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The geographic problem in cephalopod genomics

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Publications describing genomes of various cephalopod species have recently proliferated. Some papers have involved large geographic distances between the collection locality of sequenced specimens and the type locality of the presumed species. However, cryptic species have been demonstrated in many cephalopods. Therefore, even if the sequenced specimen is very similar morphologically to the species in question, the likelihood that it is a member of the species in question decreases with increasing distance from the type locality. An associated problem is that many publications do not provide information adequate to determine the source locality for the genomic sequence. We reviewed a decade of literature on mitochondrial genomes of cephalopods and found a total of 43 publications containing 48 species within 23 genera. Of the 48 species, only 17 could be evaluated for our geographic question. Distances between sampling locality and type locality of the named species ranged from 0 nautical miles (sampled at type locality) to half-way around the world. Where data were present for distance calculation, the average for the 17 species was 3785 km (2044 nmi).

KEYWORDS

biogeography, genomics, species complex, type locality, sampling

Introduction

Determination of genetic sequences has revolutionized understanding of evolutionary relationships. Increasingly sophisticated methods have allowed this revolution to progress greatly throughout the last few decades to include inferences about entire genomes. Accordingly, the literature describing cephalopod genomes, especially those of the mitochondria, has increased greatly over the past 10 years (O'Brien, 2018). The primary goal of most of these publications has been to resolve phylogenetic relationships within extant Cephalopoda.

Another result of widespread use of genetic sequencing, including "barcodes" and other sequences shorter than entire genomes, has been an increasing recognition that

species with distributions once considered to be very broad or even global were actually complexes of morphologically similar species with geographic ranges resembling a patchwork within the broad range of the species complex. Some examples include taxa within the families Sepiolidae (Fernandez-Alvarez et al., 2021), Loliginidae (Sales et al., 2017), Chtenopterygidae (Escanez et al. (2018), Ommastrephidae (Fernandez-Alvarez et al., 2020; Xu et al., 2020a), Spirulidae (Hoffmann et al., 2021), and Octopodidae (Avendano et al., 2020; Amor and Hart, 2021). Because of these species complexes, both currently recognized and possibly to be discovered in the future, a substantial potential exists for misidentification of specimens collected for genomic sequencing (e.g., Lima et al., 2017; Salvi et al., 2021). This misidentification potential is especially true if the genomic specimen is not collected within the normal range of nominal sequenced species (i.e., named based on morphological identification). We are concerned that authors, using specimens from the nearest convenient area to sample a presumed species or from sources where the actual collection locality cannot be verified (e.g., fish markets, aquarium dealers), could be using a different species than what they report and, as a result, sequences in genomic databases may be misidentified.

Materials and methods

We surveyed the past decade of genomic literature on Cephalopoda for comparison of collection locality with designated type localities of the nominal species to determine the extent of this potential problem. Only publications describing complete mitochondrial genomes were analyzed. Each publication was examined to determine the collection locality of the specimen used for genomic analysis. Type localities for the nominal species are available online. We converted both sample locality and type locality to latitude/longitude and then calculated distance between them using NOAA Latitude/Longitude Distance Calculator (https://www.nhc.noaa.gov/gccalc.shtml). The repository and accession numbers for published genome sequences were also recorded (Table 1).

TABLE 1 Mitochondrial genome sequences for cephalopods in recent literature.

Species	Reference	Genome repository	Locality of specimen	Type locality of species	Separation*
Nautilus pompilius	Wang et al., 2018	GenBank KY794928	Not given	Ambon Island, Indonesia	n/a
Sepia officinalis	Akasaki et al., 2006	GenBank AB240155	Tsukiji Fishery Market, Japan	"Oceano" [NE Atl. O.]	n/a
Sepia aculeata	Guo et al., 2016	GenBank KF690633	Not given	Java, Indonesia	n/a
Sepia apama	Kawashima et al., 2013	GenBank AP013073	Not given	Port Adelaide, Australia	n/a
Sepia esculenta	Yokobori et al., 2007	DDBJ genbank AB266516	Tsukiji Fishery Market, Japan	Yokohama Fishery Market, Japan	n/a
Sepia latimanus	Kawashima et al., 2013	GenBank AP013074	Not given	Port Dorey, New Guinea	n/a
Sepia latimanus	Lu et al., 2019	GenBank MK347498	Naozhou, China	Port Dorey, New Guinea	1890
Sepia lycidas	Kawashima et al., 2013	GenBank AP013075	Not given	Canton Fishery Market, China	n/a
Sepia lycidas	Guo et al., 2018	GenBank KJ162574	Zhanjiang fishing grounds, SE China	Canton Fishery Market, China	n/a
Sepia pharaonis	Kawashima et al., 2013	GenBank AP013076	Not given	Gulf of Suez, Red Sea	n/a
Sepia pharaonis	Song et al., 2021	ERZ1300763	Ningbo City fishfarm, China	Gulf of Suez, Red Sea	n/a
Sepia pharaonis	Wang et al., 2014	GenBank KC632521	Not given	Gulf of Suez, Red Sea	n/a
Metasepia tullbergi	Lee et al., 2021	GenBank MT974497	NE Taiwan	Nagasaki, Japan	632
Sepiella inermis	Wang et al., 2015	GenBank KF040369	Not given	Bombay [Mumbai], India	n/a
Sepiella maindroni	Zheng et al., 2016	GenBank KR912215.1	Not given	Pondichery, India	n/a
Semirossia patagonica	Kawashima et al., 2013	GenBank AP012226	Not given	Portland Bay, Patagonia	n/a
Spirula spirula	Strugnell et al., 2017	GenBank KU893141	Queensland, Australia	America	n/a
Loligo beka	Jiang et al., 2016e & 2018	GenBank KT254309	30.1°N 122.4°E, E. China Sea	Kojima Bay, Japan	585
Loligo chinensis	Jiang et al., 2017b & 2018	GenBank KT362380	30.1°N 122.4°E, E. China Sea	Canton Fishery Market, China	n/a
Loligo duvauceli	Jiang et al., 2016a & 2018	GenBank KR051264	30.1°N 122.4°E, E. China Sea	Syntypes India & Sumatra	n/a
Loligo edulis f. budo	Takemoto & Yamashita, 2012	GenBank AB675081	Multiple locations	Multiple locations, Japan	n/a
Loligo edulis f. kensaki	Takemoto & Yamashita, 2012	GenBank AB675080	Multiple locations	Multiple locations, Japan	n/a
Loligo japonica	Jiang et al., 2017b & 2018	GenBank KU568467	Hakodate, Japan	Yokohama Fishery Market, Japan	n/a

