



Vascular Targeting to Increase the Efficiency of Immune Checkpoint Blockade in Cancer

Maria Georganaki[†], Luuk van Hooren[†] and Anna Dimberg^{*}

Department of Immunology, Genetics and Pathology, Science for Life Laboratory, Uppsala University, The Rudbeck Laboratory, Uppsala, Sweden

Boosting natural immunity against malignant cells has had a major breakthrough in clinical cancer therapy. This is mainly due to the successful development of immune checkpoint blocking antibodies, which release a break on cytolytic anti-tumor-directed T-lymphocytes. However, immune checkpoint blockade is only effective for a proportion of cancer patients, and a major challenge in the field is to understand and overcome treatment resistance. Immune checkpoint blockade relies on successful trafficking of tumor-targeted T-lymphocytes from the secondary lymphoid organs, through the blood stream and into the tumor tissue. Resistance to therapy is often associated with a low density of T-lymphocytes residing within the tumor tissue prior to treatment. The recruitment of leukocytes to the tumor tissue relies on up-regulation of adhesion molecules and chemokines by the tumor vasculature, which is denoted as endothelial activation. Tumor vessels are often poorly activated due to constitutive pro-angiogenic signaling in the tumor microenvironment, and therefore constitute barriers to efficient leukocyte recruitment. An emerging possibility to enhance the efficiency of cancer immunotherapy is to combine pro-inflammatory drugs with anti-angiogenic therapy, which can enable tumor-targeted T-lymphocytes to access the tumor tissue by relieving endothelial anergy and increasing adhesion molecule expression. This would pave the way for efficient immune checkpoint blockade. Here, we review the current understanding of the biological basis of endothelial anergy within the tumor microenvironment, and discuss the challenges and opportunities of combining vascular targeting with immunotherapeutic drugs as suggested by data from key pre-clinical and clinical studies.

OPEN ACCESS

Edited by:

Patrik Andersson, Harvard Medical School, United States

Reviewed by:

Lasse Dahl Ejby Jensen, Linköping University, Sweden Krithika Kodumudi, Moffitt Cancer Center, United States

> *Correspondence: Anna Dimberg anna.dimberg@igp.uu.se

[†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Cancer Immunity and Immunotherapy, a section of the journal Frontiers in Immunology

> Received: 28 September 2018 Accepted: 12 December 2018 Published: 21 December 2018

Citation:

Georganaki M, van Hooren L and Dimberg A (2018) Vascular Targeting to Increase the Efficiency of Immune Checkpoint Blockade in Cancer. Front. Immunol. 9:3081. doi: 10.3389/fimmu.2018.03081 Keywords: angiogenesis, cancer, checkpoint blockade, PD-1, PD-L1, CTLA-4, VEGF, endothelial activation

INTRODUCTION

The field of cancer immunotherapy has made significant improvements during the last decade due to the development of new effective means to boost tumor immune responses and achieve long-term remission or even cures in patients that were previously deemed to be untreatable. A major breakthrough was the development of antibodies targeting negative regulators of T-cell activation, termed immune checkpoints. Ipilimumab, an antagonistic antibody targeting cytotoxic T-lymphocyte-associated protein 4 (CTLA-4) improved overall survival in metastatic melanoma patients in 2010 (1). Following the success of anti-CTLA-4 therapy, antibodies targeting programmed cell death protein 1 (PD-1), or its ligand PD-L1,

1

proved to be effective at improving overall survival in a wide variety of cancers (2–7). Importantly, a proportion of patients achieve long-term remission, highlighting the potential of immune checkpoint blockade to induce durable responses (8). The encouraging results of these studies has sparked an interest from the cancer research field and inspired further investigations into targeting of alternative immune checkpoint molecules.

While checkpoint blockade represents a breakthrough in cancer therapy, a majority of cancer patients do not respond and some tumor types appear to be intrinsically resistant. The treatment is designed to boost an ongoing immune response and is inefficient in cases where initial immune activation is lacking, including tumors that are devoid of infiltrating T-cells (3, 9). Development of therapeutic strategies to enhance immune cell recruitment may therefore increase the proportion of patients responding to immune checkpoint blockade. Circulating Tcells are recruited through expression of adhesion molecules and chemokines on the endothelial cells, collectively mediating capture, rolling, and transmigration of leukocytes from the blood stream into the inflamed tissue (10). In many types of cancer, constitutive stimulation by pro-angiogenic factors secreted in the tumor microenvironment renders the vasculature morphologically and functionally abnormal, constituting a barrier to efficient leukocyte recruitment. In this mini-review we summarize phenotypical differences between normal vessels and tumor vessels in mediating leukocyte recruitment, the molecular mechanisms that underlie these functional changes and current efforts to improve immune checkpoint blockade through vascular targeting.

IMMUNE CHECKPOINT BLOCKADE THERAPY RELIES ON EFFICIENT T-LYMPHOCYTE RECRUITMENT

Immune checkpoint blockade works through inhibiting negative feedback loops that downregulate T-cell activation following an initial immune response. T-cell activation and T-cell receptor signaling has recently been reviewed in detail (11, 12). T-cells remain naïve until they encounter licensed antigen-presenting cells (APC)s that present the correct peptide antigen on major histocompatibility complex (MHC) molecules together with the appropriate co-stimulatory molecules. T-cell activation requires recognition of the MHC-antigen complex displayed on an APC, engagement of co-stimulatory molecules such as CD28 on the T cell with B7 family members on the APC and stimulation by inflammatory cytokines. In response to T-cell activation, other co-stimulatory molecules such as ICOS and OX40 are expressed, but also molecules that instigate negative feedback loops to prevent over-activation of T-cells. One of those negative feedback molecules is CTLA-4, which competes with CD28 for binding to B7 family members expressed on the surface of APCs (13-16). CTLA-4 is also highly expressed on regulatory T cells, and antibodies targeting CTLA-4 have been suggested to deplete them from the tumor microenvironment through Fc effector functions (17). Although the relative importance of the immune checkpoint and regulatory depletion mechanisms for therapeutic efficacy is still under active debate (16), blocking CTLA-4 in cancer enhances T-cell activation, but can also lead to autoimmune responses.

