



Th17 and Th1 Lymphocytes in Oligoarticular Juvenile Idiopathic Arthritis

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In the last years much attention has focused on the Th17 and Th1 phenotypes and on their pathogenic role in juvenile idiopathic arthritis, investigating how the cytokines produced by T helper cells act on resident cells on the synovia and which signal transduction pathways regulate Th17 cells proliferation and plasticity. In this context, an important milestone was represented by the identification of the non-classic Th1 phenotype, developed from the shift of Th17 cells. The cytokine TNF- α , beyond its well-known proinflammatory activity is involved in this process and this is one of the reasons why the TNF- α inhibitors are widely used in the treatment of juvenile idiopathic arthritis patients.

Keywords: Th17, Th1, CD161+ T cells, TNF- α , synoviocytes

INTRODUCTION

Juvenile idiopathic arthritis (JIA) is one of the most common chronic conditions of childhood, comprising several forms of arthritis characterized by persistent joint inflammation for at least 6 weeks, with an onset before the age of 16 years and with unknown cause (1, 2). The term JIA covers seven pathologic conditions that differ for clinical presentation, disease course and treatment response; in particular it includes systemic arthritis (sJIA), oligoarthritis, polyarthritis (both rheumatoid factor positive or negative), psoriatic arthritis (JPsA), enthesitis-related arthritis (ERA), and undifferentiated arthritis (1, 2). Although the cause of disease is unknown, immune cells, including T and B lymphocytes, infiltrate the synovial membrane of inflamed joints, suggesting that the adaptive immune system is involved in the pathogenesis of JIA (3). Human effector CD4+ T lymphocytes can be classified in three main subsets based mainly on their immunological functions, their cytokines production profile and their typical transcription factor expression (4). Th1 lymphocytes express the transcription factor T-bet, produce interferon (IFN)- γ , and defend the body from intracellular infections. Th2 cells express the transcription factor GATA-3, produce type 2 cytokines (interleukin (IL)-4, IL-5, IL-9, and IL-13) and are important to protect against helminths (5, 6). Finally, the Th17 subset produce IL-17A, IL-17F and IL-22 (7–10), express the transcription factor ROR- γ T (11–13) and the lectin receptor CD161 (14), as typical surface marker. Beyond their protective role against extracellular bacterial and fungal infections, Th17 cells have been demonstrated to be important in the pathogenesis of several autoimmune and inflammatory diseases, including multiple sclerosis, inflammatory bowel disease (IBD), psoriasis, rheumatoid arthritis (RA), and JIA (15, 16). In humans, Th17 lymphocytes are included within the CD161+ cell fraction of circulating and tissue-infiltrating CD4+ T cells, and they develop from a CD161+ T cell precursor found in umbilical cord blood and neonatal thymus (14, 17, 18).

Since JIA can be considered as an immune-mediated disorder, the pharmacologic therapy is essentially based on immunosuppressive drugs, at least when the usage of non-steroidal anti-inflammatory drugs (NSAIDs) does not control symptoms. Among these disease-modifying antirheumatic drugs (DMARDs) methotrexate (MTX), is considered the first line treatment, since its positive clinical effects are associated with low toxic effects (19). For those patients with a suboptimal response to non-biologic DMARDs, in particular those with polyarthritis, the usage of biologic drugs such as tumor necrosis factor- α (TNF- α) inhibitors (etanercept, infliximab and adalimumab), IL-1 inhibitors (anakinra, canakinumab, and rilonacept), IL-6 inhibitor (tocilizumab), CD20/B-cell targeted (rituximab) and T-cell co-stimulatory signal blocker (abatacept), has been proven to be effective (19–22).

T HELPER EFFECTOR CELLS IN OLIGOARTICULAR JIA

The synovial membrane of JIA inflamed joints shows high degree of infiltrating mononuclear cells, including T and B lymphocytes, dendritic cells and macrophages (3, 18, 23). Among T cells, Th1 are the most represented since these cells can migrate in the synovia in response to the chemokine CXCL10 (24). For this reason, these cells were thought to play a key role in the pathogenesis of oligoarticular JIA (25, 26), at least until 10 years ago. More recently, after the identification of the Th17 subset, many experimental data suggested their potential pathogenic role both in adult and childhood arthritis as well as in other inflammatory and autoimmune diseases (27, 28). Indeed, increased levels of IL-17A and of the transcription factor ROR- γ T, as well as of Th17 cells were reported in the synovial fluid (SF) of oligoarticular JIA patients (26, 28–31).

Moreover, Th17 cells have been demonstrated to be pathogenic in several murine models of chronic inflammatory disorders (16), such as experimental autoimmune encephalomyelitis (7), collagen-induced arthritis (32), and IBD (10–12).

However, despite their supposed pathogenicity, Th17 are very rare at inflammatory sites if compared to Th1 cells (28, 33). A first explanation for this rarity is a self-regulatory mechanism that controls Th17 cells clonal expansion. In particular, ROR- γ T favors the up-regulation of the interleukin (IL)-4 induced gene 1 (IL4I1), which encodes an l-phenylalanine oxidase that down-regulates CD3 ϵ expression on T cells via the production of H₂O₂ (28, 34). By this way, Th17 cells display an impaired signaling pathway downstream of the T-cell receptor (TCR), leading to inappropriate proliferation and reduced IL-2 production upon TCR triggering (28, 34). In addition, high IL4I1 expression in Th17 cells induces up-regulation of Tob1, a member of the Tob/BTG anti-proliferative protein family, involved in the negative control of the cell cycle (35).

It has also been recently described that Th17 cells show reduced IL-2 responsiveness since they express Muscadin (MSC), a member of the basic helix-loop-helix transcription factors, dependent by ROR- γ T, which negatively regulates the

phosphorylation level of STAT5B upon IL-2 signaling (36). In agreement with these findings, both IL4I1 and MSC were found to be selectively expressed by CD161+ T cells obtained from SF of oligoarticular JIA inflamed joints (34, 36).

