Check for updates

OPEN ACCESS

EDITED BY Mohamed Ghoneim, Mansoura University, Egypt

REVIEWED BY

Josefina M. Alberu, Tecnológico de Monterrey, Mexico Kirsten Thus, Princess Maxima Center for Pediatric Oncology, Netherlands

*CORRESPONDENCE Michiel G. H. Betjes m.g.h.betjes@erasmusmc.nl

SPECIALTY SECTION This article was submitted to Alloimmunity and Transplantation, a section of the journal Frontiers in Immunology

RECEIVED 20 June 2022 ACCEPTED 15 August 2022 PUBLISHED 30 August 2022

CITATION

Betjes MGH, Peereboom ETM, Otten HG and Spierings E (2022) The number of donor HLA-derived T cell epitopes available for indirect antigen presentation determines the risk for vascular rejection after kidney transplantation. *Front. Immunol.* 13:973968. doi: 10.3389/fimmu.2022.973968

COPYRIGHT

© 2022 Betjes, Peereboom, Otten and Spierings. This is an open-access article distributed under the terms of the Creative Commons Attribution License

(CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author (s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in

accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

The number of donor HLAderived T cell epitopes available for indirect antigen presentation determines the risk for vascular rejection after kidney transplantation

Michiel G. H. Betjes^{1*}, Emma T. M. Peereboom², Henny G. Otten² and Eric Spierings²

¹Department of Internal Medicine, Section of Nephrology and Transplantation, Erasmus Medical Center (MC), University Medical Center, Rotterdam, Netherlands, ²Center for Translational Immunology, University Medical Center, Utrecht, Netherlands

The role of the indirect T-cell recognition pathway of allorecognition in acute T cell-mediated rejection (aTCMR) is not well defined. The amount of theoretical T-cell epitopes available for indirect allorecognition can be quantified for donor-recipient combinations by the Predicted Indirectly ReCognizable HLA Epitopes algorithm (PIRCHE-II). The PIRCHE-II score was calculated for 688 donor kidney-recipient combinations and associated with the incidence of first-time diagnosed cases of TCMR. A diagnosis of TCMR was made in 182 cases; 121 cases of tubulo-interstitial rejection cases (79 cases of borderline TCMR, 42 cases of TCMR IA-B) and 61 cases of vascular TCMR (TCMR II-III). The PIRCHE-II score for donor HLA-DR/DQ (PIRCHE-II DR/DQ) was highly associated with vascular rejection. At one year after transplantation, the cumulative percentage of recipients with a vascular rejection was 12.7%, 8.6% and 2.1% within respectively the high, medium and low tertile of the PIRCHE-II DR/DQ score (p<0.001). In a multivariate regression analysis this association remained significant (p<0.001 for PIRCHE-II DR/DQ tertiles). The impact of a high PIRCHE-II DR/DQ score was mitigated by older recipient age and a living donor kidney. In conclusion, indirect antigen presentation of donor HLApeptides may significantly contribute to the risk for acute vascular rejection.

KEYWORDS

T cell-mediated acute cellular rejection, PIRCHE-II, vascular rejection, transplantation, kidney

Introduction

Although the incidence of acute T cell-mediated rejection (aTCMR) after kidney transplantation has decreased over the last decades by more potent immune suppressive drugs, aTCMR still contribute significantly to the incidence of graft loss (1, 2). The immunological mechanism of aTCMR is characterized by cytotoxic CD8⁺ cells that recognize intact mismatched HLA class-I proteins on donor cells, a mode of recognition known as the direct pathway of allorecognition. In parallel, alloreactive CD4⁺ T cells may also play an important role in acute rejection as they either can directly invade the transplanted organ and cause rejection (3), or support the activation of the directly recognizing alloreactive cytotoxic CD8⁺ T cells. In the latter process, the indirect pathway of allorecognition is involved (4). In this pathway, mismatched donor HLA is internalized by the recipient's antigen presenting cells (APC) and processed into peptide fragments. Such donor-HLA derived peptides can then be presented by the HLA-II molecules on the cell surface of the APC of the recipient. These activated CD4⁺ T cells subsequently support the activation and proliferation of direct alloreactive CD8⁺ T cells (5-7). At present, the relative contribution of the indirect alloreactive CD4⁺ T cells to the risk for aTCMR after kidney transplantation is largely unknown.

In order to model the indirect pathway of allorecognition by CD4⁺ T cells, an algorithm has been developed to identify the Predicted Indirectly ReCognizable HLA Epitopes that can be presented by recipient HLA class II (PIRCHE-II) (8). This algorithm allows for calculation of the T-cell epitope load score based on donor HLA peptides that can be presented by HLA class II molecules of the recipient, but are not found in the recipient's own HLA-A, -B, -C, -DRB1 and -DQB1 alleles. The potential advantage of calculating such a score is an evaluation of antigenicity of the donor HLA molecules at greater detail, as the number of potential donor HLA-derived antigenic peptides that can be presented by the recipient HLA class II to the recipient T cells may vary substantially for a given HLA mismatch. Recent studies have shown that the PIRCHE-II score associates with de novo donor-specific anti-HLA antibody development and the risk for long-term graft failure after kidney transplantation (9-13). In addition, an association between the PIRCHE-II score and TCMR after liver transplantation (14, 15) and the risk for TCMR after kidney transplantation (16, 17) has been found. The latter findings are indicative of a role for indirect CD4⁺ T cell alloreactivity in TCMR as discussed by Vionet et al. (15).

