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Influence of gut microbial metabolites on tumor immunotherapy: mechanisms and potential natural products

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In recent years, tumor immunotherapy has made significant breakthroughs in the treatment of malignant tumors. However, individual differences in efficacy have been observed in clinical practice. There is increasing evidence that gut microbial metabolites influence the efficacy of distal tumor immunotherapy via the gut-liver axis, the gut-brain axis and the gut-breast axis, a process that may involve modulating the expression of immune cells and cytokines in the tumor microenvironment (TME). In this review, we systematically explore the relationship between gut microbial metabolites and tumor immunotherapy, and examine the corresponding natural products and their mechanisms of action. The in-depth exploration of this research area will provide new ideas and strategies to enhance the efficacy of tumor immunotherapy and mitigate adverse effects.

KEYWORDS

gut microbial metabolites, tumor, natural products, immune, mechanism

1 Introduction

Tumor immunotherapy has recently emerged as a breakthrough approach to the treatment of malignant tumors, and unlike traditional cancer treatments such as radiotherapy and chemotherapy, immunotherapy harnesses the host's immune system to target and eliminate cancer cells (1). In recent decades, with the continuous progress of our understanding of the cancer immunosuppressive microenvironment, immunotherapy has become the key pillar of cancer treatment, including immunosuppressive, chimeric antigen receptor (CAR) therapy, cancer vaccination and oncolytic virus therapy (1). However, long-term chronic stimulation of immune cells by tumor antigens and the uncontrolled inflammation associated with carcinogenesis ultimately impairs anti-tumor immunity and

promotes tumor progression (2), resulting in a situation where not all patients benefit and some may even suffer serious immunerelated adverse events (3).

The gut flora is the most important metabolic organ in the human body and consists of a wide range of microorganisms, including bacteria, archaea, viruses, unicellular eukaryotes and fungi (4). Numerous studies have shown that gut microbes influence cancer development through a variety of mechanisms, in which bacterial metabolites may play an important role (5, 6). Bacterial metabolites such as short-chain fatty acids (SCFAs), bile acids, lactic acid, spermidine, indole and retinoic acid have been shown to link the gut microbiota to systemic immunity, which in turn influences cancer development (7). For example, microbial metabolites have been shown to influence the initiation and progression of endocrine tumors by altering various signaling pathways (8-10). Acetic acid improves pancreatitis and its sequelae and may reduce the incidence of pancreatic cancer (11). Lactate-mediated epigenetic reprogramming regulates the formation of human pancreatic cancer-associated fibroblasts and the ultrastructural differentiation of sodium butyrate-treated human pancreatic cancer cell lines, alterations that have the potential to increase the invasiveness of pancreatic cancer cells (12, 13). It has also been shown that butyrate has an inhibitory effect on the proliferation of pancreatic cancer cells by inducing a specific secretory phenotype through structural changes (14). A study of patients with pancreatic cancer showed that all diagnosed patients had elevated bile acid levels (15). Other studies have shown that patients with ovarian cancer have reduced blood levels of tryptophan and indolepropionic acid, a trend that becomes more pronounced as the disease progresses (16-19).

In recent years, microbial metabolites have been shown to play an important role in tumor immunotherapy by activating the immune system to eliminate tumor cells and prevent the development of drug resistance (20). Therefore, in-depth exploration of the crosstalk between gut microbial metabolites and tumor immunotherapy, as well as the mining of related natural products and their potential roles, is of great importance to further alleviate the side effects of cancer therapies and improve the cure rate. In this paper, we will systematically review the relationship between gut microbial metabolites and tumor immunotherapy, and mine related natural products to explore their potential challenges and directions.

2 Sources and classification of gut microbial metabolites

Depending on the food source, gut microbial metabolites can be classified as SCFAs, bile acids, phenolics, vitamins, polyamines, tryptophan and lipids (21), which play an important role in tumorigenesis and progression (22–24). SCFAs are saturated fatty acids with chain lengths of one to six carbon atoms, which are the main product of fiber fermentation in the colon and are not absorbed in the small intestine (25), and are metabolized mainly by the phylum *Thick Walled Bacteria*, including *Fusobacterium harryi*, *Fusobacterium*

rectum and E. fecalis prausnitzii, Clostridium sporotrichum, Bifidobacterium bifidum, Propionibacterium sp, Pseudomonas sp and Lactobacillus sp (26, 27). SCFAs include mainly acetate, propionate and butyrate, with formate, valerate and hexanoate being the least abundant (28, 29), of which butyrate is the most studied SCFA, which not only maintains the balance of the intestinal flora, but also has an inhibitory effect on tumorigenesis, which may reduce the risk of colorectal cancer (30-33). Tryptophan is one of the essential amino acids and the only amino acid with an indole structure, which is found mainly in protein-rich foods such as poultry, fish, dairy products and soya beans. Studies have shown that tryptophan can be metabolized by the intestinal microflora to produce a variety of metabolites such as lactic acid, propionic acid, acrylic acid, tryptamine, etc., which may be involved in the immune response process (34). In addition, indole and indole metabolites produced by bacterial pathways, including indole-3acetic acid (IAA), indole-3-aldehyde (IAld), indole-3-propionic acid (IPA), indole-3-acetamide (34-36), which can act as activators of aryl hydrocarbon receptor (AHR) or pregnane X receptor (PXR) signaling and play an important role in the regulation of the host immune response (37). AHR or PXR signaling activators play key roles in regulating immune responses, metabolic processes, and cellular responses to environmental signals (37, 38). Most of the intestinal bacteria involved in metabolism are of the Bacillus genus. Escherichia coli (E. coli) metabolizes tryptophan to indole and pyruvate (39), and Bacillus sphaericus metabolizes tryptophan to indole, which is further metabolized to form IAld, which can be oxidized to IAA by oxidoreductases or converted to tryptamine by decarboxylases. Inosine is mainly derived from endogenous purine nucleosides formed from the deamination metabolism of the adenine portion of nucleic acid by intestinal microorganisms and has the function of regulating the intestinal microbiota and protecting the intestinal mucosal barrier (40). Trimethylamine oxides are derived from the large amounts of choline or carnitine present in foods such as fish, eggs and meat products, which can be metabolized by the intestinal microflora Fusobacterium, Anaplasma phagocytophilum, Aeromonas phagocytophilum, Clostridium phagocytophilum and Aspergillus phagocytophilum to produce trimethylamine (TMA), which passes through the portal circulation to the liver where it is catalyzed to produce trimethylamine oxide (TMAO) (41). Secondary bile acids are converted from primary bile acids through metabolism by the gut microbiota, whereas primary bile acids are synthesized in the liver and are involved in fat metabolism in the hepatointestinal circulation (42, 43). Polyphenols are water-soluble organic compounds found in fruits, vegetables and herbs, and metabolites of polyphenols in the gut flora, such as phenolic acids and flavonoid derivatives, have been shown to directly or indirectly influence cancer development by stimulating the host immune response and reducing oxidative stress and inflammation (44).

3 The basic principles of tumor immunotherapy

The immune system plays a continuous role in surveillance and protection in the human body. During the early stages of tumor development, a large number of white blood cells, including various subsets of T cells and dendritic cells, typically accumulate in the TME. These cells can specifically recognize and eliminate tumor cells (45). However, during tumor development, multiple mechanisms of immune evasion and suppression evolve, including altering tumor antigens to prevent recognition by T cells (46, 47), blocking T cell recruitment to the TME (48), and utilizing immunosuppressive white blood cells such as Tregs and macrophages (49). *James Allison* and *Tasuku Honjo*'s discovery of T-cell inhibitory signaling pathways has brought immunotherapy to widespread attention. When these pathways are activated, they prevent effective cancer immunization.

3.1 Immune checkpoint inhibitors

Immune checkpoint inhibitors (ICIs) can effectively restore or enhance the reactivity of anti-tumor T cells, preventing immune escape by cancer cells. The most prominent of these are cytotoxic Tlymphocyte-associated protein 4 (CTLA-4) monoclonal antibodies and programmed cell death protein 1 (PD-1) monoclonal antibodies. CTLA-4 is a protein receptor expressed on T cells, which binds to CD80 and CD86 on dendritic cells with higher affinity than CD28 (50). This binding triggers intracellular inhibitory signaling that leads T cells into a state of nonresponsiveness (51). CTLA-4 monoclonal antibodies can block the immunosuppressive effects of CTLA-4, reducing Treg cells and increasing effector T cells (52). In several animal model experiments, blocking CTLA-4 has been shown to effectively enhance anti-tumor effects (53, 54). PD-1 is a surface receptor for its homologous ligands programmed cell death ligand 1 (PD-L1) and programmed cell death ligand 2 (PD-L2). It is primarily expressed on activated T cells, but also present on other white blood cell subsets, including activated B cells, dendritic cells (DC), monocytes, and natural killer (NK) cell (55, 56). PD-L1 is found on activated T cells, B cells, dendritic cells, macrophages, and many tissue cells (57), whereas PD-L2 is only expressed on dendritic cells, macrophages, and some specialized cells (57). PD-1 can inhibit T cell activation through multiple pathways. In effector T cells, PD-1 activation leads to the dephosphorylation of key signaling molecules downstream of the T cell receptor (TCR), thereby suppressing TCRmediated T cell activation (58). Other studies have shown that the PD-1/PD-L1 interaction preferentially triggers dephosphorylation of CD28, which is one of the main mechanisms of T cell suppression (59). PD-1 also enhances T cell motility, hindering T cell interactions with dendritic cells. Additionally, it has been reported that PD-1 can bind to CD80 and potentially compete with CD28 like CTLA-4, thereby inhibiting T cell activation. Cancer cells can utilize the PD-1 pathway to upregulate its ligands, PD-L1 and PD-L2, to inhibit T cell-mediated apoptosis. Multiple studies have been shown that upregulated PD-1 expression on tumor-infiltrating lymphocytes is associated with poor prognosis in many human cancers (60). CTLA-4 and PD-1 provide key physiological immune regulatory mechanisms, and extensive experimental data have shown that blocking these checkpoints with antibodies can enhance anti-tumor immune responses in animal models. This discovery has led to the development of humanized monoclonal antibodies targeting these molecules. These novel immunotherapies are known as ICIs. In 2011, the U.S. approved the first CTLA-4 inhibitor, Ipilimumab, for the treatment of metastatic melanoma (61). In a phase III clinical trial, 600 melanoma patients previously treated with other therapies were divided into three experimental groups: Ipilimumab combined with the gp100 vaccine, Ipilimumab alone, and gp100 alone. The results showed that the combination therapy group had the longest median overall survival (OS) (62). Currently, ICIs are widely used in cancer immunotherapy and have been approved for melanoma, non-small cell lung cancer (NSCLC), renal cell carcinoma, bladder cancer, and head and neck squamous cell carcinoma. Although ICIs can activate T cells and induce durable anti-tumor responses, this treatment can also lead to specific immune activation in non-tumor organs, causing immune-related adverse events (irAEs). Some adverse events, such as rashes, rheumatism, and others, are not life-threatening but can reduce the patient's quality of life, while other irAEs, such as pneumonia, myocarditis, hepatitis, and irAEs affecting the nervous and hematologic systems, can be fatal (63). Kyoko et al. reported a case of a metastatic breast cancer patient receiving combination treatment with Atezolizumab and paclitaxel, who developed a rare (1%) neurological irAE, leading to severe, permanent peripheral neuropathy (64). Furthermore, ICIs largely rely on the pre-existing presence of tumor-infiltrating cytotoxic T lymphocytes to exert their anti-tumor effects. In some patients, the immune cell concentration in the TME is low, and even when activated, the immune response is minimal.

3.2 Chimeric antigen receptor-T cell therapy

CAR T cell therapy has opened a new era in cancer treatment (65). Synthetic CAR constructs typically include an extracellular domain derived from the single-chain variable fragment of an antibody, as well as a hinge and transmembrane domain (53). The functional end of CAR usually contains activation and costimulatory domains, known as intracellular domains (66, 67). Commonly used co-stimulatory molecules include CD28 and 4-1BB. When the antigen activates this receptor, it triggers TCR signaling, co-stimulatory signaling, and cytokine signaling. The synergistic transmission of these three signals fully satisfies T cell activation and proliferation (68). CAR-T cell therapy has achieved tremendous success in treating B cell malignancies, including leukemia and lymphoma. The CAR-T cell therapies Tisagenlecleucel and Axicabtagene ciloleucel were approved by the FDA and the European Medicines Agency in 2017 and 2018, respectively, for the treatment of acute lymphoblastic leukemia, targeting the CD19 antigen on B cells (69). In 2021, the FDA approved the first cellular therapy for multiple myeloma (MM), idecabtagene vicleucel (70). CAR-T cells have also shown remarkable effects and prospects in lymphoma treatment. Axi-Cel, a CD19+ CAR-T cell therapy, has demonstrated significant efficacy in refractory large B cell lymphoma, showing good response levels in 28 treated patients (71). This drug was approved for the

treatment of follicular lymphoma (FL), as 94% of the 80 evaluable FL patients in clinical trials showed a response, with 79% achieving complete remission (CR) (72). The FDA approved the CAR-T cell therapy Breyanzi[®] for the treatment of mantle cell lymphoma (MCL), which has been shown to be effective and safe in phase II clinical trials in patients with relapsed/refractory MCL (73). Despite the significant progress has been shown in CAR-T cell therapy clinical trials, the presence of adverse events during treatment has affected its effectiveness. The most common is off-target effects, where the same target antigen is expressed on normal cells, causing CAR-T cells to attack healthy tissue, leading to adverse reactions. Notably, most of these toxic reactions are reversible if the patient receives timely intervention or treatment (74). Antigen escape is another major challenge faced by CAR-T therapy. Tumor cells escape killing by promoting mutations in genes encoding antigens, leading to downregulation or loss of the alternative antigen that lacks CAR-T cell targeting epitopes (75). For example, although 70% to 90% of relapsed or refractory ALL patients exhibit long-term responses to CD19-targeting CAR-T cell therapy, recent follow-up data indicate a common resistance mechanism, with 30% to 70% of patients who relapse after treatment showing downregulation or

loss of the CD19 antigen (76). Furthermore, in the solid TME, immune suppression and fibrosis greatly limit the targeted therapeutic effects of CAR-T cells. To date, no CAR-T cell therapy has been approved for solid tumors.

3.3 Therapeutic vaccines

Therapeutic cancer vaccination is an active immunization strategy aimed at stimulating adaptive immune responses against tumor antigens and generating tumor-specific functional immune effector cells, such as cytotoxic T lymphocytes (77). Broadly, cancer vaccines can be classified into cell-based vaccines, viral vector vaccines, and molecular vaccines composed of peptides, deoxyribonucleic acid (DNA), or ribonucleic acid (RNA). Cancer vaccines targeting cells use non-replicating cancer cells or antigenpresenting cells (APCs) carrying cancer antigens (78). Sipuleucel-T (Provenge[®]) is the first FDA-approved therapeutic cancer vaccine, which improved OS in patients with metastatic castration-resistant prostate cancer in the Phase 3 trial (79). Cancer vaccines can mimic natural immune processes, demonstrating good safety and therapeutic efficacy in numerous clinical trials (80). In an early study, Hoover et al. showed survival benefits of autologous tumor cell vaccines in colon cancer, highlighting the broad potential of cancer vaccines (80). In melanoma, Mackiewicz et al. observed the highest proportion of complete responders in a trial using allogeneic whole-cell vaccines (81). In 2020, Lv et al. conducted a Phase II randomized controlled trial of DC vaccines glioblastoma multiforme (GBM) and observed improved survival rates (82). Yamanaka et al., Cho et al., and Yao et al. compared the effects of dendritic cell vaccines and conventional therapies in glioblastoma patients, with the vaccine treatment group showing significant survival benefits (83). However, it must be acknowledged that the anticipated efficacy of cancer vaccines has not been perfectly realized in clinical settings. Despite extensive preclinical and clinical work, their effectiveness remains unpredictable, and there are significant negative feedbacks.