It has also been demonstrated that the development of JIA and of other autoimmune diseases depends not only on the amount and the phenotype of Th effector cells, but also on their balance with Treg cells (37). Indeed, several papers demonstrated an accumulation of Th17 and Treg cells in SF and PB of JIA patients (31, 38, 39), in particular in active versus inactive JIA (31), hypothesizing that the joint inflammatory status persists despite the high frequency of Treg cells because Th17 cells show a reduced susceptibility to their regulatory function. This could be also related to the low proliferation rate of Th17 cells (33–36).

ROLE OF Th17 PLASTICITY IN THE PATHOGENESIS OF OLIGOARTICULAR JIA

A second explanation for human Th17 cells rarity at inflamed tissues is their phenotype plasticity (28). Indeed, in presence of local inflammatory cytokines such as IL-12 and TNF- α , Th17 cells acquire the ability to produce IFN- γ . At a first stage, cells display an intermediate phenotype known as Th17/Th1 and produce both IFN- γ and IL-17, but they can also rapidly loose IL-17 secretion and become IFN- γ single producers (12, 18, 40). These Th17-derived Th1 cells are defined as non-classic Th1 cells because, differently from classic Th1 cells, they maintain the expression of ROR- γ T, CD161, and CCR6 (17, 18, 28, 40), typical molecules of the Th17 subset (**Figure 1**).

Th17/Th1 and non-classic Th1 cells were found to be enriched in the SF of oligoarticular JIA children compared to their peripheral blood (18, 40). Moreover, a positive correlation between the frequencies of CD4+CD161+ Th17/Th1 cells in the SF of inflamed joints and disease activity parameters was described (18, 28). The shift of Th17 cells toward the non-classic Th1 phenotype is driven by IL-12, that has been found to be elevated in the SF of oligoarticular JIA patients (18, 40). Moreover, the finding that SF-derived Th17 clones share similar TCR V β spectra with Th1 CD161+ clones but not with Th1 CD161- ones (18, 40), strongly supported the data of the shift of Th17 toward non-classic Th1 cells. These findings are in agreement with several studies describing an accumulation of Th1 cells in the SF of JIA patients (25, 26), characterized as Th cells producing IFN- γ but without any distinction between the classic and non-classic phenotypes. Additional data at epigenetic level confirm that non-classic Th1 cells originate from Th17 (41); indeed it was observed that non-classic Th1 cells exhibit demethylation of RORC2 and IL17A genes, as reported for Th17 cells, whereas classic Th1 cells are completely methylated at these loci (41).

Th17 plasticity consists not only in the acquisition of the ability to produce IFN- γ , but also GM-CSF (42). In fact, it has been described an enrichment of GM-CSF producing T cells with a non-classic Th1 phenotype in the SF of oligoarticular JIA patients (42) and induced *in vitro* by IL-12. This data suggests a possible involvement not only of IFN- γ but also

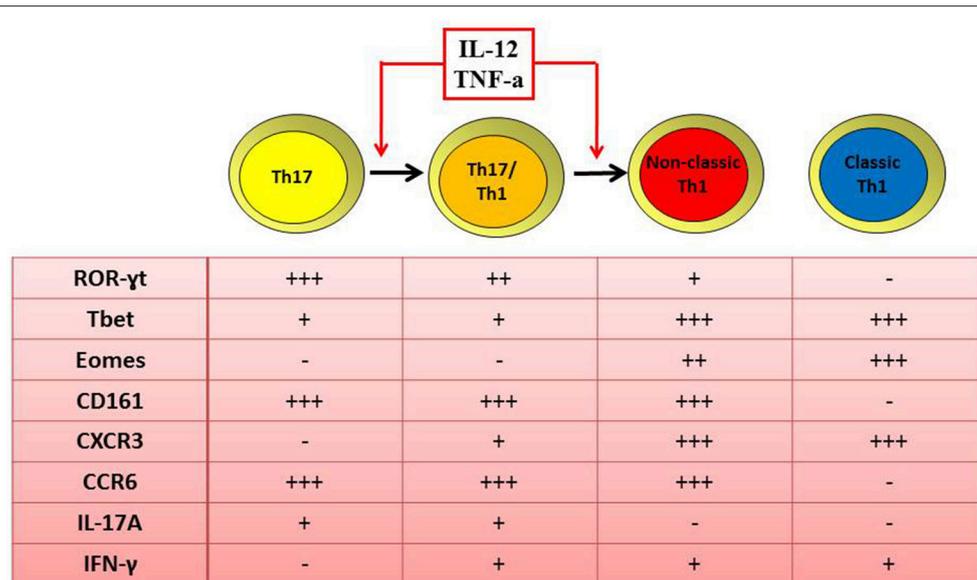


FIGURE 1 | Similar and distinct features of human Th1 and Th17 cells. Both classic and non-classic Th1 cells produce IFN- γ , express the transcription factor Tbet and Eomes and the surface marker CXCR3. Differently from classic, non-classic Th1 cells maintain also the expression of ROR- γ t, CD161, and CCR6 as typical transcription factor and surface markers of the Th17 phenotype. In fact, non-classic Th1 cells originate from the shift of Th17 toward the Th17/Th1 phenotype when in the presence of IL-12 and/or TNF- α .

of GM-CSF in JIA pathogenesis, and, accordingly, a positive correlation between GM-CSF protein levels in the SF and the serum parameters of disease activity was described (42).

Moreover it has been recently described that human non-classic Th1 cells development is promoted by the transcription factors Eomes (43), which induces and reinforces IFN- γ production, maintains the Th1 phenotype stability by inhibiting and preventing the re-expression of ROR- γ T and IL-17A and promotes GM-CSF secretion (43). Finally, it was shown that Eomes induces, *in vitro*, the production of IFN- γ and GM-CSF by human CD4+ T cells and that cells with this cytokines profile were enriched in the SF of inflamed joints of children with oligoarticular JIA (43). All these data further support the pathogenic role of Th17-derived non-classic Th1 cells.