In this study, we investigated whether the PIRCHE-II score, as a measure for the potential indirect CD4⁺ T cell alloreactivity, is related to the incidence of aTCMR in a large cohort of kidney transplant recipients with a complete clinical database and long-term follow-up.

Material and methods

Study population

In this retrospective study, we evaluated all kidney transplant recipients from the Erasmus MC between 1995 and 2005. This is a sub-cohort of the PROCARE study which included all kidney transplantations performed in the 7 transplantation centers in the Netherlands within that period (18). All transplants were performed after a negative T-cell complement dependent cytotoxicity (CDC) crossmatch. Pre-transplant sera were retrospectively assessed for the presence of HLA antibodies as previously described (18, 19). In total, 734 recipients were transplanted in our center within that period. Of all the kidney transplant donors and recipients included in this study (n=688, 46 not included because of insufficient serological or clinical data), the HLA typing was available at the serological level for a minimum of HLA-A, -B and -DR. The relevant clinical data were obtained from the Dutch Organ Transplant Registry and supplemented if needed with data from our local database, yielding a completely filled database (Table 1). The percentage of panel-reactive antibodies (PRA) was determined pretransplantation and a value >4% was considered as positive. All kidney graft biopsies were performed because of clinical suspicion for rejection and no per protocol biopsies were done. The classification of acute T cell-medicated rejection was made using the Banff 2019 criteria. Per patient, only the data of the first kidney biopsy with a diagnosis of TCMR were used for analysis. Within the first year after transplantation, 99% of all recipients suspected of a possible rejection (decrease in eGFR or unexplained proteinuria) underwent a kidney biopsy.

Patients were seen in our out-patient clinic at a regular basis at least weekly in the first month after transplantation and at least at a yearly basis beyond the first year after transplantation until death, graft loss or loss to follow-up (n=3 in the first year after transplantation and n=37 in total for lost to follow-up). The last date of follow-up was December 2021. Graft failure was defined as loss of kidney function necessitating dialysis or retransplantation. Informed consent was obtained from all participants. The clinical and research activities reported are consistent with the Principles of the Declaration of Istanbul as outlined in the 'Declaration of Istanbul on Organ Trafficking and Transplant Tourism' and in accordance with the declaration of Helsinki. The use of clinical data and assessment of donorspecific antibodies in stored serum samples was approved by the Research Ethics Committee for Biobanks at the Erasmus MC and the Medical Ethics Committee of the University Medical Center Utrecht.

TABLE 1 Clinical and demographic characteristics of 688 recipients and kidney donors.

Age recipient (average +/- SD)	45.6 years (13.6)		
Age donor in years (average +/- SD)	46.6 years (14.3)		
Recipient male/female ratio	48/52%		
Deceased/living donor kidney	48/52%		
Cold ischaemia time (average +/- SD)	10.9 hours (10.9)		
Re-transplantation	18%		
Mean PRA at transplantation	10%		
Total HLA mismatches (mean)	2.5		
HLA mismatches class I	1.7		
HLA mismatches class II	0.9		
Follow-up (median and IQR)	13.2 years (6-15)		
Recipients with anti-HLA DSA at time transplantation	21%		
Induction therapy			
- Anti-IL-2 receptor antibody	5%		
Maintenance immune suppressive medication			
- steroids	90%		
- tacrolimus/cyclosporine	60%/38%		
- MMF/azathioprine	70%/0.5%		
- other	0.9%		
T cell-mediated rejection, total number	182		
- borderline rejection	79 (43%)		
- tubulo-interstitial rejection (TCMR1a-b)	42 (23%)		
- vascular rejection (TCMR2-3)	61 (34%)		

PRA:panel reactive antibodies, DSA:donor specific anti-HLA antibodies.

Identification of PIRCHE-II peptides

The PIRCHE-II peptides originating from the donor's HLA alleles that can be presented by the recipient's HLA class II, were identified using the PIRCHE-II algorithm version 3.3.43 (PIRCHE AG, Berlin, Germany, available *via* www.pirche. com). HLA-A, -B, -C, -DRB1, -DRB3/4/5, and -DQA1/DQB1 were taken into consideration as presented loci, and HLA-DRB1 was considered as the presenting locus. As only HLA typing data at serological level were available in our cohort, the PIRCHE-II peptides and their weights were calculated based on serological typing data, as described previously in detail (11, 20). The PIRCHE-II scores for HLA class I were calculated by adding the epitope counts originating from donor HLA-A, -B, and -C, and the PIRCHE-II scores for HLA-DR/DQ were calculated by adding the epitope counts originating from donor HLA-A, -B, and -C, HLA-DR/DQ.

