



Current Insights in Cutaneous Lupus Erythematosus Immunopathogenesis

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Cutaneous Lupus Erythematosus (CLE) is a clinically diverse group of autoimmune skin diseases with shared histological features of interface dermatitis and autoantibodies deposited at the dermal–epidermal junction. Various genetic and environmental triggers of CLE promote infiltration of T cells, B cells, neutrophils, antigen presenting cells, and NK cells into lesional skin. In this mini-review, we will discuss the clinical features of CLE, insights into CLE immunopathogenesis, and novel treatment approaches.

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Cutaneous Lupus Erythematosus (CLE) is an autoimmune disease primarily affecting skin and mucosal tissue. Total CLE disease incidence is \sim 4.3 per 100,000 (1, 2). CLE exhibits a strong sex bias toward females much like systemic lupus erythematosus (SLE) (3–5). Certain CLE subtypes may progress to SLE (6, 7), which can have health consequences, including kidney and brain involvement leading to renal failure and neurologic disease (8). Successful treatment of cutaneous disease may significantly decrease the risk of systemic involvement (9). Treatment options for CLE are limited, with anti-malarials as the most commonly prescribed drugs, followed by calcineurin inhibitors, mycophenolate mofetil, methotrexate, and steroids (10–13).

CLINICAL FEATURES OF CLE

CLE encompasses a heterogeneous group of photodermatoses with varying degrees of association with systemic disease (SLE) [reviewed in (14)]. It is typically classified into three main subtypes based on the disease chronicity, clinical morphology and distribution: acute (ACLE), subacute (SCLE), and chronic (CCLE) (15, 16). All CLE subtypes are characterized histopathologically by interface dermatitis (with the exception of tumid lupus and lupus panniculitis) and lupus band reaction, which consist of infiltration of immune cells and deposition of autoantibodies at the dermal-epidermal junction (DEJ) (17).

ACLE presents as transient erythematous patches that correspond to flares in SLE patients. A well-known example of ACLE is the malar rash, or butterfly rash, that classically crosses both cheeks but spares the nasolabial folds. This helps to distinguish it from other clinical mimickers such as rosacea and seborrheic dermatitis. ACLE can affect the entire body in some patients with bad flares, and is considered a criterion for the diagnosis of SLE. Up to 80% of SLE patients experience malar rash, which typically flares with UV exposure but does not leave any scar or dyspigmentation. Histologically, ACLE manifests as lymphoplasmacytic interface dermatitis with vacuolar changes at the DEJ associated with mucin deposition.

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Lesions of SCLE last longer than the malar rash of ACLE, but systemic lupus occurs in a significantly lower percentage of the affected individuals. SCLE most often occurs on the photo-exposed areas of the upper chest, back, and external upper extremities (**Figure 1**). When active, it typically presents as papulosquamous lesions and/or annular plaques (psorasiform) with central clearing and raised erythematous scaly edges. Upon resolution, SCLE can leave dyspigmentation but no permanent scarring. SCLE is highly associated with anti-Ro/SSA autoantibodies. In patients with new-onset SCLE, it is important to carefully review medications, as SCLE can be induced by nonsteroidal anti-inflammatory drugs (NSAIDs), proton pump inhibitors, antihypertensives, and antifungals (18).

CCLE has multiple subtypes, and the most common is discoid LE (DLE). DLE commonly presents as red, atrophic, and hypopigmented plaques with a characteristic rim of hyperpigmentation. DLE typically only affects areas above the neck such as scalp, ears, nose, and cheeks (localized DLE). Presence of DLE is one of the diagnostic criteria of SLE and is observed in 20% of the patients with systemic disease; however, only 5-10% of cases with isolated DLE eventually progress to develop systemic disease (4). When it occurs on the scalp, DLE can cause irreversible scarring alopecia (19). Though rare, DLE may affect larger areas of the body including trunk and upper extremities (generalized DLE), which confers an increased risk of systemic involvement. DLE exhibits more prominent histopathological findings including follicular plugging, periadnexal lymphoplasmacytic infiltrate, and pigment incontinence. Other variants of CCLE, including tumid lupus, chilblain lupus, and lupus panniculitis, are significantly less frequent.

AUTOANTIBODIES/AUTOANTIGENS

Like SLE, many CLE patients develop autoantibodies including anti-nuclear antibodies (ANAs). Antibodies against Ro/SSA and La/SSB are detected frequently and have been associated with SCLE and neonatal lupus erythematosus (NLE), in addition to SLE and Sjogren's Syndrome (20-24). Most CLE patients exhibit anti-Ro/SSA autoantibody reactivity patterns (25). Ro refers to ribonucleoproteins that are encoded by two separate gene products, resulting in 52 kDa (also known as TRIM21) and 60 kDa (also known as TROVE2) protein isotypes (26). CLE patients react more frequently to the 60 kDa form than the 52 kDa form (27). Anti-Ro autoantibodies may also be mechanistically involved in CLE pathogenesis, as commensal bacteria that produce a Ro60 kDa ortholog can trigger lupus development in mice (28). Infusion of anti-Ro/SSA into human skin grafted mice results in lupus band reaction similar to what is observed in CLE specimens (29).

Other autoantigens in CLE include SSB/La, ribonucleoprotein, smith (sm) antigen (30), C1q (31), and HMGB1 (32). UV damage can induce translocation of these autoantigens to the surface of keratinocytes, thereby making them bioavailable to the immune system (33–36). HMGB1 appears to play a significant role in the development of CLE:

HMGB1 is highly expressed in the epidermis of CLE skin biopsies (37) and expression level correlates with clinically active photoinduced CLE lesions (38, 39).

A recent study in a cohort of Italian CLE patients found strong correlations between autoantibody specificities and CLE subtypes (40). CCLE is negatively associated with anti-extractable nuclear antigens (ENA), anti-Ro/SSA, and anti-dsDNA. SCLE positively correlates with ENA, anti-Ro/SSA, anti-Smith, and anti-RNP. ACLE is strongly associated with anti-dsDNA and ANA, though this may be due to the finding that these autoantibodies are found in higher frequencies in females and SLE patients.

NLE is a condition characterized by cutaneous, cardiac, and multi-systemic abnormalities observed in 5–16% of newborn infants whose mothers have autoantibodies against Ro/SSA, La/SSB, and U1-ribonucleoprotein (41–44), regardless of whether the mothers are symptomatic or not (45). Autoantibodies against Ro/SSA and La/SSB were detected in 98% of affected infants (45). Anti-Ro52/60-kDA Ro/SSA and 48-kD La/SSB auto-antibodies contribute to heart block (46), whereas 50-kD La/SSB are associated with cutaneous disease (47, 48), which is thought to self-resolve but can have long-term cutaneous changes (49). A study of 186 antibody exposed fetuses and infants indicates a direct correlation between the amount of maternal anti-Ro and anti-La antibodies and fetal tissue injury (48).

CLE IMMUNOPATHOGENESIS

Factors Contributing to Onset

Like many complex diseases, CLE is thought to arise from a combination of genetics and environment. Several immune genes have been implicated in subtypes of CLE, including cytokine genes, complement genes, and innate immune genes [reviewed in (50)]. Polymorphisms in the transcription factor IRF5, the signaling molecule TYK2, and the immune regulator CTLA4 were identified in a Finnish cohort of CLE patients (51). Familial chilblain lupus, a rare form of CLE characterized by acral lesions, is caused by gain-of-function mutations in the DNA sensor protein STING (52, 53), or mutations that decrease the exonuclease activity of TREX1 (54–56).

CLE and cutaneous involvement in SLE can be induced by UV radiation, and photosensitivity is a criterion used by the American College of Rheumatology for lupus diagnosis [reviewed in (57, 58)]. The pathogenic wavelengths of UV radiation remain unclear. However, UVB is considered an instigating factor, as the dosage required to induce erythema is 1,000 fold less than that of UVA in lupus patients (59). Lupus keratinocytes are more sensitive to UV light than healthy keratinocytes (60), and exhibit aberrant apoptosis thereby generating cellular debris and activating the immune system (57). Some groups have reported impaired clearance and accumulation of apoptotic keratinocyte debris in afflicted skin (61), while others have found inflammatory clearance with no evidence of impaired clearance (62). Regardless of clearance efficacy, it is clear that UV damaged, apoptotic keratinocytes are one of the main instigating factors in CLE lesion formation. A recent review by Wolf et al. (63) summarized known aspects of UV-induced CLE.



FIGURE 1 | Posterior view of the trunk in a Hispanic patient with Cutaneous Lupus Erythematosus (CLE). (A) Arrows point to two lupus patches on the upper back and lower back. (B) Closer view of the upper back patch in the intrascapular area showing scaling, erythema, dyspigmentation, and scarring.

Over 100 drugs have been identified that induce CLE or CLE flares (18, 64–66), with SCLE being the most common clinical subtype induced by prescribed medications such as antihypertensives and antifungals (67). The blood pressure medication hydrochlorothorazide induces SCLE in a photosensitive manner (68). The chemotherapeutic 5-fluorouracil can cause SCLE or DLE (69, 70). Proton pump inhibitors such as omeprazole are triggers of SCLE (71, 72). There are a few case reports of CLE resulting after anti-TNF therapy, though the etiology is not fully understood (73).

Smoking is a trigger of CLE (74), particularly DLE (75), and reduces responsiveness to anti-malarial treatments (76, 77). Counseling patients to quit smoking can significantly improve CLE and other autoimmune conditions (77–80). Case reports demonstrate environmental or occupational exposures can also induce CLE, such as silica exposure (81, 82). Lastly, microbes may trigger CLE: recent studies found *Staphylococcus aureus* can colonize skin following IFN-mediated barrier disruption (83), and is enriched in CLE lesional skin (83).

