



Pleiotropic Role and Bidirectional Immunomodulation of Innate Lymphoid Cells in Cancer

Zhengwen An^{1*}, Fabian Flores-Borja², Sheeba Irshad¹, Jinhai Deng³ and Tony Ng^{1,3,4*}

¹ KCL Breast Cancer Now Research Unit, Guys Cancer Centre, King's College London, London, United Kingdom, ² Centre for Immunobiology and Regenerative Medicine, Barts and The London School of Medicine and Dentistry, Queen Mary University of London, London, United Kingdom, ³ Richard Dimbleby Department of Cancer Research, Comprehensive Cancer Centre, Kings College London, London, United Kingdom, ⁴ UCL Cancer Institute, University College London, London, United Kingdom

Innate lymphoid cells (ILCs) are largely tissue resident and respond rapidly toward the environmental signals from surrounding tissues and other immune cells. The pleiotropic function of ILCs in diverse contexts underpins its importance in the innate arm of immune system in human health and disease. ILCs derive from common lymphoid progenitors but lack adaptive antigen receptors and functionally act as the innate counterpart to T-cell subsets. The classification of different subtypes is based on their distinct transcription factor requirement for development as well as signature cytokines that they produce. The discovery and subsequent characterization of ILCs over the past decade have mainly focused on the regulation of inflammation, tissue remodeling, and homeostasis, whereas the understanding of the multiple roles and mechanisms of ILCs in cancer is still limited. Emerging evidence of the potent immunomodulatory properties of ILCs in early host defense signifies a major advance in the use of ILCs as promising targets in cancer immunotherapy. In this review, we will decipher the non-exclusive roles of ILCs associated with both protumor and antitumor activities. We will also dissect the heterogeneity, plasticity, genetic evidence, and dysregulation in different cancer contexts, providing a comprehensive understanding of the complexity and diversity. These will have implications for the therapeutic targeting in cancer.

Keywords: ILCs, cancer, tumor immune microenvironment, immunosurveillance, heterogenity, plasticicity, immunothearpy

INTRODUCTION

Innate lymphoid cells (ILCs) derive from common lymphoid progenitors (CLPs) (1, 2) and are regulated by multiple endogenous signals including neuropeptides, hormones, cytokines and other alarmins (3). Tissue-resident ILCs represent a heterogeneous group of cells that interact with a wide variety of hematopoietic and non-hematopoietic cells through direct or indirect communication. These ILCs act as functional effectors contributing to the integration of innate and adaptive immune responses and the orchestration of physiological and pathological processes throughout the body (1). ILCs are considered to be counterparts of T-cell subsets resembling both their phenotypical and functional characteristics. Five subgroups of ILCs have been classified as natural killer (NK) cells and helper-like ILCs (ILC1s, ILC2s, and ILC3s) and lymphoid tissue inducer (LTi) cells in terms of their lineage-determining transcription factors and cytokine secretion profiles according to a redefined nomenclature (4) (Figure 1). NK cells derive from NK-cell precursors (NKP), which directly differentiate from common innate lymphoid progenitors (CILPs),

OPEN ACCESS

Edited by:

Nicolas Jacquelot, Walter and Eliza Hall Institute of Medical Research, Australia

Reviewed by:

Giuseppe Sciumè, Sapienza University of Rome, Italy Laura Chiossone, Innate Pharma, France

*Correspondence:

Zhengwen An zhengwen.an@kcl.ac.uk Tony Ng tony.ng@kcl.ac.uk

Specialty section:

This article was submitted to NK and Innate Lymphoid Cell Biology, a section of the journal Frontiers in Immunology

> Received: 30 September 2019 Accepted: 20 December 2019 Published: 04 February 2020

Citation:

An Z, Flores-Borja F, Irshad S, Deng J and Ng T (2020) Pleiotropic Role and Bidirectional Immunomodulation of Innate Lymphoid Cells in Cancer. Front. Immunol. 10:3111. doi: 10.3389/fimmu.2019.03111

1

whereas other ILCs derive from common helper innate lymphoid progenitors (CHILPs) (2, 5-7). CHILP can differentiate to ILC precursors (ILCPs) and LTi precursors (LTiPs) that give rise to all ILC subsets (ILC1, ILC2, and ILC3) and LTis, respectively (4, 8). The transcriptional repressor, inhibitor of DNA binding 2 (Id2), is sequentially expressed in the ILC lineage framework, and Id2-dependent precursors can further differentiate with lineagespecific transcription factors (9). Data from both humans and mice demonstrate that LTi cells and conventional NK (cNK) cells are developmentally related yet represent distinct lineages (10-12). Recent studies reveal the previously unappreciated ILCP heterogeneity by using polychromic reporter mice to identify the ILCP lineage. The results suggest that a fraction of CHILP highly expresses PLZF (Zbtb16) and PD-1 and retains potential for all CD127^{hi}ILCs but not LTis. In addition, these cells can also generate NK cells that express Eomes and perforin, indicating additional lineage potential (13, 14). Likewise, an analysis of Id2 and PLZF reporter mice reveals that Id2+Zbtb16+ILCPs define multipotent NK and/or ILCPs and are associated with loss of LTi potential at a clonal level, suggesting a revised model for ILC differentiation that redefines the cell-fate potential of helper-ILC-restricted Zbtb16⁺ILCPs (15) (Figure 1).

NK cells and ILC1s share the same transcription factor T-bet and produce type 1 cytokines, including interferongamma (IFNy) and the tumor necrosis factor-alpha (TNFa), in response to interleukin (IL)-12 and IL-18, whereas NK cells also require the transcription factor eomesodermin (Eomes) and produce cytotoxic proteins, perforin, and granzymes in mice and granulysin in humans (16). NK cells and ILC1s are functionally analogous to cytotoxic CD8⁺ T cells and CD4⁺ T helper1 (Th1) cells, respectively. ILC2s depend on GATA3 and produce IL-4, IL-5, IL-9, IL-13 (17, 18), and amphiregulin (AREG) (19) in response to IL-25, IL-33, and thymic stromal lymphopoietin (TSLP), which resembles the Th2 cells response (4). ILC3s are heterogeneous but consistently require transcription factor RORyt for their development and function. LTis are related to ILC3s with distinct functions in the secondary lymphoid organ formation during both embryonic and adult stages of development (8, 11, 20). In general, these cells found in adult mice are termed LTilike cells, which have a similar phenotype to LTis (21). Both ILC3s and LTis are capable of producing IL-17, IL-22 and granulocyte-macrophage colony-stimulating factor (GM-CSF) in response to IL-23, IL-1β, or natural cytotoxicity receptor ligands (NCR-L) that reflect Th17 response (3, 22). Regardless of the functional association with ILC3s, LTis are considered a separate ILC lineage. Fetal liver-derived LTiPs express α4β7 integrin and C-C Motif Chemokine Receptor 6 (CCR6), whereas adult LTis derived from bone marrow precursors display upregulated RORyt in a Notch-dependent manner (9, 20). Adult mice also have T-bet-expressing CCR6⁻NKp46⁺ILC3s derived from CCR6⁻NKp46⁻ILC3s (23) (Figure 1).

In contrast to the polarized T-cell subsets that take over several days for activation and clonal expansion, most ILCs can produce significant amounts of cytokines upon stimulation without further differentiation. Hence, ILCs are considered to be the first line of defense to confront and sense the changes in the local environment and then react rapidly as the host response in peripheral tissue (24). Dysregulation of ILCs manifested by the changes in cell numbers or subset proportions is associated with diverse inflammatory diseases and cancer (25–28). The role of ILCs in cancer is ambivalent based on both protumor and antitumorigenic activities depending on their phenotype, the variety of cancer, and the tumor microenvironment (TME) context. The remarkable heterogeneity, distinct signature cytokines they produce, the various surface markers they express, and the plasticity among all the different subsets make ILC functions divergent and less comprehensible.

Given that the role of NK cells in cancer has been extensively reviewed (29, 30), this review will mainly focus on helper-like ILCs (ILC1s, ILC2s, and ILC3s) and LTis in the context of cancer. The plasticity of ILCs and their interactions with the extracellular matrix (ECM) and adaptive immune cells shape the TME. In return, changes in the secretion of cytokines by these cells also polarize ILC functions and influence plasticity under pathological conditions in the TME. These bidirectional interactions are crucial for controlling tumor growth and metastasis. Untangling the pleotropic roles and bidirectional regulations of ILCs in cancer will ultimately help to provide a rationale for the design of therapeutic strategies for cancer treatment.

INNATE LYMPHOID CELLS AND TUMOR IMMUNE MICROENVIRONMENTS

Tumor-infiltrating leukocytes were first discovered in the 1800s, suggesting a functional relationship between immune cells and cancer (31). These infiltrating immune cells had been considered to have antitumor properties only until the last three decades, when the adverse role of promoting tumor progression has come into light. Tumor development and progression are profoundly influenced by a variety of resident host cells, stromal cells, the ECM, the blood and lymphatic vascular networks, infiltrating immune cells, and signaling molecules including growth factors, cytokines, and chemokines, collectively known as the TME (32-35). Recognizing the importance of the TME in cancer has revolutionized cancer treatment from targeting tumor cells to deciphering the tumor ecosystem complexity. This fundamentally determines the fate of the primary tumors whether it is eradicated or a premetastatic niche is established that favors metastatic dissemination (36, 37). Substantial immune cell infiltration varying in size, composition and distribution is a distinct characteristic present in almost all types of malignant tumors. These infiltrating immune cells, including tumor-associated macrophages (TAMs), mast cells, T cells and ILCs within the TME, comprise the tumor immune microenvironment (TIME) (26, 38-43).

Each of these immune components is believed to be involved in tumorigenesis, tumor invasion, and metastasis (44, 45). From immunosurveillance to tumor escape, infiltrating immune cells mutually interact with other cells within the TME niche and play a dual role that can either promote or attenuate malignant outgrowths. Given that tumor-induced immunological changes affect metastatic progression even before disseminated tumor cells reach secondary organs (38), targeting the premetastatic



TIME is believed to be critical for the success of therapeutic strategies (40, 42, 46).

ILCs are tissue-resident cells in lymphoid and non-lymphoid organs and functionally act as sentinels to maintain and shape tissue microenvironment. They respond rapidly to environmental changes by secreting significant amounts of cytokines and expanding locally under physiologic and pathological conditions, suggesting a role for these cells in the early phases of tumorigenesis and in shaping the TME (46).

ILCs in Extracellular Matrix Remodeling

ECM is a major component of the local TME, and its remodeling plays a prominent role in tumor development. ECM dysregulation promotes cancer cell invasion, induces angiogenesis and facilitates immune cell infiltration (47). Activated immune cells including ILCs closely interact with ECM, which may decisively affect the outcome of tumor progression (42).

Among the ILC family members, NK cells were first identified as being involved in the remodeling of the tumor ECM. NKp46mediated IFNy secretion increases the expression of fibronectin 1 (FN1) by tumor cells leading to ECM structural changes in the primary tumors and decreased metastasis (48). Reciprocally, FN1 stabilizes NK cell survival and facilitates NK cell migration by inducing anti-apoptotic protein B-cell leukemia 2 (Bcl-2) (49). These data suggest a bidirectional regulation between NK cells and ECM remodeling. IL-12-responding ILC1s induce a signature TME characterized by strong upregulation of IFN γ and type I-associated chemokine receptors (CXCR6, CCR5, CXCR3, and CCR1) to recruit tumor invading *Rorc^{fm+}*ILCs in a mouse model of melanoma (50). Adipose ILC1s were also found to contribute to adipose tissue fibrogenesis in humans (51). Other studies of ILC2s show that activation by IL-33 induces IL-13 secretion, which in turn stimulates hepatic stellate cells to produce ECM proteins leading to pathologic tissue remodeling in mouse liver (52) (Figure 2). Depletion of RORyt-dependent LTis and RORyt-independent ILCs impairs lung function and tissue repair, whereas IL-33/IL-33R signaling is critical for lung



ILC response through the production of AREG via IL-13/ IL-22-independent mechanisms (53).

As described above, the remodeling of ECM causes organ dysfunction and contributes to chronic disease mechanisms including cancer. Innate immune cytokines derived from ILCs subsets directly regulate fibroblast functions that are independent of adaptive immunity (54). Taken together, these studies demonstrate a critical role for ILCs as important regulators of cytokine networks involved in ECM remodeling and the consequences for health and disease. This also suggests the clinical relevance of controlling pathologic remodeling to prevent metastasis.

ILCs in Tumor Angiogenesis and Lymphatic Vascular Networks

Tumoral angiogenesis is a characteristic feature of tumor survival and progression, which differs considerably from the vascular structure generated for the regular blood vessels (55, 56). The formation of new blood vessels in tumors is driven by hypoxic tumor cells, tumor-associated stromal cells (TASCs), and the ECM in which they are embedded, producing vascular endothelial growth factor A (VEGFA) to initiate tumor angiogenesis (57). Tumor-infiltrating immune cells, such as ILCs, are also important pro-angiogenic mediators to increase VEGFA bio-availability and signaling during the angiogenic switch (58). ILCs stimulate endothelial cell proliferation and upregulation of adhesion molecules by releasing pro-angiogenic factors, contributing to immune cell recruitment and perpetuation of inflammation (42, 55, 56, 59).

