



Alterations of Striatal Subregions in a Prion Protein Gene V180I Mutation Carrier Presented as Frontotemporal Dementia With Parkinsonism

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Objective: To explore the roles of striatal subdivisions in the pathogenesis of frontotemporal dementia with parkinsonism (FTDP) in a patient resulting from prion protein gene (PRNP) mutation.

Methods: This patient received clinical interviews and underwent neuropsychological assessments, genetic testing, [¹⁸F]-fluorodeoxyglucose positron emission tomography ([¹⁸F]-FDG PET)/MRI, and [¹⁸F]-dihydrotetrabenazine positron emission tomography ([¹⁸F]-DTBZ PET)/CT. Region-of-interest analysis was conducted concerning metabolism, and dopamine transport function between this patient and 12 controls, focusing on the striatum subregions according to the Oxford-GSK-Imanova Striatal Connectivity Atlas.

Results: A 64-year-old man initially presented with symptoms of motor dysfunction and subsequently behavioral and personality changes. FTDP was initially suspected. Sequence analysis disclosed a valine to isoleucine at codon 180 in *PRNP*. Compared to controls, this patient had a severe reduction (> 2SD) of standard uptake value ratio (SUVr) in the limbic and executive subregions but relative retention of metabolism in rostral motor and caudal motor subregions using [¹⁸F]-FDG PET/MRI, and the SUVr decreased significantly across the striatal in [¹⁸F]-DTBZ PET/CT, especially in the rostral motor and caudal motor subregions.

Conclusion: The alteration of frontal striatal loops may be involved in cognitive impairment in FTDP, and the development of parkinsonism in FTDP may be primarily due to the involvement of the presynaptic nigrostriatal loops in *PRNP V180I* mutation.

Keywords: frontotemporal dementia with parkinsonism, prion protein gene, striatum, dopamine, positron emission tomography

Abbreviations: FTDP, frontotemporal dementia with parkinsonism; *PRNP*, prion protein gene; *MAPT*, microtubule-associated protein tau; *PGRN*, progranulin; PD, Parkinson's disease; CB, connectivity-based; SUVr, standard uptake value ratio; [¹⁸F]-FDG PET, [¹⁸F]-fluorodeoxyglucose positron emission tomography; [¹⁸F]-DTBZ PET, [¹⁸F]-dihydrotetrabenazine positron emission tomography.

INTRODUCTION

Frontotemporal dementia with parkinsonism (FTDP) is a group of degenerative disorders that can occur sporadically or in families. Mutations in the genes encoding the microtubule-associated protein tau (*MAPT*) and progranulin (*PGRN*) on chromosome 17 have been linked to familial FTDP (Boeve and Hutton, 2008). There has been an increase in the number of cases with a clinical diagnosis of FTDP that were shown to be genetic prion diseases caused by prion protein gene (*PRNP*) mutations (Nitrini et al., 2001; Woulfe et al., 2005; Jansen et al., 2010; Bernardi et al., 2014). However, the underlying mechanism behind this remains elusive.

The striatum, aside from its fundamental movement function evidenced by parkinsonian deficits, is involved in processing of closely related non-motor, cognitive and reward information. Previous research found that the caudate was the most vulnerable to lesion in sporadic amyotrophic lateral sclerosis-frontotemporal dementia (FTD) continuum patients associated with cognitive decline with FTD features (Masuda et al., 2016). Therefore, we hypothesize that alterations in striatal regions may be related to the mechanisms of cognitive and motor impairment in FTDP. In recent years, multiple imaging modalities have been used to explore the pathogenesis of the disease. For example, [¹⁸F]-fluorodeoxyglucose positron emission tomography ([¹⁸F]-FDG PET) is a technique for measuring changes in energy metabolism that reflect underlying cellular events. [¹⁸F]-dihydrotrabectedin positron emission tomography ([¹⁸F]-DTBZ PET) reflects structural brain changes caused by pathology in the nigrostriatal dopamine system. Previous studies using [¹⁸F]-N-3-fluoropropyl-2β-carboxymethoxy-3β-(4-iodophenyl)-nortropine positron emission tomography ([¹⁸F]-FP-CIT PET), in patients with Parkinson's disease (PD) and parkinsonism based on the anatomical subregions of the striatum found that the loss of dopamine transporters differed across the striatal subregions among diseases and can be used for disease differentiation (Oh et al., 2012; Sung et al., 2017; Kong et al., 2020). However, anatomical subregions may not provide the best annotation of the striatum. Tziortzi et al. (2014) classified the striatum into functional subregions based on anatomical links between the striatum and the cortex, and the homogeneity of dopamine release was significantly higher than the structural subregion. It remains unclear whether genetic prion diseases develop FTDP as a result of striatal functional area involvement.

