



# **COVID-19, Pre-Eclampsia, and Complement System**

Chiara Agostinis<sup>1†</sup>, Alessandro Mangogna<sup>1†</sup>, Andrea Balduit<sup>2\*</sup>, Azin Aghamajidi<sup>3</sup>, Giuseppe Ricci<sup>1,4</sup>, Uday Kishore<sup>5</sup> and Roberta Bulla<sup>2</sup>

<sup>1</sup> Institute for Maternal and Child Health, Istituto di Ricovero e Cura a Carattere Scientifico (IRCCS) Burlo Garofolo, Trieste, Italy, <sup>2</sup> Department of Life Sciences, University of Trieste, Trieste, Italy, <sup>3</sup> Department of Immunology, School of Medicine, Iran University of Medical Sciences, Tehran, Iran, <sup>4</sup> Department of Medical, Surgical and Health Science, University of Trieste, Trieste, Italy, <sup>5</sup> Biosciences, College of Health, Medicine and Life Sciences, Brunel University London, Uxbridge, United Kingdom

### **OPEN ACCESS**

### Edited by:

Taruna Madan, National Institute for Research in Reproductive Health (ICMR), India

#### Reviewed by:

Eleni Gavriilaki, G. Papanikolaou General Hospital, Greece Christian Drouet, U1016 Institut Cochin (INSERM), France

### \*Correspondence:

Andrea Balduit abalduit@units.it orcid.org/0000-0001-5902-3205

<sup>†</sup>These authors have contributed equally to this work and share first authorship

#### Specialty section:

This article was submitted to Molecular Innate Immunity, a section of the journal Frontiers in Immunology

Received: 13 September 2021 Accepted: 25 October 2021 Published: 17 November 2021

#### Citation:

Agostinis C, Mangogna A, Balduit A, Aghamajidi A, Ricci G, Kishore U and Bulla R (2021) COVID-19, Pre-Eclampsia, and Complement System. Front. Immunol. 12:775168. doi: 10.3389/fimmu.2021.775168 COVID-19 is characterized by virus-induced injury leading to multi-organ failure, together with inflammatory reaction, endothelial cell (EC) injury, and prothrombotic coagulopathy with thrombotic events. Complement system (C) via its cross-talk with the contact and coagulation systems contributes significantly to the severity and pathological consequences due to SARS-CoV-2 infection. These immunopathological mechanisms overlap in COVID-19 and pre-eclampsia (PE). Thus, mothers contracting SARS-CoV-2 infection during pregnancy are more vulnerable to developing PE. SARS-CoV-2 infection of ECs, via its receptor ACE2 and co-receptor TMPRSS2, can provoke endothelial dysfunction and disruption of vascular integrity, causing hyperinflammation and hypercoagulability. This is aggravated by bradykinin increase due to inhibition of ACE2 activity by the virus. C is important for the progression of normal pregnancy, and its dysregulation can impact in the form of PE-like syndrome as a consequence of SARS-CoV-2 infection. Thus, there is also an overlap between treatment regimens of COVID-19 and PE. C inhibitors, especially those targeting C3 or MASP-2, are exciting options for treating COVID-19 and consequent PE. In this review, we examine the role of C, contact and coagulation systems as well as endothelial hyperactivation with respect to SARS-CoV-2 infection during pregnancy and likely development of PE.

### Keywords: COVID-19, complement system, SARS-CoV-2, pregnancy, pre-eclampsia

# INTRODUCTION

The first case of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection, responsible for the coronavirus disease 2019 (COVID-19) outbreak, was reported in the Chinese town of Wuhan in the late 2019 (1). The emerging coronavirus spread worldwide over the following months has been officially recognized as a global pandemic since 11<sup>th</sup> March 2020 (2).

SARS-CoV-2 resembles several characteristics and pathways of infection with other two members of the *Coronaviridae* family: the SARS pandemic in 2002 and the Middle East respiratory syndrome (MERS) in 2012, with a fatality rate of 10% and 36%, respectively (3). CoVs encompass a group of enveloped and single-stranded RNA viruses identified in birds and mammals, and can cause gastrointestinal, central nervous system, and respiratory tract infections (4). The main structural proteins, encoded by specific genes in open reading frame (ORF)-1

1

downstream regions, include Spike (S), Envelope (E), Membrane (M), and Nucleocapsid (N), S protein being responsible for SARS-CoV-2 invasion of the host cells (5, 6). Among the different receptors identified as cellular entry mediators, the main target receptor of the SARS-CoV-2 virus is the angiotensin-converting enzyme 2 (ACE2) (7), which plays an important role in the renin-angiotensin-aldosterone system (RAAS) for the regulation of blood pressure and electrolyte homeostasis (8), and is widely expressed by most tissues, accounting for the high tropism of the virus. SARS-CoV-2 cell entry also depends on the cellular transmembrane serine protease 2 (TMPRSS2), responsible for S protein priming (7). Extracellular matrix metalloproteinase inducer (EMMPRIN, also known as basigin or CD147), a cell surface glycoprotein that belongs to the immunoglobulin superfamily and activates metalloproteases, has been regarded as a target for SARS-CoV-2 attachment and entry into the host cell (9, 10). In addition, the receptor for Semaphorin-3, named neuropilin-1, has also been demonstrated to facilitate SARS-CoV-2 infection (11).

The SARS-CoV-2 infection damages various organs via different pathogenic mechanisms, including direct viral damage to the host cells/tissues through pneumocyte syncytia formation (12) and Golgi apparatus rupture (13), RAAS disruption and endothelial cell (EC) damage resulting in inflammation, endotheliitis and thrombosis (14). Moreover, an unfavorable dysregulation of immune response characterized by lymphopenia and cytokine storm has been reported as a key pathogenic mechanism in COVID-19 (15, 16). In this context, the complement system (C) primary role as a first line of defense against infectious agents would suggest for a protective function in enhancing virus neutralization and phagocytosis. However, C activation has been proposed as a contributor in disease progression to a more severe and lethal stage, which shares important pathophysiological features, in particular endothelial damage, with a pregnancy disorder called pre-eclampsia (PE) (17, 18). This review aims to shed light on the role of the C in pregnant women with COVID-19 developing PE.

## THE COMPLEMENT SYSTEM

The C, as a powerful arm of the innate immunity, has a pivotal role in the recognition of potential danger signals and in the clearance of pathogens, apoptotic and necrotic cells (19). The C acts as a functional bridge between innate and adaptive immunity, being a system that "complements" the function of antibodies in the modulation of an integrated host defense (20). The C is comprised of over 50 plasma and cell surface proteins, including activation effectors, regulators and cell surface receptors (21). These proteins are organized to take part in three independent but interactive activation pathways (**Figure 1**): classical, lectin and alternative, converging on the common activation of the major component C3 and in the production of proinflammatory mediators, opsonization, membrane attack complex (MAC) formation, and target cell lysis (22).

## **Classical Pathway**

The classical pathway is mainly triggered by the interaction between the C1 and immune complexes, apoptotic and necrotic cells (23). C1 complex consists of three sub-components: the classical pathway recognition molecule C1q and the serine proteases C1r and C1s. The activation process is initiated by C1q recognition of the Fc domain of an array of IgM or IgG bound to an antigen. The consequent C1q conformational change is responsible for the activation of C1r serine-protease activity (24), which, in turn, acts as a trigger to the proteolytic activity of C1s, inducing C4 and C2 cleavage into two active components (C4b and C2a) and two small fragments released in the fluid phase (C4a and C2b) (25). C4b is able to covalently bind to membrane proteins and carbohydrates (26), and non-covalently to C2a inducing the C4b2a complex formation, also known as C3 convertase of the classical pathway (25). C3 convertase, attached to the target surface, is responsible for C3 cleavage into C3b and C3a anaphylatoxin (Figure 1).

## **Lectin Pathway**

The lectin pathway is an antibody-independent route in which mannan-binding lectin (MBL), ficolins (ficolin-1, -2 and -3) or collectin-11 act as pattern-recognition molecules through their binding to mannose residues and other carbohydrate ligands in the form of pathogen-associated molecular patterns (PAMPs), or damage-associated molecular patterns (DAMPs) (27). This binding triggers MBL interaction with MBL-associated serine proteases (MASPs), a family of serine proteases including three enzymatic proteins (MASP-1, MASP-2 and MASP-3) and two non-enzymatic factors (Map19 and Map44). MASP-3 is mainly involved in the alternative pathway activation (28), whilst MASP-2 activation by MASP-1, forming a dimer, activate the lectin pathway *via* C4 and C2 cleavage and C3 convertase formation (29).

## **Alternative Pathway**

The initiation of the alternative pathway is independent of immune complexes, being constitutively active at basal levels in the so-called "tick-over" mechanism and assuring a rapid and robust C activation in the presence of pathogens (30). Under normal physiological conditions, C3 undergoes constant low-grade activation by spontaneous hydrolysis, producing the C3(H<sub>2</sub>O) molecule, which is rapidly inactivated in the circulation (31). After its binding to C3 (H<sub>2</sub>O), the plasma protein factor B (FB) is cleaved by the serine protease factor D (FD), losing the small fragment Ba, whilst the residual fragment Bb remains bound to C3(H<sub>2</sub>O) forming the fluid phase C3 convertase, C3(H<sub>2</sub>O)Bb. C3(H<sub>2</sub>O)Bb has the ability to cleave large amounts of C3 molecules into C3a and C3b. C3b is partly inactivated by hydrolysis; however, the interaction of C3b with surface components of microbial agents and damaged host cells can accelerate the alternative pathway and induce the association with FB, further cleaved by FD, and generation of the amplification loop convertase C3bBb. The rapid amplification loop is also boosted by the C3b molecules generated by either the classical or lectin pathway. The alternative pathway C3 convertases are highly labile, so they need to be stabilized by an up-regulator called factor P or properdin, increasing its half-life by 10-fold (32).



FIGURE 1 | Overview of the complement system (C) and its interplay with the coagulation and kallikrein-kinin pathways. The activation of C via three pathways: classical, alternative and lectin. The recognition and activation phases in each pathway converge on the formation of C3 convertase. The amplification phase is initiated when the C3 convertase cleaves C3, and C3b attachment to C3 convertases changes their substrate specificity, allowing them to become C5 convertase. The C5 convertase cleaves C5, starting the last phase (Terminal-Attack phase), in which C5b forms the membrane-attack complex with C6, C7, C8, and C9, resulting in cell perturbation. Kallikrein exerts its activity on C activation in all these three phases. Factor (F) XIIa activates the classical pathway, whereas plasmin enhances the amplification loop. Thrombin, FIXa, FXa, FXIa, and kallikrein directly activate C5. AT, Antithrombin; C1-INH, C1-inhibitor; F, Factor; MAC, Membrane attack complex; MASP, MBL associated serine protease; MBL, Mannose binding lectin; iTCC, Inactive terminal complement complex; TAFI, Thrombin activatable fibrinolysis inhibitor.

## Membrane Attack Complex Formation

All the three C activation pathways converge on the common C3 convertase formation and C3 cleavage into the anaphylatoxin C3a and the opsonin C3b (19, 33). The incorporation of the C3b fragment in the C3 convertase gives rise to the production of C5 convertase, which in turn, cleaves C5 to yield C5b and the anaphylatoxin C5a. Then, C5b sequentially interacts with the terminal C components C6, C7, C8, and C9, resulting in the formation of the C5b-9 terminal C complex (TCC). If TCC is fully inserted into a cell plasma membrane, it is called the MAC. MAC-mediated cell death can release DAMPs, which can result in further C activation. In many clinical conditions associated with massive C activation, in plasma or other fluids, the formation of a cytolytically inactive TCC (iTCC)' also called soluble C5b-9 (sC5b-9) complex, is helpful in several pro-inflammatory responses acting directly on endothelium (34).

## Main Regulators of Complement Activation

Abnormal C activation can be responsible for severe inflammatory conditions, as hereditary angioedema, paroxysmal nocturnal haemoglobinuria, and haemolytic uremic syndrome (25, 35). Several cell membrane-bound or soluble C regulators exist to control this system. C1-inhibitor (C1-INH) inhibits both classical and lectin pathway initiation through its action on C1r, C1s and MASP-2, disassembling C1 or ficolin/MBL-MASP complexes' respectively (36, 37). Moreover, C1-INH is also able to downregulate the alternative pathway convertase by interacting with C3b and inhibiting its binding to FB (38). At the C3 convertase level, the main regulator is FI (39)' a serine protease responsible for C3b and C4b cleavage. It needs several cofactors for enhancing its activity. C4b-binding protein (C4bp) represents the main cofactor of FI for C4b cleavage (40). Another cofactor of FI is FH that can bind to polyanionic molecules exposed on the membrane such as glycosaminoglycans, heparin and sialic acid. FH interacts with C3b promoting its hydrolysis by FI (39). In addition, the membrane-bound regulatory proteins, C Receptor 1 (CR1, CD35) (41) and Decay Accelerating Factor (DAF; CD55) (42), participate in C3b and C4b degradation. Membrane Cofactor Protein (MCP; also called CD46) (43) is involved in the acceleration of their decay of the C3- and C5-convertases. CD59 (also known as protectin), by binding to C5b-8, limits the incorporation of the C9 molecules, and consequently, formation of the MAC (44). If the cell is protected from lysis by CD59 (45), the sublytic attack can induce the release of inflammatory mediators (46, 47). When C5b-7, C5b-8 or C5b-9 assemble in plasma, the binding of the plasma proteins' vitronectin (alternative names S-protein) (48) and clusterin (49) (also known as SP40.40)' can lead to iTCC formation, preventing insertion into lipid bilayers and MAC formation.

# ROLE OF COMPLEMENT SYSTEM IN VIRAL INFECTIONS INCLUDING SARS-CoV-2

Acting in concert, the three pathways of the C cascade are effective in targeting both cell-free viral particles and virus-

infected cells, boosting the anti-viral innate and adaptive immune responses (50). The antiviral activity of the C system usually takes place through four different, but not exclusive, mechanisms of action: C deposition and opsonization, MAC formation and viral cell lysis, production of pro-inflammatory anaphylatoxins, as well as enhancement of adaptive immunity (**Figure 2**) (51). The 'eat me' signal by C1q, C3b, and C4b components on the non-self-agents prompts opsonization and enhanced phagocytosis; this process can progress to MAC assembly and viral envelope lysis. Moreover, C3a and C5a anaphylatoxins are able to recruit neutrophils, mast cells, monocytes, macrophages, basophils, eosinophils, T and B cells, giving an important contribution to chemotaxis, NETosis, degranulation, production of cytokines, inflammation' and reactive oxygen species (ROS) production (52–54).