4 Crosstalk between gut microbial metabolites and tumor immunity

Gut microbial metabolites can diffuse across the epithelium and lamina propria into the somatic circulation, and studies have shown that a large number of microbial molecules have been detected in the human bloodstream, of which 5-10% are derived from the gut microbiota, and that these microbial molecules may act as regulators of cellular functions in distal organs (84, 85). In addition, gut microbial metabolites can enter the circulation along the gut-X axis to reach other target organs or target cells, act as human hormone-like signaling mediators to influence the homeostasis of the local environment (86–90), participate in the release of cytokines in the TME and in the development and differentiation of immune cells, thereby inhibiting immune escape from tumors and indirectly influencing the response to various classical immunotherapies (91–95) (Figure 1).

4.1 Immunoregulation of the gut-liver axis

Studies have shown that the gut microbiome of patients with liver pathology differs significantly from that of healthy individuals (96), and gut microbiology and liver function have been found to be linked to physiological anatomy. Fecal microbiota transplantation (FMT) using fecal samples from patients with hepatitis C virus-associated chronic liver disease was found to promote liver tumor growth in mice (97). Ponziani et al. investigated the changes in microbial populations in patients with hepatocellular carcinoma (HCC) as well as in healthy patients and found that the abundance of Eosinophilus spp., Enterococcus spp., Streptococcus spp., Dictyococcus spp. and Cholera spp. were higher in HCC patients than in controls, and that there was a decrease in Bifidobacterium Akerman, Bifidobacterium bifidum and Bifidobacterium minutissimus spp. and an increase in plasma interleukin-8 and interleukin-13 (98). Interestingly, gut microbes and their metabolites play a crucial role in maintaining the physiological state and metabolic homeostasis of the host. For example, Clostridium difficile metabolically alters the balance of primary and secondary bile acids, which in turn induces the production of C-X-C motif chemokine ligand 16 (CXCL16) in liver sinusoidal endothelial cells, and CXCL16 can induce NK cells to accumulate in the liver and generate anti-tumor immunity (99). In addition, deoxycholic acid (DCA) induces hepatic stellate cells to express a senescence-associated secretory phenotype, which promotes immune expression to eliminate senescent cells and induces fibrosis and carcinogenesis (100, 101). In addition, inulin can produce anti-inflammatory SCFAs through intestinal microbial metabolism, and oral inulin administration promotes CD8⁺ T-cell expression and enhances the therapeutic effect of PD-1 checkpoint immunosuppressants (102). Lee et al. (103) conducted a fecal analysis of 41 patients with



unresectable liver cancer, including 20 ICI responders and 21 nonresponders. The results revealed that the concentration of ursodeoxycholic acid (UDCA) in the feces of ICI responders was significantly higher, while the concentration of lithocholic acid was increased in the feces of patients. This suggests that bile acids are associated with the response of HCC patients to ICI treatment. Han et al. (104) found that the intestinal microbial metabolite Dlactic acid (DL) reaches the liver via the portal vein and converts tumor-associated macrophages (TAMs) from M2 to M1, enhancing the phagocytic function of Kupffer cells. The mechanism involves the inhibition of the phosphoinositide 3kinase (PI3K)/protein kinase B (Akt) pathway and the activation of the nuclear factor kappa B (NF-κB) pathway. Zheng et al. (105) performed dynamic analysis of HCC patients receiving anti-PD-1 immunotherapy in the 6th week and found significant differences in the gut microbiome diversity between immune responders and non-responders. The fecal samples of immune responders showed higher taxonomic richness and gene counts, enriched in Myxobacteria and Ruminococcaceae, which influenced the efficacy of anti-PD-1 immunotherapy and disease prognosis in HCC patients. Dysbiosis of the gut microbiome can also impact resistance to immunotherapy. Arielle et al. (106) demonstrated that antibiotic-induced dysbiosis reduces ICI activation of the immune system, leading to resistance and poor treatment response. Vetizou (107) showed that Bacteroides fragilis and Bacteroides thetaiotaomicron can regulate specific T cell responses. FMT

from ICI responders helped non-responders recover the anticancer effects of PD-1 and CTLA-4 immune checkpoint blockade (108).

4.2 Immunomodulation of the gutendocrine axis

The gut microbiota is recognized as an endocrine organ capable of influencing distal organs and related biological pathways. Dysbiosis of the gut microbiome can lead to diseases such as breast, cervical and ovarian cancer in women, and gut microbial metabolites are important mediators of associated diseases (109). Gastrointestinal flora can influence non-ovarian estrogen levels via the hepatointestinal cycle (110), for example, butyrate has been shown to regulate luteinizing hormone and estradiol secretion via the cyclic adenosine monophosphate (cAMP) signaling pathway (111). Liu et al. showed that butyrate supplementation alleviated non-alcoholic fatty liver disease in ovariectomized mice (112). Another study showed higher levels of Prevotella in healthy women compared to breast cancer patients (113). James et al. (114) also demonstrated that the diversity of the gut microbiome is reduced in breast cancer patients, with an increased relative abundance of Clostridia. Miko and Luu et al. (115, 116) found that the reduction in microbial diversity is most pronounced in earlystage breast cancer (Stage 0 and Stage 1). The reduction in gut

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microbiome diversity directly affects the levels of metabolic products and increases the incidence of breast cancer (117). In a mouse study, the use of ampicillin exacerbated the reduction in gut microbiome diversity and induced tumor formation, which may be related to the decreased abundance of Odoribacter and Anaeotruncus, both of which are bacteria that produce butyrate (118). In addition, Mikoah et al. found that lithocholic acid (LCA) inhibited the proliferation of MCF7, SKBR3, and 4T1 breast cancer cells, and tested the cytostatic properties of LCA in mice transplanted with 4T1 breast cancer cells, and found a significant reduction in the ability of the primary tumors to infiltrate the surrounding tissues and metastasize after LCA treatment, which suggests that LCA can be transferred through the bloodstream to the breast and may play an important role in promoting the antiproliferative effects of breast cancer (119). In addition to affecting tumor progression, gut microbiome metabolites also influence the responsiveness to immunotherapy. In a human study, strains were extracted from the feces of breast cancer patients who responded and did not respond to trastuzumab treatment. Mice receiving FMT from responders also showed a response to trastuzumab, whereas those receiving FMT from nonresponders did not. This was associated with a higher abundance of Clostridia in the gut of trastuzumab responders, which is capable of producing SCFAs (120).

4.3 Immunomodulation of the gutbrain axis

The complex relationship between the gut microbiota and host health can be further understood through the gut-brain axis (GBA) and its associated biological activities. Gut microbial metabolites can cross the blood-brain barrier through the body's circulation and, because of their small molecular size, play an immunomodulatory role in the gut-brain axis. Gut microbial tryptophan metabolites can signal to the brain, suggesting a potential role for metabolites in communication between gut microbes and the central nervous system (CNS) (121, 122), and indole derivatives act as AHR ligands, including IPA and IAA, and have the ability to cross the blood-brain barrier, giving them a key role in the GBA. For example, indole, IPA and IAld can activate AHR signaling in astrocytes and suppress inflammation in the CNS (123). IPA has a strong free radical scavenging and antioxidant capacity, protecting primary neurons and neuroblastoma cells from oxidative damage (124). Circulating SCFAs produced by gut microbiota metabolism affect the integrity of the blood-brain barrier by increasing the production of tight junction proteins, and increased blood-brain barrier integrity reduces the entry of unwanted metabolites into brain tissue and enhances bloodbrain barrier defense mechanisms (125). Compounds produced by gut microbiota metabolism, such as lipoproteins and lipopolysaccharides, affect autoimmune function by stimulating immune cells to release cytokines that can cross the blood-brain barrier and activate neurons, altering neurological function and leading to mood and behavioral changes (126). Zhou et al. used an in situ GBM model and found that gut dysbiosis leads to an increased proportion of M2-like macrophages in the TME. They also observed reduced levels of SCFAs in the blood and glioma tissues of the brain. Oral supplementation with SCFAs induced the differentiation of macrophages into the M1 type in the TME, enhancing the immune response. The mechanism may be related to SCFA activation of glycolysis in macrophages (127). In a study investigating the changes in the gut microbiota of brain tumor patients and the association between the two, 16S rRNA gene amplicon sequence sequencing was used to characterize the gut microflora in 158 patients, and the results showed that the abundance and homogeneity of the gut microbial ecosystem in brain tumor patients was significantly lower than that in healthy controls, as evidenced by an increase in the abundance of pathogenic bacteria and a decrease in the abundance of probiotic bacteria.

4.4 Others

Several studies have shown that gut microbial metabolites also play a critical role in immune modulation in many other types of tumor disease. For example, in one study, supplementation of mice with long-chain fatty acid metabolizing enzymes to reduce levels of butyrate metabolites reduced prostate cancer (PCa) growth and worsening of PCa in mice prior to prostatectomy, demonstrating a cross-talk between the gut microbial metabolite butyrate and prostate cancer (128). Butyrate, as one of the main components of SCFAs, can reduce the risk of pancreatic ductal adenocarcinoma (PDAC), inhibit the proliferation of pancreatic cancer cells, and induce their differentiation into a secretory phenotype with ultrastructural changes. Studies have shown that hyaluronic acid conjugates of butyrate can significantly inhibit cell proliferation in pancreatic cancer cell cultures (129). Another study showed that inosine improved the efficacy of monoclonal antibodies against CTLA-4 and against PD-L1 in mouse models of bladder and small bowel cancer (130). Liu et al. (131) conducted gut microbiome analysis, including metagenomic and metabolomic sequencing, on 54 lung cancer patients who initially received PD-1/PD-L1 treatment. They found that patients with higher levels of acetate, propionate, and butyrate had longer progression-free survival and lower tumor progression risk, suggesting a crosstalk between gut microbial metabolites and the lungs. Andrea et al. (132) characterized the metabolic features of the gut microbiome in 11 NSCLC patients treated with the PD-1 monoclonal antibody, nivolumab. They found that 4 patients with early progression were significantly associated with the gut metabolites 2-pentanone and tridecane, whereas propionate and butyrate in SCFAs were linked to long-term beneficial outcomes in the remaining 7 patients. Motoo Nomura et al. (133) measured SCFA concentrations in feces and plasma from 52 patients with solid tumors receiving PD-1 treatment and found that high concentrations of acetate, propionate, butyrate, and valerate in feces were significantly associated with longer progression-free survival. SCFAs exhibited immune-modulatory functions in the host, possibly through the inhibition of histone deacetylases (HDAC) (134). Two phase I clinical trials showed that FMT could improve response to ICIs in

resistant metastatic melanoma (91). Additionally, choline metabolites TMAO and TMA were found to enhance PDAC response to immune checkpoint blockade and improve survival in tumor-bearing mice. Gauri Mirji et al. treated PDAC model mice with macrophages induced by TMAO and found that tumor burden was reduced by more than 2.4 times compared to mice receiving control macrophage treatment, with significant upregulation of Interferon-y, CD103, and CD44 on CD8+ and CD4+ cells. This suggests that TMAO induces macrophages to differentiate into a phenotype that enhances T-cell effects and suppresses PDAC growth. Similar effects were observed in human macrophages. The authors then used TMAO in combination with PD-1 or Tim-3 antibodies to treat PDAC model mice and found that, compared to the PD-1 and TMAO monotherapy groups, the combination therapy enhanced the immune activation status in the PDAC TME, with stronger activation of bone marrow cells and T cells (135). Gut microbial metabolites not only improve cancer immunotherapy prognosis by enhancing immune responses, but also alleviate immune-related adverse effects. For example, CTLA-4 antibody treatment is often associated with gut-related side effects, such as diarrhea or colitis, while PD-1/PD-L1 antibodies are more commonly associated with thyroid dysfunction or pulmonary toxicity. As a result, many patients can only use these treatments for a short period of time. SCFAs, as nutrients for intestinal epithelial cells, can help repair the gut mucosa, reduce gastrointestinal adverse effects, and help extend the treatment cycle of CTLA-4. Oral probiotics such as Bacteroides fragilis and Burkholderia cepacia can also help alleviate adverse effects (108). The impact of gut microbial metabolites on tumor immunity is not always beneficial and can sometimes promote the occurrence and progression of cancer. For example, secondary bile acids such as DCA and LCA, which are metabolized by gut microbes, are associated with the proliferation, survival, and metastasis of cancer cells (129). In rodent model experiments, it has been shown that elevated serum concentrations of DCA and LCA increase the risk of liver cancer (136). The mechanism is related to the activation of nuclear receptors such as FXR and PXR, which lead to changes in gene expression (137). Clélia Coutzac et al. (138) used a CT26 tumor model in mice treated with intraperitoneal CTLA-4 antibody and administered sodium butyrate in their drinking water to observe the effects of SCFAs on the anti-tumor effect of CTLA-4. The results showed that there was no significant reduction in tumor growth compared to the control group, suggesting that butyrate may inhibit the anti-tumor efficacy of CTLA-4 in mice. The underlying reason may be related to its impact on dendritic cell maturation and T-cell function, although the exact mechanism still needs further exploration. Gut microbes and their metabolites have a double-edged sword effect on tumor immunotherapy. Under the influence of factors such as diet, medication, and disease, the balanced distribution of gut microbiota is disrupted, leading to an imbalance in the types of metabolites. This change can affect gut mucosal function, increase intestinal permeability, and allow a large number of small molecular metabolites to pass through the intestinal wall, enter the bloodstream, and accumulate in the TME. These metabolites then regulate immune function, potentially having either positive or

negative effects on tumor immunotherapy. Numerous metabolites have been confirmed to play a role in tumor development. By orally supplementing or transplanting isolated strains, it is possible to artificially intervene in metabolite levels and influence the effectiveness of tumor immunotherapy. This also opens up new avenues for cancer drug development.