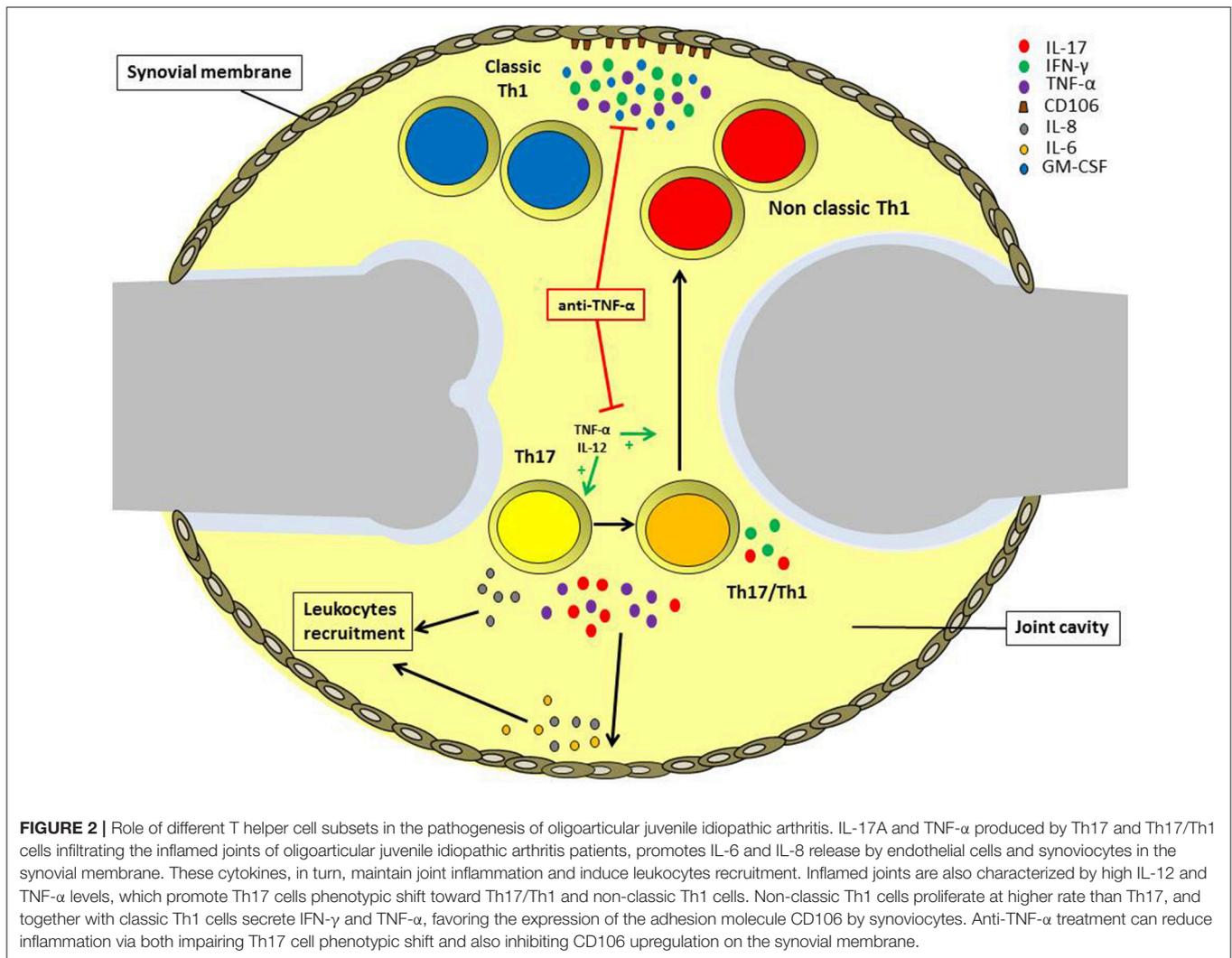
Additional evidence of the pathogenic features of CD4+CD161+ T cells, including pure Th17 cells and their derivative phenotypes Th17/Th1 and non-classic Th1, derived from a recent paper (44), describing that all these subsets express CHI3L1. This chitinase-like protein without enzymatic activity is defined in the literature as a well-known marker of disease activity and inflammation in several immune-mediated disorders (44–47): its levels are elevated in SF of children affected by oligoarticular JIA and positively correlated to inflammatory parameters (44).

These data of Th17 plasticity, partially solve the literature debate on the pathogenic or protective role of Th17 cells in immunomediated disorders (48, 49), supporting the hypothesis that, at least in JIA, the Th1 subset, both classic and non-classic, is directly involved in the active phase of the disease when the clinical manifestations are evident, and that the pure Th17 cells acquire a pathogenic feature when they start to produce IFN- γ

shifting toward the Th17/Th1 and the non-classic Th1 phenotype (18, 20, 40). Anyway, it cannot be excluded that even pure Th17 cells may play a key role in the pathogenesis of JIA during the onset and/or the early-phase of the disease.

CROSS TALK BETWEEN T HELPER CELLS AND RESIDENT SYNOVIAL CELLS

CD4+ T helper lymphocytes orchestrate both RA and JIA chronic inflammation producing cytokines that initiate and maintain the process of synovial proteolysis and proliferation as well as the angiogenesis related to the inflammatory status (50, 51). Synovial fibroblasts (SFbs) are the main tissue resident cell population in the synovia and it has been demonstrated that in adult RA SFbs produce cytokines and matrix-degrading enzymes, crucial to promote cartilage destruction and to mediate inflammation (28, 52). In particular, it has been reported (51) that SFbs derived from SF of oligoarticular JIA patients express high levels of CD106 (VCAM), a sialoglycoprotein which mediates leukocyte-endothelial cell adhesion and signal transduction (51), and whose upregulation is critical to favor leukocytes retention in the inflamed synovia (51). SFbs also showed a peculiar morphology consisting in polygonal cell body, large, and oval-shaped nucleus, many slender protrusions and branches extended out of the cell body (51, 53). This peculiar phenotype of oligoarticular JIA-derived SFbs, was resembled *in vitro* stimulating healthy-derived SFbs with culture supernatants from activated classic and non-classic Th1, but not from Th17, lymphocytes. Indeed, also in these experimental conditions SFbs upregulated CD106 expression and underwent morphological