To address this issue, we performed a series of assessments of metabolic and dopamine transporter function in a genetic prion disease caused by *PRNP* V180I mutation presented with FTDP, with a focus on the functional regions of the striatum.

METHODS

Ethics Statement

The study was approved by the Ethics Committees of the Xuanwu Hospital of Capital Medical University and carried out in accordance with the principles of the Helsinki Declaration. Each patient or their guardian provided written informed consent.

Study Design

A patient with clinical diagnosis of behavioral variant FTD and progressive parkinsonism confirmed to be genetic prion disease (*PRNP* V180I) was enrolled. Thereafter, 12 gender- and age-matched healthy controls and 3 cases with PD who fulfilled the UK Parkinson's Disease Society Brain Bank clinical diagnostic criteria admitted to the Department of Neurology at Xuanwu Hospital between 1 January 2019 and 31 January 2021 were recruited to our cohort.

The patient with genetic prion disease underwent genetic analysis, cerebrospinal fluid (CSF) 14-3-3 protein levels and prion real-time quaking-induced conversion (RT-QuIC) testing, electroencephalography (EEG) monitoring, and brain magnetic resonance imaging (MRI). Genomic DNA was extracted from fresh peripheral blood leukocytes, and whole-exome sequencing (WES) libraries were generated using the Agilent SureSelect Human All Exon V6 Kit (Agilent Technologies, Santa Clara, CA, United States). CSF protein 14-3-3 levels and RT-QuIC were detected at the National Reference Laboratory for Human Prion Diseases, CDC, China, according to the working procedures described previously (Gao et al., 2011; Xiao et al., 2019). Two-hour EEG monitoring was performed using a 21-lead EEG transducer (Micromed, Italy). The EEG electrodes were placed according to the International 10–20 system. MRI was performed at 3.0 T (Erlangen, Germany) with the following sequences: T1 weighted image (T1WI), T2 weighted image (T2WI), fluid-attenuated inversion recovery (FLAIR), diffusion-weighted imaging (DWI), and apparent diffusion coefficient (ADC) values.

All the subjects underwent clinical interviews and examination, including neuropsychological assessments and cerebral [¹⁸F]-FDG PET/MRI examinations. Six controls and all the cases with PD underwent [¹⁸F]-DTBZ PET/computed tomography (CT).

Neuroimaging Acquisition and Preprocessing

[¹⁸F]-Fluorodeoxyglucose Positron Emission Tomography/Magnetic Resonance Imaging Acquisition and Preprocessing

The [¹⁸F]-FDG PET/MRI scans were carried out with the help of a GE Signa PET/MR 3.0 Tesla scanner (GE Healthcare, Milwaukee, WI, United States). A 30-min dynamic scan was acquired approximately 45 min after intravenous injection of 3.7 MBq/kg of [¹⁸F]-FDG. Attenuation correction, scattering correction, random correction, decay correction, and dead-time correction were performed on the PET data. Corrected PET data were obtained using a time-of-flight, point spread function, ordered subset expectation maximization (TOF-PSF-OSEM) algorithm with 4 iterations and 16 subsets. Finally, all images were spatially normalized to the PET Montreal Neurological Institute (MNI) brain space template, scaled, averaged, and then smoothed using 8-mm full width at half maximum (FWHM) Gaussian kernel using SPM12 (Statistical Parametric Mapping 12)¹ running under MATLAB 7.11 (Mathworks Inc., Sherborn, MA, United States) on the CentOS 6.5. The intensity of

