



# Role of Tissue-Resident Memory in Intra-Tumor Heterogeneity and Response to Immune Checkpoint Blockade

#### Kavita M. Dhodapkar\*

Aflac Cancer Center of Children's Healthcare of Atlanta, Department of Pediatrics, Emory University, Atlanta, GA, United States

Tissue-resident memory T ( $T_{RM}$ ) cells are a distinct subset of memory T cells that reside in non-lymphoid tissues for prolonged periods of time without significant recirculation providing continued immune surveillance at these sites. Recent studies suggest that  $T_{RM}$ cells are also enriched within tumor tissue. Expression of inhibitory immune checkpoints (ICPs) is particularly enriched on this subset of tumor-infiltrating T cells, suggesting that they are major targets for newer therapies targeting ICPs such as the programmed death-1 pathway. Recent studies suggest that tissue restriction of these cells without recirculation may also lead to heterogeneity of  $T_{RM}$  cells within individual metastatic lesions, ultimately leading to inter-lesional diversity. Thus, individual metastatic lesions may contain genomically distinct immune microenvironments that impact both evolution of tumors as well as the mechanisms underlying response and resistance to immune therapies. Understanding the biology of  $T_{RM}$  cells infiltrating tumors will be essential to improving immune-based approaches in diverse settings.

#### **OPEN ACCESS**

#### Edited by:

Fathia Mami-Chouaib, Institut National de la Santé et de la Recherche Médicale (INSERM), France

#### Reviewed by:

Kimberly Sue Schluns, University of Texas MD Anderson Cancer Center, United States Kim Klonowski, University of Georgia, United States

\*Correspondence: Kavita M. Dhodapkar kavita.dhodapkar@emory.edu

#### Specialty section:

This article was submitted to Immunological Memory, a section of the journal Frontiers in Immunology

> Received: 19 April 2018 Accepted: 04 July 2018 Published: 16 July 2018

#### Citation:

Dhodapkar KM (2018) Role of Tissue-Resident Memory in Intra-Tumor Heterogeneity and Response to Immune Checkpoint Blockade. Front. Immunol. 9:1655. doi: 10.3389/fimmu.2018.01655 Keywords: tissue-resident memory cells, immune checkpoint blockade, tumor heterogeneity, cancer immunotherapy, immunity to cancer

Immune-based approaches, particularly those based on the blockade of inhibitory immune checkpoints (ICPs) on T cells have emerged as among the most promising new strategies to treat cancer (1). An important aspect of immune therapies is their potential ability to mediate long-term control of tumors. The capacity of the immune system to mediate long-term protection, particularly against pathogens, such as in the context of vaccines, is mediated in large part by immunologic memory (2). Therefore, understanding immunologic memory mediated by T cells is likely to be important for deeper understanding of immune-mediated long-term control of tumors. It is thought that uptake of antigens from dying tumor cells by antigen-presenting cells leads to activation of anti-tumor T cells in the lymph nodes, and resultant effector memory T cells traffic back to the tumor to mediate anti-tumor effects, creating a tumor-immunity cycle (3). Several studies have shown that infiltration of primary and metastatic lesions by immune cells, particularly T cells and myeloid cells impacts outcome (4). Studies in both mice and humans suggest that there are differences in the memory T cell subsets that provide immune surveillance within lymphoid and non-lymphoid tissues (NLTs). As tumor-related mortality in most solid tumors is not due to growth of primary tumors, but rather due to the growth of metastatic tumor cells in NLTs, it is the immune surveillance in NLTs that may be critical for protective tumor immunity. In this review, we discuss newer insights into spatial aspect of immunologic memory and particularly memory T cells within NLTs in the context of tumor immunity. We will discuss emerging evidence suggesting that the biology of these tissue-resident memory  $(T_{RM})$  T cells may not only be critical for understanding and improving clinical responses to ICP

blockade, but may also contribute to the complexity of immune microenvironment by creating inter-lesional heterogeneity in the setting of metastatic cancer.