After activation, which generally occurs in secondary lymphoid organs, T-cells circulate and extravasate through the vasculature at sites of inflammation to locate and kill target cells displaying the cognate peptide antigen on their MHC molecules. At the tumor site, T-cell activity can be hampered by several types of immunosuppression, including engagement of PD-1 expressed on T-cells by its ligand PD-L1 expressed on stromal cells and/or malignant cells (18, 19). Thus, anti-cancer immunity can be enhanced by antibodies that block the PD-1/PD-L1 interaction. Although manipulating T-cell activation status by blocking inhibitory receptors or enhancing co-stimulatory molecules has proven to be efficacious in boosting anti-tumor immune responses, these treatments strictly rely on efficient transport of lymphocytes from the site of T-cell activation to the tumor tissue. It is therefore not surprising that tumors that are not infiltrated by T-cells, and tumors where T-cell infiltration is only observed at the tumor border but not in the core, do not respond well to immune checkpoint blockade (3, 9). Several mechanisms contribute to regulating the inflammatory state, including the mutational landscape of the tumor, expression of chemokines, and checkpoint molecules and recruitment of immunosuppressive cells (20, 21). In cases where an immune response is correctly mounted but where lymphocyte recruitment to the tumor tissue is lacking, pharmacologically altering vascular phenotype to allow efficient leukocyte trafficking may sensitize resistant tumors for immunotherapy.

LYMPHOCYTE RECRUITMENT INVOLVES LEUKOCYTE/ENDOTHELIAL INTERACTION

Leukocyte recruitment by activated endothelial cells and subsequent migration through the vessel wall is mediated by direct molecular interactions between proteins expressed by leukocytes and endothelial cells (Figure 1A). This finely tuned process, known as the leukocyte adhesion cascade, involves leukocyte capture, rolling, adhesion, arrest, and transendothelial migration (10). This is enabled by up-regulation of adhesion molecules and chemokines on the surface of endothelial cells, denoted "endothelial activation." Leukocyte capture and rolling are mainly mediated by interaction between selectins expressed on endothelial cells (P-selectin and Eselectin) and leukocytes (L-selectin) to carbohydrate ligands including P-selectin glycosylated ligand 1. Firm adhesion of leukocytes is mediated through interaction of leukocyte integrins with endothelial adhesion molecules. For T-lymphocytes, firm adhesion is mainly induced by lymphocyte function-associated antigen (LFA)-1 and very late activation antigen (VLA)-4 binding to endothelial intercellular adhesion molecule (ICAM)-1 and vascular cell adhesion molecule (VCAM)-1, respectively. Activation of integrins via inside-out signaling associated with chemokine stimulation triggers leukocyte arrest to the endothelium (10). Blood flow-derived shear stress contributes



tumor microenvironment gives rise to an anergic endothelium with reduced pericyte coverage, disrupted endothelial cell junctions, and suboptimal activation status. Anti-angiogenic therapy reverts those defects and permits for enhanced leukocyte recruitment, through the leukocyte adhesion cascade. Chemokines and adhesion molecules on the activated endothelial surface allow for leukocyte capture, rolling, arrest, and transendothelial migration into the tumor tissue. **(B)** Aberrant pro-angiogenic signaling in tumors is associated with dysfunctional and anergic tumor vessels, which are not capable of recruiting tumor-targeted leukocytes (left panel). Vascular targeting can relieve endothelial anergy, improve perfusion and increase the recruitment of leukocytes into the tumor microenvironment (right panel).

to efficient leukocyte capture and integrin activation through mechanical forces (22). Transendothelial migration can occur through either through paracellular or transcellular pathways (10, 23). Finally, leukocytes migrate through the basement membrane and pericyte layer to reach the inflamed tissue (10). Recruitment of lymphocytes to the tumor tissue strictly depends on efficient regulation of molecules required for cell-cell interactions during capture, rolling, adhesion, and transendothelial migration.

TUMOR ANGIOGENESIS RESULTS IN MORPHOLOGICALLY AND FUNCTIONALLY DISTINCT VESSELS

Tumors need access to capillary network to proliferate, and the ability of tumors to stimulate angiogenesis is recognized as

one of the hallmarks of cancer (24). Angiogenesis is induced as a result of enhanced growth factor secretion in the tumor microenvironment, shifting the balance from predominantly angiostatic to pro-angiogenic signaling (25). This "angiogenic switch," observed as a shift from avascular to vascular tumors, can occur in dormant, and slow growing tumors and be associated with tumor progression to higher malignancy grades.

Several mechanisms can trigger neovascularization in tumors, including hypoxia, genetic alterations in tumor cells, expression of cytokines, and growth factorsm and recruitment of bone marrow-derived circulating cells (26–28). When proliferation of malignant cells results in a tumor mass that cannot be sufficiently oxygenated by pre-existing vasculature this leads to hypoxia. Hypoxia-induced stabilization of hypoxia-inducible factor (HIF)-1 α triggers up-regulation of its target genes, including several pro-angiogenic genes such as vascular endothelial growth factor (VEGF) (29). VEGF secreted by

tumor cells diffuses through the tissue and activates its receptor VEGFR2 expressed on endothelial cells (30). Downstream of VEGFR2 activation, multiple intracellular pathways are induced that regulate cell division, survival, sprouting, and migration of endothelial cells (30). Several other pro-angiogenic growth factors contribute to tumor angiogenesis, including the family of angiopoietins and their cognate receptor TIE-2 and the fibroblast growth factor family (31). Some tumors harbor mutations of the gene coding for the von Hippel-Lindau protein, a crucial member of the ubiquitin ligase complex that degrades HIF-1 α (32). These mutations stabilize HIF-1 α , allowing expression of pro-angiogenic factors under normoxic conditions. Myeloid cells, including macrophages, neutrophils, and myeloid derived suppressor cells (MDSCs), can also stimulate vessel formation through expression of pro-angiogenic factors and/or matrix metalloproteases that release VEGF from extracellular matrix (33).

Physiological angiogenesis is a well-controlled process that is attenuated when the need for new vessels have been met, but tumor angiogenesis is deregulated and continuous due to excessive expression of pro-angiogenic factors (34). Tumor angiogenesis can give rise to disorganized vessels that are tortuous, dilated and poorly covered by pericytes (35). The tumor vasculature is often leaky due to endothelial junctional defects, blood flow is generally slow and perfusion is irregular (25). Gene expression analyses have shown that tumor vessels differ molecularly from their normal counterparts and have revealed a high level of vessel heterogeneity depending on the resident tumor tissue (36–40). Importantly, tumor vessels can have multiple phenotypes ranging from normal to dysfunctional and the morphology and functionality significantly differ depending on tumor type and anatomical site.

