



A Prospective Cohort Study on the Prevalent and Recurrent Tuberculosis Isolates Using the MIRU-VNTR Typing

Beibei Qiu^{1†}, Bilin Tao^{1†}, Qiao Liu^{1,2}, Zhongqi Li¹, Huan Song¹, Dan Tian¹, Jizhou Wu¹, Zhuchao Wu¹, Mengyao Zhan¹, Wei Lu^{2*} and Jianming Wang^{1*}

¹ Department of Epidemiology, Center for Global Health, School of Public Health, Nanjing Medical University, Nanjing, China, ² Department of Chronic Communicable Disease, Center for Disease Control and Prevention of Jiangsu Province, Nanjing, China

OPEN ACCESS

Edited by:

Qinning Wang, New South Wales Health Pathology, Australia

Reviewed by:

Hami Kaboosi, Islamic Azad University of Ayatollah Amoli, Iran Ulziijargal Gurjav, Mongolian National University of Medicine, Mongolia

*Correspondence:

Wei Lu jsjkmck@163.com Jianming Wang jmwang@njmu.edu.cn

[†]These authors have contributed equally to this work and share first authorship

Specialty section:

This article was submitted to Infectious Diseases – Surveillance, Prevention and Treatment, a section of the journal Frontiers in Medicine

Received: 30 March 2021 Accepted: 23 August 2021 Published: 14 September 2021

Citation:

Qiu B, Tao B, Liu Q, Li Z, Song H, Tian D, Wu J, Wu Z, Zhan M, Lu W and Wang J (2021) A Prospective Cohort Study on the Prevalent and Recurrent Tuberculosis Isolates Using the MIRU-VNTR Typing. Front. Med. 8:685368. doi: 10.3389/fmed.2021.685368

The study aims to describe the clustering characteristics of Mycobacterium tuberculosis (M.tb) strains circulating in eastern China and determine the ratio of relapse and reinfection in recurrent patients. We recruited sputum smear-positive pulmonary tuberculosis cases from five cities of Jiangsu Province, China, during August 2013 and December 2015. Patients were followed for the treatment outcomes and recurrence based on a cohort design. *M.tb* strains were isolated and genotyped using the 12-locus MIRU-VNTR. The Beijing family was identified by the extended Region of Difference (RD) analysis. The Hunter-Gaston Discriminatory Index (HGDI) was used to judge the resolution ability of MIRU-VNTR. The odds ratio (OR) together with 95% confidence interval (CI) were used to estimate the strength of association. We performed a cluster analysis on 2098 M.tb isolates and classified them into 545 genotypes and five categories (I, 0.19%; II, 0.43%; III, 3.34%; IV, 77.46%; V, 18.59%). After adjusting for potential confounders, the Beijing family genotype (OR = 118.63, 95% CI: 79.61-176.79, P = 0.001) was significantly related to the dominant strain infections. Patients infected with non-dominant strains had a higher risk of the pulmonary cavity (OR = 1.39, 95% CI: 1.01–1.91, P = 0.046). Among 37 paired recurrent cases, 22 (59.46%) were determined as endogenous reactivation, and 15 (40.54%) were exogenous reinfection. The type of *M.tb* strains prevalent in Jiangsu Province is relatively single. Beijing family strains infection is dominant in local tuberculosis cases. Endogenous reactivation appears to be a major cause of recurrent tuberculosis in Eastern China. This finding emphasizes the importance of case follow-up and monitoring after the completion of antituberculosis treatment.

Keywords: tuberculosis, molecular epidemiology, genotyping, recurrence, reactivation, reinfection, Beijing family

INTRODUCTION

Tuberculosis (TB) is a common chronic infectious and consumptive disease that damages the organs of the whole body. It has been ranked as one of the top 10 causes of death and the greatest killer of a single infectious disease in the world (1). According to the 2020 Global Tuberculosis Report (https://www.who.int/), an estimated 10.0 million (range 9.0–11.1 million) people fell ill

1

with TB in 2019, a number that has been relatively stable in recent years. Thirty high TB burden countries, including India, China, Indonesia, Philippines, and other countries, account for 87% of the global cases.

Patients with recurrent TB are less likely to complete treatment and suffer higher fatality than those with the first episode of TB (2). Recurrence of TB can be due to either relapse of an initial infection or exogenous reinfection with new *Mycobacterium tuberculosis* (*M.tb*) strains (3). The causes of such two types of recurrence are different (4). Poor treatment adherence (5), positive sputum smear, pulmonary cavity (6), drug resistance (7), old age (8), or Beijing family *M.tb* strains (9) are the risk factors of endogenous reactivation, while human immunodeficiency virus (HIV) infection (10) and diabetes (11) can increase the risk of exogenous reinfection. Because TB recurrence is usually associated with drug resistance, it is reasonable to distinguish the types of TB recurrence and to understand the mechanism of endogenous reactivation and exogenous reinfection.