# T<sub>RM</sub> T CELLS: LOCAL POLICEMEN

Initial models of T cell memory classified effector/central memory  $(T_{EM}/T_{CM})$  T cells, with the effector subset implicated in surveying NLTs (5). Recent studies have identified a third subset, termed  $T_{\mbox{\tiny RM}}\,T$  cells that reside for prolonged periods in NLTs and play an important role in protective immunity (6). Mouse  $T_{RM}$  cells have been described in diverse tissues, including lung, liver, brain, as well as barrier tissues (6, 7). Murine  $T_{RM}$  cells haven been shown to mediate rapid in situ protection against viral, bacterial, and parasitic infections and are more effective in this regard than their circulating counterparts, including central memory T cells (7, 8). An important aspect of T<sub>RM</sub>-mediated immune surveillance is its regional nature. Thus in parabiotic mice that share systemic circulation, T<sub>RM</sub> cells remain localized within tissues and do not cross over to equilibrate in the paired mouse carrying antigenic stimulus (6). T<sub>RM</sub> cells express CD69, which is implicated in tissue retention by sequestration of the sphingosine-1-phosphate receptor (9).

Tissue-resident memory cells have also been identified in several human tissues and implicated in tissue-restricted pathology particularly in the skin, such as fixed drug eruptions (10-12). As in the mouse, human T<sub>RM</sub> cells have been identified by the expression of CD69 on memory T cells within tissues, which is generally lacking in blood memory T cells (13). In humans, CD103 is expressed only in a subset of CD69+CD8+ memory T cells in some barrier tissues, but not by CD4+ memory T cells in any tissue, indicating that CD69 may be a more universal marker distinguishing both CD4+ and CD8+ memory T cells in tissues from their blood counterparts. It is notable that the proportion of T<sub>RM</sub> cells differs in different tissues, with enrichment in some barrier tissues such as skin. Recent studies have also characterized transcriptional profiles of human T<sub>RM</sub> cells, which resemble their murine counterparts and also illustrate that these are a distinct subset of human memory T cells (14, 15).

The pathways that regulate generation, recruitment, retention, and long-term maintenance of these T cells in NLTs remain an active area of research. New insights into transcriptional regulation of the T<sub>RM</sub> differentiation are emerging and may differ between humans and mice. For example, the transcription factor Hobit/ ZNF683 is exclusively expressed and required for the generation of murine T<sub>RM</sub> cells after infection, but expressed at low/negligible levels on human  $T_{RM}$  cells (14, 16). In recent studies, we have shown that human and murine T<sub>RM</sub> cells express NR4A1/ nur77, which is also essential for T<sub>RM</sub> differentiation in several murine tissues (17). Runx3 is another transcription factor that promotes the differentiation of T cells with  $T_{RM}$  phenotype (18). Retention and maintenance of T<sub>RM</sub> cells may also depend on the availability of local antigen, interactions with myeloid cells as well as cytokines like TGF $\beta$  and IL-15 in NLTs (19, 20). Tissue distribution of T<sub>RM</sub> cells, at least against pathogens may depend on the site of initial exposure. For example, human influenzaspecific T<sub>RM</sub> cells are preferentially found in the lung (21) and hepatitis-B specific  $T_{RM}$  cells particularly in the liver (22). Human bone marrow may also be a particularly interesting compartment for long-lived memory T cells with phenotype of  $T_{RM}$  cells (17, 23, 24).

### T<sub>RM</sub> CELLS IN TUMORS

Several studies have now documented that a large proportion of T cells infiltrating human tumors have T<sub>RM</sub> phenotype, at least based on the expression of CD69 and CD103 (11, 12, 25-27). In some studies, these T cells were also shown to have genomic signatures consistent with those described for  $T_{RM}$  cells (11, 25, 26). This includes altered expression of genes involved in tissue retention/homing (such as downregulation of S1PR1, S1PR5, and KLF2; increase in CD69 and CD103) as well as transcription factors now functionally implicated in this phenotype (such as NR4A1, NR4A2, and Runx3) in several tissues. It is notable that some of the genes (such as Hobit) critically implicated in the biology of murine T<sub>RM</sub> cells are not expressed at high levels in their human counterparts. It is notable that in mouse models of viral infections such as lymphocytic choriomeningitis virus (LCMV), T cell memory has been largely studied when the underlying viral antigen is depleted. However, the biology in human tumors or other states of persistent viral infection may differ from LCMV models and local antigen may have important implications for T<sub>RM</sub> biology. Indeed, recent studies suggest that local antigen may drive proliferation of T<sub>RM</sub> cells *in situ* (28, 29).