5 Mechanisms by which gut microbial metabolites influence tumor immunity

5.1 Regulation of the innate immune response

The gut microbial tryptophan metabolite indole and its derivatives can directly regulate the growth and differentiation of non-specific immune cells through activation of the AHR pathway (139). For example, it has been shown that IAld and IAA promote the differentiation of monocytes into dendritic cells and enhance the phagocytic activity of neutrophils and macrophages (140, 141), and that tryptophan activates the AHR pathway in NK cells, which enhances cytotoxicity against tumor cells (8, 142, 143). Gut microbial tryptophan metabolites can also mediate the inflammatory response by regulating cytokine expression, in particular stimulating a key role for macrophages. A study in mouse liver showed that IAA attenuated high fat diet-induced hepatotoxicity and inhibited interleukin-1ß (IL-1ß), interleukin-23 (IL-23) and tumor necrosis factor- α (TNF- α), monocyte chemoattractant protein-1 (MCP-1) in a dose-dependent manner, interleukin-17A (IL-17A), interleukin-6 (IL-6), and other inflammatory factors, while increasing the levels of the antiinflammatory factor interleukin-10 (IL-10) and decreasing the proinflammatory factor/anti-inflammatory factor ratio (37, 144, 145). Indole also has a similar function in attenuating the expression of key proteins in the NF-KB pathway, thereby inhibiting the expression of inflammatory factors while increasing the expression of antiinflammatory factors (146, 147), inhibiting the inflammatory response and promoting tumor cell growth and metastasis (148). Tryptophan reduces TNF transcription by mediating a decrease in IL-6 signaling capacity via AHR targets (149), whereas IAA neutralizes free radicals, thereby attenuating the inflammatory response of RAW264.7 macrophages to lipopolysaccharide (LPS) and increasing interleukin-8 (IL-8) signaling (150), and similarly, indole-3-methanol inhibited LPS-mediated inflammatory cytokine production in mouse bone marrow-derived macrophages (BMM) (151). Studies have shown that propionic acid and butyric acid in SCFA can inhibit HDAC and increase histone acetylation to play an anti-inflammatory and anti-cancer role (126). In HCC, gut microbial metabolites, including lipoteichoic acid (LTA) and DCA, upregulate cyclooxygenase-2 (COX-2) expression via toll-like receptor 2 (TLR2) on the membrane surface of tumor cells, which increases prostaglandin E2 (PGE2) expression. Activation of the COX2-PGE2 pathway inhibits DCs, NK and T cells, which promotes immune escape and affects the efficacy and prognosis of tumor immunotherapy. TAMs are the major type of infiltrating immune cells in the TME (152). In a study by Parida S et al. in a mouse model

of breast cancer, the number of pro-tumorigenic M2-like macrophages was significantly increased in advanced tumors of mice with gut microbiota dysbiosis compared to non-ecological dysbiosis controls with the same tumor burden (153), suggesting that gut microbes can induce macrophage polarization to either M1 or M2, thereby altering the TME and promoting or hindering the effects of tumor immunotherapy. LPS, a metabolite of E. coli, can induce M2 polarization of TAMs, thus leading to rapid cancer development, for example, in the colorectal cancer (CRC) mouse model, with the increase of E. coli, its metabolite LPS can upregulate the secretion of cathepsin K by CRC cells, which can induce M2 polarization of TAMs, leading to rapid CRC development (154), while a high-fat diet can induce gut microbiota dysbiosis, reduce SCFA levels, activate the MCP-1/C-C motif chemokine receptor 2 (CCR2) axis, and promote TAMs recruitment and polarization towards M2, ultimately leading to CRC development (155).

5.2 Regulation of adaptive immune responses

Numerous studies have shown that the gut microbial tryptophan metabolite indole and indole derivatives can regulate Treg/T helper cell 17 (Th17) differentiation and thereby suppress the inflammatory response (156). For example, indole-3-carbinol (I3C) treatment reduces colorectal cancer by decreasing Th17 cells and increasing Treg (109). In addition, I3C increased the production of CD4⁺Foxp3⁺ cells and ultimately reduced CD4⁺IL-17⁺ cells that induce neuroinflammation in mice, and IAA and IPA promote Treg differentiation and function by upregulating the expression of Foxp3 (Forkhead Box P3) and other Treg-related genes (37, 41, 157). IAA promotes Treg differentiation by activating the AHR pathway (37). In addition, tryptamine has been shown to activate the mechanistic target of rapamycin (mTOR) in Treg cells in vitro, to increase the expression of phospho-eukaryotic translation initiation factor 4E binding protein 1 (P4EBP1), and to increase the expression of phosphorylated ribosomal protein S6 kinase 1 (P-S6K1) in Tem cells (158), suggesting that exogenous tryptophan promotes cytotoxic T cell glycolysis, thereby affecting the regulation of CD4⁺ T cell function (159). In addition to the effects of IPA on T lymphocytes, indole derivatives have been shown to inhibit and stimulate B cells. I3C can regulate B cell function by inhibiting the production of the immunoglobulins immunoglobulin M (IgM) and immunoglobulin G (IgG) and decreasing the expression of the cell surface antigen CD69. CD69, inducing B cell apoptosis and inhibiting B cell proliferation (160, 161). Tryptamine has been shown to stimulate IgA production by B cells and activate the transcription factor AHR to regulate B cell function (162). Although several studies have demonstrated the inhibitory and stimulatory effects of indole derivatives on B cells, further research into the mechanism of action of these compounds is needed before they can be used in the potential treatment of immune diseases.

Gut microbial bile metabolites also play an important role in T cell expression. DAC has been shown to inhibit the Ca²⁺-nuclear factor of activated T cells-2 (NFAT2) signaling pathway by targeting the plasma membrane Ca²⁺ATPase (PMCA), and in CRC patients, the effective function of CD8⁺ T cells is negatively correlated with

DCA concentration and expression (163). Another study showed that the tryptophan metabolite indole activates the AHR and also promotes the conversion of TAMs to an immunosuppressive phenotype in pancreatic ductal adenocarcinoma and inhibits the accumulation of $CD8^+$ T cells in the tumor (9). Researchers have found that selective depletion of TAMs in a mouse model of breast cancer increases $CD8^+$ T cell infiltration into the TME and stimulates anti-tumor immune responses (164, 165).

Specific types of gut metabolites, including SCFA, tryptophan, etc., show a compelling role in cancer inhibition. This is because these compounds show significant activity on immune signaling and cell division processes (95). SCFA recognizes specific G protein-coupled receptors (GPCRs) on the surface of immune cells, including GPR41 and GPR43, which leads to an increase in the total number of regulatory T cells, transforming growth factor- β 1 (TGF β 1) and levels of the anti-inflammatory cytokine IL-10 in the host (166). An inhibitory effect on T-cell mediated autoimmune responses was also found with diets high in SCFA. In contrast, another study found that *Lactobacillus* metabolizes dietary tryptophan to indole, which activates the AHR and inhibits the accumulation of CD8⁺ T cells in tumors. In human CRC, the gut microbiota stimulates tumor cells to produce chemokines to recruit anti-tumor T cells into tumor tissue and play an immune role (167).

Indole derivatives can regulate T cell differentiation by modulating the expression of cytokines and transcription factors. For example, the indole derivative I3C has been shown to promote Th1 cell differentiation by upregulating interferon gamma and $T\beta$ transcription factors (161). In addition, I3C inhibits T helper 2 cell (Th2) differentiation by downregulating the expression of IL-4 related genes and the transcription factor GATA binding protein 3 (GATA3) (161). Another indole derivative, Indoxyl sulfate (IS), has also shown the ability to regulate T cell activation and proliferation. IS inhibits the expression of CD25 and CD69 surface markers (162), which are key indicators of early T cell activation (168, 169). In addition, IS reduced the frequency of IL-4-producing CD4⁺ T cells and inhibited Th2 differentiation. This effect was attributed to inhibition of phosphorylation of signal transducer and activator of transcription 5 (STAT5) and signal transducer and activator of transcription 6 (STAT6), transcription factors involved in Th2 differentiation. Indoxyl 3-sulfate (I3S) can also induce T cell apoptosis or programmed cell death by upregulating the expression of proapoptotic proteins (170). Other indole derivatives IAld and IPA have also been shown to modulate the immune response by regulating T cell function (171, 172). IAld induces T-cell apoptosis and inhibits T-cell activation by regulating the expression of pro- and anti-apoptotic proteins (173) (Figure 2).

6 Potential application of natural products to microbial metabolites in tumor immunotherapy

Natural products have unique advantages such as a wide range of sources, low side effects and diverse biological activities. Natural products not only have broad anti-tumor activity (174). They also



have the ability to modulate complex host-microbe interactions. Furthermore, an increasing amount of evidence indicates that natural products can influence the immune microenvironment of tumors by regulating microbial metabolites. Natural products can also target the microbiome to improve the efficacy of chemotherapy, overcome drug resistance and provide new therapeutic ideas and strategies for tumor immunotherapy (Table 1).

6.1 Alkaloids

In recent years, alkaloid chemicals extracted from plants have been found to have good anti-tumor activity, high efficacy, good tolerability and few toxic side effects (175). Berberine (BBR) is an isoquinoline alkaloid extracted from the roots and bark of plants in the *Coptis chinensis* Franch. Dextran sulfate sodium (DSS) can break down the intestinal mucosal barrier, leading to intestinal inflammation and ulcers. Previous studies have shown that BBR can ameliorate the bile acid imbalance induced by DSS in both the liver and the intestine. It achieves this by restoring the perturbed gut microbiota and activating the FXR and TGR5 signaling pathways, thereby alleviating DSSinduced ulcerative colitis in mice (176). Prolonged ulcerative colitis causes high levels of pro-inflammatory cytokines to accumulate in the colonic mucosa, and this accumulation, which in turn leads to proliferative lesions, is considered to be a major risk factor for the development of colorectal cancer (177). BBR was further investigated by Wang et al. and found to be therapeutically useful for potential protection against oxidative azomethane (AOM)/DSS-induced colitis and tumors in mice. Transcriptome analysis revealed that BBR regulates colonic epithelial cell signaling pathways in colitisassociated colon cancer mice through tryptophan metabolism and Wnt signaling pathways, which may affect fecal metabolites and SCFA metabolism (178). In addition, BBR also synergized with exercise to slow the progression of breast cancer in 4T1 hormone mice, and this synergistic effect was able to enhance the body's immune function, significantly increase levels of SCFAs, and activate the mitochondrial apoptotic pathway as well as the Fas death receptor apoptotic pathway, resulting in an anti-cancer effect (179). Furthermore, it has also been shown that Matrine increases SCFA levels in the gut and prevents intestinal damage through the Toll-like receptor 4 (TLR4)/NF-KB/ mitogen-activated protein kinase (MAPK) signaling pathway, demonstrating its great potential as a potential therapeutic agent for the treatment of cancer (180).

6.2 Glycosides

Glycosides, as a class of natural products widely found in plants, have a variety of biological activities, mainly through the interaction with intestinal flora and metabolites to exert a medicinal effect,

TABLE 1 Mechanisms of tumor therapy by natural products and their active ingredients via gut microbial metabolites.

Natural product	Active ingredient	Microbial metabolites	Mechanism	References
Coptis chinensis	Berberine	SCFAs	Inhibits intestinal inflammatory response and modulates colon epithelial cell signaling pathways in CAC mice via tryptophan metabolism and Wnt signaling pathway. Regulates intestinal microbiota imbalance, improves intestinal mucosal barrier function. Increases NK cell infiltration, increases levels of SCFAs, and activates mitochondrial apoptosis pathway as well as Fas death receptor apoptosis pathway for breast cancer.	(178) (179)
Sophora flavescens	Matrine	SCFAs	Increased SCFA levels in the intestine via the TLR4/NF- κ B/MAPK signaling pathway.	(180)
Pulsatilla chinensis	Pulsatilla chinensis saponin	SCFAs	Increased levels of colonic SCFAs, activation of the GPR43-NLRP3 signaling pathway, and decreased levels of IL-1 β , IL-6, and TNF- α pro-inflammatory cytokines.	(182)
Panax ginseng	Ginseng glucosyl oleanolate	SCFAs	Regulation of the MAPK signaling pathway inhibits Hep G2 cell proliferation, rebalances the intestinal flora, increases the level of total SCFAs improves intestinal inflammation and inhibits tumor growth.	(183)
Gynostemma pentaphyllum	Jiaogulan saponins	SCFAs	Improvement of the inflamed intestinal barrier, polarization of colonic M1 macrophages to M2 macrophages, positive restoration of the E-calmodulin/N-calmodulin ratio and down-regulation of oncogenic signaling molecules, and promotion of SCFAs-producing bacteria in a time-dependent manner.	(186)
Brown algae	Fucoidan	SCFAs	Regulate tryptophan metabolism and SCFAs to enhance CD8 ⁺ T cell function, increase the production of IFN- γ and TNF α , reduce the inhibitory effect of Treg in the circulatory system, remodel the intestinal microbiota, and cooperate with PD-1 monoclonal antibody to exert anti-tumor effects.	(187)
Cichorium glandulosum	Inulin	SCFAs	Regulation of intestinal flora, production of SCFAs, and inhibition of EMT transition processes.	(189)
Tetrastigma hemsleyanum	Tetrastigma hemsleyanum polysaccharide	SCFAs	Increased ileal secretion of immunoglobulin A and SCFAs restored tumor- induced intestinal microflora disturbances and promoted macrophage phagocytic index, NK cell activation.	(191)
Tremella fuciformis	Tremella fuciformis polysaccharides	Tryptophan metabolites (xanthurenic acid and kynurenic acid), Bile acids (DCA)	Stimulation of Foxp 3 ⁺ T cells promoted the production of anti-inflammatory cytokines, significantly increased intestinal flora diversity, and modulated tyrosine biosynthesis, tryptophan metabolism, and bile acid metabolism pathways.	(192)
Kaempferia galanga	Kaempferol	Bile acids (chenodesoxycholic acid)	Reducing tumor load, restoring the damaged intestinal barrier, down- regulating the secondary bile acid synthesis pathway, and increasing the protein expression of FXR to inhibit the activation of the Wnt/ β -catenin pathway lead to the inhibition of CRC.	(194)
Dioscorea esculenta	Sitosterols	SCFAs	Decreases PI 3K/Akt expression, promotes Bad activation, decreases Bcl-xl, and increases cyto-c release, leading to caspase-9 and caspase-3 activation, PARP cleavage, and apoptosis. Increased levels of SCFAs led to apoptosis of cancer cells <i>in vitro</i> .	(195)
Houpoea officinalis	Magnolol	Tryptophan metabolites (kynurenic acid, 5- hydroxyindoleacetic acid, IAA, indolelactic acid and indoxylsulfuric acid)	Anti-inflammatory, restores tryptophan metabolites and enhances AHR activation inhibited by colonic inflammation thereby inhibiting colonic inflammation.	(196)
Vaccinium macrocarpon	Proanthocyanidins	Bile acids (chenodesoxycholic acid)	Targeting the gut microbiome-esophageal metabolome axis, modulating the TLR/NF- κ B/TP 53 signaling pathway to inhibit EAC, and specifically reversing reflux-induced bacterial, inflammatory, and immune-related proteins and genes.	(198)

especially in the antitumor and immunomodulatory aspects showing significant potential (181). Furthermore, Pulsatilla chinensis saponin (PRS) is a natural saponin analogue isolated from *Pulsatilla chinensis* (Bunge) Regel. PRS protects against DSSinduced inflammatory bowel disease by increasing the levels of SCFAs, activating the GPR 43-NOD-like receptor thermal protein domain associated protein 3 (NLRP 3) signaling pathway and decreasing the levels of pro-inflammatory factors IL-1 β , IL-6 and TNF- α *in vivo* (182). Among these, *Panax ginseng* C. A. Mey. has been widely used as a functional food and medicine. Ginseng glucosyl oleanolate (GGO) was enzymatically prepared from ginsenoside Ro. GGO inhibited the proliferation of Hep G2 cells

through phosphorylation of the MAPK signaling pathway, rebalanced the intestinal flora, increased the concentration of total SCFA, and increased the levels of acetic and propionic acids in the colon, thereby delaying the progression of liver tumors (183). Jiaogulan saponins are extracted from the dried above-ground part of Cucurbitaceae gynostemma, with hypoglycemic and anti-tumor effects (184, 185). *Imran Khan* et al. found that the combination of jiaogulan saponin and *Ganoderma lucidum* polysaccharides to improve the inflamed intestinal barrier in Apc^{Min/+} mice polarized colonic M1 macrophages to M2 macrophages, positively restored the E-calmodulin/N-calmodulin ratio and downregulated oncogenic signaling molecules, and increased the production of SCFAs in bacteria in a time-dependent manner (186).

6.3 Polysaccharides

Polysaccharides are macromolecular compounds widely found in plants, fungi, marine organisms and other natural sources, usually consisting of monosaccharides or oligosaccharides linked by glycosidic bonds, with diverse structures and rich biological activities. Polysaccharides can influence the tumor immune microenvironment by regulating the metabolic activity of the gut microbiota and increasing the synthesis of microbial metabolites.