With the development of molecular biological techniques, we can genotype the strains of *M.tb* that broke out circulating in a particular area to analyze the epidemiological characteristics of the pathogen and clarify factors related to the transmission. Molecular genotyping methods, such as IS6110 restriction fragment length polymorphism (IS6110-RFLP), spacer typing oligonucleotide (Spoligo-typing), mycobacterial interspersed repetitive units (MIRU), and the variable number of tandem repeats (VNTR), have been applied to genotype TB isolates (12). TB genotyping helps to identify the outbreak and epidemic process, trace the source of infection, establish a phylogenetic relationship and identify patients whose disease is the result of reactivation of an infection that was acquired in the past or involved in the same chain of recent transmission (13). MIRU-VNTR is a PCR-based typing method that calculates the size and repeated number of units in each locus by amplifying mycobacterial interspersed repetitive units (12). Various combinations of MIRU-VNTR loci have been used to give better differentiation of *M.tb* strains. The typical combinations include 12, 15, and 24-loci. The 15 and 24-loci of MIRU-VNTR include the previous 12 loci, together with several additional high-resolution loci (14). However, the allelic diversity of loci can vary between areas or between *M.tb* complex species (15).

Jiangsu is an area with a higher population density and a province with higher TB mobility in Eastern China. To describe the molecular typing characteristics and epidemic trends of *M.tb* is of great significance for the targeted disease prevention and control strategies. In the current study, we selected five cities in Jiangsu Province as the study sites, continuously recruited TB cases as study subjects, and genotyped clinical isolates of *M.tb* using the MIRU-VNTR. TB patients were followed to record the treatment outcomes and monitor the recurrence. This study had two purposes. Firstly, we attempted to analyze the clustering characteristics of TB isolates and related factors. Secondly, we aimed to determine the ratio of relapse and reinfection in recurrent patients and explore the molecular types of recurrent infections.

MATERIALS AND METHODS

Patients

We selected Lianyungang, Xuzhou, Nantong, Changzhou, and Taizhou in Jiangsu Province, China, as the study sites. Patients diagnosed during August 2013 and December 2015 were recruited as study subjects and followed to record the disease's recurrence. We defined the recurrence as a TB episode occurring within the study period after treatment completion or cure of a previous episode. The interval of two disease episodes should be longer than 1 year. Newly diagnosed sputum smear-positive TB cases were included. Extrapulmonary TB or HIV-positive cases were excluded.

Samples Collection

Sputum smear microscopy test, culture, and strain identification were performed following the guideline of the Tuberculosis Diagnosis Laboratory Test Regulations of China Tuberculosis Association. *M.tb* strains were harvested from the Lowenstein-Jensen medium and preserved in the freezer at -80° C. The DNA of *M.tb* strains was extracted by a modified CTAB (hexadecyltrimethylammonium bromide) method. In brief, the colony of the 10 ul quantitative inoculation ring was scraped in the biosafety cabinet, suspended in the EP tube equipped with 200 ul sterilized TE, and inactivated in a water bath at 80° C for 30 min. The inactivated bacteria were boiled at 100° C for 10 min and centrifuged with 12,000 r/min for 15 min. The DNA template was obtained by absorbing the supernatant and stored at -20° C.

Drug Sensitivity Test

A drug sensitivity test was carried out by using the proportional method (16, 17). Samples to be tested were diluted to 10^{-2} and 10^{-4} g/L bacterial suspensions. We dipped 1 ring (0.01 ml) of the bacterial suspensions with a 22 standard wire gauge (SWG) quantitative inoculation ring and inoculated them on the surface of the control and drug-containing medium. The bacterial liquid was dispersed on the slope of the culture medium as evenly as possible, and the final inoculation amount was 10^{-4} mg and 10^{-6} mg, respectively. After inoculation, the culture medium was placed at 37° C, and the number of colonies was read after 4 weeks to calculate the resistance percentage, where $\leq 1\%$ was judged as sensitive, and >1% was considered resistant.

Beijing Family Identification

The Beijing family isolates were identified by extended genotyping Region of Difference (RD) analysis. The detection of the genomic deletions RD105 was done by PCR using primers previously described (18). If the strain had the RD105 genomic deletion, it was defined as the Beijing family strain.

MIRU-VNTR Genotyping

Although there are various combinations of MIRU-VNTR loci, no specific set of loci have been agreed upon as a standard. Considering the time-consuming and economic cost, some researchers recommended using 12-loci MIRU-VNTR. Besides, the resolution of 12-loci MIRU-VNTR is relatively high, and there may be over-clustering by 15- or 24-loci MIRU-VNTR for Beijing strains (19). Thus, in this study, we applied the



12-loci MIRU-VNTR to genotype the strains isolated from recurrent patients. Each MIRU locus was amplified individually with primers specific for sequences flanking the MIRU units (**Supplementary Table S1**). In the current study, we used a set of 12 VNTR loci (20, 21). The PCR reaction mixture for all loci was listed in **Supplementary Table S2**. The amplification profile consisted of pre-denaturation at 94°C for 5 min, followed by 25 cycles of 94°C for 45 s, 60°C for 50 s, and 72°C for 60 s, and 72°C extensions for 10 min. We used the H37Rv strain as a standard control to determine the size of the amplified fragment.