While the infiltration of tumors by T cells has in general emerged as a strong indicator of improved prognosis, the presence of T<sub>RM</sub> cells within tumor-infiltrating lymphocytes (TILs) may be a particular driver of this correlation. The proportion of TILs that have T<sub>RM</sub> phenotype differs between studies (for example, from 25 to 75%) and may depend in part on the nature of specific markers utilized to identify these cells as well as the specific tissue/ organ studied. This subset of cells may also be enriched for tumor reactivity, which is also consistent with other studies showing enrichment of tumor reactivity such as against tumor-associated neoantigens in CD8+ memory T cells with PD1+ phenotype (26, 30). Recent studies in murine models also suggest that these cells are important contributors to protective tumor immunity (31). In this study, the presence of  $T_{\text{RM}}$  cells was modeled in the setting of autoimmune vitiligo and melanoma-specific T<sub>RM</sub> cells infiltrating these lesions were shown to mediate strong tumor protection. To date, most of the data relating to the biology of  $T_{RM}$ cells in human tumor tissues are largely based on patients with solid tumors. Further studies are needed to better characterize this subset of T cells within hematologic malignancies. Below, we particularly focus on two aspects of the biology of tumor-associated T<sub>RM</sub> cells, their contribution to clinical responses to ICP blockade therapies and emergence of inter-lesional heterogeneity.

#### ARE T<sub>RM</sub> CELLS A CRITICAL TARGET FOR ICP BLOCKADE?

Antibody-mediated blockade of inhibitory ICPs such as programmed death-1 (PD-1) have led to impressive and durable clinical regressions in several cancers (32). This is remarkable as the expression of ICPs such as PD-1 is limited to only a subset of TILs (33). The principle of ICP blockade is based on the concept of unleashing the activity of pre-existing anti-tumor T cells against the tumor (34). Studies of T cell receptor (TCR) sequencing of T cells from patients receiving anti-PD1 therapy suggests that this therapy leads to in situ proliferation of CD8+ T cells within tumors of patients who respond to therapy (35). The ICP expressing T cells were found to include most of the tumor reactive T cells. While such tumor-reactive T cells can be detected in peripheral blood, these cells are predominantly present within the tumor tissue. In recent studies, we and others have shown that T<sub>RM</sub> cells are the dominant T cell subset expressing ICPs within the tumor microenvironment (11, 25). While most studies have described the presence of T<sub>RM</sub> cells within adult tumors recent data suggest that T<sub>RM</sub> cells are also enriched within pediatric tumors like glioma and are the T cell subset within these tumors that predominantly expresses ICPs (36). While  $T_{RM}$  cells were initially identified in the tumor tissue based on the expression of classic T<sub>RM</sub> markers such as CD69 or CD103, gene expression studies confirmed that these T cells are a distinct subset of TILs with a genomic signature overlapping with  $T_{RM}$  signature. Importantly, although CD69 is well studied as a T cell activation marker, the genomic profiles of CD69+  $T_{RM}$  cells are distinct from activated T cells and instead enriched for tissue retention genes (25). Therefore, while tumor tissue contains antigens recognized by these cells, and  $T_{\text{RM}}$  cells express CD45RO consistent with memory T cells, they are genomically distinct from simply activated effector memory T cells. Recent studies in murine tumor models also support the importance of tumor-infiltrating T<sub>RM</sub> cells in mediating long-term control of melanoma tumors (31). The relationships between T<sub>RM</sub> cells and other populations such as stem memory cells implicated as targets of proliferative burst after PD-1 blockade need further study (37). Further studies are also needed to better characterize the proportion of tumor infiltrating  $T_{RM}$  cells that are truly tumor specific.