changes (50). It has been demonstrated that TNF- α is the main cytokine involved in this process and that IFN- γ exerts a synergic effect (51, 54). The concept that cytokines produced by T cells play an important role on the activation of SFBs has been confirmed also by the paper of Lavocat et al. (55). It demonstrates with *in vitro* experiments that IL-17A and TNF- α alone are able to induce the expression of IL-6 and IL-8 (55) by both endothelial cells and synoviocytes (even if with different kinetics on each cell type), and that a synergistic effect can be achieved from the use of both cytokines (55). Similar results were obtained also by stimulating endothelial cells and synoviocytes in the presence of culture supernatants from activated T cell clones or recombinant cytokines. Indeed, the main increase in IL-6 and IL-8 production was observed when cells were cultured in presence of supernatants from Th17/Th1 T cell clones that contained both IL-17A and TNF- α (55). The early expression of IL-8 in inflamed joints, directly produced also by Th17 cells itself (9), might explain the massive neutrophil recruitment in the acute phase (56). On the other hand, IL-6 production

might be important to sustain the pro-inflammatory process since it is involved in the differentiation and expansion of Th17 cell (57), in VEGF production [thus mediating angiogenesis (58)], as well as in antibody production (59) and in osteoclast activation (55).

The IL-17 signature, which is typical of JIA, is important also for bone and cartilage erosion. In fact, it has been demonstrated that IL-17A acts on SFBs increasing the expression of different types of matrix metalloproteinases, MMP-1, MMP-3 (60). Finally, it is important to note that IL-17A production is not strictly associated to Th17 cells, since it is produced also by additional cells of the immune system enriched in SF of JIA patients, such as CD3⁺CD8⁺ and CD3⁺CD4⁻CD8⁻ T cells (17, 61, 62) and innate lymphoid cells (62). Collectively, these data suggest that mechanisms actively contributing to joint inflammation in the synovia of JIA patients depend on the final balance and cross-talk between tissue resident cells and immune cells from both the adaptive and innate immune systems.

EFFECTS OF BIOLOGICAL DRUGS IN THE TREATMENT OF JIA: *EX-VIVO* AND *IN-VITRO* OBSERVATIONS

Cytokines produced by immune cells (in particular T cells and monocytes) and by tissue resident cells in the synovia contribute to the development of JIA and are responsible for most of the clinical manifestations of the disease. In this view, pro-inflammatory cytokines represent a key therapeutic target for biological treatment. The drugs mainly used and effective in JIA inhibit the activity of TNF- α , IL-1, or IL-6. TNF- α has pleiotropic effects in the inflamed environment of affected joints, acting on different cell populations (51): TNF- α mediates monocyte, macrophage and SFb activation, and it is also responsible for inflammation induction, cartilage degradation, bone erosion and tissue damage (51). Moreover, as previously stated, TNF- α acts on SFbs inducing the upregulation of CD106, thus favoring leukocytes retention within the synovia and increasing joint inflammatory status (51). TNF- α is also involved in the neovascularization process, leading to synovial membrane growth, and in the process of osteoclast-containing 'pannus' formation (51). Additionally, TNF- α interferes with T helper cells phenotype plasticity, mediating the shifting of Th17 lymphocytes toward non-classic Th1 cells (20, 51). Nowadays, JIA patients are treated with non-steroidal antiinflammatory drugs, corticosteroids, and disease modifying antirheumatic drugs including TNF- α antagonists (28). Among these antagonists etanercept is a soluble dimeric fusion protein binding soluble TNF- α . Etanercept has been reported to induce improvement of clinical symptoms (as measured by radiological progression and laboratory parameters of disease activity) in patients affected by immune-mediated arthritis, including RA, JIA, and psoriatic arthritis (28, 63). Moreover, etanercept efficacy in JIA treatment has been demonstrated in randomized clinical trials, as well as in long-term observational registries (28, 64). These clinical effects were sustained by its well-known anti-inflammatory properties on the innate and adaptive immune responses. Moreover, it has been reported a new mechanism of action of etanercept, defining its inhibitory role in the plasticity of Th17 cells toward the non-classic Th1 phenotype mediated by TNF- α (18, 20, 28). In fact, etanercept reduces the proportion of circulating non-classic Th1 cells (20), supposed to play a key role in the pathogenesis of oligoarticular JIA and leads to an increased frequency of Th17 cells (20). Similar evidence was observed also during the treatment of RA patients with adalimumab, a fully humanized monoclonal IgG1 antibody against TNF- α (65), supporting again the important role of this cytokine in the pathogenesis of such diseases. In fact, after 12 weeks of treatment an increase was found in the frequency of IL-17A producing cells that significantly correlated with a reduction in joint inflammation (65, 66). These data support again the important role of this cytokine in the pathogenesis of such diseases. Moreover, it was demonstrated that etanercept acts also on the regulation of CD106 expression on SFbs, in fact *in vitro* administration of this drug negatively interferes with the ability of both classic and non-classic Th1 cells supernatants to significantly induce CD106 expression

on SFbs (51). Taking into account the important role of CD106 expression on SFbs to mediate leukocytes adhesion, these recent data define also the role of etanercept in interfering with the adhesion of immune cells on SFbs. Even if these data were obtained with *in-vitro* experimental models (51), they may suggest that the reduction of inflammatory cells in the synovia, occurring during etanercept treatment of oligoarticular JIA, may be driven by the reduced retention of immune cells within inflamed joints.

All these data define the immunomodulatory properties of TNF- α inhibitors, especially of etanercept and adalimumab, which could further explain its disease-modifying effect in JIA (20, 51, 66). Regarding infliximab (a human-mouse chimeric anti-TNF- α antibody) and golimumab (a fully humanized monoclonal anti-TNF- α antibody), their use is related mainly to polyarticular JIA and reported in case reports and open-label trials (22).

Among additional drugs inhibiting inflammatory cytokines, the IL-6 receptor antagonist (tocilizumab) and the IL-1 antagonists (anakinra, canakinumab and rilonacept), are currently used in systemic JIA. Anyway these drugs were also tested in clinical practice for oligoarticular or polyarticular JIA (22, 67), and their efficacy may be due, at least in part, to the interference with Th17 expansion and differentiation in the synovia mediated by IL-1 and IL-6.