Statistical Analysis

Cluster analyses were performed using R software for Windows version 3.5.3 (https://www.r-project.org/) with "hclust," "phangorn," "ape," and "tidyverse" package. We used the MIRU-VNTRplus web application (https://www.miru-vntrplus. org) to compare the strains with the reference strains. We used iTOLs (https://itol.embl.de/) to modify and style the phylogenetic tree. The Hunter-Gaston Discriminatory Index (HGDI) (22) was used to judge the resolution ability of MIRU-VNTR. $HGDI = 1 - \frac{1}{N(N-1)} \sum_{j=1}^{s} nj (nj-1)$. N stands for the total number of strains, nj is the number of strains with the *j*th genotype, and s is the number of different genotypes at the MIRU-VNTR loci. HGDI ≥0.60 was judged as a high polymorphism, 0.30 \leq HGDI <0.60 showed a medium polymorphism, and HGDI $<\!0.30$ indicated a low polymorphism. We applied a logistic regression model to explore factors related to dominant and non-dominant isolates and used the Fisher exact test to compare the characteristics of recurrent types using SPSS software (version 25.0, IBM Corporation, Armonk, NY). A two-tailed *P*-value of <0.05 was regarded as statistically significant.

RESULTS

Data Collection

We recruited 2,098 TB cases from August 2013 to December 2015. After the standard antituberculosis therapy, 1,897 patients were treated successfully (including cured and completed treatment), the other 201 patients were including death, treatment failure, adverse reactions, refuse treatment, change treatment regime, MDR-TB treatment, and others. After follow-up on cases that were cured and completed treatment, we observed 141 recurrent cases. After excluding contaminated isolates, incomplete retention of primary and recurrent isolates, and samples with failed DNA extraction, 37 pairs of recurrent cases were used for MIRU-VNTR genotyping comparison (**Figure 1**).

Cluster Analysis of All Isolates

We performed a cluster analysis on 2,098 isolates to observe the classification tree and clustering (**Figure 2**). Red represents the branches of the phylogenetic tree, and green means the



FIGURE 2 | Clustering tree of 2,098 tuberculosis isolates. Red represents the branches of the phylogenetic tree, and green means the ID of the strains.

ID of the strains. All isolates could be divided into 545 genotypes, including 156 clustered genotypes. Three hundred eighty-nine isolates had independent genotypes. Among 1,709 clustered isolates, the largest cluster had 586 isolates (27.93%). We further divided 2,098 isolates into five categories (I, 4, 0.19%; II, 9, 0.43%; III, 70, 3.34%; IV, 1,625, 77.46%; V, 390, 18.59%). We defined the strains which belonged to a category that exceeded 70% of the total number of strains as the dominant strain. Infection with dominant strains was related to Beijing family genotype (OR = 115.95, 95% CI: 78.21-171.91, P = 0.001) and drinking (OR = 0.67, 95% CI: 0.53-0.86, P = 0.002). After adjusting for potential confounders, such as sex, age, smoking, drinking, Beijing family genotype, contact history, MDR-TB, treatment outcomes, and pulmonary cavity, Beijing family genotype (OR = 118.63, 95% CI: 79.61-176.79, P = 0.001) remained significant. Patients with the pulmonary cavity (OR = 1.39, 95% CI: 1.01–1.91, P = 0.046) had a higher risk of infection with non-dominant strains (Table 1).

For 12 sites used for genotyping, MIRU 26 had a high polymorphism (HGDI = 0.6262), MIRU 10, 31, 39, and 40 had a moderate polymorphism ($0.30 \le \text{HGDI} < 0.60$), while MIRU 2, 4, 16, 20, 23, 24, and 27 had a low polymorphism (HGDI < 0.30) (**Table 2**).

Cluster Analysis of Recurrent Cases

To explore the proportion of relapse and reinfection among recurrent cases, we compared the MIRU-VNTR genotypes of 37 paired isolates. The copy number and HGDI index were shown in **Table 3**. We have also listed the total copy numbers of 37 paired recurrent strains and Beijing family genotypes obtained by the RD105 test in **Supplementary Table S3**. Endogenous or reactivation paired cases have the same lineages. The polymorphism of MIRU 10, 26, 31, and 39 kept at a moderate level ($0.30 \le \text{HGDI} < 0.60$), while MIRU 2, 4, 16, 20, 23, 24, 27 and 40 had low polymorphisms (HGDI < 0.30). The isolates with similar characteristics were grouped into one genotype by cluster analysis (**Figure 3**). Red and blue labels represent reinfection and reactivation, respectively.

Among 37 paired recurrent cases, 22 (59.46%) were regarded as endogenous reactivation, and 15 (40.54%) were exogenous reinfection. In the endogenous relapse, males (86.36%) and the elderly (68.18%) accounted for a higher proportion than exogenous reinfection. The proportion of tobacco smoking history was also higher in patients with endogenous relapse (50.00%) than in patients with exogenous reinfection (26.67%). Patients with pulmonary cavities were more likely to suffer from exogenous reinfection (66.67%) than endogenous relapse (45.45%). Recurrent cases with rifampicin



resistance (1 case) and isoniazid resistance (2 cases) were attributed to endogenous relapse. The comparison of the characteristics between endogenous reactivation and exogenous reinfection by Fisher's exact test were shown in **Table 4**. The *P*-value of these factors was >0.05 with no significant difference.

DISCUSSION

In this molecular epidemiological study, we genotyped 2,098 strains circulating in Eastern China. We observed that the types of prevalent *M.tb* strains were relatively single, with the dominant strain of the Beijing family. Endogenous reactivation appeared to be a major cause of recurrent TB, emphasizing the importance of case management and long-term follow-up. Although MIRU-VNTR typing has been widely used to examine the genetic diversity and transmission of *M.tb* strains, this study has a relatively large sample size and provided robust statistical power.