Sensing Damage: Innate Immune Cells

UVB radiation and/or drugs cause keratinocyte damage and death, which is sensed by the innate immune system to create a feed-forward loop driving pathogenesis. Keratinocytes themselves can respond to TLR-independent nucleic acid ligands via MDA5, RIG-I, c-GAS and STING, and can activate the AIM2 inflammasome, which can initiate the interferon response [reviewed in (84)]. Thus, they can respond to bystander damage to alert the immune system.

Langerhans cells are specialized dendritic cell (DC) populations that live in the epidermis, and migrate to skin draining lymph nodes upon antigen encounter (85). UVB-induced keratinocyte damage is sensed by Langerhans cells in both lupus-prone [MRL/lpr and B6.SLE1yaa (86)] and wild type mice (87). Shipman et al. (86) demonstrated that ADAM17 is upregulated in Langerhans cells following UVB exposure, which in turn increases conversion of EGFR ligands into an active

form in an attempt to protect keratinocytes from further UVB damage. In murine SLE models, Langerhans cells have a reduced ability to process EGFR ligands into an active form, resulting in a dysfunctional LC-KC axis in CLE lesions. LCs are subsequently replaced by other inflammatory DC subsets, thereby promoting further inflammation (88, 89) (**Figure 2A**). These data are supported by a recent microarray study of CLE biopsies that demonstrated decreased EGFR signaling pathways (90).

One particularly important inflammatory DC subset contributing to CLE pathogenesis is plasmacytoid DCs (pDCs). pDCs are common in DLE, and are used by dermatopathologists to assist in diagnosis (91). When presented with DNA, pDCs potently upregulate Type-1 IFN, mainly IFN α . pDCs are a key source of Type-1 IFN in lupus lesional skin (92, 93), and UVB promotes their recruitment to the skin (94). A first-in-human study of BDCA2 antibody (BIIB059), which targets pDCs, for SLE decreased expression of IFN response genes in blood, normalized MxA expression, reduced immune infiltrates in skin lesions, and decreased CLASI-A score [(95), NCT02847598]. BIIB059 is now in a phase-2 clinical trial for SLE and active CLE treatment (NCT02847598).

In addition to apoptotic keratinocytes, another potential source of DNA that could activate pDCs in the skin is from neutrophils. Some neutrophils have the ability to produce neutrophil extracellular traps (NETs), which are comprised of DNA, chromatin and various proteins. NETs have been found in various CLE subtypes including: lupus panniculitis, ACLE, DLE, and to a lesser extent, SCLE (96). Though the study by Safi et al. (96) included a cohort of only 30 patients, their work indicates the presence and contribution of NETs in CLE is worth further investigation. A subclass of neutrophils, called low density granulocytes (LDGs), have an increased propensity for producing NETs. LDGs have been reported in the skin of SLE patients (97). LDGs NETs provide a source of autoantigens, and may interact with nucleic acids from UV-B damaged KCs. Subsequent accumulation of apoptotic DNA provides a potential mechanism by which skin lesions are initiated or sustained (98) (Figure 2B).



Like DC populations, macrophages and monocytes are also involved in debris clearance and sensing of DAMPs. Immunohistochemical studies demonstrated that CD68⁺ macrophages express FasL and are densely populated near hair follicles in CLE lesions (99). Inflammasome activity in blood monocytes from SLE patients is enhanced via type I IFN-mediated upregulation of IRF1 (100), though the functional capacity of macrophages in CLE has not been well-studied. A trial of macrophage colony-stimulating factor (MCSF) antibody failed to reduce immune infiltrates or activation in CLE lesions and did not improve CLASI score (101). Thus, it is possible that tissue macrophages in CLE lesions perform an immune regulatory function, require different cytokines for their function/survival, or are dispensable for CLE.

Another aspect of innate immune involvement in lupus is the uptake and processing of cellular debris for both clearance and presentation of autoantigens. The scavenger receptor C1q binds to keratinocyte apoptotic blebs to assist their clearance (102). A silent single nucleotide polymorphism (SNP) in C1QA gene (Gly70_{GGG/GGA}) results in lower serum C1q and is associated with SCLE (103).

Promoting Damage: Lymphocytic Infiltrates

Inflammatory infiltrates in CLE are comprised mainly of T lymphocytes, with other infiltrating cells including B cells/plasma

cells, NK cells, dendritic cells (104, 105), and in some subtypes, neutrophils (106), implicating these populations as key drivers of inflammation in CLE lesions. The T cell specificities in CLE skin are unknown, but SLE studies identified T cells reactive to nucleosomes/histones (107), which can induce anti-dsDNA antibody production (108). TCR-VB38 and VB13 were enriched in skin of CCLE patients, implicating oligoclonal expansion (109). Several studies have observed expression of cytotoxic markers characteristic of T cell function, such as Th1-related cytokines and granzyme B (110, 111). Interestingly, the perform promoter (112), CD70 promoter (113), and other loci (114) are hypomethylated in CD4⁺ T cells from SCLE patients, making them poised to transcribe effector molecules. Both CD4⁺ and CD8⁺ circulating T cells have higher HLA-DR expression in CLE patients, and higher CD25 in DLE patients specifically, compared to healthy controls (115). Tregs are lower (116, 117), and $\gamma\delta$ T cells are higher (118), in CLE lesions. Skin-infiltrating CD4⁺ T cells express FasL have the potential to ligate Fas and can induce apoptosis in keratinocytes and other infiltrating immune cells (99).

Plasma cells in lupus skin lesions are often observed as "clockface" cells (119). It is not clear whether lupus band reactions arise from local production of autoantibodies in the skin by plasma cells, or if they deposit in the skin from the circulation. Interestingly, while B cells are required for development of lupus in mice (120, 121), autoantibody formation

appears to be a byproduct of lupus immune responses and not a driving factor: MRL-lpr mice with B cells incapable of secreting immunoglobulin develop nephritis, implicating B cells' antigen presenting function as being critical for lupus development (122). It is unclear which B cell functions are required for CLE, though preclinical studies using chimeric antigen receptor T cells (CAR-Ts) directed against B cells ameliorated skin disease in lupus-prone mice (123). Furthermore, B cell depletion with rituximab showed efficacy in a case study of 4 lupus panniculitis patients with childhood onset who were refractory to other standard treatments (124), and ameliorated skin symptoms in a retrospective case study of 14 consecutive SLE, one CCLE and two SCLE patients with recalcitrant skin involvement (125).

The atypical lymphocyte marker CD38 (ADP-ribosyl cyclase/cyclic ADP-ribose hydrolase) is highly expressed in CLE lesional skin. CD38 was recently shown to be important for Tfh-B cell collaboration in response to recurrent influenza vaccination (126). Polymorphisms in intron 1 of CD38 are associated with the development of DLE in a Spanish patient population (127). It is unclear what role CD38 plays in CLE pathogenesis, though knocking CD38 out of MRL-lpr mice accelerates lupus (128).

NK cells have been studied in peripheral blood from SLE patients, though their precise roles in CLE and skin are not known due to a paucity of mechanistic studies. In the SLE studies, blood NK numbers decrease with increased lupus disease activity and/or exhibit defects in traditional killing functions (129-131); though rare populations are often expanded and secrete higher levels of IFNy compared to healthy controls (132-135). Given their roles in sensing cellular stress (136), clearing tumors (137, 138), and keratinocyte-derived tumors in particular (139), it is likely that NK cells are also able to kill stressed keratinocytes following UV or drug injury to promote CLE lesions. It is also possible that T cells expressing killer receptors contribute to keratinocyte death in CLE, as exemplified by NKG2D ligation on mouse dendritic epidermal T cells (DETCs): in the absence of TCR signaling, NKG2D ligation on DETCs induces IFNy production and causes keratinocyte cytotoxicity (140). In line with this, one study found invariant NK T cells (iNKTs) were enriched in SCLE and DLE patient blood by flow cytometry and in lesional skin by immunohistochemistry (141): they expressed IFNγ *in situ*, and were Ki67+, indicating they were proliferating in skin lesions.

Chemokine Recruitment of Leukocytes to CLE Lesions

Type 1 IFN and type 2 IFN signaling stimulates expression of CXCL9, CXCL10, and CXCL11 which mediate leukocyte migration to peripheral tissue via CXCR3. IFN and CXCR3 have been postulated to drive the pathogenesis of all subtypes CLE (142), mediating the recruitment of the aforementioned immune cell types and providing positive feedback loops for T cell and pDC recruitment. Previous studies reported CXCR3 ligand expression in the skin in all the different subtypes of CLE (57, 58, 104, 143). Further, UV light induces upregulation of CXCR3 ligands in keratinocytes, linking the environmental insult to the recruitment of pathogenic immune cells (58). Type 1 IFN also induces CXCL13 (144), which can support germinal center formation through migration of Tfh cells and B cells [reviewed in (145)]. While CXCL13 serves as a biomarker of SLE but not CLE (146), epidermal injury can accelerate nephritis in NZM2328 mice via upregulation of CXCL13 (147). The receptor for CXCL13, CXCR5, can influence B cell function by enhancing antigen uptake via membrane ruffling and LFA-1mediated adhesion, and integrating BCR signaling in motile cells (148). The precise role of CXCL13 in CLE is not known.

CCL17 is expressed by keratinocytes in CLE lesions and has been hypothesized to recruit $CD8^+$ T cells bearing the cognate receptor CCR4 to the skin in scarring CLE (149). iNKT cells in CLE lesions also express CCR4, and blood iNKTs express higher levels of CCR4 and CCR6 than healthy controls (141). Higher CCR5 and lower CCR3 expression on peripheral CD4+ T cells is associated with higher disease activity in CLE (150).