Tumor-infiltrating NK cells, transformed from CD16⁺NK to CD16⁻NK subset upon transforming growth factor-beta (TGFβ) stimulation, predominate in non-small-cell lung carcinoma (NSCLC) and produce elevated levels of pro-angiogenic factors such as VEGF and placenta growth factor (PIGF) to sustain tumor progression (55, 60, 61). This pro-angiogenic effect can be reversed by transcriptional factor STAT5 via repressing VEGFA transcription in NK cells, in both mice and humans (62). The secretion of VEGF by NK cells is associated with angiogenesis in human melanoma, and breast and colon carcinoma (42). These studies suggest that the TIME is able to functionally affect NK cells by inhibiting their cytotoxic ability or promoting pro-angiogenic phenotypes by a wide array of cytokines and soluble factors such as TGF- β , PGE2, VEGF, and adenosine (55). Likewise, tumor-infiltrating ILC1s in mouse fibrosarcoma also showed upregulation of angiogenesis gene sets that also expressed in NK cells (63). ILC1s impaired tumor neovascularization mediated by IL-12, which required the presence of NK cells to induce endothelial cell cytotoxicity in lymphomas (64). ILC1s produce two signature cytokines, IFN γ and TNF α , which are associated with cell proliferation and angiogenesis. IFNy activates the transcription factor STAT1 to inhibit tumor cell proliferation and angiogenesis (65, 66). TNF α can either destroy tumor vasculature and induce apoptosis as an antitumor effector or stimulate the expression of angiogenic and growth factors to promote tumor formation and growth (38). ILC2s in response to IL-33, the cytokine that induces angiogenesis and vascular permeability through ST2 receptor binding (67), can enhance the re-epithelialization and promote the restoration of skin integrity after injury (68). ILC3s contribute to preserving epithelial integrity and maintaining tissue homeostasis by the release of IL-22 (59). Secretion of IL-17 by ILC3s promotes angiogenesis via stimulation of vascular endothelial cell migration and cord formation, resembling those indirect angiogenic stimulators such as TGF-B and platelet-derived growth factor B subunit homodimer (PDGF BB) in vivo (69). The indirect role of ILC3s in tumor angiogenesis is also manifested by their recruitment of myeloid-derived suppressor cells (MDSCs) and regulatory T cell (Treg) cells, which in turn promote M2-like macrophages in the TIME (70, 71). Apart from IL-17 and IL-22, the LTi-like neuropilin (NRP)1⁺ILC3 subset was also found to release CSF2, TNFα, B-cell-activating factor, and CXCL8, in association with VEGF production that might contribute to angiogenesis (59) (Figure 3).

The other prominent feature of tumor angiogenesis is the expression of adhesion molecules such as vascular cell adhesion molecule (VCAM) and intercellular adhesion molecule (ICAM), which conveys the apparent tumor-immune privilege. In a subcutaneous melanoma mouse model, NKp46⁺LTi cells alter the tumor microvasculature upon IL-12 stimulation, which leads to upregulation of VCAM and tumor suppression (72). Indeed, LTis modulate not only blood vasculature but also



FIGURE 3 | Innate lymphoid cells (ILCs) in tumor angiogenesis. ILCs act as tumor angiogenesis modulators by releasing pro-angiogenic factors and by inducing the recruitment and infiltration of immune cells to affect tumor-related inflammation. Transforming growth factor-beta (TGF-β) secreted by tumor cells activate natural killer (NK) cell to produce vascular endothelial growth factor (VEGF) and placenta growth factor (PIG) to induce tumor angiogenesis; conversely, the transcription factor STAT5 represses the expression of VEGF resulting in the inhibition of angiogenesis and tumor growth. ILC1s produce two signature cytokines, interferon-gamma (IFN γ) and tumor necrosis factor-alpha (TNFα), that are associated with cell proliferation and angiogenesis. TNFa secreted by ILC1s increases vascular cell adhesion molecule (VCAM)1 expression causing tumor vascular formation, whereas in a different context, TNFa-producing ILC1s can either destroy tumor vasculature or induce apoptosis acting as antitumor effectors. Furthermore, IFNy released from ILC1s causes STAT1 activation, thereby inhibiting angiogenesis formation. ILC2s respond to IL-33 and induce angiogenesis and vascular permeability through ST2 receptor binding. IL-17 and IL-22 released by ILC3s promote angiogenesis via stimulation of vascular endothelia cell migration and cord formation. The indirect role of ILC3s in tumor angiogenesis is also shown in the recruitment of myeloid-derived suppressor cells (MDSCs), regulatory T cell (Treg) cells, and the promotion of M2-like macrophages in the tumor immune microenvironment (TIME).

the lymphatic vascular system. LTis induce mesenchymal stem cells (MSCs) to produce chemokines, CCL19, CCL21, or CXCL13, which promote lymphocyte recruitment and spatial compartmentalization (73). This cross talk also plays a role in promoting lymph node metastasis in breast cancer. In the 4T1.2 triple-negative breast cancer (TNBC) mouse model, ILC3s are recruited to the primary tumors by CCL21 and stimulate tumor stromal cells to release CXCL13, leading to enhanced tumor cell motility, lymphangiogenesis, and lymph node invasion by tumor cells (74). These data suggest that the number of infiltrating ILCs within the primary breast tumors could be used as a predictor of metastatic and malignancy potential (74).

Tumor angiogenesis and lymphatic vascular formation prompt tumor invasion and metastasis, the landmark events that transform a locally growing tumor into a systemic metastatic and life-threatening disease. As tumor-infiltrating ILCs can polarize the TME to either protumor or antitumor effects by the modulation of angiogenic activities and lymphatic vascular networks, these cells represent valid targets for antitumor immunotherapy and cancer preventive strategies (55).

Interplay Between ILCs and Cytokines, Chemokines and Growth Factors in Tumor Immune Microenvironment

Initiation of ILC response relies on sensing the cytokines, alarmins, and inflammatory mediators that are derived from tissue sentinels such as myeloid cells, dendritic cells (DCs) and macrophages, or epithelial cells to translate environmental signals into a specific cytokine profile (75). The complex, diverse and dynamic interplay with surrounding environments amplifies ILC signaling and determines their function. Tumor-infiltrating immune cells engage in an extensive and dynamic interaction with TIME and shape the TME, whereas tumor cells also induce an immunosuppressive microenvironment by the secretion of the cytokines and other soluble factors (33).

In a model of subcutaneous melanoma, ILC1s respond to IL-12, produced by tissue sentinels such as DCs and macrophages, and alter the TME at an early stage of tumor development to facilitate tumor suppression by infiltrating immune cells (72). The production of TGF- β in the TME drives ILC fate outcomes between the ILC subsets from NCR⁻ILC3s to NCR⁺ILC3s (76). In human ovarian cancer, IL-22-producing ILC3s prevent tumorinfiltrating lymphocyte activation and proliferation (27), whereas the release of IL-22 and IL-17 by ILC3s modulates intestinal immune pathology (77). In a preclinical mouse model of human breast cancer, RORyt+ILC3 promoted lymphatic metastasis by modulating the local chemokine milieu in the TME (74). Genetic mapping of the fate of ILC3 revealed that different subsets of ILC3 are phenotypically adapted to the local tissue environment they invade, indicating that the function of ILCs is shaped not only during their lineage commitment but also by the TME (50). Although ILCs are characterized by the distinct signature cytokine they express, tissue-derived cytokines can extrinsically shape ILC function (50), and this cross talk can drive both positive and negative consequences in tumor progression (78).

Interestingly, exosomes (extracellular vesicles, 50-150 nm in size released from the endocytic compartment) (79) derived from both tumors and stromal cells have been implicated as signaling mediators between cancer cells and surrounding cells that comprise the TME and contribute to therapy resistance. Several studies have identified the role of exosomes in premetastatic niche establishment that is favorable for future dissemination and metastatic seeding through the communication with immune cells like NK cells, macrophages, B cells, and T cells. Recent studies also revealed that chemotherapy-elicited extracellular vesicles in breast cancer cells can promote premetastatic niche formation in lung by delivering annexin 6 to induce CCL2, which promotes monocyte activation (80). The direct correlation of exosomes with ILCs has not been explored yet, although AREG, which is produced by ILC2s, can be found in distinct nanoparticles (DNPs), a subset of extracellular vesicles that are smaller than 50 nm with a dot-shaped morphology (81). Further investigation of ILCs and exosome communication would potentially allow the development of effective clinical applications that may overcome the therapeutic resistance.

Interplay Between ILCs and Adaptive Immunity

Emerging evidence indicates that ILCs play an important role in coordinating tissue-specific adaptive immunity and act as master regulators in homeostasis and diseases. Substantial studies have demonstrated the direct communication between ILC2s and ILC3s with the adaptive immune system, whereas with ILC1s, it remains less well-investigated (21).

ILCs share several developmental and transcriptional signatures with T-cell subsets but have additional and/or unique, non-redundant functions in T-cell polarization and effector functions by secretion of regulatory cytokines, antigen presentation, or direct cellular interactions within complex tissue microenvironments. ILC2s functionally resemble Th2 cells and play a critical role in the type II immune response. ILC2-derived cytokine IL-13 stimulates tissue DC recruitment to the draining lymph node for eliciting production of Th2 cell-attracting chemokine CCL17, causing the Th2 response (82). The protumor activity of IL-13 is also involved in the activation and differentiation of MDSCs by induction of Treg cell expansion to establish an immune-suppressive TME (83). These studies may indicate that the role of ILC2/IL-13 axis in the initiation of cancers is linked to extensive tissue remodeling. During intestinal or lung inflammation, tissuespecific IL-33 secretion upregulates OX40L expression on ILC2s resulting in protective Th2 immunity and Treg cell expansion, which could be related to the induction of a tolerant microenvironment and inhibition of antitumor responses (84). In addition, ILC2-derived AREG limits antitumor immunity by stimulating Treg cell expansion, supporting the establishment of an immune-suppressive TME (85). ILC2s have been suggested as a key source of Th2 master regulator IL-4 (86), and the previously unrecognized role of ILC2-derived IL-4 in Th2 differentiation may indicate the potential involvement of ILC2/T-cell interactions in promoting tumor growth and metastasis (38). Notwithstanding the protumor role of ILC2s in cancer, ILC2-derived cytokines such as IL-5 are involved in an antitumor function. In a melanoma model, increased IL-5 production by ILC2 resulted in the recruitment and activation of eosinophils, which was correlated with lung tumor metastasis. Moreover, ILC2-derived IL-9 has recently gained attention in the context of tumor biology and has functional importance as growth factor for ILC2 and survival factor for other cell types (83). In a melanoma model, IL-9 deficiency promoted tumor growth, whereas IL-9 treatment decreased metastasis (87). It is possible that the role of IL-9 in cancer is associated with the promotion of infiltrating DC antigen presentation (88). Conversely, high expression of IL-9 was associated with T-cell transformation in humans, indicating that the anti-apoptotic properties on transformed cells promote tumorigenesis (89) (Figure 4).

Increasing evidence over the past decade also indicated the role of ILC3s in bridging innate and adaptive immune responses.

In a bone marrow chimeric mouse model, RORyt⁺ILCs (LTis) express OX40L and CD30L to regulate memory CD4⁺ T-cell survival but not that of CD8⁺ T cells in the homeostatic state (90), whereas in normal intestines ILC3s limit Tcell activation but decrease the regulatory effects under an inflammatory microenvironment through MCH-II presentation (91), indicating that the function of ILCs can be altered by mutual interaction with the microenvironment. ILC3s localize at the boundary between B-cell follicles and T-cell zone within intestinal draining lymph nodes to regulate T follicular helper cells (Tfh) responses and B-cell class switching via antigen presentation (92). In contrast to peripheral lymph nodes, resident ILC3s at mucosal draining lymph nodes express MHC-II and contribute to a distinct microenvironment constraining CD4⁺ T-cell response to commensal bacteria in a contact-dependent manner dependent on CCR7 (93). Commensal microbiota and IL-1ß stimulate the release of GM-CSF and IL-22 by ILC3 through MHC-II-dependent interactions to promote intestinal Treg cell expansion (94, 95). In addition, IL-1β stimulation leads to activation of peripheral ILC3s and consequently primes the CD4⁺ T-cell immune response, whereas T cell-derived effector cytokines also cross-regulate the ILC response. The cognate interaction of ILC3 and CD4⁺ T cells is bidirectional, indicating an unappreciated role of ILC3s in T cell-mediated immunity (96). ILC3 also activates local and systemic inflammatory T cells by increasing TNF-like ligand 1A (TL1A) and IL-1β (82, 95). In the peritoneal B-cell compartment, RORyt+ILCs produce soluble lymphotoxin a3 (sLTa3) and membranebound lymphotoxin β (LT β), both trimeric cytokines of the TNF superfamily, resulting in T cell-dependent and T cellindependent IgA production, respectively (97). Moreover, the adherent microbiota stimulates ILC3 to secrete IL-22 that triggers epithelial serum amyloid A protein (SAA) production to promote local Th17 responses in a STAT3-dependent manner (98) (Figure 4). The proliferative and anti-apoptotic capacities of ILC3-derived IL-22 may support malignant transformation in chronic inflammatory diseases. Those studies revealed the potential function of ILC3s as antigen-presenting cells and directly modulate adaptive immune responses. Even though ILCs show striking similarities to T-cell subsets, the non-redundant functions go beyond those shared with T cells through indirect and direct interaction with adaptive immune cells. In turn, adaptative immunity reciprocally regulates ILCs, which indicates that these bidirectional regulations are a crucial determinant of immune response within tissues. Given that the abundant evidence is mostly established under inflammatory conditions, a functional analysis between ILC3s and adaptive immunity in chronic inflammatory diseases may also imply the role of these interactions in the malignant transformations.

Collectively, these studies demonstrate key innate mechanisms in the regulation of the adaptive immune response and shed light on the tissue ILC-mediated control of adaptive immunity in orchestrating inflammatory and restorative tissue responses (99). Delineating the complex network of ILCs with other immune cells interactions will allow the manipulation of their specific function and ultimately open a way to innovative therapeutic strategies (41).



