



Performance Evaluation of BD Phoenix NMIC-413 Antimicrobial Susceptibility Testing Panel for Imipenem, Meropenem, and Ertapenem Against Clinical Carbapenem-Resistant and Carbapenem-Susceptible Enterobacterales

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Purpose: The infection of carbapenem-resistant *Enterobacterales* (CRE) has become a major clinical and healthcare problem worldwide. The screening methods of CRE have been extensively developed but still need improving [e.g., tests with accurate and simple minimum inhibitory (MICs)]. In this study, the performance of the BD Phoenix NMIC-413 AST panel was evaluated against clinical CRE and carbapenem-susceptible *Enterobacterales* (CSE) in China. The panel was first evaluated in the Chinese clinical lab.

Methods: Antimicrobial susceptibility testing of 303 clinical *Enterobacterales* isolates were conducted by broth microdilution (BMD), Phoenix NMIC-413 AST panel, and disk diffusion method for imipenem, ertapenem, and meropenem. Considering BMD is a gold standard, essential agreement (EA), categorical agreement (CA), minor error (MIE), major error (ME), and very major error (VME) were determined according to CLSI guidelines. CA and EA > 90%, ME < 3%, and VME < 1.5% were considered as acceptable criteria. Polymerase chain reaction and sanger sequencing were performed to determine the β -lactamase genotypes of CRE isolates.

Results: Three hundred and three isolates included 195 CREs and 108 CSEs were enrolled according to the BMD-MIC values of three carbapenems. Tested CREs showing 100 $bl_{A\text{KPC}-2}$ -positive organisms, 31 $bl_{a\text{IMP}}$ -positive organisms, 28 $bl_{a\text{NDM}}$ -positive organisms, 5 $bl_{a\text{VIM}}$ -positive organisms, 2 both $bl_{a\text{IMP}}$ and $bl_{a\text{VIM}}$ -positive organisms, 2 $bl_{a\text{OXA}-48}$ -positive organisms, and 27 isolates without carbapenemase genes. For the Phoenix NMIC-413 method, CA and EA rates >93%, MIE rates <5%, ME rates <1.75%, and VME rates were 0%, across the three drugs. For the disk diffusion method, the CA rates for three drugs were all >93%, while the MIE and ME rates were all <5 and <3%,

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respectively. VME rate was 3.28% for imipenem, exceeded the cut-off value specified by CLSI M52, 0 and 0.56% for ertapenem and meropenem, separately.

Conclusion: Based on the genomic data, the detection of CRE and CSE was more reliable using the BD Phoenix NMIC-413 panel compared to the BMD and disk approaches. Therefore, our study supports the use of BD Phoenix NMIC-413 panel as a suitable alternative to BMD for the detection of carbapenem resistant isolates in a clinical setting.

Keywords: BD Phoenix NMIC-413, CRE, broth microdilution, disk diffusion, evaluation

INTRODUCTION

Carbapenem-resistant *Enterobacterales* (CRE) is a major clinical and public health issue worldwide, which can cause infections associated with high mortality and have limited treatment options (1, 2). CREs are generally resistant to all β -lactams, including carbapenems such as imipenem, meropenem, ertapenem, doripenem (3), and other antibiotics such as cephalosporins, quinolones, and aminoglycosides, which further restrict the choice of antibiotic treatment. After the initial report of KPC-1 (*Klebsiella pneumoniae* carbapenemase-1) from a strain of *K. pneumoniae* discovered in North Carolina in 2001 (4), CRE has been widely reported in almost every state (1). In China, the incidence of carbapenem-resistant *Escherichia coli* and *K. pneumoniae* increased from 0 and 0.7% in 2004 to 1.0 and 13.4% in 2014 (5, 6).

The mechanism of Carbapenem resistance can be divided into two types: carbapenemase factor and carbapenemase-nonproducing factor. Carbapenem resistance in *Enterobacterales* is mainly mediated by the horizontal transfer of genes encoding carbapenemases, although porin mutations or overexpression of efflux pumps can lead to carbapenem resistance, especially in combination with the hyperproduction of β -lactamase (7). Carbapenemases consisted of different molecular classes: A, B, and D of the Ambler classification (8). The clinically most important and frequent carbapenemases in *Enterobacterales* are class A (KPCs), class B metallo- β -lactamases (VIM, IMP, and NDM), and class D (OXA-48) subgroups and their variants (9–12).