Gene expression analysis of human DLE and SCLE skin biopsies, as well as a mouse model of CLE, exhibited increased CCL3, CCL4, CCL7, and CCL8 (151). While the precise roles of these chemokines in CLE have yet to be elucidated, inferences can be made based on their previously established biological activities. Future studies will need to be conducted to determine how these chemokines guide specific immune populations to and through the skin during CLE.

Paracrine Signals: Cytokines, Hormones, and Master Regulators

The inflammatory signatures of CLE are interferon (IFN)-based: a recent microarray study of 90 CLE biopsies found increased IFN pathways in all CLE subtypes, and DLE samples had a unique IFN γ node (90). Interestingly, the authors found no differences between skin biopsies of patients with and without systemic involvement. Blocking IFN α receptor improved CLASI scores for SLE patients with cutaneous involvement in a phase II trial [(152, 153), reviewed in (154)]; however IFN γ blockade did not improve CLASI scores in DLE patients (155). Single cell resolution may be necessary to elucidate the precise roles of different IFNs in pathogenesis.

Keratinocytes from CLE patients exhibit an enhanced response to both type-1 and type-2 IFNs (156), and produce IFN κ (157), IL-6 (158), and Type-III IFN (IFN λ) following UVB damage. Cytokine dysregulation in UVB treated CLE keratinocytes provides a link between UV initiation factors and immunopathogenesis (156). UV treatment of keratinocytes induces upregulation of IFN κ , which plays a key homeostatic role in maintaining IFN balance in skin (158). Lupus keratinocytes derived from active CLE lesions and nonlesional skin constitutively overexpress IFN κ , which increases photosensitivity via plasmacytoid DC production of IFN κ in response to IFN κ signaling (157).

Higher levels of Th1 and Th17 cells in CLE lesions have been reported, along with IL-21 (159). IL-21 activates pDCs to produce granzyme B (160), thereby enhancing keratinocyte killing by NK cells. Interestingly, Salvi et al. (159) found that type I IFN served as a negative regulatory loop for granzyme B production by pDCs. This implies that high levels of type I IFN in skin are not pathogenic, but rather represent an attempt to counter inflammation.

The contribution of female sex hormones in CLE remains unclear. One epidemiological study noted premenstrual and perimenopausal flares (161), indicating that a certain level of estrogen is protective. Nevertheless, the addition of estrogen to keratinocyte cultures doubles Ro/SSA surface expression following UVB exposure (162) and enhances binding of anti-Ro/SSA and La/SSB autoantibodies to the plasma membrane (163). A patient who received estrogen as part of sex reassignment surgery developed tumid lupus following UV exposure (164). Estrogen can positively regulate the IFN γ promoter (165) and NK cell activity (166), possibly explaining other roles in CLE pathogenesis.

Vgll3 is a transcription co-factor that governs expression of inflammatory genes associated with autoimmunity, including CLE (167). Vgll3 is more highly expressed in female skin and in lupus patient skin regardless of gender. Overexpression of Vgll3 in male mice makes skin appear more "female-like" and promotes both skin and systemic autoimmune attack (168).

CONCLUSION

The complexity of CLE has made it difficult to fully elucidate pathogenesis, with genetic and environmental triggers causing both innate and adaptive immune activation that creates diverse clinical manifestations. Nevertheless, increased knowledge in this field has paved the wave for promising new drugs in CLE. Blocking IFNAR antibody (Anifrolumab) improved CLASI scores in a phase IIb double-blind trial (>50% improvement, p = 0.013) (152, 153). Rituximab B cell depletion showed efficacy in case reports of CLE subtypes and SLE with skin involvement (124). The JAK/STAT pathways mediate signaling for a myriad of cytokines, including IFN, and other biological processes. JAK inhibition with tofacitinib (JAK3>JAK1>JAK2), baricitinib (JAK1/2) or ruxolitinib (JAK1/2) showed efficacy for familial chilblain lupus in human case studies and small clinical trials (53, 169-171). However, filgotinib (JAK1) did not reach its primary endpoint in a phase II double blind study, underscoring the need for a better understanding of which JAKs and STATs drive CLE (172-174). Similarly, the roles of IL-12 and IL-23 need to be revisited, as ustekinumab has been reported to both treat (175-178) and cause CLE (179).

REFERENCES

- Hejazi EZ, Werth VP. Cutaneous lupus erythematosus: an update on pathogenesis, diagnosis and treatment. Am J Clin Dermatol. (2016) 17:135– 46. doi: 10.1007/s40257-016-0173-9
- Durosaro O, Davis MDP, Reed KB, Rohlinger AL. Incidence of cutaneous lupus erythematosus, 1965-2005: a population-based study. Arch Dermatol. (2009) 145:249–53. doi: 10.1001/archdermatol.2009.21
- Lockshin MD, Mary Kirkland Center for Lupus Research Consortium. Biology of the sex and age distribution of systemic lupus erythematosus. *Arthritis Rheum.* (2007) 57:608–11. doi: 10.1002/art.22676
- 4. Merola JF, Prystowsky SD, Iversen C, Gomez-Puerta JA, Norton T, Tsao P, et al. Association of discoid lupus erythematosus with other clinical

Another approach to treat CLE is to enhance Treg function. The immune-dampening peptide Edratide, which stimulates Tregs, was safe in a phase II trial for SLE and improved BILAG score, but did not meet its primary endpoint (180). CAR-Tregs, which have been shown to be efficacious in a mouse model of asthma (181), may provide another approach, though they have yet to be tested for lupus (182). Of note, a challenge that remains in all CLE and lupus trials is the fact that patients are maintained on immunosuppressive drugs to keep their autoimmunity in check, which may have the unintended consequence of masking true efficacy. Ultimately, novel immunotherapies will need to be tested and developed for treatment of all CLE subtypes.

ETHICS STATEMENT

Written informed consent was obtained from the individual for the publication of any potentially identifiable images or data included in this article.

AUTHOR CONTRIBUTIONS

MAR: patient photos and consent and figure generation. JR: artwork/figure generation. MR: clinical section. ZR-O: innate immune genes/receptors of cellular debris section. PN: autoantibodies/autoantigens section. CG and JR: all other sections. All authors approved the final manuscript. All authors contributed to the article and approved the submitted version.

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manifestations among patients with systemic lupus erythematosus. *J Am Acad Dermatol.* (2013) 69:19–24. doi: 10.1016/j.jaad.2013. 02.010

- Wahie S, Daly AK, Cordell HJ, Goodfield MJ, Jones SK, Lovell CR, et al. Clinical and pharmacogenetic influences on response to hydroxychloroquine in discoid lupus erythematosus: a retrospective cohort study. J Invest Dermatol. (2011) 131:1981–6. doi: 10.1038/jid.2011.167
- Chong BF. Understanding how cutaneous lupus erythematosus progresses to systemic lupus erythematosus. *JAMA Dermatol.* (2014) 150:296. doi: 10.1001/jamadermatol.2013.9030
- Kirchhof MG, Dutz JP. The immunopathology of cutaneous lupus erythematosus. *Rheum Dis Clin North Am.* (2014) 40:455–74. doi: 10.1016/j.rdc.2014.04.006