Statistical analysis

Differences between groups of date were assessed by the Fisher's exact test for categorical variables, the Mann-Whitney U test for not-normally distributed continuous variables and the Kruskall-Wallis test with post-hoc comparisons for comparing multiple groups with non-normally distributed value. All p-values were 2-tailed and the level of statistical significance was set to a p-value<0.05.

The PIRCHE-II score was used both as continuous variable and structured into tertiles for Kaplan-Meier analysis with logrank statistics for difference between strata and as categorical variable in the Cox proportional hazards analysis. Univariate Cox proportional hazards analysis was used to identify clinical and demographic variables as given in Table 1 for their association with TCMR-free survival for the different types of TCMR (tubulo-interstitial rejection (borderline TCMR and TCMR Banff grade IA-B) and vascular rejection (TCMR Banff grade II-III)). Variables with a p-value of 0.1 or less were considered for further analysis in a multivariate analysis to calculate hazard ratios. The type of calcineurin inhibitor used was included as strata in the model. Absence of collinearity in the model covariates was formally assessed by calculating the variance inflation factor. Statistical analysis was performed with software IBM SPSS statistics 21.

Results

In this cohort, a diagnosis of TCMR was made in 182 cases. The cohort included 121 cases of tubulo-interstitial rejection cases (79 cases of borderline rejection, 42 cases of TCMR IA-B) and 61 cases of vascular TCMR (TCMR II-III) (Table 1 for clinical characteristics). Vascular TCMR occurred almost exclusively in the first weeks after transplantation and was rarely diagnosed thereafter. A significant difference was observed between the PIRCHE-II scores of recipients with TCMR and those with no episode of TCMR (median score 52 vs.46, p=0.01). Subsequently, the PIRCHE-II scores for the donor HLA class I derived- (loci A, B and C) and HLA class II-derived epitopes (loci DR and DQ) were separately analyzed for their relation to different types of TCMR. The PIRCHE-II HLA class I score was not significantly associated with borderline rejection, tubulo-interstitial and vascular rejection (Figure 1). However, the median PIRCHE-II score for HLA-DR and DQ (PIRCHE-II DR/DQ score) was significantly higher among patients with vascular rejection compared to patients with tubulo-interstitial rejection and compared to patients without rejection(median score 30 vs.18, p<0.001).

To assess a possible dose-response effect, the PIRCHE-II scores for HLA class I and II were divided into tertiles. Rejection-free survival curves (Figure 2) were made for borderline and tubulo-interstitial TCMR (TCMR Banff grade <2) and vascular rejection (TCMR grade 2-3). In particular for the PIRCHE-II DR/DQ score, a significant association was found between the tertiles and vascular rejection-free survival. One year after transplantation, the cumulative percentage of recipients with a vascular rejection was 12.7%, 8.6% and 2.1% within respectively

the high, medium and low tertile of the PIRCHE-II DR/DQ score (p<0.001). In addition, a significant difference was found between the tubulo-interstitial rejection-free survival curves of patients within the lowest and the middle PIRCHE-II DR/DQ tertile (Figure 2, log-rank test;p=0.03). A trend for a better tubulo-interstitial rejection-free survival for the patients within the lowest PIRCHE-II DR/DQ tertile as compared to those within the highest PIRCHE-II DR/DQ tertile was observed (p=0.07).

The PIRCHE-II DR/DQ score is an independent risk factor for vascular rejection

A uni- and multivariate logistic regression analysis was performed to calculate the hazard ratios of relevant clinical parameters for the TCMR incidence after transplantation (Table 2 and 3). The known risk factors for TCMR, like younger age at transplantation, deceased donor kidney vs. living donor kidney and a positive PRA (>4%) were all confirmed to have a statistically significant hazard ratio in the univariate regression analysis. In accordance with a previous publication (21), the presence of pre-transplant DSA for HLA did not affect the incidence of TCMR. An increasing PIRCHE-II DR/DQ score was strongly related to TCMR, in particular TCMR Banff grade 2-3. In a multivariate analysis, which included all univariate variables with a p-value <0.1, the hazard ratio (HR) for the highest PIRCHE-II DR/ DQ tertile as compared to the lowest tertile was 1.75 for tubulointerstitial rejection (p=0.02), and 5.34 for vascular rejection (p<0.001). Such a relation was not significantly present for the PIRCHE-II HLA I score (tubulo-interstitial rejection HR 1.10, p=0.37; vascular rejection HR 1.23, p=0.17).

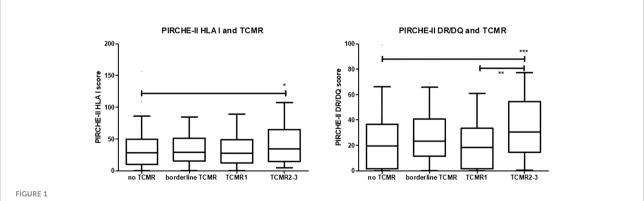
A sensitivity analysis was done by assessing the hazard ratio per tertile of PIRCHE-II DR/DQ score in a multivariate logistic regression analysis with strata for relevant risk factors for