Recently, different methods were developed to detect CRE, such as the disk diffusion method, Brilliance TM CRE Agar, chromID Carba, and molecular methods (13, 14). However, phenotypic antimicrobial susceptibility assay which can accurately determine minimum inhibitory concentrations (MICs) is still the key method to guide clinical medication quickly and precisely. Most of the products in the market used to measure the MICs were based on the broth microdilution (BMD) or improved BMD method, such as BioMérieux VITEK 2, Beckman Coulter MicroScan WalkAway, and BD Phoenix. BD Phoenix NMIC-413 panel is a new panel that has recently been marketed, and covered the main cephalosporins and carbapenems such as imipenem, meropenem, and ertapenem. However, its performance to detect carbapenem susceptibility was not well-evaluated yet. In this study, the performance of the BD Phoenix NMIC-413 panel were evaluated using carbapenem-resistant and carbapenem-susceptible clinical isolates in China.

MATERIALS AND METHODS

Isolates

The Human Research Ethics Committee of our hospital approved the study protocols (Et. Number: S-K677). Three hundred and three clinical Enterobacterales isolates from Peking Union Medical College Hospital from 2010-2019 were evaluated in this study. The majority of the specimens were taken from sputum (74, 24.42%), blood (59, 19.47%), urine (58, 19.14%), bronchoalveolar lavage fluid (33, 10.89%), peritoneal fluid (29, 9.57%), gall bladder (19, 6.27%), abscess (12, 3.96%), wound (6, 1.98%), and others (13, 4.29%). Strains were isolated from surgery department (115, 37.95%), medicine department (96, 31.68%), ICU (65, 21.45%), emergency department (21, 6.93%), and pediatric department (6, 1.98%) (Supplementary Table 1). Isolates were identified using MALDI-TOF MS (Vitek MS, BioMérieux, France). All duplicate isolates (the same genus and species from the same patient) were excluded. Isolates were stored at -80°C in a cryotube with 20% (w/v) skimmed milk until subcultured on Blood Agar Plate (Oxoid, Basingstoke, United Kingdom). *Klebsiella pneumoniae* BAA 1705 (*bla*_{KPC-2}), Es. coli ATCC 2452 (bla_{NDM-1}), K. pneumoniae ATCC 700603, and Es. coli ATCC 25922 were used as quality control strains.

BD Phoenix System

the BD Phoenix NMIC-413 panel (BD Catalog Number: 448442) was used to determined MICs of imipenem (range: 0.25-8 mg/L), meropenem (range: 0.125-8 mg/L), and ertapenem (range: 0.25-2 mg/L) according to the manufacturer's instructions. In short, the identification broth was regulated with bacterial colonies from Blood Agar Plates to 0.5 McFarland standard using a spectrophotometric device. Transferred 25 μ L 0.5 McFarland identification broth suspension to the Phoenix Antibiotic susceptibility testing (AST) broth, which was supplemented with 50 μ L of the Phoenix AST indicator for the organism growth detecting before added to the panels. The panels were loaded into the Phoenix device (M50). The results were analyzed using Epicenter data management software version 6.61A (BD Diagnostic Systems) after 16 h of incubation (15, 16).

BMD Method

The susceptibility of strains to imipenem (range: 0.12–128 mg/L), meropenem (range: 0.12–128 mg/L), and ertapenem (range: 0.12–128 mg/L) were tested and analyzed. Three antimicrobial powders were obtained from National Institutes for Food and Drug Control (Beijing, China). A 0.5 McFarland standard suspension was prepared and used to inoculate the reference BMD panel according to CLSI M100 (17). Incubated these panels at 35°C for 16–20 h. Clinical carbapenem breakpoints for susceptibility/resistance were $\leq 1/\geq 4$ mg/L for imipenem and meropenem and $\leq 0.5/\geq 2$ mg/L for ertapenem.

Disk Diffusion Method

Disk diffusion tests for imipenem, meropenem, and ertapenem (Oxoid, Basingstoke, United Kingdom) were carried out according to CLSI M2 (18). The content of three antimicrobials in each disk was 10 µg. The disk was pasted to the MH agar plate using sterile tweezers and inoculated with 0.5 McFarland standard suspension. Incubated these plates at 35°C for 16–20 h, the diameter of the inhibition zone was measured with a vernier caliper. The zone diameter \geq 23 mm indicated that the strain was susceptible to imipenem and meropenem and \geq 22 mm to ertapenem, whereas the zone diameter \leq 19 mm indicated that the strain was resistant to imipenem and meropenem and \leq 18 mm to ertapenem.

Screening of Carbapenemase Genes

Polymerase chain reaction (PCR) and Sanger sequencing were used to screen out carbapenemase genes, including KPC, NDM, VIM, IMP, and OXA-48. The oligonucleotide sequences of the primers were listed in **Supplementary Table 2** (9, 19, 20). The QIAquick PCR Purification Kit was used to purify the PCR product, #REF is 28104. The PCR products were sequenced and analyzed using BLAST (http://www.ncbi.nlm.nih.gov/BLAST). Our BLAST cut-off is that the percent identity must be 100%.

