96. doi:10.3748/wjg.v20.i16.4586

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