(Continued)

Species	Reference	Genome repository	Locality of specimen	Type locality of species	Separation*
Loligo opalescens	Jiang et al., 2016d & 2018	GenBank KP336703	30.10N 122.40E, E. China Sea	Puget Sound, Washington, USA	4975
Uroteuthis chinensis	Xu et al., 2020b	GenBank MN687903	Minnan–Taiwan Bank	Canton Fishery Market, China	n/a
Loliolus (N.) uyii	Jiang et al., 2016b & 2018	GenBank KP265013	30.1°N 122.4°E, E. China Sea	Kagoshima Bay, Japan	432
Sepioteuthis lessoniana	Akasaki et al., 2006	GenBank AB240154	Tsukiji Fishery Market, Japan	Not designated	n/a
Watasenia scintillans	Akasaki et al., 2006	GenBank AB240152	Tsukiji Fishery Market, Japan	Misaki[?], Japan	n/a
Watasenia scintillans	Hayashi et al., 2016	GenBank KJ845633	Toyama Bay, Japan	Misaki[?], Japan	n/a
Chiroteuthis picteti	H. Kim et al., 2018	GenBank MG833837	east sea of Korea	Ambon Island, Indonesia	2450
Bathyteuthis abyssicola	Kawashima et al., 2013	GenBank AP012225	Not given	46016'S 48027'E, Southern Ocean	n/a
Thysanoteuthis rhombus	Tang et al., 2021	GenBank MT733875	South China Sea	Strait of Messina, Sicily	5070
Illex argentinus	Jiang et al., 2016c	GenBank KP336702	Not given	Patagonia, 390S 550W	n/a
Todarodes pacificus	Akasaki et al., 2006	GenBank AB240153	Tsukiji Fishery Market, Japan	Hokodate, Japan	n/a
Sthenoteuthis oualaniensis	Xu et al., 2020c	GenBank MT661575	17059'N 111059'E, China Sea	Oualan Island, Caroline Islands	3080
Vampyroteuthis infernalis	Yokobori et al., 2007	DDBJ genbank AB266515	Ogasawara Island Chain, Japan	1056.7'S 7040.6'E, Atlantic Ocean	7715
Amphioctopus aegina	Zhang et al., 2017	GenBank KT428877	Haikou Fishery Market, China	Not designated	n/a
Amphioctopus fangsiao	Lashari et al., 2020	GenBank MF029678- 029691	9 separate localities in China	Japan	n/a
Amphioctopus marginatus	Tang et al., 2018	GenBank KY646153	Haikou Fishery Market, China	Kamae, Oita Prefecture, Japan	n/a
Amphioctopus neglectus	Tang et al., 2019	GenBank MF447873	Nanning Fishery Market, China	Ko Phuket, Thailand	n/a
Amphioctopus rex	Tang et al., 2019	GenBank MF447874	Wenzhon Fishery Market, China	Ko Food, Trat Province, Thailand	n/a
Octopus bimaculatus	Dominguez et al., 2016	GenBank KT581981	N. Gulf of California, Mexico	Syntypes; 3 localities	n/a
Octopus conispadiceus	Ma et al., 2016	GenBank KJ789854	Haishenwai, Amur Bay, Russia	Sapparo Fishery Market, Japan	n/a
Octopus dollfusi	Yan et al., 2018	GenBank KX108697	Zhanjiang, Guangdong, China	"Indochina"	n/a
Octopus fitchi	Magallon-Gayon et al., 2020	GenBank MK450541	Bahia Magdalena, Mexico	N. Gulf of California, Mexico	990
Octopus minor	B. Kim et al., 2018	SRA database SRX3462978	Not given	Suruga Bay, Japan	n/a
Octopus minor	Cheng et al., 2012	GenBank HQ638215	Weihai, Shandong Province, China	Suruga Bay, Japan	785
Octopus mimus	Magallon-Gayon et al., 2020	GenBank MN078094	Zihuatanejo, Guerrero, Mexico	Iquique, Chile	2925
Octopus ocellatus	Akasaki et al., 2006	GenBank AB240156	Tsukiji Fishery Market, Japan	"China Sea"	n/a
Octopus sinensis	Li et al., 2021	GenBank MT712046	Zhoushan, China	Oyano Island, Ariake Sea, Japan	446
Octopus vulgaris	Zarrella et al., 2019	Not listed	Bay of Naples, Italy	"Mediterranean Sea"	n/a
Cistopus chinensis	Cheng et al., 2013	GenBank KF017606	coastal Xiamen, China	Xiamen, China	0!!
Cistopus taiwanicus	Cheng et al., 2013	GenBank KF017605	"coastal Taiwan"	Miaoli, Taiwan	close+/-
Hapalochlaena fasciata	Kim et al., 2020	GenBank MT497543	Southern coastal Korea	Port Jackson, Australia	1165
Hapalochlaena maculosa	Morse et al., 2018	Not listed	8 South Australia localities	"Australia"	n/a
Argonauta argo	Hirota et al., 2021	DDBJ genbank LC596061	Oki Island, Sea of Japan	Syntypes; Red Sea + Mediterranean	n/a
Argonauta hians	Chiu et al., 2018a	GenBank KY649285	Kenting, Taiwan	Ambon Island, Indonesia	1600
Tremoctopus violaceus	Chiu et a.l, 2018b	GenBank KY649286	Taiwan	Not designated	n/a