Moreover, the use of anti-IL-12/IL-23 p40 inhibitors could improve the course of JIA, since both cytokines are involved in Th1 and Th17 differentiation and are important regulators of Th17 plasticity. Ustekinumab, the human monoclonal antibody anti- p40 subunit, is often used in the treatment of psoriatic arthritis and ankylosing spondylarthritis in adults and children (68) and its use in oligoarticular JIA is poor. These data suggest that although these cytokines contribute to joint inflammation, they may not be the principal factors responsible and it is likely that other key mediators are involved (i.e., TNF- α).

Similarly, the use of anti-IL-17A in the treatment of JIA has been explored. In fact data from clinical trials show that secukinumab, the high-affinity fully human monoclonal antibody neutralizing the activity of IL-17A, can be effective in the treatment of JPsA and ERA, suggesting that IL-17 cytokine and Th17 cells play a key role in the pathogenesis of these subtypes of JIA and their role is instead marginal in oligoarticular JIA (18, 20, 40). These data suggest that the use of the appropriate treatment and its effectiveness are related to the different biological conditions found in the different subtypes of JIA (69).

Finally, since oligoarticular JIA is mainly considered to arise due to a dysregulated adaptive immunity, involving Th1 and Th17 effector cells and Treg cells, abatacept was used for its treatment. Abatacept is a chimeric CTLA4 and IgG Fc fusion protein, that, binding to CD80/86 molecules instead of CD28, reduces T helper cells activation. In the treatment of oligoarticular JIA, it has been demonstrated that abatacept reduced proliferation of CD4+ T cells and their cytokines production (mainly IFN- γ and TNF- α) (21) and reduced the induction of Ig production by B cells (70).

CONCLUSION

In this review, we analyzed the role of different types of T helper cell subsets in the pathogenesis of JIA with particular attention to Th17 and Th1 phenotype (Figure 2). Both Th1 and Th17 cells are critical for the pathogenesis of the disease: Th1 lymphocytes through the production of pro-inflammatory cytokines IFN- γ and TNF- α ; Th17 lymphocytes thanks to the shift toward the non-classic Th1 phenotype. Th1 cells subsets exert their function through inducing the expression of CD106 on SFbs, which is crucial in mediating immune cells retention in inflamed synovia. Intriguingly the TNF- α inhibitors are the main biological drugs used in JIA and interfere both with the shift of Th17 to Th1 cells

and the TNF- α mediated CD106 upregulation on SFbs. All these data give an explanation at both cellular and molecular level for the efficacy of etanercept treatment in JIA and represent the beginning for further investigation with the aim to identify more specific therapeutic targets. In this view, it is important to underline that different subtypes of JIA are characterized by different inflammatory conditions, whose characterization is crucial for the choice of the efficacious biological treatment.

AUTHOR CONTRIBUTIONS

LM and LC wrote the paper. LM and AM prepared the figure. RC, FL, FA, and LC revised the manuscript.