¹<http://www.fil.ion.ucl.ac.uk/spm>

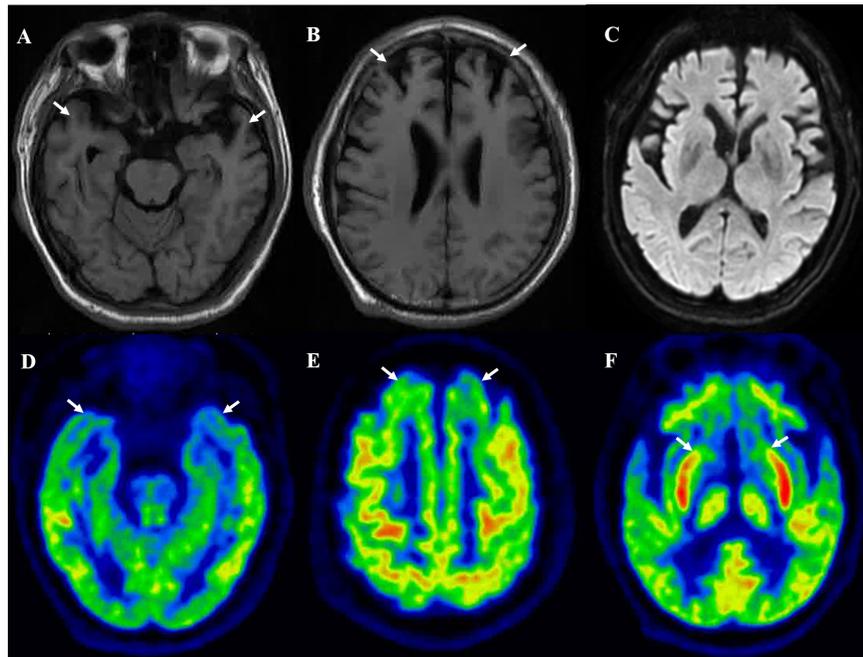


FIGURE 1 | Brain MRI suggested bilateral frontal (**B**) and temporal (**A**) lobes atrophy, and no hyperintensity was detected in diffusion-weighted imaging (**C**). [^{18}F]-FDG-PET/MRI suggested bilateral frontal lobe (**E**), temporal lobe (**D**), and caudate nucleus (**F**) hypometabolism.

[^{18}F]-FDG PET/MRI scans were normalized using the entire cerebellar reference region to generate standard uptake value ratio (SUVr) images.

[^{18}F]-Dihydrotetrabenazine Positron Emission Tomography/Computed Tomography Acquisition and Preprocessing

The [^{18}F]-DTBZ PET/CT scans were performed on a Gemini GXL 16 PET/CT scanner (Philips, Amsterdam, the Netherlands) in three-dimensional iterative mode. Ninety minutes after injecting approximately 250 MBq of [^{18}F]-DTBZ radiotracer ([^{18}F]-AV133), 15 min of brain scans were obtained. Using the [^{18}F]-AV133 PET template, the florbenzamine images were spatially normalized to the standard atlas. The target regions were the assigned regions of interest based on the atlas (caudate, anterior and posterior putamen, and the entire striatum). The SUVr was calculated by calculating the ratio of tracer activity in the target region relative to the occipital cortex as the reference region.