rejection. Recipients with a cycloporine (HR 0.48, 95% CI 0.30-0.66, p=0.002) or tacrolimus-based immune suppressive regime (HR 0.44, 95% CI 0.28-0.72 p=0.002); for recipients of a living (HR 0.49, 95% CI 0.29-0.81 p=0.001) or deceased donor kidney (HR 0.50, 95% CI 0.22-0.76, p=0.018); and for different recipient age groups when divided in young (18-39 years, HR 0.46, 95% CI 0.28-0.72, p=0.005), middle age (40-55 years, HR 0.6695% CI 0.28-0.72, p=0.08) and older age (>50 years, HR 0.28, 95% CI 0.22-0.67, p=0.01). As shown in Figure 3, the impact of the PIRCHE-II DR/DQ score is related to the risk factors like younger age and deceased kidney donor. For instance, the risk for vascular rejection in recipients having a PIRCHE-II DR/DQ score in the highest tertile is relatively little mitigated by older age and increased by a deceased donor kidney. In contrast, no cases of vascular rejection in elderly recipients having a PIRCHE-II DR/DQ score in the lowest tertile were observed, independent of the type of donor kidney.

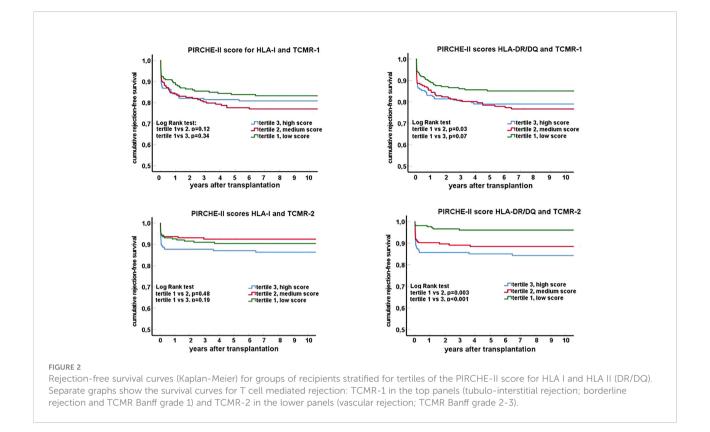
Finally, the effect of the PIRCHE-II score for the risk of rejection was evaluated in the fully matched recipient-donor kidney combination by serological HLA typing (n=95 transplantations). Eight cases of borderline rejection, 6 cases of TCMR Banff grade I and 2 cases of TCMR Banff grade IIA rejections were documented. These recipients had a significantly higher PIRCHE-II DR/DQ scores compared to the non-rejecting recipients (5.8 vs. 2.0, p=0.018).

Discussion

Recently, there is a renewed interest in the long-term negative effects of TCMR. Even after T cell-depleting therapy, a definitive resolution of the rejection is sometimes not achieved and graft loss may follow in the years thereafter (1, 2, 22). Hence, a better understanding of clinical relevant HLA mismatches and mechanisms involved is still needed. In this study, we investigated whether the PIRCHE-II score, as a parameter for



The PIRCHE-II score for donor HLA I and HLA II (DR/DQ) given in box-whisker plots for the different groups of recipients; no T cell-mediated rejection (TCMR), borderline TCMR, tubule-interstitial TCMR grade 1 and vascular TCMR grade 2-3. The p-value for the Kruskal-Waliss test for difference between multiple groups was <0.001. Post-hoc analysis for comparison between groups: *p<0.05, **p< 0.01, ***p<0.001.



the potential of indirect CD4⁺ T-cell alloreactivity, is associated with aTCMR after kidney transplantation. A strong association between the PIRCHE-II score for predicted donor HLA IIderived epitopes and the incidence of TCMR was observed, in particular vascular rejection. Sensitivity analysis showed that this association was consistently found in different strata of groups of recipients based upon known risk factors for TCMR like a deceased donor kidney, a younger recipient age and cyclosporine instead of tacrolimus as calcineurin inhibitor (23, 24). Our results are in accordance and extend previous work on the relation between the PIRCHE-II score and TCMR after liver and kidney transplantation (14–17). Vionnet *et al.* showed a significant association between PIRCHE-II and the probability score for TCMR in transplanted livers based on transcript levels of TCMR-related genes (15).

Lezoeva *et al.* showed that patients with borderline kidney rejection have a higher PIRCHE-II score compared to the entire study population, in particular in association with a high score for HLA-A (16). In contrast, Ono *et al.* observed in liver

TABLE 2 Uni- and multivariate Cox proportional hazard analysis for T-cell mediated rejection borderline and Banff grade 1.

