



Differential expression of large-conductance Ca^{2+} -activated K channels in the carotid body between DBA/2J and A/J strains of mice

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The carotid body (CB) is a primary chemosensory organ for arterial hypoxia. Inhibition of K channels in chemosensory glomus cells (GCs) are considered to be responsible for hypoxic chemoreception and/or chemotransduction of the CB. Hypoxic sensitivity of large-conductance calcium-activated K (BK) channels has been established in the rat CB. Our previous work has shown the BK channel $\beta 2$ subunits are more expressed in the CB of the DBA/2J mouse than that of the A/J mouse. Because the DBA/2J mouse is more sensitive to hypoxia than the A/J mouse, our general hypothesis is that BK channels play a role in the sensitivity of the mouse CB to mild hypoxia. We performed vigorous analysis of the gene expression of α , $\beta 2$, and $\beta 4$ subunits of BK channels in the CB. We found that α and $\beta 2$ subunits were expressed more in the CB of the DBA/2J mice than that of the A/J mice. No differences were found in the $\beta 4$ subunit expression. These differences were not seen in the neighboring tissues, the superior cervical ganglion and the carotid artery, suggesting that the differences are CB specific. Further, the sensitivity of BK channels in GCs to mild hypoxia was examined in patch clamp experiments using undissociated CBs. Iberiotoxin significantly inhibited K current of GCs in the DBA/2J mice, but not in the A/J mice. When reducing PO_2 to ~ 70 mmHg, K current reversibly decreased in GCs of the DBA/2J, but not of the A/J mice. In the presence of iberiotoxin, mild hypoxia did not inhibit K current in either strains. Thus, the data suggest that BK channels in GCs of the DBA/2J mice are sensitive to mild hypoxia. Differential expression of BK channel β subunits in the CBs may, at least in part, explain the different hypoxic sensitivity in these mouse strains.

Keywords: chemoreceptor, glomus cell, hypoxia, patch clamp, oxygen, RT-PCR

INTRODUCTION

The carotid body (CB) is a primary chemosensory organ for arterial hypoxia. Its excitation by hypoxia induces a variety of responses in the cardiovascular, respiratory, renal, and endocrine systems to compensate decreased oxygen levels. The mechanisms of hypoxic chemoreception and chemotransduction in the CB are still not totally understood, but a general consensus is that the glomus cell (GC, or Type I cell) is a major chemosensory element (Gonzalez et al., 1994; Shirahata and Sham, 1999; Kumar and Prabhakar, 2007). A widely accepted model assumes that hypoxic inhibition of oxygen-sensitive K channels induces depolarization of GCs, followed by an activation of voltage-dependent Ca^{2+} channels, an influx of Ca^{2+} , and a release of neurotransmitters from GCs. Neurotransmitters bind to their receptors in the afferent nerve endings, which generates increased action potentials (Gonzalez et al., 1994; Peers, 1997; Shirahata and Sham, 1999; Lopez-Lopez and Perez-Garcia, 2007). The efforts for defining oxygen-sensitive K channels in the CB reveal several molecular identities. For example, in the rabbit GC, Kv4 family, in particular Kv4.3 α subunits, appears to be the molecular correlate of the oxygen-sensitive K channel (Sanchez et al., 2002). On the other hand, large-conductance calcium-activated K channels (BK channels; Peers, 1997) and background TASK-like K channels (Williams and Buckler, 2004) were identified

to be oxygen sensitive in the rat GC. In the cat GC, oxygen-sensitive K current is mediated via a delayed rectifier, but neither BK channels nor fast-inactivating K channels (Kv4 family) appear to be responsible (Chou and Shirahata, 1996). Further, Kv3 family in GCs of C57BL/6J mice is inhibited by hypoxia (Perez-Garcia et al., 2004). These differences have been attributed to species differences (Shirahata and Sham, 1999; Lopez-Lopez and Perez-Garcia, 2007; Peers and Wyatt, 2007). We can also consider these species differences as variable protein expression in the CB due to background genetic differences. Another factor that needs to be contemplated is oxygen tension. Some studies have indicated that the inhibition of K channels is PO_2 dependent (Lopez-Lopez et al., 1989; Montoro et al., 1996; Shirahata and Sham, 1999), although many studies have applied only severe hypoxia. Thus, it is possible that each oxygen-sensitive K channel contributes to GCs' function at specific levels of oxygen tension.

Recent studies in inbred strains of mice have suggested that the CB in the DBA/2J strain is more sensitive to moderate hypoxia than that in the A/J strain (Tankersley et al., 1994; Rubin et al., 2003; Campen et al., 2004). Investigating the differences in the CB between these inbred strains of mice could offer deeper understanding of the CB function and its relation to genetics. Our previous study found that gene expression of BK channel $\beta 2$

subunits are more expressed in the CB of the DBA/2J mice compared to the A/J mice (Balbir et al., 2007). Further, K current in the CB of DBA/2J mice was inhibited by mild hypoxia (Yamaguchi et al., 2004). Thus, our general hypothesis is that BK channels play a role in the sensitivity of the mouse CB to mild hypoxia. In this study, we performed more vigorous analysis of gene expression of α , β_2 , and β_4 subunits of BK channels. Further, we have examined if the differences in BK channel expression between the DBA/2J and the A/J mice correlate with the response of GCs to mild hypoxia.