Fucoidan, mainly derived from marine brown algae and marine invertebrates, is a sulfate -rich functional polysaccharide unique to the oceans that may enhance anti-PD-1 immunotherapy by modulating breast cancer-induced alterations in tryptophan metabolism and glycerophospholipid metabolism pathways, which are significantly increased by SCFAs, particularly acetic acid and butyric acid (187). They act as metabolic immunomodulators. They enhance the function of the T-lymphocyte immune response and anti-tumor immunity against breast cancer (188). Inulin increases the relative abundance of Bifidobacterium, Lactobacillus and Leptospira and restores the levels of acetic, propionic, isobutyric and butyric acids, thereby inhibiting the process of epithelial-mesenchymal transition (EMT) and inhibiting metastasis of colorectal cancer (189). Inulin is a natural soluble functional fructan. It has been found that inulin produces SCFA and exerts anti-tumor effects through the action of the intestinal microbiota (190). Zhou et al. evaluated the antitumor effects of Tetrastigma hemsleyanum polysaccharide (THP) using a lung tumor model. THP was able to increase the levels of secretory immunoglobulin A (SIgA) in the ileum and SCFAs in the cecum, and to improve the diversity of the intestinal microbial community, thereby restoring tumor-induced intestinal dysbiosis (191). Tremella fuciformis polysaccharides (TPs) are acidic heteropolysaccharides extracted from Tremella fuciformis. Berk. TPs exert a palliative effect on DSS-induced colitis by modulating tyrosine biosynthesis, tryptophan metabolism, and bile acid metabolism, while restoring the balance of the intestinal flora and the normal level of its microbial metabolites (192).

6.4 Flavonoids

Kaempferol is mainly derived from the ginger plant *Kaempferia* galanga L., which has received widespread attention for its

anticancer and antioxidant properties (193). Kaempferol regulates intestinal flora by significantly increasing the abundance of beneficial bacteria and inhibiting the growth of potentially pathogenic bacteria, downregulating the secondary bile acid synthesis pathway, increasing the activity of G-protein-coupled receptors, and decreasing NOD-like receptor activity (194).

6.5 Terpenoids

Sterols maintain a diverse gut microbial environment and enrich the beneficial bacterium *Lactobacillus pentosus*. Significantly increased levels of SCFAs were found in fecal samples analyzed from mice treated with sterols. In addition, sterols reduced the transactivation effects of the PI3K/Akt signaling pathway, which regulated the expression levels of several apoptosis-related proteins, ultimately inducing apoptosis in colon cancer tumor cells (195).

6.6 Phenolic compounds

Inflammation is one of the most important factors in the development of intestinal tumors, and the close relationship between inflammation and tumors has made inflammation one of the important targets for anticancer therapy. Magnolol exhibits a significant anti-inflammatory effect on DSS-induced colitis, and the mechanism may be related to the restoration of tryptophan metabolites that inhibit colonic inflammation (196). Cranberries are rich in polyphenolic compounds, one of the most important being proanthocyanidins (C-PACs). Among other things, cranberry has been shown to attenuate animal diet-induced increases in secondary bile acids and decreases in SCFAs (197), suggesting that cranberry compounds have a positive effect on the regulation of intestinal microbial metabolites. C-PAC targeted the gut microbiome-esophageal metabolome axis to inhibit esophageal adenocarcinoma (EAC) progression by increasing the abundance of the beneficial bacteria Lactococcus, Lactobacillus and Bifidobacterium. C-PAC has been shown to reverse refluxinduced microbial dysregulation, attenuate bile acid metabolism and transport, and ultimately significantly inhibit EAC via the TLR/ NF- κ B/TP 53 signaling cascade (198).

6.7 Natural product extracts

Patchouli Essential Oil (PEO), derived from Pogostemon cablin (PC), and its derivatives, patchouli alcohol (PA) and pogostone (PO), stimulate SCFA producers in $Apc^{Min/+}$ colon cancer mouse models and activate key SCFA-sensitive receptors (GPR 41, GPR 43 and GPR 109a). The gut microbiota of PEO-treated mice changed, with a significant reduction in the abundance of the *Bacteroidetes phylum* and a notable increase in the abundance of the *Firmicutes phylum* (199). In addition, PA and PO significantly promoted the growth of a probiotic, eosinophilic Ackermannia, which has anti-inflammatory effects and protects the host intestinal mucosa (200).

6.8 Traditional Chinese medicine compound prescription

Phytopharmaceuticals have shown broad application prospects in the field of tumor therapy due to their multi-target and multi-link synergistic regulatory properties. Currently, the specific mechanism of this process has become a hot topic of research regarding how botanicals mediate tumor therapy by regulating the function of the gut flora and its microbial metabolites. (Table 2).

For example, some conventional drugs such as Astragalus mongholicus Bunge.-Curcuma aromatica Salisb. (ACE), which effectively delayed tumor progression in CT26 colon cancerbearing mice, increased levels of SCFAs such as the gut microbial metabolites propionic acid and butyric acid in colon cancer cells and mediated the gut-derived stromal cell-derived factor-1 (SDF-1)/C-X-C chemokine receptor type 4 (CXCR 4) signaling pathway to inhibit tumor growth and metastasis (201). Xianlian Jiedu Decoction (XLJDD) treatment was effective in reducing the level of inflammatory response in a mouse model of colorectal cancer, the ability to reduce serum levels of inflammatory cytokines, and the ability to reduce levels of β-conjugated proteins, COX-2 and inducible nitric oxide synthase (iNOS) protein expression in colorectal tissue. Mechanistically, XLJDD improved gut dysbiosis and associated metabolic levels of SCFAs, sphingolipids and glycerophospholipids. It also increased the abundance of Enterobacteriaceae and Zeylococcus-like probiotics, as well as butyric and isovaleric acid levels (202). Shen-Bai-Jie-Du-Decoction (SBJDD) may promote the production of SCFAs by modulating the composition of intestinal flora, a process that further induces the polarization of M2-like macrophages, attenuates intestinal inflammation, restores intestinal barrier function and inhibits colorectal cancer cell proliferation (203). PRM 1201 inhibits CRC metastasis by regulating the abundance and metabolism of SCFA-producing bacteria, effectively suppressing histone deacetylation and inhibiting EMT in metastatic lesions (204). Zhu et al. used 16 S rRNA sequencing and gas chromatography-mass spectrometry (GC-MS) to detect changes in intestinal flora and fecal SCFAs after Huangqin Decoction (HQD) administration, respectively. The results showed that HQD improved intestinal flora dysbiosis, increased Clostridium abundance and fecal butyric acid levels, and inhibited the activity of PI3K/Akt pathway (205). Jianpi Yangzheng decoction (JPYZ) inhibits the formation of the pre-metastatic microenvironment of gastric cancer (GC) by remodeling the intestinal flora and increasing the production of its metabolite SCFA, which inhibits the infiltration of myeloid-derived suppressor cells (MDSC) and the production of inflammatory factors (206). Moreover, Jianpi Jiedu Decoction (JPJDF) inhibits the secretion of tryptophan metabolites, effectively reduces inflammation and significantly restores intestinal barrier function in colon cancer mice. It inhibited the expression of AhR and M2type tumor associated macrophages polarization, thereby promoting tumor immunity and inhibiting the growth of colitisassociated colorectal cancer (CAC) caused by colonic inflammation (207). Wumei Wan(WMW) can restore the balance between pathogenic and probiotic bacteria in the intestinal tract. Early administration of WMW significantly regulated serum 3-indole propionic acid levels in mice with colon cancer and prevented colon cancer by inhibiting MDSCs by decreasing the PI3K/Akt signaling pathway (208). Pien-Tze-Huang (PZH) dose-dependently inhibited colorectal tumorigenesis in AOM/DSS-treated and Apcmin/+ mice, promoted the production of beneficial metabolites such as taurine and hypotaurine, increased the levels of bile acids and unsaturated fatty acids, and significantly enhanced intestinal barrier function. Transcriptomic analysis further revealed that PZH inhibited signaling pathways such as PI3K-Akt, IL-17, TNF and cytokinechemokines, effectively blocking oncogenic and pro-inflammatory pathways, which in turn inhibited colorectal carcinogenesis (209). Bile acids are mainly synthesized in the liver and secreted into the intestine, and the conversion of primary bile acids into secondary bile acids occurs with the help of the intestinal microbiota. Secondary bile acids are also able to be reabsorbed into the liver via the enterohepatic circulation and participate in the process of bile acid resynthesis and excretion (210). Certain secondary bile acids (e.g., DCA and lithocholic acid) are thought to have procancer effects. They can alter gene expression by activating nuclear receptors, promote cell proliferation and survival, and may even induce cancer cell invasion and metastasis. Xianglian Pill (XLP) can be used for the treatment of gastrointestinal disorders of the dampheat type, and Ye et al. assessed the protective effect of XLP using AOM and DSS-induced CRC models in mice exposed to high-fat diet. XLP significantly reduced a microbial-derived metabolite of bile acids called fecal DCA by restoring ecological dysbiosis in the gut microbiota, thereby inhibiting the TLR4/Myeloid Differentiation Primary Response Protein 88 (MyD88) pathway and reducing M1 macrophage infiltration (211). Sanhuang xiexin decoction (SXD) was able to modulate the gut-liver axis immunomodulation to ameliorate DSS-induced colitis secondary to hepatic injury, and attenuate hepatic inflammation and cholestasis by improving TLR4-NF-KB and bile acid metabolic pathways (212). Treatment of feed-induced methionine- and choline-deficient diet-induced nonalcoholic steatohepatitis (NASH) mice with Qiang-Gan formula (QGE) showed that QGE treatment altered the gut microbiota of NASH mice and led to an increase in fecal lithocholic acid content (213). In addition, the ability of Siwu-Yin (SWY) to improve the intestinal flora of rats with esophageal precancerous lesions is also related to the regulation of bile acid synthesis and secretion (214) (Figure 3).

Phytopharmaceuticals have demonstrated significant efficacy and benefits in the treatment of tumors, and their actions involve a variety of complex mechanisms. However, research into the role of phytopharmaceuticals in regulating microbial metabolites is still at a preliminary stage and more in-depth studies are needed to clarify their mechanism of action and precise efficacy.

7 Challenges and perspectives

The relationship between microbial metabolites and tumor immunotherapy is complex. Several studies have shown that microorganisms and their metabolites have a non-negligible impact on tumorigenesis and development, as well as on the

TABLE 2	Mechanisms of	f traditional	Chinese I	medicine	compoundin	a for	the '	treatment	of tu	umors via	intestinal	microbial	metabolites.
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Chinese medicine compound prescription	Microbial metabolites	Mechanism	References
Astragalus mongholicus Bunge- Curcuma aromatica Salisb.	SCFAs	Increasing the content of SCFAs such as propionic acid and butyric acid in colon cancer cells, mediating the intestinal-derived SDF-1/CXCR 4 signaling pathway to repair the integrity of the intestinal barrier, decreasing the expression of Cyclin D1 and C-myc, and inhibiting tumor growth and metastasis.	(201)
Xianlian Jiedu Decoction	SCFAs	Reduced levels of inflammatory cytokines and decreased expression of β - conjugated proteins, COX-2, and iNOS proteins in colorectal tissues. Improves gut microbes and metabolic levels of associated SCFAs, sphingolipids and glycerophospholipids.	(202)
Shen-Bai-Jie-Du decoction	SCFAs	Increased production of SCFAs, activated G protein-coupled receptors, and inhibited HDAC. Decreased the proportion of M1-type macrophages, increased M2-type macrophages, down-regulated the expression levels of IL-1 β , IL-6 pro-inflammatory factors, and up-regulated the expression levels of IL-10 anti-inflammatory factors.	(203)
PRM 1201	SCFAs	Regulates the composition of intestinal flora, increases the number of SCFA-producing bacteria and SCFA production, inhibits histone deacetylation and inhibits EMT in metastatic lesions thereby inhibiting CRC metastasis.	(204)
Huangqin Decoction	SCFAs	Improvement of intestinal dysbiosis, increase of <i>Clostridium</i> abundance and fecal butyric acid level, inhibition of microbial butyric acid-mediated PI3K/Akt pathway, induction of apoptosis, attenuation of intestinal inflammation, and reduction of tumor load in CRC.	(205)
Jianpi Yangzheng decoction	SCFAs	Remodeling the structure of intestinal flora, enhancing the production of SCFAs, inhibiting the recruitment of MDSCs and reducing the production of inflammatory inhibitory factors thereby inhibiting the formation of pre-metastatic microenvironment of gastric cancer.	(206)
Jianpi Jiedu decoction	Tryptophan metabolites (indole-3- acetaldehyde, 3-methylindole, 5- hydroxyindoleacetic acid, kynurenine, 5- methoxyindoleacetate, and L-tryptophan.)	Altering the composition of the intestinal flora, increasing the abundance of beneficial intestinal bacteria, decreasing tryptophan metabolites, decreasing inflammation inhibiting the expression of AhR and M2-type tumor-associated macrophages, and enhancing anti-tumor immunity.	(207)
Wumei Wan	Tryptophan metabolites (3- indolepropionic acid)	Modulation of 3-indolepropionic acid in serum of CAC mice reduces the PI3K/Akt signaling pathway thereby inhibiting MDSCs	(208)
Pien-Tze-Huang	Bile acids (cholic acid, taurocholic acid)	Promotes the production of taurine and taurine, increases the content of bile acids and unsaturated fatty acids, and strengthens the function of the intestinal barrier. Inhibits PI 3 K-Akt, IL-17, TNF and cytokine- chemokine signaling	(209)
Xianglian Pill	Bile acids (DCA)	Reducing IL-6 and TNF- α expression and infiltration of pro- inflammatory macrophages, remodeling gut microbiota composition and bile acid metabolism to inhibit the TLR4/MyD 88 pathway, and thereby inhibiting colorectal cancer associated with a high-fat diet.	(211)
Sanhuang xiexin decoction	Bile acids	Reduced hepatocyte swelling in mice with colitis, inhibited IL-1 β and TNF- α expression, and attenuated hepatic inflammation and cholestasis by regulating TLR4 NF- κ B and bile acid metabolic pathways.	(212)
Qiang-Gan formula	Bile acids (Cholic Acid, Chenodeoxycholic Acid, DCA and LCA)	Decreased hepatic and serum bile acid concentrations, increased fecal lithobionic acid production, improved the structure of intestinal flora, promoted TGR5 expression, and inhibited NF-кB activation.	(213)
Siwu-Yin	Bile acids	Improvement of the intestinal flora in precancerous esophageal lesions and modulation of bile acid synthesis and secretion, thereby modulating macrophage polarization.	(214)

efficacy and prognostic ability of tumor immunotherapy, and that their role includes regulation of immune cells and modulation of anti-inflammatory pathways. At the same time, several studies have shown that microbial metabolites not only affect the development of gastrointestinal tumors, but also influence the TME outside the gastrointestinal tract via the gut-X axis, which in turn affects tumor immunotherapy. Therefore, many researchers have also actively explored the use of the positive aspects of microbial metabolites in conjunction with tumor immunotherapy, including probiotic supplementation therapy,



fecal microbiota transplantation methods etc., with the aim of increasing bacterial diversity, attenuating acute and long-term treatment-related toxicity, and achieving amazing results in improving immunotherapy response and prognosis, and individualizing treatment. However, we should also be aware of the increasing interest in the relationship between gut flora and anti-tumor immunotherapy, the essence of which is that the flora modulates the immune response through metabolic pathways, while the mechanisms of action between identified microbial metabolites and tumors are unclear, especially for tumors outside the gastrointestinal tract, leading to an inability to explain the differences in the number of species of gut microbial metabolites in different types of cancer. For example, studies have shown that butyrate in SCFA maintains gut integrity, preserves normal metabolic immune function and is beneficial for immunotherapeutic response, but a study showed that serum butyrate and juxtate concentrations did not significantly correlate with longer progression-free survival in French and Italian melanoma patients. Current studies of gut microbiota regulation of anti-tumor immunity have focused on the regulation of immune cells and inflammatory factors outside tumor cells, and there are no reports of gut microbes causing changes inside tumor cells to evade immune surveillance. On the other hand, most studies have focused on the relationship between the gut microbiota and cancer development, and metabolites have not been sufficiently studied; many microbial metabolites have not yet been identified. Meanwhile, the role of the gut microbiota in other immunotherapies, such as cytokine therapy and immunovaccine therapy, remains understudied. With further elucidation of the mechanism of influence of gut flora metabolites on tumor immune escape, targeted drug therapy and individualized anti-tumor therapy will be further developed, and gut flora modulation will become an effective and necessary adjuvant anti-tumor immunotherapy.