We categorized M.tb strains into five groups, and the dominant isolates were category IV, accounting for 77.46% of all isolates. We observed that the RD105 deletion (Beijing family) contributed to the predominant infection, which was consistent with findings by Larry D. Teeter (23) and Carlo Garzelli (24). In addition, we found a significant association

between non-dominant strains and pulmonary cavitation, which was consistent with the report by Lazzarini (25). In China, although the main lineage of M.tb is the Beijing family, non-dominant strains mainly characterized by the LAM family still keeps an important composition (26). Liu et al. reported that the Beijing family was the dominant genotype (83.3%) in Beijing and linked to the recent spread of TB (27). The Beijing family strain was also found to be the most common genotype in Shanxi (81.5%) and Yunnan (70.5%), which was related to an increased risk of MDR-TB (28, 29). In Xinjiang, the Beijing lineage strains accounted for 57.48% of total M.tb isolates (30).

We observed that the recurrence rate of TB was 7.43%, which was higher than the proportion reported by Xie (6.5%) (31) and slightly lower than that reported by Ahmad (32) (8.0%). Other studies in China showed that the recurrence rate was 7.6% (33) or 6.9% (34). A prospective study and paired analysis emphasized that endogenous reactivation (62.16%) was the leading cause of recurrent TB in Eastern China, which was lower than that in India (35) (93%) and Australia (36) (87%). Endogenous reactivation mainly occurred in high-burden countries and was related to Beijing family genotype (9), infection with drug-resistant TB (37), low socioeconomic status, and foreign birth (38). Previous

Risk factors		N	Domina	Dominant strains		ominant strains	Univariate analy	vsis	Multivariate analysis	
			n	%	n	%	cOR (95% CI)	P-value	aOR (95% CI)	P-value
Sex	Male	1,631	1,252	77.05	379	80.13	0.83 (0.65–1.07)	0.157	1.00 (0.66–1.53)	0.996
	Female	467	373	22.95	94	19.87	Reference		Reference	
Age (years)	<=25	316	252	15.51	64	13.53	Reference		Reference	
	26–54	685	534	32.86	151	31.92	0.90 (0.65-1.25)	0.522	1.10 (0.67–1.80)	0.722
	≥55	1,097	839	51.63	258	54.55	0.83 (0.61-1.12)	0.223	0.98 (0.61–1.57)	0.931
Smoking	Yes	831	634	39.02	197	41.65	0.90 (0.73-1.10)	0.303	1.23 (0.85–1.77)	0.280
	No	1,267	991	60.98	276	58.35	Reference		Reference	
Drinking	Yes	389	278	17.11	111	23.47	0.67 (0.53–0.86)	0.002	0.72 (0.47-1.10)	0.126
	No	1,709	1,347	82.89	362	76.53	Reference		Reference	
Beijing family	Yes	1,727	1,591	97.91	136	28.75	115.95 (78.21–171.91)	0.001	118.63 (79.61–176.79)	0.001
	No	371	34	2.09	337	71.25	Reference		Reference	
Contact history	Yes	159	118	7.26	41	8.67	0.83 (0.57-1.20)	0.310	0.93 (0.52-1.66)	0.801
	No	1,939	1,507	92.74	432	91.33	Reference		Reference	
MDR-TB	Yes	87	73	4.49	14	2.96	1.54 (0.86–2.76)	0.144	1.16 (0.48–2.79)	0.742
	No	2,011	1,552	95.51	459	97.04	Reference		Reference	
Recurrence	No	1,756	1,358	83.57	398	84.14	Reference		Reference	
	Yes	141	114	7.02	27	5.71	1.24 (0.80-1.91)	0.336	0.97 (0.51-1.82)	0.920
	Others	201	153	9.41	48	10.15	0.93 (0.66-1.32)	0.697	0.82 (0.47-1.45)	0.499
Pulmonary cavity	Yes	987	751	46.22	236	49.89	Reference		Reference	
	No	1,111	874	53.78	237	50.11	1.16 (0.94–1.42)	0.158	1.39 (1.01–1.91)	0.046

Dominant strains are defined as a category that exceeds 70% of the total number of strains. MIRU-VNTR, Mycobacterial Interspersed Repetitive Units – Variable Number of Tandem Repeats; MDR-TB, Multi-drug resistant tuberculosis; cOR, Crude odds ratio; aOR, Adjusted odds ratio; CI, Confidence interval.

Index	MIRU-VNTR locus													
	2	4	10	16	20	23	24	26	27	31	39	40		
Copy number														
0														
1	9	59	52	15	37	3	2,092	48	7	3	15	89		
2	2,073	1,836	497	151	2,054	6	6	10	227	32	414	264		
3	16	27	1,466	1,890	7	5		62	1,836	328	1,617	1,619		
4		122	62	42		18		86	27	192	50	114		
5		40	16			2,035		305	1	1,460	2	12		
6		4	1			25		230		74				
7		3				5		1,215		7				
8		3	1			1		96		2				
9		3	3					39						
10		1						7						
HGDI	0.0236	0.2296	0.4543	0.1829	0.0412	0.0589	0.0057	0.6262	0.2224	0.4816	0.3666	0.3841		

TABLE 2 | Resolution of each polymorphic site for 2,098 isolates.