The most striking evidence supporting the importance of C in viral infection outcome is provided by the evolution of specific evasion mechanisms employed by viruses for subverting C (55). First of all, viruses can recruit and exploit soluble and membrane-bound host C regulators, as well as encode their own C regulatory proteins (56). Moreover, they are also capable of using C regulators and receptors for cellular entry or even to modulate C protein expression through the upregulation of C regulators or the downregulation of C activators (57).

It is now widely accepted that, during the initial stages of the infection, SARS-CoV-2 proteins are able to directly activate all three pathways of C (58): N protein is able to induce the MASP-2-mediated activation of lectin pathway (59) and S protein is responsible for the alternative pathway activation (60), whilst classical pathway is usually activated at advanced stages *via* immune complexes and C-reactive protein involvement (61, 62). Evidence so far suggests that C may be beneficial in the early stages of SARS-CoV-2 infection due to its participation in virus elimination; however, C activation may be severely harmful in later phases.

In COVID-19, the initial viral invasion phase is usually followed by an immunopathological phase, which is characterized by an uncontrolled immunological response causing pulmonary, and sometimes, systemic inflammation, in which C is also involved. In particular, recent findings indicate that excessive or deregulated C activation may occur in COVID-19 patients (62, 63), triggering anaphylatoxin generation and binding to their receptors (64), with subsequent inflammatory cell recruitment in the lungs and other organs (65). This contributes to cytokine storm, EC injury, intravascular coagulation and thrombosis (66, 67).

## ENDOTHELIITIS, COMPLEMENT SYSTEM, AND PATHOGENESIS OF COVID-19

COVID-19 pathogenesis is characterized by an initial virusinduced injury and consequent multi-organ failure, coupled with an intense inflammatory reaction, EC injury, and a prothrombotic coagulopathy with thrombotic events. The progression from mild to severe COVID-19 is characterized by the transition from an epithelial to an endothelial disease (14, 68).



FIGURE 2 | The recognition and binding of S protein to the ACE2 receptor are fundamental key events in SARS-CoV-2 invasion of the host cells. The engagement of ACE2 receptor and S protein of SARS-CoV-2 induces the fusion of virus and target cell. This process is orchestrated by the cleavage of S protein into S1 and S2 subunits *via* transmembrane serine protease priming known as TMPRSS. Subsequently, prompt viral replication occurs inside the infected cell, contributing to the dissemination of infection. The C as a first line of innate immune defence could recognize the virus. SARS-CoV-2 is capable of directly activating the C through the classical, alternative, and especially lectin pathways. Dysregulated C activation during the SARS-CoV-2 infection can cause serious damage to different organs. The blockade of C effectors has a potential therapeutic effect through the prevention of the recruitment of immune cell as well as immune activation. ACE2, Angiotensin-converting enzyme 2; LDA, Low-dose aspirin; LMWH, Low-molecular weight heparin; MAC, Membrane attack complex; MASP, MBL (mannose binding lectin) associated serine protease; SARS-CoV-2, Severe acute respiratory syndrome coronavirus 2; TMPRSS2, Transmembrane serine protease 2.

Indeed, ECs, playing a pivotal role in the regulation of immune response, initiation, and maintenance of inflammatory process (69), coagulation, and platelet function, are key players in various pathological manifestations associated with COVID-19 (**Figure 3**) (70, 71).

Direct viral infection of ECs, via SARS-CoV-2 receptors, ACE2 and TMPRSS2, present on their surface (7), is able to provoke endothelial dysfunction and disruption of vascular integrity, leading to hyperinflammation and hypercoagulability (72). The binding of SARS-CoV-2 to ACE2 hampers its enzymatic activity, with consequent enhanced vascular permeability (72) associated with activation of kallikrein-kinin system and bradykinin (BK) accumulation (7, 73). Furthermore, reduced ACE2 expression by binding of SARS-CoV-2 on ECs limits the degradation of des-Arg<sup>9</sup>-BK, the active metabolite of BK, into inactive peptides, increasing prothrombotic signaling via the activation of BK receptor 1, expressed during inflammatory conditions (72, 74). ACE2 downregulation determines angiotensin II accumulation, which enhances vascular permeability through AT1 receptor and promotes tissue damage, but also reduces Mas activation by angiotensin 1-7, supporting a local pro-inflammatory and prothrombotic EC phenotype (75, 76).

Not only a direct virus-dependent effect on ECs has been observed, but also host-specific factors seem to contribute to

systemic endothelial dysfunction in COVID-19. The C seems primarily involved in this process, the activation of its three pathways and MAC formation as contributors to EC swelling and even disruption (77). Indeed, in severe disease conditions, elevated levels of serum MAC (62), as well as a strong immunohistochemical staining for deposited C5b-9 in the microvasculature, have been detected (61) in co-localization studies with SARS-CoV-2 N protein (62, 78). MBL, MASP-2, C4a and C3 deposits have also been observed (78).

Immune complexes comprising SARS-CoV-2 specific antibodies and viral antigens may lead to EC injury through the activation of C1 complex of the classical pathway and induction of antibody-dependent cytotoxicity. Increased levels of C3a and C5a, due to C hyperactivation, amplify the vicious cycle of vascular integrity disruption, promoting infiltration of neutrophils which potentiates ROS production, degranulation and NETosis, ultimately provoking further injury to ECs (79). Elevated serum levels of C5a, the most potent C anaphylatoxin, have also been reported in severe COVID-19 patients, whereas circulating C5a levels in patients with mild manifestations are similar to those of the healthy controls (62). Furthermore, a close association of C5a-C5aR axis with inflammation and endotheliitis has been observed in the pathogenesis of severe COVID-19 (64).



FIGURE 3 | Summary of the elements involved in endotheliitis. COVID-19 pathogenesis is characterized by an initial virus-induced injury leading to multi-organ failure, coupled with an intense inflammatory reaction, endothelial cell (EC) injury, and a prothrombotic coagulopathy with thrombotic events. The hypercytokinemia induced by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) causes the expression by ECs of adhesion molecules such as intercellular adhesion molecule 1 (ICAM-1), vascular cell adhesion molecule 1 (VCAM-1) and P-selectin. Neutrophils are recruited and contribute to this process through the release of neutrophil extracellular traps (NETs), which directly activate factor XII and, thus, the contact-dependent pathway of coagulation. NETs also bind von Wilebrand factor (vWF) and help to recruit platelets. The feed-forward loop is amplified by the effect of P-selectin, wWF, and fibrinogen expression by ECs in response to hypercytokinemia, causing platelets to directly bind to ECs leading to their activation and hypercontractility, which can lead to disruption of cell-cell junctions and vascular leakage. ACE2, Angiotensin-converting enzyme 2; Ang II, Angiotensin II; BK, Bradykinin; MAC, Membrane attack complex; iTCC, Inactive terminal complement complex; ROS, Reactive oxygen species.

EC damage, characterized by disruption of cell-cell junctions and vascular leakage, exposes basement membrane to circulatory platelets, initiating platelet aggregation, hypercoagulable state and thrombotic events (72) (**Figure 3**), which are frequently observed in severe COVID-19 patients (80).

## COMPLEMENT SYSTEM AND ITS ROLE IN THE THROMBOTIC EVENTS IN COVID-19

The C activation has been associated with COVID-19-related coagulopathy and thromboembolia (81), suggesting an interplay between C and coagulation system (**Figure 1**). Interestingly, SARS-CoV-2 infection is able to induce the transcription of C (C1r, C1s, factor B and C3) and coagulation genes (fibrinogen) in pneumocytes and hepatocytes (81).

C and coagulation systems are evolutionarily linked in terms of functional similarities and shared structural motifs (82). The process of coagulation is activated through the contact (intrinsic) and the Tissue Factor (TF) (extrinsic) pathways, both converging on factor (F) X activation. The intrinsic pathway starts from FXII activation by negatively charged surfaces, such as phospholipids present on activated platelets and exposed subendothelial collagen (83) (**Figure 4**). The extrinsic pathway is responsible for a quick and efficient *in vivo* hemostasis, once activated by TF release by damaged cells or expression on the surface of activated monocytes, ECs, and other non-vascular cells. TF then converts FVII to FVIIa (82). The common factor X, upon activation, is then responsible for prothrombin (FII) activation in thrombin (FIIa), which in turn, triggers the formation of fibrin from the soluble fibrinogen (84) (**Figure 4**).

When generated, FXIIa is also responsible of the C activation *via* C1r and C1s (85), and that of the inflammatory kallikreinkinin pathway by converting pre-kallikrein into active plasma kallikrein, which cleaves FXII into FXIIa and high molecular weight kininogen to BK and the fibrinolytic system by plasma kallikrein activation of pro-urokinase into urokinase, which in turn, cleaves plasminogen into plasmin, an enzyme that degrades fibrin clots (86, 87) (**Figure 5**).

The coagulation system is also involved in inflammation and tissue remodelling through the interaction of coagulation proteases with four distinct protease-activated receptors (PAR1, PAR2, PAR3 and PAR4) (83). The interplay between thrombosis and inflammation is ensured by PAR1 (previously known as the thrombin receptor) cleavage, upon thrombin-PAR1 interaction, resulting into an overall pro-inflammatory state characterized by the release of P-selectin, von Willebrand factor (vWF) and the disruption of the endothelial barrier function (88).

### **Complement and Tissue Factor Expression**

TF (also known also thromboplastin), a transmembrane receptor for FVII/VIIa, acts as initiator of the extrinsic coagulation pathway (89). Several C activation factors induce TF expression on leukocytes and ECs (90–92). C5a increases TF activity in the circulation (93). C5a, by interacting with C5aR, mediates the expression of TF in neutrophils enhancing their procoagulant activity (93) The TF expression in monocytes can also be induced by the membrane insertion of the C5b-7 (94). In addition, the terminal components of the C cascade stimulate the synthesis and release of TF (90); in fact, sublytic MAC (92) and iTCC (91) have been shown to stimulate the expression of TF on ECs,



triggering a prothrombotic state through FVII-dependent activation of FX.

A C-linked release of TF has been observed in COVID-19, mainly due to a direct virus-dependent effect on ECs, in which viral infection of the endothelium provokes release of viral proteins able to activate C with consequent stimulation of TF production by neutrophils, monocytes and ECs, as well as causes endothelial injury that would expose subendothelial TF (66, 67, 95, 96).

# Complement and von Willebrand Factor Interaction

vWF is a complex multimeric plasma glycoprotein critical for normal haemostatic function; under physiological conditions, it is synthesized by ECs and stored in the Weibel–Palade bodies, and by megakaryocytes, being primarily stored in  $\alpha$ -granules of platelets, as ultra-large vWF (97, 98). vWF has two main roles in haemostasis: firstly, to recruit and tether platelets at sites of vascular injury, facilitating aggregation; secondly, vWF acts as a protective carrier molecule for procoagulant FVIII. The assembly of C5b-9 on human ECs results in the secretion of high molecular weight multimers of vWF and release of membrane particles from the EC surface, which express binding sites for FVa, supporting prothrombinase activity (99, 100). Moreover, ultra-large vWF offers an activating surface for the assembly of the alternative pathway convertase; FH is able to reduce ultralarge vWF to smaller forms (101).

Plasma vWF levels are significantly increased in patients with COVID-19 (102), due to EC activation, thereby facilitating recruitment and aggregation of platelets (103) and tethering of leucocytes to the vessel wall (104). A C-linked release of vWF has been observed in response to sublytic MAC addition to ECs (99, 105).



**FIGURE 5** | Interplay between coagulation, fibrinolytic, contact and complement systems. The kinin system is comprised of proteins that participate in the coagulation and inflammation. In the kinin/contact system activation may occur *via* auto-activation of FXII in contact with negatively charged surfaces, or the plasma kallikrein-kinin system, as activation in plasma may be driven by pre-kallikrein activation. Kinins are vasoactive peptides produced by the action of a specific protease, called plasma kallikrein on kininogens. Plasma kallikrein, through the cleavage of the plasma glycoprotein precursor high-molecular-weight kininogen (HMWK), produces bradykinin. Bradykinin (BK) causes increase in vascular permeability, contraction of smooth muscle, dilation of blood vessels, and pain when injected into the skin, with similar effects to histamine. It exerts profibrinolytic properties by stimulating release of tissue plasminogen activator from endothelial cells. BK is quickly inactivated by redundant membrane and soluble kininase. The coagulation intrinsic pathway of the coagulation system is composed by plasma proteins activated by FXII, a protein synthesized by the liver that can be activated by collagens, basal membrane and activated platelets. The coagulation system culminates in the formation of thrombin (FII) and in the formation of fibrin from the soluble fibrinogen. When generated, FXIIa is also responsible of the activation of other three systems: (a) *C, via* activation of C11 and C11; (b) inflammatory kallikrein-kinin pathway by converting pre-kallikrein activation, in turn cleaving plasma kallikrein, and kallikrein, and kallikrein, and kallikrein, and kallikrein, and kallikrein, and high molecular weight kininogen to BK; and (c) fibrinolytic system by plasma kallikrein activation, in turn cleaving plasminogen into plasmin, an enzyme that degrades fibrin clots. Several coagulation system enzymes, including thrombin, FIXa, FXa, FXIa, and kallikrein, directly activate C5. Kallikrein also activates C3 a

## **Complement and Thrombin Generation**

MASPs associated with surface-bound MBL or ficolins exert their activity also in the coagulation process by cleaving coagulation factors, despite a slower kinetics as compared to coagulation proteases (27). MASP-1 and MASP-2 directly cleave prothrombin causing thrombin generation; MASP-1 is also able to cleave fibrinogen to generate fibrin monomers. Furthermore, MASP-1 is able to induce the formation of FXIIIa, a fibrin stabilizing factor, and that of thrombin-activatable fibrinolysis inhibitor (TAFI), an attenuator of the fibrinolytic rate. Among thrombin inhibitors, C1-INH and anti-thrombin III+heparin exert their inhibitory effect also on MASP-1 and MASP-2, whereas α2-macroglobulin does not abolish lectin pathway activation (37). The key TF Pathway Inhibitor (TFPI), expressed by microvascular ECs, can interfere with the lectin pathway by blocking MASP-2 (106). In addition to MASPs, sC5b-9 can also be involved in thrombin generation and in the flipping of EC and platelet phospholipid membranes, supporting prothrombinase assembly (FXa and FVa binding) (107).