REFERENCES

- Petty RE, Southwood TR, Manners P, Baum J, Glass DN, Goldenberg J, et al. International league of associations for rheumatology classification of juvenile idiopathic arthritis: second revision, Edmonton, 2001. *J Rheumatol.* (2004) 31:390–2.
- Ferrara G, Mastrangelo G, Barone P, La Torre F, Martino S, Pappagallo G, et al. Methotrexate in juvenile idiopathic arthritis: advice and recommendations from the MARAJIA expert consensus meeting. *Pediatr Rheumatol Online J.* (2018) 16:46. doi: 10.1186/s12969-018-0255-8
- Murray KJ, Luyrink L, Grom AA, Passo MH, Emery H, Witte D, et al. Immunohistological characteristics of T cell infiltrates in different forms of childhood onset chronic arthritis. *J Rheumatol.* (1996) 23:2116–24.
- Annunziato F, Cosmi L, Liotta F, Maggi E, Romagnani S. Defining the human T helper 17 cell phenotype. *Trends Immunol.* (2012) 33:505–12. doi: 10.1016/j.it.2012.05.004
- Mosmann TR, Coffman RL. TH1 and TH2 cells: different patterns of lymphokine secretion lead to different functional properties. *Annu Rev Immunol.* (1989) 7:145–73. doi: 10.1146/annurev.iy.07.040189.001045
- Romagnani S. Lymphokine production by human T cells in disease states. *Annu Rev Immunol.* (1994) 12:227–57. doi: 10.1146/annurev.iy.12.040194.001303
- Cua DJ, Sherlock J, Chen Y, Murphy CA, Joyce B, Seymour B, et al. Interleukin-23 rather than interleukin-12 is the critical cytokine for autoimmune inflammation of the brain. *Nature.* (2003) 421:744–8. doi: 10.1038/nature01355
- Oppmann B, Lesley R, Blom B, Timans JC, Xu Y, Hunte B, et al. Novel p19 protein engages IL-12p40 to form a cytokine, IL-23, with biological activities similar as well as distinct from IL-12. *Immunity.* (2000) 13:715–25. doi: 10.1016/S1074-7613(00)00070-4
- Pelletier M, Maggi L, Micheletti A, Lazzari E, Tamassia N, Costantini C, et al. Evidence for a cross-talk between human neutrophils and Th17 cells. *Blood.* (2010) 115:335–43. doi: 10.1182/blood-2009-04-216085
- Ouyang W, Koli JK, Zheng Y. The biological functions of T helper 17 cell effector cytokines in inflammation. *Immunity.* (2008) 28:454–67. doi: 10.1016/j.immuni.2008.03.004
- Ivanov II, McKenzie BS, Zhou L, Tadokoro CE, Lepelley A, Lafaille JJ, et al. The orphan nuclear receptor ROR γ t directs the differentiation program of proinflammatory IL-17+T helper cells. *Cell.* (2006) 126:1121–31. doi: 10.1016/j.cell.2006.07.035
- Annunziato F, Cosmi L, Santarlasci V, Maggi L, Liotta F, Mazzinghi B, et al. Phenotypic and functional features of human Th17 cells. *J Exp Med.* (2007) 204:1849–61. doi: 10.1084/jem.20070663
- Acosta-Rodriguez EV, Rivino L, Geginat J, Jarrossay D, Gattorno M, Lanzavecchia A, et al. Surface phenotype and antigenic specificity of human interleukin 17-producing T helper memory cells. *Nat Immunol.* (2007) 8:639–46. doi: 10.1038/ni1467
- Cosmi L, De Palma R, Santarlasci V, Maggi L, Capone M, Frosali F, et al. Human interleukin-17-producing cells originate from a CD161+ CD4+ T-cell precursor. *J Exp Med.* (2008) 205:1903–16. doi: 10.1084/jem.20080397
- Fouser LA, Wright JF, Dunussi-Joannopoulos K, Collins M. Th17 cytokines and their emerging roles in inflammation and autoimmunity. *Immunol Rev.* (2008) 226:87–102. doi: 10.1111/j.1600-065X.2008.00712.x
- Cosmi L, Liotta F, Maggi E, Romagnani S, Annunziato F. Th17 and non-classic Th1 cells in chronic inflammatory disorders: two sides of the same coin. *Int Arch Allergy Immunol.* (2014) 164:171–7. doi: 10.1159/000363502
- Maggi L, Santarlasci V, Capone M, Rossi MC, Querci V, Mazzoni A, et al. Distinctive features of classic and nonclassic (Th17 derived) human Th1 cells. *Eur J Immunol.* (2012) 42:3180–8. doi: 10.1002/eji.201242648
- Cosmi L, Cimaz R, Maggi L, Santarlasci V, Capone M, Borriello F, et al. Evidence of the transient nature of the Th17 phenotype of CD4+CD161+ T cells in the synovial fluid of patients with juvenile idiopathic arthritis. *Arthritis Rheum.* (2011) 63:2504–15. doi: 10.1002/art.30332
- Ungar WJ, Costa V, Burnett HF, Feldman BM, Laxer RM. The use of biologic response modifiers in polyarticular-course juvenile idiopathic arthritis: a systematic review. *Semin Arthritis Rheum.* (2013) 42:597–618. doi: 10.1016/j.semarthrit.2012.10.006
- Maggi L, Cimaz R, Capone M, Santarlasci V, Querci V, Simonini G, et al. Brief report: etanercept inhibits the tumor necrosis factor α -driven shift of Th17 lymphocytes toward a nonclassic Th1 phenotype in juvenile idiopathic arthritis. *Arthritis Rheumatol.* (2014) 66:1372–7. doi: 10.1002/art.38355
- Maggi L, Cimaz R, Capone M, Santarlasci V, Rossi MC, Mazzoni A, et al. Immunosuppressive activity of abatacept on circulating T helper lymphocytes from juvenile idiopathic arthritis patients. *Int Arch Allergy Immunol.* (2016) 171:45–53. doi: 10.1159/000450948
- Vanoni F, Minoia F, Malattia C. Biologics in juvenile idiopathic arthritis: a narrative review. *Eur J Pediatr.* (2017) 176:1147–53. doi: 10.1007/s00431-017-2960-6
- Gregorio A, Gambini C, Gerloni V, Parafioriti A, Sormani MP, Gregorio S, et al. Lymphoid neogenesis in juvenile idiopathic arthritis correlates with ANA positivity and plasma cells infiltration. *Rheumatology.* (2007) 46:308–13. doi: 10.1093/rheumatology/ke1225
- Aggarwal A, Agarwal S, Misra R. Chemokine and chemokine receptor analysis reveals elevated interferon-inducible protein-10 (IP)-10/CXCL10 levels and increased number of CCR5+ and CXCR3+ CD4 T cells in synovial fluid of patients with enthesitis-related arthritis. *Clin Exp Immunol.* (2007) 148:515–9. doi: 10.1111/j.1365-2249.2007.03377.x
- De Jager W, Hoppenreijns EP, Wulfraat NM. Blood and synovial fluid cytokine signatures in patients with juvenile idiopathic arthritis: a cross-sectional study. *Ann Rheum Dis.* (2007) 66:589–98. doi: 10.1136/ard.2006.061853
- Mahendra A, Misra R, Aggarwal A. Th1 and Th17 Predominance in the enthesitis-related arthritis form of juvenile idiopathic arthritis. *J Rheumatol.* (2009) 36:1730–6. doi: 10.3899/jrheum.081179