	Univariate analysis		Multivariate analysis		
	Hazard ratio (95% CI)	p-value	Hazard ratio (95% CI)	p-value	
Recipient age per decade	0.81 (0.71-0.93)	0.003	0.80 (0.71-0.92)	0.002	
Donor age per decade	1.09 (0.95-1.26)	0.19	-	-	
Deceased vs living kidney donor	1.14 (0.79-1.63)	0.47	-	-	
Re-transplantation	0.93 (0.57-1.50)	0.77	-	-	
PRA >4%	1.29 (0.88-1.90)	0.18	-	-	
DSA pre-transplantation present	0.77 (0.48-1.24)	0.29	-	-	
PIRCHE-II score HLA class-I per tertile	1.10 (0.88-1.37)	0.37	-	-	
PIRCHE-II score HLA-DR/DQ per tertile		0.08		0.04	
- Lowest tertile vs middle tertile	1.51 (0.95-2.40)	0.08	1.61 (1.01-2.57)	0.04	
- Lowest tertile vs highest tertile	1.62 (1.03-1.54)	0.03	1.75 (1.09-2.69)	0.02	

	Univariate analysis		Multivariate analysis	
	Hazard ratio (95% CI)	p-value	Hazard ratio (95% CI)	p-value
Recipient age per decade	0.86 (0.71-1.01)	0.10	0.81 (0.66-0.96)	0.03
Donor age per decade	1.06 (0.87-1.28)	0.54	-	-
Deceased vs living kidney donor	2.01 (1.20-3.37	0.008	2.39 (1.42-4.04)	0.002
Re-transplantation	1.25 (0.68-2.32)	0.46	-	-
PRA >4%	1.80 (1.09-3.03)	0.02	1.65 (0.99-2.77)	0.05
DSA pre-transplantation present	1.49 (0.86-2.58)	0.15	-	-
PIRCHE-II score HLA class-I per tertile	1.23 (0.98-1.67)	0.18	-	-
PIRCHE-II score HLA-DR/DQ per tertile		0.001		< 0.001
- lowest tertile vs middle tertile	3.13 (1.4-7.0)	0.005	3.36 (1.49-7.58)	0.003
- lowest tertile vs highest tertile	4.46 (2.05-9.71)	< 0.001	5.34 (2.44-11.7)	< 0.001

Incidence of TCMR2-3 after living donor kidney transplantation % 25 20 15 10 5 0 18-39 yrs 40-55 yrs >55 yrs = PIRCHE DR/DQ low PIRCHE DR/DQ medium = PIRCHE DR/DQ high Incidence of TCMR2-3 after deceased donor kidney transplantation % 25 20 15 10 5 0 18-39 yrs 40-55 yrs >55 yrs = PIRCHE DR/DQ low = PIRCHE DR/DQ medium = PIRCHE DR/DQ high FIGURE 3 Incidence of vascular rejection (TCMR Banff grade 2-3) in relation to recipient age groups and PIRCHE-II DR/DQ tertiles for living and deceased

TABLE 3 Uni- and multivariate Cox proportional hazard analysis for T-cell mediated rejection Banff grade 2-3.

donor kidneys.

transplant recipients that the PIRCHE-II scores for HLA-DQB1 was significantly higher in the TCMR group than in the no-TCMR group, corresponding to our findings that specifically the PIRCHE-II score for the HLA-DR/DQ locus was associated with vascular rejection (14). Finally, Naef et al. observed an association between PIRCHE-II scores for HLA class II and TCMR as well; kidney transplant recipient with low-level BK polyomavirus-DNAemia who developed TCMR had significantly higher PIRCHE-II scores for HLA-DR compared to those who did not develop TCMR (17). A very recently published paper also investigated the relation between PIRCHE II scores and TCMR in a large cohort of kidney transplant patients (25). Similar to the results of this study, only high PIRCHE II scores for HLA class II but not class I were found to be associated with TCMR. However, whether vascular TCMR or not was specifically related to the PIRCHE II score was not stated. In addition, an unknown number of recipients were given T cell depleting induction therapy which can obscure such a relation.

This study emphasizes the importance of the PIRCHE-II score for HLA II donor-derived epitopes in relation to the incidence of aTCMR, especially vascular rejection, which has in general the worst prognosis of the different types of TCMR. To specifically start a vascular and not a tubulo-interstitial rejection, it seems likely that the donor endothelial cells are involved. Renal endothelial cells constitutively express HLA-II

which increases shortly after transplantation and provide the source for donor HLA-II and the peptides (26, 27). Recently, Abrahimi et al, observed that aTCMR mediated by CD8⁺ T cells requires help from CD4⁺ T cells activated by recognition of HLA class II on the surface of donor endothelial cells (28). Following interaction with these donor HLA II molecules on the endothelial cells, the alloreactive CD4⁺ T cells become activated and can migrate across the endothelium (28, 29) (Figure 4). However, since the activation of these directly activated CD4⁺ T cells does not require priming by APC's (30) it seems unlikely that the indirect pathway of allorecognition plays a role in this process. Therefore, this mode of action does not explain the association between PIRCHE-II and aTCMR.