Data Analysis

Using BMD as a gold stand, the categorical agreement (CA), essential agreement (EA), minor error (MIE), major error (ME), and very major error (VME) were calculated (21, 22). Results were considered CA when isolates had the same susceptible, intermediate, susceptible-dose dependent, and resistant category with the BMD method category result. Results were considered EA when the MIC obtained with the BD Phoenix NMIC-413 panel was within one doubling dilution step (two-fold serial) of the MIC value established by the BMD method. Results were considered ME when the BMD method result was susceptible and the BD Phoenix NMIC-413 panel is resistant. Results were considered MIE when one result was intermediate and the other was susceptible or resistant. Results were considered VME when the BMD method result was resistant and the BD Phoenix NMIC-413 panel was susceptible. The calculation formulas of related parameters were shown in Supplementary Table 3. The Spearman correlation coefficients (P-value) were calculated by SPSS 26.0. The linear regression curve was performed using GraphPad Prism8, and the R^2 -value was obtained at same time.

RESULTS

Isolates Information

Klebsiella pneumoniae (n = 142) accounted for the highest proportion, followed by *Enterbacter cloacae* (n = 62), *Enterbacter* coli (n = 59), Klebsiella aerogenes (n = 19), Citrobacter freundii (n = 9), Serratia marcescens (n = 6), Klebsiella oxytoca (n = 6)5), and Proteus mirabilis (n = 1) (Figure 1). There were 195 strains resistant to at least one of the 3 antimicrobials: imipenem, meropenem, and ertapenem. The susceptibility of 303 isolates to 3 carbapenems with different methods were shown in Figure 2. The numbers of imipenem, ertapenem, and meropenem resistant strains were 182, 194, and 179 by BMD, respectively. All three antibiotics were resistant to 177 strains, and at least one of them was resistant to 195 strains. There were 195 CRE and 108 carbapenem-susceptible Enterobacterales (CSE) in this study. The resistance rate of the BD Phoenix NMIC-413 (61.06, 65.35, and 58.75%) was in general agreement with BMD (60.07, 64.03, and 59.08%), and better than disk diffusion (55.45, 65.35, and 56,12%) (P > 0.05).

Genotype Determination

The genotypes determined by PCR and Sanger sequencing demonstrated different resistance mechanisms which were shown in **Table 1**. The highest proportion was the KPC-2 carbapenemase gene (100/195), followed by IMP (33/195), NDM (28/195), VIM (7/195), and OXA-48(2/195). Two isolates produced both IMP-1 and VIM-1. A total of 27 CRE isolates were carbapenemase gene negative in this study. KPC-2 was the most prevalent carbapenemase gene in *K. pneumoniae* strains, while IMP and NDM accounted for the majority gene type in *E. cloacae* and *C. freundii*. All carbapenem-susceptible isolates (N = 108) were proved no carbapenemase genes by PCR method.

Performance of BD NMIC-413 Panel vs. Disk Diffusion Method

Figure 3 and **Table 2** showed CAs, EAs, and the number of MIE, ME, and VME for 303 clinical *Enterobacterales* isolates. CA and EA were all above 90%, and the CA of the three antibiotics in the BD Phoenix NMIC-413 was greater than the disk diffusion.

For imipenem, CA and MIE of the BD Phoenix NMIC-413 were similar with the disk diffusion. However, the VME of BD Phoenix NMIC-413 was 0%, while the disk diffusion was 3.28%. The ME rate of disk diffusion was close to 3%, while the BD Phoenix NMIC-413 was only 0.99%. For meropenem, the CA, MIE, ME, VME of BD NMIC-413 and disk diffusion was 97.69 vs. 95.38%, 1.98 vs. 3.30%, 0.83 vs. 1.65%, 0 vs. 0.56%. All the indexes of BD Phoenix NMIC-413 were better than disk diffusion. The performance of ertapenem of both methods is comparable, the CA, MIE, ME, VME was 95.05 vs. 94.06%, 4.62 vs. 4.95%, 0.99 vs. 2.97%, 0 vs. 0%.

Figures 4A–C showed the linear regression curve between the MICs determined by BD NMIC-413 and BMD. The R^2 value of imipenem, ertapenem, and meropenem was 0.97, 0.99, and 0.97, respectively. The *P*-values were all <0.001. Therefore, the BD NMIC-413 had the best performance in the detection of ertapenem. **Figures 4D–F** showed the linear regression curve





between disk diffusion and BMD. The R^2 -values were 0.81–0.87, of which the R^2 -value of ertapenem was the largest (0.87).