- Yu C, Gershwin ME, Chang C. Diagnostic criteria for systemic lupus erythematosus: a critical review. J Autoimmun. (2014) 48–9:10–3. doi: 10.1016/j.jaut.2014.01.004
- Stannard JN, Kahlenberg JM. Cutaneous lupus erythematosus: updates on pathogenesis and associations with systemic lupus. *Curr Opin Rheumatol.* (2016) 28:453–9. doi: 10.1097/BOR.00000000000308
- Schultz HY, Dutz JP, Furukawa F, Goodfield MJ, Kuhn A, Lee LA, et al. From pathogenesis, epidemiology, and genetics to definitions, diagnosis, and treatments of cutaneous lupus erythematosus and dermatomyositis: a report from the 3rd International Conference on Cutaneous Lupus Erythematosus (ICCLE) 2013. J Invest Dermatol. (2015) 135:7–12. doi: 10.1038/jid.2014.316
- Reich A, Werth VP, Furukawa F, Kuhn A, Szczech J, Samotij D, et al. Treatment of cutaneous lupus erythematosus: current practice variations. *Lupus*. (2016) 25:964–72. doi: 10.1177/0961203316628997
- 12. Chang AY, Werth VP. Treatment of cutaneous lupus. *Curr Rheumatol Rep.* (2011) 13:300–7. doi: 10.1007/s11926-011-0180-z
- Kuhn A, Ruland V, Bonsmann G. Cutaneous lupus erythematosus: update of therapeutic options part II. J Am Acad Dermatol. (2011) 65:e195– 213. doi: 10.1016/j.jaad.2010.06.017
- 14. Werth VP. Clinical manifestations of cutaneous lupus erythematosus. *Autoimmun Rev.* (2005) 4:296–302. doi: 10.1016/j.autrev.2005.01.003
- Obermoser G, Sontheimer RD, Zelger B. Overview of common, rare and atypical manifestations of cutaneous lupus erythematosus and histopathological correlates. *Lupus.* (2010) 19:1050–70. doi: 10.1177/0961203310370048
- Okon LG, Werth VP. Cutaneous lupus erythematosus: diagnosis and treatment. *Best Pract Res Clin Rheumatol.* (2013) 27:391–404. doi: 10.1016/j.berh.2013.07.008
- Bennion SD, Middleton MH, David-Bajar KM, Brice S, Norris DA. In three types of interface dermatitis, different patterns of expression of intercellular adhesion molecule-1 (ICAM-1) indicate different triggers of disease. *J Invest Dermatol.* (1995) 105:71S-9S. doi: 10.1111/1523-1747.ep12316107
- Michaelis TC, Sontheimer RD, Lowe GC. An update in druginduced subacute cutaneous lupus erythematosus. *Dermatol Online J.* (2017) 23:13030/qt55x42822.
- Wilson CL, Burge SM, Dean D, Dawber RP. Scarring alopecia in discoid lupus erythematosus. Br J Dermatol. (1992) 126:307– 14. doi: 10.1111/j.1365-2133.1992.tb00670.x
- Chan EK, Andrade LE. Antinuclear antibodies in Sjögren's syndrome. *Rheum Dis Clin North Am.* (1992) 18:551–70.
- García-Carrasco M, Fuentes-Alexandro S, Escárcega RO, Salgado G, Riebeling C, Cervera R. Pathophysiology of Sjögren's syndrome. Arch Med Res. (2006) 37:921–32. doi: 10.1016/j.arcmed.2006.08.002
- Kobayashi R, Mii S, Nakano T, Harada H, Eto H. Neonatal lupus erythematosus in Japan: a review of the literature. *Autoimmun Rev.* (2009) 8:462-6. doi: 10.1016/j.autrev.2008.12.013
- Sontheimer RD, Maddison PJ, Reichlin M, Jordon RE, Stastny P, Gilliam JN. Serologic and HLA associations in subacute cutaneous lupus erythematosus, a clinical subset of lupus erythematosus. *Ann Intern Med.* (1982) 97:664– 71. doi: 10.7326/0003-4819-97-5-664
- 24. Wasicek CA, Reichlin M. Clinical and serological differences between systemic lupus erythematosus patients with antibodies to Ro versus patients with antibodies to Ro and La. *J Clin Invest.* (1982) 69:835–43. doi: 10.1172/JCI110523
- 25. Patsinakidis N, Gambichler T, Lahner N, Moellenhoff K, Kreuter A. Cutaneous characteristics and association with antinuclear antibodies in 402 patients with different subtypes of lupus erythematosus. J Eur Acad Dermatol Venereol. (2016) 30:2097–104. doi: 10.1111/jdv.13769
- Itoh K, Itoh Y, Frank MB. Protein heterogeneity in the human Ro/SSA ribonucleoproteins. The 52- and 60-kD Ro/SSA autoantigens are encoded by separate genes. J Clin Invest. (1991) 87:177–86. doi: 10.1172/ JCI114968
- 27. Popovic K, Brauner S, Ek M, Wahren-Herlenius M, Nyberg F. Fine specificity of the Ro/SSA autoantibody response in relation to serological and clinical findings in 96 patients with self-reported cutaneous symptoms induced by the sun. *Lupus.* (2007) 16:10–7. doi: 10.1177/0961203306073135
- Greiling TM, Dehner C, Chen X, Hughes K, Iñiguez AJ, Boccitto M, et al. Commensal orthologs of the human autoantigen Ro60

as triggers of autoimmunity in lupus. *Sci Transl Med.* (2018) 10:eaan2306. doi: 10.1126/scitranslmed.aan2306

- Lee LA, Gaither KK, Coulter SN, Norris DA, Harley JB. Pattern of cutaneous immunoglobulin G deposition in subacute cutaneous lupus erythematosus is reproduced by infusing purified anti-Ro (SSA) autoantibodies into human skin-grafted mice. J Clin Invest. (1989) 83:1556–62. doi: 10.1172/JCI114052
- Vera-Recabarren MA, García-Carrasco M, Ramos-Casals M, Herrero C. Cutaneous lupus erythematosus: clinical and immunological study of 308 patients stratified by gender. *Clin Exp Dermatol.* (2010) 35:729– 35. doi: 10.1111/j.1365-2230.2009.03764.x
- Bălănescu E, Tănăsescu C, Bălănescu P, Olteanu R, Badea C, Petrutescu B, et al. Anti C1q antibodies in cutaneous lupus erythematosus. *Rom J Intern Med.* (2010) 48:159–63.
- Hayashi A, Nagafuchi H, Ito I, Hirota K, Yoshida M, Ozaki S. Lupus antibodies to the HMGB1 chromosomal protein: epitope mapping and association with disease activity. *Mod Rheumatol.* (2009) 19:283– 92. doi: 10.3109/s10165-009-0151-7
- 33. Lawley W, Doherty A, Denniss S, Chauhan D, Pruijn G, van Venrooij WJ, et al. Rapid lupus autoantigen relocalization and reactive oxygen species accumulation following ultraviolet irradiation of human keratinocytes. *Rheumatology*. (2000) 39:253–61. doi: 10.1093/rheumatology/39.3.253
- Caricchio R, McPhie L, Cohen PL. Ultraviolet B radiationinduced cell death: critical role of ultraviolet dose in inflammation and lupus autoantigen redistribution. J Immunol. (2003) 171:5778–86. doi: 10.4049/jimmunol.171.11.5778
- Abdulahad DA, Westra J, Limburg PC, Kallenberg CGM, Bijl M. HMGB1 in systemic lupus erythematosus: its role in cutaneous lesions development. *Autoimmun Rev.* (2010) 9:661–5. doi: 10.1016/j.autrev.2010.05.015
- Abdulahad DA, Westra J, Reefman E, Zuidersma E, Bijzet J, Limburg PC, et al. High mobility group box1 (HMGB1) in relation to cutaneous inflammation in systemic lupus erythematosus (SLE). *Lupus*. (2013) 22:597– 606. doi: 10.1177/0961203313483377
- 37. Popovic K, Ek M, Espinosa A, Padyukov L, Harris HE, Wahren-Herlenius M, et al. Increased expression of the novel proinflammatory cytokine high mobility group box chromosomal protein 1 in skin lesions of patients with lupus erythematosus. *Arthritis Rheum.* (2005) 52:3639–45. doi: 10.1002/art.21398
- Garcia-Romo GS, Caielli S, Vega B, Connolly J, Allantaz F, Xu Z, et al. Netting neutrophils are major inducers of type I IFN production in pediatric systemic lupus erythematosus. *Sci Transl Med.* (2011) 3:73ra20. doi: 10.1126/scitranslmed.3001201
- Barkauskaite V, Ek M, Popovic K, Harris HE, Wahren-Herlenius M, Nyberg F. Translocation of the novel cytokine HMGB1 to the cytoplasm and extracellular space coincides with the peak of clinical activity in experimentally UV-induced lesions of cutaneous lupus erythematosus. *Lupus.* (2007) 16:794–802. doi: 10.1177/0961203307 081895
- Verdelli A, Coi A, Marzano AV, Antiga E, Cozzani E, Quaglino P, et al. Autoantibody profile and clinical patterns in 619 Italian patients with cutaneous lupus erythematosus. J Eur Acad Dermatol Venereol. (2019) 33:742–52. doi: 10.1111/jdv.15147
- Garcia S, Campos-de-Carvalho AC. Neonatal lupus syndrome: the heart as a target of the immune system. An Acad Bras Cienc. (2000) 72:83– 9. doi: 10.1590/S0001-376520000010012
- Hornberger LK, Al Rajaa N. Spectrum of cardiac involvement in neonatal lupus. Scand J Immunol. (2010) 72:189– 97. doi: 10.1111/j.1365-3083.2010.02437.x
- Hon KL, Leung AKC. Neonatal lupus erythematosus. Autoimmune Dis. (2012) 2012:301274. doi: 10.1155/2012/301274
- Barsalou J, Costedoat-Chalumeau N, Berhanu A, Fors-Nieves C, Shah U, Brown P, et al. Effect of in utero hydroxychloroquine exposure on the development of cutaneous neonatal lupus erythematosus. *Ann Rheum Dis.* (2018) 77:1742–9. doi: 10.1136/annrheumdis-2018-213718
- Shahian M, Khosravi A, Anbardar M-H. Early cholestasis in neonatal lupus erythematosus. Ann Saudi Med. (2011) 31:80–2. doi: 10.5144/0256-4947.2011.80
- Eftekhari P, Sallé L, Lezoualc'h F, Mialet J, Gastineau M, Briand JP, et al. Anti-SSA/Ro52 autoantibodies blocking the cardiac 5-HT4 serotoninergic

receptor could explain neonatal lupus congenital heart block. *Eur J Immunol.* (2000) 30:2782–90. doi: 10.1002/1521-4141(200010)30:10<2782:AID-IMMU2782>3.0.CO;2-9