TUMOR BLOOD VESSELS ARE BARRIERS TO EFFICIENT LEUKOCYTE RECRUITMENT

Immune cells in the circulation are dependent on the vascular network to reach the tumor and kill malignant cells. However, functional abnormalities of tumor blood vessels represent difficult hurdles for leukocyte recruitment. The architectural defects of tumor vessels limit perfusion and alter sheer stress, and differential protein expression in tumor endothelial cells can dampen the immune response (34, 41-44). Tumor endothelial cell respond inefficiently to pro-inflammatory signaling, and fail to express sufficient levels of molecules involved in the leukocyte capture, adhesion and extravasation process (Figure 1). Downregulation or ineffective clustering of adhesion molecules on tumor endothelial cells limits T-cell infiltration and inhibit anti-tumor immunity (45-47). Reduced expression of adhesion molecules in tumor vessels has been observed in several types of human cancer (48-50). Endothelial activation is generally induced by binding of pro-inflammatory cytokines such as tumor necrosis factor (TNF)a and interleukin (IL)-1 to their endothelial receptors, leading to activation of the transcription factor nuclear factor-KB (NF-KB) and up-regulation of selectins, adhesion molecules and chemokines (51). Pro-inflammatory cytokines are abundantly expressed in many cancers, but proangiogenic factors present in the tumor microenvironment can suppress expression of adhesion molecules and chemokines that attract cytolytic T-cells and NK cells such as CXCL10 and CXCL11 (41, 52, 53). VEGF-induced signaling pathways can directly interfere with TNF-α-induced NF-κB activation, globally repressing TNF-a-induced gene expression in endothelial cells (53). Consistent with this, antagonizing VEGFR2 signaling sensitizes endothelial cells to TNF- α (54). However, the interplay between angiogenesis and inflammation is context dependent. TNF-a stimulation synergistically primes endothelial cells for VEGF-induced angiogenesis (55). Notably, VEGF stimulation can induce leukocyte infiltration in some systems, and pathways downstream of VEGF signaling can both induce and repress adhesion molecule expression (56-58). Nitric oxide and molecules such as epidermal growth factor-like domain 7 can also regulate adhesion molecule expression and clustering in tumors (59, 60). Another less studied feature of endothelial regulation of tumor immunity is the selective recruitment of immunosuppressive leukocytes through expression of specific adhesion molecules such as the common lymphatic endothelial and vascular endothelial receptor-1 (CLEVER-1) (41).

In addition to regulating leukocyte entry, tumor endothelial cells can alter the anti-tumor immune response by modulating immune cell activity or viability. This can occur as a response of endothelial cells to tumor-derived growth factors (61). The concept of a "tumor endothelial barrier" refers to molecules expressed on endothelial cells that inhibit promote T-cell arrest. An example of this is tumor endothelial upregulation of FasL in response to tumor-derived VEGF, IL-10 and prostaglandin E2, which has been shown to selectively kill effector CD8 Tcells but not Treg cells (44, 62). Endothelial cells can express several inhibitory molecules including immune checkpoint molecules [PD-L1, T-cell immunoglobulin domain and mucin domain (TIM3), B7-H3 and B7-H4], death receptor-ligands (TNF-related apoptosis-inducing ligand (TRAIL) and secreted immunomodulatory factors (IL-6, prostaglandin E (PGE) 2, IL-10, and TGF-β) (44, 62). The relative importance of endothelial expression of these molecules in immunosuppression and their regulation in tumor vessels need further investigation. Antigen presentation by endothelial cells suggests that they can function as potential antigen presenting cells (63). Whether tumor endothelial cells present antigen and if this is sufficient for activation of T-cells, or alternatively induces T-cell anergy, is still unknown. As discussed below, anti-angiogenic therapies can alleviate endothelial anergy and enhance T-cell recruitment in tumors (53, 64-67). Immunosuppressive molecules expressed on tumor endothelial cells represent new potential targets for novel combination treatments with immunotherapy.

SUCCESSES AND FAILURES OF ANTI-ANGIOGENIC THERAPY

The idea that anti-angiogenic therapy could block tumor progression by depriving the tumor cells of oxygen and

TABLE 1 Selected studies combining anti-angiogenic therapy with immune checkpoint blockade in preclinical models and clinical trials.

Anti-angiogenic target	Immune checkpoint target	Cancer model	Survival	References
PRECLINICAL MODELS				
VEGF (B20-4.1.1)	PD-L1 (6E11)	SCLC	+ **	(81)
VEGFR2 (DC101)	PD-1 (RMPI-14)	Colon-26 adenocarcinoma	+	(82)
VEGF and ANG2 (Vanucizumab)	PD-1 (RMPI-14)	MMTV-PyMT, RIP1-Tag2, Melanoma, Neuroendocrine	+ ***	(83)
VEGFR-1,-2 and—3 (Axitinib)	CTLA-4 (9H10)	Melanoma	+	(84)
VEGFR2* (Sunitinib)	PD-1 (RMPI-14)	Colon cancer	+	(85)
VEGFR2 (DC101)	PD-L1 (10F.9G2)	Pancreatic cancer, breast cancer and glioblastoma	+	(86)
VEGF + ANG2 (10F.9G2 + CVX-241)	PD-L1 (10F.9G2)	Breast cancer	+/-	(87)
Anti-angiogenic target	Immune checkpoint target	Cancer type	Trial	References
CLINICAL TRIALS				
VEGFR-1,-2 and—3 (Axitinib)	PD-1 (Pembrolizumab)	Renal cell cancer	Phase 3	(88)
VEGF (Bevacizumab)	CTLA-4 (Ipilimumab)	Metastatic melanoma	Phase 1	(89)
				(90)
				(91)
VEGFR-1,-2 and-3 (Axitinib)	PD-L1 (Avelumab)	Advanced clear-cell renal cell carcinoma	Phase 1b	(92)
VEGFR-1,-2 and-3 (Lenvatinib)	PD-1 (Pembrolizumab)	Renal cell cancer	Retrospective	(93)
VEGF (Bevacizumab)	PD-L1 (Atezolizumab)	Metastatic renal cell carcinoma	Phase 1b	(94)

Antibody clone or brand name in brackets. SCLC = small-cell lung cancer, * broad tyrosine kinase inhibitor, ** increased T-cell exhaustion, *** increased T-cell numbers and endothelial activation. Ongoing clinical trials are available at www.clinicaltrials.gov and were recently reviewed by Fukumura et al. (95).

nutrients (68) led to intense research efforts and sparked numerous clinical trials. A number of anti-angiogenic drugs have been approved to date, several of which are antibodies or small tyrosine kinase inhibitors that target VEGF/VEGFR signaling (69). The first clinically approved drug was a humanized antibody targeting VEGF named Bevacizumab. Treatment with Bevacizumab slows tumor growth in patients with non-small cell lung and colorectal cancer, though with only a marginal improvement of longterm survival (70, 71). It has also been approved for patients with cervical cancer, glioblastoma, ovarian cancer and renal cell carcinoma (72). In breast, melanoma, pancreatic, and prostate cancer no improvement of overall survival has been observed (73).

Treatment of colorectal cancer patients with Bevacizumab results in an initial response with decreased tumor growth or even regression. However, relapse is common, associated with rapid rebound angiogenesis, and tumor regrowth is often more aggressive than before anti-angiogenic treatment (74). Several mechanisms have been proposed for the resistance to anti-angiogenic treatment, including co-option of normal vessels in the surrounding tissue, recruitment of pro-angiogenic myeloid cells and upregulation of alternative pro-angiogenic factors (25). Notably, anti-angiogenic treatment can increase invasiveness and promote metastasis formation in experimental models of cancer (75, 76). Although metastasis-promoting effects of anti-angiogenic therapy have not been observed in clinical studies, the pre-clinical work has cautioned the field and questioned how anti-angiogenic therapy should best be administrated (77).