Analysis at the Striatal Subregion Level

The striatum was subdivided using the Oxford-GSK-Imanova Striatal Connectivity Atlas, a probabilistic atlas that divides the striatum into seven subregions based on white matter connections to cortical regions (Tziortzi et al., 2014). To begin, the striatum was divided into four subregions, each of which was linked to the frontal, parietal, occipital, and temporal lobes. The frontal subregion is further subdivided into four subregions (limbic, executive, rostral motor, and caudal motor). Limbic subregion is linked to the anterior orbital gyrus, posterior orbital

gyrus, medial orbital gyrus, gyrus rectus, and subcallosal gyrus-ventral anterior cingulate; executive subregion is linked to the rostral superior and middle frontal gyri and the dorsal prefrontal cortex; rostral motor subregion is linked to the caudal portions of lateral and medial superior gyrus as well as the caudal middle and inferior frontal gyri; caudal motor subregion is linked to the precentral gyrus that corresponds functionally to the primary motor cortex (Area 4) and the caudal premotor area (Caudal area 6).

Statistical Analysis

Analyses were performed using SPSS version 22.0 (IBM, Armonk, NY, United States). Continuous data are represented as the mean \pm SD. Two or more standard deviations from the mean between groups were considered statistically significant.

RESULTS

Case Presentation

A 64-year-old man presented with initial symptoms of limb rigidity, bradykinesia. In the following months, he developed involuntary limb tremors and gait abnormality, but was still able to walk independently; these symptoms can be partially alleviated by levodopa. One year after the onset of symptoms, he developed apathy, behavior abnormality, loss of sympathy or empathy, hyperorality, and disinhibition, and had difficulty in daily activities. The symptoms of parkinsonism also became aggravated; he had severe bilateral

TABLE 1 | Comparison of the striatal functional distribution in this case and controls using [¹⁸F]-FDG-PET/MRI.

Striatal subregion	[¹⁸ F]-FDG-PET/MRI (SUVR value)	
	Controls (N = 12)	Case with <i>PRNP</i> mutation
Limbic subregion	1.43 ± 0.16	1.06†
Executive subregion	1.37 ± 0.18	0.99†
Rostral motor subregion	1.37 ± 0.18	1.20
Caudal motor subregion	1.41 ± 0.20	1.26
Parietal subregion	1.48 ± 0.19	1.45
Occipital subregion	1.34 ± 0.17	1.36
Temporal subregion	1.02 ± 0.12	0.94

† Indicated values in patient with V180I mutation are 2SD lower than the mean value for the controls in the same brain regions.

lower extremity rigidity with inability to flex, leading to a bed-bound state, and was accompanied by fecal incontinence and sweating. He showed no improvement to antiparkinson and dystonia-relieving drugs. He had no family history concerning similar neurodegenerative diseases. Neurological examination 26 months after onset revealed akinetic mutism, marked rigidity in all of the extremities, bilateral hyporeflexia, and extensor Babinski signs with the Hoehn and Yahr of stage V. Cognitive assessment (mini-mental state examination and Montreal Cognitive Assessment scores) did not cooperate. Brain MRI displayed bilateral frontotemporal lobe atrophy (Figures 1A,B) and no hyperintensity was detected in diffusion-weighted imaging or fluid attenuation inversion recovery imaging (Figure 1C). FTDP was initially suspected. Genomic DNA extracted from peripheral blood was used to identify mutations associated with FTDP. It turned out that the patient carried a point mutation of valine to isoleucine at codon 180, as well as an MM polymorphism at codon 129 in *PRNP*. No mutations were detected in FTD-related genes, such as *MAPT*, *GRN*, *TARDBP*, *FUS*, *C9orf72*, *VCP*, *CHMP2B*, *SQSTM1*, and *TBK1*.

Ancillary tests related to prion diseases were implemented further, and CSF analyses revealed a negative 14-3-3 protein assay. The CSF prion RT-QuIC assay was negative. EEG showed non-specific slow waves but lacking a definite periodism.