It has been shown that thrombin can cleave C3 and C5, producing biologically active anaphylatoxins, C3a and C5a (108). Within the context of an overall increased thrombin generation, as observed in COVID-19, this mechanism accounts for the amplification of the feed-forward loop between C and coagulation, linking both cascades *via* multiple direct interactions.

## **Reciprocal Regulation of Complement and Coagulation Pathways**

Several C factors are able to control different steps of the coagulation pathway and vice versa (**Figures 1**, **5**). C1-INH can exert a dual inhibitory function on C and coagulation, by respectively inhibiting C1 complex and FXIIa, which is also responsible for C activation through C1r and C1s; regulation of contact system is also C1-INH-dependent through inactivation of plasma kallikrein (109). Furthermore, C1-INH is able to inhibit FXIa (110) and thrombin (111). Interestingly, the interaction of SARS-CoV proteins with C1-INH during viral infection determines C1-INH blockage (112, 113). Low C1-INH serum levels were shown as a predictive factor of progression to respiratory distress in COVID-19 (114).

Among coagulation regulators, an important cross-talk is represented by thrombomodulin (TM), a cell-bound regulator with anticoagulant properties, which can interact with FH enhancing its regulatory activity and consequently accelerating the degradation of C3b into inactive iC3b (115). Several coagulation system enzymes, such as thrombin, factor IXa, factor Xa, factor XIa, and kallikrein, are responsible for direct C5 activation. Kallikrein also activates C3 and factor B. In animal models of arterial and venous thrombosis, plasmin also exerts C5 convertase activity (116) (Figure 5). TAFI, also known as plasma carboxypeptidase B2 or R, suppresses fibrinolysis in physiological conditions. Following activation by thrombin/TM and/or plasmin, it can inactivate C3a and C5a, potentiating the action of the constitutive carboxypeptidase N as a supplementary inhibitor (117). Interestingly, markedly elevated circulating TAFI levels are reported in COVID-19 patients, being implicated in microvascular fibrin deposition (118).

## **Complement-Platelet Crosstalk**

Platelet activation is also responsible for the release of C components, including C1q, C3, C4, and C5b-9 (119, 120). C3a, C5a and, to a lesser extent, C4a, promote platelet aggregation and activation through their binding to cognate receptors C3aR or C5aR1 and C5aR2 (121, 122). Despite the absence of a specifically recognized receptor for C4a, it is able to bind to PAR1 and PAR4, participating in platelet activation. Activated platelets expose P-selectin, which is a receptor for C3b, providing a site for the assembly of the alternative pathway C3 convertase (123). Concurrently, C5a and the C5b-9 induce the expression of P-selectin and vWF by ECs (105), promoting platelet adhesion and aggregation, and release of TM from cell surface (124), triggering the coagulation cascade.

MAC is able to activate platelets and promote platelet aggregation (107, 125). The assembly of MAC on human platelets also results in a dose-dependent increase in the binding of FVa and FXa, which increases platelet prothrombinase activity.

# PRE-ECLAMPSIA IN COVID-19 AND THE ROLE OF COMPLEMENT SYSTEM

PE is a frequent pregnancy-related disease (2-6% incidence in healthy nulliparous women), contributing to 20-25% of perinatal mortality, characterized by the onset of hypertension, proteinuria and multi-organ impairments. PE has been defined by the International Society for the Study of Hypertension in Pregnancy as the manifestation of arterial hypertension and proteinuria (300 mg/d) occurring after 20 weeks of gestation or as new-onset arterial hypertension combined with organ dysfunctions, such as renal failure, liver dysfunction, hematological or neurological abnormalities, intrauterine growth restriction, or uteroplacental insufficiency (126, 127). Approximately 10-20% of women with severe PE develop HELLP (hemolysis, elevated liver enzymes and low platelets) syndrome as a further complication (128).

From an etiological perspective, PE is commonly distinct in early onset PE (EOPE) and late onset PE (LOPE), occurring before or after 34 weeks of gestation, respectively. EOPE is associated with poor trophoblast invasion and inadequate arterial remodeling and its pathophysiology is considered to be related to the placental development (129, 130). In physiological placentation, the maternal spiral arteries are invaded by extravillous trophoblast cells, which through a process named endovascular migration, gradually replace decidual ECs, leading to spiral artery remodeling (131-133) (Figure 6). When this process is impaired, it causes placental hypo-perfusion, tissue ischemia, vascular endothelium dysfunction, microangiopathic thrombosis, oxidative stress, and inflammatory response (134-136). In general, the cause of PE can be ascribed to an excessive maternal systemic inflammation in response to pregnancy through both innate and adaptive immune activation (137). Excessive inflammation during PE has also been confirmed through the measurement of procalcitonin, as a marker of sepsis or severe inflammation (138). The cause of the LOPE is more debatable, but the basal inflammatory state of the mother



**FIGURE 6** | Schematic representation of the placenta architecture in pre-eclampsia (PE). Overview of the structural organization of the placenta and feto/maternal interface during the abnormal placentation in PE. The diagram was designed using the Blender 3D (Blender Foundation, Stichting Blender Foundation, Buikslotermeerplein, Amsterdam, the Netherlands). Edited with permission from Balduit et al. (133).

and the incompatibility between maternal supply and metabolic needs of the developing fetus (130) are considered key precipitating factors.

## Important Contribution of the Complement System in the Pathogenesis of Pre-Eclampsia

The C is an important component of the inflammatory process in PE (17, 18). Increased levels of C components and their activation products, including C1q, C3a, C5a and C5b-9 complex, in the circulation of PE patients as compared to normal pregnancy have been reported (17, 139–141), although the results in this field are quite discordant (141). C activation products have also been found in the urine of severe PE patients and are considered a marker of C-mediated renal damage (142). Elevated levels of the activation fragment Bb of the alternative pathway have been proposed as a predictive marker for the development of PE (143). Furthermore, increased C activation was demonstrated in several models of PE (144–146).

## Pre-Eclampsia: Diagnostic Criteria

Despite not sharing common pathological pathways, EOPE and LOPE are diagnosed by mutual clinical criteria (147). Uteroplacental under-perfusion, measured by the uterine artery

pulsatility index (UtAPI), induces placental ischemia, which in turn, gives rise to oxidative-inflammation cascade activation, increased production of antiangiogenic factors, such as soluble fms-like tyrosine kinase 1 (sFlt-1) and soluble endoglin (sEng), and reduced production of angiogenic factors, such as placental growth factor (PlGF) (148, 149), both in the intrauterine environment and maternal endothelium. Interestingly, low maternal circulating levels of PIGF are measured prior to the clinical manifestation of PE and intrauterine growth restriction, as a marker of abnormal placentation (150). sFlt-1 is a soluble form of the vascular-endothelial growth factor (VEGF) and a receptor for PIGF. In the maternal circulation, sFlt-1 is able to bind to free VEGF and PIGF, reducing their bioavailability for membrane receptors (151). The use of a diagnostic test based on sFlt-1/PlGF ratio for the prediction of the short-term risk of PE may be a useful tool for patient management (152). In fact, with a sFlt-1/PlGF ratio of  $\leq$  38 PE occurrence in the next week can be excluded with a negative predictive value of 99.3% (97.9% for ruling out within 2 weeks), whereas a ratio of >38 is associated to an enhanced risk of developing PE in the next 4 weeks (152, 153).

## Pre-Eclampsia in COVID-19 Pregnancy

Despite similar clinical manifestations between pregnant and non-pregnant women with COVID-19 (154), an increased rate of preterm delivery, PE, and cesarean section has been noticed in COVID-19 pregnant women (155-158). The INTERCOVID prospective longitudinal study showed an increased incidence of PE in pregnant women with COVID-19 (8.1%), as compared with non-diagnosed COVID-19 (4.4%), especially in nulliparous women; the association appeared to be independent of preexisting conditions and other risk factors, such as obesity, diabetes and hypertension (159). Mendoza et al. reported that a PE-like syndrome could be manifested in some pregnancies with critical COVID-19, according to the presence of severe pneumonia (160). The authors used the term PE-like syndrome to highlight some diverging aspects as compared to classical PE; in their cohort' the PE-like syndrome patients showed PE clinical signs and symptoms but normal parameters, such as sFlt-1/PlGF ratio, UtAPI and LDH <600 IU/l. This information could improve the management of these pregnancies since PE-like syndrome alone may not be considered as an obstetric indication for delivery (160).

Differential diagnosis in COVID-19 pregnant women developing hypertension, thrombocytopenia, proteinuria, as well as increased levels of liver enzymes, might be challenging, since misdiagnosis may occur due to COVID-19 and PE overlapping clinical features (147). In fact, a recent study suggested a two-fold increased risk of developing hypertensive disorders of pregnancy in patients who manifested COVID-19, especially early in their pregnancy (161); this is considered as a consequence of COVID-19-mediated modulation of placental ACE2 expression (162).

A common denominator in the pathophysiology of PE or PElike syndrome and COVID-19 is the endothelial injury, due to disrupted placentation in PE (135, 163) and to directly or indirectly SARS-CoV-2-mediated damage to ECs in COVID-19 (71, 96, 164, 165). To clarify the extent of endothelial injury, the evaluation of two well-studied markers, such as the antiangiogenic factor sFlt-1 and the angiogenetic factor PIGF (166-168) can be useful. An imbalance between angiogenic and anti-angiogenic factors has been observed in COVID-19. Negro et al. indicated that an increase in sFlt-1 levels could be considered a good biomarker to predict survival and thrombotic events in COVID-19 patients (169). Smadja et al. considered PIGF increase as a relevant predictive factor for inhospital mortality to discriminate COVID-19 severity (170), whereas Giardini et al. have used sFlt-1/PlGF ratio as a tool to stratify the severity of endothelial dysfunction (171). Interestingly, in PE-like syndrome, Mendoza et al. noted a normal sFlt-1/PlGF ratio and UtAPI assessment (160), suggesting normal values of sFlt-1/PlGF and UtAPI in COVID-19 patients with normal early phase of placental implantation, despite their symptomatic manifestations. Thus, an interplay between two different phenomena can be proposed in the clinical setting of SARS-CoV-2-infected obstetric patients at risk of developing COVID-19. At one hand, COVID-19 may mimic PE, particularly in early pregnancy; on the other hand, an already established PE may act as a risk factor for the development of severe or critical COVID-19. These two separate clinical conditions merit further investigation when clinical and epidemiological criteria indicate patients at risk of one or the other, or both conditions (172).

# Common Pathophysiology of Pre-Eclampsia and Severe COVID-19

Endothelial damage can be responsible for multi-organ dysfunction in both PE and COVID-19 (165, 173), as well as for an augmented risk of non-cardiogenic pulmonary oedema and venous thromboembolism (165). An increased hypercoagulable state characterizes PE women as compared to normal pregnancies, showing a rise in factor VIII, vWF, thrombin-antithrombin complex, D-dimers, soluble fibrin and TM levels (174). At the same time, the fibrinolytic system also plays an important role in PE, considering the significant increase in plasma plasminogen activator inhibitor type-1 (PAI-1) (174, 175). COVID-19 is also linked to a thrombogenic coagulopathy with a wide range of manifestations. COVID-19 patients commonly manifest mild thrombocytopenia (176) and increased D dimer levels (177) in accordance with disease severity, whereas other coagulation measurements are more variable (178, 179). Another feature shared between PE and COVID-19 patients is represented by an overall inflammatory microenvironment, characterized by an increase in serum and placental levels of pro-inflammatory and decrease of anti-inflammatory cytokines (180, 181).

In the context of endothelial damage, three common players have been proposed in PE and severe COVID-19 pathophysiological mechanisms: NETosis, anti-phospholipid antibodies (aPLAs) and  $\alpha$ -1-antitrypsin (182). However, only the first two aspects concern an engagement of C.

There is an overwhelming case for the involvement of neutrophil extracellular traps (NETs) in immunothrombosis through several mechanisms: (a) NETs bind to vWF and recruit platelets; (b) NETs are able to trigger platelet activation; (c) NETs binding to TF provokes extrinsic pathway activation and thrombin generation; (d) Cleavage induced by neutrophil elastase and other neutrophil serine proteases inactivate anticoagulants, including TFPI and TM; and (e) NETs can directly support FXII activation mediated by platelet-derived polyphosphates (183). Interestingly, NETs have also been reported as contributors to PE pathogenesis, usually associated with maternal vasculitis, maternal-fetal interface hemorrhage and laminar decidual necrosis (184), and COVID-19-related endothelial damage and immunothrombosis through plateletneutrophil interactions (185). NETs formation due to SARS-CoV-2 infection contains C3, factor B and properdin, triggering and stabilizing the alternative pathway convertase (67). An hyper-inflammatory immune state in response to abnormal neutrophil activation and NET formation, together with excessive or deregulated C activation, contributes to the welldocumented clinical manifestations observed in severe COVID-19 (186). Moreover, NETs induce an excessive production of thrombin and the subsequent generation of C3a and C5a (108, 186). Hence, a feed-forward loop beginning with C activation may proceed with NETosis, consequently increased thrombin production, further stimulation of the C, and enhanced NET formation (67).

Another potential link between PE and COVID-19 is the presence of anti-phospholipid antibodies (aPLAs), since they have been indicated as an important risk factor for PE, especially EOPE (187). A recent study has reported elevated aPLA levels in nearly 52% of COVID-19 patients (188). In placenta, aPLAs promote platelet and EC activation, directly inducing procoagulant activity by interacting with factors of the coagulation pathway. This activity, however, was greatly reduced in C3 gene knock-out mice (189). Anti- $\beta$  2-glycoprotein-I, the primary pathogenic antibody in anti-phospholipid syndrome (190), is associated with an increased C activation (191), amplifying the production of other mediators of effector cell activation, including C3a, C5a, and MAC, with consequent thrombosis, tissue hypoxia, and inflammation within the placenta.