ANTI-ANGIOGENIC THERAPY CAN IMPROVE THE EFFECT OF IMMUNE CHECKPOINT BLOCKADE

The importance of a functional vasculature for immune cell recruitment justifies efforts of combining immunotherapy with vascular targeting to improve vessel function and enhance up-regulation of adhesion molecules and chemokines. Inhibition of angiogenic signaling using sub-maximal doses of antiangiogenic drugs may result in a normalization of vascular function and improve the efficacy of other anti-cancer drugs, as proposed by Jain (78). Anti-angiogenic therapy provides relief of continuous angiogenic signaling, which at sub-maximal doses can result in vessel pruning, maturation, and improved perfusion (69). For cancer immunotherapy, there is an added benefit that anti-angiogenic drugs enhance expression of adhesion molecules and chemokines involved in T-cell recruitment (53, 64-67). Therefore, combining immunotherapy with antiangiogenic drugs may relieve endothelial anergy and induce lymphocyte infiltration into tumors that prior to treatment were of an immune-excluded phenotype (Figure 1B). Indeed, by combining adoptive T-cell transfer with anti-VEGF therapy in murine melanoma, tumor T-cell infiltration was increased and survival was prolonged (79). An important challenge in this concept is that the dosing of anti-angiogenic drugs is crucial for normalizing vessels and improving T-cell recruitment, and that the optimal dose may differ between patients (80). Nevertheless, the combination of immunotherapy and anti-angiogenic therapy has shown benefit in various therapeutic settings (Table 1).

Drugs targeting VEGF/VEGFR2 signaling have been observed to enhance the response to immune checkpoint antibodies in pre-clinical tumor models. The combination of anti-VEGF and anti-VEGFR2 antibodies prolonged survival in a murine model of adenocarcinoma in combination with PD-1 blockade (82). Similarly, an antibody targeting both VEGF and Angiopoeitin-2 improved responses to PD-1 inhibition in preclinical cancer models (83). The VEGFR inhibitor axitinib combined with anti-CTLA-4, but neither monotherapy, prolonged survival of mice bearing murine melanoma (84). This was associated with increased numbers of CD4⁺ and CD8⁺ T cells in the tumor after the combination treatment. In addition to their effect on vessel phenotype, therapies targeting pro-angiogenic factors can alleviate immunosuppression by directly affecting the immune cells. For example, the tyrosine kinase inhibitor Sunitinib can decrease MDSCs and Tregs (67, 96, 97).

The first phase I clinical trial combining anti-angiogenic therapy with immune checkpoint blockade was a study using Bevacizumab and ipilimumab (anti-CTLA-4). The combination therapy modulated tumor vessel morphology and induced endothelial activation, associated with increased infiltration of dendritic cells and cytotoxic T-cells in melanoma tumors (89, 98). Similarly, combining atezolizumab (anti-PD-L1) with Bevacizumab in patients with metastatic renal cell carcinoma resulted in enhanced trafficking of lymphocytes, and increased cytotoxic T cells (94). Following these promising results, several clinical trials with the same therapeutic rationale have been initiated (95, 98, 99).

FUTURE DIRECTIONS BEYOND NORMALIZATION AND ENDOTHELIAL ACTIVATION

An emerging concept is that vascular targeting in combination with immune checkpoint blockade may promote tumor immunity by inducing formation of high-endothelial venules (HEV)s. HEVs are specialized vessels found in secondary lymphoid organs that are adapted for lymphocyte trafficking (100). The combination of anti-VEGFR2 antibodies with PD-L1 antibodies induced formation of HEVs and improved T-cell infiltration in the polyoma middle T oncoprotein (PyMT) breast cancer model and the Rip1-Tag2 pancreatic neuroendocrine tumor model (RT2-PNET) (86). Formation of HEVs in glioblastoma models required further stimulation using a lymphotoxin β receptor agonistic antibody, resulting

REFERENCES

- Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, et al. Improved survival with ipilimumab in patients with metastatic melanoma. N Engl J Med. (2010) 363:711–23. doi: 10.1056/NEJMoa1003466
- Hamid O, Robert C, Daud A, Hodi FS, Hwu WJ, Kefford R, et al. Safety and tumor responses with lambrolizumab (anti-PD-1) in melanoma. N Engl J Med. (2013) 369:134–44. doi: 10.1056/NEJMoa1305133
- Herbst RS, Soria JC, Kowanetz M, Fine GD, Hamid O, Gordon MS, et al. Predictive correlates of response to the anti-PD-L1 antibody MPDL3280A

in enhanced T-cell infiltration and reduced tumor growth (86). Vessel normalization in combination with a vascular targeting peptide coupled to LIGHT, a ligand for the lymphotoxin β receptor, induced HEVs and tertiary lymphoid structures in Rip1-Tag5 pancreatic neuroendocrine tumors. Importantly, this therapeutic approach sensitized these tumors to anti-PD-1 and anti-CTLA-4 antibody therapy (101). These studies indicate that beyond normalizing vessels, transforming tumor vessels to HEVs can be of additional benefit in enhancing the response to cancer immunotherapy. Furthermore, HEVs may promote formation of tertiary lymphoid structures which have been associated with a beneficial response to cancer immunotherapy in several types of cancer (100, 102).

Current efforts in vascular targeting aim to improve the efficacy of cancer immunotherapy through inhibition of proangiogenic signaling. However, several immunosuppressive molecules that contribute to the tumor endothelial barrier are regulated through alternative pathways, and may be induced secondary to immune activation. This aspect has not yet been sufficiently explored. An increased understanding of the crosstalk between tumor cells, endothelial cells, and immune cells during immune checkpoint blockade therapy may lead to new combinatorial treatment regimens that enhance the abundance of activated T-cells in tumor tissue. This can ultimately increase the proportion of patients that respond to immune checkpoint blockade.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

FUNDING

Swedish Cancer Society (CAN 2017/502), the Swedish Childhood Cancer Society (PR2015-0133, NCP2015-0075), the Swedish Research Council (Dnr 2016-02495), the Emil and Wera Cornells Stiftelse foundation, Senior Investigator Award from the Swedish Cancer Society (CAN 2015/1216) to AD.