The presentation of intact and processed MHC class I alloantigens by recipient dendritic cells (the semi-direct pathway of allorecognition (31, 32) could provide an explanation for the association between PIRCHE-II and aTCMR, since it allows linked help to be delivered by indirect-pathway CD4⁺ T cells for generating destructive cytotoxic $CD8^+$ T-cell alloresponses (5, 6). An intact donor HLA molecule is acquired by the recipient APC and presented as an intact protein on the cell surface. The recipient APC loaded with donor-derived peptides on their HLA II molecules can stimulate recipient CD4⁺ T cells in e.g. the lymph node. When these APCs are simultaneously cross dressed with intact donor HLA molecules via the semi-direct pathway, the indirectly activated CD4⁺ T cells can provide help for

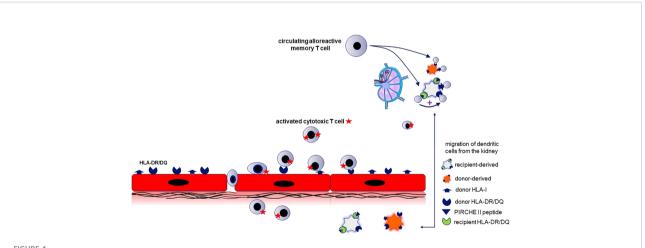


FIGURE 4

The relation between the PIRCH-II scores and acute vasculair rejection indicates that indirect antigen presentation of donor-derived MHC molecules potentiates the alloreactive cytotoxic T cell response. The mechanism proposed is semi-direct antigen presentation combined with direct antigen presentation. This requires that recipient dendritic cells (DC) cross-dressed with donor HLA-molecules (causing direct antigen presentation) is combined with indirect antigen presentation by the same DC to recipient T cells by via immunogenic donor HLA-II peptides (predicted by the PIRCHE score) loaded on the recipient HLA-II molecules. Renal endothelial cells (in red) constitutively express HLA-I and -II and are the source for intact donor-derived HLA molecules and the donor HLA-derived peptides that can be presented by recipient HLA-II (PIRCHE-II peptide). The recipient kidney DC (light grey cell) loaded with donor HLA II peptides on their HLA II molecules migrates to lymphoid tissue and stimulates recipient CD4⁺ T cells (indirect pathway). When the recipient dendritic cell is simultaneously cross-dressed with intact donor HLA molecules, the indirectly activated CD4⁺ T cells can provide help for directly alloreactive recipient CD4⁺ or CD8⁺ T cells (known as the semidirect pathway). The indirectly activated recipient $CD4^+T$ cells may also promote the activation of alloreactive T cells recognizing donor HLA on the donor kidney-derived DC (direct pathway; orange cell). The activated alloreactive T cells attack the donor endothelial cells and migrate into the subendothelial tissue (vascular rejection).

directly alloreactive recipient T cells, and thereby lead to aTCMR (7). This hypothesis is illustrated in Figure 4.

The observed association between PIRCHE-II scores and aTCMR was significant for donor HLA DR/DQ and not for donor HLA class I. Potentially, HLA-DR/DQ could be more immunogenic than HLA class I. Although studies have shown a correlation between the degree of HLA-DR/DQ eplet molecular mismatch and T cell-mediated rejection (33, 34), it remains to be investigated whether HLA class II is more or less immunogenic than HLA class I. However, the strong association between PIRCHE-II scores and early vascular aTCMR may aid in the decision which recipients could benefit from T cell depleting induction therapy.

A limitation of this study is that we were not able to correct well for the number of HLA mismatches between donor and recipient. As the PIRCHE-II score is calculated based on HLA alleles, this score is by definition highly correlated with the number of HLA mismatches (34). Therefore, a comparative evaluation of these two tightly-associated parameters cannot be performed by standard regression analysis. One solution is to analyze the impact of the PIRCHE-II score in subcategories of the HLA-mismatch score, but this requires a database containing a sufficient large number of recipients in each subcategory. Such a cross-analysis was recently performed for the large database of the Collaborative Transplant Study and showed an independent effect of the PIRCHE-II score on graft survival, but varying in strength between different categories of HLA-mismatches (35). Our database is too small to perform reliably such an analysis but we did observe in the zero HLA mismatch stratum a significantly higher PIRCHE-II score for recipients with TCMR compared to no TCMR. In addition, kidney biopsies were only performed for cause and not per protocol. This could lead to an underestimation of the TCMR frequency. However, vascular rejection is in virtually all cases associated with a rapid decline in graft function and is unlikely not to be diagnosed.

In conclusion, the PIRCHE-II DR/DQ score is significantly related to TCMR, in particular acute vascular rejection. This observation indicates that an increased availability of donor HLA class II-derived epitopes for indirect antigen presentation, leading to activation of indirectly alloreactive CD4⁺ T cells, favors the development of this subtype of TCMR.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding author.

Ethics statement

The studies involving human participants were reviewed and approved by Research Ethics Committee for Biobanks at the Erasmus MC and the Medical Ethics Committee of the University Medical Center Utrecht. The patients/participants provided their written informed consent to participate in this study.

Author contributions

MB: Writing, study design, data analysis. EP: Writing, data analysis. HO: Writing, data collection, data analysis. ES: Writing, data analysis. All authors contributed to the article and approved the submitted version.

Funding

This study was supported by research funding from the Dutch Kidney Foundation Project code CP12.23 Risk assessment of kidney graft failure by HLA antibody profiling.