Figures 3A–C displayed the MICs distribution of imipenem, ertapenem, and meropenem by BMD and the BD Phoenix NMIC-413. Most MEs were clustered near the susceptibility breakpoint. For imipenem and ertapenem, the resistance rates detected by BD Phoenix NMIC-413 were higher than BMD.

Figures 3D-F displayed the MICs and disk zoom diameter of three antibiotics by BMD and disk diffusion. Only in ertapenem, the number of drug resistance measured by the disk diffusion method was more than that of BMD (198 vs. 194), and the number of drug resistance of the other two drugs were nearly 10 less (imipenem: 168 vs. 182; meropenem: 170 vs. 179).

TABLE 1	The enzyme t	vpe and	strain types	of 195	CRE strains
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Organism	Number and percentage of isolates with different resistance mechanisms [n (%)]							
	KPC	IMP	NDM	VIM	OXA-48	Carbapenemase negative		
Klebsiella pneumoniae (110)	84 (76.36)	10 (9.09)	6 (5.45)	3 (2.73)	0	7 (6.36)		
Enterobacter cloacae (38)	4 (10.53)	11 (28.95)	14 (36.84)	0	0	9 (23.68)		
Escherichia coli (21)	6 (28.57)	2 (9.53)	6 (28.57)	1 (4.76)	2 (9.52)	4 (19.05)		
Klebsiella aerogenes (8)	3 (37.50)	1 (12.50)	0	0	0	4 (50.00)		
Citrobacter freundii (7)	0	6 (85.71)	1 (14.29)	0	0	0		
Serratia marcescrns (5)	1 (20.00)	0	0	1 (20.00)	0	3 (60.00)		
Klebsiella oxytoca (5)	1 (20.00)	3 (60.00)	1(20.00)	0	0	0		
Proteus mirabilis (1)	1 (100.00)	0	0	0	0	0		

KPC, Klebsiella pneumoniae carbapenemase; NDM, New Delhi metallo-beta-lactamase; VIM, Verona integron-borne metallo-beta-lactamase; IMP, imipenemase; OXA, oxacillinase.



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Performance Evaluation Against CRE Strains With Different Genotypes

For imipenem, the BD NMIC-413 and disk diffusion method showed a CA rate of 62.96 and 74.07% separately against carbapenemase-non-producing CREs. The disk diffusion performed not well in IMP-producing CREs, with CA of 72.73% and VME of 9.09%. There was a KPC-producing *K. pneumoniae*, an OXA-48-producing *Es. coli*, and an IMP-producing *K. aerogenes* in the BD NMIC-413 that belong to MIE, the MIC detected by BMD and BD NMIC-413 were 4 and 2 mg/L, respectively.

For ertapenem, two methods performed well in the detection of different enzyme types, only disk diffusion had a slightly lower CA (93.94%) in the detection of IMP type

isolates. Ertapenem was the best to detect CRE among the three antibiotics.

For meropenem, the EA of IMP-producing CREs was <90% (87.88%) when detected by the BD Phoenix NMIC-413. When testing the IMP/VIM producing isolates or carbapenemase-non-producing isolates, the disk diffusion method showed that the CA was <90%, while the VME of IMP-producing isolates was >3% (3.13%). There was one KPC-producing *Es. coli*, one carbapenemase-non-producing *E. cloacae*, and one carbapenemase-non-producing *K. pneumoniae* in BD NMIC-413 belong to MIE. These results of BMD were resistant (MIC was 8, 64, and 4 mg/L, respectively), while BD NMIC-413 were intermediate (MIC all was 2 mg/L). Meanwhile, there was also one IMP-producing *K. pneumoniae* and one OXA-48-producing

TABLE 2 Performance of the BD Phoenix NMIC-413 and disk diffusion compared with BMD for Enterobacterales isolates.

Antimicrobial agent	BD Phoenix NMIC-413 vs. BMD N (%)				Disk diffusion vs. BMD N (%)				
	CA	EA	MIE	ME	VME	CA	MIE	ME	VME
Imipenem	284 (93.73)	284 (93.73)	16 (5.28)	2 (1.75)	0	283 (93.40)	14 (4.62)	0	6 (3.28)
Ertapenem	288 (95.05)	296 (97.69)	14 (4.62)	1 (0.99)	0	285 (94.06)	15 (4.95)	3 (2.97)	0
Meropenem	296 (97.69)	292 (96.37)	6 (1.98)	1 (0.83)	0	289 (95.38)	10 (3.30)	2 (1.65)	1(0.56)

S, susceptible; I, intermediate; R, resistant; BMD, broth microdilution; CA, categorical agreement; EA, essential agreement; MIE, minor error; ME, major error; VME, very major error.