ACKNOWLEDGMENTS

We apologize to the authors of original work that was not cited in this mini-review due to space constraint.

in cancer patients. *Nature* (2014) 515:563-7. doi: 10.1038/nature 14011

- Powles T, Eder JP, Fine GD, Braiteh FS, Loriot Y, Cruz C, et al. MPDL3280A (anti-PD-L1) treatment leads to clinical activity in metastatic bladder cancer. *Nature* (2014) 515:558–62. doi: 10.1038/nature13904
- Topalian SL, Sznol M, McDermott DF, Kluger HM, Carvajal RD, Sharfman WH, et al. Survival, durable tumor remission, and long-term safety in patients with advanced melanoma receiving nivolumab. J Clin Oncol. (2014) 32:1020–30. doi: 10.1200/JCO.2013. 53.0105

- Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N Engl J Med.* (2012) 366:2443–54. doi: 10.1056/NEJMoa1200690
- Ribas A, Wolchok JD. Cancer immunotherapy using checkpoint blockade. Science (2018) 359:1350–5. doi: 10.1126/science.aar4060
- Schadendorf D, Hodi FS, Robert C, Weber JS, Margolin K, Hamid O, et al. Pooled analysis of long-term survival data from phase II and phase III trials of ipilimumab in unresectable or metastatic melanoma. *J Clin Oncol.* (2015) 33:1889–94. doi: 10.1200/JCO.2014.56.2736
- Tumeh PC, Harview CL, Yearley JH, Shintaku IP, Taylor EJ, Robert L, et al. PD-1 blockade induces responses by inhibiting adaptive immune resistance. *Nature* (2014) 515:568–71. doi: 10.1038/nature13954
- Ley K, Laudanna C, Cybulsky MI, Nourshargh S. Getting to the site of inflammation: the leukocyte adhesion cascade updated. *Nat Rev Immunol.* (2007) 7:678–89. doi: 10.1038/nri2156
- Gaud G, Lesourne R, Love PE. Regulatory mechanisms in T cell receptor signalling. Nat Rev Immunol. (2018) 18:485–97. doi: 10.1038/s41577-018-0020-8
- 12. Golstein P, Griffiths GM. An early history of T cell-mediated cytotoxicity. *Nat Rev Immunol.* (2018) 18:527–35. doi: 10.1038/s41577-018-0009-3
- Walunas TL, Lenschow DJ, Bakker CY, Linsley PS, Freeman GJ, Green JM, et al. CTLA-4 can function as a negative regulator of T cell activation. *Immunity* (1994) 1:405–13. doi: 10.1016/1074-7613(94)90071-X
- Linsley PS, Greene JL, Brady W, Bajorath J, Ledbetter JA, Peach R. Human B7-1 (CD80) and B7-2 (CD86) bind with similar avidities but distinct kinetics to CD28 and CTLA-4 receptors. *Immunity* (1994) 1:793-801. doi: 10.1016/S1074-7613(94)80021-9
- van der Merwe PA, Bodian DL, Daenke S, Linsley P, Davis SJ. CD80 (B7–1) binds both CD28 and CTLA-4 with a low affinity and very fast kinetics. *J Exp Med.* (1997) 185:393–403. doi: 10.1084/jem.185.3.393
- Wei SC, Duffy CR, Allison JP. Fundamental mechanisms of immune checkpoint blockade therapy. *Cancer Discov.* (2018) 8:1069–86. doi: 10.1158/2159-8290.CD-18-0367
- Arce Vargas F, Furness AJS, Litchfield K, Joshi K, Rosenthal R, Ghorani E, et al. Fc effector function contributes to the activity of human Anti-CTLA-4 antibodies. *Cancer Cell.* (2018) 33:649–63 e4. doi: 10.1016/j.ccell.2018.02.010
- Dong H, Zhu G, Tamada K, Chen L. B7-H1, a third member of the B7 family, co-stimulates T-cell proliferation and interleukin-10 secretion. *Nat Med.* (1999) 5:1365–9. doi: 10.1038/70932
- Freeman GJ, Long AJ, Iwai Y, Bourque K, Chernova T, Nishimura H, et al. Engagement of the PD-1 immunoinhibitory receptor by a novel B7 family member leads to negative regulation of lymphocyte activation. *J Exp Med.* (2000) 192:1027–34. doi: 10.1084/jem.192.7.1027
- Popovic A, Jaffee EM, Zaidi N. Emerging strategies for combination checkpoint modulators in cancer immunotherapy. J Clin Invest. (2018) 128:3209–18. doi: 10.1172/JCI120775
- Binnewies M, Roberts EW, Kersten K, Chan V, Fearon DF, Merad M, et al. Understanding the tumor immune microenvironment (TIME) for effective therapy. *Nat Med.* (2018) 24:541–50. doi: 10.1038/s41591-018-0014-x
- 22. Alon R, Dustin ML. Force as a facilitator of integrin conformational changes during leukocyte arrest on blood vessels and antigen-presenting cells. *Immunity* (2007) 26:17–27. doi: 10.1016/j.immuni.2007.01.002
- 23. Filippi MD. Mechanism of diapedesis: importance of the transcellular route. *Adv Immunol.* (2016) 129:25–53. doi: 10.1016/bs.ai.2015.09.001
- Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell* (2000) 100:57–70. doi: 10.1016/S0092-8674(00)81683-9
- Bergers G, Benjamin LE. Tumorigenesis and the angiogenic switch. Nat Rev Cancer. (2003) 3:401–10. doi: 10.1038/nrc1093
- Ferrara N, Gerber H-P, LeCouter J. The biology of VEGF and its receptors. Nat Med. (2003) 9:669–76. doi: 10.1038/nm0603-669
- Yadav L, Puri N, Rastogi V, Satpute P, Sharma V. Tumour angiogenesis and angiogenic inhibitors: a review. J Clin Diagn Res. (2015) 9:XE01-XE5. doi: 10.7860/JCDR/2015/12016.6135
- Fang S, Salven P. Stem cells in tumor angiogenesis. J Mol Cell Cardiol. (2011) 50:290–5. doi: 10.1016/j.yjmcc.2010. 10.024
- Liao D, Johnson RS. Hypoxia: a key regulator of angiogenesis in cancer. Cancer Metastasis Rev. (2007) 26:281–90. doi: 10.1007/s10555-007-9066-y