Acknowledgments

The authors wish to acknowledge the contribution of the renal pathology team over the years, in particular Prof. Weening (in memoriam), I Bajema, J von der Thussen and M. Clahsenvan Groningen.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

1. Ho J, Okoli GN, Rabbani R, Lam OLT, Reddy VK, Askin N, et al. Effectiveness of T cell-mediated rejection therapy: A systematic review and metaanalysis. *Am J Transplant* (2021) 22(3):772–85. doi: 10.1111/ajt.16907

2. Betjes MGH, Roelen DL, van Agteren M, Kal-van Gestel J. Causes of kidney graft failure in a cohort of recipients with a very long-time follow-up after transplantation. *Front Med (Lausanne)* (2022) 9:842419. doi: 10.3389/fmed.2022.842419

3. Grazia TJ, Pietra BA, Johnson ZA, Kelly BP, Plenter RJ, Gill RG. A two-step model of acute CD4 T-cell mediated cardiac allograft rejection. *J Immunol* (2004) 172(12):7451–8. doi: 10.4049/jimmunol.172.12.7451

4. Gallon L, Watschinger B, Murphy B, Akalin E, Sayegh MH, Carpenter CB. The indirect pathway of allorecognition. the occurrence of self-restricted T cell recognition of allo-MHC peptides early in acute renal allograft rejection and its inhibition by conventional immunosuppression. *Transplantation* (1995) 59 (4):612–6. doi: 10.1097/00007890-199502270-00029

5. Harper SJ, Ali JM, Wlodek E, Negus MC, Harper IG, Chhabra M, et al. CD8 T-cell recognition of acquired alloantigen promotes acute allograft rejection. *Proc Natl Acad Sci U.S.A.* (2015) 112(41):12788–93. doi: 10.1073/pnas.1513533112

6. Marino J, Paster J, Benichou G. Allorecognition by T lymphocytes and allograft rejection. Front Immunol (2016) 7:582. doi: 10.3389/fimmu.2016.00582

7. Siu JHY, Surendrakumar V, Richards JA, Pettigrew GJ. T Cell allorecognition pathways in solid organ transplantation. *Front Immunol* (2018) 9:2548. doi: 10.3389/fimmu.2018.02548

8. Geneugelijk K, Spierings E. PIRCHE-II: an algorithm to predict indirectly recognizable HLA epitopes in solid organ transplantation. *Immunogenetics* (2020) 72(1-2):119–29. doi: 10.1007/s00251-019-01140-x

9. Tomosugi T, Iwasaki K, Sakamoto S, Niemann M, Spierings E, Nahara I, et al. Clinical significance of shared T cell epitope analysis in early *De novo* donor-specific anti-HLA antibody production after kidney transplantation and comparison with shared b cell epitope analysis. *Front Immunol* (2021) 12:621138. doi: 10.3389/fimmu.2021.621138

10. Geneugelijk K, Niemann M, Drylewicz J, van Zuilen AD, Joosten I, Allebes WA, et al. PIRCHE-II is related to graft failure after kidney transplantation. *Front Immunol* (2018) 9:321. doi: 10.3389/fimmu.2018.00321

11. Peereboom ETM, Matern BM, Tomosugi T, Niemann M, Drylewicz J, Joosten I, et al. T-Cell epitopes shared between immunizing HLA and donor HLA associate with graft failure after kidney transplantation. *Front Immunol* (2021) 12:784040. doi: 10.3389/fimmu.2021.784040

12. Lachmann N, Niemann M, Reinke P, Budde K, Schmidt D, Halleck F, et al. Donor-recipient matching based on predicted indirectly recognizable HLA epitopes independently predicts the incidence of *De novo* donor-specific HLA antibodies following renal transplantation. *Am J Transplant* (2017) 17(12):3076–86. doi: 10.1111/aji.14393

13. Sakamoto S, Iwasaki K, Tomosugi T, Niemann M, Spierings E, Miwa Y, et al. Analysis of T and b cell epitopes to predict the risk of *de novo* donor-specific antibody (DSA) production after kidney transplantation: A two-center retrospective cohort study. *Front Immunol* (2020) 11:2000. doi: 10.3389/fimmu.2020.02000

14. Ono K, Ide K, Tanaka Y, Ohira M, Tahara H, Tanimine N, et al. Molecular mismatch predicts T cell-mediated rejection and *De novo* donor-specific antibody formation after living donor liver transplantation. *Liver Transplant* (2021) 27 (11):1592–602. doi: 10.1002/lt.26238

15. Vionnet J, Miquel R, Abraldes JG, Wall J, Kodela E, Lozano JJ, et al. Noninvasive alloimmune risk stratification of long-term liver transplant recipients. *J Hepatol* (2021) 75(6):1409–19. doi: 10.1016/j.jhep.2021.08.007

16. Lezoeva E, Nilsson J, Wüthrich R, Mueller TF, Schachtner T. High PIRCHE scores may allow risk stratification of borderline rejection in kidney transplant recipients. *Front Immunol* (2022) 13. doi: 10.3389/fimmu.2022.788818

17. Naef B, Nilsson J, Wuethrich RP, Mueller TF, Schachtner T. Intravenous immunoglobulins do not prove beneficial to reduce alloimmunity among kidney transplant recipients with BKV-associated nephropathy. *Transplant Int* (2021) 34 (8):1481–93. doi: 10.1111/tri.13882