[¹⁸F]-Fluorodeoxyglucose Positron Emission Tomography Analysis

The [¹⁸F]-FDG PET showed diffuse cortical and subcortices hypometabolism (Figures 1D–F). Compared to controls, this patient had a severe reduction of SUVR in the limbic subregion (25.9%) and executive subregion (27.7%), but the motor subregion did not differ between this patient and controls using functional subdivision. Detailed data are shown in Table 1.

[¹⁸F]-Dihydrotrabenazine Positron Emission Tomography Analysis

[¹⁸F]-DTBZ PET revealed reduced radioactivity in bilateral putamen, predominantly in the posterior part. Asymmetric decline of vesicular transporter availability with a posterior-to-anterior gradient can be seen in cases with PD (Figure 2). The functional subdivision analysis suggested the SUVR across

the striatal decreased (> 2SD) significantly, with a reduction of 40.5% in rostral motor region and 47.7% in caudal motor subregion compared to controls. Detailed data are provided in Table 2.

DISCUSSION

In this study, we present the first case of a *PRNP* V180I carrier presenting with the clinical syndrome of FTDP. Furthermore, we investigated the pathogenesis of FTDP related to striatal subdivision in this patient using multiple imaging modalities. Our findings showed that this case had hypometabolism in cognition-related limbic and executive subregions, while the motor-related rostral and caudal motor areas were mismatched for metabolism and dopamine transport function. These findings suggest that disruption of frontal striatal loops may be involved in cognitive impairment in FTDP, and that disruption of nigrostriatal loops may be a strong argument in favor of *PRNP* mutation-associated parkinsonism, providing a more complete understanding of the mechanisms underlying striatal-associated FTDP in subjects with *PRNP* mutations.

Genetic prion diseases carrying V180I mutation have been widely reported in East Asia. Patients carrying this mutation usually have atypical clinical and laboratory features (Qina et al., 2014). Previous studies have reported that V180I mutation can mimic Alzheimer's disease (AD) (Bagyinszky et al., 2019a) or dementia with Lewy bodies (DLB) (Tomizawa et al., 2020). FTD or FTDP is a rare genetic prion disease phenotype. To date, 20 point mutations and 3 insertional mutations in *PRNP* have been reported to be associated with this phenotype (Supplementary Table 1). Patients with the FTD phenotype often lack the typical ancillary findings of prion disease. Of the published prion patients with the FTD/FTDP phenotype, it is roughly estimated that no patients have periodic sharp-wave complexes on EEG, three quarters had no hyperintensity on MRI, and two-thirds were negative for CSF 14-3-3 protein (Nitrini et al., 2001; Hall et al., 2005; Woulfe et al., 2005; Clerici et al., 2008; Giovagnoli et al., 2008; Alzualde et al., 2010; Bernardi et al., 2010, 2014; Jansen et al., 2010, 2011; Kumar et al., 2011; Beck et al., 2013; Cupidi et al., 2013; McKnight et al., 2013; San Millán et al., 2013; Riudavets et al., 2014; Mano et al., 2016; Oldoni et al., 2016; Kenny et al., 2017; Ghetti et al., 2018; Sun et al., 2018; Takayanagi et al., 2018; Bagyinszky et al., 2019b; Di Fede et al., 2019; Priemer et al., 2019; Townley et al., 2020).

The striatum is widely acknowledged to have connections with the cortex and to play an important role in the striato-cortical circuitry, which has been characterized by several functionally segregated subcircuits that are anatomically different but functionally adjacent. In this study, a connectivity-based (CB) functional striatum atlas was used to provide optimal subdivision of the striatum. In the CB functional striatum atlas, the frontal lobe connections dominate the total volume of the striatum and are the main factor influencing the functional organization of the striatum (Tziortzi et al., 2014). We presumed that the reduction of SUVR in the limbic and executive subregions of the striatum may be secondary to the hypometabolism of the frontal lobes through the frontal-striatal loops in this patient.