MIRU-VNTR, Mycobacterial Interspersed Repetitive Units-Variable Number of Tandem Repeats; HGDI, Hunter-Gaston, Discriminatory Index.

TABLE 3 | Resolution of each polymorphic site for 37 paired cases.

Index	MIRU-VNTR locus												
	2	4	10	16	20	23	24	26	27	31	39	40	
Copy number													
0					1			1	1				
1		1	2	2			74			2			
2	74	66	15	4	73				6	1	14	4	
3		1	56	68		1		3	66	8	59	65	
4		4	1					3	1	3	1	5	
5		2				71		9		55			
6								11		5			
7						2		46					
8								1					
HGDI	0	0.2033	0.3906	0.1540	0.0270	0.0796	0	0.5809	0.2002	0.4317	0.3328	0.2240	

MIRU-VNTR, Mycobacterial Interspersed Repetitive Units-Variable Number of Tandem Repeats; HGDI, Hunter-Gaston, Discriminatory Index.

studies have shown that most recurrent TB cases in HIV uninfected patients were due to endogenous reactivation, while most TB recurrence in HIV-infected patients was exogenous reinfection (39, 40). In our study, we found that the proportion of males and patients with tobacco smoking history were at a higher risk of endogenous reactivation. Patients with MDR, rifampicin resistance, and isoniazid resistance were related to an increased risk of endogenous reactivation, indicating a potential correlation with antituberculosis drug resistance (37, 41).

At present, a variety of genotyping methods have been developed, such as IS6110-RFLP, Spoligotyping, MIRU-VNTR, and WGS. Although the gold standard for TB genotyping is IS6110-RFLP, it is labor-intensive, and the interpretation of data from this method can be susceptible to errors (42). Although the 15- and 24-loci MIRU-VNTR has a greater

discriminatory power (42, 43), the 12-loci MIRU-VNTR can trace the recent outbreak and identify dominant isolates, requiring less time and labor (19, 44). To distinguish the endogenous reactivation and exogenous reinfection, 12-loci MIRU-VNTR is an alternative methodological choice with easy-to-apply, costeffective and good performance. Comparing the consistency of strains isolated from paired recurrent cases can clarify the type of recurrence and take targeted clinical intervention measures (44, 45).

In this study, we used the RD105 deletion to identify the Beijing family strains. There were 1,727 (82.32%) strains being judged as Lineage 2 Beijing genotype. Previous studies have assumed that Beijing family strains were more likely to develop into MDR-TB (46). Moreover, men and older adults were also observed to be more susceptible to Beijing genotype infection (27). Using MIRU-VNTR and detection of the deletion of RD105

TABLE 4 Comparison of the characteristics between endogenous reactivation and exogenous reinfection.

Factors		Ν	Endoge	enous reactivation	Exoge	P-value ³	
			n	%	n	%	
Sex	Male	31	19	86.36	12	80.00	0.670
	Female	6	3	13.64	3	20.00	
Age (years)	<=25	4	2	9.09	2	13.33	0.611
	26–54	10	5	22.73	5	33.33	
	≥55	23	15	68.18	8	53.33	
Contact history	Yes	2	1	4.55	1	0.00	1.000
	No	35	21	95.45	14	100.00	
Smoking	Yes	15	11	50.00	4	26.67	0.190
	No	22	11	50.00	11	73.33	
Drinking	Yes	7	5	22.73	2	13.33	0.677
	No	30	17	77.27	13	86.67	
Pulmonary cavity	Yes	20	10	45.45	10	66.67	0.315
	No	17	12	54.55	5	33.33	
Beijing family	Yes	33	19	86.36	14	93.33	0.633
	No	4	3	13.64	1	6.67	
Rifampicin resistance	Yes	1	1	4.55	0	0.00	1.000
	No	36	21	95.45	15	100.00	
Isoniazid resistance	Yes	2	2	9.09	0	0.00	0.505
	No	35	20	90.91	15	100.00	

*Fisher's exact test.

synthetically provides a simple, fast, and effective method for identification of *M.tb* genotyping in China (47). The Beijing family genotype obtained by the RD105 test is also mutually confirmed with the reactivation cases by 12-loci MIRU-VNTR.

Several limitations should not be ignored to explain our findings. Firstly, MIRU-VNTR genotyping is usually done by amplifying a panel of 12, 15, or 24 loci. Although the 12 loci method has a low workload and was proved to have good discrimination in the Chinese population, some scientists still recommend using the standard MIRU-15 and MIRU-24, which have considerably increased discriminatory power. Secondly, we used the MIRU-VNTR to determine the type of recurrence. However, the microevolution and mutation of the strain may lead to misjudgment of exogenous reinfection and endogenous reactivation. In the next step, it is necessary to sequence the strains to identify the common mutation sites and accurately distinguish the recurrence type.

CONCLUSION

The genotypes of M.tb strains prevalent in Eastern China are relatively single. Beijing family strain was the dominant strain. Endogenous reactivation was the leading cause of recurrent TB. Endogenous reactivation appeared to be a major cause of recurrent TB in Eastern China. This finding emphasizes the importance of case follow-up and monitoring after the completion of antituberculosis treatment.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding author/s.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Ethics Committee of Nanjing Medical University. Written informed consent to participate in this study was provided by the participants' legal guardian/next of kin.