FIGURE 4 Innate lymphoid cells (ILCs) modulate adaptive immunity by direct or indirect interactions with other cells in the tissue. ILC2s functionally resemble Th2 cells and play a critical role in the type 2 immune response. The role of ILC2s in the establishment of an by direct interactions with Th2 cells through OX40L/OX40 under IL-33 stimulation or through indirect interactions by the section of IL-13 or amphiregulin (AREG), which then causes regulatory T cell (Treg) cell expansion and myeloid-derived suppressor cell (MDSC) activation. ILC2-derived IL-13 also stimulates dendritic cell (DC) recruitment and CCL17 production to provoke a Th2 response. ILC2s produce IL-4 to regulate Th2 differentiation, which may suggest a potential role in promoting tumor growth and metastasis. In terms of antitumor function, the release of IL-5 and IL-9 by ILC2s restrains tumor growth through eosinophils recruitment and activation and DC antigen presentation. Conversely, high expression of IL-9 is associated with T-cell transformation in humans, indicating that the anti-apoptotic properties on transformed cells promote tumorigenesis. ILC3s are important in the Th17 immune response. The direct interactions with CD4+T cells are either through MHC II/TCR or through OX40/OX40 and CD30L/CD30 to regulate T-cell survival or B-cell class switch, respectively. ILC3 migrate to the draining mesenteric lymph node to initiate an adaptive response that requires CCR7 expression. ILC3s promote IgA production in a T cell-dependent manner via the regulation of CD4+T-cell homing by sLTa3 secretion and also in a T cell-independent manner via the regulates macrophages and DCs to release IL-10, and this consequently results in regulatory T cell (Treg) cell expansion. In addition, IL-18 produced by macrophages activates ILC3 and leads to CD4+T-cell priming, whereas T cell-derived cytokines also cross-regulate ILC response, indicating the bidirectional interactions between ILC3 and T cell-mediated immunity.

INNATE LYMPHOID CELLS AND CANCER-RELATED INFLAMMATION

Inflammation is a well-known hallmark of cancer that sustains cancer progression by providing growth factors, proangiogenic factors, and ECM modifying enzymes. During the different stages of tumor development, the inflammatory response plays decisive roles in tumor initiation, promotion, malignant conversion, invasion, and metastasis (100). Inflammatory conditions can be present before or after a malignant change occurs, which induces an inflammatory microenvironment to promote tumor development (101). Cancer-related inflammation in the presence of inflammatory cells and their mediators, such as cytokines, chemokines and prostaglandins, in tumor tissue is similar to that observed in a chronic inflammatory response (101). The character of the inflammatory response and the composition of the immune cell infiltration are indispensable to shape the TIME (33).

ILCs are believed to be resident, whereas NK cells recirculate. Given their distribution throughout the body, ILCs play a critical role in coordinating inflammation and cancer (75). ILC1s are highly abundant in the small intestine, liver, uterus, and salivary glands; ILC2s are enriched in the skin and lung; and ILC3s are abundant in the colon and tonsils (27, 102). Mucosal ILC1s are important in barrier defense through escalating Th1-like responses against pathogenic infections (99). During inflammation, a higher frequency of IFNy-producing ILC1 subsets in mucosal tissue is positively correlated with patients having Crohn's disease compared with the normal tissue, indicating a role for ILC1 in the pathogenesis of gut mucosal inflammation (103). ILC2s secrete the type 2 cytokines IL-5 and IL-13 to control helminths, restrain parasitic infections (99), and drive airway hyper-reactivity in mouse models of allergic asthma (103, 104). Tissue-derived cytokines can also shape ILC function in inflammation, and this is largely achieved by the IL-22 pathway (78). In response to IL-23 stimulation, IL-22-producing ILC3s facilitate tissue protection from the acute phase of Citrobacter rodentium-induced colitis (105). ILC3-dependent protection also occurs at other mucosal sites, including the lung and oropharyngeal barrier through IL-17 and IL-22 (106-108). ILCs are important in tissue repair and remodeling after infection. IL-22⁺ILC3s limit inflammatory colitis, and their frequency correlates with mucosal repair in inflammatory bowel disease (IBD) (99). LTis are involved in tissue repair after clearance of lymphocytic choriomeningitis virus infection (109), and ILC2s assist lung tissue repair after influenza virus infection (53).

Indeed, ILCs act as a two-edged sword in inflammation. The same factors that inhibit acute inflammation and promote tissue repair can have pathogenic effects during chronic inflammation activation. IL-5-producing ILC2s boost lung inflammation by recruiting eosinophils that are the main allergic effectors, whereas maintenance of IL-13 production stimulates collagen deposition leading to chronic fibrotic tissue damage and pulmonary fibrosis in hyper-reactive lung airways (110-112). In the colon, chronic infection-induced IL-23 triggers RORyt+ILC accumulation and increases IL-17 and IFNy production that results in the bacteriadriven colitis (77). Furthermore, the protumor activities of ILC3 are mainly associated with chronic inflammation by IL-23-induced secretion of IL-17 and IL-22 (113). Antigenpresenting ILC3s act as a checkpoint to control Tfh and B cells response toward mucosal-dwelling microbiota (92). Interestingly, regulatory ILCs (ILCregs) are found in both human and mouse to protect intestines from inflammation through inhibition of ILC1 and ILC3 activation regulated by IL-10 secretion (114).

ILCs as mediators of inflammation in various organs play a key role in tissue remodeling and repair after recruitment and activation, which are controlled by cytokines and growth factors that are selective for each ILC member. Hence, targeting the regulation of the tissue-specific response of ILCs in inflammation may be beneficial in the setting of inflammation-driven cancer.

INNATE LYMPHOID CELLS IN PROTUMOR AND ANTITUMOR EFFECTS

The role of ILCs in cancer is ambivalent as a double-edged sword by engaging in cancer immune surveillance or enhancing the unique tumor-promoting milieu, depending on the conceptual framework. We review here the different subsets of ILCs in both antitumor and protumor activities in the diverse contexts of cancer (**Table 1**).

ILC1s

ILC1s are defined by the expression of T-bet and the production of the signature cytokine IFN γ . The prototypical member of ILC1s is the NK cell. NK cells recirculate and show strong cytotoxicity, whereas ILC1s are tissue resident and only exhibit weak cytotoxicity but produce more IFN γ and GM-CSF (131) than NK cells do (26). ILC1s in response to IL-12, a potent antitumor cytokine (115), produce the effector cytokines, IFN γ and TNF α , to limit tumor growth in a melanoma mouse model (132). The antitumor properties of ILC1s also depend on IL-15. The expanded ILC1s in mouse mammary pre-cancerous lesions exhibit potent cytotoxicity to limit tumor growth in response to IL-15 (116).

However, ILC1s also show a more complex behavior and display a detrimental role in cancer. Alteration of ILC subsets composition in patients with acute myeloid leukemia (AML), compared with the controls, could change the protective function of ILC1s by increasing their frequencies and reducing IFN γ and TNF α (118). Conversely, even though IFN γ is currently used as a clinical target for treatment of malignancies, enhanced IFN γ

production by ILC1s is not associated with a better immune response but shows an immunosuppressive effect in the presence of IL-12-producing chronic lymphocytic leukemia (CLL) cells (117, 124). Moreover, ILC1s are highly abundant in colorectal cancer and produce IFN γ in response to IL-15 and IL-18, thus inducing an immunosuppressive environment (102, 133). These data suggest a dark side of IFN γ in tumor-promoting effects in cancer, and further investigation might help to understand the complex mechanisms of cancer progression in different contexts.

ILC2s

As an alarmin, IL-33 is released from nuclei to the extracellular milieu upon cell injury caused by cell stress or damage and activates ILC2s to initiate a type 2 immune response (120). IL-33-dependent tumor-infiltrating ILC2s mobilize from lung and recruit DCs to promote the adaptive T-cell response (119). ILC2 antitumor activity under IL-33 stimulation is also involved in recruiting eosinophils to limit tumor growth and in shaping chemokine profiles in the TME under IL-5 release (134). IL-33 promotes ILC2 secretion of CXCR2 ligands that bind to CXCR2-expressing tumor cells, reinforcing tumor cell-specific apoptosis independent of adaptive immunity (135). Secretion of IL-13 by ILC2s activates CD8⁺ T cells through DC recruitment (120). In addition, ILC2s are major sources of IL-9, which can decrease tumor metastasis in a mouse model of melanoma (87).

Conversely, IL-13-producing ILC2s also show protumor immunity and are associated with a negative outcome in cancer. The ILC2/IL-13 axis modulates T cell-to-MDSC balance to drive an immunosuppressive microenvironment, whereas the inhibition of ILC2/IL-13 axis improves bladder cancer treatment (125). Similarly, the IL-33/ILC2 axis also accelerates tumor growth and promotes lung and liver metastasis by recruiting MDSCs to produce IL-13 in a mouse model of breast cancer (136), acute promyelocytic leukemia (APL), and prostate cancer (123). ILC2s impair IL-33-mediated tumor suppression by antagonizing the NK cell function during tumor growth via CD73 independently of adaptive immunity in a melanoma mouse model (119). Indeed, ILC2-derived IL-33 promotes tumor formation and metastasis in breast cancer by upregulating Treg cells through inducible co-stimulator (ICOS)/ICOS ligand (ICOSL) interaction (121), whereas IL-33 also activates ILC2s to recruit eosinophils through ICOS/ICOSL interaction during lung inflammation (137). Moreover, ILC2-derived AREG expression favors lung tumor growth and enhances resistance to apoptosis (126). Increased numbers of ILC2s were found in inflamed colonic tissue of patients with ulcerative colitis, a condition associated with a high risk for development of inflammationdriven colorectal cancer (122). Clinical studies have shown that ILC2s and their cytokines (IL-33 and IL-4) accumulate in peripheral blood and contribute to immune-suppressive environment formation in gastric cancer patients (138). The mediation of resistance to apoptosis indicates a poor prognosis in a number of cancers (139, 140).

All these data indicate that the function of ILC2s and their related cytokines are cell context dependent to facilitate either protumor or antitumor activities.

TABLE 1 | ILCs in different cancer types with anti-/pro-turmeric effects.

	Cancer types	Functions (anti-/pro-tumor)	References
ILC1s	Melanoma	Anti-tumor: ILC1s in response to IL-12, produce IFN $\!\gamma$ and TFN $\!\alpha$ to limit tumor growth.	(115)
	Colorectal Cancer	Anti-tumor: intraepithelial CD127 ⁻ ILC1s (ieILC1) produce cytotoxic granules to repress tumor growth.	(102)
	Mammary pre-lesion	Anti-tumor: ILC1s number expansion exhibits potent cytotoxicity to limit tumor growth in response to IL-15.	(116)
	CLL	Pro-tumor: CLL cells induce ILC1s produce IFN $\!\gamma$ and TNF $\!\alpha$ and form immunosuppressive environment.	(117)
	AML	Pro-tumor: reduce IFN γ and TNF α production.	(118)
ILC2s	Melanoma	Anti-tumor: IL-9-producing ILC2s inhibit tumor metastasis in mouse melanoma model.	(87)
	Lymphoma	Anti-tumor: IL-33 stimulates ILC2s to secrete CXCR2 ligands that bind to CXCR2-expressing tumor cells to induce tumor cell-specific apoptosis independently of adaptive immunity.	(119)
	Lung cancer	Anti-tumor: IL-33 dependent tumor-infiltrating ILC2s mobilize from lung and facilitate dendritic cells to promote adaptive T cell response. Pro-tumor: IL-33 actives ILC2s to recruit eosinophils through ICOS/ICOSL interaction.	(120) (121)
	Colorectal cancer	Pro-tumor: Increased ILC2 are associated with high risk for development of inflammation-related CRC.	(122)
	Breast cancer	Pro-tumor: IL-33/ILC2 axis facilitates tumor growth and metastasis by recruiting MDSCs to produce IL-13. Pro-tumor: ILC2s promote breast cancer growth by upregulating Tregs through ICOS/ICOSL interaction.	(123) (124)
	Bladder cancer	Pro-tumor: ILC2/IL-13 axis modulate T cell-to-MDSC balance driving an immunosuppressive TME.	(125)
	Gastric cancer	Pro-tumor: Accumulation of ILC2s and their cytokines, IL-33 and IL-4 contribute to immune-suppressive environment.	(126)
	Prostate cancer	Pro-tumor: ILC2 elevated tumor-derived PGD2 and B7H6 and activated MDSCs via IL-13 secretion to promote tumor growth.	(123)
	APL	Pro-tumor: ILC2 elevate tumor-derived PGD2 and B7H6 to active MDSCs via IL-13 thus inducing immunosuppressive effects.	(123)
ILC3s	Melanoma	Anti-tumor: IL-12 promotes NKp46 ⁺ CD49b ⁻ RORγt ⁺ ILC3 expansion and upregulation of VCAM1 to facilitate immune leukocyte infiltration resulting in tumor suppression.	(72)
	Lung (NSCLC)	Anti-tumor: NCR ⁺ ILC3s produce IL-22, TNF α , IL-8 and IL-2 to activate endothelial cells forming protective tumor-associated tertiary lymphoid structures.	(127)
	Colorectal cancer	Pro-tumor: ILC3s sustain colon cancer via production of IL-22.	(128)
	Breast cancer	Pro-tumor: CCL21-mediated recruitment of ILC3 triggers CXCL13 secretion by TME stromal cells thereby enhancing tumor cell motility and promoting lymph node metastasis.	(74)
	Hepatocellular carcinoma	Pro-tumor: NCR ⁻ ILC3s initiate IL-17 production upon IL-23 stimulation and promote hepatocellur carcinoma development.	(129)
	Cervical carcinoma	Pro-tumor: IL-17-producing ILC3s are also associated with poor survival in early stages.	(130)

ILC3s

Same as the other ILC subsets, ILC3s are also associated with both antitumor and protumor activities. NCR⁺ILC3 concentrates in human NSCLC and associates with intratumoral lymphoid structures, which contribute to the formation of an antitumor environment and act as predictors of favorable clinical outcome (127). In a mouse model of melanoma, increased IL-12 promotes NKp46⁺CD49b⁻RORγt⁺ILC3 expansion leading to upregulation of VCAM1 to facilitate leukocyte infiltration and the mediation of tumor suppression (72).