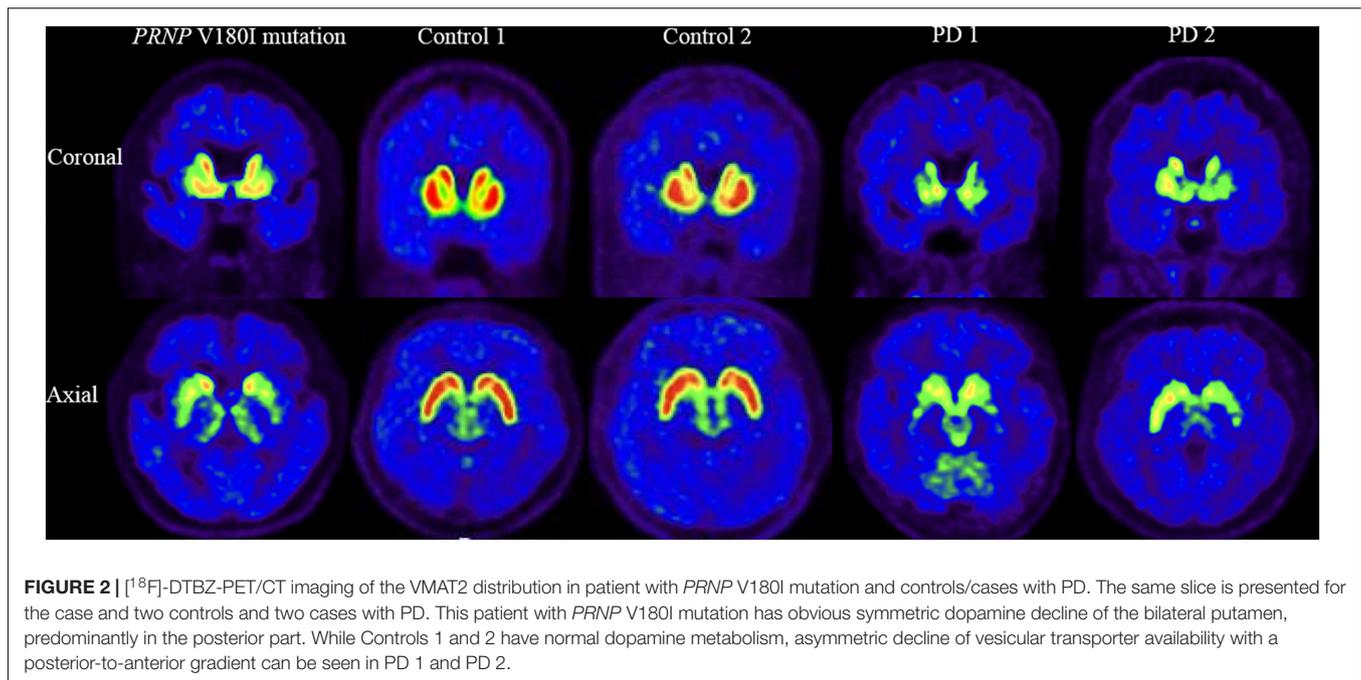


TABLE 2 | Comparison of the striatal functional distribution in this case and controls/cases with PD using [¹⁸F]-DTBZ-PET/CT.

Striatal subregion	[¹⁸ F]-DTBZ-PET/CT (SUVR value)		
	Controls (N = 6)	PD (N = 3)	Case with <i>PRNP</i> mutation
Limbic subregion	3.35 ± 0.11	2.49 ± 0.07	2.40 [†]
Executive subregion	2.96 ± 0.08	2.08 ± 0.18	1.97 [†]
Rostral motor subregion	2.42 ± 0.10	1.66 ± 0.17	1.44 [†]
Caudal motor subregion	2.83 ± 0.16	1.64 ± 0.15	1.48 [†]
Parietal subregion	3.01 ± 0.14	1.63 ± 0.26	1.61 [†]
Occipital subregion	2.60 ± 0.31	2.00 ± 0.21	2.24
Temporal subregion	–	–	–

[†]Indicated values in patient with V180I mutation are 2SD lower than the mean value for the controls in the same brain regions.