- Olsson AK, Dimberg A, Kreuger J, Claesson-Welsh L. VEGF receptor signalling - in control of vascular function. *Nat Rev Mol Cell Biol.* (2006) 7:359–71. doi: 10.1038/nrm1911
- Welti J, Loges S, Dimmeler S, Carmeliet P. Recent molecular discoveries in angiogenesis and antiangiogenic therapies in cancer. J Clin Invest. (2013) 123:3190–200. doi: 10.1172/JCI70212
- Miller F, Kentsis A, Osman R, Pan ZQ. Inactivation of VHL by tumorigenic mutations that disrupt dynamic coupling of the pVHL.hypoxia-inducible transcription factor-1alpha complex. J Biol Chem. (2005) 280:7985–96. doi: 10.1074/jbc.M413160200
- Rivera LB, Bergers G. Intertwined regulation of angiogenesis and immunity by myeloid cells. *Trends Immunol.* (2015) 36:240–9. doi: 10.1016/j.it.2015.02.005
- De Bock K, Cauwenberghs S, Carmeliet P. Vessel abnormalization: another hallmark of cancer? Molecular mechanisms and therapeutic implications. *Curr Opin Genet Dev.* (2011) 21:73–9. doi: 10.1016/j.gde.2010.10.008
- Baluk P, Hashizume H, McDonald DM. Cellular abnormalities of blood vessels as targets in cancer. *Curr Opin Genet Dev.* (2005) 15:102–11. doi: 10.1016/j.gde.2004.12.005
- Langenkamp E, Molema G. Microvascular endothelial cell heterogeneity: general concepts and pharmacological consequences for antiangiogenic therapy of cancer. *Cell Tissue Res.* (2009) 335:205–22. doi: 10.1007/s00441-008-0642-4
- Dieterich LC, Mellberg S, Langenkamp E, Zhang L, Zieba A, Salomäki H, et al. Transcriptional profiling of human glioblastoma vessels indicates a key role of VEGF-A and TGFβ2 in vascular abnormalization. *J Pathol.* (2012) 228:378–90. doi: 10.1002/path.4072
- Masiero M, Simões FC, Han HD, Snell C, Peterkin T, Bridges E, et al. A core human primary tumor angiogenesis signature identifies the endothelial orphan receptor ELTD1 as a key regulator of angiogenesis. *Cancer Cell* (2013) 24:229–41. doi: 10.1016/j.ccr.2013.06.004
- Aird WC. Endothelial cell heterogeneity. Cold Spring Harb Perspect Med. (2012) 2:a006429. doi: 10.1101/cshperspect.a006429
- Zhang L, He L, Lugano R, Roodakker K, Bergquist M, Smits A, et al. IDH-mutation status is associated with distinct vascular gene expression signatures in lower grade gliomas. *Neuro Oncol.* (2018) 20:1505–16. doi: 10.1093/neuonc/noy088
- Mauge L, Terme M, Tartour E, Helley D. Control of the adaptive immune response by tumor vasculature. *Front Oncol.* (2014) 4:61. doi: 10.3389/fonc.2014.00061
- 42. Slaney CY, Kershaw MH, Darcy PK. Trafficking of T cells into tumors. *Cancer Res.* (2014) 74:7168–74. doi: 10.1158/0008-5472.CAN-14-2458
- Peske JD, Woods AB, Engelhard VH. Control of CD8 T-cell infiltration into tumors by vasculature and microenvironment. *Adv Cancer Res.* (2015) 128:263–307. doi: 10.1016/bs.acr.2015.05.001
- Lanitis E, Irving M, Coukos G. Targeting the tumor vasculature to enhance T cell activity. *Curr Opin Immunol.* (2015) 33:55–63. doi: 10.1016/j.coi.2015.01.011
- Bouzin C, Brouet A, De Vriese J, Dewever J, Feron O. Effects of vascular endothelial growth factor on the lymphocyte-endothelium interactions: identification of caveolin-1 and nitric oxide as control points of endothelial cell anergy. J Immunol. (2007) 178:1505–11. doi: 10.4049/jimmunol.178.3.1505
- Buckanovich RJ, Facciabene A, Kim S, Benencia F, Sasaroli D, Balint K, et al. Endothelin B receptor mediates the endothelial barrier to T cell homing to tumors and disables immune therapy. *Nat Med.* (2008) 14:28–36. doi: 10.1038/nm1699
- Clark RA, Huang SJ, Murphy GF, Mollet IG, Hijnen D, Muthukuru M, et al. Human squamous cell carcinomas evade the immune response by downregulation of vascular E-selectin and recruitment of regulatory T cells. *J Exp Med.* (2008) 205:2221–34. doi: 10.1084/jem.20071190
- Griffioen AW, Damen CA, Martinotti S, Blijham GH, Groenewegen G. Endothelial intercellular adhesion molecule-1 expression is suppressed in human malignancies: the role of angiogenic factors. *Cancer Res.* (1996) 56:1111–17.
- Piali L, Fichtel A, Terpe HJ, Imhof BA, Gisler RH. Endothelial vascular cell adhesion molecule 1 expression is suppressed by melanoma and carcinoma. *J Exp Med.* (1995) 181:811–6. doi: 10.1084/jem.181.2.811