MATERIALS AND METHODS

TISSUE PREPARATION

All animal experiments were conducted in accordance with the NIH guidelines (Guide for the Care and Use of Laboratory Animals, National Academy Press Washington, DC, 1996), and the protocols were approved by the Animal Care and Use Committee of the Johns Hopkins University. DBA/2J and A/J mice were purchased from Jackson Laboratories (Bar Harbor, ME, USA). Some mice were bred in the facility of the Johns Hopkins Bloomberg School of Public Health where the temperature ($\sim 22^\circ\text{C}$) and the light cycle (12 h light and 12 h dark) were controlled. Water and mouse chow (Harlan TEKLAD, Madison) were provided *ad libitum* before all experiments. Preparation of the CB was performed as described before (Yamaguchi et al., 2004) with some modifications. In short, male mice (4–6 weeks old) were deeply anesthetized by peritoneal injection of 1.5 g/kg urethane. The carotid bifurcations were exposed. After the heart was removed to avoid bleeding in the neck, the CBs were quickly harvested together with the carotid bifurcation and immersed in ice-cold phosphate-buffered saline (for RT-PCR studies) or modified Krebs solution (for patch clamp experiments). Fat and connective tissues were removed using a stereoscopic microscope.

RT-PCR ANALYSIS OF BK CHANNEL SUBUNITS

The CBs were isolated in ice-cold phosphate-buffered saline, collected in ice-cold TRIzol solution (Invitrogen, CA, USA), and frozen at -80°C until use. Total RNA isolation and DNase treatment were similar to what has been described previously (Balbir et al., 2007). RNA was reverse-transcribed to obtain first strand cDNA using SuperScript III™ (Invitrogen). We performed real-time PCR targeting BK channel subunits (α , β_2 , β_4) together with β -actin (Act) as a reference gene. TaqMan7 gene expression assays (Applied Biosystems) were used for PCR amplification of cDNA following the manufacture's instruction. The primers were purchased from Applied Biosystems: Mm00516078_m1 for BK α , Mm00511481_m1 for BK β_2 , Mm00465684_m1 for BK β_4 , and 4352341E for Act. ABI PRIZM7 7000HT Sequence Detection System (Applied Biosystems) was used. The protocol for PCR amplification was: 2 min at 50°C , 10 min at 95°C , 50 cycles of 95°C for 15 s followed by 60°C for 1 min. Individual cDNA samples were run in triplicates for each gene. After completion of real-time amplification, the amplification efficiency, crossing point, and corrected crossing point (cCP) of each run were determined using the methods of data analysis for real-time (DART) PCR (Peirson et al., 2003) and four-parameter logistic model (FPLM; Tichopad et al., 2003). CAMPER 1.2 software (Center for Biotechnology, Bielefeld

University¹) provided a simple and computerized tool for these analytical methods. Subsequently, average and SD of all readings in efficiency and cCP were calculated. The readings outside of mean $\pm 3\text{SD}$ were eliminated from further analysis of the data. Also, if one reading in the triplicates was three cCP away from other two readings, it was eliminated from further analysis. A mean of the triplicates was calculated to represent a gene transcript of a particular sample. Statistical analysis was performed with Student's *t*-test using Prism 4.0 (GraphPad Software Inc.) and with the Relative Expression Software Tool (REST 2009; Qiagen). REST is based on the efficiency-calibrated model and employs randomization tests to determine differences in target gene expression between experiments. Differences were considered significant if $p < 0.05$.

EXPRESSION OF BK CHANNEL SUBUNITS IN THE SUPERIOR CERVICAL GANGLION AND THE CAROTID ARTERY

We further examined if gene expression of BK channel subunits differ in the superior cervical ganglion (SCG) and the carotid artery between the two strains of mice. RNA extraction and RT were performed as described above, and a conventional multiplex PCR was performed following the manufacture's instruction (Qiagen). Primers were designed based on the published nucleotide sequences of mice BK channel subunits (GenBank²). The primers' sequences were: BK α subunit (forward: CTCATCATCCCGGTGACCAT; reverse: GGATTC-TATTGGGTTTGACGAG); BK β_2 subunit (forward: TTTATATG-GACCAGTGGCCG; reverse: TGTTGATCCGTTGGATCCCTCT); BK β_4 subunit (forward: GGTGTCTTACGAGTACACGGAA; reverse: TTGCGTTTCTTCATGGCTTC); Act (forward: CGT-GAAAAGATGACCCAGATCA; reverse: ATTTCCCTCTCAGCT-GTGGTG). The final concentrations of primers were $1\ \mu\text{M}$ for β -actin and $5\ \mu\text{M}$ for other primers. The conditions for PCR amplification were as follows. The template was denatured at 95°C for 15 min followed by 25 cycles of denaturation at 94°C for 30 s, annealing of primers at 60°C for 1.5 min, extension at 72°C for 1.5 min. Final cycle was 72°C for 10 min. The PCR products were purified with QIAquick PCR Purification Kit (Qiagen), and the second multiplex PCR was performed. The conditions for PCR amplification were the same as the first PCR except for applying 20 cycles. The final PCR products were separated by 2% agarose gel electrophoresis. The image of gel electrophoresis was captured and analyzed by Kodak Electrophoresis Documentation and Analysis System 290 (Eastman Kodak Co.). The density of the each band was measured and the expression of each BK channel subunit was normalized relative to the expression of β -actin in the same sample.