AUTHOR CONTRIBUTIONS

BQ and BT: conceptualization, methodology, and software. JW: validation, writing—review and editing, and funding acquisition. BQ, BT, and QL: formal analysis and visualization. QL, ZL, HS, DT, JW, ZW, and MZ: investigation and data curation. QL and WL: resources. BQ, BT, QL, ZL, HS, DT, JW, ZW, MZ, and WL: writing—original draft preparation. WL and JW: supervision and project administration. All authors have given final approval of the version to be published and agreed on the journal to which the article has been submitted.

FUNDING

This study was funded by the National Natural Science Foundation of China (81973103), National Key R&D Program of China (2017YFC0907000), Medical Research Project of Jiangsu Health Commission (ZDB2020013), and Priority Academic Program Development of Jiangsu Higher Education Institutions (PAPD). The funding agencies had no role in the study design, data collection, analysis, decision to publish, or preparation of the manuscript.

REFERENCES

- Pai M, Behr MA, Dowdy D, Dheda K, Divangahi M, Boehme CC, et al. Tuberculosis. Nat Rev Dis Primers. (2016) 2:16076. doi: 10.1038/nrdp.2016.76
- Mutembo S, Mutanga JN, Musokotwane K, Kanene C, Dobbin K, Yao X, et al. Urban-rural disparities in treatment outcomes among recurrent TB cases in Southern Province, Zambia. *BMC Infect Dis.* (2019) 19:1087. doi: 10.1186/s12879-019-4709-5
- van Rie A, Warren R, Richardson M, Victor TC, Gie RP, Enarson DA, et al. Exogenous reinfection as a cause of recurrent tuberculosis after curative treatment. N Engl J Med. (1999) 341:1174–9. doi: 10.1056/NEJM199910143411602
- Zong Z, Huo F, Shi J, Jing W, Ma Y, Liang Q, et al. Relapse versus reinfection of recurrent tuberculosis patients in a national tuberculosis specialized hospital in Beijing, China. *Front Microbiol.* (2018) 9:1858. doi: 10.3389/fmicb.2018.01858
- Alipanah N, Jarlsberg L, Miller C, Linh NN, Falzon D, Jaramillo E, et al. Adherence interventions and outcomes of tuberculosis treatment: a systematic review and meta-analysis of trials and observational studies. *PLoS Med.* (2018) 15:e1002595. doi: 10.1371/journal.pmed.1002595
- Casha AR, Camilleri L, Manche A, Gatt R, Attard D, Wolak W, et al. A hypothesis for reactivation of pulmonary tuberculosis: how thoracic wall shape affects the epidemiology of tuberculosis. *Clin Anat.* (2015) 28:614–20. doi: 10.1002/ca.22544
- Bastos ML, Cosme LB, Fregona G, do Prado TN, Bertolde AI, Zandonade E, et al. Treatment outcomes of MDR-tuberculosis patients in Brazil: a retrospective cohort analysis. *BMC Infect Dis.* (2017) 17:718. doi: 10.1186/s12879-017-2810-1
- Wu X, Yang J, Tan G, Liu H, Liu Y, Guo Y, et al. Drug resistance characteristics of *Mycobacterium tuberculosis* isolates from patients with tuberculosis to 12 antituberculous drugs in China. *Front Cell Infect Microbiol.* (2019) 9:345. doi: 10.3389/fcimb.2019.00345
- Gao M, Gao J, Du J, Liu Y, Zhang Y, Ma L, et al. Efficacy of ultra-short course chemotherapy for new smear positive drug susceptible pulmonary tuberculosis: study protocol of a multicenter randomized controlled clinical trial. *BMC Infect Dis.* (2017) 17:435. doi: 10.1186/s12879-017-2505-7
- McIvor A, Koornhof H, Kana BD. Relapse, re-infection and mixed infections in tuberculosis disease. *Pathog Dis.* (2017) 75:1–16. doi: 10.1093/femspd/ftx020
- Sola E, Rivera C, Mangual M, Martinez J, Rivera K, Fernandez R. Diabetes mellitus: an important risk factor for reactivation of tuberculosis. *Endocrinol Diabetes Metab Case Rep.* (2016) 2016:1–4. doi: 10.1530/EDM-16-0035
- Xu G, Mao X, Wang J, Pan H. Clustering and recent transmission of Mycobacterium tuberculosis in a Chinese population. Infect Drug Resist. (2018) 11:323–30. doi: 10.2147/IDR.S156534
- Mears J, Abubakar I, Cohen T, McHugh TD, Sonnenberg P. Effect of study design and setting on tuberculosis clustering estimates using Mycobacterial Interspersed Repetitive Units-Variable Number Tandem Repeats (MIRU-VNTR): a systematic review. *BMJ Open.* (2015) 5:e005636. doi: 10.1136/bmjopen-2014-005636
- Ali A, Hasan Z, Tanveer M, Siddiqui AR, Ghebremichael S, Kallenius G, et al. Characterization of *Mycobacterium tuberculosis* Central Asian Strain 1 using mycobacterial interspersed repetitive unit genotyping. *BMC Microbiol.* (2007) 7:76. doi: 10.1186/1471-2180-7-76
- Yang L, Wang C, Wang H, Meng Q, Wang Q. Evaluation of MIRU-VNTR for typing of *Mycobacterium bovis* isolated from Sika deer in Northeast China. *BMC Vet Res.* (2015) 11:93. doi: 10.1186/s12917-015-0402-0
- 16. Faksri K, Kaewprasert O, Ong RT, Suriyaphol P, Prammananan T, Teo YY, et al. Comparisons of whole-genome sequencing and phenotypic