Es. coli, the results detected by BMD were intermediate (MIC for both of them was 2 mg/L), while BD NMIC-413 were resistant and susceptible (MIC was 4 and 1 mg/L). Another one KPC-producing *K. pneumoniae* was also MIE, and the result of BMD was susceptible (MIC was ≤ 0.12 mg/L) while NMIC413 results were intermediate (MIC was 2 mg/L).

Table 3 showed the rates of CAs, EAs, and the number of MIE and VME for 195 resistant isolates with the three antibiotics.

DISCUSSION

In prior studies, the BD Phoenix NMIC-500 and NMIC-203 panels have been reported for the performance evaluation of negative bacilli, and PMIC-84 panel has reported for positive cocci (15, 23), including identification and antimicrobial

susceptibility testing evaluation (24). In this study, the performance of the BD Phoenix NMIC-413 system used for CRE testing was compared with the traditional disk diffusion method, while BMD was used as the reference method. Compared with BMD and disk diffusion, the BD Phoenix NMIC-413 panel had higher CA/EA and lower ME/MIE/VME. For carbapenemase genes negative CRE isolates, the CA rates of imipenem were <75%, while the other two antibiotics were all >90%. Imipenem was often degraded by the enzyme dehydropeptidase-1 (DHP-1), hence the co-administration with a DHP-1 inhibitor such as cilastatin was required (25). Later meropenem and ertapenem demonstrated increased stability to DHP-1 and were administered without a DHP-1 inhibitor (25). So meropenem and ertapenem are more stable than Imipenem. For intermediate isolates, the MICs obtained by BD

TABLE 3 | Performance of the BD Phoenix NMIC-413 and disk diffusion compared with BMD for 195 CREs.

Enzyme		BD Phoenix NMIC-41	3 vs. BMD <i>N</i> (%)	Disk diffusion vs. BMD N (%)			
	CA	EA	MIE	VME	CA	MIE	VME
Imipenem							
KPC (100)	99 (99.00)	99 (99.00)	1 (1.00)	0	99 (99.00)	1 (1.00)	0
NDM (28)	28 (100)	28 (100)	0	0	28 (100)	0	0
IMP (33)	32 (96.97)	33 (100)	1 (3.03)	0	24 (72.73)	6 (18.18)	3 (9.09)
VIM (7)	7 (100)	7 (100)	0	0	7 (100)	0	0
OXA-48 (2)	1 (50.00)	2 (100)	1(50.00)	0	2 (100)	0	0
no enzyme (27)	17 (62.96)	26 (96.30)	10 (37.04)	0	20 (74.07)	5 (18.52)	3 (11.11)
Ertapenem							
KPC (100)	99 (99.00)	99 (99.00)	0	0	99 (99.00)	0	0
NDM (28)	28 (100)	28 (100)	0	0	28 (100)	0	0
IMP (33)	33 (100)	33 (100)	0	0	31 (93.94)	2 (6.06)	0
VIM (7)	7 (100)	7 (100)	0	0	7 (100)	0	0
OXA-48 (2)	2 (100)	2 (100)	0	0	2 (100)	0	0
No enzyme (27)	27 (100)	27 (100)	0	0	27 (100)	0	0
Meropenem							
KPC (100)	99 (99.00)	96 (96.00)	1 (1.00)	0	98 (98.00)	1 (1.00)	0
NDM (28)	28 (100)	28 (100)	0	0	28 (100)	0	0
IMP (33)	31 (93.94)	29 (87.88)	2 (6.06)	0	27 (81.82)	5 (15.15)	1 (3.13)
VIM (7)	7 (100)	7 (100)	0	0	6 (85.71)	1 (14.29)	0
OXA-48 (2)	1 (50.00)	2 (100)	1 (50.00)	0	2 (100)	0	0
No enzyme (27)	25 (92.59)	25 (92.59)	2 (7.41)	0	23 (85.19)	3 (11.11)	0

S, susceptible; I, intermediate; R, resistant; BMD, broth microdilution; CA, categorical agreement; EA, essential agreement; MIE, minor error; VME, very major error; KPC, Klebsiella pneumoniae carbapenemase; NDM, New Delhi metallo-beta-lactamase; VIM, Verona integron-borne metallo-beta-lactamase; IMP, imipenemase; OXA, oxacillinase.

Phoenix system NMIC-413 panel were probably higher than BMD. Considering that the BD Phoenix system is a broth-based microdilution method that not only measures turbidity but also detects the bacterial growth utilizing the redox indicator (26), enabling it to detect resistant bacteria with high sensitivity. In addition, the BD Phoenix system had fewer manual operations, which was simpler and more convenient compared with the BMD method. The advantages and disadvantages of the three methods were shown in **Supplementary Table 4**.