- Buyon JP. Neonatal lupus: bedside to bench and back. *Scand J Rheumatol.* (1996) 25:271–6. doi: 10.3109/03009749609104057
- Jaeggi E, Laskin C, Hamilton R, Kingdom J, Silverman E. The importance of the level of maternal anti-Ro/SSA antibodies as a prognostic marker of the development of cardiac neonatal lupus erythematosus a prospective study of 186 antibody-exposed fetuses and infants. *J Am Coll Cardiol.* (2010) 55:2778–84. doi: 10.1016/j.jacc.2010.02.042
- Levy R, Briggs L, Silverman E, Pope E, Lara-Corrales I. Cutaneous sequelae in neonatal lupus: a retrospective cohort study. J Am Acad Dermatol. (2019). doi: 10.1016/j.jaad.2019.09.083. [Epub ahead of print].
- Hersh AO, Arkin LM, Prahalad S. Immunogenetics of cutaneous lupus erythematosus. Curr Opin Pediatr. (2016) 28:470-5. doi: 10.1097/MOP.00000000000383
- 51. Järvinen TM, Hellquist A, Koskenmies S, Einarsdottir E, Koskinen LLE, Jeskanen L, et al. Tyrosine kinase 2 and interferon regulatory factor 5 polymorphisms are associated with discoid and subacute cutaneous lupus erythematosus. *Exp Dermatol.* (2010) 19:123–31. doi: 10.1111/j.1600-0625.2009.00982.x
- Fiehn C. Familial chilblain lupus what can we learn from type I interferonopathies? *Curr Rheumatol Rep.* (2017) 19:61. doi: 10.1007/s11926-017-0689-x
- König N, Fiehn C, Wolf C, Schuster M, Costa EC, Tüngler V, et al. Familial chilblain lupus due to a gain-of-function mutation in STING. *Ann Rheum Dis.* (2017) 76:468–472. doi: 10.1136/annrheumdis-2016-209841
- Rice G, Newman WG, Dean J, Patrick T, Parmar R, Flintoff K, et al. Heterozygous mutations in TREX1 cause familial chilblain lupus and dominant aicardi-goutières syndrome. *Am J Hum Genet.* (2007) 80:811– 5. doi: 10.1086/513443
- Zimmermann N, Wolf C, Schwenke R, Lüth A, Schmidt F, Engel K, et al. Assessment of clinical response to janus kinase inhibition in patients with familial chilblain lupus and TREX1 mutation. *JAMA Dermatol.* (2019) 155:342–6. doi: 10.1001/jamadermatol.2018.5077
- Günther C, Berndt N, Wolf C, Lee-Kirsch MA. Familial chilblain lupus due to a novel mutation in the exonuclease III domain of 3' repair exonuclease 1 (TREX1). JAMA Dermatol. (2015) 151:426– 31. doi: 10.1001/jamadermatol.2014.3438
- Kuhn A, Wenzel J, Weyd H. Photosensitivity, apoptosis, and cytokines in the pathogenesis of lupus erythematosus: a critical review. *Clin Rev Allergy Immunol.* (2014) 47:148–62. doi: 10.1007/s12016-013-8403-x
- Meller S, Winterberg F, Gilliet M, Müller A, Lauceviciute I, Rieker J, et al. Ultraviolet radiation-induced injury, chemokines, and leukocyte recruitment: an amplification cycle triggering cutaneous lupus erythematosus. *Arthritis Rheum.* (2005) 52:1504–16. doi: 10.1002/art.21034
- Kochevar IE. Action spectrum and mechanisms of UV radiation-induced injury in lupus erythematosus. J Invest Dermatol. (1985) 85:S140– S3. doi: 10.1111/1523-1747.ep12275658
- 60. Furukawa F, Itoh T, Wakita H, Yagi H, Tokura Y, Norris DA, et al. Keratinocytes from patients with lupus erythematosus show enhanced cytotoxicity to ultraviolet radiation and to antibody-mediated cytotoxicity. *Clin Exp Immunol.* (1999) 118:164–70. doi: 10.1046/j.1365-2249.1999.01026.x
- Kuhn A, Herrmann M, Kleber S, Beckmann-Welle M, Fehsel K, Martin-Villalba A, et al. Accumulation of apoptotic cells in the epidermis of patients with cutaneous lupus erythematosus after ultraviolet irradiation. *Arthritis Rheum.* (2006) 54:939–50. doi: 10.1002/art.21658
- 62. Reefman E, de Jong MC, Kuiper H, Jonkman MF, Limburg PC, Kallenberg CGM, et al. Is disturbed clearance of apoptotic keratinocytes responsible for UVB-induced inflammatory skin lesions in systemic lupus erythematosus? *Arthritis Res Ther.* (2006) 8:R156. doi: 10.1186/ar2051
- Wolf SJ, Estadt SN, Gudjonsson JE, Kahlenberg JM. Human and murine evidence for mechanisms driving autoimmune photosensitivity. *Front Immunol.* (2018) 9:2430. doi: 10.3389/fimmu.2018.02430
- Laurinaviciene R, Sandholdt LH, Bygum A. Drug-induced cutaneous lupus erythematosus: 88 new cases. *Eur J Dermatol.* (2017) 27:28– 33. doi: 10.1684/ejd.2016.2912

- Sandholdt LH, Laurinaviciene R, Bygum A. Proton pump inhibitor-induced subacute cutaneous lupus erythematosus. Br J Dermatol. (2014) 170:342– 51. doi: 10.1111/bjd.12699
- Borucki R, Werth VP. Cutaneous lupus erythematosus induced by drugs - novel insights. *Expert Rev Clin Pharmacol.* (2019) 13:35– 42. doi: 10.1080/17512433.2020.1698290
- Lowe GC, Henderson CL, Grau RH, Hansen CB, Sontheimer RD. A systematic review of drug-induced subacute cutaneous lupus erythematosus. *Br J Dermatol.* (2011) 164:465–72. doi: 10.1111/j.1365-2133.2010.10110.x
- Reed BR, Huff JC, Jones SK, Orton PW, Lee LA, Norris DA. Subacute cutaneous lupus erythematosus associated with hydrochlorothiazide therapy. *Ann Intern Med.* (1985) 103:49– 51. doi: 10.7326/0003-4819-103-1-49
- Kluger N, Bessis D, Guillot B. Chronic cutaneous lupus flare induced by systemic 5-fluorouracil. J Dermatolog Treat. (2006) 17:51–3. doi: 10.1080/09546630500475591
- Weger W, Kränke B, Gerger A, Salmhofer W, Aberer E. Occurrence of subacute cutaneous lupus erythematosus after treatment with fluorouracil and capecitabine. J Am Acad Dermatol. (2008) 59:S4–S6. doi: 10.1016/j.jaad.2007.06.040
- Aggarwal N. Drug-induced subacute cutaneous lupus erythematosus associated with proton pump inhibitors. *Drugs Real World Outcomes.* (2016) 3:145–54. doi: 10.1007/s40801-016-0067-4
- Reich A, Maj J. Subacute cutaneous lupus erythematosus due to proton pump inhibitor intake: case report and literature review. *Arch Med Sci.* (2012) 8:743–7. doi: 10.5114/aoms.2012.30300
- Vedove CD, Simon JC, Girolomoni G. Drug-induced lupus erythematosus with emphasis on skin manifestations and the role of anti-TNFα agents. J Dtsch Dermatol Ges. (2012) 10:889–97. doi: 10.1111/j.1610-0387.2012.08000.x
- Boeckler P, Cosnes A, Francès C, Hedelin G, Lipsker D. Association of cigarette smoking but not alcohol consumption with cutaneous lupus erythematosus. *Arch Dermatol.* (2009) 145:1012–6. doi: 10.1001/archdermatol.2009.199
- Miot HA, Bartoli Miot LD, Haddad GR. Association between discoid lupus erythematosus and cigarette smoking. *Dermatology*. (2005) 211:118– 22. doi: 10.1159/000086440
- Rahman P, Gladman DD, Urowitz MB. Smoking interferes with efficacy of antimalarial therapy in cutaneous lupus. J Rheumatol. (1998) 25:1716–9.
- 77. Chasset F, Francès C, Barete S, Amoura Z, Arnaud L. Influence of smoking on the efficacy of antimalarials in cutaneous lupus: a meta-analysis of the literature. J Am Acad Dermatol. (2015) 72:634– 9. doi: 10.1016/j.jaad.2014.12.025
- Wattiaux A, Bettendorf B, Block L, Gilmore-Bykovskyi A, Ramly E, Piper ME, et al. Patient perspectives on smoking cessation and interventions in rheumatology clinics. *Arthritis Care Res.* (2019) 72:369–77. doi: 10.1002/acr. 23858
- Cheng HM, Liu WC, Chua G, Liew CF, Li W, Choo W, et al. Impact of a pharmacy-led smoking cessation clinic in a dermatology centre. *Singapore Med J.* (2019) 60:31–3. doi: 10.11622/smedj.2018063
- Naranjo A, Khan NA, Cutolo M, Lee S-S, Lazovskis J, Laas K, et al. Smoking cessation advice by rheumatologists: results of an international survey. *Rheumatology*. (2014) 53:1825–9. doi: 10.1093/rheumatology/keu213
- Masson C, Audran M, Pascaretti C, Chevailler A, Subra JF, Tuchais E, et al. Silica-associated systemic erythematosus lupus or mineral dust lupus? *Lupus*. (1997) 6:1–3. doi: 10.1177/096120339700600101
- Vukicevic JS, Milobratovic DJ. Discoid lupus erythematosus of the eyelid. *Indian J Dermatol Venereol Leprol.* (2010) 76:418–20. doi: 10.4103/0378-6323.66592
- Sirobhushanam S, Parsa N, Reed TJ, Berthier CC, Sarkar MK, Hile GA, et al. *Staphylococcus aureus* colonization is increased on lupus skin lesions and is promoted by interferon-mediated barrier disruption. *J Invest Dermatol.* (2019) 140:1066–74.e4. doi: 10.1016/j.jid.2019.11.016
- Wenzel J. Cutaneous lupus erythematosus: new insights into pathogenesis and therapeutic strategies. Nat Rev Rheumatol. (2019) 15:519–32. doi: 10.1038/s41584-019-0272-0
- 85. Nagao K, Kobayashi T, Moro K, Ohyama M, Adachi T, Kitashima DY, et al. Stress-induced production of chemokines by hair follicles regulates

the trafficking of dendritic cells in skin. Nat Immunol. (2012) 13:744-52. doi: 10.1038/ni.2353