The concept that  $T_{RM}$  cells may be major targets of ICP blockade therapies is consistent with emerging insights into their functional properties. T<sub>RM</sub> cells seem to provide a dual role that encompasses both protection and regulation. Thus, while human T<sub>RM</sub> cells in NLTs can produce higher levels of effector cytokines, such as IFN $\gamma$ , IL2, and TNF, they also produce higher levels of immune regulatory cytokines such as IL10 (14, 15). Moreover, T<sub>RM</sub> cells also express higher levels of ICPs, such as CTLA4, PD-1, TIM-3, and LAG-3 (14, 25). T<sub>RM</sub> cells also seem to have a quiescent phenotype, which may be essential for their ability to survive long-term in tissues, being poised for activation but not harming tissues (17). Antibody-mediated blockade of ICPs such as PD-1, therefore, provides a mechanism for activation of these T cells in situ. The precise nature of the activation signal may differ between CTLA4 and PD-1 blockade (or combination thereof) (38).

The concept that  $T_{RM}$  cells within tumors may be major targets of ICP blockade has several implications for immune therapies. Vaccines that foster the generation of  $T_{RM}$  cells may be best suited for combination with ICP blockade (39). The ability of  $T_{RM}$  cells to mediate long-term residence in tissues may help to explain why clinical responses to ICP blockade have been durable. Along these lines, strategies that help to maintain or even enrich these  $T_{RM}$  pools may allow enhanced durability of responses. It would also be important to better understand the nature of antigenic targets on tumors recognized by these T cells, and the impact of tumor genetics as well as other cells in the tumor microenvironment on the functional properties and retention of these cells.

## DO T<sub>RM</sub> CELLS CONTRIBUTE TO INTRA-TUMOR HETEROGENEITY OF TUMORS?

Advances in cancer genomics and particularly the capacity to sequence multiple lesions in the same patient or even different parts of the same tumor have demonstrated a complex and heterogeneous landscape with varying sub-clonal architecture; studies have also suggested a potential impact of such intra-tumoral heterogeneity on clinical outcome (40, 41). However, the degree to which the genetics of the microenvironment contributes to intra-tumoral heterogeneity is less clear. Diversity within the immune microenvironment may in principle not only impact the mechanisms underlying response or resistance to immune therapies but also evolution of tumors in individual metastases. Advances in TCR sequencing provide an opportunity to gain some insights into the nature and genetics of T cells infiltrating tumor lesions. While the same antigenic epitope may in principle be recognized by different TCRs, they are likely to differ in terms of their affinity or functional properties.

In the setting of advanced or metastatic cancer, tumor cells grow as discrete lesions in diverse NLTs. These lesions by definition share the systemic circulation of the host and could in principle be likened to the situation in parabiotic mice that share systemic circulation. As discussed earlier, a characteristic feature of  $T_{RM}$ cells is tissue residence without recirculation, revealed by lack of equilibration in parabiotic mice. We hypothesized that if  $T_{RM}$  cells within individual tissues (e.g., lung or liver or skin lesions) indeed remain local, then dominant TCRs within individual metastatic lesions in the same patient would not equilibrate even if the oncogenic mutations or neoantigen-load were largely shared between these lesions (Figure 1). Concurrent sequencing of tumor cells as well as TCRs from individual lesions in patients with advanced melanoma supported this hypothesis; as expected, the interlesional diversity of TCRs was mostly accounted for by TCRs from  $T_{RM}$  subset of TILs (25). Differences in dominant TCRs between individual lesions from the same patient is consistent with lack of equilibration of TCRs between individual metastatic lesions even though they may share a major component of neoantigen load. However, the mechanisms that limit this equilibration need to be better defined; our current hypothesis is that it may relate to the lack of recirculation of tissue-resident TCRs, or their relative tissue retention, both consistent with  $T_{RM}$  biology.

The concept that  $T_{RM}$  cells infiltrating tumor tissues may exhibit local residence and little recirculation has several implications for immune therapies, immune monitoring, and cancer biology. If the individual metastatic lesions are established early, and carry different TCRs, then the level of immune pressure in individual lesions may differ and provide a pathway for divergent



genomic evolution (42). Along the same lines, it may be important to carefully consider the specific site of tissue biopsy when evaluating the results of immune monitoring. It should be noted, however, that the impact of ICP blockade on T<sub>RM</sub> homeostasis and redistribution in vivo in humans remains understudied and may add additional layers of complexity. Studies harvesting TILs for adoptive transfer are now entering the clinic in diverse cancers. If the dominant TCRs differ between individual lesions, it may be desirable to harvest and pool T cells from more than one lesion to optimize efficacy of such cell therapies. Finally, if the T cells in individual lesions differ, then it raises the potential that multiple mechanisms of immune resistance may be simultaneously operative in the same patient (43); along these lines, isolated progression at a single site in the face of continued regression at other sites may not reflect systemic loss of tumor control in the context of immune therapies. Clinicians have already come to appreciate this difference between immune therapies as compared to