BK CHANNEL ACTIVITY

Patch clamp experiments were performed using undissociated CBs as described before (Yamaguchi et al., 2004). The CB was localized by gently pulling the SCG, and then, the SCG was removed. Subsequently, the tissue was warmed in normoxic Krebs solution (in mM: NaCl, 118; KCl, 4.7; CaCl₂, 1.8; KH₂PO₄, 1.2; MgSO₄·7H₂O, 1.2; NaHCO₃, 22; glucose, 11.1; pH, 7.4 with 5% CO₂/Air) at 37°C for 60 min, and then placed in a recording chamber which was

¹<http://camper.cebitec.uni-bielefeld.de/>

²<http://www.ncbi.nlm.nih.gov/Genbank/>

attached to a stage of an upright microscope (Axioskop 2, Zeiss). After the tissue was superfused with Krebs solution containing 0.0375% collagenase (Type IX, Sigma) for 40 min, the CB was visualized using a water immersion lens (ACHROPLAN, 40 \times , Zeiss) combined with an infrared differential interference video camera (DAGE MTI Inc., Michigan City, IN, USA; **Figures 3A,B**). GCs were easily identified from their morphology as seen in **Figure 3B**. All experiments were performed at 37.0°C while the tissue was continuously superfused with Krebs solution (1.22 mL/min) using a peristaltic pump (Minipuls 3, Gilson). A conventional tight seal whole cell recording was applied. The patch electrodes (Corning 8161, Warner Instrument Co.) were pulled using a puller (Sutter Instrument Co.), and their resistance was between 4 and 6 M Ω . The composition of the internal solution was: (in mM) K gluconate, 90; KCl, 33; NaCl, 10; CaCl₂, 1; EGTA, 10; MgATP, 5; HEPES, 10; pH 7.2 with KOH. Voltage-dependent whole cell current was evoked by voltage clamp pulses from -80 mV of holding potential to $+60$ mV with 10 mV increments for 100 ms. Single pulse protocol (from -80 mV of holding potential to $+20$ mV) was also used. A specific BK channel blocker, iberiotoxin (Alomone Labs), was included in Krebs solution. Hypoxia was applied by switching normoxic Krebs to hypoxic Krebs solution saturated with 5% CO₂/95% N₂. PO₂ in the chamber was measured with a small oxygen electrode (MI-730 Oxygen Electrode, OM-4 Oxygen Meter; Microelectrodes, Inc.) in separate experiments in which the patch clamp experiments were simulated. The current was amplified and filtered at 2 kHz with a low pass Bessel filter using an Axopatch 200B patch clamp amplifier (Axon Instruments). The signals were digitized by a Digidata 1320A (Axon Instruments). A window based personal computer and pCLAMP 8.1 (Axon Instrument) were used for acquisition and processing of data.

All chemicals were obtained from Sigma Aldrich Inc., unless otherwise indicated. For data analysis, current size was measured between 93 and 97 ms from the start of a step pulse application, and the data were expressed as mean \pm SEM (pA or % of control). Prism 4.0 was used to determine statistical differences, and Student's *t*-test or ANOVA was applied. Values were considered significant if $p < 0.05$.

RESULTS

EXPRESSION OF BK CHANNEL SUBUNITS

A total of 16 samples from DBA/2J mice and 15 samples from A/J mice were used for this set of experiments. Not all genes

were examined from a single cDNA sample, but the expression of Act was always analyzed as a control gene together with genes of interest. **Table 1** shows the cCP values of each experimental set (BK subunits with Act). Small discrepancies were observed in the final number of samples between the DART and FPLM analyses. Because DART and FPLM use different criteria for determining amplification efficiency and CP, some samples were excluded by one method, but not necessarily by another method. Thus, the numbers of tests included for the statistics differ between the DART method and the FPLM method.

Comparisons between the two strains were made using the differences of cCP (Δ cCP) between the gene of interest and Act (**Figures 1A,B**). REST analysis was also performed for the comparison between the two strains using Act as a reference gene (**Figure 1C**). The results of both methods (Δ cCP and REST) clearly showed that BK α subunits and BK β 2 subunits were expressed significantly less in the CB of the A/J mice compared to that of the DBA/2J mice. No differences were observed in BK β 4 subunit expression between the strains.

EXPRESSION OF BK CHANNEL SUBUNITS IN THE SCG AND THE CAROTID ARTERY

We examined five SCGs and four carotid arteries in each strain. To show the difference of the gene expression pattern, CBs were also examined. Clear bands of expected sizes for the BK α , BK β 2, and BK β 4 subunits together with Act were observed in the CB samples (**Figure 2A**), showing the validity of the methods. The density of the each band was normalized relative to the band of Act in the same sample (**Figure 2B**). The decreased expression in BK α and BK β 2 subunits in the CB of the A/J mice compared to the DBA/2J mice was observed (data not shown). BK β 4 subunits were the dominant β subunits in the SCG and the band of BK β 2 subunits was weakly seen. No significant differences in the expression of BK channel subunits were observed between the strains. In the carotid artery, the BK α subunit was expressed without significant differences between the strains. As expected (Gribkoff et al., 2001; Orio et al., 2002), no expression of BK β 2 and BK β 4 subunits were observed in the artery.