- Madhavan M, Srinivas P, Abraham E, Ahmed I, Vijayalekshmi NR, Balaram P. Down regulation of endothelial adhesion molecules in node positive breast cancer: possible failure of host defence mechanism. *Pathol Oncol Res.* (2002) 8:125–8. doi: 10.1007/BF03033721
- Pober JS, Sessa WC. Evolving functions of endothelial cells in inflammation. Nat Rev Immunol. (2007) 7:803–15. doi: 10.1038/nri2171
- Griffioen AW, Damen CA, Blijham GH, Groenewegen G. Tumor angiogenesis is accompanied by a decreased inflammatory response of tumor-associated endothelium. *Blood* (1996) 88:667–73.
- Huang H, Langenkamp E, Georganaki M, Loskog A, Fuchs PF, Dieterich LC, et al. VEGF suppresses T-lymphocyte infiltration in the tumor microenvironment through inhibition of NF-kappaB-induced endothelial activation. *FASEB J.* (2015) 29:227–38. doi: 10.1096/fj.14-2 50985
- Cindrova-Davies T, Sanders DA, Burton GJ, Charnock-Jones DS. Soluble FLT1 sensitizes endothelial cells to inflammatory cytokines by antagonizing VEGF receptor-mediated signalling. *Cardiovasc Res.* (2011) 89:671–9. doi: 10.1093/cvr/cvq346
- Sainson RC, Johnston DA, Chu HC, Holderfield MT, Nakatsu MN, Crampton SP, et al. TNF primes endothelial cells for angiogenic sprouting by inducing a tip cell phenotype. *Blood* (2008) 111:4997–5007. doi: 10.1182/blood-2007-08-108597
- Melder RJ, Koenig GC, Witwer BP, Safabakhsh N, Munn LL, Jain RK. During angiogenesis, vascular endothelial growth factor and basic fibroblast growth factor regulate natural killer cell adhesion to tumor endothelium. *Nat Med.* (1996) 2:992–7. doi: 10.1038/nm0996-992
- Goebel S, Huang M, Davis WC, Jennings M, Siahaan TJ, Alexander JS, et al. VEGF-A stimulation of leukocyte adhesion to colonic microvascular endothelium: implications for inflammatory bowel disease. *Am J Physiol Gastrointest Liver Physiol.* (2006) 290:G648–54. doi: 10.1152/ajpgi.00466.2005
- Kim I, Moon SO, Kim SH, Kim HJ, Koh YS, Koh GY. Vascular endothelial growth factor expression of intercellular adhesion molecule 1 (ICAM-1), vascular cell adhesion molecule 1 (VCAM-1), and E-selectin through nuclear factor-kappa B activation in endothelial cells. *J Biol Chem.* (2001) 276:7614– 20. doi: 10.1074/jbc.M009705200
- Gehad AE, Lichtman MK, Schmults CD, Teague JE, Calarese AW, Jiang Y, et al. Nitric oxide-producing myeloid-derived suppressor cells inhibit vascular E-selectin expression in human squamous cell carcinomas. J Invest Dermatol. (2012) 132:2642–51. doi: 10.1038/jid.2012.190
- Delfortrie S, Pinte S, Mattot V, Samson C, Villain G, Caetano B, et al. Egfl7 promotes tumor escape from immunity by repressing endothelial cell activation. *Cancer Res.* (2011) 71:7176–86. doi: 10.1158/0008-5472.CAN-11-1301
- Mulligan JK, Lathers DM, Young MR. Tumors skew endothelial cells to disrupt NK cell, T-cell and macrophage functions. *Cancer Immunol Immunother*. (2008) 57:951–61. doi: 10.1007/s00262-007-0425-x
- 62. Motz GT, Santoro SP, Wang LP, Garrabrant T, Lastra RR, Hagemann IS, et al. Tumor endothelium FasL establishes a selective immune barrier promoting tolerance in tumors. *Nat Med.* (2014) 20:607–15. doi: 10.1038/nm.3541
- Mai J, Virtue A, Shen J, Wang H, Yang X-F. An evolving new paradigm: endothelial cells-conditional innate immune cells. *J Hematol Oncol.* (2013) 6:61. doi: 10.1186/1756-8722-6-61
- Griffioen AW, Damen CA, Mayo KH, Barendsz-Janson AF, Martinotti S, Blijham GH, et al. Angiogenesis inhibitors overcome tumor induced endothelial cell anergy. *Int J Cancer* (1999) 80:315–9.
- 65. Dirkx AEM, oude Egbrink MGA, Castermans K, van der Schaft DWJ, Thijssen VLJL, Dings RPM, et al. Anti-angiogenesis therapy can overcome endothelial cell anergy and promote leukocyte-endothelium interactions and infiltration in tumors. *FASEB J.* (2006) 20:621–30. doi: 10.1096/fj.05-4493com
- 66. Dings RPM, Vang KB, Castermans K, Popescu F, Zhang Y, Oude Egbrink MGA, et al. Enhancement of T-cell-mediated antitumor response: angiostatic adjuvant to immunotherapy against cancer. *Clin Cancer Res.* (2011) 17:3134–45. doi: 10.1158/1078-0432.CCR-1 0-2443
- 67. van Hooren L, Georganaki M, Huang H, Mangsbo SM, Dimberg A. Sunitinib enhances the antitumor responses of agonistic CD40-antibody by reducing

MDSCs and synergistically improving endothelial activation and T-cell recruitment. *Oncotarget* (2016) 7:50277–89. doi: 10.18632/oncotarget.10364

- Folkman J. Tumor angiogenesis: therapeutic implications. New Eng J Med. (1971) 285:1182–6. doi: 10.1056/NEJM197111182852108
- Jain RK. Antiangiogenesis strategies revisited: from starving tumors to alleviating hypoxia. *Cancer Cell.* (2014) 26:605–22. doi: 10.1016/j.ccell.2014.10.006
- Manegold C. Bevacizumab for the treatment of advanced nonsmall-cell lung cancer. *Expert Rev Anticancer Ther.* (2008) 8:689–99. doi: 10.1586/14737140.8.5.689
- Arkenau HT, Brunetto AT, Barriuso J, Olmos D, Eaton D, de Bono J, et al. Clinical benefit of new targeted agents in phase I trials in patients with advanced colorectal cancer. *Oncology* (2009) 76:151–6. doi: 10.1159/000195884
- Calmes-Miller J. FDA approves first biosimilar to treat cancer. *Cancer Discov.* (2017) 7:1206. doi: 10.1158/2159-8290.CD-NB2017-183
- Vasudev NS, Reynolds AR. Anti-angiogenic therapy for cancer: current progress, unresolved questions and future directions. *Angiogenesis* (2014) 17:471–94. doi: 10.1007/s10456-014-9420-y
- 74. Ruegg C, Mutter N. Anti-angiogenic therapies in cancer: achievements and open questions. *Bull Cancer.* (2007) 94:753–62. doi: 10.1684/bdc.2007.0451
- Paez-Ribes M, Allen E, Hudock J, Takeda T, Okuyama H, Vinals F, et al. Antiangiogenic therapy elicits malignant progression of tumors to increased local invasion and distant metastasis. *Cancer Cell* (2009) 15:220– 31. doi: 10.1016/j.ccr.2009.01.027
- Ebos JM, Lee CR, Cruz-Munoz W, Bjarnason GA, Christensen JG, Kerbel RS. Accelerated metastasis after short-term treatment with a potent inhibitor of tumor angiogenesis. *Cancer Cell* (2009) 15:232–9. doi: 10.1016/j.ccr.2009.01.021
- De Bock K, Mazzone M, Carmeliet P. Antiangiogenic therapy, hypoxia, and metastasis: risky liaisons, or not? *Nat Rev Clin Oncol.* (2011) 8:393–404. doi: 10.1038/nrclinonc.2011.83
- Jain RK. Normalization of tumor vasculature: an emerging concept in antiangiogenic therapy. *Science* (2005) 307:58–62. doi: 10.1126/science.1104819
- Shrimali RK, Yu Z, Theoret MR, Chinnasamy D, Restifo NP, Rosenberg SA. Antiangiogenic agents can increase lymphocyte infiltration into tumor and enhance the effectiveness of adoptive immunotherapy of cancer. *Cancer Res.* (2010) 70:6171–80. doi: 10.1158/0008-5472.CAN-10-0153
- Huang Y, Yuan J, Righi E, Kamoun WS, Ancukiewicz M, Nezivar J, et al. Vascular normalizing doses of antiangiogenic treatment reprogram the immunosuppressive tumor microenvironment and enhance immunotherapy. *Proc Natl Acad Sci USA*. (2012) 109:17561–6. doi: 10.1073/pnas.1215397109
- Meder L, Schuldt P, Thelen M, Schmitt A, Dietlein F, Klein S, et al. Combined VEGF and PD-L1 blockade displays synergistic treatment effects in an autochthonous mouse model of small cell lung cancer. *Cancer Res.* (2018) 78:4270–81. doi: 10.1158/0008-5472.CAN-17-2176
- 82. Yasuda S, Sho M, Yamato I, Yoshiji H, Wakatsuki K, Nishiwada S, et al. Simultaneous blockade of programmed death 1 and vascular endothelial growth factor receptor 2 (VEGFR2) induces synergistic anti-tumour effect *in vivo*. *Clin Exp Immunol*. (2013) 172:500–6. doi: 10.1111/cei.12069
- Schmittnaegel M, Rigamonti N, Kadioglu E, Cassara A, Wyser Rmili C, Kiialainen A, et al. Dual angiopoietin-2 and VEGFA inhibition elicits antitumor immunity that is enhanced by PD-1 checkpoint blockade. *Sci Transl Med.* (2017) 9:eaak9670. doi: 10.1126/scitranslmed.aak9670
- 84. Du Four S, Maenhout SK, Niclou SP, Thielemans K, Neyns B, Aerts JL. Combined VEGFR and CTLA-4 blockade increases the antigen-presenting function of intratumoral DCs and reduces the suppressive capacity of intratumoral MDSCs. *Am J Cancer Res.* (2016) 6:2514–31.
- Voron T, Colussi O, Marcheteau E, Pernot S, Nizard M, Pointet AL, et al. VEGF-A modulates expression of inhibitory checkpoints on CD8+ T cells in tumors. J Exp Med. (2015) 212:139–48. doi: 10.1084/jem.201 40559
- Allen E, Jabouille A, Rivera LB, Lodewijckx I, Missiaen R, Steri V, et al. Combined antiangiogenic and anti-PD-L1 therapy stimulates tumor immunity through HEV formation. *Sci Transl Med.* (2017) 9:aak9679. doi: 10.1126/scitranslmed.aak9679