Interestingly, several findings suggest that genetic susceptibility may be involved in the dysregulation of C activation frequently observed in the progression to moderate/ severe form of COVID-19 and in PE. Pathogenetic mutations or deletions in C factor and regulatory genes, which predispose to an increased C activation, have been identified both in pregnant women with PE/HELLP syndrome and in patient with COVID-19. C gene mutations were attributed to FH, MCP, FI and C3 (192–194). In COVID-19, gender differences were also observed in C-related variants, with women more genetically susceptible to C dysregulation (194).

## THERAPEUTIC CONSIDERATIONS

Emerging evidence suggests that C is constantly activated in severe COVID-19 as well as PE; due to its ubiquity, potency and rapidity, it is reasonably considered as a potential drug target.

At present, the only effective treatment for PE remains parturition, since therapeutic approaches are mainly symptomatic. However,

several preventative therapies are effective if administered early in the pregnancy (before 16 weeks of gestation): Low-dose Aspirin (LDA) and low-molecular weight heparin (LMWH) are the most common preventive treatments for PE. Both these drugs also modulate the C activity: LDA is able to down-regulate the levels of C3 and factor B expression in placenta (195, 196), whereas LMWH inhibits the activation of the alternative pathway and directly C5a (197-199). For its anti-thrombotic, anti-inflammatory, analgesic, and antipyretic effects, aspirin was also proposed in COVID-19 treatment, despite its clinical use for preventive or curative purposes has not been accepted yet (200-202). According to the beneficial anticoagulation effects of heparin observed in COVID-19 patients, with moderate or severe illness (203), pregnant women with severe COVID-19 should undergo thromboprophylaxis during hospitalization and at least until discharge (155). Moreover, in pregnant women with COVID-19, the use of steroids (dexamethasone followed by methylprednisolone) is recommended if clinically indicated (155). Several studies have reported the ability of methylprednisolone to inhibit C activation, particularly by acting on alternative pathway amplification (204, 205).

Other drugs are potentially used to prevent PE, especially targeting immunological conditions. For instance, it is well known that systemic lupus erythematosus (SLE) and antiphospholipid Syndrome (APS) predispose to a higher risk of developing PE. Hydroxychloroquine (HCQ), an antimalarial drug, is often used in the treatment of SLE (206), resulting in a lower incidence of PE (196, 206). The mechanisms of action of HCQ on the C is debatable, so is its efficacy in the treatment of COVID-19. Most clinical trials have failed to demonstrate the efficacy of HCQ treatment in COVID-19 (207).

Recently, Lefkou et al. reported that the use of LDA+LMWH in addition to pravastatin (HMG-CoA reductase inhibitor) in APS-affected women at risk of developing PE reduced the onset of adverse outcomes, increasing placenta perfusion, reducing PE onset and improving neonatal outcomes (208). In murine studies, pravastatin inhibited C activation (C5a) by increasing the expression of C inhibitor, DAF (209).

The first C inhibitor to be approved for clinical trials was eculizumab, a monoclonal antibody able to block C5 and decrease C5a and C5b-9 formation (210). The use of eculizumab for C5 inhibition is a reasonable therapeutic option also in PE (196, 210), as suggested by a case report of a patient with severe PE/HELLP at 26 weeks of gestation, showing an improvement of woman and foetus clinical endpoints after eculizumab treatment (pregnancy was prolonged by 17 days, resulting in a reduction of neonatal morbidity).

There are currently 13 clinical studies investigating the effect of C inhibitors in the treatment of severe forms of COVID-19, but none of them include pregnant patients. These are mainly C3 and C5 inhibitors, including zilucoplan, AMY101, APL-9, eculizumab, and ravulizumab (211–214). In particular, eculizumab and ruxolitinib (JAK1/2 inhibitor) treatment resulted in clinical improvement within 3 days; this may be also due to their inhibitory effect on pathways responsible for local and hepatic C synthesis, such as NF-κB and STAT1/2 (215). Compstatin-based C3 inhibitor, AMY-101, showed a satisfactory efficacy in the treatment of severe COVID-19 pneumonia (63, 212). The lectin pathway inhibitor, narsoplimab, which exerts its activity by blocking MASP-2, has been shown to prevent EC damage and thrombotic microangiopathy; recovery and survival was observed in all COVID-19 patients treated with narsoplimab (216).

Despite the promising results in clinical trials, the potential use of both C3- and C5-targeted therapies in COVID-19 patients is undermined by some limitations as comparted to selective inhibitors (217). First, the relevance of C3 inhibitors is dependent on the timing: blocking the activation of all three C pathways may undesirably reduce viral clearance during the early disease, whilst they may be useful in advanced phases preventing uncontrolled C activation (77). On the other hand, blocking C5 activation prevents the proinflammatory and prothrombotic actions of the terminal products of the C cascade (C5a and C5b-9) activated by SARS-CoV-2, whilst preserving the activity of early C components involved in viral clearance and activation of the adaptive immune response (218); although, this exposes patients to the risk of developing other infections, especially bacterial (219).

Thus, a selective blockage of C5a (vilobelimab) or C5aR (avdoralimab) could be more beneficial since it preserves the formation of C5b-9 complex as a crucial player in pathogen elimination (220, 221). The blockage of C5a-C5aR1 axis limits the infiltration of myeloid cells in damaged organs, hampers the production of pro-thrombotic factors by immune cells, platelets, and ECs, as well as prevents the excessive lung inflammation and endotheliitis associated with acute respiratory distress syndrome in patients with COVID-19 (58, 64, 222).

Another interesting therapeutic option under clinical investigation is the recombinant human C1–INH conestat alfa, which yielded encouraging results (223). Due to its multifaceted inhibitory action, C1–INH prevents all three C pathways' activation (36–38, 113) and inhibits components of the coagulation cascade, plasmin and kallikrein, reducing C-driven inflammation and coagulation (113).

## REFERENCES

- Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, et al. A Novel Coronavirus From Patients With Pneumonia in China, 2019. N Engl J Med (2020) 382 (8):727–33. doi: 10.1056/NEJMoa2001017
- Lai CC, Shih TP, Ko WC, Tang HJ, Hsueh PR. Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) and Coronavirus Disease-2019 (COVID-19): The Epidemic and the Challenges. *Int J Antimicrob Agents* (2020) 55(3):105924. doi: 10.1016/j.ijantimicag.2020.105924
- Lu L, Zhong W, Bian Z, Li Z, Zhang K, Liang B, et al. A Comparison of Mortality-Related Risk Factors of COVID-19, SARS, and MERS: A Systematic Review and Meta-Analysis. J Infect (2020) 81(4):e18–25. doi: 10.1016/j.jinf.2020.07.002
- Paules CI, Marston HD, Fauci AS. Coronavirus Infections-More Than Just the Common Cold. JAMA (2020) 323(8):707–8. doi: 10.1001/ jama.2020.0757
- Mousavizadeh L, Ghasemi S. Genotype and Phenotype of COVID-19: Their Roles in Pathogenesis. J Microbiol Immunol Infect (2021) 54(2):159–63. doi: 10.1016/j.jmii.2020.03.022
- Hu B, Guo H, Zhou P, Shi ZL. Characteristics of SARS-CoV-2 and COVID-19. Nat Rev Microbiol (2021) 19(3):141–54. doi: 10.1038/s41579-020-00459-7

## **CONCLUSIONS AND PERSPECTIVES**

A link between PE and COVID-19 has clearly emerged due to overarching pathophysiological mechanisms involved, which are triggered by an interplay between C and coagulation. Mothers or women who have recovered from COVID-19 need to be monitored closely during subsequent pregnancies for immune deviations. A very important question that is not yet solved and described in the use of C inhibitors for COVID-19 patients is the definition of the right window of opportunity for treatment. In these high-risk patients, with a major genetic susceptibility associated to complement polymorphisms, genetic testing might be an important tool aiming at intensified monitoring and early initiation of specific treatment with C inhibitors. Furthermore, the exact role of the C in severe COVID-19 development is yet to be clarified, which includes C-driven cytokines and viral protein activation of C-coagulation crosstalk.

## **AUTHOR CONTRIBUTIONS**

CA, AM, AB, AA, GR, UK, and RB reviewed the literature and wrote sections of the review article. CA and AA created figures. UK critically reviewed the entire manuscript. All authors contributed to the article and approved the submitted version.

## FUNDING

This research was supported by Ferring COVID-19 Investigational Grant (GRAVISAR to RB) and by the Institute for Maternal and Child Health, IRCCS Burlo Garofolo, Trieste, Italy (RC24/19 to GR and 09/21 to CA). The funder Ferring Pharmaceuticals was not involved in the study design, collection, analysis, interpretation of data, the writing of this article or the decision to submit it for publication.

- Hoffmann M, Kleine-Weber H, Schroeder S, Kruger N, Herrler T, Erichsen S, et al. SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. *Cell* (2020) 181(2):271– 80.e8. doi: 10.1016/j.cell.2020.02.052
- Patel S, Rauf A, Khan H, Abu-Izneid T. Renin-Angiotensin-Aldosterone (RAAS): The Ubiquitous System for Homeostasis and Pathologies. *BioMed Pharmacother* (2017) 94:317–25. doi: 10.1016/j.biopha.2017.07.091
- Yan R, Zhang Y, Li Y, Xia L, Guo Y, Zhou Q. Structural Basis for the Recognition of SARS-CoV-2 by Full-Length Human ACE2. *Science* (2020) 367(6485):1444–8. doi: 10.1126/science.abb2762
- Varadarajan S, Balaji TM, Sarode SC, Sarode GS, Sharma NK, Gondivkar S, et al. EMMPRIN/BASIGIN as a Biological Modulator of Oral Cancer and COVID-19 Interaction: Novel Propositions. *Med Hypotheses* (2020) 143:110089. doi: 10.1016/j.mehy.2020.110089
- Cantuti-Castelvetri L, Ojha R, Pedro LD, Djannatian M, Franz J, Kuivanen S, et al. Neuropilin-1 Facilitates SARS-CoV-2 Cell Entry and Infectivity. *Science* (2020) 370(6518):856–60. doi: 10.1126/science.abd2985
- Bussani R, Schneider E, Zentilin L, Collesi C, Ali H, Braga L, et al. Persistence of Viral RNA, Pneumocyte Syncytia and Thrombosis Are Hallmarks of Advanced COVID-19 Pathology. *EBioMedicine* (2020) 61:103104. doi: 10.1016/j.ebiom.2020.103104

- Saraste J, Prydz K. Assembly and Cellular Exit of Coronaviruses: Hijacking an Unconventional Secretory Pathway From the Pre-Golgi Intermediate Compartment via the Golgi Ribbon to the Extracellular Space. *Cells* (2021) 10(3):503. doi: 10.3390/cells10030503
- Gupta A, Madhavan MV, Sehgal K, Nair N, Mahajan S, Sehrawat TS, et al. Extrapulmonary Manifestations of COVID-19. *Nat Med* (2020) 26(7):1017– 32. doi: 10.1038/s41591-020-0968-3
- Jamal M, Bangash HI, Habiba M, Lei Y, Xie T, Sun J, et al. Immune Dysregulation and System Pathology in COVID-19. *Virulence* (2021) 12 (1):918–36. doi: 10.1080/21505594.2021.1898790
- Garcia LF. Immune Response, Inflammation, and the Clinical Spectrum of COVID-19. Front Immunol (2020) 11:1441. doi: 10.3389/fimmu.2020.01441
- Derzsy Z, Prohaszka Z, Rigo J Jr, Fust G, Molvarec A. Activation of the Complement System in Normal Pregnancy and Preeclampsia. *Mol Immunol* (2010) 47(7-8):1500–6. doi: 10.1016/j.molimm.2010.01.021
- Girardi G, Bulla R, Salmon JE, Tedesco F. The Complement System in the Pathophysiology of Pregnancy. *Mol Immunol* (2006) 43(1-2):68–77. doi: 10.1016/j.molimm.2005.06.017
- Walport MJ. Complement. First of Two Parts. N Engl J Med (2001) 344 (14):1058–66. doi: 10.1056/NEJM200104053441406
- Hajishengallis G, Reis ES, Mastellos DC, Ricklin D, Lambris JD. Novel Mechanisms and Functions of Complement. *Nat Immunol* (2017) 18 (12):1288–98. doi: 10.1038/ni.3858
- Ghebrehiwet B. The Complement System: An Evolution in Progress. F1000Res (2016) 5:2840. doi: 10.12688/f1000research.10065.1
- Merle NS, Church SE, Fremeaux-Bacchi V, Roumenina LT. Complement System Part I - Molecular Mechanisms of Activation and Regulation. *Front Immunol* (2015) 6:262. doi: 10.3389/fimmu.2015.00262
- Mortensen SA, Sander B, Jensen RK, Pedersen JS, Golas MM, Jensenius JC, et al. Structure and Activation of C1, the Complex Initiating the Classical Pathway of the Complement Cascade. *Proc Natl Acad Sci USA* (2017) 114 (5):986–91. doi: 10.1073/pnas.1616998114
- Lu J, Kishore U. C1 Complex: An Adaptable Proteolytic Module for Complement and Non-Complement Functions. *Front Immunol* (2017) 8:592. doi: 10.3389/fimmu.2017.00592
- Noris M, Remuzzi G. Overview of Complement Activation and Regulation. Semin Nephrol (2013) 33(6):479–92. doi: 10.1016/j.semnephrol.2013.08.001
- Ehrnthaller C, Ignatius A, Gebhard F, Huber-Lang M. New Insights of an Old Defense System: Structure, Function, and Clinical Relevance of the Complement System. *Mol Med* (2011) 17(3-4):317–29. doi: 10.2119/ molmed.2010.00149
- Garred P, Genster N, Pilely K, Bayarri-Olmos R, Rosbjerg A, Ma YJ, et al. A Journey Through the Lectin Pathway of Complement-MBL and Beyond. *Immunol Rev* (2016) 274(1):74–97. doi: 10.1111/imr.12468
- Iwaki D, Kanno K, Takahashi M, Endo Y, Matsushita M, Fujita T. The Role of Mannose-Binding Lectin-Associated Serine Protease-3 in Activation of the Alternative Complement Pathway. J Immunol (2011) 187(7):3751–8. doi: 10.4049/jimmunol.1100280
- Heja D, Kocsis A, Dobo J, Szilagyi K, Szasz R, Zavodszky P, et al. Revised Mechanism of Complement Lectin-Pathway Activation Revealing the Role of Serine Protease MASP-1 as the Exclusive Activator of MASP-2. *Proc Natl Acad Sci USA* (2012) 109(26):10498–503. doi: 10.1073/pnas.1202588109
- Dunkelberger JR, Song WC. Complement and Its Role in Innate and Adaptive Immune Responses. *Cell Res* (2010) 20(1):34–50. doi: 10.1038/ cr.2009.139
- Fishelson Z, Pangburn MK, Muller-Eberhard HJ. Characterization of the Initial C3 Convertase of the Alternative Pathway of Human Complement. *J Immunol* (1984) 132(3):1430–4.
- Lesher AM, Nilsson B, Song WC. Properdin in Complement Activation and Tissue Injury. Mol Immunol (2013) 56(3):191–8. doi: 10.1016/j.molimm.2013.06.002
- Walport MJ. Complement. Second of Two Parts. N Engl J Med (2001) 344 (15):1140–4. doi: 10.1056/NEJM200104123441506
- 34. Dobrina A, Pausa M, Fischetti F, Bulla R, Vecile E, Ferrero E, et al. Cytolytically Inactive Terminal Complement Complex Causes Transendothelial Migration of Polymorphonuclear Leukocytes In Vitro and In Vivo. Blood (2002) 99(1):185–92. doi: 10.1182/blood.v99.1.185
- Wong EKS, Kavanagh D. Diseases of Complement Dysregulation-An Overview. Semin Immunopathol (2018) 40(1):49–64. doi: 10.1007/s00281-017-0663-8