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmed. 2021.685368/full#supplementary-material

drug susceptibility testing for *Mycobacterium tuberculosis* causing MDR-TB and XDR-TB in Thailand. *Int J Antimicrob Agents*. (2019) 54:109–16. doi: 10.1016/j.ijantimicag.2019.04.004

- Maurya AK, Singh AK, Kumar M, Umrao J, Kant S, Nag VL, et al. Changing patterns and trends of multidrug-resistant tuberculosis at referral centre in Northern India: a 4-year experience. *Indian J Med Microbiol.* (2013) 31:40–6. doi: 10.4103/0255-0857.108720
- Hu Y, Mathema B, Zhao Q, Zheng X, Li D, Jiang W, et al. Comparison of the socio-demographic and clinical features of pulmonary TB patients infected with sub-lineages within the W-Beijing and non-Beijing *Mycobacterium tuberculosis*. *Tuberculosis*. (2016) 97:18–25. doi: 10.1016/j.tube.2015.11.007
- Zheng C, Zhao YD, Zhu GQ, Li S, Sun HH, Feng Q, et al. Suitability of IS6110-RFLP and MIRU-VNTR for differentiating spoligotyped drugresistant *Mycobacterium tuberculosis* clinical isolates from Sichuan in China. *Biomed Res Int.* (2014) 2014. doi: 10.1155/2014/763204
- You NN, Zhu LM, Li GL, Martinez L, Lu W, Liu Q, et al. A tuberculosis school outbreak in China, 2018: reaching an often overlooked adolescent population. *Epidemiol Infect.* (2019) 147:e303. doi: 10.1017/S0950268819001882
- 21. Armas F, Camperio C, Coltella L, Selvaggini S, Boniotti MB, Pacciarini ML, et al. Comparison of semi-automated commercial rep-PCR fingerprinting, spoligotyping, 12-locus MIRU-VNTR typing and single nucleotide polymorphism analysis of the embB gene as molecular typing tools for *Mycobacterium bovis. J Med Microbiol.* (2017) 66:1151–7. doi: 10.1099/jmm.0.000536
- Hunter PR, Gaston MA. Numerical index of the discriminatory ability of typing systems: an application of Simpson's index of diversity. *J Clin Microbiol.* (1988) 26:2465–6. doi: 10.1128/jcm.26.11.2465-2466.1988
- 23. Teeter LD, Vempaty P, Nguyen DT, Tapia J, Sharnprapai S, Ghosh S, et al. Validation of genotype cluster investigations for *Mycobacterium tuberculosis*: application results for 44 clusters from four heterogeneous United States jurisdictions. *BMC Infect Dis.* (2016) 16:594. doi: 10.1186/s12879-016-1937-9
- Garzelli C, Lari N, Rindi L. Genomic diversity of *Mycobacterium tuberculosis* Beijing strains isolated in Tuscany, Italy, based on large sequence deletions, SNPs in putative DNA repair genes and MIRU-VNTR polymorphisms. *Tuberculosis*. (2016) 97:147–53. doi: 10.1016/j.tube.2015.10.008
- Lazzarini LC, Spindola SM, Bang H, Gibson AL, Weisenberg S, da Silva Carvalho W, et al. RDRio *Mycobacterium tuberculosis* infection is associated with a higher frequency of cavitary pulmonary disease. *J Clin Microbiol.* (2008) 46:2175–83. doi: 10.1128/JCM.00065-08
- Mokrousov I, Jiao WW, Wan K, Shen A. Stranger in a strange land: Ibero-American strain of *Mycobacterium tuberculosis* in Tibet, China. *Infect Genet Evol.* (2014) 26:323–6. doi: 10.1016/j.meegid.2014.06.011
- 27. Liu Y, Zhang X, Zhang Y, Sun Y, Yao C, Wang W, et al. Characterization of *Mycobacterium tuberculosis* strains in Beijing, China: drug susceptibility phenotypes and Beijing genotype family transmission. *BMC Infect Dis.* (2018) 18:658. doi: 10.1186/s12879-018-3578-7
- Li Y, Pang Y, Zhang T, Xian X, Yang J, Wang R, et al. Genotypes of Mycobacterium tuberculosis isolates circulating in Shaanxi Province, China. PLoS ONE. (2020) 15:e0242971. doi: 10.1371/journal.pone.0242971
- Li D, Song Y, Yang P, Li X, Zhang AM, Xia X. Genetic diversity and drug resistance of *Mycobacterium tuberculosis* in Yunnan, China. J Clin Lab Anal. (2019) 33:e22884. doi: 10.1002/jcla.22884
- Yuan L, Mi L, Li Y, Zhang H, Zheng F, Li Z. Genotypic characteristics of Mycobacterium tuberculosis circulating in Xinjiang, China. Infect Dis. (2016) 48:108–15. doi: 10.3109/23744235.2015.1087649
- Xie Z, Wang T, Chen H, Wang D, Gao X, Hui Y. Factors associated with diagnostic delay in recurrent TB. *BMC Public Health*. (2020) 20:1207. doi: 10.1186/s12889-020-09005-9