Author contributions

DL: Writing – original draft. XL: Writing – review & editing. LX: Writing – original draft. SZ: Writing – original draft. HL: Writing – original draft. XZ: Writing – original draft. WY: Writing – original draft. YY: Writing – review & editing. XF: Writing – review & editing.

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References

1. Topalian SL, Drake CG, Pardoll DM. Immune checkpoint blockade: a common denominator approach to cancer therapy. *Cancer Cell*. (2015) 27:450–61. doi: 10.1016/j.ccell.2015.03.001

2. Rooks MG, Garrett WS. Gut microbiota metabolites and host immunity. Nature reviews. *Immunology*. (2016) 16:341–52.

3. Zhou L, Wei X. Ocular immune-related adverse events associated with immune checkpoint inhibitors in lung cancer. *Front Immunol.* (2021) 12:701951. doi: 10.3389/fimmu.2021.701951

4. Hanahan D. Hallmarks of cancer: new dimensions. . *Cancer Discovery*. (2022) 12:31-46. doi: 10.1158/2159-8290.CD-21-1059

5. Arthur JC, Perez-Chanona E, Mühlbauer M, Tomkovich S, Uronis JM, Fan TJ, et al. Intestinal inflammation targets cancer-inducing activity of the microbiota. *Science (New York N.Y.).* (6103) 2012 338:120–3.

6. Duan H, Wang L, Huangfu M, Li H. The impact of microbiota-derived shortchain fatty acids on macrophage activities in disease: Mechanisms and therapeutic potentials. *Biomedicine pharmacotherapy = Biomedecine pharmacotherapie*. (2023) 165:115276. doi: 10.1016/j.biopha.2023.115276

7. Levy M, Thaiss CA, Elinav E. Metabolites: messengers between the microbiota and the immune system. *Genes Dev.* (2016) 30:1589-97. doi: 10.1101/gad.284091.116

8. Sipos A, Ujlaki G, Mikó E, Maka E, Szabó J, Uray K, et al. The role of the microbiome in ovarian cancer: mechanistic insights into oncobiosis and to bacterial metabolite signaling. *Mol medicine(Cambridge Mass.)*. (2021) 27:33.

9. Hezaveh K, Shinde RS, Klötgen A, Halaby MJ, Lamorte S, Ciudad MT, et al. Tryptophan-derived microbial metabolites activate the aryl hydrocarbon receptor in tumor-associated macrophages to suppress anti-tumor immunity. *Immunity*. (2022) 55:324–340.e8. doi: 10.1016/j.immuni.2022.01.006

10. Kiss B, Mikó E, Sebő É, Toth J, Ujlaki G, Szabó J, et al. Oncobiosis and microbial metabolite signaling in pancreatic adenocarcinoma. *Cancers*. (2020) 12. doi: 10.3390/ cancers12051068

11. Bhagat TD, Von Ahrens D, Dawlaty M, Zou Y, Baddour J, Achreja A, et al. Lactate-mediated epigenetic reprogramming regulates formation of human pancreatic cancer-associated fibroblasts. *eLife*. (2019) 8. doi: 10.7554/eLife.50663

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12. Mullins TD, Kern HF, Metzgar RS. Ultrastructural differentiation of sodium butyrate-treated human pancreatic adenocarcinoma cell lines. *Pancreas*. (1991) 6:578–87. doi: 10.1097/00006676-199109000-00012

13. Chen AN, Luo Y, Yang YH, Fu JT, Geng XM, Shi JP, et al. Lactylation a novel metabolic reprogramming code: current status and prospects. *Front Immunol.* (2021) 12:688910. doi: 10.3389/fimmu.2021.688910

14. Rees DO, Crick PJ, Jenkins GJ, Wang Y, Griffiths WJ, Brown TH, et al. Comparison of the composition of bile acids in bile of patients with adenocarcinoma of the pancreas and benign disease. *J Steroid Biochem Mol Biol.* (2017) 174:290–5. doi: 10.1016/j.jsbmb.2017.10.011

15. Plewa S, Horała A, Dereziński P, Klupczynska A, Nowak-Markwitz E, Matysiak J, et al. Usefulness of amino acid profiling in ovarian cancer screening with special emphasis on their role in cancerogenesis. *Int J Mol Sci.* (2017) 18. doi: 10.3390/ ijms18122727

16. Hilvo M, de Santiago I, Gopalacharyulu P, Schmitt WD, Budczies J, Kuhberg M, et al. Accumulated metabolites of hydroxybutyric acid serve as diagnostic and prognostic biomarkers of ovarian high-grade serous carcinomas. *Cancer Res.* (2016) 76:796–804. doi: 10.1158/0008-5472.CAN-15-2298

17. Zhou M, Guan W, Walker LD, Mezencev R, Benigno BB, Gray A, et al. Rapid mass spectrometric metabolic profiling of blood sera detects ovarian cancer with high accuracy. *Cancer Epidemiol Biomarkers Prev Publ Am Assoc Cancer Res cosponsored by Am Soc Prev Oncol.* (2010) 19:2262–71. doi: 10.1158/1055-9965.EPI-10-0126

18. Ke C, Hou Y, Zhang H, Fan L, Ge T, Guo B, et al. Large-scale profiling of metabolic dysregulation in ovarian cancer. *Int J Cancer.* (2015) 136:516–26. doi: 10.1002/ijc.v136.3

19. Yang Q, Wang B, Zheng Q, Li H, Meng X, Zhou F, et al. A review of gut microbiota-derived metabolites in tumor progression and cancer therapy. *Advanced science(Weinheim Baden-Wurttemberg Germany).* (2023) 10:e2207366. doi: 10.1002/advs.202207366

20. Aghamajidi A, Maleki Vareki S. The effect of the gut microbiota on systemic and anti-tumor immunity and response to systemic therapy against cancer. *Cancers*. (2022) 14. doi: 10.3390/cancers14153563

21. Li Y, Jiang H, Wang X, Liu X, Huang Y, Wang Z, et al. Crosstalk between the gut and brain: importance of the fecal microbiota in patient with brain tumors. *Front Cell infection Microbiol.* (2022) 12:881071. doi: 10.3389/fcimb.2022.881071

22. Tremaroli V, Bäckhed F. Functional interactions between the gut microbiota and host metabolism. *Nature*. (7415) 2012 489:242–9.

23. González-Sánchez P, DeNicola GM. The microbiome(s) and cancer: know thy neighbor(s). J Pathol. (2021) 254:332-43. doi: 10.1002/path.v254.4

24. Oliphant K, Allen-Vercoe E. Macronutrient metabolism by the human gut microbiome: major fermentation by-products and their impact on host health. *Microbiome.* (2019) 7:91. doi: 10.1186/s40168-019-0704-8

25. den Besten G, Lange K, Havinga R, van Dijk TH, Gerding A, van Eunen K, et al. Gut-derived short-chain fatty acids are vividly assimilated into host carbohydrates and lipids. American journal of physiology. *Gastrointestinal liver Physiol.* (2013) 305: G900–10. doi: 10.1152/ajpgi.00265.2013

26. Wang G, Huang S, Wang Y, Cai S, Yu H, Liu H, et al. Bridging intestinal immunity and gut microbiota by metabolites. *Cell Mol Life Sci CMLS*. (2019) 76:3917–37. doi: 10.1007/s00018-019-03190-6

27. Wong CC, Yu J. Gut microbiota in colorectal cancer development and therapy. Nature reviews. *Clin Oncol.* (2023) 20:429–52.

28. Cummings JH, Pomare EW, Branch WJ, Naylor CP, Macfarlane GT. Short chain fatty acids in human large intestine portal hepatic and venous blood. *Gut.* (1987) 28:1221–7. doi: 10.1136/gut.28.10.1221

29. Topping DL, Clifton PM. Short-chain fatty acids and human colonic function: roles of resistant starch and nonstarch polysaccharides. *Physiol Rev.* (2001) 81:1031–64. doi: 10.1152/physrev.2001.81.3.1031

30. Kelly CJ, Zheng L, Campbell EL, Saeedi B, Scholz CC, Bayless AJ, et al. Crosstalk between microbiota-derived short-chain fatty acids and intestinal epithelial HIF augments tissue barrier function. *Cell Host Microbe*. (2015) 17:662–71. doi: 10.1016/j.chom.2015.03.005

31. Lewis K, Lutgendorff F, Phan V, Söderholm JD, Sherman PM, McKay DM. Enhanced translocation of bacteria across metabolically stressed epithelia is reduced by butyrate. *Inflammatory bowel Dis.* (2010) 16:1138–48. doi: 10.1002/ibd.21177

32. Singh N, Gurav A, Sivaprakasam S, Brady E, Padia R, Shi H, et al. Activation of Gpr109a receptor for niacin and the commensal metabolite butyrate suppresses colonic inflammation and carcinogenesis. *Immunity*. (2014) 40:128–39. doi: 10.1016/j.immuni.2013.12.007

33. Ben Q, Sun Y, Chai R, Qian A, Xu B, Yuan Y. Dietary fiber intake reduces risk for colorectal adenoma: a meta-analysis. *Gastroenterology*. (2014) 146:689–699.e6. doi: 10.1053/j.gastro.2013.11.003

34. Sipe LM, Chaib M, Pingili AK, Pierre JF, Makowski L. Microbiome bile acids and obesity: How microbially modified metabolites shape anti-tumor immunity. *Immunol Rev.* (2020) 295:220–39. doi: 10.1111/imr.v295.1

35. Shen J, Yang L, You K, Chen T, Su Z, Cui Z, et al. Indole-3-acetic acid alters intestinal microbiota and alleviates ankylosing spondylitis in mice. *Front Immunol.* (2022) 13:762580. doi: 10.3389/fimmu.2022.762580

36. Xue C, Li G, Zheng Q, Gu X, Shi Q, Su Y, et al. Tryptophan metabolism in health and disease. *Cell Metab.* (2023) 35:1304–26. doi: 10.1016/j.cmet.2023.06.004

37. Sun M, Ma N, He T, Johnston LJ, Ma X. Tryptophan(Trp) modulates gut homeostasis via aryl hydrocarbon receptor(AhR). Crit Rev Food Sci Nutr. (2020) 60:1760-8. doi: 10.1080/10408398.2019.1598334

38. Wei GZ, Martin KA, Xing PY, Agrawal R, Whiley L, Wood TK, et al. Tryptophan-metabolizing gut microbes regulate adult neurogenesis via the aryl hydrocarbon receptor. *Proc Natl Acad Sci United States America*. (2021) 118. doi: 10.1073/pnas.2021091118

39. Bhattarai Y, Jie S, Linden DR, Ghatak S, Mars RAT, Williams BB, et al. Bacterially derived tryptamine increases mucus release by activating a host receptor in a mouse model of inflammatory bowel disease. *iScience*. (2020) 23:101798. doi: 10.1016/j.isci.2020.101798

40. Cho CE, Taesuwan S, Malysheva OV, Bender E, Tulchinsky NF, Yan J, et al. Trimethylamine-N-oxide(TMAO) response to animal source foods varies among healthy young men and is influenced by their gut microbiota composition: A randomized controlled trial. *Mol Nutr Food Res.* (2017) 61. doi: 10.1002/mnfr.201600324

41. Roager HM, Licht TR. Microbial tryptophan catabolites in health and disease. *Nat Commun.* (2018) 9:3294. doi: 10.1038/s41467-018-05470-4

42. Sanchez-Azofra A, Gu W, Masso-Silva JA, Sanz-Rubio D, Marin-Oto M, Cubero P, et al. Inflammation biomarkers in OSA chronic obstructive pulmonary disease and chronic obstructive pulmonary disease/OSA overlap syndrome. *J Clin sleep Med JCSM Off Publ Am Acad Sleep Med*. (2023) 19:1447–56. doi: 10.5664/jcsm.10600

43. Fan Y, Pedersen O. Gut microbiota in human metabolic health and disease. Nature reviews. *Microbiology*. (2021) 19:55–71.

44. Zhang Z, Zhang H, Chen T, Shi L, Wang D, Tang D. Regulatory role of shortchain fatty acids in inflammatory bowel disease. *Cell communication Signaling CCS*. (2022) 20:64. doi: 10.1186/s12964-022-00869-5

45. Wilson RAM, Evans TRJ, Fraser AR, Nibbs RJB. Immune checkpoint inhibitors: new strategies to checkmate cancer. *Clin Exp Immunol.* (2018) 191:133–48. doi: 10.1111/cei.13081

46. Igney FH, Krammer PH. Immune escape of tumors: apoptosis resistance and tumor counterattack. J leukocyte Biol. (2002) 71:907–20. doi: 10.1189/jlb.71.6.907

47. Vinay DS, Ryan EP, Pawelec G, Talib WH, Stagg J, Elkord E, et al. Immune evasion in cancer: Mechanistic basis and therapeutic strategies. *Semin Cancer Biol.* (2015) 35 Suppl S185-s198. doi: 10.1016/j.semcancer.2015.03.004

48. Bouzin C, Brouet A, De Vriese J, Dewever J, Feron O. Effects of vascular endothelial growth factor on the lymphocyte-endothelium interactions: identification of caveolin-1 and nitric oxide as control points of endothelial cell anergy. *J immunology* (*Baltimore Md. 1950*). (2007) 178:1505–11. doi: 10.4049/jimmunol.178.3.1505

49. Oleinika K, Nibbs RJ, Graham GJ, Fraser AR. Suppression subversion and escape: the role of regulatory T cells in cancer progression. *Clin Exp Immunol.* (2013) 171:36-45. doi: 10.1111/j.1365-2249.2012.04657.x

50. Krummel MF, Allison JP. CD28 and CTLA-4 have opposing effects on the response of T cells to stimulation. J Exp Med. (1995) 182:459–65. doi: 10.1084/ jem.182.2.459

51. Alegre ML, Frauwirth KA. T-cell regulation by CD28 and CTLA-4. Nat Rev Immunol. (2001) 1:220–8. doi: 10.1038/35105024

52. Leach DR, Krummel MF, Allison JP. Enhancement of antitumor immunity by CTLA-4 blockade. *Science(New York N.Y.).* (5256) 1996 271:1734–6.