- 36. Kajdacsi E, Jandrasics Z, Veszeli N, Mako V, Koncz A, Gulyas D, et al. Patterns of C1-Inhibitor/Plasma Serine Protease Complexes in Healthy Humans and in Hereditary Angioedema Patients. *Front Immunol* (2020) 11:794. doi: 10.3389/fimmu.2020.00794
- Parej K, Dobo J, Zavodszky P, Gal P. The Control of the Complement Lectin Pathway Activation Revisited: Both C1-Inhibitor and Antithrombin Are Likely Physiological Inhibitors, While Alpha2-Macroglobulin Is Not. *Mol Immunol* (2013) 54(3-4):415–22. doi: 10.1016/j.molimm.2013.01.009
- Jiang H, Wagner E, Zhang H, Frank MM. Complement 1 Inhibitor Is a Regulator of the Alternative Complement Pathway. J Exp Med (2001) 194 (11):1609–16. doi: 10.1084/jem.194.11.1609
- Parente R, Clark SJ, Inforzato A, Day AJ. Complement Factor H in Host Defense and Immune Evasion. *Cell Mol Life Sci* (2017) 74(9):1605–24. doi: 10.1007/s00018-016-2418-4
- Fujita T, Gigli I, Nussenzweig V. Human C4-Binding Protein. II. Role in Proteolysis of C4b by C3b-Inactivator. J Exp Med (1978) 148(4):1044–51. doi: 10.1084/jem.148.4.1044
- Iida K, Nussenzweig V. Complement Receptor Is an Inhibitor of the Complement Cascade. J Exp Med (1981) 153(5):1138–50. doi: 10.1084/jem.153.5.1138
- 42. Nicholson-Weller A, Wang CE. Structure and Function of Decay Accelerating Factor CD55. J Lab Clin Med (1994) 123(4):485–91.
- Liszewski MK, Post TW, Atkinson JP. Membrane Cofactor Protein (MCP or CD46): Newest Member of the Regulators of Complement Activation Gene Cluster. Annu Rev Immunol (1991) 9:431–55. doi: 10.1146/annurev.iy.09. 040191.002243
- 44. Tedesco F, Bulla R, Fischetti F. Terminal Complement Complex: Regulation of Formation and Pathophysiological Functions. In: J Szebeni, editor. *The Complement System: Novel Roles in Health and Disease*. Boston, MA: Springer US (2004). p. 97–127.
- Morgan BP, Meri S. Membrane Proteins That Protect Against Complement Lysis. Springer Semin Immunopathol (1994) 15(4):369–96. doi: 10.1007/BF01837366
- Morgan BP, Campbell AK. The Recovery of Human Polymorphonuclear Leucocytes From Sublytic Complement Attack Is Mediated by Changes in Intracellular Free Calcium. *Biochem J* (1985) 231(1):205–8. doi: 10.1042/ bj2310205
- Zhang J, Li Y, Shan K, Wang L, Qiu W, Lu Y, et al. Sublytic C5b-9 Induces IL-6 and TGF-Beta1 Production by Glomerular Mesangial Cells in Rat Thy-1 Nephritis Through P300-Mediated C/EBPbeta Acetylation. FASEB J (2014) 28(3):1511–25. doi: 10.1096/fj.13-242693
- Podack ER, Kolb WP, Muller-Eberhard HJ. The C5b-6 Complex: Formation, Isolation, and Inhibition of Its Activity by Lipoprotein and the S-Protein of Human Serum. J Immunol (1978) 120(6):1841–8.
- Murphy BF, Kirszbaum L, Walker ID, d'Apice AJ. SP-40,40, a Newly Identified Normal Human Serum Protein Found in the SC5b-9 Complex of Complement and in the Immune Deposits in Glomerulonephritis. J Clin Invest (1988) 81(6):1858–64. doi: 10.1172/JCI113531
- Murugaiah V, Varghese PM, Beirag N, De Cordova S, Sim RB, Kishore U. Complement Proteins as Soluble Pattern Recognition Receptors for Pathogenic Viruses. Viruses (2021) 13(5):824. doi: 10.3390/v13050824
- Mellors J, Tipton T, Longet S, Carroll M. Viral Evasion of the Complement System and Its Importance for Vaccines and Therapeutics. *Front Immunol* (2020) 11:1450. doi: 10.3389/fimmu.2020.01450
- Markiewski MM, Lambris JD. The Role of Complement in Inflammatory Diseases From Behind the Scenes Into the Spotlight. *Am J Pathol* (2007) 171 (3):715–27. doi: 10.2353/ajpath.2007.070166
- 53. Köhl J. Self, Non-Self, and Danger: A Complementary View. Adv Exp Med Biol (2006) 586:71–94. doi: 10.1007/0-387-34134-X\_6
- Zhou W. The New Face of Anaphylatoxins in Immune Regulation. Immunobiology (2012) 217(2):225–34. doi: 10.1016/j.imbio.2011.07.016
- Agrawal P, Nawadkar R, Ojha H, Kumar J, Sahu A. Complement Evasion Strategies of Viruses: An Overview. *Front Microbiol* (2017) 8:1117. doi: 10.3389/fmicb.2017.01117
- Sinha A, Singh AK, Kadni TS, Mullick J, Sahu A. Virus-Encoded Complement Regulators: Current Status. *Viruses* (2021) 13(2):208. doi: 10.3390/v13020208
- Agrawal P, Sharma S, Pal P, Ojha H, Mullick J, Sahu A. The Imitation Game: A Viral Strategy to Subvert the Complement System. *FEBS Lett* (2020) 594 (16):2518–42. doi: 10.1002/1873-3468.13856

- Noris M, Benigni A, Remuzzi G. The Case of Complement Activation in COVID-19 Multiorgan Impact. *Kidney Int* (2020) 98(2):314–22. doi: 10.1016/j.kint.2020.05.013
- Ali YM, Ferrari M, Lynch NJ, Yaseen S, Dudler T, Gragerov S, et al. Lectin Pathway Mediates Complement Activation by SARS-CoV-2 Proteins. Front Immunol (2021) 12:714511. doi: 10.3389/fimmu.2021.714511
- Yu J, Yuan X, Chen H, Chaturvedi S, Braunstein EM, Brodsky RA. Direct Activation of the Alternative Complement Pathway by SARS-CoV-2 Spike Proteins Is Blocked by Factor D Inhibition. *Blood* (2020) 136(18):2080–9. doi: 10.1182/blood.2020008248
- Macor P, Durigutto P, Mangogna A, Bussani R, De Maso L, D'Errico S, et al. Multiple-Organ Complement Deposition on Vascular Endothelium in COVID-19 Patients. *Biomedicines* (2021) 9(8):1003. doi: 10.3390/ biomedicines9081003
- Gao T, Hu M, Zhang X, Li H, Zhu L, Liu H, et al. Highly Pathogenic Coronavirus N Protein Aggravates Lung Injury by MASP-2-Mediated Complement Over-Activation. *medRxiv* (2020). doi: 10.1101/ 2020.03.29.20041962 2020.03.29.20041962.
- Risitano AM, Mastellos DC, Huber-Lang M, Yancopoulou D, Garlanda C, Ciceri F, et al. Complement as a Target in COVID-19? *Nat Rev Immunol* (2020) 20(6):343–4. doi: 10.1038/s41577-020-0320-7
- Carvelli J, Demaria O, Vely F, Batista L, Chouaki Benmansour N, Fares J, et al. Association of COVID-19 Inflammation With Activation of the C5a-C5aR1 Axis. *Nature* (2020) 588(7836):146–50. doi: 10.1038/s41586-020-2600-6
- Fodil S, Annane D. Complement Inhibition and COVID-19: The Story So Far. Immunotargets Ther (2021) 10:273–84. doi: 10.2147/ITT.S284830
- Lo MW, Kemper C, Woodruff TM. COVID-19: Complement, Coagulation, and Collateral Damage. J Immunol (2020) 205(6):1488–95. doi: 10.4049/ jimmunol.2000644
- Java A, Apicelli AJ, Liszewski MK, Coler-Reilly A, Atkinson JP, Kim AH, et al. The Complement System in COVID-19: Friend and Foe? *JCI Insight* (2020) 5(15):e140711. doi: 10.1172/jci.insight.140711
- Chouaki Benmansour N, Carvelli J, Vivier E. Complement Cascade in Severe Forms of COVID-19: Recent Advances in Therapy. *Eur J Immunol* (2021) 51 (7):1652–9. doi: 10.1002/eji.202048959
- Goveia J, Rohlenova K, Taverna F, Treps L, Conradi LC, Pircher A, et al. An Integrated Gene Expression Landscape Profiling Approach to Identify Lung Tumor Endothelial Cell Heterogeneity and Angiogenic Candidates. *Cancer Cell* (2020) 37(1):21–36.e13. doi: 10.1016/j.ccell.2019.12.001
- Siddiqi HK, Libby P, Ridker PM. COVID-19 A Vascular Disease. Trends Cardiovasc Med (2021) 31(1):1–5. doi: 10.1016/j.tcm.2020.10.005
- Libby P, Luscher T. COVID-19 Is, in the End, an Endothelial Disease. Eur Heart J (2020) 41(32):3038–44. doi: 10.1093/eurheartj/ehaa623
- Teuwen LA, Geldhof V, Pasut A, Carmeliet P. COVID-19: The Vasculature Unleashed. Nat Rev Immunol (2020) 20(7):389–91. doi: 10.1038/s41577-020-0343-0
- Abassi Z, Skorecki K, Hamo-Giladi DB, Kruzel-Davila E, Heyman SN. Kinins and Chymase: The Forgotten Components of the Renin-Angiotensin System and Their Implications in COVID-19 Disease. *Am J Physiol Lung Cell Mol Physiol* (2021) 320(3):L422–9. doi: 10.1152/ajplung.00548.2020
- Dagnino APA, Campos MM, Silva RBM. Kinins and Their Receptors in Infectious Diseases. *Pharmaceut (Basel)* (2020) 13(9):215. doi: 10.3390/ ph13090215
- Simoes e Silva AC, Silveira KD, Ferreira AJ, Teixeira MM. ACE2, Angiotensin-(1-7) and Mas Receptor Axis in Inflammation and Fibrosis. *Br J Pharmacol* (2013) 169(3):477–92. doi: 10.1111/bph.12159
- Perico L, Benigni A, Casiraghi F, Ng LFP, Renia L, Remuzzi G. Immunity, Endothelial Injury and Complement-Induced Coagulopathy in COVID-19. Nat Rev Nephrol (2021) 17(1):46–64. doi: 10.1038/s41581-020-00357-4
- Song WC, FitzGerald GA. COVID-19, Microangiopathy, Hemostatic Activation, and Complement. J Clin Invest (2020) 130(8):3950–3. doi: 10.1172/JCI140183
- Magro C, Mulvey JJ, Berlin D, Nuovo G, Salvatore S, Harp J, et al. Complement Associated Microvascular Injury and Thrombosis in the Pathogenesis of Severe COVID-19 Infection: A Report of Five Cases. *Transl Res* (2020) 220:1–13. doi: 10.1016/j.trsl.2020.04.007
- 79. Barrett CD, Hsu AT, Ellson CD, Miyazawa BY, Kong YW, Greenwood JD, et al. Blood Clotting and Traumatic Injury With Shock Mediates

Complement-Dependent Neutrophil Priming for Extracellular ROS, ROS-Dependent Organ Injury and Coagulopathy. *Clin Exp Immunol* (2018) 194 (1):103–17. doi: 10.1111/cei.13166