Conversely, ILC3s can be associated with tumor growth under different contexts. In a mouse model of colorectal cancer, NCR⁺IL-22⁺ILC3s, previously known as NK22 cells that are phenotypically distinct from LTis, sustain colon carcinogenesis in chronic inflammation (141). IL-22-producing ILC3s are crucial for the IL-22-mediated innate immune response. Notably, other subsets of ILC3s can also produce IL-22 such as CD4⁺ILC3s, which lack NCR expression in the gut (142). NCR⁻ILC3s promote hepatocellular carcinoma (HCC) development in response to IL-23 to produce IL-17, which directly limits CD8⁺ T-cell immunity by enhancing lymphocyte apoptosis and inhibiting their proliferation (129). By using an immunodeficient Rag^{-/-} mouse model, it has been shown that NKp46⁻CD4⁻ILC3 accumulation leads to IL-17 and IL-22 production, which promotes tumor development, whereas depletion of ILCs with anti-Thy1 mAbs limits tumor growth in colon cancer (141). Similarly, RORyt⁺ILC3s are associated with an increased incidence of metastasis in human breast cancer, as it was shown that depletion of ILC3 using an anti-CD90.2 antibody reduced tumor cells metastasis to lymph nodes in a syngeneic 4T1.2 mouse model (74). IL-17-producing ILC3s are also associated with poor survival in early stages of squamous cervical carcinoma (130). Interestingly, one study showed that a previously uncharacterized ILC population, ILCregs, can produce IL-22 and exhibit low cytotoxicity with a gene expression profile that overlaps with NK cells and other ILCs showing the same immunosuppressive capacity as Treg cells in human ovarian cancer studies (39).

Nevertheless, emerging evidence indicates the balance of power of ILCs in the context of cancer, and many ILC functions appear to be regulated by mechanisms distinct from those of other innate and adaptive immune cells. Notably, many studies of ILCs in cancer are based on the Rag-deficient mouse models lacking adaptive immunity. Although these experiments were crucial to analyze the fundamental features of ILC biology, these approaches have hampered the ability to study ILCs in the context of adaptive immunity and the potential interplay between ILCs and adaptive lymphocytes. The challenge now is to understand their roles and contribution within a complete immune system. The extent to which ILCs regulate the immune response (with antitumor or protumor activities) in a multitude of contexts need to be further analyzed. This is important for the understanding of their roles in cancer and for identifying targets for use in clinical treatments. In summary, although current studies reveal an important role of ILCs during tumor development, their indistinct functions in humans are incompletely understood.

INNATE LYMPHOID CELL HETEROGENEITY AND PLASTICITY

The pleiotropic roles of ILCs in cancer are dictated by their heterogeneity and plasticity, the noteworthy capacity of ILC subsets to convert into one another. Like helper T cells, the function and phenotype of ILCs can be modulated by extrinsic signals. In the context of cancer, mature ILCs exhibit substantial plasticity depending on the distinct cytokines present in the microenvironment milieus (143) (**Figure 5**).

NK Cells to ILC1s and ILC3s

In a mouse model of fibrosarcoma and melanoma, TGF- β in the TME induced mouse NK cells to convert into non-cytotoxic ILC1s, thus weakening the control over tumor growth and metastasis (63, 144, 145). Recent studies revealed that NK cells can permanently transform to Eomes⁻T-bet-dependent ILC1-like cells independently of ongoing infection, which may explain the discrepancies of ILC1-like cells with either having antitumor function or as an inflammatory subpopulation. This exemplifies the current limitations of NK cells and ILC categorization on the basis of only their phenotypes as discrete ILC lineages (146). NK cells can also be converted into ILC3s. IL-1 β stimulation prevents stage 3 NK cells from differentiating into stage 4 mature NK cells, thus keeping an ILC3-like phenotype with IL-22 production and IL-1R1 expression (147).

ILC1s to NK Cells, ILC2s, and ILC3s

In TGF- β knock-out mice, the ILC1 number decreased in salivary glands concurrent with increased expression of CD49a, CD103, and CD69, suggesting the potential for ILC1 to NK cell plasticity (148). IL-4 accumulation leads to a shift from ILC1s toward ILC2s, which can be reversed by IL-12, in patients with severe chronic obstructive pulmonary disease (COPD) or chronic rhinosinusitis with nasal polyps, suggesting the induction of IL-12 and IL-4 signatures in ILC1s and ILC2s, respectively (149). Differentiation of ILC1s to ILC3s is driven by IL-23 (103), and this can be enhanced by IL-1 β and retinoic acid produced by CD14⁻DC (103, 149, 150). The enhanced accumulation of ILC3s likely promotes tumorigenesis within different tissue locations (151).

ILC2s to ILC1s and ILC3s

Conversion of ILC2s into ILC1s has been shown in the intestines of patients with Crohn's disease (152) and in the lungs of patients with COPD (25). Inflammation may increase ILC plasticity, thereby increasing the risk of cancer development. IL-1 β also converts ILC2s into ILC1s, resulting in the increase of T-bet and IL-12R β 2 expression, which promotes ILC1s in response to IL-12 (153). Mouse lung inflammatory ILC2s, in response to IL-25, can differentiate into IL-17-producing ILC3s that play an important role in antihelminths and antifungal immunity (154).

ILC3s to NK Cells, ILC1s, and ILC2s

Human ILC3s treated with IL-15 and IL-12 initiate the expression of NK cell cytotoxicity markers (Eomes, CD94, CD56 NKG2A, and NKG2C), suggesting a beneficial role for ILC3-NK plasticity in cancer (155, 156). Plasticity between ILC3 and ILC1 mediated by IL-23 and also IL-12 produced by CD14⁺DC promotes polarization from ILC3s toward ILC1s, which ultimately affects tumor immunosurveillance in response to various environmental changes (2, 103, 150). IL-23-induced activation of STAT4 in NCR⁺ILC3s is a key determinant of plasticity from NCR⁺ILC3s toward ILC1s, as demonstrated by a transcriptomic analysis (157). Moreover, ILC3-to-ILC1 transition was tissue dependent and relied on transcription factor Aiolos and T-bet cooperation to repress regulatory elements active in ILC3s. This highlights the relevance of the tissue niche in creating a microenvironment that promotes ILC diversity and functional adaptation to local stimuli (158). ILC3s convert into ILC2s after activation with IL-2/TLR2 signals. In humans, IL-23 stimulation induces IL-22 secretion by ILC3s, which display clonal heterogeneity for IL-13 and IL-5 production indicating polarization in vivo (159).

NCR⁻ILC3s to NCR⁺ILC3s

Mouse NCR⁻ILC3s that differentiate into NCR⁺ILC3s need Notch signals; conversely, TGF- β can convert NCR⁺ILC3s into NCR⁻ILC3s, indicating that bidirectional plasticity of ILC3s is regulated by the balance between the opposing effects of Notch and TGF- β signaling in the face of challenges (76).

Interestingly, genetic evidence shows that the differences in specific chromatin regions such as *cis*-acting enhancers and silencers that bind to transcription factors may determine the apparent plasticity of ILCs compared with the T-helper subsets. With the use of ATAC-sequencing to evaluate chromatin regions accessibility, the regulatory regions that control the distinct cytokine gene expression in ILCs exhibit poised or active status, whereas the identical regions in CD4 T helper cell counterparts become accessible only after stimulation (160, 161). These results indicate that ILC regulomes are prone to dynamic changes, and the resilient feature of ILCs directly affects their plasticity in response to the TME.

As discussed above, ILC plasticity, the ability to modify their functional phenotypes, is a fundamental phenomenon that can contribute to tumor escape mechanisms by altering the ILCdependent tumor-surveillance system. Fully understanding the ILC plasticity control mechanisms is essential for manipulation of ILC function in the TME and designing novel therapeutic targets to prevent tumor metastasis.



FIGURE 5 The plasticity between innate lymphoid cell (ILC) subsets. Mature ILCs display substantial plasticity depending on the distinct cytokines, the microenvironment milieus, and the diverse context of cancer. Natural killer (NK) cells convert into non-cytotoxic ILC1s upon transforming growth factor-beta (TGF- β) stimulation in the tumor microenvironment (TME) in mouse fibrosarcoma and melanoma. NK cells also convert into ILC3s in response to IL-1 β , preventing stage 3 NK cells from stage 4 mature NK cells and retaining ILC3-like phenotype. ILC1s show plasticity toward all other ILCs subtypes, including NK cells, ILC2s, and ILC3s with the absence of TGF- β , the accumulation of IL-4, and under the stimulation of IL-23, respectively. Conversion of ILC2s to ILC1s needs IL-1 β , whereas ILC2s in response to IL-17-producing ILC3s. ILC3 treatment with IL-15 and IL-12 leads to NK cell conversion, whereas plasticity from ILC3s toward ILC1s and ILC2s depends on IL-23, IL-12, and IL-27LR2 signals. NCR⁻ILC3 and NCR⁺ILC3 conversion needs Notch and TGF- β signals, indicating the bidirectional plasticity of ILC3s in the face of different challenges.

TARGETING INNATE LYMPHOID CELLS FOR CANCER IMMUNOTHERAPY

Over the last two decades, cancer treatment has been revolutionized from targeting the tumor itself with traditional methods like surgery, chemotherapy, and radiotherapy to boosting the immune system coordinating with TIME to control cancers. Diverse approaches of immunotherapy including targeting cytokines, immune checkpoint blockade (ICB), and administration of depleting or agonistic antibodies have been used to enhance antitumor activities (27).

ILCs were first identified at the mucosal barriers, but more recent data confirm that ILCs exist in most, if not all, tissue types (75). ILCs are among the first immune cells to react to changes in environmental signaling and to shape TIME to coordinate the adaptive immune response, providing promising targets for cancer immunotherapy strategies.

Targeting Cytokines

As immunomodulators, ILCs produce their signature cytokines and respond to various cytokines produced from other cell types and the surrounding environments. Cytokine-based immunotherapies such as IL-2 family, IL-10, IL-12, TGF-β, and several other immunosuppressive cytokines are currently under ongoing clinical trials for cancer treatment (30, 162, 163). IL-2 is a key cytokine in promoting the expansion of cytotoxic lymphocytes. IL-2 receptors, CD25, and CD122 are expressed on mouse ILC2s, whereas human ILC2s and ILC3s show high expression of CD25 only. In response to IL-2 stimulation, ILCs can be expanded and activated to facilitate antitumor effects (128, 164). However, the systemic administration of this cytokine frequently causes grade 3 and 4 adverse effects owing to its toxic profile even at the recommended doses. Several secondgeneration IL-2-based compounds have been engineered to reduce the high-affinity binding to IL-2Ra/CD25 and increase the half-life in circulation (163). Directed conjugation of polyethylene glycol (PEGylation) generates an inactive cytokine with a long half-life in circulation. PEGylated IL-2 (NKTR-214) has been shown to preferentially activate CD122, and this modified cytokine is currently being evaluated in phase I/II clinical trials for the treatment of various tumors (27, 165). IL-15 and IL-15 super-agonist ALT803 exhibit capabilities to increase NK cell cytotoxicity in AML patients (166) and other tumor models such as advanced solid tumors (NCT01946789), multiple myeloma (NCT02099539), relapsed hematologic malignancy (NCT01885897) (27) and metastatic NSCLC (NCT02989844) (167). IL-10 is released by innate and adaptive immune cells to regulate pro-inflammatory cytokines activity. IL-10 exhibits a context-dependent outcome in cancer, and administration of PEGylated IL-10 increases its half-life to avoid grade 3-4 immune-related adverse effects. This feature has been evaluated in a phase I clinical trial in advanced, treatment-refractory tumors (NTC02009449) (163). The antitumor and protumor effects of TGF- β in cancer are based on the control of epithelial cell growth and promoting epithelial-mesenchymal transition (EMT), respectively. Although their efficacy as monotherapy agents is limited, combined blockade of TGF-B using small molecules or antagonistic monoclonal antibodies with immune checkpoint inhibitors such as PD-1/PD-L1 (NCT02423343 and NCT02734160) shows great potential for blocking the functions of this cytokine (163). TGF-β receptor type 1 (TGFBR1) inhibitors are currently used in phase I clinical trials to treat advanced solid tumors (NCT02160106) or in combination with pomalidomide for multiple myeloma (NCT03143985) (30). Because of the intimate links between cancer and inflammation, future cancer immunotherapies may benefit from repurposing of anti-inflammatory treatments that are already in human trials for chronic inflammatory diseases such as IBD. For instance, anti-IL-12 and IL-23 blocking antibody ustekinumab has been approved for clinical trials in Crohn's disease (168). Targeting

IL-17 or IFN- γ alone failed to show any effects in clinical trials, whereas targeting both provided favorable results in a preclinical model on IBD (77). Antibodies against IL-5 and anti-IL-4 receptor α subunit that block ILC2 function have provided encouraging results in patients with chronic rhinosinusitis (169). Similarly, inhibition of ILC2 function by chemoattractant receptor-homologous molecule expressed on Th2 cells (CTTH2) antagonists restores lung function in patients with asthma (170).