Haffler et al. (27) have evaluated one panel of the BD Phoenix system and showed that the CA of ertapenem and meropenem were 94 and 50%. Hogan et al. (28) have evaluated the AST of VITEK 2 (bioMérieux, France) with Gram-negative bacteria by blood culture and reported that the CA and EA were 86.5 and 84.6, 96.2, and 96.2% for meropenem and ertapenem, respectively. In our study, the CA and EA of the BD Phoenix system NMIC-413 panel were ranged from 93 to 98%.

However, the BD Phoenix system NMIC-413 panel can only obtain MICs of the designated antibiotics, so it cannot be used for taxonomic identification of the selected isolates. However, the BD Phoenix CPO panel can detect the MIC and classification of CRE at the same time. It can divide CRE into three categories: A (KPC), B (NDM, IMP, and VIM), and D (OXA-48). Saad Albichr et al. (29) evaluated the performance of the automated BD Phoenix CPO Detect-test for detection and Ambler classification of carbapenemases in *Enterobacterales, P. aeruginosa*, and *A. baumannii* complex, the overall sensitivity and specificity were 89.7 and 83.5%, respectively, 68.9 and 62.1% for *P. aeruginosa*, respectively. Although the BD Phoenix system can detect the classification of CRE expediently, the accuracy needed to improve, it also needs other methods to verify.

KPC type enzyme is the most prevalent carbapenemase in China (30). A. Antonelli has investigated the sensitivity of 6 different commercial methods (Sensititre, Microscan, Vitek2, Etest, Kirby-Bauer, and MIC strip) on KPC-producing isolates (31). In that study, the CA rates of imipenem, meropenem, and ertapenem were 16.7–51.8%, 14.8–79.6%, and 83.3–96.3%, respectively, which were much lower than NMIC-413 panel evaluated in this study.

The BD phoenix NMIC-413 panel also had a notable limitation in this study. There are not enough $bla_{\rm VIM}$ and $bla_{\rm OXA}$ producing strains among the isolates tested in this study. Mainly because these two carbapenemase types are very rare in China (32, 33). From 2010 to 2019, we isolated only 7 $bla_{\rm VIM}$ and 2 $bla_{\rm OXA}$ producing strains in our laboratory. Now we do not have enough VIM and OXA-48 to make up for this limitation. In the next work, we will deliberately save these two types of strains to facilitate future research.

CONCLUSION

In conclusion, for the BD Phoenix NMIC-413 panel, CA and EA rates > 93%, MIE rates < 5%, ME rates < 1.75%, and

VME rates were 0%, across the three drugs. It showed acceptable performance as alternatives to the BMD method for clinical treatment explanation.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding authors.

ETHICS STATEMENT

The protocol has been reviewed by the Human Research Ethics Committee of the Institutional Review Board (IRB) of the Peking Union Medical College Hospital and since all bacterial strains were from residual samples used in clinical diagnosis or strains from their subcultures, it met the criteria for exemption. This project did not involve any patient information nor did it affect the normal diagnosis and treatment of patients, and after consultation with the IRB, formal ethical approval was reviewed and waived; written patient consent was not required (Ethics Approval Number: S-K677).