BK CHANNEL ACTIVITY

After collagenase perfusion, GCs can be readily visualized as shown in **Figure 3B** and giga ohm seals were routinely obtained. The whole cell configuration was established, Cm was measured by

Table 1 | Gene expression of BK channel subunits: cCP values.

Genes	BK α		Act		BK β 2		Act		BK β 4		Act	
	DBA	A/J	DBA	A/J	DBA	A/J	DBA	A/J	DBA	A/J	DBA	A/J
DART METHOD												
<i>n</i>	10	9	10	9	10	7	10	7	10	7	10	7
Mean	28.99	30.83	19.41	19.57	27.96	30.67	20.06	19.76	32.81	31.62	20.57	20.21
SE	0.64	0.88	0.35	0.68	0.46	1.08	0.41	0.80	0.69	1.05	0.55	0.81
FPLM METHOD												
<i>n</i>	11	9	11	9	10	7	10	7	8	7	8	7
Mean	29.26	31.41	19.85	20.06	26.8	28.51	19.81	19.63	32.20	32.12	20.09	20.70
SE	0.68	1.02	0.25	0.74	0.51	0.83	0.18	0.72	0.73	0.69	0.27	0.92

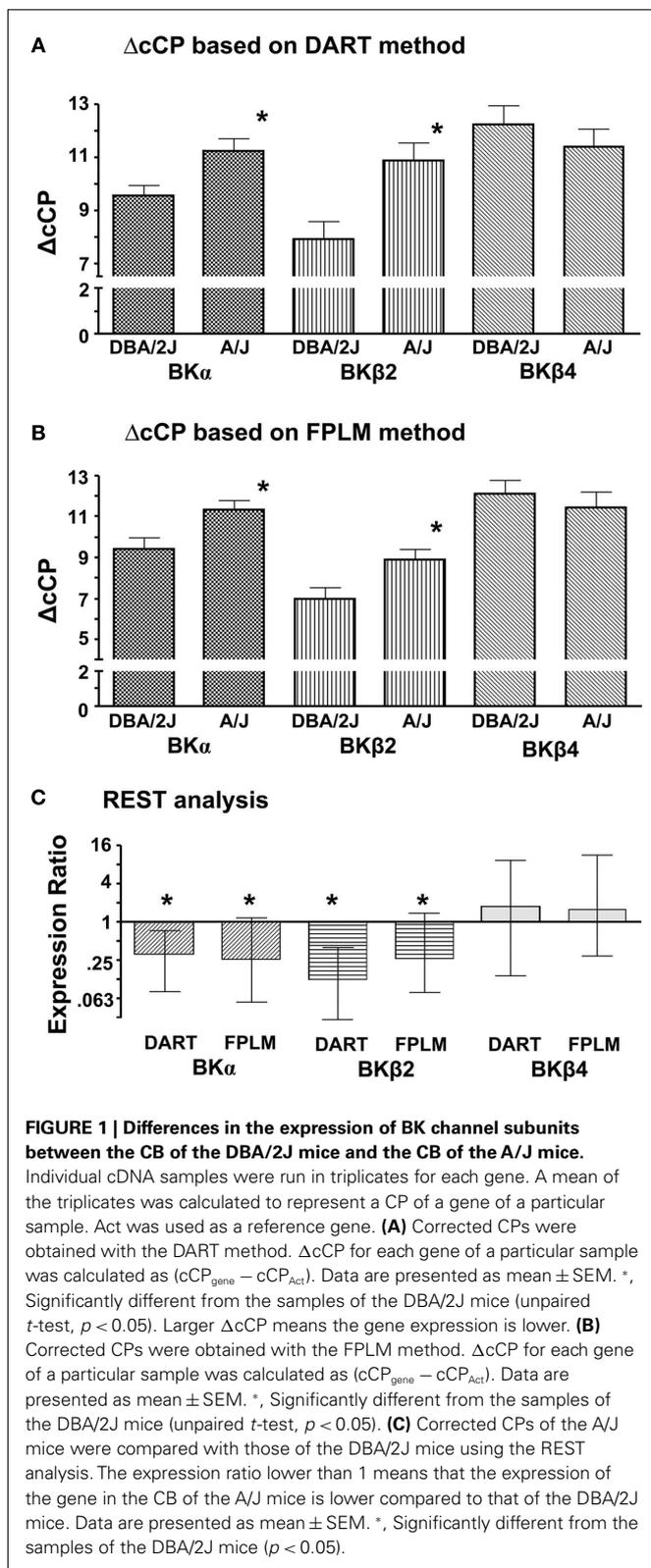


FIGURE 1 | Differences in the expression of BK channel subunits between the CB of the DBA/2J mice and the CB of the A/J mice. Individual cDNA samples were run in triplicates for each gene. A mean of the triplicates was calculated to represent a CP of a gene of a particular sample. Act was used as a reference gene. **(A)** Corrected CPs were obtained with the DART method. ΔcCP for each gene of a particular sample was calculated as $(cCP_{gene} - cCP_{Act})$. Data are presented as mean \pm SEM. *, Significantly different from the samples of the DBA/2J mice (unpaired *t*-test, $p < 0.05$). Larger ΔcCP means the gene expression is lower. **(B)** Corrected CPs were obtained with the FPLM method. ΔcCP for each gene of a particular sample was calculated as $(cCP_{gene} - cCP_{Act})$. Data are presented as mean \pm SEM. *, Significantly different from the samples of the DBA/2J mice (unpaired *t*-test, $p < 0.05$). **(C)** Corrected CPs of the A/J mice were compared with those of the DBA/2J mice using the REST analysis. The expression ratio lower than 1 means that the expression of the gene in the CB of the A/J mice is lower compared to that of the DBA/2J mice. Data are presented as mean \pm SEM. *, Significantly different from the samples of the DBA/2J mice ($p < 0.05$).