- Panigada M, Bottino N, Tagliabue P, Grasselli G, Novembrino C, Chantarangkul V, et al. Hypercoagulability of COVID-19 Patients in Intensive Care Unit: A Report of Thromboelastography Findings and Other Parameters of Hemostasis. *J Thromb Haemost* (2020) 18(7):1738–42. doi: 10.1111/jth.14850
- Bosmann M. Complement Control for COVID-19. Sci Immunol (2021) 6 (59):eabj1014. doi: 10.1126/sciimmunol.abj1014
- Markiewski MM, Nilsson B, Ekdahl KN, Mollnes TE, Lambris JD. Complement and Coagulation: Strangers or Partners in Crime? *Trends Immunol* (2007) 28(4):184–92. doi: 10.1016/j.it.2007.02.006
- Madhusudhan T, Kerlin BA, Isermann B. The Emerging Role of Coagulation Proteases in Kidney Disease. Nat Rev Nephrol (2016) 12(2):94–109. doi: 10.1038/nrneph.2015.177
- Weidmann H, Heikaus L, Long AT, Naudin C, Schluter H, Renne T. The Plasma Contact System, a Protease Cascade at the Nexus of Inflammation, Coagulation and Immunity. *Biochim Biophys Acta Mol Cell Res* (2017) 1864 (11 Pt B):2118–27. doi: 10.1016/j.bbamcr.2017.07.009
- Amara U, Rittirsch D, Flierl M, Bruckner U, Klos A, Gebhard F, et al. Interaction Between the Coagulation and Complement System. Adv Exp Med Biol (2008) 632:71–9. doi: 10.1007/978-0-387-78952-1\_6
- Schmaier AH, Stavrou EX. Factor XII What's Important But Not Commonly Thought About. *Res Pract Thromb Haemost* (2019) 3(4):599– 606. doi: 10.1002/rth2.12235
- Maas C. Plasminflammation-An Emerging Pathway to Bradykinin Production. Front Immunol (2019) 10:2046. doi: 10.3389/fimmu.2019.02046
- Rovai ES, Alves T, Holzhausen M. Protease-Activated Receptor 1 as a Potential Therapeutic Target for COVID-19. *Exp Biol Med (Maywood)* (2021) 246(6):688–94. doi: 10.1177/1535370220978372
- Cimmino G, Cirillo P. Tissue Factor: Newer Concepts in Thrombosis and Its Role Beyond Thrombosis and Hemostasis. *Cardiovasc Diagn Ther* (2018) 8 (5):581–93. doi: 10.21037/cdt.2018.10.14
- Tedesco F, Fischetti F, Pausa M, Dobrina A, Sim RB, Daha MR. Complement-Endothelial Cell Interactions: Pathophysiological Implications. *Mol Immunol* (1999) 36(4-5):261-8. doi: 10.1016/s0161-5890(99)90054-8
- Tedesco F, Pausa M, Nardon E, Introna M, Mantovani A, Dobrina A. The Cytolytically Inactive Terminal Complement Complex Activates Endothelial Cells to Express Adhesion Molecules and Tissue Factor Procoagulant Activity. J Exp Med (1997) 185(9):1619–27. doi: 10.1084/jem.185.9.1619
- Saadi S, Holzknecht RA, Patte CP, Stern DM, Platt JL. Complement-Mediated Regulation of Tissue Factor Activity in Endothelium. J Exp Med (1995) 182(6):1807–14. doi: 10.1084/jem.182.6.1807
- Ritis K, Doumas M, Mastellos D, Micheli A, Giaglis S, Magotti P, et al. A Novel C5a Receptor-Tissue Factor Cross-Talk in Neutrophils Links Innate Immunity to Coagulation Pathways. *J Immunol* (2006) 177(7):4794–802. doi: 10.4049/jimmunol.177.7.4794
- 94. Langer F, Spath B, Fischer C, Stolz M, Ayuk FA, Kroger N, et al. Rapid Activation of Monocyte Tissue Factor by Antithymocyte Globulin Is Dependent on Complement and Protein Disulfide Isomerase. *Blood* (2013) 121(12):2324–35. doi: 10.1182/blood-2012-10-460493
- Escher R, Breakey N, Lammle B. Severe COVID-19 Infection Associated With Endothelial Activation. *Thromb Res* (2020) 190:62. doi: 10.1016/ j.thromres.2020.04.014
- Varga Z, Flammer AJ, Steiger P, Haberecker M, Andermatt R, Zinkernagel AS, et al. Endothelial Cell Infection and Endotheliitis in COVID-19. *Lancet* (2020) 395(10234):1417–8. doi: 10.1016/S0140-6736(20)30937-5
- Sadler JE. Biochemistry and Genetics of Von Willebrand Factor. Annu Rev Biochem (1998) 67:395–424. doi: 10.1146/annurev.biochem.67.1.395
- Patmore S, Dhami SPS, O'Sullivan JM. Von Willebrand Factor and Cancer; Metastasis and Coagulopathies. J Thromb Haemost (2020) 18(10):2444–56. doi: 10.1111/jth.14976
- Hattori R, Hamilton KK, McEver RP, Sims PJ. Complement Proteins C5b-9 Induce Secretion of High Molecular Weight Multimers of Endothelial Von Willebrand Factor and Translocation of Granule Membrane Protein GMP-140 to the Cell Surface. J Biol Chem (1989) 264(15):9053–60. doi: 10.1016/ S0021-9258(18)81901-9

- 100. Hamilton KK, Hattori R, Esmon CT, Sims PJ. Complement Proteins C5b-9 Induce Vesiculation of the Endothelial Plasma Membrane and Expose Catalytic Surface for Assembly of the Prothrombinase Enzyme Complex. J Biol Chem (1990) 265(7):3809–14. doi: 10.1016/S0021-9258 (19)39666-8
- 101. Nolasco L, Nolasco J, Feng S, Afshar-Kharghan V, Moake J. Human Complement Factor H Is a Reductase for Large Soluble Von Willebrand Factor Multimers-Brief Report. Arterioscler Thromb Vasc Biol (2013) 33 (11):2524–8. doi: 10.1161/ATVBAHA.113.302280
- 102. Goshua G, Pine AB, Meizlish ML, Chang CH, Zhang H, Bahel P, et al. Endotheliopathy in COVID-19-Associated Coagulopathy: Evidence From a Single-Centre, Cross-Sectional Study. *Lancet Haematol* (2020) 7(8):e575–82. doi: 10.1016/S2352-3026(20)30216-7
- Ruggeri ZM, Mendolicchio GL. Adhesion Mechanisms in Platelet Function. *Circ Res* (2007) 100(12):1673–85. doi: 10.1161/01.RES.0000267878.97021.ab
- 104. Petri B, Broermann A, Li H, Khandoga AG, Zarbock A, Krombach F, et al. Von Willebrand Factor Promotes Leukocyte Extravasation. *Blood* (2010) 116 (22):4712–9. doi: 10.1182/blood-2010-03-276311
- 105. Hattori R, Hamilton KK, Fugate RD, McEver RP, Sims PJ. Stimulated Secretion of Endothelial Von Willebrand Factor Is Accompanied by Rapid Redistribution to the Cell Surface of the Intracellular Granule Membrane Protein GMP-140. *J Biol Chem* (1989) 264(14):7768–71. doi: 10.1016/S0021-9258(18)83104-0
- 106. Keizer MP, Pouw RB, Kamp AM, Patiwael S, Marsman G, Hart MH, et al. TFPI Inhibits Lectin Pathway of Complement Activation by Direct Interaction With MASP-2. Eur J Immunol (2015) 45(2):544–50. doi: 10.1002/eji.201445070
- 107. Sims PJ, Faioni E, Wiedmer T, Shattil S. Complement Proteins C5b-9 Cause Release of Membrane Vesicles From the Platelet Surface That Are Enriched in the Membrane Receptor for Coagulation Factor Va and Express Prothrombinase Activity. J Biol Chem (1988) 263(34):18205–12. doi: 10.1016/S0021-9258(19)81346-7
- Amara U, Flierl MA, Rittirsch D, Klos A, Chen H, Acker B, et al. Molecular Intercommunication Between the Complement and Coagulation Systems. *J Immunol* (2010) 185(9):5628–36. doi: 10.4049/jimmunol.0903678
- Davis AE3rd. Biological Effects of C1 Inhibitor. Drug News Perspect (2004) 17(7):439–46. doi: 10.1358/dnp.2004.17.7.863703
- Meijers JC, Vlooswijk RA, Bouma BN. Inhibition of Human Blood Coagulation Factor XIa by C-1 Inhibitor. *Biochemistry* (1988) 27(3):959– 63. doi: 10.1021/bi00403a018
- 111. Cugno M, Bos I, Lubbers Y, Hack CE, Agostoni A. In Vitro Interaction of Cl-Inhibitor With Thrombin. Blood Coagul Fibrinolysis (2001) 12(4):253–60. doi: 10.1097/00001721-200106000-00005
- 112. Thomson TM, Toscano-Guerra E, Casis E, Paciucci R. C1 Esterase Inhibitor and the Contact System in COVID-19. Br J Haematol (2020) 190(4):520–4. doi: 10.1111/bjh.16938
- 113. Adesanya TMA, Campbell CM, Cheng L, Ogbogu PU, Kahwash R. C1 Esterase Inhibition: Targeting Multiple Systems in COVID-19. J Clin Immunol (2021) 41(4):729–32. doi: 10.1007/s10875-021-00972-1
- 114. Shen B, Yi X, Sun Y, Bi X, Du J, Zhang C, et al. Proteomic and Metabolomic Characterization of COVID-19 Patient Sera. *Cell* (2020) 182(1):59–72 e15. doi: 10.1016/j.cell.2020.05.032
- 115. Heurich M, Preston RJ, O'Donnell VB, Morgan BP, Collins PW. Thrombomodulin Enhances Complement Regulation Through Strong Affinity Interactions With Factor H and C3b-Factor H Complex. *Thromb Res* (2016) 145:84–92. doi: 10.1016/j.thromres.2016.07.017
- 116. Foley JH, Walton BL, Aleman MM, O'Byrne AM, Lei V, Harrasser M, et al. Complement Activation in Arterial and Venous Thrombosis Is Mediated by Plasmin. *EBioMedicine* (2016) 5:175–82. doi: 10.1016/j.ebiom.2016.02.011
- 117. Campbell WD, Lazoura E, Okada N, Okada H. Inactivation of C3a and C5a Octapeptides by Carboxypeptidase R and Carboxypeptidase N. *Microbiol Immunol* (2002) 46(2):131–4. doi: 10.1111/j.1348-0421.2002.tb02669.x
- 118. Nougier C, Benoit R, Simon M, Desmurs-Clavel H, Marcotte G, Argaud L, et al. Hypofibrinolytic State and High Thrombin Generation may Play a Major Role in SARS-COV2 Associated Thrombosis. J Thromb Haemost (2020) 18(9):2215–9. doi: 10.1111/jth.15016
- 119. Speth C, Rambach G, Wurzner R, Lass-Florl C, Kozarcanin H, Hamad OA, et al. Complement and Platelets: Mutual Interference in the Immune

Network. Mol Immunol (2015) 67(1):108-18. doi: 10.1016/j.molimm. 2015.03.244

- 120. Kim H, Conway EM. Platelets and Complement Cross-Talk in Early Atherogenesis. Front Cardiovasc Med (2019) 6:131. doi: 10.3389/ fcvm.2019.00131
- 121. Klos A, Tenner AJ, Johswich KO, Ager RR, Reis ES, Kohl J. The Role of the Anaphylatoxins in Health and Disease. *Mol Immunol* (2009) 46(14):2753– 66. doi: 10.1016/j.molimm.2009.04.027
- 122. Polley MJ, Nachman RL. Human Platelet Activation by C3a and C3a Des-Arg. J Exp Med (1983) 158(2):603–15. doi: 10.1084/jem.158.2.603
- 123. Del Conde I, Cruz MA, Zhang H, Lopez JA, Afshar-Kharghan V. Platelet Activation Leads to Activation and Propagation of the Complement System. *J Exp Med* (2005) 201(6):871–9. doi: 10.1084/jem.20041497
- 124. Bettoni S, Galbusera M, Gastoldi S, Donadelli R, Tentori C, Sparta G, et al. Interaction Between Multimeric Von Willebrand Factor and Complement: A Fresh Look to the Pathophysiology of Microvascular Thrombosis. *J Immunol* (2017) 199(3):1021–40. doi: 10.4049/jimmunol.1601121
- Wiedmer T, Esmon CT, Sims PJ. Complement Proteins C5b-9 Stimulate Procoagulant Activity Through Platelet Prothrombinase. *Blood* (1986) 68 (4):875–80. doi: 10.1182/blood.V68.4.875.875
- 126. Mol BWJ, Roberts CT, Thangaratinam S, Magee LA, de Groot CJM, Hofmeyr GJ. Pre-Eclampsia. *Lancet* (2016) 387(10022):999-1011. doi: 10.1016/S0140-6736(15)00070-7
- 127. Sjaus A, McKeen DM, George RB. Hypertensive Disorders of Pregnancy. Can J Anaesth (2016) 63(9):1075–97. doi: 10.1007/s12630-016-0689-8
- Haram K, Svendsen E, Abildgaard U. The HELLP Syndrome: Clinical Issues and Management. A Review. BMC Pregnancy Childbirth (2009) 9:8. doi: 10.1186/1471-2393-9-8
- Redman CW, Sargent IL, Staff AC. IFPA Senior Award Lecture: Making Sense of Pre-Eclampsia - Two Placental Causes of Preeclampsia? *Placenta* (2014) 35 Suppl:S20–5. doi: 10.1016/j.placenta.2013.12.008
- Burton GJ, Redman CW, Roberts JM, Moffett A. Pre-Eclampsia: Pathophysiology and Clinical Implications. *BMJ* (2019) 366:l2381. doi: 10.1136/bmj.l2381
- 131. Bulla R, Villa A, Bossi F, Cassetti A, Radillo O, Spessotto P, et al. VE-Cadherin Is a Critical Molecule for Trophoblast-Endothelial Cell Interaction in Decidual Spiral Arteries. *Exp Cell Res* (2005) 303(1):101–13. doi: 10.1016/ j.yexcr.2004.09.015
- 132. Agostinis C, Bulla R, Tripodo C, Gismondi A, Stabile H, Bossi F, et al. An Alternative Role of C1q in Cell Migration and Tissue Remodeling: Contribution to Trophoblast Invasion and Placental Development. *J Immunol* (2010) 185(7):4420–9. doi: 10.4049/jimmunol.0903215
- 133. Balduit A, Mangogna A, Agostinis C, Zito G, Romano F, Ricci G, et al. Zinc Oxide Exerts Anti-Inflammatory Properties on Human Placental Cells. *Nutrients* (2020) 12(6):1822. doi: 10.3390/nu12061822
- Bulla R, Fischetti F, Bossi F, Tedesco F. Feto-Maternal Immune Interaction at the Placental Level. *Lupus* (2004) 13(9):625–9. doi: 10.1191/ 0961203304lu2010oa
- 135. Roland CS, Hu J, Ren CE, Chen H, Li J, Varvoutis MS, et al. Morphological Changes of Placental Syncytium and Their Implications for the Pathogenesis of Preeclampsia. *Cell Mol Life Sci* (2016) 73(2):365–76. doi: 10.1007/s00018-015-2069-x
- Bulla R, Bossi F, Radillo O, de Seta F, Tedesco F. Placental Trophoblast and Endothelial Cells as Target of Maternal Immune Response. *Autoimmunity* (2003) 36(1):11–8. doi: 10.1080/0891693031000067331
- 137. Saito S, Shiozaki A, Nakashima A, Sakai M, Sasaki Y. The Role of the Immune System in Preeclampsia. *Mol Aspects Med* (2007) 28(2):192–209. doi: 10.1016/j.mam.2007.02.006
- Mangogna A, Agostinis C, Ricci G, Romano F, Bulla R. Overview of Procalcitonin in Pregnancy and in Pre-Eclampsia. *Clin Exp Immunol* (2019) 198(1):37–46. doi: 10.1111/cei.13311
- Haeger M, Bengtson A, Karlsson K, Heideman M. Complement Activation and Anaphylatoxin (C3a and C5a) Formation in Preeclampsia and by Amniotic Fluid. *Obstet Gynecol* (1989) 73(4):551–6.
- 140. Denny KJ, Coulthard LG, Finnell RH, Callaway LK, Taylor SM, Woodruff TM. Elevated Complement Factor C5a in Maternal and Umbilical Cord Plasma in Preeclampsia. J Reprod Immunol (2013) 97(2):211–6. doi: 10.1016/ j.jri.2012.11.006