REFERENCES

- Guh AY, Bulens SN, Mu Y, Jacob JT, Reno J, Scott J, et al. Epidemiology of carbapenem-resistant Enterobacteriaceae in 7 US communities, 2012-2013. *JAMA*. (2015) 314:1479–87. doi: 10.1001/jama.2015.12480
- Zhang Y, Wang Q, Yin Y, Chen H, Jin L, Gu B, et al. Epidemiology of carbapenem-resistant Enterobacteriaceae infections report from the China CRE network. *Antimicrob Agents Chemother*. (2018) 62:e01882– 17. doi: 10.1128/AAC.01882-17
- Miao M, Wen H, Xu P, Niu S, Lv J, Xie X, et al. Genetic diversity of carbapenem-resistant Enterobacteriaceae (CRE) clinical isolates from a tertiary hospital in Eastern China. *Front Microbiol.* (2018) 9:3341. doi: 10.3389/fmicb.2018.03341
- Wei ZQ, Du XX, Yu YS, Shen P, Chen YG, Li LJ. Plasmid-mediated KPC-2 in a *Klebsiella pneumoniae* isolate from China. *Antimicrob Agents Chemother*. (2007) 51:763–5. doi: 10.1128/AAC.01053-06
- Hu FP, Guo Y, Zhu DM, Wang F, Jiang XF, Xu YC, et al. Resistance trends among clinical isolates in China reported from CHINET surveillance of bacterial resistance, 2005-2014. *Clin Microbiol Infect.* (2016) 22(Suppl. 1):S9– 14. doi: 10.1016/j.cmi.2016.01.001
- Yang Y, Chen J, Lin D, Xu X, Cheng J, Sun C. Prevalence and drug resistance characteristics of carbapenem-resistant Enterobacteriaceae in Hangzhou, China. Front Med. (2018) 12:182–8. doi: 10.1007/s11684-017-0529-4
- van Loon K, Voor In 't Holt AF, Vosa MC. A systematic review and meta-analyses of the clinical epidemiology of carbapenemresistant Enterobacteriaceae. *Antimicrob Agents Chemother*. (2018) 62:e01730–17. doi: 10.1128/AAC.01730-17
- Boutal H, Vogel A, Bernabeu S, Devilliers K, Creton E, Cotellon G, et al. A multiplex lateral flow immunoassay for the rapid identification of NDM-, KPC-, IMP- and VIM-type and OXA-48-like carbapenemaseproducing Enterobacteriaceae. J Antimicrob Chemother. (2018) 73:909– 15. doi: 10.1093/jac/dkx521
- Queenan AM, Bush K. Carbapenemases: the versatile β-lactamases. Clin Microbiol Rev. (2007) 20:440–58. doi: 10.1128/CMR.00001-07
- 10. Tzouvelekis LS, Markogiannakis A, Psichogiou M, Tassios PT, Daikos GL. Carbapenemases in *Klebsiella pneumoniae* and other Enterobacteriaceae: an

AUTHOR CONTRIBUTIONS

QY and YX conceived and designed the experiments. JZ, GZ, PJ, and YZ performed the experiments, analyzed the data, and wrote the paper. All authors approved the final version of the manuscript.

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SUPPLEMENTARY MATERIAL

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evolving crisis of global dimensions. Clin Microbiol Rev. (2012) 25:682-707. doi: 10.1128/CMR.05035-11

- Dortet L, Agathine A, Naas T, Cuzon G, Poirel L, Nordmann P. Evaluation of the RAPIDEC(R) CARBA NP, the Rapid CARB Screen(R) and the Carba NP test for biochemical detection of carbapenemaseproducing Enterobacteriaceae. J Antimicrob Chemother. (2015) 70:3014– 22. doi: 10.1093/jac/dkv213
- Ong CH, Ratnayake L, Ang MLT, Lin RTP, Chana DSG. Diagnostic accuracy of BD phoenix CPO detect for carbapenemase production in 190 Enterobacteriaceae isolates. J Clin Microbiol. (2018) 56:e01043– 18. doi: 10.1128/JCM.01043-18
- Bracco S, Migliavacca R, Pini B, Corbo E, Nucleo E, Brigante G, et al. Evaluation of Brilliance CRE Agar for the detection of carbapenem-resistant Gram-negative bacteria. *New Microbiol.* (2013) 36:181–6.
- Hinic V, Amrein I, Stammler S, Heckendorn J, Meinel D, Frei R, et al. Comparison of two rapid biochemical tests and four chromogenic selective media for detection of carbapenemase-producing Gram-negative bacteria. J Microbiol Methods. (2017) 135:66–8. doi: 10.1016/j.mimet.2017.01.012
- Park BY, Mourad D, Hong JS, Yoon EJ, Kim D, Lee H, et al. Performance evaluation of the newly developed BD phoenix NMIC-500 panel using clinical isolates of gram-negative bacilli. *Ann Lab Med.* (2019) 39:470– 7. doi: 10.3343/alm.2019.39.5.470
- Simon M, Gatermann S, Pfeifer Y, Reischl U, Gessner A, Jantsch J. Evaluation of the automated BD Phoenix CPO Detect panel in combination with the beta-CARBA assay for detection and classification of carbapenemase-producing Enterobacterales. J Microbiol Methods. (2019) 156:29–33. doi: 10.1016/j.mimet.2018.11.024
- CLSI. Performance Standards for Antimicrobial Susceptibility Testing: Thirty Informational Supplement M100-ED30. CLSI supplement M100. Wayne, PA: Clinical and Laboratory Standards Institute (2020).
- CLSI. Performance Standards for Antimicrobial Disk Susceptibility Tests. 13th ed. CLSI standard M02. Wayne, PA: Clinical and Laboratory Standards Institute (2018).
- Yigit H, Queenan AM, Anderson GJ, Domenech-Sanchez A, Biddle JW, Steward CD, et al. Novel carbapenem-hydrolyzing beta-lactamase, KPC-1, from a carbapenem-resistant strain of

Klebsiella pneumoniae. Antimicrob Agents Chemother. (2001) 45:1151-61. doi: 10.1128/AAC.45.4.1151-1161.2001