applying a short 10 mV pulse from holding potential -80 mV, and current–voltage relationships for K current was obtained as described above. Mean membrane capacitance of GCs in the

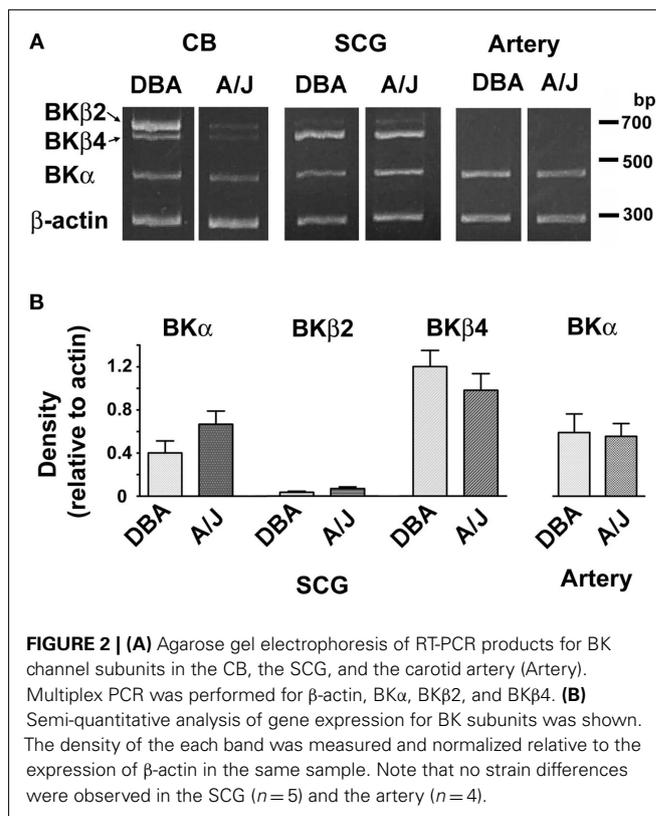
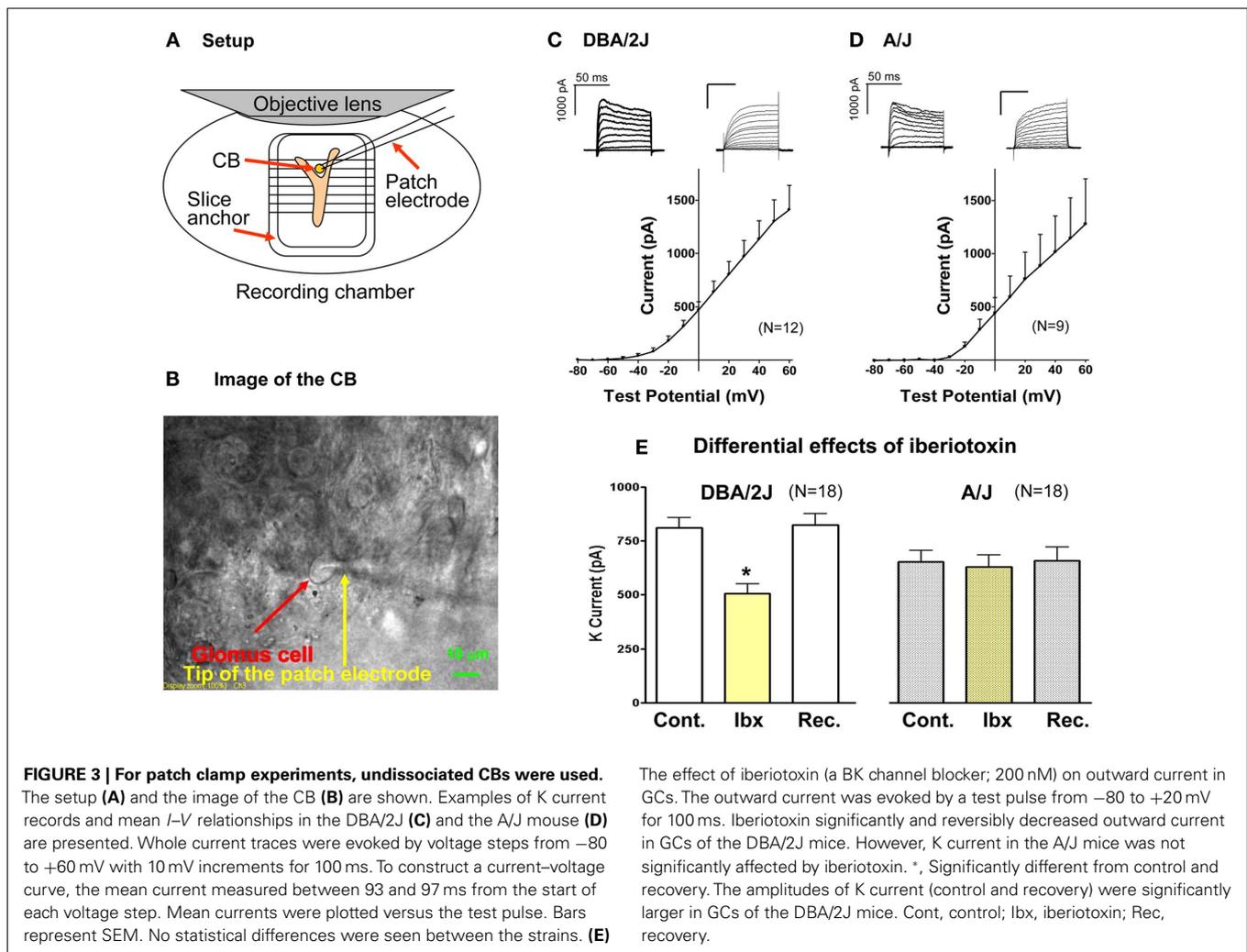