- Shipman WD, Chyou S, Ramanathan A, Izmirly PM, Sharma S, Pannellini T, et al. A protective Langerhans cell-keratinocyte axis that is dysfunctional in photosensitivity. *Sci Transl Med.* (2018) 10:eaap9527. doi: 10.1126/scitranslmed.aap9527
- Hatakeyama M, Fukunaga A, Washio K, Taguchi K, Oda Y, Ogura K, et al. Anti-inflammatory role of langerhans cells and apoptotic keratinocytes in ultraviolet-B-induced cutaneous inflammation. *J Immunol.* (2017) 199:2937– 47. doi: 10.4049/jimmunol.1601681
- Sontheimer RD, Bergstresser PR. Epidermal Langerhans cell involvement in cutaneous lupus erythematosus. J Invest Dermatol. (1982) 79:237– 43. doi: 10.1111/1523-1747.ep12500069
- Mori M, Pimpinelli N, Romagnoli P, Bernacchi E, Fabbri P, Giannotti B. Dendritic cells in cutaneous lupus erythematosus: a clue to the pathogenesis of lesions. *Histopathology*. (1994) 24:311–21. doi: 10.1111/j.1365-2559.1994.tb00531.x
- Berthier CC, Tsoi LC, Reed TJ, Stannard JN, Myers EM, Namas R, et al. Molecular profiling of cutaneous lupus lesions identifies subgroups distinct from clinical phenotypes. J Clin Med Res. (2019) 8:1244. doi: 10.3390/jcm8081244
- McNiff JM, Kaplan DH. Plasmacytoid dendritic cells are present in cutaneous dermatomyositis lesions in a pattern distinct from lupus erythematosus. J Cutan Pathol. (2008) 35:452–6. doi: 10.1111/j.1600-0560.2007.00848.x
- Farkas L, Beiske K, Lund-Johansen F, Brandtzaeg P, Jahnsen FL. Plasmacytoid dendritic cells (natural interferon-α/β-producing cells) accumulate in cutaneous lupus erythematosus lesions. *Am J Pathol.* (2001) 159:237– 43. doi: 10.1016/S0002-9440(10)61689-6
- 93. Tomasini D, Mentzel T, Hantschke M, Cerri A, Paredes B, Rütten A, et al. Plasmacytoid dendritic cells: an overview of their presence and distribution in different inflammatory skin diseases, with special emphasis on Jessner's lymphocytic infiltrate of the skin and cutaneous lupus erythematosus. J Cutan Pathol. (2010) 37:1132–9. doi: 10.1111/j.1600-0560.2010.01587.x
- Yin Q, Xu X, Lin Y, Lv J, Zhao L, He R. Ultraviolet B irradiation induces skin accumulation of plasmacytoid dendritic cells: a possible role for chemerin. *Autoimmunity.* (2014) 47:185–92. doi: 10.3109/08916934.2013.866105
- Furie R, Werth VP, Merola JF, Stevenson L, Reynolds TL, Naik H, et al. Monoclonal antibody targeting BDCA2 ameliorates skin lesions in systemic lupus erythematosus. J Clin Invest. (2019) 129:1359– 71. doi: 10.1172/JCI124466
- Safi R, Al-Hage J, Abbas O, Kibbi A-G, Nassar D. Investigating the presence of neutrophil extracellular traps in cutaneous lesions of different subtypes of lupus erythematosus. *Exp Dermatol.* (2019) 28:1348– 52. doi: 10.1111/exd.14040
- Villanueva E, Yalavarthi S, Berthier CC, Hodgin JB, Khandpur R, Lin AM, et al. Netting neutrophils induce endothelial damage, infiltrate tissues, and expose immunostimulatory molecules in systemic lupus erythematosus. J Immunol. (2011) 187:538–52. doi: 10.4049/jimmunol.1100450
- Guo X, Fang X, He G, Zaman MH, Fei X, Qiao W, et al. The role of neutrophils in skin damage induced by tissue-deposited lupus IgG. *Immunology*. (2018) 154:604–12. doi: 10.1111/imm.12908
- Nakajima M, Nakajima A, Kayagaki N, Honda M, Yagita H, Okumura K. Expression of Fas ligand and its receptor in cutaneous lupus: implication in tissue injury. *Clin Immunol Immunopathol.* (1997) 83:223–9. doi: 10.1006/clin.1997.4352
- 100. Liu J, Berthier CC, Kahlenberg JM. Enhanced inflammasome activity in systemic lupus erythematosus is mediated via type I interferon upregulation of interferon regulatory factor 1. *Arthritis Rheumatol.* (2017) 69:1840– 9. doi: 10.1002/art.40166
- 101. Masek-Hammerman K, Peeva E, Ahmad A, Menon S, Afsharvand M, Peng Qu R, et al. Monoclonal antibody against macrophage colonystimulating factor suppresses circulating monocytes and tissue macrophage function but does not alter cell infiltration/activation in cutaneous lesions or clinical outcomes in patients with cutaneous lupus erythematosus. *Clin Exp Immunol.* (2016) 183:258–70. doi: 10.1111/cei.12705
- Korb LC, Ahearn JM. C1q binds directly and specifically to surface blebs of apoptotic human keratinocytes: complement deficiency and systemic lupus erythematosus revisited. *J Immunol.* (1997) 158:4525–8.

- 103. Racila DM, Sontheimer CJ, Sheffield A, Wisnieski JJ, Racila E, Sontheimer RD. Homozygous single nucleotide polymorphism of the complement C1QA gene is associated with decreased levels of C1q in patients with subacute cutaneous lupus erythematosus. *Lupus.* (2003) 12:124–32. doi: 10.1191/0961203303lu3290a
- 104. Wenzel J, Zahn S, Mikus S, Wiechert A, Bieber T, Tüting T. The expression pattern of interferon-inducible proteins reflects the characteristic histological distribution of infiltrating immune cells in different cutaneous lupus erythematosus subsets. *Br J Dermatol.* (2007) 157:752–7. doi: 10.1111/j.1365-2133.2007.08137.x
- 105. Sinha AA, Dey-Rao R. Genomic Investigation of Lupus in the Skin. J Investig Dermatol Symp Proc. (2017) 18:S75–S80. doi: 10.1016/j.jisp.2016.09.002
- Lipsker D, Saurat J-H. Neutrophilic cutaneous lupus erythematosus. Dermatology. (2008) 216:283. doi: 10.1159/000113940
- Lu L, Kaliyaperumal A, Boumpas DT, Datta SK. Major peptide autoepitopes for nucleosome-specific T cells of human lupus. *J Clin Invest.* (1999) 104:345– 55. doi: 10.1172/JCI6801
- 108. Voll RE, Roth EA, Girkontaite I, Fehr H, Herrmann M, Lorenz H-M, et al. Histone-specific Th0 and Th1 clones derived from systemic lupus erythematosus patients induce double-stranded DNA antibody production. *Arthritis Rheum.* (1997) 40:2162–71. doi: 10.1002/art.1780401210
- 109. Furukawa F, Tokura Y, Matsushita K, Iwasaki-Inuzuka K, Onagi-Suzuki K, Yagi H, et al. Selective expansions of T cells expressing Vβ38 and Vβ13 in skin lesions of patients with chronic cutaneous lupus erythematosus. *J Dermatol.* (1996) 23:670–6. doi: 10.1111/j.1346-8138.1996.tb02679.x
- Wenzel J, Zahn S, Bieber T, Tüting T. Type I interferon-associated cytotoxic inflammation in cutaneous lupus erythematosus. *Arch Dermatol Res.* (2009) 301:83–6. doi: 10.1007/s00403-008-0892-8
- 111. Grassi M, Capello F, Bertolino L, Seia Z. Identification of granzyme Bexpressing CD-8-positive T cells in lymphocytic inflammatory infiltrate in cutaneous lupus erythematosus and in dermatomyositis. *Clin Exp Dermatol.* (2009) 34:910–4. doi: 10.1111/j.1365-2230.2009.03297.x
- 112. Luo Y, Zhang X, Zhao M, Lu Q. DNA demethylation of the perforin promoter in CD4⁺ T cells from patients with subacute cutaneous lupus erythematosus. *J Dermatol Sci.* (2009) 56:33–6. doi: 10.1016/j.jdermsci.2009.06.010
- 113. Luo Y, Zhao M, Lu Q. Demethylation of promoter regulatory elements contributes to CD70 overexpression in CD4⁺ T cells from patients with subacute cutaneous lupus erythematosus. *Clin Exp Dermatol.* (2010) 35:425– 30. doi: 10.1111/j.1365-2230.2009.03611.x
- 114. Luo Y, Li Y, Su Y, Yin H, Hu N, Wang S, et al. Abnormal DNA methylation in T cells from patients with subacute cutaneous lupus erythematosus. Br J Dermatol. (2008) 159:827–33. doi: 10.1111/j.1365-2133.2008.08758.x
- 115. Wenzel J, Henze S, Brähler S, Bieber T, Tüting T. The expression of human leukocyte antigen-DR and CD25 on circulating T cells in cutaneous lupus erythematosus and correlation with disease activity. *Exp Dermatol.* (2005) 14:454–9. doi: 10.1111/j.0906-6705.2005.00301.x
- 116. Franz B, Fritzsching B, Riehl A, Oberle N, Klemke C-D, Sykora J, et al. Low number of regulatory T cells in skin lesions of patients with cutaneous lupus erythematosus. *Arthritis Rheum*. (2007) 56:1910–20. doi: 10.1002/art.22699
- 117. Gambichler T, Pätzholz J, Schmitz L, Lahner N, Kreuter A. FOXP3⁺ and CD39⁺ regulatory T cells in subtypes of cutaneous lupus erythematosus. J Eur Acad Dermatol Venereol. (2015) 29:1972–7. doi: 10.1111/jdv.13123
- Volc-Platzer B, Anegg B, Milota S, Pickl W, Fischer G. Accumulation of γδ cells in chronic cutaneous lupus erythematosus. J Invest Dermatol. (1993) 100:S84–S91. doi: 10.1038/jid.1993.29
- 119. Walsh NM, Kutzner H, Requena L, Cerroni L. Plasmacytic cutaneous pathology: a review. J Cutan Pathol. (2019) 46:698– 708. doi: 10.1111/cup.13499
- Chan O, Shlomchik MJ. A new role for B cells in systemic autoimmunity: B cells promote spontaneous T cell activation in MRL-lpr/lpr mice. *J Immunol.* (1998) 160:51–9.
- 121. Shlomchik MJ, Madaio MP, Ni D, Trounstein M, Huszar D. The role of B cells in lpr/lpr-induced autoimmunity. J Exp Med. (1994) 180:1295– 306. doi: 10.1084/jem.180.4.1295
- 122. Chan OT, Hannum LG, Haberman AM, Madaio MP, Shlomchik MJ. A novel mouse with B cells but lacking serum antibody reveals an antibodyindependent role for B cells in murine lupus. *J Exp Med.* (1999) 189:1639– 48. doi: 10.1084/jem.189.10.1639