- 32. Collaborative Group for the Meta-Analysis of Individual Patient Data in MDRTBt, Ahmad N, Ahuja SD, Akkerman OW, Alffenaar JC, Anderson LF, et al. Treatment correlates of successful outcomes in pulmonary multidrugresistant tuberculosis: an individual patient data meta-analysis. *Lancet*. (2018) 392:821–34. doi: 10.1016/S0140-6736(18)31644-1
- 33. Chen Q, Peng L, Xiong G, Peng Y, Luo D, Zou L, et al. recurrence is a noticeable cause of rifampicin-resistant *Mycobacterium tuberculosis* in the elderly population in Jiangxi, China. *Front Public Health.* (2019) 7:182. doi: 10.3389/fpubh.2019.00182
- 34. Du J, Zhang L, Ma Y, Chen XY, Ge QP, Tian XZ, et al. Treatment and recurrence on re-treatment tuberculosis patients: a randomized clinical trial and 7-year perspective cohort study in China. Eur J Clin Microbiol Infect Dis. (2020) 39:93–101. doi: 10.1007/s10096-019-03696-8
- 35. Velayutham B, Chadha VK, Singla N, Narang P, Gangadhar Rao V, Nair S, et al. Recurrence of tuberculosis among newly diagnosed sputum positive pulmonary tuberculosis patients treated under the Revised National Tuberculosis Control Programme, India: a multi-centric prospective study. *PLoS ONE.* (2018) 13:e0200150. doi: 10.1371/journal.pone.02 00150
- Parvaresh L, Crighton T, Martinez E, Bustamante A, Chen S, Sintchenko V. Recurrence of tuberculosis in a low-incidence setting: a retrospective crosssectional study augmented by whole genome sequencing. *BMC Infect Dis.* (2018) 18:265. doi: 10.1186/s12879-018-3164-z
- Rosser A, Marx FM, Pareek M. Recurrent tuberculosis in the pre-elimination era. Int J Tuberc Lung Dis. (2018) 22:139–50. doi: 10.5588/ijtld.17.0590
- Jasmer RM, Bozeman L, Schwartzman K, Cave MD, Saukkonen JJ, Metchock B, et al. Recurrent tuberculosis in the United States and Canada: relapse or reinfection? *Am J Respir Crit Care Med.* (2004) 170:1360–6. doi: 10.1164/rccm.200408-1081OC
- Narayanan S, Swaminathan S, Supply P, Shanmugam S, Narendran G, Hari L, et al. Impact of HIV infection on the recurrence of tuberculosis in South India. *J Infect Dis.* (2010) 201:691–703. doi: 10.1086/650528
- Bruchfeld J, Correia-Neves M, Kallenius G. Tuberculosis and HIV coinfection. Cold Spring Harb Perspect Med. (2015) 5:a017871. doi: 10.1101/cshperspect.a017871
- Yang C, Gao Q. Recent transmission of *Mycobacterium tuberculosis* in China: the implication of molecular epidemiology for tuberculosis control. *Front Med.* (2018) 12:76–83. doi: 10.1007/s11684-017-0609-5
- Christianson S, Wolfe J, Orr P, Karlowsky J, Levett PN, Horsman GB, et al. Evaluation of 24 locus MIRU-VNTR genotyping of *Mycobacterium tuberculosis* isolates in Canada. *Tuberculosis*. (2010) 90:31–8. doi: 10.1016/j.tube.2009.12.003

- 43. Iwamoto T, Yoshida S, Suzuki K, Tomita M, Fujiyama R, Tanaka N, et al. Hypervariable loci that enhance the discriminatory ability of newly proposed 15-loci and 24-loci variable-number tandem repeat typing method on *Mycobacterium tuberculosis* strains predominated by the Beijing family (vol 270, pg 67, 2007). *Fems Microbiol Lett.* (2007) 272:282–3. doi: 10.1111/j.1574-6968.2007.00777.x
- Asante-Poku A, Nyaho MS, Borrell S, Comas I, Gagneux S, Yeboah-Manu D. Evaluation of customised lineage-specific sets of MIRU-VNTR loci for genotyping *Mycobacterium tuberculosis* complex isolates in Ghana. *PLoS ONE*. (2014) 9:e92675. doi: 10.1371/journal.pone.0092675
- 45. Supply P, Lesjean S, Savine E, Kremer K, van Soolingen D, Locht C. Automated high-throughput genotyping for study of global epidemiology of *Mycobacterium tuberculosis* based on mycobacterial interspersed repetitive units. *J Clin Microbiol.* (2001) 39:3563–71. doi: 10.1128/JCM.39.10.3563-3571.2001
- 46. Zhou Y, van den Hof S, Wang S, Pang Y, Zhao B, Xia H, et al. Association between genotype and drug resistance profiles of *Mycobacterium tuberculosis* strains circulating in China in a national drug resistance survey. *PLoS ONE.* (2017) 12:e0174197. doi: 10.1371/journal.pone.0174197
- 47. Tian LL, Si HY, Mu TJ, Fan WB, Wang J, Jiang WM, et al. Molecular epidemiology of *Mycobacterium tuberculosis* in Gansu province of China. *Chinese Med J Peking.* (2012) 125:3458–64. doi: 10.3760/cma.j.issn.0366-6999.2012.19.018

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Qiu, Tao, Liu, Li, Song, Tian, Wu, Wu, Zhan, Lu and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.