53. Dagar G, Gupta A, Masoodi T, Nisar S, Merhi M, Hashem S, et al. Harnessing the potential of CAR-T cell therapy: progress challenges and future directions in hematological and solid tumor treatments. *J Trans Med.* (2023) 21:449. doi: 10.1186/s12967-023-04292-3

54. Cheng W, Kang K, Zhao A, Wu Y. Dual blockade immunotherapy targeting PD-1/PD-L1 and CTLA-4 in lung cancer. *J Hematol Oncol.* (2024) 17:54. doi: 10.1186/ s13045-024-01581-2

55. Ishida Y, Agata Y, Shibahara K, Honjo T. Induced expression of PD-1 a novel member of the immunoglobulin gene superfamily upon programmed cell death. *EMBO J.* (1992) 11:3887–95. doi: 10.1002/j.1460-2075.1992.tb05481.x

 Ortega MA, Boaru DL, De-Leon-Oliva D, Fraile-Martinez O, García-Montero C, Rios L, et al. PD-1/PD-L1 axis: implications in immune regulation cancer progression and translational applications. *J Mol medicine(Berlin Germany)*. (2024) 102:987–1000. doi: 10.1007/s00109-024-02463-3

57. Keir ME, Butte MJ, Freeman GJ, Sharpe AH. PD-1 and its ligands in tolerance and immunity. *Annu Rev Immunol*. (2008) 26:677-704. doi: 10.1146/ annurev.immunol.26.021607.090331

58. Latchman Y, Wood CR, Chernova T, Chaudhary D, Borde M, Chernova I, et al. PD-L2 is a second ligand for PD-1 and inhibits T cell activation. *Nat Immunol.* (2001) 2:261–8. doi: 10.1038/85330

59. Hui E, Cheung J, Zhu J, Su X, Taylor MJ, Wallweber HA, et al. T cell costimulatory receptor CD28 is a primary target for PD-1-mediated inhibition. *Science(New York N.Y.).* (6332) 2017 355:1428-33.

60. Thompson RH, Gillett MD, Cheville JC, Lohse CM, Dong H, Webster WS, et al. Costimulatory B7-H1 in renal cell carcinoma patients: Indicator of tumor aggressiveness and potential therapeutic target. *Proc Natl Acad Sci United States America.* (2004) 101:17174–9. doi: 10.1073/pnas.0406351101

61. Curiel TJ, Wei S, Dong H, Alvarez X, Cheng P, Mottram P, et al. Blockade of B7-H1 improves myeloid dendritic cell-mediated antitumor immunity. *Nat Med.* (2003) 9:562–7. doi: 10.1038/nm863

62. Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, et al. Improved survival with ipilimumab in patients with metastatic melanoma. *New Engl J Med.* (2010) 363:711–23. doi: 10.1056/NEJMoa1003466

63. Inoue Y, Inui N. Associations between immune-related adverse events and prognosis in cancer patients receiving immune checkpoint inhibitor therapy. *Internal medicine(Tokyo Japan)*. (2024). doi: 10.2169/internalmedicine.4654-24

 Shimoyama K, Nakajima A, Minari Y. The first report of bickerstaff brainstem encephalitis induced by atezolizumab for metastatic breast cancer. *Acta Med Okayama*. (2024) 78:407–12.

65. Dana H, Chalbatani GM, Jalali SA, Mirzaei HR, Grupp SA, Suarez ER, et al. CAR-T cells: Early successes in blood cancer and challenges in solid tumors. A. *cta Pharm Sinica. B.* (2021) 11:1129–47.

66. Bird RE, Hardman KD, Jacobson JW, Johnson S, Kaufman BM, Lee SM, et al. Single-chain antigen-binding proteins. *Science(New York N.Y.).* (4877) 1988 242:423-6.

67. Zmievskaya E, Valiullina A, Ganeeva I, Petukhov A, Rizvanov A, Bulatov E. Application of CAR-T cell therapy beyond oncology: autoimmune diseases and viral infections. *Biomedicines*. (2021) 9. doi: 10.3390/biomedicines9010059

68. Huston JS, Levinson D, Mudgett-Hunter M, Tai MS, Novotný J, Margolies MN, et al. Protein engineering of antibody binding sites: recovery of specific activity in an anti-digoxin single-chain Fv analogue produced in Escherichia coli. *Proc Natl Acad Sci United States America*. (1988) 85:5879–83. doi: 10.1073/pnas.85.16.5879

69. Gupta A, Dagar G, Rehmani MU, Prasad CP, Saini D, Singh M, et al. CAR T-cell therapy in cancer: Integrating nursing perspectives for enhanced patient care. *Asia-Pacific J Oncol Nurs.* (2024) 11:100579. doi: 10.1016/j.apjon.2024.100579

70. Ferment B, Arnulf B. CAR-T cells immunotherapy in multiple myeloma: Present and future. *Bull du Cancer*. (2021) 108:S65–s72.

71. Locke FL, Neelapu SS, Bartlett NL, Lekakis LJ, Jacobson CA, Braunschweig I, et al. Tocilizumab prophylaxis following axicabtagene ciloleucel in relapsed or refractory large B-cell lymphoma. *Transplant Cell Ther.* (2024) 30:1065–79. doi: 10.1016/j.jtct.2024.08.018

72. Neelapu SS, Chavez JC, Sehgal AR, Epperla N, Ulrickson M, Bachy E, et al. Three-year follow-up analysis of axicabtagene ciloleucel in relapsed/refractory indolent non-Hodgkin lymphoma(ZUMA-5). *Blood.* (2024) 143:496–506. doi: 10.1182/ blood.2023021243

73. Wang M, Siddiqi T, Gordon LI, Kamdar M, Lunning M, Hirayama AV, et al. Lisocabtagene maraleucel in relapsed/refractory mantle cell lymphoma: primary analysis of the mantle cell lymphoma cohort from TRANSCEND NHL 001 a phase I multicenter seamless design study. *J Clin Oncol Off J Am Soc Clin Oncol.* (2024) 42:1146–57. doi: 10.1200/JCO.23.02214

74. Cohen AD. CAR T-cell therapy against B-cell maturation antigen in multiple myeloma. *Clin Adv Hematol Oncol H&O*. (2018) 16:804-6.

75. Majzner RG, Mackall CL. Tumor antigen escape from CAR T-cell therapy. Cancer Discovery. (2018) 8:1219–26. doi: 10.1158/2159-8290.CD-18-0442

76. Maude SL, Teachey DT, Porter DL, Grupp SA. CD19-targeted chimeric antigen receptor T-cell therapy for acute lymphoblastic leukemia. *Blood.* (2015) 125:4017–23. doi: 10.1182/blood-2014-12-580068

77. Kim JM, Chen DS. Immune escape to PD-L1/PD-1 blockade: seven steps to success(or failure). Ann Oncol Off J Eur Soc Med Oncol. (2016) 27:1492–504. doi: 10.1093/annonc/mdw217

78. Hollingsworth RE, Jansen K. Turning the corner on therapeutic cancer vaccines. *NPJ Vaccines.* (2019) 4:7. doi: 10.1038/s41541-019-0103-y

79. Kantoff PW, Higano CS, Shore ND, Berger ER, Small EJ, Penson DF, et al. Sipuleucel-T immunotherapy for castration-resistant prostate cancer. *New Engl J Med.* (2010) 363:411–22. doi: 10.1056/NEJMoa1001294

80. Tiwari A, Alcover K, Carpenter E, Thomas K, Krum J, Nissen A, et al. Utility of cell-based vaccines as cancer therapy: Systematic review and meta-analysis. *Hum Vaccines immunotherapeutics*. (2024) 20:2323256. doi: 10.1080/21645515.2024.2323256

81. Xi HB, Wang GX, Fu B, Liu WP, Li Y. Survivin and PSMA loaded dendritic cell vaccine for the treatment of prostate cancer. *Biol Pharm Bull.* (2015) 38:827–35. doi: 10.1248/bpb.b14-00518

82. Han CL, Yan YC, Yan LJ, Meng GX, Yang CC, Liu H, et al. Efficacy and security of tumor vaccines for hepatocellular carcinoma: a systemic review and meta-analysis of the last 2 decades. *J Cancer Res Clin Oncol.* (2023) 149:1425–41. doi: 10.1007/s00432-022-04008-y

83. Yamanaka R, Homma J, Yajima N, Tsuchiya N, Sano M, Kobayashi T, et al. Clinical evaluation of dendritic cell vaccination for patients with recurrent glioma: results of a clinical phase I/II trial. *Clin Cancer Res an Off J Am Assoc Cancer Res.* (2005) 11:4160–7. doi: 10.1158/1078-0432.CCR-05-0120

84. Agus A, Clément K, Sokol H. Gut microbiota-derived metabolites as central regulators in metabolic disorders. *Gut.* (2021) 70:1174–82. doi: 10.1136/gutjnl-2020-323071

85. Chen L, Zhernakova DV, Kurilshikov A, Andreu-Sánchez S, Wang D, Augustijn HE, et al. Influence of the microbiome diet and genetics on inter-individual variation in the human plasma metabolome. *Nat Med.* (2022) 28:2333–43. doi: 10.1038/s41591-022-02014-8

86. Burcelin R, Serino M, Chabo C, Garidou L, Pomié C, Courtney M, et al. Metagenome and metabolism: the tissue microbiota hypothesis. *Diabetes Obes Metab.* (2013) 15 Suppl 3:61–70.

87. Wikoff WR, Anfora AT, Liu J, Schultz PG, Lesley SA, Peters EC, et al. Metabolomics analysis reveals large effects of gut microflora on mammalian blood metabolites. *Proc Natl Acad Sci United States America*. (2009) 106:3698–703. doi: 10.1073/pnas.0812874106

88. Dumas ME. The microbial-mammalian metabolic axis: beyond simple metabolism. *Cell Metab.* (2011) 13:489–90. doi: 10.1016/j.cmet.2011.04.005

89. Puertollano E, Kolida S, Yaqoob P. Biological significance of short-chain fatty acid metabolism by the intestinal microbiome. *Curr Opin Clin Nutr Metab Care*. (2014) 17:139–44. doi: 10.1097/MCO.00000000000025

90. Mikó E, Kovács T, Sebő É, Tóth J, Csonka T, Ujlaki G, et al. Microbiomemicrobial metabolome-cancer cell interactions in breast cancer-familiar but unexplored. *Cells.* (2019) 8.

91. Davar D, Dzutsev AK, McCulloch JA, Rodrigues RR, Chauvin JM, Morrison RM, et al. Fecal microbiota transplant overcomes resistance to anti-PD-1 therapy in melanoma patients. *Science(New York N.Y.).* (6529) 2021 371:595–602.

92. Sivan A, Corrales L, Hubert N, Williams JB, Aquino-Michaels K, Earley ZM, et al. Commensal Bifidobacterium promotes antitumor immunity and facilitates anti-PD-L1 efficacy. *Science(New York N.Y.).* (6264) 2015 350:1084–9.

93. Smith M, Dai A, Ghilardi G, Amelsberg KV, Devlin SM, Pajarillo R, et al. Gut microbiome correlates of response and toxicity following anti-CD19 CAR T cell therapy. *Nat Med.* (2022) 28:713–23. doi: 10.1038/s41591-022-01702-9

94. Zhou CB, Zhou YL, Fang JY. Gut microbiota in cancer immune response and immunotherapy. *Trends Cancer.* (2021) 7:647–60. doi: 10.1016/j.trecan.2021.01.010

95. Tsvetikova SA, Koshel EI. Microbiota and cancer: host cellular mechanisms activated by gut microbial metabolites. *Int J Med Microbiol IJMM*. (2020) 310:151425. doi: 10.1016/j.ijmm.2020.151425

96. Qin N, Yang F, Li A, Prifti E, Chen Y, Shao L, et al. Alterations of the human gut microbiome in liver cirrhosis. *Nature*. (7516) 2014 513:59–64.

97. Iida N, Mizukoshi E, Yamashita T, Yutani M, Seishima J, Wang Z, et al. Chronic liver disease enables gut Enterococcus faecalis colonization to promote liver carcinogenesis. *Nat Cancer.* (2021) 2:1039–54. doi: 10.1038/s43018-021-00251-3

98. Ponziani FR, Bhoori S, Castelli C, Putignani L, Rivoltini L, Del Chierico F, et al. Hepatocellular carcinoma is associated with gut microbiota profile and inflammation in nonalcoholic fatty liver disease. *Hepatology(Baltimore Md.).* (2019) 69:107–20.

99. Ma C, Han M, Heinrich B, Fu Q, Zhang Q, Sandhu M, et al. Gut microbiomemediated bile acid metabolism regulates liver cancer via NKT cells. *Science(New York* N.Y.). (2018) 360. doi: 10.1126/science.aan5931

100. Yoshimoto S, Loo TM, Atarashi K, Kanda H, Sato S, Oyadomari S, et al. Obesity-induced gut microbial metabolite promotes liver cancer through senescence secretome. *Nature*. (7456) 2013 499:97–101.

 Yamagishi R, Kamachi F, Nakamura M, Yamazaki S, Kamiya T, Takasugi M, et al. Gasdermin D-mediated release of IL-33 from senescent hepatic stellate cells promotes obesity-associated hepatocellular carcinoma. *Sci Immunol.* (2022) 7:eabl7209. doi: 10.1126/sciimmunol.abl7209

102. Han K, Nam J, Xu J, Sun X, Huang X, Animasahun O, et al. Generation of systemic antitumour immunity via the in situ modulation of the gut microbiome by an orally administered inulin gel. *Nat Biomed Eng.* (2021) 5:1377–88. doi: 10.1038/s41551-021-00749-2

103. Lee PC, Wu CJ, Hung YW, Lee CJ, Chi CT, Lee IC, et al. Gut microbiota and metabolites associate with outcomes of immune checkpoint inhibitor-treated unresectable hepatocellular carcinoma. *J immunotherapy Cancer*. (2022) 10. doi: 10.1136/jitc-2022-004779

104. Han S, Bao X, Zou Y, Wang L, Li Y, Yang L, et al. d-lactate modulates M2 tumor-associated macrophages and remodels immunosuppressive tumor microenvironment for hepatocellular carcinoma. *Sci Adv.* (2023) 9:eadg2697. doi: 10.1126/sciadv.adg2697

105. Zheng Y, Wang T, Tu X, Huang Y, Zhang H, Tan D, et al. Gut microbiome affects the response to anti-PD-1 immunotherapy in patients with hepatocellular carcinoma. *J immunotherapy Cancer.* (2019) 7:193. doi: 10.1186/ s40425-019-0650-9

106. Elkrief A, El Raichani L, Richard C, Messaoudene M, Belkaid W, Malo J, et al. Antibiotics are associated with decreased progression-free survival of advanced melanoma patients treated with immune checkpoint inhibitors. *Oncoimmunology*. (2019) 8:e1568812. doi: 10.1080/2162402X.2019.1568812

107. Vétizou M, Pitt JM, Daillère R, Lepage P, Waldschmitt N, Flament C, et al. Anticancer immunotherapy by CTLA-4 blockade relies on the gut microbiota. *Science (New York N.Y.).* (6264) 2015 350:1079–84.

108. Routy B, Le Chatelier E, Derosa L, Duong CPM, Alou MT, Daillère R, et al. Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. *Science(New York N.Y.).* (6371) 2018 359:91–7.

109. Su X, Gao Y, Yang R. Gut microbiota-derived tryptophan metabolites maintain gut and systemic homeostasis. *Cells.* (2022) 11. doi: 10.3390/cells11152296

110. Flores R, Shi J, Fuhrman B, Xu X, Veenstra TD, Gail MH, et al. Fecal microbial determinants of fecal and systemic estrogens and estrogen metabolites: a cross-sectional study. *J Trans Med.* (2012) 10:253. doi: 10.1186/1479-5876-10-253

111. Liu L, Fu Q, Li T, Shao K, Zhu X, Cong Y, et al. Gut microbiota and butyrate contribute to nonalcoholic fatty liver disease in premenopause due to estrogen deficiency. *PloS One.* (2022) 17:e0262855. doi: 10.1371/journal.pone.0262855

112. Wang MY, Sang LX, Sun SY. Gut microbiota and female health. World J Gastroenterol. (2024) 30:1655–62. doi: 10.3748/wjg.v30.i12.1655

113. Hosseini E, Grootaert C, Verstraete W, Van de Wiele T. Propionate as a healthpromoting microbial metabolite in the human gut. *Nutr Rev.* (2011) 69:245–58. doi: 10.1111/j.1753-4887.2011.00388.x

114. Wu Z, Pfeiffer RM, Byrd DA, Wan Y, Ansong D, Clegg-Lamptey JN, et al. Associations of circulating estrogens and estrogen metabolites with fecal and oral microbiome in postmenopausal women in the Ghana breast health study. *Microbiol Spectr.* (2023) 11:e0157223. doi: 10.1128/spectrum.01572-23

115. Goedert JJ, Jones G, Hua X, Xu X, Yu G, Flores R, et al. Investigation of the association between the fecal microbiota and breast cancer in postmenopausal women: a population-based case-control pilot study. *J Natl Cancer Institute*. (2015) 107.