- 123. Kansal R, Richardson N, Neeli I, Khawaja S, Chamberlain D, Ghani M, et al. Sustained B cell depletion by CD19-targeted CAR T cells is a highly effective treatment for murine lupus. *Sci Transl Med.* (2019) 11:eaav1648. doi: 10.1126/scitranslmed.aav1648
- Correll CK, Miller DD, Maguiness SM. Treatment of childhood-onset lupus erythematosus panniculitis with rituximab. *JAMA Dermatol.* (2020) 156:566–9. doi: 10.1001/jamadermatol.2019.4984
- 125. Hofmann SC, Leandro MJ, Morris SD, Isenberg DA. Effects of rituximab-based B-cell depletion therapy on skin manifestations of lupus erythematosus-report of 17 cases and review of the literature. *Lupus*. (2013) 22:932–9. doi: 10.1177/0961203313497115
- 126. Herati RS, Muselman A, Vella L, Bengsch B, Parkhouse K, Del Alcazar D, et al. Successive annual influenza vaccination induces a recurrent oligoclonotypic memory response in circulating T follicular helper cells. *Sci Immunol.* (2017) 2:eaag2152 doi: 10.1126/sciimmunol.aag2152
- 127. González-Escribano MF, Aguilar F, Torres B, Sánchez-Román J, Núñez-Roldán A. CD38 polymorphisms in Spanish patients with systemic lupus erythematosus. *Hum Immunol.* (2004) 65:660–4. doi: 10.1016/j.humimm.2004.02.032
- Viegas MS, Silva T, Monteiro MM, do Carmo A, Martins TC. Knocking out of CD38 accelerates development of a lupus-like disease in lpr mice. *Rheumatology*. (2011) 50:1569–77. doi: 10.1093/rheumatology/ker178
- 129. Erkeller-Yuksel FM, Lydyard PM, Isenberg DA. Lack of NK cells in lupus patients with renal involvement. *Lupus*. (1997) 6:708–12. doi: 10.1177/096120339700600905
- 130. Green MRJ, Kennell ASM, Larche MJ, Seifert MH, Isenberg DA, Salaman MR. Natural killer cell activity in families of patients with systemic lupus erythematosus: demonstration of a killing defect in patients. *Clin Exp Immunol.* (2005) 141:165–73. doi: 10.1111/j.1365-2249.2005.02822.x
- 131. Henriques A, Teixeira L, Inês L, Carvalheiro T, Gonçalves A, Martinho A, et al. NK cells dysfunction in systemic lupus erythematosus: relation to disease activity. *Clin Rheumatol.* (2013) 32:805–13. doi: 10.1007/s10067-013-2176-8
- 132. Park Y-W, Kee S-J, Cho Y-N, Lee E-H, Lee H-Y, Kim E-M, et al. Impaired differentiation and cytotoxicity of natural killer cells in systemic lupus erythematosus. *Arthritis Rheum.* (2009) 60:1753–63. doi: 10.1002/art. 24556
- 133. Schepis D, Gunnarsson I, Eloranta M-L, Lampa J, Jacobson SH, Kärre K, et al. Increased proportion of CD56bright natural killer cells in active and inactive systemic lupus erythematosus. *Immunology.* (2009) 126:140–6. doi: 10.1111/j.1365-2567.2008.02887.x
- 134. Huang Z, Fu B, Zheng SG, Li X, Sun R, Tian Z, et al. Involvement of CD226⁺ NK cells in immunopathogenesis of systemic lupus erythematosus. *J Immunol.* (2011) 186:3421–31. doi: 10.4049/jimmunol.1000569
- 135. Hervier B, Beziat V, Haroche J, Mathian A. Phenotype and function of natural killer cells in systemic lupus erythematosus: excess interferonγ production in patients with active disease. *Arthritis.* (2011) 63:1698– 706. doi: 10.1002/art.30313
- Raulet DH, Guerra N. Oncogenic stress sensed by the immune system: role of natural killer cell receptors. *Nat Rev Immunol.* (2009) 9:568– 80. doi: 10.1038/nri2604
- 137. Swann JB, Hayakawa Y, Zerafa N, Sheehan KCF, Scott B, Schreiber RD, et al. Type I IFN contributes to NK cell homeostasis, activation, and antitumor function. *J Immunol.* (2007) 178:7540–9. doi: 10.4049/jimmunol.178.12.7540
- Smyth MJ, Hayakawa Y, Takeda K, Yagita H. New aspects of natural-killercell surveillance and therapy of cancer. *Nat Rev Cancer*. (2002) 2:850– 61. doi: 10.1038/nrc928
- 139. Sullivan TP, Dearaujo T, Vincek V, Berman B. Evaluation of superficial basal cell carcinomas after treatment with imiquimod 5% cream or vehicle for apoptosis and lymphocyte phenotyping. *Dermatol Surg.* (2003) 29:1181– 6. doi: 10.1097/00042728-200312000-00007
- 140. Nitahara A, Shimura H, Ito A, Tomiyama K, Ito M, Kawai K. NKG2D ligation without T cell receptor engagement triggers both cytotoxicity and cytokine production in dendritic epidermal T cells. J Invest Dermatol. (2006) 126:1052–8. doi: 10.1038/sj.jid.57 00112
- 141. Hofmann SC, Bosma A, Bruckner-Tuderman L, Vukmanovic-Stejic M, Jury EC, Isenberg DA, et al. Invariant natural killer T cells are enriched at the site

of cutaneous inflammation in lupus erythematosus. J Dermatol Sci. (2013) 71:22–8. doi: 10.1016/j.jdermsci.2013.04.012

- 142. Wenzel J, Wörenkämper E, Freutel S, Henze S, Haller O, Bieber T, et al. Enhanced type I interferon signalling promotes Th1-biased inflammation in cutaneous lupus erythematosus. *J Pathol.* (2005) 205:435–42. doi: 10.1002/path.1721
- 143. Flier J, Boorsma DM, van Beek PJ, Nieboer C, Stoof TJ, Willemze R, et al. Differential expression of CXCR3 targeting chemokines CXCL10, CXCL9, and CXCL11 in different types of skin inflammation. J Pathol. (2001) 194:398–405. doi: 10.1002/1096-9896(200108)194:4<397::AID-PATH899>3. 0.CO;2-S
- 144. Denton AE, Innocentin S, Carr EJ, Bradford BM, Lafouresse F, Mabbott NA, et al. Type I interferon induces CXCL13 to support ectopic germinal center formation. J Exp Med. (2019) 216:621–37. doi: 10.1084/jem.20181216
- Moser B. CXCR5, the defining marker for follicular B helper T (TFH) Cells. Front Immunol. (2015) 6:296. doi: 10.3389/fimmu.2015.00296
- 146. Niederkorn A, Frühauf J, Schwantzer G, Wutte N, Painsi C, Werner S, et al. CXCL13 is an activity marker for systemic, but not cutaneous lupus erythematosus: a longitudinal cohort study. *Arch Dermatol Res.* (2018) 310:485–93. doi: 10.1007/s00403-018-1836-6
- 147. Clark KL, Reed TJ, Wolf SJ, Lowe L, Hodgin JB, Kahlenberg JM. Epidermal injury promotes nephritis flare in lupus-prone mice. J Autoimmun. (2015) 65:38–48. doi: 10.1016/j.jaut.2015.08.005
- Sáez de Guinoa J, Barrio L, Mellado M, Carrasco YR. CXCL13/CXCR5 signaling enhances BCR-triggered B-cell activation by shaping cell dynamics. *Blood.* (2011) 118:1560–9. doi: 10.1182/blood-2011-01-332106
- 149. Wenzel J, Henze S, Wörenkämper E, Basner-Tschakarjan E, Sokolowska-Wojdylo M, Steitz J, et al. Role of the chemokine receptor CCR4 and its ligand thymus- and activation-regulated chemokine/CCL17 for lymphocyte recruitment in cutaneous lupus erythematosus. J Invest Dermatol. (2005) 124:1241–8. doi: 10.1111/j.0022-202X.2005.23755.x
- 150. Freutel S, Gaffal E, Zahn S, Bieber T, Tüting T, Wenzel J. Enhanced CCR5⁺/CCR3⁺ T helper cell ratio in patients with active cutaneous lupus erythematosus. *Lupus*. (2011) 20:1300–4. doi: 10.1177/0961203311 409267
- 151. Mande P, Zirak B, Ko W-C, Taravati K, Bride KL, Brodeur TY, et al. Fas ligand promotes an inducible TLR-dependent model of cutaneous lupus-like inflammation. *J Clin Invest.* (2018) 128:2966–78. doi: 10.1172/JCI98219
- 152. Merrill JT, Furie R, Werth VP, Khamashta M, Drappa J, Wang L, et al. Anifrolumab effects on rash and arthritis: impact of the type I interferon gene signature in the phase IIb MUSE study in patients with systemic lupus erythematosus. *Lupus Sci Med.* (2018) 5:e000284. doi: 10.1136/lupus-2018-000284
- 153. Furie R, Khamashta M, Merrill JT, Werth VP, Kalunian K, Brohawn P, et al. Anifrolumab, an anti–interferon-α receptor monoclonal antibody, in moderate-to-severe systemic lupus erythematosus. *Arthritis Rheumatol.* (2017) 69:376–86. doi: 10.1002/art.39962
- 154. Felten R, Scher F, Sagez F, Chasset F, Arnaud L. Spotlight on anifrolumab and its potential for the treatment of moderate-to-severe systemic lupus erythematosus: evidence to date. *Drug Des Devel Ther.* (2019) 13:1535– 43. doi: 10.2147/DDDT.S170969
- 155. Werth VP, Fiorentino D, Sullivan BA, Boedigheimer MJ, Chiu K, Wang C, et al. Brief report: pharmacodynamics, safety, and clinical efficacy of AMG 811, a human anti-interferon-γ antibody, in patients with discoid lupus erythematosus. *Arthritis Rheumatol.* (2017) 69:1028– 34. doi: 10.1002/art.40052
- 156. Tsoi LC, Hile GA, Berthier CC, Sarkar MK, Reed TJ, Liu J, et al. Hypersensitive IFN responses in lupus keratinocytes reveal key mechanistic determinants in cutaneous lupus. *J Immunol.* (2019) 202:2121–30. doi: 10.4049/jimmunol.1800650
- 157. Sarkar MK, Hile GA, Tsoi LC, Xing X, Liu J, Liang Y, et al. Photosensitivity and type I IFN responses in cutaneous lupus are driven by epidermal-derived interferon kappa. Ann Rheum Dis. (2018) 77:1653– 64. doi: 10.1136/annrheumdis-2018-213197
- 158. Stannard JN, Reed TJ, Myers E, Lowe L, Sarkar MK, Xing X, et al. Lupus Skin is primed for IL-6 inflammatory responses through a keratinocytemediated autocrine type I interferon loop. *J Invest Dermatol.* (2017) 137:115– 22. doi: 10.1016/j.jid.2016.09.008