## REFERENCES

- Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. Nat Rev Cancer (2012) 12(4):252–64. doi:10.1038/nrc3239
- Farber DL, Netea MG, Radbruch A, Rajewsky K, Zinkernagel RM. Immunological memory: lessons from the past and a look to the future. *Nat Rev Immunol* (2016) 16(2):124–8. doi:10.1038/nri.2016.13
- Chen DS, Mellman I. Oncology meets immunology: the cancer-immunity cycle. *Immunity* (2013) 39(1):1–10. doi:10.1016/j.immuni.2013.07.012
- Bindea G, Mlecnik B, Angell HK, Galon J. The immune landscape of human tumors: implications for cancer immunotherapy. *Oncoimmunology* (2014) 3(1):e27456. doi:10.4161/onci.27456

chemotherapies and often utilize localized therapies to tackle such lesions.

## SUMMARY

In summary,  $T_{RM}$  cells within tumor lesions are likely to gain increasing importance as targets of immune therapies as well as deeper understanding of cancer biology and evolution. It is likely that optimal integration of these immune therapies will require attention to the unique biology of these immune cells and exploit their regional nature of enhance tumor immunity with reduced systemic toxicity.

# **AUTHOR CONTRIBUTIONS**

The author confirms being the sole contributor of this work and approved it for publication.

- Mueller SN, Gebhardt T, Carbone FR, Heath WR. Memory T cell subsets, migration patterns, and tissue residence. *Annu Rev Immunol* (2013) 31:137–61. doi:10.1146/annurev-immunol-032712-095954
- Schenkel JM, Masopust D. Tissue-resident memory T cells. *Immunity* (2014) 41(6):886–97. doi:10.1016/j.immuni.2014.12.007
- Gebhardt T, Wakim LM, Eidsmo L, Reading PC, Heath WR, Carbone FR. Memory T cells in nonlymphoid tissue that provide enhanced local immunity during infection with herpes simplex virus. *Nat Immunol* (2009) 10(5):524–30. doi:10.1038/ni.1718
- Schenkel JM, Fraser KA, Masopust D. Cutting edge: resident memory CD8 T cells occupy frontline niches in secondary lymphoid organs. *J Immunol* (2014) 192(7):2961–4. doi:10.4049/jimmunol.1400003