FIGURE 2 | (A) Agarose gel electrophoresis of RT-PCR products for BK channel subunits in the CB, the SCG, and the carotid artery (Artery). Multiplex PCR was performed for β -actin, BK α , BK $\beta 2$, and BK $\beta 4$. **(B)** Semi-quantitative analysis of gene expression for BK subunits was shown. The density of the each band was measured and normalized relative to the expression of β -actin in the same sample. Note that no strain differences were observed in the SCG ($n = 5$) and the artery ($n = 4$).

DBA/2J mice was 5.9 ± 1.7 pF ($n = 12$) and that in the A/J mice was 5.9 ± 1.3 pF ($n = 11$). These values were not significantly different. Depolarization induced outward currents with activation threshold around -40 to -30 mV in both strains. Variations in current size and kinetics were seen in both strains (**Figures 3C,D**). When inactivation was seen, it was slight and slow during the 100-ms depolarizing pulse in both strains. In some cells no inactivation was observed. Examples of the current records and mean $I-V$ relationships were shown in **Figures 3C,D**. No statistical differences in the $I-V$ relationships were found between the strains. To examine the BK channel component, a single step protocol was applied with or without iberiotoxin (200 nM; Alomone Laboratories), a selective blocker for BK channels. After 5 min superfusion with iberiotoxin-containing Krebs, K current in GCs of the DBA/2J mice was significantly attenuated ($63 \pm 5\%$ of control; $n = 18$) and the effect was reversible ($104 \pm 6\%$ of control; **Figure 3E**). On the other hand, K current in GCs of the A/J mice was not significantly affected by iberiotoxin 200 nM ($98 \pm 5\%$ of control; **Figure 3E**).

In other set of GCs, hypoxic sensitivity of voltage-dependent outward K current was tested by applying hypoxic Krebs solution. Mild hypoxia ($PO_2 \sim 70$ mmHg) clearly inhibited outward K current in GCs of the DBA/2J mice, but this level of hypoxia did not affect outward K current in GCs of the A/J mice (**Figure 4A**). The inhibitory effect was reversible. The effect of O_2 tension on K current was further studied using single pulse protocol (**Figure 4B**). K current was evoked at 1, 3, and 5 min after changing Krebs from normoxia (equilibrated with 21% $O_2/5\%$ CO_2) to hypoxia (equilibrated with 0% $O_2/5\%$ CO_2). PO_2 in the chamber was



measured in the experiments simulating the patch clamp experiments. PO_2 reached 70 mmHg after 5 min of hypoxic superfusion. The degree of inhibition of K current in the DBA/2J mice was dependent on PO_2 values in the chamber (Figure 4B, solid line) between the range of 150 and 70 mmHg, while no significant changes were noted in the A/J mice. To evaluate the contribution of BK channels to hypoxic sensitivity, the effect of hypoxia on K current was monitored in the presence of iberiotoxin (200 nM). Mild hypoxia ($PO_2 \sim 70$ mmHg) did not significantly influence K current in the presence of iberiotoxin in either strain of mice (Figure 4C).

DISCUSSION

In this study we have found that gene expression of BK channel α and β_2 subunits in the CB differs between the DBA/2J and the A/J strains of mice. BK channels consist of the four α subunits, which form the pore of the channel, and the auxiliary β subunits (β_1-4). Although α subunits alone can form functional channels when expressed in some cell lines, in native tissues they are likely associated with β subunits. Beta subunits significantly influence pharmacology and kinetics of BK channels, and the tissue distribution is distinct. Beta1 mRNA is high in smooth

The effect of iberiotoxin (a BK channel blocker; 200 nM) on outward current in GCs. The outward current was evoked by a test pulse from -80 to $+20$ mV for 100 ms. Iberiotoxin significantly and reversibly decreased outward current in GCs of the DBA/2J mice. However, K current in the A/J mice was not significantly affected by iberiotoxin. *, Significantly different from control and recovery. The amplitudes of K current (control and recovery) were significantly larger in GCs of the DBA/2J mice. Cont, control; Ibx, iberiotoxin; Rec, recovery.

muscle; β_2 , in chromaffin cells and brain; β_3 , in testis, pancreas, and spleen; and β_4 , in brain (Gribkoff et al., 2001; Orio et al., 2002). Because the parenchymal cells of the CB originate from the neural crest, we focused on α , β_2 , and β_4 mRNA levels in the CB of the two strains of mice. Real-time PCR analysis of these subunits showed that mRNA for α and β_2 subunits was significantly more expressed in the CB of the DBA/2J than those of the A/J mice (Figure 1).