- 159. Salvi V, Vermi W, Cavani A, Lonardi S, Carbone T, Facchetti F, et al. IL-21 may promote granzyme B-dependent NK/plasmacytoid dendritic cell functional interaction in cutaneous lupus erythematosus. *J Invest Dermatol.* (2017) 137:1493–500. doi: 10.1016/j.jid.2017.03.016
- 160. Karrich JJ, Jachimowski LCM, Nagasawa M, Kamp A, Balzarolo M, Wolkers MC, et al. IL-21-stimulated human plasmacytoid dendritic cells secrete granzyme B, which impairs their capacity to induce T-cell proliferation. *Blood.* (2013) 121:3103–11. doi: 10.1182/blood-2012-08-452995
- 161. Yell JA, Burge SM. The effect of hormonal changes on cutaneous disease in lupus erythematosus. Br J Dermatol. (1993) 129:18–22. doi: 10.1111/j.1365-2133.1993.tb03305.x
- 162. Jones SK. The effects of hormonal and other stimuli on cell-surface Ro/SSA antigen expression by human keratinocytes in vitro: their possible role in the induction of cutaneous lupus lesions. *Br J Dermatol.* (1992) 126:554–60. doi: 10.1111/j.1365-2133.1992.tb00099.x
- 163. Furukawa F, Lyons MB, Lee LA. Estradiol enhances binding to cultured human keratinocytes of antibodies specific for SS-A/Ro and SS-B/La. Another possible mechanism for estradiol influence of Lupus Erythematosus. *J Immunol.* (1988) 141:1480–8.
- Zandman-Goddard G, Solomon M, Barzilai A, Shoenfeld Y. Lupus erythematosus tumidus induced by sex reassignment surgery. J Rheumatol. (2007) 34:1938–40.
- 165. Fox HS, Bond BL, Parslow TG. Estrogen regulates the IFN-gamma promoter. *J Immunol.* (1991) 146:4362–7.
- 166. Sorachi K-I, Kumagai S, Sugita M, Yodoi J, Imura H. Enhancing effect of 17β-estradiol on human NK cell activity. *Immunol Lett.* (1993) 36:31– 5. doi: 10.1016/0165-2478(93)90065-A
- 167. Liang Y, Tsoi LC, Xing X, Beamer MA, Swindell WR, Sarkar MK, et al. A gene network regulated by the transcription factor VGLL3 as a promoter of sex-biased autoimmune diseases. *Nat Immunol.* (2016) 18:152. doi: 10.1038/ni.3643
- Billi AC, Gharaee-Kermani M, Fullmer J, Tsoi LC, Hill BD, Gruszka D, et al. The female-biased factor VGLL3 drives cutaneous and systemic autoimmunity. JCI Insight. (2019) 4:e127291. doi: 10.1172/jci.insight.127291
- 169. Klaeschen AS, Wolf D, Brossart P, Bieber T, Wenzel J. JAK inhibitor ruxolitinib inhibits the expression of cytokines characteristic of cutaneous lupus erythematosus. *Exp Dermatol.* (2017) 26:728–30. doi: 10.1111/exd.13253
- 170. Dörner T, Tanaka Y, Petri M, Smolen JS, Dow ER, Higgs RE, Benschop RJ, Abel A, Silk ME, de Bono S, et al. 185 Baricitinib-associated changes in type I interferon gene signature during a 24-week phase 2 clinical SLE trial. *Lupus Sci Med.* (2019) 6:A141. doi: 10.1136/lupus-2019-lsm.185
- 171. Wallace DJ, Furie RA, Tanaka Y, Kalunian KC, Mosca M, Petri MA, et al. Baricitinib for systemic lupus erythematosus: a double-blind, randomised, placebo-controlled, phase 2 trial. *Lancet.* (2018) 392:222– 31. doi: 10.1016/S0140-6736(18)31363-1
- 172. Namour F, Desrivot J, Van der Aa A, Harrison P, Tasset C, van't Klooster G. Clinical confirmation that the selective JAK1 inhibitor filgotinib (GLPG0634) has a low liability for drug-drug interactions. *Drug Metab Lett.* (2016) 10:38–48. doi: 10.2174/1872312810666151223103353
- 173. Baker KF, Isaacs JD. Novel therapies for immune-mediated inflammatory diseases: what can we learn from their use in rheumatoid arthritis, spondyloarthritis, systemic lupus erythematosus, psoriasis, Crohn's disease and ulcerative colitis? *Ann Rheum Dis.* (2018) 77:175–87. doi: 10.1136/annrheumdis-2017-211555

- 174. Pohlmeyer C, Cui Z-H, Han P, Clarke AS, Jones RM, Mollova N, et al. AB0484 Monotherapy with filgotinib, a jak1-selective inhibitor, reduces disease severity and alters immune cell subsets in the nzb/w fl murine model of lupus. *Ann Rheum Dis.* (2018) 77:1403. doi: 10.1136/annrheumdis-2018-eular.3367
- 175. Van Vollenhoven RF, Hahn BH, Tsokos GC, Wagner CL, Lipsky P, Touma Z, et al. Efficacy and safety of ustekinumab, an IL-12 and IL-23 inhibitor, in patients with active systemic lupus erythematosus: results of a multicentre, double-blind, phase 2, randomised, controlled study. *Lancet.* (2018) 392:1330–9. doi: 10.1016/S0140-6736(18)3 2167-6
- 176. Romero-Maté A, García-Donoso C, Hernández-Núñez A, Martínez-Morán C, Moreno-Torres A, Borbujo-Martínez J. Successful treatment of recalcitrant discoid lupus erythematosus with ustekinumab. *Dermatol Online* J. (2017) 23:13030/qt206538zm.
- 177. Dahl C, Johansen C, Kragballe K, Olesen AB. Ustekinumab in the treatment of refractory chronic cutaneous lupus erythematosus: a case report. Acta Derm Venereol. (2013) 93:368–9. doi: 10.2340/0001555 5-1467
- De Souza A, Ali-Shaw T, Strober BE, Franks AG Jr. Successful treatment of subacute lupus erythematosus with ustekinumab. *Arch Dermatol.* (2011) 147:896–8. doi: 10.1001/archdermatol.2011.185
- 179. Tierney E, Kirthi S, Ramsay B, Ahmad K. Ustekinumabinduced subacute cutaneous lupus. *JAAD Case Rep.* (2019) 5:271–3. doi: 10.1016/j.jdcr.2019.01.015
- 180. Urowitz MB, Isenberg DA, Wallace DJ. Safety and efficacy of hCDR1 (Edratide) in patients with active systemic lupus erythematosus: results of phase II study. *Lupus Sci Med.* (2015) 2:e000104. doi: 10.1136/lupus-2015-000104
- 181. Skuljec J, Chmielewski M, Happle C, Habener A, Busse M, Abken H, et al. Chimeric antigen receptor-redirected regulatory T cells suppress experimental allergic airway inflammation, a model of asthma. *Front Immunol.* (2017) 8:1125. doi: 10.3389/fimmu.2017.01125
- Mizui M, Tsokos GC. Targeting regulatory T cells to treat patients with systemic lupus erythematosus. *Front Immunol.* (2018) 9:786. doi: 10.3389/fimmu.2018.00786

Conflict of Interest: JR is an inventor on patent application #62489191, "Diagnosis and Treatment of Vitiligo" which covers targeting IL-15 and Trm for the treatment of vitiligo; and on patent application #15/851,651, "Anti-human CXCR3 antibodies for the Treatment of Vitiligo" which covers targeting CXCR3 for the treatment of vitiligo. JR is a reviewer for Frontiers Immunology journal.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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