- Mackay LK, Braun A, Macleod BL, Collins N, Tebartz C, Bedoui S, et al. Cutting edge: CD69 interference with sphingosine-1-phosphate receptor function regulates peripheral T cell retention. *J Immunol* (2015) 194(5):2059–63. doi:10.4049/jimmunol.1402256
- Sathaliyawala T, Kubota M, Yudanin N, Turner D, Camp P, Thome JJ, et al. Distribution and compartmentalization of human circulating and tissue-resident memory T cell subsets. *Immunity* (2013) 38(1):187–97. doi:10.1016/j.immuni.2012.09.020
- Djenidi F, Adam J, Goubar A, Durgeau A, Meurice G, de Montpreville V, et al. CD8+CD103+ tumor-infiltrating lymphocytes are tumor-specific tissue-resident memory T cells and a prognostic factor for survival in lung cancer patients. *J Immunol* (2015) 194(7):3475–86. doi:10.4049/jimmunol. 1402711
- Webb JR, Milne K, Nelson BH. Location, location, location: CD103 demarcates intraepithelial, prognostically favorable CD8 tumor-infiltrating lymphocytes in ovarian cancer. *Oncoimmunology* (2014) 3:e27668. doi:10.4161/ onci.27668
- Clark RA. Resident memory T cells in human health and disease. Sci Transl Med (2015) 7(269):269rv1. doi:10.1126/scitranslmed.3010641
- Kumar BV, Ma W, Miron M, Granot T, Guyer RS, Carpenter DJ, et al. Human tissue-resident memory T cells are defined by core transcriptional and functional signatures in lymphoid and mucosal sites. *Cell Rep* (2017) 20(12):2921–34. doi:10.1016/j.celrep.2017.08.078
- Hombrink P, Helbig C, Backer RA, Piet B, Oja AE, Stark R, et al. Programs for the persistence, vigilance and control of human CD8(+) lung-resident memory T cells. *Nat Immunol* (2016) 17(12):1467–78. doi:10.1038/ ni.3589
- Mackay LK, Minnich M, Kragten NA, Liao Y, Nota B, Seillet C, et al. Hobit and Blimp1 instruct a universal transcriptional program of tissue residency in lymphocytes. *Science* (2016) 352(6284):459–63. doi:10.1126/science. aad2035
- Boddupalli CS, Nair S, Gray SM, Nowyhed HN, Verma R, Gibson JA, et al. ABC transporters and NR4A1 identify a quiescent subset of tissue-resident memory T cells. *J Clin Invest* (2016) 126(10):3905–16. doi:10.1172/ JCI85329
- Milner JJ, Toma C, Yu B, Zhang K, Omilusik K, Phan AT, et al. Runx3 programs CD8(+) T cell residency in non-lymphoid tissues and tumours. *Nature* (2017) 552(7684):253–7. doi:10.1038/nature 24993
- Iijima N, Iwasaki A. T cell memory. A local macrophage chemokine network sustains protective tissue-resident memory CD4 T cells. *Science* (2014) 346(6205):93–8. doi:10.1126/science.1257530
- Mackay LK, Wynne-Jones E, Freestone D, Pellicci DG, Mielke LA, Newman DM, et al. T-box transcription factors combine with the cytokines TGF-beta and IL-15 to control tissue-resident memory T cell fate. *Immunity* (2015) 43(6):1101–11. doi:10.1016/j.immuni.2015.11.008
- Purwar R, Campbell J, Murphy G, Richards WG, Clark RA, Kupper TS. Resident memory T cells (T(RM)) are abundant in human lung: diversity, function, and antigen specificity. *PLoS One* (2011) 6(1):e16245. doi:10.1371/ journal.pone.0016245
- 22. Pallett LJ, Davies J, Colbeck EJ, Robertson F, Hansi N, Easom NJW, et al. IL-2(high) tissue-resident T cells in the human liver: sentinels for hepatotropic infection. *J Exp Med* (2017) 214(6):1567–80. doi:10.1084/jem.20162115
- 23. Okhrimenko A, Grun JR, Westendorf K, Fang Z, Reinke S, von Roth P, et al. Human memory T cells from the bone marrow are resting and maintain long-lasting systemic memory. *Proc Natl Acad Sci U S A* (2014) 111(25): 9229–34. doi:10.1073/pnas.1318731111
- Sercan Alp O, Durlanik S, Schulz D, McGrath M, Grun JR, Bardua M, et al. Memory CD8(+) T cells colocalize with IL-7(+) stromal cells in bone marrow and rest in terms of proliferation and transcription. *Eur J Immunol* (2015) 45(4):975–87. doi:10.1002/eji.201445295
- Boddupalli CS, Bar N, Kadaveru K, Krauthammer M, Pornputtapong N, Mai Z, et al. Interlesional diversity of T cell receptors in melanoma with immune checkpoints enriched in tissue-resident memory T cells. *JCI Insight* (2016) 1(21):e88955. doi:10.1172/jci.insight.88955
- Ganesan AP, Clarke J, Wood O, Garrido-Martin EM, Chee SJ, Mellows T, et al. Tissue-resident memory features are linked to the magnitude of cytotoxic T cell responses in human lung cancer. *Nat Immunol* (2017) 18(8):940–50. doi:10.1038/ni.3775