27. Singh RP, Hasan S, Sharma S, Nagra S, Yamaguchi DT, Wong DT, et al. Th17 cells in inflammation and autoimmunity. *Autoimmun Rev.* (2014)13:1174–81 doi: 10.1016/j.autrev.2014.08.019
28. Maggi L, Cosmi L, Simonini G, Annunziato F, Cimaz R. T cell subpopulations in juvenile idiopathic arthritis and their modifications after biotherapies. *Autoimmun Rev.* (2016) 15:1141–4. doi: 10.1016/j.autrev.2016.09.012
29. Nistala K, Moncrieffe H, Newton KR, Varsani H, Hunter P, Wedderburn LR. Interleukin-17-producing T cells are enriched in the joints of children with arthritis, but have a reciprocal relationship to regulatory T cell numbers. *Arthritis Rheum.* (2008) 58:875–87. doi: 10.1002/art.23291
30. Olivito B, Simonini G, Ciullini S, Moriondo M, Betti L, Gambineri E, et al. Th17 transcription factor RORC2 is inversely correlated with FOXP3 expression in the joints of children with juvenile idiopathic arthritis. *J Rheumatol.* (2009) 36:2017–24. doi: 10.3899/jrheum.090066
31. Wu SA, Yeh KW, Lee WI, Yao TC, Huang JL. Persistent improper upregulation of Th17 and TReg cells in patients with juvenile idiopathic arthritis. *J Microbiol Immunol Infect.* (2016) 49:402–8. doi: 10.1016/j.jmii.2014.07.002
32. Murphy CA, Langrish CL, Chen Y, Blumenschein W, McClanahan T, Kastelein RA, et al. Divergent pro- and anti-inflammatory roles for IL-23 and IL-12 in joint autoimmune inflammation. *J Exp Med.* (2003)198:1951–8. doi: 10.1084/jem.20030896
33. Annunziato F, Santarlasci V, Maggi L, Cosmi L, Liotta F, Romagnani S. Reasons for rarity of Th17 cells in inflammatory sites of human disorders. *Semin Immunol.* (2013) 25:299–304. doi: 10.1016/j.smim.2013.10.011
34. Santarlasci V, Maggi L, Capone M, Querci V, Beltrame L, Cavalieri D, et al. Rarity of human T helper 17 cells is due to retinoic acid orphan receptor-dependent mechanisms that limit their expansion. *Immunity.* (2012)36:201–14. doi: 10.1016/j.immuni.2011.12.013
35. Santarlasci V, Maggi L, Mazzoni A, Capone M, Querci V, Rossi MC, et al. IL-4-induced gene 1 maintains high Tob1 expression that contributes to TCR unresponsiveness in human T helper 17 cells. *Eur J Immunol.* (2014) 44:654–61. doi: 10.1002/eji.201344047
36. Santarlasci V, Mazzoni A, Capone M, Rossi MC, Maggi L, Montaini G, et al. Musculin inhibits human T helper 17 cells response to interleukin 2 by controlling STAT5B activity. *Eur J Immunol.* (2017) 47:1427–42. doi: 10.1002/eji.201746996
37. Nguyen LT, Jacobs J, Mathis D, Benoist C. Where FoxP3- dependent regulatory T cells impinge on the development of inflammatory arthritis. *Arthritis Rheum.* (2007)56:509–20. doi: 10.1002/art.22272
38. De Kleer IM, Wedderburn LR, Taams LS, Patel A, Varsani H, Klein M, et al. CD4^bCD25^{bright} regulatory T cells actively regulate inflammation in the joints of patients with the remitting form of juvenile idiopathic arthritis. *J Immunol.* (2004) 172:6435–43. doi: 10.4049/jimmunol.172.10.6435
39. Grose R, Millard D, Mavrangelos C, Barry S, Zola H, Nicholson IC, et al. Comparison of blood and synovial fluid Th17 and novel peptidase inhibitor 16 treg cell subsets in juvenile idiopathic arthritis. *J Rheumatol.* (2012)39:2021–31. doi: 10.3899/jrheum.111421
40. Nistala K, Adams S, Cambrook H, Ursu S, Olivito B, de Jager W, et al. Th17 plasticity in human autoimmune arthritis is driven by the inflammatory environment. *Proc Natl Acad Sci USA.* (2010) 107:14751–6. doi: 10.1073/pnas.1003852107
41. Mazzoni A, Santarlasci V, Maggi L, Capone M, Rossi MC, Querci V, et al. Demethylation of the RORC2 and IL17A in Human CD4⁺ T lymphocytes defines Th17 origin of nonclassical Th1 cells. *J Immunol.* (2015) 194:3116–26. doi: 10.4049/jimmunol.1401303
42. Piper C, Pesenacker AM, Bending D, Thirugnanabalan B, Varsani H, Wedderburn LR, et al. T Cell expression of granulocyte-macrophage colony-stimulating factor in juvenile arthritis is contingent upon Th17 plasticity. *Arthritis Rheumatol.* (2014) 66:1955–60. doi: 10.1002/art.38647
43. Mazzoni A, Maggi L, Siracusa F, Ramazzotti M, Rossi MC, Santarlasci V, et al. Eomes controls the development of Th17-derived (non-classic) Th1 cells during chronic inflammation. *Eur J Immunol.* (2019) 49:79–95. doi: 10.1002/eji.201847677
44. Capone M, Maggi L, Santarlasci V, Rossi MC, Mazzoni A, Montaini G, et al. Chitinase 3-like-1 is produced by human Th17 cells and correlates with the level of inflammation in juvenile idiopathic arthritis patients. *Clin Mol Allergy.* (2016) 14:16. doi: 10.1186/s12948-016-0053-0
45. Chupp GL, Lee CG, Jarjour N, Shim YM, Holm CT, He S, et al. A chitinase-like protein in the lung and circulation of patients with severe asthma. *N Engl J Med.* (2007) 357:2016–27. doi: 10.1056/NEJMoa073600
46. Vos K, Steenbakkers P, Miltenburg AM, Bos E, van Den Heuvel MW, van Hogeand RA, et al. Raised human cartilage glycoprotein-39 plasma levels in patients with rheumatoid arthritis and other inflammatory conditions. *Ann Rheum Dis.* (2000) 59:544–8. doi: 10.1136/ard.59.7.544
47. Baeten D, Steenbakkers PG, Rijnders AM, Boots AM, Veys EM, De Keyser F. Detection of major histocompatibility complex/human cartilage of MHC/HC gp-39 complexes in rheumatoid arthritis synovitis as a specific and independent histological marker. *Arthritis Rheum.* (2004)50:444–51. doi: 10.1002/art.20012
48. Marwaha AK, Leung NJ, McMurchy AN, Levings MK. TH17 cells in autoimmunity and immunodeficiency: protective or pathogenic? *Front Immunol.* (2012) 3:129. doi: 10.3389/fimmu.2012.00129
49. Zielinski CE, Mele F, Aschenbrenner D, Jarrossay D, Ronchi F, Gattorno M, et al. Pathogen-induced human TH17 cells produce IFN- γ or IL-10 and are regulated by IL-1 β . *Nature.* (2012) 484:514–8. doi: 10.1038/nature10957
50. Guiducci S, Del Rosso A, Cinelli M, Margheri F, D'Alessio S, Fibbi G, et al. Rheumatoid synovial fibroblasts constitutively express the fibrinolytic pattern of invasive tumor-like cells. *Clin Exp Rheumatol.* (2005)23:364–72.
51. Maggi L, Margheri F, Luciani C, Capone M, Rossi MC, Chilà A, et al. Th1-Induced CD106 expression mediates leukocytes adhesion on synovial fibroblasts from juvenile idiopathic arthritis patients. *PLoS ONE.* (2016) 11:e0154422. doi: 10.1371/journal.pone.0154422
52. Brescia AC, Simonds MM, McCahan SM, Fawcett PT, Rose CD. The role of transforming growth factor β signaling in fibroblast-like synoviocytes from patients with oligoarticular juvenile idiopathic arthritis: dysregulation of transforming growth factor β signaling, including overexpression of bone morphogenetic protein 4, may lead to a chondrocyte phenotype and may contribute to bony hypertrophy. *Arthritis Rheumatol.* (2014) 66:1352–62. doi: 10.1002/art.38336
53. Casnici C, Lattuada D, Tonna N, Crotta K, Storini C, Bianco F, et al. Optimized “in vitro” culture conditions for human rheumatoid arthritis synovial fibroblasts. *Mediators Inflamm.* (2014) 2014:702057. doi: 10.1155/2014/702057
54. Lombardi A, Cantini G, Mello T, Francalanci M, Gelmini S, Cosmi L, et al. Molecular mechanisms underlying the pro-inflammatory synergistic effect of tumor necrosis factor alpha and interferon gamma in human microvascular endothelium. *Eur J Cell Biol.* (2009) 88:731–42. doi: 10.1016/j.ejcb.2009.07.004
55. Lavocat F, Maggi L, Annunziato F, Miossec P. T-cell clones from Th1, Th17 or Th1/17 lineages and their signature cytokines have different capacity to activate endothelial cells or synoviocytes. *Cytokine.* (2016) 88:241–50. doi: 10.1016/j.cyto.2016.09.019
56. Griffin GK, Newton G, Tarrío ML, Bu DX, Maganto-García E, Azcutia V, et al. IL-17 and TNF α sustain neutrophil recruitment during inflammation through synergistic effects on endothelial activation. *J Immunol.* (2012) 188:6287–99. doi: 10.4049/jimmunol.1200385
57. Yang L, Anderson DE, Baecher-Allan C, Hastings WD, Bettelli E, Oukka M, et al. IL-21 and TGF- β are required for differentiation of human T(H)17 cells. *Nature.* (2008) 454:350–2. doi: 10.1038/nature07021
58. Pickens SR, Volin MV, Mandelin AM II, Kolls JK, Pope RM, Shahrara S. IL-17 contributes to angiogenesis in rheumatoid arthritis. *J Immunol.* (2010) 184:3233–41. doi: 10.4049/jimmunol.0903271
59. Gottenberg JE, Dayer JM, Lukas C, Ducot B, Chiochia G, Cantagrel A, et al. Serum IL-6 and IL-21 are associated with markers of B cell activation and structural progression in early rheumatoid arthritis: results from the ESPOIR cohort. *Ann Rheum Dis.* (2012) 71:1243–8. doi: 10.1136/annrheumdis-2011-200975
60. Agarwal S, Misra R, Aggarwal A. Interleukin 17 levels are increased in juvenile idiopathic arthritis synovial fluid and induce synovial fibroblasts to produce proinflammatory cytokines and matrix metalloproteinases. *J Rheumatol.* (2008) 35:515–9.
61. Omoyinmi E, Hamaoui R, Pesenacker A, Nistala K, Moncrieffe H, Ursu S, et al. Th1 and Th17 cell subpopulations are enriched in the peripheral blood of patients with systemic juvenile idiopathic arthritis. *Rheumatology.* (2012) 51:1881–6. doi: 10.1093/rheumatology/kes162