- Armand-Lefevre L, Leflon-Guibout V, Bredin J, Barguellil F, Amor A, Pages JM, et al. Imipenem resistance in *Salmonella enterica* serovar Wien related to porin loss and CMY-4 beta-lactamase production. *Antimicrob Agents Chemother*. (2003) 47:1165–8. doi: 10.1128/AAC.47.3.1165-1168.2003
- 21. CLSI. Verification of Commercial Microbial Identification and Antimicrobial Susceptibility Testing Systems. 1st ed. CLSI guideline M52. Wayne, PA: Clinical and Laboratory Standards Institute (2015).
- Humphries RM, Ambler J, Mitchell SL, Castanheira M, Dingle T, Hindler JA, et al. CLSI methods development and standardization working group best practices for evaluation of antimicrobial susceptibility tests. *J Clin Microbiol.* (2018) 56:e01934–17. doi: 10.1128/JCM.01934-17
- Hong JS, Kim D, Kang DY, Park BY, Yang S, Yoon EJ, et al. Evaluation of the BD phoenix M50 automated microbiology system for antimicrobial susceptibility testing with clinical isolates in Korea. *Microb Drug Resist.* (2019) 25:1142–8. doi: 10.1089/mdr.2018.0370
- Carroll KC, Borek AP, Burger C, Glanz B, Bhally H, Henciak S, et al. Evaluation of the BD Phoenix automated microbiology system for identification and antimicrobial susceptibility testing of staphylococci and enterococci. J Clin Microbiol. (2006) 44:2072–7. doi: 10.1128/JCM.02636-05
- Zhangel GG, Wiebe R, Dilay L, Thomson K, Rubinstein E, Haban DJ, et al. Comparative review of the carbapenems. *Drugs.* (2007) 67:1027– 52. doi: 10.2165/00003495-200767070-00006
- Snyder JW, Munier GK, Johnson CL. Direct comparison of the BD phoenix system with the microscan walkaway system for identification and antimicrobial susceptibility testing of Enterobacteriaceae and nonfermentative gram-negative organisms. J Clin Microbiol. (2008) 46:2327–33. doi: 10.1128/JCM.00075-08
- Haffler ZJ, Kulengowski B, Ribes JA, Burgess DS. Evaluation of the BD Phoenix automated system for determining antimicrobial susceptibility against carbapenem-resistant Enterobacteriaceae compared with broth microdilution. *Int J Antimicrob Agents*. (2019) 54:249–54. doi: 10.1016/j.ijantimicag.2019.05.002
- Hogan CA, Watz N, Budvytiene I, Banaei N. Rapid antimicrobial susceptibility testing by VITEK(R)2 directly from blood cultures in patients

with Gram-negative rod bacteremia. *Diagn Microbiol Infect Dis.* (2019) 94:116–21. doi: 10.1016/j.diagmicrobio.2019.01.001

- Saad Albichr I, Anantharajah A, Dodemont M, Hallin M, Verroken A, Rodriguez-Villalobos H. Evaluation of the automated BD Phoenix CPO Detect test for detection and classification of carbapenemases in Gram negatives. *Diagn Microbiol Infect Dis.* (2020) 96:114911. doi: 10.1016/j.diagmicrobio.2019.114911
- Zhang R, Liu L, Zhou H, Chan EW, Li J, Fang Y, et al. Nationwide surveillance of clinical carbapenem-resistant Enterobacteriaceae (CRE) strains in China. *EBioMedicine*. (2017) 19:98–106. doi: 10.1016/j.ebiom.2017.04.032
- Antonelli A, Coppi M, Camarlinghi G, Parisio EM, Nardone M, Riccobono E, et al. Variable performance of different commercial systems for testing carbapenem susceptibility of KPC carbapenemase-producing *Escherichia coli. Clin Microbiol Infect.* (2019) 25:1432.e1431-e1434. doi: 10.1016/j.cmi.2019.08.005
- 32. Menglan Zhou DW, Kudinha T, Yang Q, Yu S, Xu Y-C. Comparative evaluation of four phenotypic methods for detection of class A and B carbapenemase-producing Enterobacteriaceae in China. J Clin Microbiol. (2018) 56:e00395–18. doi: 10.1128/JCM.00395-18
- 33. Wei Q, Sun J, Wang Z, Yan L, Zhang C, Xu X. Evaluation of modified rapid carbapenem inactivation method (mrCIM) combined with rapid EDTAmodified carbapenem inactivation method (reCIM) to detect carbapenemase and distinguish metallo-carbapenemase in Enterobacteriaceae within four hours. *Infect Drug Resist.* (2020) 13:1919–27. doi: 10.2147/IDR.S249570

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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