- Wu FT, Man S, Xu P, Chow A, Paez-Ribes M, Lee CR, et al. Efficacy of Cotargeting Angiopoietin-2 and the VEGF pathway in the adjuvant postsurgical setting for early breast, colorectal, and renal cancers. *Cancer Res.* (2016) 76:6988–7000. doi: 10.1158/0008-5472.CAN-16-0888
- Rexer H, Bedke J. [First-line therapy in advanced renal cell carcinoma : A randomized, open-label phase III study evaluating the efficacy and safety of pembrolizumab (MK-3475) in combination with axitinib compared to sunitinib monotherapy as first-line treatment for locally advanced or metastatic renal cell carcinoma (mRCC) (Keynote-426) - AN 39/16 of the AUO]. Urologe A. (2017) 56:385–6. doi: 10.1007/s00120-017-0335-2
- Hodi FS, Lawrence D, Lezcano C, Wu X, Zhou J, Sasada T, et al. Bevacizumab plus ipilimumab in patients with metastatic melanoma. *Cancer Immunol Res.* (2014) 2:632–42. doi: 10.1158/2326-6066.CIR-14-0053
- Wu X, Giobbie-Hurder A, Liao X, Lawrence D, McDermott D, Zhou J, et al. VEGF neutralization plus CTLA-4 blockade alters soluble and cellular factors associated with enhancing lymphocyte infiltration and humoral recognition in melanoma. *Cancer Immunol Res.* (2016) 4:858–68. doi: 10.1158/2326-6066.CIR-16-0084
- Wu X, Li J, Connolly EM, Liao X, Ouyang J, Giobbie-Hurder A, et al. Combined Anti-VEGF and Anti-CTLA-4 therapy elicits humoral immunity to galectin-1 which is associated with favorable clinical outcomes. *Cancer Immunol Res.* (2017) 5:446–54. doi: 10.1158/2326-6066.CIR-16-0385
- 92. Choueiri TK, Larkin J, Oya M, Thistlethwaite F, Martignoni M, Nathan P, et al. Preliminary results for avelumab plus axitinib as first-line therapy in patients with advanced clear-cell renal-cell carcinoma (JAVELIN Renal 100): an open-label, dose-finding and dose-expansion, phase 1b trial. *Lancet Oncol.* (2018) 19:451–60. doi: 10.1016/S1470-2045(18)30107-4
- Iyer PC, Dadu R, Gule-Monroe M, Busaidy NL, Ferrarotto R, Habra MA, et al. Salvage pembrolizumab added to kinase inhibitor therapy for the treatment of anaplastic thyroid carcinoma. *J Immunother Cancer* (2018) 6:68. doi: 10.1186/s40425-018-0378-y
- 94. Wallin JJ, Bendell JC, Funke R, Sznol M, Korski K, Jones S, et al. Atezolizumab in combination with bevacizumab enhances antigen-specific T-cell migration in metastatic renal cell carcinoma. *Nat Commun.* (2016) 7:12624. doi: 10.1038/ncomms12624
- Fukumura D, Kloepper J, Amoozgar Z, Duda DG, Jain RK. Enhancing cancer immunotherapy using antiangiogenics: opportunities and challenges. *Nat Rev Clin Oncol.* (2018) 15:325–40. doi: 10.1038/nrclinonc.2018.29

- Finke JH, Rini B, Ireland J, Rayman P, Richmond A, Golshayan A, et al. Sunitinib reverses type-1 immune suppression and decreases T-regulatory cells in renal cell carcinoma patients. *Clin Cancer Res.* (2008) 14:6674–82. doi: 10.1158/1078-0432.CCR-07-5212
- 97. Ko JS, Zea AH, Rini BI, Ireland JL, Elson P, Cohen P, et al. Sunitinib mediates reversal of myeloid-derived suppressor cell accumulation in renal cell carcinoma patients. *Clin Cancer Res.* (2009) 15:2148–57. doi: 10.1158/1078-0432.CCR-08-1332
- Ott PA, Hodi FS, Buchbinder EI. Inhibition of immune checkpoints and vascular endothelial growth factor as combination therapy for metastatic melanoma: an overview of rationale, preclinical evidence, and initial clinical data. *Front Oncol.* (2015) 5:202. doi: 10.3389/fonc.2015.00202
- Comunanza V, Bussolino F. Therapy for cancer: strategy of combining anti-angiogenic and target therapies. *Front Cell Dev Biol.* (2017) 5:101. doi: 10.3389/fcell.2017.00101
- Ager A. High endothelial venules and other blood vessels: critical regulators of lymphoid organ development and function. *Front Immunol.* (2017) 8:45. doi: 10.3389/fimmu.2017.00045
- 101. Johansson-Percival A, He B, Li ZJ, Kjellen A, Russell K, Li J, et al. De novo induction of intratumoral lymphoid structures and vessel normalization enhances immunotherapy in resistant tumors. Nat Immunol. (2017) 18:1207–17. doi: 10.1038/ni.3836
- 102. Sautes-Fridman C, Lawand M, Giraldo NA, Kaplon H, Germain C, Fridman WH, et al. Tertiary lymphoid structures in cancers: prognostic value, regulation, and manipulation for therapeutic intervention. *Front Immunol.* (2016) 7:407. doi: 10.3389/fimmu.2016.00407

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Georganaki, van Hooren and Dimberg. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.