62. Rosser EC, Lom H, Bending D, Duurland CL, Bajaj-Elliott M, Wedderburn LR. Brief report: innate lymphoid cells and T-cells contribute to the IL-17A signature detected in the synovial fluid of patients with Juvenile Idiopathic Arthritis. *Arthritis Rheumatol.* (2019) 71:460–7. doi: 10.1002/art.40731
63. Marotte H, Cimaz R. Etanercept - TNF receptor and IgG1 Fc fusion protein: is it different from other TNF blockers? *Expert Opin Biol Ther.* (2014) 14:569–72. doi: 10.1517/14712598.2014.896334
64. Lovell DJ, Giannini EH, Reiff A, Cawkwell GD, Silverman ED, Nocton JJ, et al. Etanercept in children with polyarticular juvenile rheumatoid arthritis. *N Engl J Med.* (2000) 342:763–9. doi: 10.1056/NEJM200003163421103
65. Hull DN, Cooksley H, Chokshi S, Williams RO, Abraham S, Taylor PC. Increase in circulating Th17 cells during anti-TNF therapy is associated with ultrasonographic improvement of synovitis in rheumatoid arthritis. *Arthritis Res Ther.* (2016) 18:303. doi: 10.1186/s13075-016-1197-5
66. Hull DN, Williams RO, Pathan E, Alzabin S, Abraham S, Taylor PC. Anti-tumour necrosis factor treatment increases circulating T helper type 17 cells similarly in different types of inflammatory arthritis. *Clin Exp Immunol.* (2015) 181:401–6. doi: 10.1111/cei.12626
67. Poddighe D, Romano M, Gattinara M, Gerloni V. Biologics for the treatment of juvenile idiopathic arthritis. *Curr Med Chem.* (2018) 25:5860–93. doi: 10.2174/0929867325666180522085716
68. Webb K, Wedderburn LR. Advances in the treatment of polyarticular juvenile idiopathic arthritis. *Curr Opin Rheumatol.* (2015) 27:505–10. doi: 10.1097/BOR.0000000000000206
69. Davies R, Gaynor D, Hyrich KL, Pain CE. Efficacy of biologic therapy across individual juvenile idiopathic arthritis subtypes: a systematic review. *Semin Arthritis Rheum.* (2017) 46:584–93. doi: 10.1016/j.semarthrit.2016.10.008
70. Scarsi M, Paolini L, Ricotta D, Pedrini A, Piantoni S, Caimi L, et al. Abatacept reduces levels of switched memory B cells, autoantibodies, and immunoglobulins in patients with rheumatoid arthritis. *J Rheumatol.* (2014) 41:666–72. doi: 10.3899/jrheum.130905

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