This case showed a severe hypometabolism in the caudate nucleus and a relatively preserved metabolism in the putamen in [¹⁸F]-FDG PET, which is distinguished from PD or other parkinsonism, such as hypermetabolism of the putamen in PD and metabolic reductions of the putamen in multiple system atrophy (Eckert et al., 2005; Akdemir et al., 2014; Meles et al., 2020). The symmetrical low intake of the putamen using [¹⁸F]-DTBZ PET also differs in patients with PD, who generally have asymmetrical presynaptic dopaminergic synapse selectively degenerated, causing the decrease of dopaminergic uptake with a posterior to the anterior gradient in the striatum (Nandhagopal et al., 2009). Putamen-related motor circuits are well known to be responsible for motor functions, and putamen dysfunction correlates well with the onset of cardinal motor symptoms and motor severities. It is surprising that this patient had severe motor symptoms while metabolism in this area was relatively preserved. Further, functional subdivision analysis

suggested the SUVR in motor subregions decreased significantly using dopamine transporter function imaging while relatively preserved metabolism on metabolic imaging. This mismatch pattern of metabolism and dopamine transport function of motor subregions suggests that parkinsonism caused by *PRNP* mutation may be mainly associated with presynaptic degeneration of the nigrostriatal pathway. This finding contradicts the conventional perception that parkinsonism in prion diseases is generally regarded to be postsynaptic structural deterioration caused by spongiform degeneration of basal ganglia (Maltête et al., 2006). Although the application of [¹²³I] FP-CIT in a few cases of genetic prion disease also found altered presynaptic dopamine function, none of these studies evaluated the metabolism of the striatum (Malek et al., 2017; Baiardi et al., 2020; Tomizawa et al., 2020). Of course, no response to levodopa therapy observed in our patient could be explained by post-synaptic dysfunction of the nigrostriatal pathway. There may also be deterioration of the postsynaptic nigrostriatal pathway in this patient due to lower SUVR in motor subregions than in controls, although no significant difference was reached. Indeed, a combination of presynaptic and postsynaptic cell loss in the substantia nigra system has been observed in pathological studies of patients with sporadic prion diseases (Vital et al., 2009).

There were some limitations to this study. First, the negative results of Creutzfeldt-Jakob disease (CJD) supportive investigation (CSF 14-3-3, EEG, MRI, and RT-QuIC) and the lack of pathology data to support a diagnosis of prion disease in this patient do not rule out the potential of an incidental V180I combination, as V180I carriers have only a weak to moderate elevated risk of developing prion disease (Minikel et al., 2016). Second, because genetic prion diseases are extremely rare, the results were hampered by small sample size. Third, the enrolled patient was in a relatively

advanced stage of the disease with severe clinical manifestations and lack of imaging of dynamic changes. Hence, more research into striatal subdivisions in larger genetic prion diseases with FTDP or parkinsonism is required.

In summary, the frontal striatal loops may be involved in cognitive impairment of FTDP and the movement dysfunction of FTDP may be primarily due to the involvement of the presynaptic nigrostriatal loops. Measuring metabolism and dopaminergic changes in a functionally defined striatal region could provide a more sensitive tool for detecting PRNP-associated specific striatal changes.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by the Ethics Committees of the Xuanwu Hospital of Capital Medical University. The patients/participants provided their written informed consent to participate in this study.

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AUTHOR CONTRIBUTIONS

LW: study concept and design. JM, LL, SL, ZC, MC, and ZW: acquisition of data analysis and interpretation of data. ZC: drafting of the manuscript. JM, LL, SL, MC, JZ, ZW, PC, and LW: critical revision of the manuscript for important intellectual content. All authors read and approved the final manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fnagi.2022.830602/full#supplementary-material>

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