Cytokines are complex immune mediators with great potential in clinical cancer immunotherapy. The safety and effective use of cytokine-based drugs require a thorough knowledge of cytokine biology and advanced biotechnology in order to exploit their antitumor activity while minimizing their toxicity. In a combination with other immunomodulatory drugs such as immune checkpoint inhibitors and chimeric antigen receptor T cells, cytokine immunotherapy might become a more effective strategy in cancer treatment.

Immune Checkpoint Blockade

ILCs share some activating and inhibitory receptors with NK and T cells, suggesting potential targets for immunotherapeutic applications. ICB is an emerging antibody-based immunotherapy providing promising strategies for cancer treatment. Antibodies targeting immune checkpoints including CTLA-4, PD-1, and PD-L1 have now been approved and used in the clinics for diverse cancer treatments (171, 172). Activation of mouse ILCs shows upregulation of PD-1, whereas increased PD-1 expression on ILC2s and ILC3 has been found in human gastrointestinal tumors, and highly expressed PD-1 and CTLA-4 were shown in human breast cancer (133, 173). Strikingly, only a minority of treated patients have benefited from of PD-1 blockade, with some others displaying drug resistance. Therefore, understanding how to shape the TME in order to increase the sensitivity to PD-1 blockade becomes indispensable. The preclinical and clinical studies in TNBC have shown that low-dose chemotherapy with doxorubicin or cisplatin or irradiation induces a favorable TME that increases the sensitivity to nivolumab treatment for PD-1 blockade to stimulate an anticancer immune response (174).

Another immune checkpoint activation protein, inducible co-stimulator (ICOS) expressed on CD4+T cells as well as on ILC2s and ILC3s, was identified as a crucial player in CTLA-4 blockade antitumor effects. Engagement of the ICOS pathway markedly enhances the efficacy of CTLA-4 blockade in cancer immunotherapy in a mouse model of melanoma and prostate cancer (175). Currently, an anti-ICOS agonist antibody (GSK3359609) is being used in a phase I clinical trial aimed at patients with solid tumors. In addition, therapeutic antibodies against other immune checkpoints have also been developed, such as TIM-3, LAG-3 (expressed on ILC1s only), and TIGIT (expressed on both ILC1 and ILC3s) (176). Moreover, monalizumab, an antibody against NK cell checkpoint protein NKG2A, showed the capability to enhance antitumor immunity by promoting both NK and CD8⁺ T-cell function. Based on these findings, monalizumab in combination with cetuximab is being used in head and neck squamous cell carcinoma phase II clinical trial (177).

Depleting or Agonistic Antibody Application and Others

Both experimental mouse models and human samples showed that therapeutic targeting of ILC3 could be beneficial for autoimmune diseases. For example, depletion of ILCs by anti-Thy1 antibody improved the treatment for *Helicobacter hepaticus*-induced colitis (77). In the context of nonsolid tumors, human ILCs can be depleted before hematopoietic stem cell transplantation to treat acute leukemia (178).

Adenosine signaling belongs to a metabolic pathway that plays an important role in the immunosuppressive microenvironment favorable for cancer development. ILC1s express CD39 and CD73, activating the adenosine pathway in mouse salivary glands and increasing an immunosuppressive TIME for tumor growth, suggesting a potential target for therapeutic approaches (148).

So far, comprehensive information on the phenotypic and functional characterization of ILCs has been gathered. After the initial observation of the antitumor cytotoxicity of NK cells, other ILCs have become attractive targets for immunotherapy based on their abilities to rapidly sense and respond to signaling changes in the TME. The current challenge in cancer biology is to harness the cytokines, growth factors, and chemokines that regulate the specific function of ILCs within the TME, and further investigations will help to develop the exploitable targets for clinical applications.

CONCLUSIONS AND PERSPECTIVES

Despite advances in immunotherapies for cancer treatment, some patients still encounter few or no clinical benefits with the same treatments. Given the unique, non-redundant roles of ILCs within cancers, development of rational therapies that specifically target ILCs is likely to boost the immunotherapeutic effects in a larger group of cancer patients.

State-of-the-art technologies like single-cell RNA-sequencing (173) and high-dimensional mass spectrometry (CyTOF) analysis (179) have allowed us to investigate ILCPs, delineate distinct ILCs subsets and their developmental pathways, and identify novel mediators of antitumor immunity at single-cell resolution levels. These studies present new perspectives for exploring the complexity of ILCs in the diverse context of cancers and allow effective manipulation of ILC immunosurveillance to achieve optimal therapeutic opportunities.

AUTHOR CONTRIBUTIONS

ZA and TN conceived the manuscript. ZA reviewed the literature and wrote the manuscript. FF-B, SI, JD, and TN revised the manuscript.

ACKNOWLEDGMENTS

We thank Simon Broad for the critical reading. This work was supported by KCL Breast Cancer Now Research Unit. FF-B was funded by grant KCL-Q2-Y5 and SI was by CRUK (grant C56773/A24869).

REFERENCES

- Artis D, Spits H. The biology of innate lymphoid cells. Nature. (2015) 517:293-301. doi: 10.1038/nature14189
- Klose CSN, Flach M, Möhle L, Rogell L, Hoyler T, Ebert K, et al. Differentiation of type 1 ILCs from a common progenitor to all helper-like innate lymphoid cell lineages. *Cell.* (2014) 157:340–56. doi: 10.1016/j.cell.2014.03.030
- Spits H, Artis D, Colonna M, Diefenbach A, Di Santo JP, Eberl G, et al. Innate lymphoid cells-a proposal for uniform nomenclature. *Nat Rev Immunol.* (2013) 13:145–9. doi: 10.1038/nri3365
- Vivier E, Artis D, Colonna M, Diefenbach A, Di Santo JP, Eberl G, et al. Innate lymphoid cells: 10 years on. *Cell.* (2018) 174:1054–66. doi: 10.1016/j.cell.2018.07.017
- Constantinides MG, Mcdonald BD, Verhoef PA, Bendelac A. A committed precursor to innate lymphoid cells. *Nature*. (2014) 508:397–401. doi: 10.1038/nature13047
- Renoux VM, Zriwil A, Peitzsch C, Michaëlsson J, Friberg D, Soneji S, Sitnicka E. Identification of a human natural killer cell lineagerestricted progenitor in fetal and adult tissues. *Immunity*. (2015) 43:394–407. doi: 10.1016/j.immuni.2015.07.011
- Scoville SD, Mundy-Bosse BL, Zhang MH, Chen L, Zhang X, Keller KA, et al. A progenitor cell expressing transcription factor RORγt Generates all human innate lymphoid cell subsets. *Immunity*. (2016) 44:1140–50. doi: 10.1016/j.immuni.2016.04.007
- Withers DR. Lymphoid tissue inducer cells. Curr Biol. (2011) 21:R381–2. doi: 10.1016/j.cub.2011.03.022
- Cherrier M, Sawa S, Eberl G. Notch, Id2, and RORγt sequentially orchestrate the fetal development of lymphoid tissue inducer cells. *J Exp Med.* (2012) 209:729–40. doi: 10.1084/jem.20111594
- Satoh-Takayama N, Lesjean-Pottier S, Vieira P, Sawa S, Eberl G, Vosshenrich CAJ, Di Santo JP. IL-7 and IL-15 independently program the differentiation of intestinal CD3-NKp46+ cell subsets from Id2-dependent precursors. J Exp Med. (2010) 207:273–80. doi: 10.1084/jem.20092029
- Mebius RE. Organogenesis of lymphoid tissues. Nat Rev Immunol. (2003) 3:292–303. doi: 10.1038/nri1054
- Crellin NK, Trifari S, Kaplan CD, Cupedo T, Spits H. Human NKp44+IL-22+ cells and LTi-like cells constitute a stable RORC+ lineage distinct from conventional natural killer cells. J Exp Med. (2010) 207:281–90. doi: 10.1084/jem.20091509
- Walker JA, Clark PA, Crisp A, Haim-Vilmovsky L, Teichmann SA, Mckenzie ANJ. Polychromic reporter mice reveal unappreciated innate lymphoid cell progenitor heterogeneity and elusive ILC3 progenitors in bone marrow correspondence. *Immunity*. (2019) 51:104–18. doi: 10.1016/j.immuni.2019.05.002
- Seillet C, Mielke LA, Amann-Zalcenstein DB, Huntington ND, Carotta S, Belz GT, et al. Deciphering the innate lymphoid cell transcriptional program. *CellReports*. (2016) 17:436–47. doi: 10.1016/j.celrep.2016.09.025
- Xu W, Cherrier DE, Chea S, Vosshenrich C, Serafini N, Petit M, et al. An Id2RFP-reporter mouse redefines innate lymphoid cell precursor potentials. *Immunity*. (2019) 50:1054–68.e3. doi: 10.1016/j.immuni.2019.02.022
- Lu C-C, Wu T-S, Hsu Y-J, Chang C-J, Lin C-S, Chia J-H, et al. NK cells kill mycobacteria directly by releasing perforin and granulysin. *J Leukoc Biol.* (2014) 96:1119–29. doi: 10.1189/jlb.4A0713-363RR
- Neill DR, Wong SH, Bellosi A, Flynn RJ, Daly M, Langford TKA, et al. Nuocytes represent a new innate effector leukocyte that mediates type-2 immunity. *Nature*. (2010) 464:1367–70. doi: 10.1038/nature 08900
- Moro K, Kabata H, Tanabe M, Koga S, Takeno N, Mochizuki M, et al. Interferon and IL-27 antagonize the function of group 2 innate lymphoid cells and type 2 innate immune responses. *Nat Immunol.* (2016) 17:76–86. doi: 10.1038/ni.3309
- Monticelli LA, Osborne LC, Noti M, Tran SV, Zaiss DMW, Artis D. IL-33 promotes an innate immune pathway of intestinal tissue protection dependent on amphiregulin-EGFR interactions. *Proc Natl Acad Sci USA*. (2015) 112:10762–7. doi: 10.1073/pnas.1509070112
- 20. Willinger T. Metabolic control of innate lymphoid cell migration. *Front Immunol.* (2019) 10:10. doi: 10.3389/fimmu.2019.02010

- Sonnenberg GF, Hepworth MR. Functional interactions between innate lymphoid cells and adaptive immunity. *Nat Rev Immunol.* (2019) 19:599–613. doi: 10.1038/s41577-019-0194-8
- 22. Mebius RE, Rennert P, Weissman IL. Developing lymph nodes collect CD4+CD3- LT β + cells that can differentiate to APC, NK cells, and follicular cells but not T or B cells. *Immunity*. (1997) 7:493–504. doi: 10.1016/S1074-7613(00)80371-4
- Klose CSN, Kiss EA, Schwierzeck V, Ebert K, Hoyler T, Göppert N, et al. A T-bet gradient controls the fate and function of CCR6 2 RORct 1 innate lymphoid cells. *Nature*. (2013) 494:261–5. doi: 10.1038/nature11813
- Bando JK, Colonna M. Innate lymphoid cell function in the context of adaptive immunity. Nat Immunol. (2016) 17:783–9. doi: 10.1038/ni.3484
- Silver JS, Kearley J, Copenhaver AM, Sanden C, Mori M, Yu L, et al. Inflammatory triggers associated with exacerbations of COPD orchestrate plasticity of group 2 innate lymphoid cells in the lungs. *Nat Immunol.* (2016) 17:626–35. doi: 10.1038/ni.3443
- Wagner M, Koyasu S. Cancer immunoediting by innate lymphoid cells. Trends Immunol. (2019) 40:415–30. doi: 10.1016/j.it.2019.03.004
- Crinier A, Vivier E, Bléry M. Helper-like innate lymphoid cells and cancer immunotherapy. *Semin Immunol.* (2019) 2019:101274. doi: 10.1016/j.smim.2019.04.002
- Walker JA, Barlow JL, McKenzie ANJ. Innate lymphoid cells-how did we miss them? *Nat Rev Immunol.* (2013) 13:75–87. doi: 10.1038/nri3349
- Morvan MG, Lanier LL. NK cells and cancer: you can teach innate cells new tricks. *Nat Rev Cancer*. (2016) 16:7–19. doi: 10.1038/nrc.2015.5
- Chiossone L, Dumas PY, Vienne M, Vivier E. Natural killer cells and other innate lymphoid cells in cancer. *Nat Rev Immunol.* (2018) 18:671–88. doi: 10.1038/s41577-018-0061-z
- 31. Balkwill F, Mantovani A. Inflammation and cancer: back to Virchow? *Lancet*. (2001) 357:539–45. doi: 10.1016/S0140-6736(00)04046-0
- Pickup MW, Mouw JK, Weaver VM. The extracellular matrix modulates the hallmarks of cancer. *EMBO Rep.* (2014) 15:1243–53. doi: 10.15252/embr.201439246
- Binnewies M, Roberts EW, Kersten K, Chan V, Fearon DF, Merad M, et al. Understanding the tumor immune microenvironment (TIME) for effective therapy. *Nat Med.* (2018) 24:541–50. doi: 10.1038/s41591-018-0014-x
- Friedl P, Alexander S. Cancer invasion and the microenvironment: plasticity and reciprocity. *Cell.* (2011) 147:992–1009. doi: 10.1016/j.cell.2011.11.016
- Weis SM, Cheresh DA. Tumor angiogenesis: molecular pathways and therapeutic targets. *Nat Med.* (2011) 17:1359–70. doi: 10.1038/nm.2537
- Lotfi R, Lee JJ, Lotze MT. Eosinophilic granulocytes and damage-associated molecular pattern molecules (DAMPs). J Immunother. (2007) 30:16–28. doi: 10.1097/01.cji.0000211324.53396.f6
- Stamatiades EG, Li MO. Tissue-resident cytotoxic innate lymphoid cells in tumor immunosurveillance. *Semin Immunol.* (2019) 41:101269. doi: 10.1016/j.smim.2019.03.001
- Mattner J, Wirtz S. Friend or Foe? The ambiguous role of innate lymphoid cells in cancer development. *Trends Immunol.* (2017) 38:29–38. doi: 10.1016/j.it.2016.10.004
- Crome SQ, Nguyen LT, Lopez-Verges S, Yang SYC, Martin B, Yam JY, et al. A distinct innate lymphoid cell population regulates tumor-associated T cells. *Nat Med.* (2017) 23:368–75. doi: 10.1038/nm.4278
- Barry KC, Hsu J, Broz ML, Cueto FJ, Binnewies M, Combes AJ, et al. A natural killer-dendritic cell axis defines checkpoint therapyresponsive tumor microenvironments. *Nat Med.* (2018) 24:1178–91. doi: 10.1038/s41591-018-0085-8
- Flores-Borja F, Irshad S, Gordon P, Wong F, Sheriff I, Tutt A, et al. Crosstalk between innate lymphoid cells and other immune cells in the tumor microenvironment. *J Immunol Res.* (2016) 2016:1–14. doi: 10.1155/2016/7803091
- Tugues S, Ducimetiere L, Friebel E, Becher B. Innate lymphoid cells as regulators of the tumor microenvironment. *Semin Immunol.* (2019) 41:101270. doi: 10.1016/j.smim.2019.03.002
- Vallentin B, Barlogis V, Piperoglou C, Cypowyj S, Zucchini N, Chene M, et al. Innate lymphoid cells in cancer. *Cancer Immunol Res.* (2015) 3:1109–14. doi: 10.1158/2326-6066.CIR-15-0222
- 44. Coussens LM, Tinkle CL, Hanahan D, Werb Z. MMP-9 supplied by bone marrow-derived cells contributes to skin