Approx. distance between specimen location and type locality calculated using https://www.nhc.noaa.gov/gccalc.shtml Separation* is calculated as nmi. (1nmi = 1.852 km). Shaded boxes: data absent or too general to be analyzed. Bold numbers in the last column highlight the publications for which included information was adequate for distance calculation.

Results

An online search of the previous ten years of Cephalopoda genomic literature found a total of 58 genomic descriptions within 43 publications containing 48 different species in 23 genera (Table 1). For many species sequenced (70%), either collection locality or type locality (from the original description) was missing or was too general (e.g., Australia). In addition, if either locality was indeterminate (e.g., Tsukiji fishery market); or there were multiple type localities (ex. syntypes); or the genome was derived from combined specimens from multiple localities, the sequence was not included in our distance analysis.

Of the 48 species sequenced, only 17 could be evaluated for our geographic question (Table 1). Distances calculated ranged from 0 km (sampled at type locality) to half-way around the world in a different ocean basin. The average distance between sampling locality and type locality for the 17 species for which data were adequate for distance calculation, was 3785 km (2044 nm).

Incidentally, as we reviewed this literature for geographic information, we also noticed that very few of the publications included any indication that voucher specimens or unprocessed tissue were preserved in established archival collections for future research. For example, of the 17 species mentioned above, only 5 (29.4%) had vouchered specimens. Thus, 10.4% of species accounts included both adequate geographic information and archived specimens.

Discussion

Our point here is not that any of these publications is wrong. Rather, we want to highlight the potential for taxonomic errors in publications where the sampling area is very distant from the species' type locality. As pointed out by one of the reviewers, for coastal cephalopod species in complex habitats, such errors are possible even at very small distances. Any taxonomic error introduced by this geographic mismatch may be compounded when the sequence is archived in a genomic database and the database is used for other investigations.

We therefore recommend selection of specimens for genomic sequencing collected from as close to the type locality of the species as possible. Although we recognize that it may not always be possible to sample the type locality, we recommend that the genomic sample be from the same biogeographic province (e.g., GOODS, 2009 or subsequent modifications by various authors) or "Large Marine Ecosystem" (LME – Sherman and Duda, 2011) as the type locality. The collecting locality should always be included in any publication resulting from DNA sequencing. Furthermore, specimens should not be selected for sequencing from a source where the actual collecting locality cannot be determined confidently (e.g., not from fishery landings, etc.). Also, although our primary purpose here is to highlight the need for sequenced specimens to come from as close to the type locality as possible, we also recommend that specimens sequenced and any unprocessed tissue be vouchered in an established archival collection. Relevant information about archived material (e.g., museum catalogue number) should be included in resulting publications.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding author.

Ethics statement

Ethical review and approval was not required for the animal study because this is a literature review. No animals (live or preserved) were used.

Author contributions

MS conceived the idea. MV and MS analyzed the data. PR accumulated the references. MV wrote the first draft. All authors contributed to the final manuscript. All authors contributed to the article and approved the submitted version.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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