In our previous comparative gene expression data, BK channel β_2 subunits are more expressed in the CB of the DBA/2J than that of the A/J mice at 4 weeks of age (Balbir et al., 2007). The current study utilized more vigorous evaluation with the unbiased estimation of amplification efficiency and cCP using the DART and the FPLM methods. The data agree with our previous observation regarding β subunits expression. The lower expression of BK α subunit gene further suggest that the BK channel numbers may be less in the CB of the A/J mice compared to that of the DBA/2J mice. However, one methodological concern in our gene expression analysis was the possible contamination with other tissues, such as the SCG and blood vessels. This is an inherent problem with the use of an extremely small tissue such as the mouse CB. Thus, we checked if the BK channel subunits were

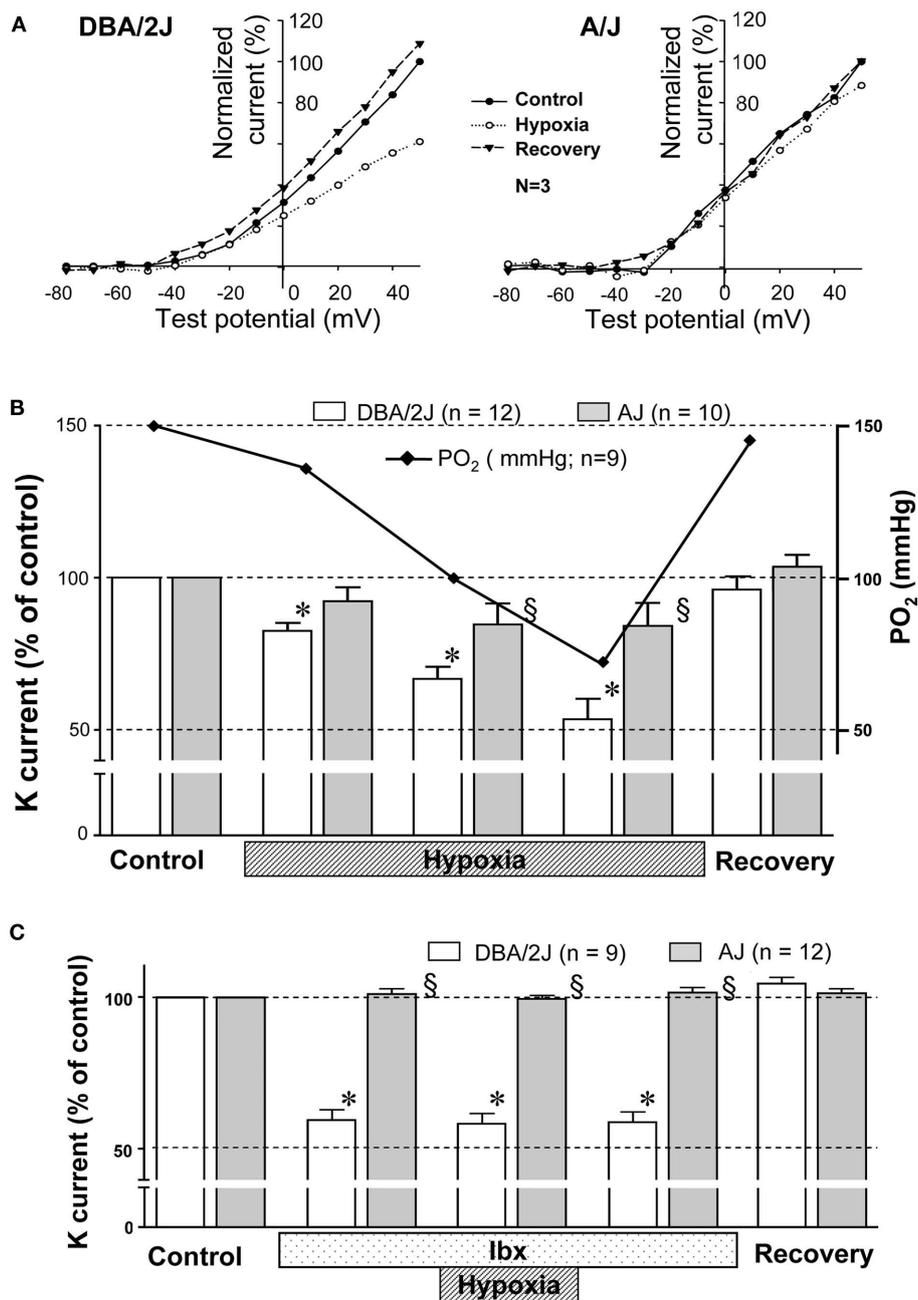


FIGURE 4 | (A) The effect of decreasing O₂ on voltage-dependent outward K current in GCs of the DBA/2J (left) and the A/J mice (right). Whole current was evoked by voltage steps from -80 to +50 mV with 10 mV increments for 100 ms. To construct a current-voltage curve, the mean current measured between 93 and 97 ms from the start of each voltage step was plotted versus the test pulse. The current during hypoxia was recorded after 5–10 min of superfusion with hypoxic Krebs solution. Each current was normalized as % of the current measured at +50 mV of test pulse during normoxia. I–V curves were mean values obtained from three GCs in each strain. **(B)** During control, the CB was superfused with Krebs equilibrated with 5% CO₂/air. Subsequently, the superfusate was switched to Krebs equilibrated with 5% CO₂/0% O₂, and K current was recorded at 1, 3, and 5 min (marked as “Hypoxia”). PO₂ in the chamber was measured in the experiments simulating the patch clamp experiments. K current was almost linearly inhibited with