carcinogenesis. Cell. (2000) 103:481-90. doi: 10.1016/S0092-8674(00)0 0139-2

- Mantovani A, Bottazzi B, Colotta F, Sozzani S, Ruco L. The origin and function of tumor-associated macrophages. *Immunol Today*. (1992) 13:265–70.
- Gasteiger G, Fan X, Dikiy S, Lee SY, Rudensky AY. Tissue residency of innate lymphoid cells in lymphoid and nonlymphoid organs. *Science*. (2015) 350:981–5. doi: 10.1126/science.aac9593
- Lu P, Weaver VM, Werb Z. The extracellular matrix: a dynamic niche in cancer progression. J Cell Biol. (2012) 196:395–406. doi: 10.1083/jcb.201102147
- Glasner A, Levi A, Enk J, Isaacson B, Viukov S, Orlanski S, et al. NKp46 receptor-mediated interferon-γ production by natural killer cells increases fibronectin 1 to alter tumor architecture and control metastasis. *Immunity*. (2018) 48:107–19.e4. doi: 10.1016/j.immuni.2017.12.007
- Zhang T, Wang J, Cao X, Liu S, Yang P, Han C, et al. Fibronectin maintains survival of mouse natural killer (NK) cells via CD11b/Src/β-catenin pathway. *Blood.* (2009) 114:4081–8. doi: 10.1182/blood-2009-05-219881
- Nussbaum K, Burkhard SH, Ohs I, Mair F, Klose CSN, Arnold SJ, et al. Tissue microenvironment dictates the fate and tumor-suppressive function of type 3 ILCs. J Exp Med. (2017) 214:2331–47. doi: 10.1084/jem.20162031
- Wang H, Shen L, Sun X, Liu F, Feng W, Jiang C, et al. Adipose group 1 innate lymphoid cells promote adipose tissue fibrosis and diabetes in obesity. *Nat Commun.* (2019) 10:1. doi: 10.1038/s41467-019-11270-1
- Mchedlidze T, Waldner M, Zopf S, Walker J, Rankin AL, Schuchmann M, et al. Interleukin-33-dependent innate lymphoid cells mediate hepatic fibrosis. *Immunity*. (2013) 39:357–71. doi: 10.1016/j.immuni.2013.07.018
- Monticelli LA, Sonnenberg GF, Abt MC, Alenghat T, Ziegler CGK, Doering TA, et al. Innate lymphoid cells promote lung-tissue homeostasis after infection with influenza virus. *Nat Immunol.* (2011) 12:1045–54. doi: 10.1038/ni.2131
- Richards CD. Innate immune cytokines, fibroblast phenotypes, and regulation of extracellular matrix in lung. J Interf Cytokine Res. (2017) 37:52–61. doi: 10.1089/jir.2016.0112
- Bruno A, Pagani A, Pulze L, Albini A, Dallaglio K, Noonan DM, et al. Orchestration of angiogenesis by immune cells. *Front Oncol.* (2014) 4:131. doi: 10.3389/fonc.2014.00131
- Stockmann C, Schadendorf D, Klose R, Helfrich I. The impact of the immune system on tumor: angiogenesis and vascular remodeling. *Front Oncol.* (2014) 4:69. doi: 10.3389/fonc.2014.00069
- De Palma M, Biziato D, Petrova TV. Microenvironmental regulation of tumour angiogenesis. *Nat Rev Cancer.* (2017) 17:457–74. doi: 10.1038/nrc.2017.51
- Bergers G, Brekken R, Mcmahon G, Vu TH, Itoh T, Tamaki K, Tanzawa K, Thorpe P, Itohara S, Werb Z, et al. Matrix metalloproteinase-9 triggers the angiogenic switch during carcinogenesis. *Nat Cell Biol.* (2000) 2:737–44. doi: 10.1038/35036374
- Albini A, Bruno A, Noonan DM, Mortara L. Contribution to tumor angiogenesis from innate immune cells within the tumor microenvironment: implications for immunotherapy. *Front Immunol.* (2018) 9:527. doi: 10.3389/fimmu.2018.00527
- Keskin DB, Allan DSJ, Rybalov B, Andzelm MM, Stern JNH, Kopcow HD, et al. TGFβ promotes conversion of CD16+ peripheral blood NK cells into CD16- NK cells with similarities to decidual NK cells. *Proc Natl Acad Sci* USA. (2007) 104:3378–83. doi: 10.1073/pnas.0611098104
- Bruno A, Focaccetti C, Pagani A, Imperatori AS, Spagnoletti M, Rotolo N, et al. The proangiogenic phenotype of natural killer cells in patients with non-small cell lung cancer. *Neoplasia*. (2013) 15:133–42. doi: 10.1593/neo.121758
- Gotthardt D, Putz EM, Grundschober E, Prchal-Murphy M, Straka E, Kudweis P, et al. STAT5 is a key regulator in NK cells and acts as a molecular switch from tumor surveillance to tumor promotion. *Cancer Discov.* (2016) 6:414–29. doi: 10.1158/2159-8290.CD-15-0732
- Gao Y, Souza-Fonseca-Guimaraes F, Bald T, Ng SS, Young A, Ngiow SF, et al. Tumor immunoevasion by the conversion of effector NK cells into type 1 innate lymphoid cells. *Nat Immunol.* (2017) 18:1004–15. doi: 10.1038/ni.3800
- 64. Sgadari C, Angiolillo AL, Tosato G. Inhibition of angiogenesis by interleukin-12 is mediated by the interferon-inducible protein 10.

Blood. (1996) 87:3877–82. doi: 10.1182/blood.V87.9.3877.bloodjournal87 93877

- Beatty GL, Paterson Y. IFN-γ-Dependent inhibition of tumor angiogenesis by tumor-infiltrating CD4 + T cells requires tumor responsiveness to IFN-γ. J Immunol. (2001) 166:2276–82. doi: 10.4049/jimmunol.166. 4.2276
- 66. Bromberg JF, Horvath CM, Wen Z, Schreiber RD, Darnell JE. Transcriptionally active Stat1 is required for the antiproliferative effects of both interferon α and interferon γ. Proc Natl Acad Sci USA. (1996) 93:7673–8. doi: 10.1073/pnas.93.15.7673
- Choi Y-S, Choi H-J, Min J-K, Pyun B-J, Maeng Y-S, Park H, Kim J, et al. Interleukin-33 induces angiogenesis and vascular permeability through ST2/TRAF6-mediated endothelial nitric oxide production. *Nat Immunol.* (2009) 114:3117–26. doi: 10.1182/blood-2009-02-203372
- Rak GD, Osborne LC, Siracusa MC, Kim BS, Wang K, Bayat A, et al. IL-33dependent group 2 innate lymphoid cells promote cutaneous wound healing. *J Invest Dermatol.* (2016) 136:487–96. doi: 10.1038/JID.2015.406
- Numasaki M, Fukushi JI, Ono M, Narula SK, Zavodny PJ, Kudo T, et al. Interleukin-17 promotes angiogenesis and tumor growth. *Blood.* (2003) 101:2620–7. doi: 10.1182/blood-2002-05-1461
- He D, Li H, Yusuf N, Elmets CA, Li J, Mountz JD, et al. IL-17 promotes tumor development through the induction of tumor promoting microenvironments at tumor sites and myeloid-derived suppressor cells. J Immunol. (2010) 184:2281–8. doi: 10.4049/jimmunol.0902574
- Li Q, Liu L, Zhang Q, Liu S, Ge D, You Z. Interleukin-17 indirectly promotes M2 macrophage differentiation through stimulation of COX-2/PGE2 pathway in the cancer cells. *Cancer Res Treat.* (2014) 46:297–306. doi: 10.4143/crt.2014.46.3.297
- 72. Eisenring M, Vom Berg J, Kristiansen G, Saller E, Becher B. IL-12 initiates tumor rejection via lymphoid tissue-inducer cells bearing the natural cytotoxicity receptor NKp46. *Nat Immunol.* (2010) 11:1030–8. doi: 10.1038/ni.1947
- Ansel KM, Ngo VN, Hyman PL, Luther SA, Förster R, Sedgwick JD, et al. A chemokine-driven positive feedback loop organizes lymphoid follicles. *Nature*. (2000) 406:309–14. doi: 10.1038/35018581
- 74. Irshad S, Flores-Borja F, Lawler K, Monypenny J, Evans R, Male V, et al. RORγt+ innate lymphoid cells promote lymph node metastasis of breast cancers. *Cancer Res.* (2017) 77:1083–96. doi: 10.1158/0008-5472.CAN-16-0598
- Klose CSN, Artis D. Innate lymphoid cells as regulators of immunity, inflammation and tissue homeostasis. *Nat Immunol.* (2016) 17:765–74. doi: 10.1038/ni.3489
- 76. Viant C, Girard-Madoux MJH, Vivier E, Rankin LC, Seillet C, Shi W, et al. Transforming growth factor-β and Notch ligands act as opposing environmental cues in regulating the plasticity of type 3 innate lymphoid cells. *Sci Signal.* (2016) 9:ra46. doi: 10.1126/scisignal.aaf2176
- Buonocore S, Ahern PP, Uhlig HH, Ivanov II, Littman DR, Maloy KJ, et al. Innate lymphoid cells drive interleukin-23-dependent innate intestinal pathology. *Nature*. (2010) 464:1371–5. doi: 10.1038/nature08949
- Almeida FF, Belz GT. Innate lymphoid cells: models of plasticity for immune homeostasis and rapid responsiveness in protection. *Mucosal Immunol.* (2016) 9:1103–12. doi: 10.1038/mi.2016.64
- van Niel G, Raposo G. Shedding light on the cell biology of extracellular vesicles. Nat Publ Gr. (2018) 19:213–28. doi: 10.1038/nrm.2017.125
- Li I, Nabet BY. Exosomes in the tumor microenvironment as mediators of cancer therapy resistance. *Mol Cancer*. (2019) 18:32. doi: 10.1186/s12943-019-0975-5
- Zhang Q, Higginbotham JN, Jeppesen DK, Franklin JL, Bellis SL, Coffey RJ, et al. Transfer of functional cargo in exomeres article transfer of functional cargo in exomeres. *CellReports*. (2019) 27:940–54.e6. doi: 10.1016/j.celrep.2019.01.009
- Halim TYF, Hwang YY, Scanlon ST, Zaghouani H, Garbi N, Fallon PG, Mckenzie ANJ. Group 2 innate lymphoid cells license dendritic cells to potentiate memory TH2 cell responses. *Nat Immunol.* (2016) 17:57–64. doi: 10.1038/ni.3294
- 83. Gabitass RF, Annels NE, Stocken DD, Pandha HA, Middleton GW. Elevated myeloid-derived suppressor cells in pancreatic, esophageal and gastric cancer are an independent prognostic factor and are associated with significant

elevation of the Th2 cytokine interleukin-13. *Cancer Immunol Immunother*. (2011) 60:1419–30. doi: 10.1007/s00262-011-1028-0