- 141. Agostinis C, Stampalija T, Tannetta D, Loganes C, Vecchi Brumatti L, De Seta F, et al. Complement Component C1q as Potential Diagnostic But Not Predictive Marker of Preeclampsia. Am J Reprod Immunol (2016) 76(6):475– 81. doi: 10.1111/aji.12586
- 142. Burwick RM, Fichorova RN, Dawood HY, Yamamoto HS, Feinberg BB. Urinary Excretion of C5b-9 in Severe Preeclampsia: Tipping the Balance of Complement Activation in Pregnancy. *Hypertension* (2013) 62(6):1040–5. doi: 10.1161/HYPERTENSIONAHA.113.01420
- 143. Lynch AM, Gibbs RS, Murphy JR, Byers T, Neville MC, Giclas PC, et al. Complement Activation Fragment Bb in Early Pregnancy and Spontaneous Preterm Birth. Am J Obstet Gynecol (2008) 199(4):354.e1–8. doi: 10.1016/ j.ajog.2008.07.044
- 144. Petitbarat M, Durigutto P, Macor P, Bulla R, Palmioli A, Bernardi A, et al. Critical Role and Therapeutic Control of the Lectin Pathway of Complement Activation in an Abortion-Prone Mouse Mating. J Immunol (2015) 195 (12):5602–7. doi: 10.4049/jimmunol.1501361
- 145. Lokki AI, Heikkinen-Eloranta J, Jarva H, Saisto T, Lokki ML, Laivuori H, et al. Complement Activation and Regulation in Preeclamptic Placenta. *Front Immunol* (2014) 5:312. doi: 10.3389/fimmu.2014.00312
- 146. Chaouat G, Petitbarat M, Bulla R, Dubanchet S, Valdivia K, Ledee N, et al. Early Regulators in Abortion and Implications for a Preeclampsia Model. *J Reprod Immunol* (2009) 82(2):131–40. doi: 10.1016/j.jri.2009.08.004
- 147. Hypertension in pregnancy. Report of the American College of Obstetricians and Gynecologists' Task Force on Hypertension in Pregnancy. Obstet Gynecol (2013) 122(5):1122–31. doi: 10.1097/01.AOG.0000437382.03963.88
- 148. Powers RW, Jeyabalan A, Clifton RG, Van Dorsten P, Hauth JC, Klebanoff MA, et al. Soluble Fms-Like Tyrosine Kinase 1 (Sflt1), Endoglin and Placental Growth Factor (PIGF) in Preeclampsia Among High Risk Pregnancies. *PloS One* (2010) 5 (10):e13263. doi: 10.1371/journal.pone.0013263
- 149. Kleinrouweler CE, Wiegerinck MM, Ris-Stalpers C, Bossuyt PM, van der Post JA, von Dadelszen P, et al. Accuracy of Circulating Placental Growth Factor, Vascular Endothelial Growth Factor, Soluble Fms-Like Tyrosine Kinase 1 and Soluble Endoglin in the Prediction of Pre-Eclampsia: A Systematic Review and Meta-Analysis. *BJOG* (2012) 119(7):778–87. doi: 10.1111/j.1471-0528.2012.03311.x
- Chau K, Hennessy A, Makris A. Placental Growth Factor and Pre-Eclampsia. J Hum Hypertens (2017) 31(12):782–6. doi: 10.1038/jbh.2017.61
- Lecarpentier E, Tsatsaris V. Angiogenic Balance (Sflt-1/PIGF) and Preeclampsia. Ann Endocrinol (Paris) (2016) 77(2):97–100. doi: 10.1016/ j.ando.2016.04.007
- 152. Zeisler H, Llurba E, Chantraine F, Vatish M, Staff AC, Sennstrom M, et al. Predictive Value of the Sflt-1:PIGF Ratio in Women With Suspected Preeclampsia. N Engl J Med (2016) 374(1):13–22. doi: 10.1056/NEJMoa 1414838
- 153. Hund M, Allegranza D, Schoedl M, Dilba P, Verhagen-Kamerbeek W, Stepan H. Multicenter Prospective Clinical Study to Evaluate the Prediction of Short-Term Outcome in Pregnant Women With Suspected Preeclampsia (PROGNOSIS): Study Protocol. BMC Pregnancy Childbirth (2014) 14:324. doi: 10.1186/1471-2393-14-324
- Zaigham M, Andersson O. Maternal and Perinatal Outcomes With COVID-19: A Systematic Review of 108 Pregnancies. Acta Obstet Gynecol Scand (2020) 99(7):823–9. doi: 10.1111/aogs.13867
- 155. Di Mascio D, Buca D, Berghella V, Khalil A, Rizzo G, Odibo A, et al. Counseling in Maternal-Fetal Medicine: SARS-CoV-2 Infection in Pregnancy. Ultrasound Obstet Gynecol (2021) 57(5):687–97. doi: 10.1002/ uog.23628
- 156. Abedzadeh-Kalahroudi M, Sehat M, Vahedpour Z, Talebian P, Haghighi A. Clinical and Obstetric Characteristics of Pregnant Women With Covid-19: A Case Series Study on 26 Patients. *Taiwan J Obstet Gynecol* (2021) 60(3):458– 62. doi: 10.1016/j.tjog.2021.03.012
- 157. Chen H, Guo J, Wang C, Luo F, Yu X, Zhang W, et al. Clinical Characteristics and Intrauterine Vertical Transmission Potential of COVID-19 Infection in Nine Pregnant Women: A Retrospective Review of Medical Records. *Lancet* (2020) 395(10226):809–15. doi: 10.1016/S0140-6736(20)30360-3
- 158. Narang K, Enninga EAL, Gunaratne M, Ibirogba ER, Trad ATA, Elrefaei A, et al. SARS-CoV-2 Infection and COVID-19 During Pregnancy: A Multidisciplinary Review. *Mayo Clin Proc* (2020) 95(8):1750–65. doi: 10.1016/j.mayocp.2020.05.011

- 159. Papageorghiou AT, Deruelle P, Gunier RB, Rauch S, Garcia-May PK, Mhatre M, et al. Preeclampsia and COVID-19: Results From the INTERCOVID Prospective Longitudinal Study. Am J Obstet Gynecol (2021) 225(3):289.e1–e17. doi: 10.1016/j.ajog.2021.05.014
- 160. Mendoza M, Garcia-Ruiz I, Maiz N, Rodo C, Garcia-Manau P, Serrano B, et al. Pre-Eclampsia-Like Syndrome Induced by Severe COVID-19: A Prospective Observational Study. *BJOG* (2020) 127(11):1374–80. doi: 10.1111/1471-0528.16339
- 161. Rosenbloom JI, Raghuraman N, Carter EB, Kelly JC. Coronavirus Disease 2019 Infection and Hypertensive Disorders of Pregnancy. Am J Obstet Gynecol (2021) 224(6):623–4. doi: 10.1016/j.ajog.2021.03.001
- 162. Jing Y, Run-Qian L, Hao-Ran W, Hao-Ran C, Ya-Bin L, Yang G, et al. Potential Influence of COVID-19/ACE2 on the Female Reproductive System. *Mol Hum Reprod* (2020) 26(6):367–73. doi: 10.1093/molehr/gaaa030
- 163. Kornacki J, Wirstlein P, Wender-Ozegowska E. Serum Levels of Soluble FMS-Like Tyrosine Kinase 1 and Endothelial Glycocalyx Components in Early- and Late-Onset Preeclampsia. J Matern Fetal Neonatal Med (2021) 1– 5. doi: 10.1080/14767058.2021.1949704
- Ackermann M, Verleden SE, Kuehnel M, Haverich A, Welte T, Laenger F, et al. Pulmonary Vascular Endothelialitis, Thrombosis, and Angiogenesis in Covid-19. N Engl J Med (2020) 383(2):120–8. doi: 10.1056/NEJMoa2015432
- 165. Smadja DM, Mentzer SJ, Fontenay M, Laffan MA, Ackermann M, Helms J, et al. COVID-19 Is a Systemic Vascular Hemopathy: Insight for Mechanistic and Clinical Aspects. *Angiogenesis* (2021) 24(4):755–88. doi: 10.1007/ s10456-021-09805-6
- 166. Shapiro NI, Schuetz P, Yano K, Sorasaki M, Parikh SM, Jones AE, et al. The Association of Endothelial Cell Signaling, Severity of Illness, and Organ Dysfunction in Sepsis. Crit Care (2010) 14(5):R182. doi: 10.1186/cc9290
- 167. Palmer KR, Tong S, Kaitu'u-Lino TJ. Placental-Specific sFLT-1: Role in Pre-Eclamptic Pathophysiology and Its Translational Possibilities for Clinical Prediction and Diagnosis. *Mol Hum Reprod* (2017) 23(2):69–78. doi: 10.1093/molehr/gaw077
- 168. Smadja DM, Nunes H, Juvin K, Bertil S, Valeyre D, Gaussem P, et al. Increase in Both Angiogenic and Angiostatic Mediators in Patients With Idiopathic Pulmonary Fibrosis. *Pathol Biol (Paris)* (2014) 62(6):391–4. doi: 10.1016/ j.patbio.2014.07.006
- 169. Negro A, Fama A, Penna D, Belloni L, Zerbini A, Giuri PG. SFLT-1 Levels in COVID-19 Patients: Association With Outcome and Thrombosis. Am J Hematol (2021) 96(2):E41–3. doi: 10.1002/ajh.26037
- 170. Smadja DM, Philippe A, Bory O, Gendron N, Beauvais A, Gruest M, et al. Placental Growth Factor Level in Plasma Predicts COVID-19 Severity and in-Hospital Mortality. J Thromb Haemost (2021) 19(7):1823–30. doi: 10.1111/jth.15339
- 171. Giardini V, Carrer A, Casati M, Contro E, Vergani P, Gambacorti-Passerini C. Increased sFLT-1/PIGF Ratio in COVID-19: A Novel Link to Angiotensin II-Mediated Endothelial Dysfunction. *Am J Hematol* (2020) 95(8):E188–91. doi: 10.1002/ajh.25882
- 172. Amorim MM, Takemoto MLS, Katz L. Re: Pre-Eclampsia-Like Syndrome Induced by Severe Coronavirus Disease 2019: A Prospective Observational Study. BJOG (2020) 127(12):1577. doi: 10.1111/1471-0528.16402
- Young BC, Levine RJ, Karumanchi SA. Pathogenesis of Preeclampsia. Annu Rev Pathol (2010) 5:173–92. doi: 10.1146/annurev-pathol-121808-102149
- 174. Dusse LM, Rios DR, Pinheiro MB, Cooper AJ, Lwaleed BA. Pre-Eclampsia: Relationship Between Coagulation, Fibrinolysis and Inflammation. *Clin Chim Acta* (2011) 412(1-2):17–21. doi: 10.1016/j.cca.2010.09.030
- Pinheiro MB, Gomes KB, Dusse LM. Fibrinolytic System in Preeclampsia. Clin Chim Acta (2013) 416:67–71. doi: 10.1016/j.cca.2012.10.060
- 176. Lippi G, Plebani M, Henry BM. Thrombocytopenia Is Associated With Severe Coronavirus Disease 2019 (COVID-19) Infections: A Meta-Analysis. *Clin Chim Acta* (2020) 506:145–8. doi: 10.1016/j.cca.2020.03.022
- 177. Lippi G, Favaloro EJ. D-Dimer Is Associated With Severity of Coronavirus Disease 2019: A Pooled Analysis. *Thromb Haemost* (2020) 120(5):876–8. doi: 10.1055/s-0040-1709650
- 178. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical Features of Patients Infected With 2019 Novel Coronavirus in Wuhan, China. *Lancet* (2020) 395(10223):497–506. doi: 10.1016/S0140-6736(20)30183-5
- Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected

Pneumonia in Wuhan, China. JAMA (2020) 323(11):1061–9. doi: 10.1001/ jama.2020.1585