116. Goedert JJ, Hua X, Bielecka A, Okayasu I, Milne GL, Jones GS, et al. Postmenopausal breast cancer and oestrogen associations with the IgA-coated and IgA-noncoated faecal microbiota. *Br J Cancer.* (2018) 118:471–9. doi: 10.1038/bjc.2017.435

117. Luu TH, Michel C, Bard JM, Dravet F, Nazih H, Bobin-Dubigeon C. Intestinal proportion of blautia sp. is associated with clinical stage and histoprognostic grade in patients with early-stage breast cancer. *Nutr Cancer.* (2017) 69:267–75. doi: 10.1080/01635581.2017.1263750

118. Yin Y, Sichler A, Ecker J, Laschinger M, Liebisch G, Höring M, et al. Gut microbiota promote liver regeneration through hepatic membrane phospholipid biosynthesis. *J Hepatol.* (2023) 78:820–35. doi: 10.1016/j.jhep.2022.12.028

119. Mikó E, Vida A, Kovács T, Ujlaki G, Trencsényi G, Márton J, et al. Lithocholic acid a bacterial metabolite reduces breast cancer cell proliferation and aggressiveness. Biochimica et biophysica acta. *Bioenergetics*. (2018) 1859:958–74.

120. Di Modica M, Gargari G, Regondi V, Bonizzi A, Arioli S, Belmonte B, et al. Gut microbiota condition the therapeutic efficacy of trastuzumab in HER2-positive breast cancer. *Cancer Res.* (2021) 81:2195–206. doi: 10.1158/0008-5472.CAN-20-1659

121. Li D, Yu S, Long Y, Shi A, Deng J, Ma Y, et al. Tryptophan metabolism: Mechanism-oriented therapy for neurological and psychiatric disorders. *Front Immunol.* (2022) 13:985378. doi: 10.3389/fimmu.2022.985378

122. Rothhammer V, Mascanfroni ID, Bunse L, Takenaka MC, Kenison JE, Mayo L, et al. Type I interferons and microbial metabolites of tryptophan modulate astrocyte activity and central nervous system inflammation via the aryl hydrocarbon receptor. *Nat Med.* (2016) 22:586–97. doi: 10.1038/nm.4106

123. Chyan YJ, Poeggeler B, Omar RA, Chain DG, Frangione B, Ghiso J, et al. Potent neuroprotective properties against the Alzheimer beta-amyloid by an endogenous melatonin-related indole structure indole-3-propionic acid. *J Biol Chem.* (1999) 274:21937–42. doi: 10.1074/jbc.274.31.21937

124. Gao J, Xu K, Liu H, Liu G, Bai M, Peng C, et al. Impact of the gut microbiota on intestinal immunity mediated by tryptophan metabolism. *Front Cell infection Microbiol.* (2018) 8:13. doi: 10.3389/fcimb.2018.00013

125. Blackmer-Raynolds LD, Sampson TR. The gut-brain axis goes viral. Cell Host Microbe. (2022) 30:283–5. doi: 10.1016/j.chom.2022.02.013

126. Wang M, Zhang L, Chang W, Zhang Y. The crosstalk between the gut microbiota and tumor immunity: Implications for cancer progression and treatment outcomes. *Front Immunol.* (2022) 13:1096551. doi: 10.3389/fimmu.2022.1096551

127. Green GBH, Cox-Holmes AN, Potier ACE, Marlow GH, McFarland BC. Modulation of the immune environment in glioblastoma by the gut microbiota. *Biomedicines*. (2024) 12. doi: 10.3390/biomedicines12112429

128. Cong J, Liu P, Han Z, Ying W, Li C, Yang Y, et al. Bile acids modified by the intestinal microbiota promote colorectal cancer growth by suppressing CD8(+) T cell effector functions. *Immunity.* (2024) 57:876–889.e11. doi: 10.1016/j.immuni.2024.02.014

129. Pant K, Venugopal SK, Lorenzo Pisarello MJ, Gradilone SA. The role of gut microbiome-derived short-chain fatty acid butyrate in hepatobiliary diseases. *Am J Pathol.* (2023) 193:1455–67. doi: 10.1016/j.ajpath.2023.06.007

130. Mager LF, Burkhard R, Pett N, Cooke NCA, Brown K, Ramay H, et al. Microbiome-derived inosine modulates response to checkpoint inhibitor immunotherapy. *Science(New York N.Y.).* (6510) 2020 369:1481–9.

131. Liu X, Lu B, Tang H, Jia X, Zhou Q, Zeng Y, et al. Gut microbiome metabolites molecular mimicry and species-level variation drive long-term efficacy and adverse event outcomes in lung cancer survivors. *EBioMedicine*. (2024) 109:105427. doi: 10.1016/j.ebiom.2024.105427

132. Botticelli A, Vernocchi P, Marini F, Quagliariello A, Cerbelli B, Reddel S, et al. Gut metabolomics profiling of non-small cell lung cancer(NSCLC) patients under immunotherapy treatment. *J Trans Med.* (2020) 18:49. doi: 10.1186/s12967-020-02231-0

133. Song B, Zhao K, Zhou S, Xue Y, Lu H, Jia X, et al. Association of the gut microbiome with fecal short-chain fatty acids lipopolysaccharides and obesity in young Chinese college students. *Front Nutr.* (2023) 10:1057759. doi: 10.3389/fnut.2023.1057759

134. Luu M, Weigand K, Wedi F, Breidenbend C, Leister H, Pautz S, et al. Regulation of the effector function of CD8(+) T cells by gut microbiota-derived metabolite butyrate. *Sci Rep.* (2018) 8:14430. doi: 10.1038/s41598-018-32860-x

135. Mirji G, Worth A, Bhat SA, El Sayed M, Kannan T, Goldman AR, et al. The microbiome-derived metabolite TMAO drives immune activation and boosts responses to immune checkpoint blockade in pancreatic cancer. *Sci Immunol.* (2022) 7:eabn0704. doi: 10.1126/sciimmunol.abn0704

136. Armstrong D, Cameron RG. Comparison of liver cancer and nodules induced in rats by deoxycholic acid diet with or without prior initiation. *Cancer Lett.* (1991) 57:153–7. doi: 10.1016/0304-3835(91)90209-Z

137. Režen T, Rozman D, Kovács T, Kovács P, Sipos A, Bai P, et al. The role of bile acids in carcinogenesis. *Cell Mol Life Sci CMLS.* (2022) 79:243. doi: 10.1007/s00018-022-04278-2

138. Coutzac C, Jouniaux JM, Paci A, Schmidt J, Mallardo D, Seck A, et al. Systemic short chain fatty acids limit antitumor effect of CTLA-4 blockade in hosts with cancer. *Nat Commun.* (2020) 11:2168. doi: 10.1038/s41467-020-16079-x

139. Zhu J, Luo L, Tian L, Yin S, Ma X, Cheng S, et al. Aryl hydrocarbon receptor promotes IL-10 expression in inflammatory macrophages through src-STAT3 signaling pathway. *Front Immunol.* (2018) 9:2033. doi: 10.3389/fimmu.2018.02033

140. Chng SH, Kundu P, Dominguez-Brauer C, Teo WL, Kawajiri K, Fujii-Kuriyama Y, et al. Ablating the aryl hydrocarbon receptor(AhR) in CD11c+ cells perturbs intestinal epithelium development and intestinal immunity. *Sci Rep.* (2016) 6:23820. doi: 10.1038/srep23820

141. Frankel TL, Pasca di Magliano M. Immune sensing of microbial metabolites: Action at the tumor. *Immunity*. (2022) 55:192-4. doi: 10.1016/j.immuni.2022.01.009

142. Ji Y, Gao Y, Chen H, Yin Y, Zhang W. Indole-3-acetic acid alleviates nonalcoholic fatty liver disease in mice via attenuation of hepatic lipogenesis and oxidative and inflammatory stress. *Nutrients.* (2019) 11. doi: 10.3390/nu11092062

143. Krishnan S, Ding Y, Saedi N, Choi M, Sridharan GV, Sherr DH, et al. Gut microbiota-derived tryptophan metabolites modulate inflammatory response in hepatocytes and macrophages. *Cell Rep.* (2018) 23:1099–111. doi: 10.1016/j.celrep.2018.03.109

144. Beaumont M, Neyrinck AM, Olivares M, Rodriguez J, de Rocca Serra A, Roumain M, et al. The gut microbiota metabolite indole alleviates liver inflammation in

mice. FASEB J Off Publ Fed Am Societies Exp Biol. (2018) 32:fj201800544. doi: 10.1096/ fj.201800544

145. Bansal T, Alaniz RC, Wood TK, Jayaraman A. The bacterial signal indole increases epithelial-cell tight-junction resistance and attenuates indicators of inflammation. *Proc Natl Acad Sci United States America*. (2010) 107:228–33. doi: 10.1073/pnas.0906112107

146. Walter K, Grosskopf H, Karkossa I, von Bergen M, Schubert K. Proteomic characterization of the cellular effects of ahR activation by microbial tryptophan catabolites in endotoxin-activated human macrophages. *Int J Environ Res Public Health*. (2021) 18. doi: 10.3390/ijerph181910336

147. Mantovani A, Sozzani S, Locati M, Allavena P, Sica A. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol.* (2002) 23:549–55. doi: 10.1016/S1471-4906(02)02302-5

148. Ji Y, Yin W, Liang Y, Sun L, Yin Y, Zhang W. Anti-inflammatory and antioxidative activity of indole-3-acetic acid involves induction of HO-1 and neutralization of free radicals in RAW264.7 cells. . *Int J Mol Sci.* (2020) 21. doi: 10.3390/ijms21051579

149. Ishihara Y, Kado SY, Hoeper C, Harel S, Vogel CFA. Role of NF-kB relB in aryl hydrocarbon receptor-mediated ligand specific effects. *Int J Mol Sci.* (2019) 20. doi: 10.3390/ijms20112652

150. Agista AZ, Tanuseputero SA, Koseki T, Budijanto S, Sultana H, Ohsaki Y, et al. Tryptamine a microbial metabolite in fermented rice bran suppressed lipopolysaccharide-induced inflammation in a murine macrophage model. *Int J Mol Sci.* (2022) 23. doi: 10.3390/ijms231911209

151. Liu YJ, Tang B, Wang FC, Tang L, Lei YY, Luo Y, et al. Parthenolide ameliorates colon inflammation through regulating Treg/Th17 balance in a gut microbiotadependent manner. *Theranostics*. (2020) 10:5225–41. doi: 10.7150/thno.43716

152. Chanmee T, Ontong P, Konno K, Itano N. Tumor-associated macrophages as major players in the tumor microenvironment. *Cancers*. (2014) 6:1670–90. doi: 10.3390/cancers6031670

153. Buchta Rosean C, Bostic RR, Ferey JCM, Feng TY, Azar FN, Tung KS, et al. Preexisting commensal dysbiosis is a host-intrinsic regulator of tissue inflammation and tumor cell dissemination in hormone receptor-positive breast cancer. *Cancer Res.* (2019) 79:3662–75. doi: 10.1158/0008-5472.CAN-18-3464

154. Li R, Zhou R, Wang H, Li W, Pan M, Yao X, et al. Gut microbiota-stimulated cathepsin K secretion mediates TLR4-dependent M2 macrophage polarization and promotes tumor metastasis in colorectal cancer. *Cell Death differentiation*. (2019) 26:2447–63. doi: 10.1038/s41418-019-0312-y

155. Liu T, Guo Z, Song X, Liu L, Dong W, Wang S, et al. High-fat diet-induced dysbiosis mediates MCP-1/CCR2 axis-dependent M2 macrophage polarization and promotes intestinal adenoma-adenocarcinoma sequence. *J Cell Mol Med.* (2020) 24:2648–62. doi: 10.1111/jcmm.14984

156. Busbee PB, Menzel L, Alrafas HR, Dopkins N, Becker W, Miranda K, et al. Indole-3-carbinol prevents colitis and associated microbial dysbiosis in an IL-22dependent manner. *JCI Insight*. (2020) 5. doi: 10.1172/jci.insight.127551

157. Zhu C, Xie Q, Zhao B. The role of AhR in autoimmune regulation and its potential as a therapeutic target against CD4 T cell mediated inflammatory disorder. *Int J Mol Sci.* (2014) 15:10116–35. doi: 10.3390/ijms150610116

158. Safa M, Tavasoli B, Manafi R, Kiani F, Kashiri M, Ebrahimi S, et al. Indole-3carbinol suppresses NF-κB activity and stimulates the p53 pathway in pre-B acute lymphoblastic leukemia cells. *Tumour Biol J Int Soc Oncodevelopmental Biol Med.* (2015) 36:3919–30. doi: 10.1007/s13277-014-3035-1

159. Yan XJ, Qi M, Telusma G, Yancopoulos S, Madaio M, Satoh M, et al. Indole-3carbinol improves survival in lupus-prone mice by inducing tandem B- and T-cell differentiation blockades. *Clin immunology(Orlando Fla.)*. (2009) 131:481–94.

160. Pierre S, Chevallier A, Teixeira-Clerc F, Ambolet-Camoit A, Bui LC, Bats AS, et al. Aryl hydrocarbon receptor-dependent induction of liver fibrosis by dioxin. *Toxicological Sci an Off J Soc Toxicol.* (2014) 137:114–24. doi: 10.1093/toxsci/kft236

161. Weng JR, Tsai CH, Kulp SK, Chen CS. Indole-3-carbinol as a chemopreventive and anti-cancer agent. *Cancer Lett.* (2008) 262:153–63. doi: 10.1016/j.canlet.2008.01.033

162. Hung SC, Kuo KL, Huang HL, Lin CC, Tsai TH, Wang CH, et al. Indoxyl sulfate suppresses endothelial progenitor cell-mediated neovascularization. *Kidney Int.* (2016) 89:574–85. doi: 10.1016/j.kint.2015.11.020

163. Zeb F, Naqeeb H, Osaili T, Faris ME, Ismail LC, Obaid RS, et al. Molecular crosstalk between polyphenols and gut microbiota in cancer prevention. *Nutr research (New York N.Y.).* (2024) 124:21–42.