- Halim TYF, Rana BMJ, Walker JA, Kerscher B, Knolle MD, Jolin HE, et al. Tissue-restricted adaptive type 2 immunity is orchestrated by expression of the costimulatory molecule OX40L on group 2 innate lymphoid cells. *Immunity*. (2018) 48:1195–207.e6. doi: 10.1016/j.immuni.2018.05.003
- Zaiss DMW, van Loosdregt J, Gorlani A, Bekker CPJ, Gröne A, Sibilia M, et al. Amphiregulin enhances regulatory T cell-suppressive function via the epidermal growth factor receptor. *Immunity*. (2013) 38:275–84. doi: 10.1016/j.immuni.2012.09.023
- Pelly VS, Kannan Y, Coomes SM, Entwistle LJ, Rückerl D, Seddon B, et al. IL-4-producing ILC2s are required for the differentiation of TH2 cells following Heligmosomoides polygyrus infection. *Mucosal Immunol.* (2016) 9:1407–17. doi: 10.1038/mi.2016.4
- Purwar R, Schlapbach C, Xiao S, Kang HS, Elyaman W, Jiang X, et al. Robust tumor immunity to melanoma mediated by interleukin-9-producing T cells. *Nat Med.* (2012) 18:1248–53. doi: 10.1038/nm.2856
- Kim IK, Kim BS, Koh CH, Seok JW, Park JS, Shin KS, et al. Glucocorticoidinduced tumor necrosis factor receptor-related protein co-stimulation facilitates tumor regression by inducing IL-9-producing helper T cells. *Nat Med.* (2015) 21:1010–7. doi: 10.1038/nm.3922
- Lange K, Uckert W, Blankenstein T, Nadrowitz R, Bittner C, Renauld JC, et al. Overexpression of NPM-ALK induces different types of malignant lymphomas in IL-9 transgenic mice. *Oncogene*. (2003) 22:517–27. doi: 10.1038/sj.onc.1206076
- Withers DR, Gaspal FM, Mackley EC, Marriott CL, Ross EA, Desanti GE, et al. Cutting edge: lymphoid tissue inducer cells maintain memory CD4 T cells within secondary lymphoid tissue. *J Immunol.* (2012) 189:2094–8. doi: 10.4049/jimmunol.1201639
- Masur SH, Kelsen JR, Laufer TM, Baldassano RN, Dubrot J, Eberl G, et al. Group 3 innate lymphoid cells mediate intestinal selection of commensal bacteria-specific CD4+ T cells. *Science*. (2015) 348:1031–5. doi: 10.1126/science.aaa4812
- Melo-Gonzalez F, Kammoun H, Evren E, Dutton EE, Papadopoulou M, Bradford BM, et al. Antigen-presenting ILC3 regulate T cell-dependent IgA responses to colonic mucosal bacteria. J Exp Med. (2019) 216:728–42. doi: 10.1084/jem.20180871
- Mackley EC, Houston S, Marriott CL, Halford EE, Lucas B, Cerovic V, et al. CCR7-dependent trafficking of RORγ+ ILCs creates a unique microenvironment within mucosal draining lymph nodes. *Nat Commun.* (2015) 6:5862. doi: 10.1038/ncomms6862
- Hepworth MR, Monticelli LA, Fung TC, Ziegler CGK, Grunberg S, Sinha R, et al. Innate lymphoid cells regulate CD4+ T-cell responses to intestinal commensal bacteria. *Nature*. (2013) 498:113–17. doi: 10.1038/nature12240
- Mortha A, Chudnovskiy A, Hashimoto D, Bogunovic M, Spencer SP, Belkaid Y, et al. Microbiota-dependent crosstalk between macrophages and ILC3 promotes intestinal homeostasis. *Science.* (2014) 343:1249288. doi: 10.1126/science.1249288
- 96. von Burg N, Chappaz S, Baerenwaldt A, Horvath E, Bose Dasgupta S, Ashok D, et al. Activated group 3 innate lymphoid cells promote T-cellmediated immune responses. *Proc Natl Acad Sci USA*. (2014) 111:12835–40. doi: 10.1073/pnas.1406908111
- Kruglov AA, Grivennikov SI, Kuprash DV, Winsauer C, Prepens S, Seleznik GM, et al. Nonredundant function of soluble ltα3 produced by innate lymphoid cells in intestinal homeostasis. *Science*. (2013) 342:1243–6. doi: 10.1126/science.1243364.
- Sano T, Hall JA, Bonneau R, Chen A, Miraldi ER, Littman DR, et al. An IL-23R/IL-22 circuit regulates epithelial serum amyloid a to promote local effector Th17 responses. *Cell.* (2016) 164:324. doi: 10.1016/j.cell.2015.12.047
- Castellanos JG, Longman RS. The balance of power: innate lymphoid cells in tissue inflammation and repair. J Clin Invest. (2019) 129:2640–50. doi: 10.1172/JCI124617
- Grivennikov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. Cell. (2010) 140:883–99. doi: 10.1016/j.cell.2010.01.025
- Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. Nature. (2008) 454:436–44. doi: 10.1038/nature07205
- 102. Simoni Y, Fehlings M, Kløverpris HN, McGovern N, Koo SL, Loh CY, et al. Human innate lymphoid cell subsets possess tissue-type based

heterogeneity in phenotype and frequency. *Immunity*. (2017) 46:148-61. doi: 10.1016/j.immuni.2016.11.005

- 103. Bernink JH, Peters CP, Munneke M, Te Velde AA, Meijer SL, Weijer K, et al. Human type 1 innate lymphoid cells accumulate in inflamed mucosal tissues. *Nat Immunol.* (2013) 14:221–9. doi: 10.1038/ni.2534
- 104. Moro K, Yamada T, Tanabe M, Takeuchi T, Ikawa T, Kawamoto H, et al. Innate production of T H 2 cytokines by adipose tissue-associated c-Kit + Sca-1 + lymphoid cells. *Nature*. (2010) 463:540–4. doi: 10.1038/nature08636
- 105. Cella M, Fuchs A, Vermi W, Facchetti F, Otero K, Lennerz JKM, et al. A human natural killer cell subset provides an innate source of IL-22 for mucosal immunity. *Nature*. (2009) 457:722–5. doi: 10.1038/nature07537
- 106. Gladiator A, Wangler N, Trautwein-Weidner K, LeibundGut-Landmann S. Cutting edge: IL-17-secreting innate lymphoid cells are essential for host defense against fungal infection. *J Immunol.* (2013) 190:521–5. doi: 10.4049/jimmunol.1202924
- 107. Gray J, Oehrle K, Worthen G, Alenghat T, Whitsett J, Deshmukh H. Intestinal commensal bacteria mediate lung mucosal immunity and promote resistance of newborn mice to infection. *Sci Transl Med.* (2017) 9:eaaf9412. doi: 10.1126/scitranslmed.aaf9412
- 108. Castellanos JG, Woo V, Viladomiu M, Putzel G, Lima S, Diehl GE, et al. Microbiota-induced TNF-like ligand 1A drives group 3 innate lymphoid cellmediated barrier protection and intestinal T cell activation during colitis. *Immunity*. (2018) 49:1077–89.e5. doi: 10.1016/j.immuni.2018.10.014
- 109. Scandella E, Bolinger B, Lattmann E, Miller S, Favre S, Littman DR, et al. Restoration of lymphoid organ integrity through the interaction of lymphoid tissue-inducer cells with stroma of the T cell zone. *Nat Immunol.* (2008) 9:667–75. doi: 10.1038/ni.1605
- 110. Karta MR, Broide DH, Doherty TA. Insights into group 2 innate lymphoid cells in human airway disease. Curr Allergy Asthma Rep. (2016) 16:8. doi: 10.1007/s11882-015-0581-6
- 111. Chang YJ, Kim HY, Albacker LA, Baumgarth N, McKenzie ANJ, Smith DE, et al. Innate lymphoid cells mediate influenza-induced airway hyperreactivity independently of adaptive immunity. *Nat Immunol.* (2011) 12:631–8. doi: 10.1038/ni.2045
- 112. Hams E, Armstrong ME, Barlow JL, Saunders SP, Schwartz C, Cooke G, et al. IL-25 and type 2 innate lymphoid cells induce pulmonary fibrosis. *Proc Natl Acad Sci USA*. (2014) 111:367–72. doi: 10.1073/pnas.1315854111
- 113. Geremia A, Arancibia-Cárcamo CV, Fleming MPP, Rust N, Singh B, Mortensen NJ, et al. IL-23–responsive innate lymphoid cells are increased in inflammatory bowel disease. J Exp Med. (2011) 208:1127–33. doi: 10.1084/jem.20101712
- 114. Wang S, Xia P, Chen Y, Qu Y, Xiong Z, Ye B, et al. Regulatory innate lymphoid cells control innate intestinal inflammation. *Cell.* (2017) 171:201–16.e18. doi: 10.1016/j.cell.2017.07.027
- Tugues S, Burkhard SH, Ohs I, Vrohlings M, Nussbaum K, vom Berg J, et al. New insights into IL-12-mediated tumor suppression. *Cell Death Differ*. (2015) 22:237–46. doi: 10.1038/cdd.2014.134
- 116. Dadi S, Chhangawala S, Whitlock BM, Franklin RA, Luo CT, Oh SA, et al. Cancer immunosurveillance by tissue-resident innate lymphoid cells and innate-like t cells. *Cell*. (2016) 164:365–77. doi: 10.1016/j.cell.2016.01.002
- 117. De Weerdt I, Van Hoeven V, Marius Munneke J, Endstra S, Hofland T, et al. Innate lymphoid cells are expanded and functionally altered in chronic lymphocytic leukemia. *Haematologica*. (2016) 101:e461-4. doi: 10.3324/haematol.2016.144725
- 118. Trabanelli S, Curti A, Lecciso M, Salomé B, Riether C, Ochsenbein A, et al. CD127+ innate lymphoid cells are dysregulated in treatment naïve acute myeloid leukemia patients at diagnosis. *Haematologica*. (2015) 100:e257– 60. ddoi: 10.3324/haematol.2014.119602
- Long A, Dominguez D, Qin L, Chen S, Fan J, Zhang M, et al. Type 2 innate lymphoid cells impede IL-33–mediated tumor suppression. J Immunol. (2018) 201:3456–64. doi: 10.4049/jimmunol.1800173
- Fournié JJ, Poupot M. The pro-tumorigenic IL-33 involved in antitumor immunity: a yin and yang cytokine. Front Immunol. (2018) 9:506. doi: 10.3389/fimmu.2018.02506
- 121. Molofsky AB, Van Gool F, Liang HE, Van Dyken SJ, Nussbaum JC, Lee J, et al. InterleuKin-33 and interferon-Γ counter-regulate group 2 innate lymphoid cell activation during immune perturbation. *Immunity*. (2015) 43:161–74. doi: 10.1016/j.immuni.2015.05.019

- 122. Forkel M, VanTol S, Höög C, Michaëlsson J, Almer S, Mjösberg J. Distinct alterations in the composition of mucosal innate lymphoid cells in newly diagnosed and established Crohn's disease and ulcerative colitis. J Crohn's Colitis. (2019) 13:67–78. doi: 10.1093/ecco-jcc/jjy119
- 123. Trabanelli S, Chevalier MF, Martinez-Usatorre A, Gomez-Cadena A, Salomé B, Lecciso M, et al. Tumour-derived PGD2 and NKp30-B7H6 engagement drives an immunosuppressive ILC2-MDSC axis. *Nat Commun.* (2017) 8:593. doi: 10.1038/s41467-017-00678-2
- Bruchard M, Ghiringhelli F. Deciphering the roles of innate lymphoid cells in cancer. *Front Immunol.* (2019) 10:656. doi: 10.3389/fimmu.2019. 00656
- 125. Chevalier MF, Trabanelli S, Racle J, Salomé B, Cesson V, Gharbi D, et al. ILC2-modulated T cell-to-MDSC balance is associated with bladder cancer recurrence. J Clin Invest. (2017) 127:2916–29. doi: 10.1172/JCI89717
- 126. Bie Q, Zhang P, Su Z, Zheng D, Ying X, Wu Y, et al. Polarization of ILC2s in peripheral blood might contribute to immunosuppressive microenvironment in patients with gastric cancer. J Immunol Res. (2014) 2014:923135. doi: 10.1155/2014/923135
- 127. Carrega P, Loiacono F, Di Carlo E, Scaramuccia A, Mora M, Conte R, et al. NCR + ILC3 concentrate in human lung cancer and associate with intratumoral lymphoid structures. *Nat Commun.* (2015) 6:8280. doi: 10.1038/ncomms9280
- Robinette ML, Bando JK, Song W, Ulland TK, Gilfillan S, Colonna M. IL-15 sustains IL-7R-independent ILC2 and ILC3 development. *Nat Commun.* (2017) 8. doi: 10.1038/ncomms14601
- 129. Liu Y, Song Y, Lin D, Lei L, Mei Y, Jin Z, et al. NCR group 3 innate lymphoid cells orchestrate IL-23/IL-17 axis to promote hepatocellular carcinoma development. *EBioMedicine*. (2019) 41:333–44. doi: 10.1016/j.ebiom.2019.02.050
- 130. Punt S, Fleuren GJ, Kritikou E, Lubberts E, Trimbos JB, Jordanova ES, Gorter A. Angels and demons: Th17 cells represent a beneficial response, while neutrophil IL-17 is associated with poor prognosis in squamous cervical cancer. *Oncoimmunology.* (2015) 4:984539. doi: 10.4161/2162402X.2014.984539
- 131. Roan F, Stoklasek TA, Whalen E, Molitor JA, Bluestone JA, Buckner JH, et al. CD4 + Group 1 Innate Lymphoid Cells (ILC) form a functionally distinct ILC subset that is increased in systemic sclerosis. *J Immunol.* (2016) 196:2051–62. doi: 10.4049/jimmunol.1501491
- 132. Kodama T, Takeda K, Shimozato O, Hayakawa Y, Atsuta M, Kobayashi K, et al. Perforin-dependent NK cell cytotoxicity is sufficient for anti-metastatic effect of IL-12. *Eur J Immunol.* (1999) 29:1390–6.
- 133. Salimi M, Wang R, Yao X, Li X, Wang X, Hu Y, et al. Activated innate lymphoid cell populations accumulate in human tumour tissues. *BMC Cancer*. (2018) 18:341. doi: 10.1186/s12885-018-4262-4
- Ikutani M, Yanagibashi T, Ogasawara M, Tsuneyama K, Yamamoto S, Hattori Y, et al. Identification of innate IL-5–producing cells and their role in lung eosinophil regulation and antitumor immunity. *J Immunol.* (2012) 188:703–13. doi: 10.4049/jimmunol.1101270
- Kim J, Kim W, Moon UJ, Kim HJ, Choi H-J, Sin J-I, et al. Intratumorally establishing type 2 innate lymphoid cells blocks tumor growth. *J Immunol.* (2016) 196:2410–23. doi: 10.4049/jimmunol.1501730
- 136. Jovanovic IP, Pejnovic NN, Radosavljevic GD, Pantic JM, Milovanovic MZ, Arsenijevic NN, et al. Interleukin-33/ST2 axis promotes breast cancer growth and metastases by facilitating intratumoral accumulation of immunosuppressive and innate lymphoid cells. *Int J Cancer.* (2014) 134:1669–82. doi: 10.1002/ijc.28481
- 137. Paclik D, Stehle C, Lahmann A, Hutloff A, Romagnani C. ICOS regulates the pool of group 2 innate lymphoid cells under homeostatic and inflammatory conditions in mice. *Eur J Immunol.* (2015) 45:2766–72. doi: 10.1002/eji.201545635
- Busser B, Sancey L, Brambilla E, Coll JL, Hurbin A. The multiple roles of amphiregulin in human cancer. *Biochim Biophys Acta Rev Cancer*. (2011) 1816:119–31. doi: 10.1016/j.bbcan.2011.05.003
- Liew FY, Girard JP, Turnquist HR. Interleukin-33 in health and disease. Nat Rev Immunol. (2016) 16:676–89. doi: 10.1038/nri.2016.95
- 140. Hallett MA, Venmar KT, Fingleton B. Cytokine stimulation of epithelial cancer cells: the similar and divergent functions of IL-4 and IL-13. *Cancer Res.* (2012) 72:6338–43. doi: 10.1158/0008-5472.CAN-12-3544