- 180. Medeiros LT, Peracoli JC, Bannwart-Castro CF, Romao M, Weel IC, Golim MA, et al. Monocytes From Pregnant Women With Pre-Eclampsia Are Polarized to a M1 Phenotype. Am J Reprod Immunol (2014) 72(1):5–13. doi: 10.1111/aji.12222
- 181. Costela-Ruiz VJ, Illescas-Montes R, Puerta-Puerta JM, Ruiz C, Melguizo-Rodriguez L. SARS-CoV-2 Infection: The Role of Cytokines in COVID-19 Disease. Cytokine Growth Factor Rev (2020) 54:62–75. doi: 10.1016/ j.cytogfr.2020.06.001
- 182. Leavitt AO, Li Q, Chan ED. Re: Pre-Eclampsia-Like Syndrome Induced by Severe COVID-19: A Prospective Observational Study: Common Pathophysiology of Pre-Eclampsia and Severe COVID-19? *BJOG* (2021) 128(3):618–9. doi: 10.1111/1471-0528.16584
- 183. Jayarangaiah A, Kariyanna PT, Chen X, Jayarangaiah A, Kumar A. COVID-19-Associated Coagulopathy: An Exacerbated Immunothrombosis Response. *Clin Appl Thromb Hemost* (2020) 26:1076029620943293. doi: 10.1177/1076029620943293
- 184. Marder W, Knight JS, Kaplan MJ, Somers EC, Zhang X, O'Dell AA, et al. Placental Histology and Neutrophil Extracellular Traps in Lupus and Pre-Eclampsia Pregnancies. *Lupus Sci Med* (2016) 3(1):e000134. doi: 10.1136/ lupus-2015-000134
- 185. Middleton EA, He XY, Denorme F, Campbell RA, Ng D, Salvatore SP, et al. Neutrophil Extracellular Traps Contribute to Immunothrombosis in COVID-19 Acute Respiratory Distress Syndrome. *Blood* (2020) 136 (10):1169–79. doi: 10.1182/blood.2020007008
- 186. de Bont CM, Boelens WC, Pruijn GJM. NETosis, Complement, and Coagulation: A Triangular Relationship. *Cell Mol Immunol* (2019) 16 (1):19–27. doi: 10.1038/s41423-018-0024-0
- 187. Zemet R, Dulitzki M, Baum M, Ofer Friedman H, Morag I, Simchen MJ. Early-Onset Preeclampsia - The Impact of Antiphospholipid Antibodies on Disease Severity. *Eur J Obstet Gynecol Reprod Biol* (2021) 263:79–84. doi: 10.1016/j.ejogrb.2021.06.006
- 188. Zuo Y, Estes SK, Ali RA, Gandhi AA, Yalavarthi S, Shi H, et al. Prothrombotic Autoantibodies in Serum From Patients Hospitalized With COVID-19. Sci Transl Med (2020) 12(570):eabd3876. doi: 10.1126/ scitranslmed.abd3876
- 189. Holers VM, Girardi G, Mo L, Guthridge JM, Molina H, Pierangeli SS, et al. Complement C3 Activation Is Required for Antiphospholipid Antibody-Induced Fetal Loss. J Exp Med (2002) 195(2):211–20. doi: 10.1084/jem.200116116
- 190. Galli M, Borrelli G, Jacobsen EM, Marfisi RM, Finazzi G, Marchioli R, et al. Clinical Significance of Different Antiphospholipid Antibodies in the WAPS (Warfarin in the Antiphospholipid Syndrome) Study. *Blood* (2007) 110 (4):1178–83. doi: 10.1182/blood-2007-01-066043
- 191. Chaturvedi S, Brodsky RA, McCrae KR. Complement in the Pathophysiology of the Antiphospholipid Syndrome. *Front Immunol* (2019) 10:449. doi: 10.3389/fimmu.2019.00449
- 192. Burwick RM, Feinberg BB. Complement Activation and Regulation in Preeclampsia and Hemolysis, Elevated Liver Enzymes, and Low Platelet Count Syndrome. Am J Obstet Gynecol (2020). doi: 10.1016/j.ajog.2020.09.038
- 193. Vaught AJ, Braunstein EM, Jasem J, Yuan X, Makhlin I, Eloundou S, et al. Germline Mutations in the Alternative Pathway of Complement Predispose to HELLP Syndrome. *JCI Insight* (2018) 3(6):e99128. doi: 10.1172/ jci.insight.99128
- 194. Gavriilaki E, Asteris PG, Touloumenidou T, Koravou EE, Koutra M, Papayanni PG, et al. Genetic Justification of Severe COVID-19 Using a Rigorous Algorithm. *Clin Immunol* (2021) 226:108726. doi: 10.1016/ j.clim.2021.108726
- 195. Roberge S, Bujold E, Nicolaides KH. Aspirin for the Prevention of Preterm and Term Preeclampsia: Systematic Review and Metaanalysis. Am J Obstet Gynecol (2018) 218(3):287–93.e1. doi: 10.1016/j.ajog.2017.11.561
- 196. Pierik E, Prins JR, van Goor H, Dekker GA, Daha MR, Seelen MAJ, et al. Dysregulation of Complement Activation and Placental Dysfunction: A Potential Target to Treat Preeclampsia? *Front Immunol* (2019) 10:3098. doi: 10.3389/fimmu.2019.03098
- 197. Wang X, Gao H. Prevention of Preeclampsia in High-Risk Patients With Low-Molecular-Weight Heparin: A Meta-Analysis. J Matern Fetal Neonatal Med (2020) 33(13):2202–8. doi: 10.1080/14767058.2018.1543656

- Roberge S, Demers S, Nicolaides KH, Bureau M, Cote S, Bujold E. Prevention of Pre-Eclampsia by Low-Molecular-Weight Heparin in Addition to Aspirin: A Meta-Analysis. Ultrasound Obstet Gynecol (2016) 47(5):548–53. doi: 10.1002/uog.15789
- 199. Kazatchkine MD, Fearon DT, Silbert JE, Austen KF. Surface-Associated Heparin Inhibits Zymosan-Induced Activation of the Human Alternative Complement Pathway by Augmenting the Regulatory Action of the Control Proteins on Particle-Bound C3b. J Exp Med (1979) 150(5):1202–15. doi: 10.1084/jem.150.5.1202
- 200. Bianconi V, Violi F, Fallarino F, Pignatelli P, Sahebkar A, Pirro M. Is Acetylsalicylic Acid a Safe and Potentially Useful Choice for Adult Patients With COVID-19? Drugs (2020) 80(14):1383–96. doi: 10.1007/s40265-020-01365-1
- 201. Protective Effect of Aspirin on COVID-19 Patients (2020). Available at: https://ClinicalTrials.gov/show/NCT04365309.
- 202. Gautret P, Lagier JC, Parola P, Hoang VT, Meddeb L, Mailhe M, et al. Hydroxychloroquine and Azithromycin as a Treatment of COVID-19: Results of an Open-Label Non-Randomized Clinical Trial. *Int J Antimicrob Agents* (2020) 56(1):105949. doi: 10.1016/j.ijantimicag.2020.105949
- Billett HH, Reyes-Gil M, Szymanski J, Ikemura K, Stahl LR, Lo Y, et al. Anticoagulation in COVID-19: Effect of Enoxaparin, Heparin, and Apixaban on Mortality. *Thromb Haemost* (2020) 120(12):1691–9. doi: 10.1055/s-0040-1720978
- Packard BD, Weiler JM. Steroids Inhibit Activation of the Alternative-Amplification Pathway of Complement. *Infect Immun* (1983) 40(3):1011– 4. doi: 10.1128/iai.40.3.1011-1014.1983
- Weiler JM, Packard BD. Methylprednisolone Inhibits the Alternative and Amplification Pathways of Complement. *Infect Immun* (1982) 38(1):122–6. doi: 10.1128/iai.38.1.122-126.1982
- 206. Seo MR, Chae J, Kim YM, Cha HS, Choi SJ, Oh S, et al. Hydroxychloroquine Treatment During Pregnancy in Lupus Patients Is Associated With Lower Risk of Preeclampsia. *Lupus* (2019) 28(6):722–30. doi: 10.1177/ 0961203319843343
- 207. Elavarasi A, Prasad M, Seth T, Sahoo RK, Madan K, Nischal N, et al. Chloroquine and Hydroxychloroquine for the Treatment of COVID-19: A Systematic Review and Meta-Analysis. J Gen Intern Med (2020) 35 (11):3308–14. doi: 10.1007/s11606-020-06146-w
- Lefkou E, Mamopoulos A, Dagklis T, Vosnakis C, Rousso D, Girardi G. Pravastatin Improves Pregnancy Outcomes in Obstetric Antiphospholipid Syndrome Refractory to Antithrombotic Therapy. J Clin Invest (2016) 126 (8):2933–40. doi: 10.1172/JCI86957
- 209. Gonzalez JM, Pedroni SM, Girardi G. Statins Prevent Cervical Remodeling, Myometrial Contractions and Preterm Labor Through a Mechanism That Involves Hemoxygenase-1 and Complement Inhibition. *Mol Hum Reprod* (2014) 20(6):579–89. doi: 10.1093/molehr/gau019
- 210. Burwick RM, Feinberg BB. Eculizumab for the Treatment of Preeclampsia/ HELLP Syndrome. *Placenta* (2013) 34(2):201-3. doi: 10.1016/ j.placenta.2012.11.014
- 211. Declercq J, Bosteels C, Van Damme K, De Leeuw E, Maes B, Vandecauter A, et al. Zilucoplan in Patients With Acute Hypoxic Respiratory Failure Due to COVID-19 (ZILU-COV): A Structured Summary of a Study Protocol for a Randomised Controlled Trial. *Trials* (2020) 21(1):934. doi: 10.1186/s13063-020-04884-0
- 212. Mastaglio S, Ruggeri A, Risitano AM, Angelillo P, Yancopoulou D, Mastellos DC, et al. The First Case of COVID-19 Treated With the Complement C3 Inhibitor AMY-101. *Clin Immunol* (2020) 215:108450. doi: 10.1016/j.clim.2020.108450
- 213. Smith K, Pace A, Ortiz S, Kazani S, Rottinghaus S. A Phase 3 Open-Label, Randomized, Controlled Study to Evaluate the Efficacy and Safety of Intravenously Administered Ravulizumab Compared With Best Supportive Care in Patients With COVID-19 Severe Pneumonia, Acute Lung Injury, or Acute Respiratory Distress Syndrome: A Structured Summary of a Study Protocol for a Randomised Controlled Trial. *Trials* (2020) 21(1):639. doi: 10.1186/s13063-020-04548-z
- 214. Diurno F, Numis FG, Porta G, Cirillo F, Maddaluno S, Ragozzino A, et al. Eculizumab Treatment in Patients With COVID-19: Preliminary Results From Real Life ASL Napoli 2 Nord Experience. *Eur Rev Med Pharmacol Sci* (2020) 24(7):4040–7. doi: 10.26355/eurrev\_202004\_20875

- Yan B, Freiwald T, Chauss D, Wang L, West E, Mirabelli C, et al. SARS-CoV-2 Drives JAK1/2-Dependent Local Complement Hyperactivation. Sci Immunol (2021) 6(58):eabg0833. doi: 10.1126/sciimmunol.abg0833
- 216. Rambaldi A, Gritti G, Mico MC, Frigeni M, Borleri G, Salvi A, et al. Endothelial Injury and Thrombotic Microangiopathy in COVID-19: Treatment With the Lectin-Pathway Inhibitor Narsoplimab. *Immunobiology* (2020) 225(6):152001. doi: 10.1016/j.imbio.2020.152001
- 217. Mastellos DC, Pires da Silva BGP, Fonseca BAL, Fonseca NP, Auxiliadora-Martins M, Mastaglio S, et al. Complement C3 vs C5 Inhibition in Severe COVID-19: Early Clinical Findings Reveal Differential Biological Efficacy. *Clin Immunol* (2020) 220:108598. doi: 10.1016/j.clim.2020.108598
- Stoermer KA, Morrison TE. Complement and Viral Pathogenesis. Virology (2011) 411(2):362–73. doi: 10.1016/j.virol.2010.12.045
- 219. Annane D, Heming N, Grimaldi-Bensouda L, Fremeaux-Bacchi V, Vigan M, Roux AL, et al. Eculizumab as an Emergency Treatment for Adult Patients With Severe COVID-19 in the Intensive Care Unit: A Proof-of-Concept Study. *EClinicalMedicine* (2020) 28:100590. doi: 10.1016/j.eclinm. 2020.100590
- 220. Vlaar APJ, de Bruin S, Busch M, Timmermans S, van Zeggeren IE, Koning R, et al. Anti-C5a Antibody IFX-1 (Vilobelimab) Treatment Versus Best Supportive Care for Patients With Severe COVID-19 (PANAMO): An Exploratory, Open-Label, Phase 2 Randomised Controlled Trial. *Lancet Rheumatol* (2020) 2(12):e764–73. doi: 10.1016/S2665-9913(20)30341-6
- Avdoralimab an Anti-C5aR Antibody, in Patients With COVID-19 Severe Pneumonia (FORCE) (2020). Available at: https://ClinicalTrials.gov/show/ NCT04371367.

- 222. Skendros P, Mitsios A, Chrysanthopoulou A, Mastellos DC, Metallidis S, Rafailidis P, et al. Complement and Tissue Factor-Enriched Neutrophil Extracellular Traps Are Key Drivers in COVID-19 Immunothrombosis. *J Clin Invest* (2020) 130(11):6151–7. doi: 10.1172/JCI141374
- 223. Urwyler P, Moser S, Charitos P, Heijnen I, Rudin M, Sommer G, et al. Treatment of COVID-19 With Conestat Alfa, a Regulator of the Complement, Contact Activation and Kallikrein-Kinin System. Front Immunol (2020) 11:2072. doi: 10.3389/fimmu.2020.02072

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Agostinis, Mangogna, Balduit, Aghamajidi, Ricci, Kishore and Bulla. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# GLOSSARY

ACE2	angiotensin-converting enzyme 2
aPLAs	anti-phospholipid antibodies
APS	anti-phospholipid antibodies
BK	bradykinin
	2
C	complement system
C1-INH	C1-inhibitor
COVID-19	coronavirus disease 2019
CR1	complement receptor 1
DAF	decay accelerating factor
DAMPs	damage-associated molecular patterns
EC	endothelial cell
EMMPRIN	extracellular matrix metalloproteinase inducer
EOPE	early onset pre-eclampsia
F	factor
HCQ	hydroxychloroquine
HELLP	hemolysis, elevated liver enzymes and low platelets
ITCC	inactive terminal complement complex
LDA	low-dose aspirin
LMWH	low-molecular weight heparin
LOPE	late onset pre-eclampsia
MAC	membrane attack complex
MASPs	MBL-associated serine proteases
MBL	mannan-binding lectin
MCP	membrane cofactor protein
MERS	•
NETS	middle east respiratory syndrome
	neutrophil extracellular traps
PAI-1	plasminogen activator inhibitor type-1
PAMPs	pathogen-associated molecular patterns
PARs	protease-activated receptors
PE	pre-eclampsia
PIGF	placental growth factor
RAAS	renin-angiotensin-aldosterone system
ROS	reactive oxygen species
SARS-CoV-2	severe acute respiratory syndrome coronavirus 2
sEng	soluble endoglin
sFlt-1	soluble fms-like tyrosine kinase 1
SLE	systemic lupus erythematosus
√WF	von Willebrand factor
TAFI	thrombin-activable fibrinolysis inhibitor
TCC	terminal complement complex
TF	tissue factor
TFPI	tissue factor pathway inhibitor
TM	thrombomodulin
TMPRSS2	transmembrane serine protease 2
UtAPI	uterine artery pulsatility index
VEGF	vascular-endothelial growth factor