164. Mitchem JB, Brennan DJ, Knolhoff BL, Belt BA, Zhu Y, Sanford DE, et al. Targeting tumor-infiltrating macrophages decreases tumor-initiating cells relieves immunosuppression and improves chemotherapeutic responses. *Cancer Res.* (2013) 73:1128–41. doi: 10.1158/0008-5472.CAN-12-2731

165. DeNardo DG, Brennan DJ, Rexhepaj E, Ruffell B, Shiao SL, Madden SF, et al. Leukocyte complexity predicts breast cancer survival and functionally regulates response to chemotherapy. *Cancer Discovery.* (2011) 1:54–67. doi: 10.1158/2159-8274.CD-10-0028

166. Qu R, Zhang Y, Ma Y, Zhou X, Sun L, Jiang C, et al. Role of the gut microbiota and its metabolites in tumorigenesis or development of colorectal cancer. *Advanced science(Weinheim Baden-Wurttemberg Germany)*. (2023) 10:e2205563. doi: 10.1002/advs.202205563

167. Jaye K, Chang D, Li CG, Bhuyan DJ. Gut metabolites and breast cancer: the continuum of dysbiosis breast cancer risk and potential breast cancer therapy. *Int J Mol Sci.* (2022) 23. doi: 10.3390/ijms23169490

168. Létourneau S, Krieg C, Pantaleo G, Boyman O. IL-2- and CD25-dependent immunoregulatory mechanisms in the homeostasis of T-cell subsets. J Allergy Clin Immunol. (2009) 123:758–62.

169. Cibrián D, Sánchez-Madrid F. CD69: from activation marker to metabolic gatekeeper. *Eur J Immunol.* (2017) 47:946–53. doi: 10.1002/eji.201646837

170. Hwang YJ, Yun MO, Jeong KT, Park JH. Uremic toxin indoxyl 3-sulfate regulates the differentiation of Th2 but not of Th1 cells to lessen allergic asthma. *Toxicol Lett.* (2014) 225:130–8. doi: 10.1016/j.toxlet.2013.11.027

171. Kamal A, Rao MP, Swapna P, Srinivasulu V, Bagul C, Shaik AB, et al. Synthesis of β -carboline-benzimidazole conjugates using lanthanum nitrate as a catalyst and their biological evaluation. *Organic biomolecular Chem.* (2014) 12:2370–87. doi: 10.1039/C3OB42236D

172. Nyström S, Govender M, Yap SH, Kamarulzaman A, Rajasuriar R, Larsson M. HIV-infected individuals on ART with impaired immune recovery have altered plasma metabolite profiles. *Open Forum Infect Dis.* (2021) 8:ofab288. doi: 10.1093/ofid/ofab288

173. Borghi M, Pariano M, Solito V, Puccetti M, Bellet MM, Stincardini C, et al. Targeting the aryl hydrocarbon receptor with indole-3-aldehyde protects from vulvovaginal candidiasis via the IL-22-IL-18 cross-talk. *Front Immunol.* (2019) 10:2364. doi: 10.3389/fimmu.2019.02364

174. Ghosh S, Das SK, Sinha K, Ghosh B, Sen K, Ghosh N, et al. The emerging role of natural products in cancer treatment. *Arch Toxicol.* (2024) 98:2353–91. doi: 10.1007/s00204-024-03786-3

175. Luo Y, Yin S, Lu J, Zhou S, Shao Y, Bao X, et al. Tumor microenvironment: a prospective target of natural alkaloids for cancer treatment. *Cancer Cell Int.* (2021) 21:386. doi: 10.1186/s12935-021-02085-6

176. Sun X, Zhang Y, Cheng G, Zhu T, Zhang Z, Xiong L, et al. Berberine improves DSS-induced colitis in mice by modulating the fecal-bacteria-related bile acid metabolism. *Biomedicine pharmacotherapy* = *Biomedecine pharmacotherapie*. (2023) 167:115430. doi: 10.1016/j.biopha.2023.115430

177. Eaden J, Abrams K, Ekbom A, Jackson E, Mayberry J. Colorectal cancer prevention in ulcerative colitis: a case-control study. *Alimentary Pharmacol Ther.* (2000) 14:145–53. doi: 10.1046/j.1365-2036.2000.00698.x

178. Wang M, Ma Y, Yu G, Zeng B, Yang W, Huang C, et al. Integration of microbiome metabolomics and transcriptome for in-depth understanding of berberine attenuates AOM/DSS-induced colitis-associated colorectal cancer. *Biomedicine pharmacotherapy = Biomedecine pharmacotherapie*. (2024) 179:117292. doi: 10.1016/j.biopha.2024.117292

179. Ma W, Zhang Y, Yu M, Wang B, Xu S, Zhang J, et al. *In-vitro* and *in-vivo* antibreast cancer activity of synergistic effect of berberine and exercise through promoting the apoptosis and immunomodulatory effects. *Int Immunopharmacol.* (2020) 87:106787. doi: 10.1016/j.intimp.2020.106787

180. Mao N, Yu Y, Lu X, Yang Y, Liu Z, Wang D. Preventive effects of matrine on LPS-induced inflammation in RAW 264.7 cells and intestinal damage in mice through the TLR4/NF- κ B/MAPK pathway. *Int Immunopharmacol.* (2024) 143:113432. doi: 10.1016/j.intimp.2024.113432

181. Zhang Y, Hao R, Chen J, Li S, Huang K, Cao H, et al. Health benefits of saponins and its mechanisms: perspectives from absorption metabolism and interaction with gut. *Crit Rev Food Sci Nutr.* (2024) 64:9311–32. doi: 10.1080/10408398.2023.2212063

182. Li Z, Song Y, Xu W, Chen J, Zhou R, Yang M, et al. Pulsatilla chinensis saponins improve SCFAs regulating GPR43-NLRP3 signaling pathway in the treatment of ulcerative colitis. *J ethnopharmacology*. (2023) 308:116215. doi: 10.1016/j.jep.2023.116215

183. Ai Z, Liu S, Zhang J, Hu Y, Tang P, Cui L, et al. Ginseng glucosyl oleanolate from ginsenoside ro exhibited anti-liver cancer activities via MAPKs and gut microbiota *in vitro*/vivo. *J Agric Food Chem.* (2024) 72:7845–60. doi: 10.1021/acs.jafc.3c08150

184. Xie J, Luo M, Chen Q, Zhang Q, Qin L, Wang Y, et al. Hypolipidemic effect and gut microbiota regulation of Gypenoside aglycones in rats fed a high-fat diet. *J* ethnopharmacology. (2024) 328:118066. doi: 10.1016/j.jep.2024.118066

185. Wu H, Lai W, Wang Q, Zhou Q, Zhang R, Zhao Y. Gypenoside induces apoptosis by inhibiting the PI3K/AKT/mTOR pathway and enhances T-cell antitumor immunity by inhibiting PD-L1 in gastric cancer. *Front Pharmacol.* (2024) 15:1243353. doi: 10.3389/fphar.2024.1243353

186. Khan I, Huang G, Li XA, Liao W, Leong WK, Xia W, et al. Mushroom polysaccharides and jiaogulan saponins exert cancer preventive effects by shaping the gut microbiota and microenvironment in Apc(Min/+) mice. *Pharmacol Res.* (2019) 148:104448. doi: 10.1016/j.phrs.2019.104448

187. Li H, Dong T, Tao M, Zhao H, Lan T, Yan S, et al. Fucoidan enhances the antitumor effect of anti-PD-1 immunotherapy by regulating gut microbiota. *Food Funct.* (2024) 15:3463–78. doi: 10.1039/D3FO04807A

188. Miller KD, O'Connor S, Pniewski KA, Kannan T, Acosta R, Mirji G, et al. Acetate acts as a metabolic immunomodulator by bolstering T-cell effector function and potentiating antitumor immunity in breast cancer. *Nat Cancer*. (2023) 4:1491–507. doi: 10.1038/s43018-023-00636-6

189. Shoaib M, Shehzad A, Omar M, Rakha A, Raza H, Sharif HR, et al. Inulin: Properties health benefits and food applications. *Carbohydr polymers*. (2016) 147:444–54. doi: 10.1016/j.carbpol.2016.04.020

190. Wang C, Lan T, Chen Z, Wang X, Han Y, Yang N, et al. The preventive effects of inulin cellulose and their mixture on colorectal cancer liver metastasis in mice by regulating gut microbiota. *J Food Sci.* (2023) 88:4705–17. doi: 10.1111/1750-3841.16772

191. Zhou F, Lu Y, Sun T, Sun L, Wang B, Lu J, et al. Antitumor effects of polysaccharides from Tetrastigma hemsleyanum Diels et Gilg via regulation of intestinal flora and enhancing immunomodulatory effects *in vivo. Front Immunol.* (2022) 13:1009530. doi: 10.3389/fimmu.2022.1009530

192. Xu Y, Xie L, Zhang Z, Zhang W, Tang J, He X, et al. Tremella fuciformis Polysaccharides Inhibited Colonic Inflammation in Dextran Sulfate Sodium-Treated Mice via Foxp3+ T Cells Gut Microbiota and Bacterial Metabolites. *Front Immunol.* (2021) 12:648162. doi: 10.3389/fimmu.2021.648162

193. Auyeung KK, Han QB, Ko JK. Astragalus membranaceus: A Review of its Protection Against Inflammation and Gastrointestinal Cancers. *Am J Chin Med.* (2016) 44:1–22. doi: 10.1142/S0192415X16500014

194. Li X, Khan I, Huang G, Lu Y, Wang L, Liu Y, et al. Kaempferol acts on bile acid signaling and gut microbiota to attenuate the tumor burden in ApcMin/+ mice. *Eur J Pharmacol.* (2022) 918:174773. doi: 10.1016/j.ejphar.2022.174773

195. Ma H, Yu Y, Wang M, Li Z, Xu H, Tian C, et al. Correlation between microbes and colorectal cancer: tumor apoptosis is induced by sitosterols through promoting gut microbiota to produce short-chain fatty acids. *Apoptosis an Int J programmed Cell Death.* (2019) 24:168–83. doi: 10.1007/s10495-018-1500-9

196. Zhao L, Xiao HT, Mu HX, Huang T, Lin ZS, Zhong LLD, et al. Magnolol a natural polyphenol attenuates dextran sulfate sodium-induced colitis in mice. *Molecules(Basel Switzerland)*. (2017) 22. doi: 10.3390/molecules22071218

197. Rodríguez-Morató J, Matthan NR, Liu J, de la Torre R, Chen CO. Cranberries attenuate animal-based diet-induced changes in microbiota composition and functionality: a randomized crossover controlled feeding trial. *J Nutr Biochem.* (2018) 62:76–86. doi: 10.1016/j.jnutbio.2018.08.019

198. Weh KM, Howard CL, Zhang Y, Tripp BA, Clarke JL, Howell AB, et al. Prebiotic proanthocyanidins inhibit bile reflux-induced esophageal adenocarcinoma through reshaping the gut microbiome and esophageal metabolome. *JCI Insight*. (2024) 9. doi: 10.1172/jci.insight.168112

199. Leong W, Huang G, Liao W, Xia W, Li X, Su Z, et al. Traditional Patchouli essential oil modulates the host's immune responses and gut microbiota and exhibits potent anti-cancer effects in Apc(Min /+) mice. *Pharmacol Res.* (2022) 176:106082. doi: 10.1016/j.phrs.2022.106082

200. Gu J, Sun R, Wang Q, Liu F, Tang D, Chang X. Standardized astragalus mongholicus bunge-curcuma aromatica salisb. Extract efficiently suppresses colon cancer progression through gut microbiota modification in CT26-bearing mice. *Front Pharmacol.* (2021) 12:714322. doi: 10.3389/fphar.2021.714322

201. de Vos WM. Microbe Profile: Akkermansia muciniphila: a conserved intestinal symbiont that acts as the gatekeeper of our mucosa. *Microbiology(Reading England)*. (2017) 163:646–8. doi: 10.1099/mic.0.000444

202. Zong S, Ye H, Ye Z, He Y, Zhang X, Ye M. Polysaccharides from Lachnum sp. Inhibited colitis-associated colon tumorigenesis in mice by modulating fecal microbiota and metabolites. *Int Immunopharmacol.* (2022) 108:108656. doi: 10.1016/j.intimp.2022.108656

203. Huang M, Zhang Y, Ni M, Shen M, Tao Y, Shen W, et al. Shen-Bai-Jie-Du decoction suppresses the progression of colorectal adenoma to carcinoma through regulating gut microbiota and short-chain fatty acids. . *Chin Med.* (2024) 19:149. doi: 10.1186/s13020-024-01019-4

204. Jia R, Shao S, Zhang P, Yuan Y, Rong W, An Z, et al. PRM1201 effectively inhibits colorectal cancer metastasis via shaping gut microbiota and short- chain fatty acids. *Phytomedicine Int J phytotherapy phytopharmacology.* (2024) 132:155795. doi: 10.1016/j.phymed.2024.155795

205. Zhu JJ, Liu HY, Yang LJ, Fang Z, Fu R, Chen JB, et al. Anti-tumour effect of Huangqin Decoction on colorectal cancer mice through microbial butyrate mediated PI3K/Akt pathway suppression. *J Med Microbiol.* (2023) 72. doi: 10.1099/jmm.0.001692

206. Zhu X, Zhang X, Shen J, Zheng S, Li H, Han B, et al. Gut microbiota-dependent modulation of pre-metastatic niches by Jianpi Yangzheng decoction in the prevention of lung metastasis of gastric cancer. *Phytomedicine Int J phytotherapy phytopharmacology*. (2024) 128:155413. doi: 10.1016/j.phymed.2024.155413

207. Chang Y, Ou Q, Zhou X, Nie K, Zheng P, Liu J, et al. Jianpi Jiedu decoction suppresses colorectal cancer growth by inhibiting M2 polarization of TAMs through the tryptophan metabolism-AhR pathway. *Int Immunopharmacol.* (2024) 138:112610. doi: 10.1016/j.intimp.2024.112610

208. Lu ZH, Ding Y, Wang YJ, Chen C, Yao XR, Yuan XM, et al. Early administration of Wumei Wan inhibit myeloid-derived suppressor cells via PI3K/ Akt pathway and amino acids metabolism to prevent colitis-associated colorectal cancer. *J ethnopharmacology*. (2024) 333:118260. doi: 10.1016/j.jep.2024.118260

209. Gou H, Su H, Liu D, Wong CC, Shang H, Fang Y, et al. Traditional medicine pien tze huang suppresses colorectal tumorigenesis through restoring gut microbiota and metabolites. *Gastroenterology*. (2023) 165:1404–19. doi: 10.1053/j.gastro.2023.08.052

210. Pushpass RG, Alzoufairi S, Jackson KG, Lovegrove JA. Circulating bile acids as a link between the gut microbiota and cardiovascular health: impact of prebiotics probiotics and polyphenol-rich foods. *Nutr Res Rev.* (2022) 35:161–80. doi: 10.1017/S0954422421000081

211. Ye C, Wu C, Li Y, Chen C, Li X, Zhang J, et al. Traditional medicine Xianglian pill suppresses high-fat diet-related colorectal cancer via inactivating TLR4/MyD88 by remodeling gut microbiota composition and bile acid metabolism. *J ethnopharmacology*. (2024) 333:118411. doi: 10.1016/j.jep.2024.118411

212. Li L, Wang Y, Zhao L, Ye G, Shi F, Li Y, et al. Sanhuang xiexin decoction ameliorates secondary liver injury in DSS-induced colitis involve regulating

inflammation and bile acid metabolism. J ethnopharmacology. (2022) 299:115682. doi: 10.1016/j.jep.2022.115682

213. Li Q, Li M, Li F, Zhou W, Dang Y, Zhang L, et al. Qiang-Gan formula extract improves non-alcoholic steatohepatitis via regulating bile acid metabolism and gut microbiota in mice. *J ethnopharmacology*. (2020) 258:112896. doi: 10.1016/j.jep.2020.112896

214. Shi HJ, Chen XY, Chen XR, Wu ZB, Li JY, Sun YQ, et al. Chinese medicine formula siwu-yin inhibits esophageal precancerous lesions by improving intestinal flora and macrophage polarization. *Front Pharmacol.* (2022) 13:812386. doi: 10.3389/fphar.2022.812386