- 141. Kirchberger S, Royston DJ, Boulard O, Thornton E, Franchini F, Szabady RL, et al. Innate lymphoid cells sustain colon cancer through production of interleukin-22 in a mouse model. J Exp Med. (2013) 210:917–31. doi: 10.1084/jem.20122308
- 142. Sonnenberg GF, Monticelli LA, Elloso MM, Fouser LA, Artis D. CD4+ lymphoid tissue-inducer cells promote innate immunity in the gut. *Immunity*. (2011) 34:122–34. doi: 10.1016/j.immuni.2010.12.009
- 143. Colonna M. Innate lymphoid cells: diversity, plasticity, and unique functions in immunity. *Immunity*. (2018) 48:1104–17. doi: 10.1016/j.immuni.2018.05.013
- Crome SQ, Ohashi PS. Immunoregulatory functions of innate lymphoid cells. J Immunother Cancer. (2018) 6:121. doi: 10.1186/s40425-018-0433-8
- 145. Cortez VS, Ulland TK, Cervantes-Barragan L, Bando JK, Robinette ML, Wang Q, et al. SMAD4 impedes the conversion of NK cells into ILC1like cells by curtailing non-canonical TGF-β signaling. *Nat Immunol.* (2017) 18:995–1003. doi: 10.1038/ni.3809
- 146. Park E, Patel S, Wang Q, Andhey P, Zaitsev K, Porter S, et al. Toxoplasma gondii infection drives conversion of NK cells into ILC1-like cells. *Elife.* (2019) 8:e47605. doi: 10.7554/eLife.47605.024
- 147. Hughes T, Briercheck EL, Freud AG, Trotta R, McClory S, Scoville SD, et al. The transcription factor AHR prevents the differentiation of a stage 3 innate lymphoid cell subset to natural killer cells. *Cell Rep.* (2014) 8:150–62. doi: 10.1016/j.celrep.2014.05.042
- 148. Cortez VS, Cervantes-Barragan L, Robinette ML, Bando JK, Wang Y, Geiger TL, et al. Transforming growth Factor-β signaling guides the differentiation of innate lymphoid cells in Salivary Glands. *Immunity*. (2016) 44:1127–39. doi: 10.1016/j.immuni.2016.03.007
- 149. Bal SM, Bernink JH, Nagasawa M, Groot J, Shikhagaie MM, Golebski K, et al. IL-1β, IL-4 and IL-12 control the fate of group 2 innate lymphoid cells in human airway inflammation in the lungs. *Nat Immunol.* (2016) 17:636–45. doi: 10.1038/ni.3444
- 150. Bernink JH, Krabbendam L, Germar K, de Jong E, Gronke K, Kofoed-Nielsen M, et al. Interleukin-12 and-23 control plasticity of Cd127+ group 1 and group 3 innate lymphoid cells in the intestinal lamina propria. *Immunity*. (2015) 43:146–60. doi: 10.1016/j.immuni.2015.06.019
- 151. Chan IH, Jain R, Tessmer MS, Gorman D, Mangadu R, Sathe M, et al. Interleukin-23 is sufficient to induce rapid de novo gut tumorigenesis, independent of carcinogens, through activation of innate lymphoid cells. *Mucosal Immunol.* (2014) 7:842–56. doi: 10.1038/mi.2013.101
- Lim AI, Menegatti S, Bustamante J, Le Bourhis L, Allez M, et al. IL-12 drives functional plasticity of human group 2 innate lymphoid cells. *J Exp Med.* (2016) 213:569–83. doi: 10.1084/jem.20151750
- Ohne Y, Silver JS, Thompson-Snipes LA, Collet MA, Blanck JP, Cantarel BL, et al. IL-1 is a critical regulator of group 2 innate lymphoid cell function and plasticity. *Nat Immunol.* (2016) 17:646–55. doi: 10.1038/ni.3447
- Huang Y, Guo L, Qiu J, Chen X, Hu-Li J, Siebenlist U, et al. IL-25-responsive, lineage-negative KLRG1 hi cells are multipotential "inflammatory" type 2 innate lymphoid cells. *Nat Immunol.* (2015) 16:161–9. doi: 10.1038/ni.3078
- 155. Raykova A, Carrega P, Lehmann FM, Ivanek R, Landtwing V, Quast I, et al. Interleukins 12 and 15 induce cytotoxicity and early NK-cell differentiation in type 3 innate lymphoid cells. *Blood Adv.* (2017) 1:2679–91. doi: 10.1182/bloodadvances.2017008839
- 156. Imai K, Matsuyama S, Miyake S, Suga K, Nakachi K. Natural cytotoxic activity of peripheral-blood lymphocytes and cancer incidence: an 11year follow-up study of a general population. *Lancet.* (2000) 356:1795–9. doi: 10.1016/S0140-6736(00)03231-1
- 157. Mikami Y, Scarno G, Zitti B, Shih HY, Kanno Y, Santoni A, et al. NCR+ ILC3 maintain larger STAT4 reservoir via T-BET to regulate type 1 features upon IL-23 stimulation in mice. *Eur J Immunol.* (2018) 48:1174–80. doi: 10.1002/eji.201847480
- 158. Cella M, Gamini R, Sécca C, Collins PL, Zhao S, Peng V, et al. Subsets of ILC3–ILC1-like cells generate a diversity spectrum of innate lymphoid cells in human mucosal tissues. *Nat Immunol.* (2019) 20:980–91. doi: 10.1038/s41590-019-0425-
- 159. Crellin NK, Trifari S, Kaplan CD, Satoh-Takayama N, Di Santo JP, Spits H. Regulation of cytokine secretion in human CD127+ LTi-like innate lymphoid cells by toll-like receptor 2. *Immunity*. (2010) 33:752–64. doi: 10.1016/j.immuni.2010.10.012

- Shih HY, Sciumè G, Mikami Y, Guo L, Sun HW, Brooks SR, et al. Developmental acquisition of regulomes underlies innate lymphoid cell functionality. *Cell.* (2016) 165:1120–33. doi: 10.1016/j.cell.2016.04.029
- Koues OI, Collins PL, Cella M, Robinette ML, Porter SI, Pyfrom SC, et al. Distinct gene regulatory pathways for human innate versus adaptive lymphoid cells. *Cell.* (2016) 165:1134–46. doi: 10.1016/j.cell.2016.04.014
- Chiossone L, Vienne M, Kerdiles YM, Vivier E. Natural killer cell immunotherapies against cancer: checkpoint inhibitors and more. *Semin Immunol.* (2017) 31:55–63. doi: 10.1016/j.smim.2017.08.003
- 163. Berraondo P, Sanmamed MF, Ochoa MC, Etxeberria I, Aznar MA, Luis Pérez-Gracia J, et al. Immunology cytokines in clinical cancer immunotherapy. *Br J Cancer*. (2018) 120:6–15. doi: 10.1038/s41416-018-0328-y
- Roediger B, Kyle R, Yip KH, Sumaria N, Guy TV., Kim BS, et al. Cutaneous immunosurveillance and regulation of inflammation by group 2 innate lymphoid cells. *Nat Immunol.* (2013) 14:564–73. doi: 10.1038/ni.2584
- 165. Charych DH, Hoch U, Langowski JL, Lee SR, Addepalli MK, Kirk PB, et al. NKTR-214, an engineered cytokine with biased IL2 receptor binding, increased tumor exposure, and marked efficacy in mouse tumor models. *Clin Cancer Res.* (2016) 22:680–90. doi: 10.1158/1078-0432.CCR-15-1631
- 166. Szczepanski MJ, Szajnik M, Welsh A, Foon KA, Whiteside TL, Boyiadzis M. Interleukin-15 enhances natural killer cell cytotoxicity in patients with acute myeloid leukemia by upregulating the activating NK cell receptors. *Cancer Immunol Immunother*. (2010) 59:73–9. doi: 10.1007/s00262-009-0724-5
- 167. Wrangle JM, Velcheti V, Patel MR, Garrett-Mayer E, Hill EG, Ravenel JG, et al. ALT-803, an IL-15 superagonist, in combination with nivolumab in patients with metastatic non-small cell lung cancer: a non-randomised, open-label, phase 1b trial. *Lancet Oncol.* (2018) 19:694–704. doi: 10.1016/S1470-2045(18)30148-7
- 168. Moschen AR, Tilg H, Raine T. IL-12, IL-23 and IL-17 in IBD: immunobiology and therapeutic targeting. Nat Rev Gastroenterol Hepatol. (2019) 16:185–96. doi: 10.1038/s41575-018-0084-8
- 169. Okubo K, Hashiguchi K, Takeda T, Baba K, Kitagoh H, Miho H, et al. A randomized controlled phase II clinical trial comparing ONO-4053, a novel DP1 antagonist, with a leukotriene receptor antagonist pranlukast in patients with seasonal allergic rhinitis. *Allergy Eur J Allergy Clin Immunol.* (2017) 72:1565–75. doi: 10.1111/all.13174
- Singh D, Ravi A, Southworth T. CRTH2 antagonists in asthma: current perspectives. *Clin Pharmacol Adv Appl.* (2017) 9:165–73. doi: 10.2147/CPAA.S119295
- Park IJ. Selection of adjuvant treatment without neoadjuvant chemoradiotherapy for patients with rectal cancer: Room for further investigation. Ann Coloproctol. (2018) 34:109–10. doi: 10.3393/ac.2018.05.27

- Hargadon KM, Johnson CE, Williams CJ. Immune checkpoint blockade therapy for cancer: an overview of FDA-approved immune checkpoint inhibitors. *Int Immunopharmacol.* (2018) 62:29–39. doi: 10.1016/j.intimp.2018.06.001
- 173. Yu Y, Tsang JCH, Wang C, Clare S, Wang J, Chen X, et al. Single-cell RNAseq identifies a PD-1hi ILC progenitor and defines its development pathway. *Nature*. (2016) 539:102–6. doi: 10.1038/nature20105
- 174. Voorwerk L, Slagter M, Horlings HM, Sikorska K, van de Vijver KK, de Maaker M, Nederlof I, Kluin RJC, Warren S, Ong SF, et al. Immune induction strategies in metastatic triple-negative breast cancer to enhance the sensitivity to PD-1 blockade: the TONIC trial. *Nat Med.* (2019) 25:920–8. doi: 10.1038/s41591-019-0432-4
- 175. Fan X, Quezada SA, Sepulveda MA, Sharma P, Allison JP. Engagement of the ICOS pathway markedly enhances efficacy of CTLA-4 blockade in cancer immunotherapy. *J Exp Med.* (2014) 211:715–25. doi: 10.1084/jem.20 130590
- 176. Burugu S, Dancsok AR, Nielsen TO. Emerging targets in cancer immunotherapy. Semin Cancer Biol. (2018) 52:39–52. doi: 10.1016/j.semcancer.2017.10.001
- 177. André P, Denis C, Soulas C, Bourbon-Caillet C, Lopez J, Arnoux T, et al. Anti-NKG2A mAb is a checkpoint inhibitor that promotes anti-tumor immunity by unleashing both T and NK cells. *Cell.* (2018) 175:1731–43.e13. doi: 10.1016/j.cell.2018.10.014
- 178. Marius Munneke J, Björklund AT, Mjösberg JM, Garming-Legert K, Bernink JH, Blom B, et al. Activated innate lymphoid cells are associated with a reduced susceptibility to graft-versus-host disease. *Blood*. (2014) 124:812–21. doi: 10.1182/blood-2013-11-536888
- 179. de Vries NL, van Unen V, Ijsselsteijn ME, Abdelaal T, van der Breggen R, Farina Sarasqueta A, et al. High-dimensional cytometric analysis of colorectal cancer reveals novel mediators of antitumour immunity. *Gut.* (2019). doi: 10.1136/gutjnl-2019-318672

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 An, Flores-Borja, Irshad, Deng and Ng. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.