18. Otten HG, Joosten I, Allebes WA, van der Meer A, Hilbrands LB, Baas M, et al. The PROCARE consortium: toward an improved allocation strategy for

kidney allografts. Transpl Immunol (2014) 31(4):184-90. doi: 10.1016/ j.trim.2014.09.008

19. Kamburova EG, Wisse BW, Joosten I, Allebes WA, van der Meer A, Hilbrands LB, et al. Differential effects of donor-specific HLA antibodies in living versus deceased donor transplant. *Am J Transplant* (2018) 18(9):2274–84. doi: 10.1111/ajt.14709

20. Geneugelijk K, Wissing J, Koppenaal D, Niemann M, Spierings E. Computational approaches to facilitate epitope-based HLA matching in solid organ transplantation. *J Immunol Res* (2017) 2017:9130879. doi: 10.1155/2017/9130879

21. Betjes MGH, Sablik KS, Otten HG, Roelen DL, Claas FH, de Weerd A. Pretransplant donor-specific anti-HLA antibodies and the risk for rejection-related graft failure of kidney allografts. *J Transplant* (2020) 2020:5694670. doi: 10.1155/2020/5694670

22. Rampersad C, Balshaw R, Gibson IW, Ho J, Shaw J, Karpinski M, et al. The negative impact of T cell-mediated rejection on renal allograft survival in the modern era. *Am J Transplant* (2021) 22(3):761–71. doi: 10.1111/ajt.16883

23. Kidney Disease: Improving Global Outcomes Transplant Work G. KDIGO clinical practice guideline for the care of kidney transplant recipients. *Am J Transplant: Off J Am Soc Transplant Am Soc Transplant Surgeons* (2009) 9:S1–S155. doi: 10.1111/j.1600-6143.2009.02834.x

24. Tullius SG, Tran H, Guleria I, Malek SK, Tilney NL, Milford E. The combination of donor and recipient age is critical in determining host immunoresponsiveness and renal transplant outcome. *Ann Surg* (2010) 252 (4):662–74. doi: 10.1097/SLA.0b013e3181f65c7d

25. Senev A, Van Loon E, Lerut E, Coemans M, Callemeyn J, Daniëls L, et al. Association of predicted HLA T-cell epitope targets and T cell-mediated rejection after kidney transplantation. *Am J Kidney Dis* (2022). doi: 10.1053/j.ajkd.2022.04.009

26. Mannam VK, Lewis RE, Cruse JM. The fate of renal allografts hinges on responses of the microvascular endothelium. *Exp Mol Pathol* (2013) 94(2):398–411. doi: 10.1016/j.yexmp.2012.06.002

27. Muczynski KA, Ekle DM, Coder DM, Anderson SK. Normal human kidney HLA-DR-expressing renal microvascular endothelial cells: characterization, isolation, and regulation of MHC class II expression. *J Am Soc Nephrol* (2003) 14(5):1336–48. doi: 10.1097/01.ASN.0000061778.08085.9F

28. Abrahimi P, Qin L, Chang WG, Bothwell AL, Tellides G, Saltzman WM, et al. Blocking MHC class II on human endothelium mitigates acute rejection. *JCI Insight* (2016) 1(1):e85293. doi: 10.1172/jci.insight.85293

29. Manes TD, Pober JS. Polarized granzyme release is required for antigendriven transendothelial migration of human effector memory CD4 T cells. *J Immunol* (2014) 193(12):5809–15. doi: 10.4049/jimmunol.1401665

30. Shiao SL, McNiff JM, Pober JS. Memory T cells and their costimulators in human allograft injury. *J Immunol* (2005) 175(8):4886–96. doi: 10.4049/jimmunol.175.8.4886

31. Sivaganesh S, Harper SJ, Conlon TM, Callaghan CJ, Saeb-Parsy K, Negus MC, et al. Copresentation of intact and processed MHC alloantigen by recipient dendritic cells enables delivery of linked help to alloreactive CD8 T cells by indirect-pathway CD4 T cells. *J Immunol* (2013) 190(11):5829–38. doi: 10.4049/jimmunol.1300458

32. Brown K, Sacks SH, Wong W. Coexpression of donor Peptide/Recipient MHC complex and intact donor MHC: Evidence for a link between the direct and indirect pathways. *Am J Transplant* (2011) 11(4):826–31. doi: 10.1111/j.1600-6143.2011.03437.x

33. Wiebe C, Rush DN, Gibson IW, Pochinco D, Birk PE, Goldberg A, et al. Evidence for the alloimmune basis and prognostic significance of borderline T cellmediated rejection. *Am J Transplant* (2020) 20(9):2499–508. doi: 10.1111/ajt.15860

34. Rampersad C, Balshaw R, Gibson IW, Ho J, Shaw J, Karpinski M, et al. The negative impact of T cell-mediated rejection on renal allograft survival in the modern era. *Am J Transplant* (2022) 22(3):761–71. doi: 10.1111/ajt.16883

35. Unterrainer C, Döhler B, Niemann M, Lachmann N, Süsal C. Can PIRCHE-II matching outmatch traditional HLA matching? *Front Immunol* (2021) 12:631246. doi: 10.3389/fimmu.2021.631246