- Edwards J, Wilmott JS, Madore J, Gide TN, Quek C, Tasker A, et al. CD103+ tumor-resident CD8+ T cells are associated with improved survival in immunotherapy naive melanoma patients and expand significantly during anti-PD1 treatment. *Clin Cancer Res* (2018) 24(13):3036–45. doi:10.1158/1078-0432. CCR-17-2257
- Park SL, Zaid A, Hor JL, Christo SN, Prier JE, Davies B, et al. Local proliferation maintains a stable pool of tissue-resident memory T cells after antiviral recall responses. *Nat Immunol* (2018) 19(2):183–91. doi:10.1038/s41590-017-0027-5
- Muschaweckh A, Buchholz VR, Fellenzer A, Hessel C, Konig PA, Tao S, et al. Antigen-dependent competition shapes the local repertoire of tissue-resident memory CD8+ T cells. *J Exp Med* (2016) 213(13):3075–86. doi:10.1084/ jem.20160888
- Gros A, Robbins PF, Yao X, Li YF, Turcotte S, Tran E, et al. PD-1 identifies the patient-specific CD8+ tumor-reactive repertoire infiltrating human tumors. *J Clin Invest* (2014) 124(5):2246–59. doi:10.1172/JCI73639
- Malik BT, Byrne KT, Vella JL, Zhang P, Shabaneh TB, Steinberg SM, et al. Resident memory T cells in the skin mediate durable immunity to melanoma. *Sci Immunol* (2017) 2(10):eaam6346. doi:10.1126/sciimmunol.aam6346
- Sznol M, Chen L. Antagonist antibodies to PD-1 and B7-H1 (PD-L1) in the treatment of advanced human cancer. *Clin Cancer Res* (2013) 19(5):1021–34. doi:10.1158/1078-0432.CCR-12-2063
- Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N Engl J Med* (2012) 366(26):2443–54. doi:10.1056/NEJMoa1200690
- Korman AJ, Peggs KS, Allison JP. Checkpoint blockade in cancer immunotherapy. Adv Immunol (2006) 90:297–339. doi:10.1016/S0065-2776(06)90008-X
- Ribas A, Shin DS, Zaretsky J, Frederiksen J, Cornish A, Avramis E, et al. PD-1 blockade expands intratumoral memory T cells. *Cancer Immunol Res.* (2016) 4(3):194–203. doi:10.1158/2326-6066.CIR-15-0210
- Vasquez JC, Huttner A, Zhang L, Marks A, Chan A, Baehring JM, et al. SOX2 immunity and tissue resident memory in children and young adults with glioma. *J Neurooncol* (2017) 134(1):41–53. doi:10.1007/s11060-017-2515-8
- Im SJ, Hashimoto M, Gerner MY, Lee J, Kissick HT, Burger MC, et al. Defining CD8+ T cells that provide the proliferative burst after PD-1 therapy. *Nature* (2016) 537(7620):417–21. doi:10.1038/nature19330
- Das R, Verma R, Sznol M, Boddupalli CS, Gettinger SN, Kluger H, et al. Combination therapy with anti-CTLA-4 and anti-PD-1 leads to distinct immunologic changes in vivo. *J Immunol* (2015) 194(3):950–9. doi:10.4049/ jimmunol.1401686
- Nizard M, Roussel H, Diniz MO, Karaki S, Tran T, Voron T, et al. Induction of resident memory T cells enhances the efficacy of cancer vaccine. *Nat Commun* (2017) 8:15221. doi:10.1038/ncomms15221
- Jamal-Hanjani M, Quezada SA, Larkin J, Swanton C. Translational implications of tumor heterogeneity. *Clin Cancer Res* (2015) 21(6):1258–66. doi:10.1158/1078-0432.CCR-14-1429
- Sanborn JZ, Chung J, Purdom E, Wang NJ, Kakavand H, Wilmott JS, et al. Phylogenetic analyses of melanoma reveal complex patterns of metastatic dissemination. *Proc Natl Acad Sci U S A* (2015) 112(35):10995–1000. doi:10.1073/ pnas.1508074112
- Gerlinger M, McGranahan N, Dewhurst SM, Burrell RA, Tomlinson I, Swanton C. Cancer: evolution within a lifetime. *Annu Rev Genet* (2014) 48:215–36. doi:10.1146/annurev-genet-120213-092314
- Sharma P, Hu-Lieskovan S, Wargo JA, Ribas A. Primary, adaptive, and acquired resistance to cancer immunotherapy. *Cell* (2017) 168(4):707–23. doi:10.1016/j.cell.2017.01.017

**Conflict of Interest Statement:** The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Dhodapkar. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.