decreasing PO₂ in GCs of the DBA/2J mice, but not those of the A/J mice. *, Significantly different from control and recovery ($p < 0.01$). §, Significantly different from the DBA/2J mice. There is no statistical difference between the control and the recovery. **(C)** The effect of decreasing O₂ on voltage-dependent K current of GCs in the DBA/2J and the A/J mice with iberiotoxin (Ibx). Initially the CB was superfused with Krebs equilibrated with 5% CO₂/air. Then, Krebs was changed to that containing 200 nM iberiotoxin for 5 min. K current was significantly reduced in GCs of the DBA/2J mice, but not of the A/J mice. Subsequently, the superfusate was switched to Krebs containing iberiotoxin and equilibrated with 5% CO₂/0% O₂ for 5 min. K current was not further affected in either strain. The effect of iberiotoxin was reversible. *, Significantly different from control and recovery ($p < 0.01$). §, Significantly different from the DBA/2J mice. There is no statistical difference between the control and the recovery.

differentially expressed in these tissues between the two strains of mice. No differences in the expression of BK α subunit were observed between the strains in the SCG and the carotid artery. Further, the expression of BK β 2 subunits in the SCG was very weak and indifferent between the strains. The expression of BK β 4 subunits in the SCG was not different between the strains as well. These β subunits were not detected in the carotid artery as shown in previous studies (Gribkoff et al., 2001; Orio et al., 2002). Based on these data, it is reasonable to conclude that the strain differences in the gene expression of BK α and β 2 subunits exist in the CB.

The electrophysiology data may reflect the differences in BK channel gene expression between the strains. Iberiotoxin, a selective BK channel antagonist, inhibited K current of GCs in the DBA/2J mice, but not in the A/J mice (Figure 3E). However, RT-PCR analysis (Figure 1; Table 1) suggests that α subunits of BK channels are present, although a smaller amount, in the GCs of the A/J mice. Although speculative, the ineffectiveness of iberiotoxin on K current in GCs of the A/J mice may be, at least in part, explained by the differences in the expression of BK β subunits. Because BK α and β 2 subunits were less expressed in GCs of the A/J mice than those of the DBA/2J mice, relative expression of β 4 subunits to β 2 subunits as well as relative expression of β 4 subunits to α subunits would be much higher in GCs of the A/J mice compared to GCs of the DBA/2J mice. Thus, β 4 subunits would contribute more as an associated protein to BK channels in GCs of the A/J mice. It is well known that β subunits have significant influence on the electrophysiological and pharmacological diversity of BK channels. Particularly, β 4 subunits have unique influence on BK channel activity. When α subunits are expressed together with β 4 subunits in HEK293 or CHO cells, $I-V$ curve of BK current is shifted to the right by 50 mV. Iberiotoxin or charybdotoxin, which inhibits α subunit alone or $\alpha+\beta$ 1 BK channel activity at low nanomolar range, is not effective in $\alpha+\beta$ 4 BK channels until high concentrations (micromolar) are used (Meera et al., 2000; Weiger et al., 2000). Further, β 4 subunits decrease BK channel openings at low calcium concentrations (Brenner et al., 2000). If this is the case in the CB, BK channels in GCs of the A/J mice may not have been well activated in our experimental conditions. Their insensitivity to iberiotoxin may be due to a close association of BK α subunits with β 4 subunits.

Hypoxic inhibition of voltage-dependent K channels, including BK channels, has been proposed as an early event leading to depolarization of GCs. However, the role of voltage-dependent K channels in GC depolarization in response to hypoxia has been still debated (Buckler, 2007; Lopez-Lopez and Perez-Garcia, 2007; Peers and Wyatt, 2007). Important questions are (1) whether voltage-dependent K channels including BK channels are active at or close to the resting membrane potential of GCs, and (2) whether the inhibition of K channels are fast enough to precede the onset of an increase in chemoreceptor discharge. Wyatt and Peers (1995) have shown that charybdotoxin, a specific BK channel blocker, depolarized rat GCs without other stimuli, indicating that BK channels are active at the resting membrane potential. Consistent with this finding, Pardal et al. (2000) reported that

TEA and iberiotoxin increased catecholamine secretion from rat GCs. In a recent review, Peers and Wyatt (2007) further provided detailed discussion which supports a role of BK channels initiating depolarization of GCs during hypoxia. They further argued that hypoxic inhibition of BK channels will maintain depolarization of GCs due to the delay of GC repolarization. In terms of the speed, hypoxic inhibition of voltage-dependent K channels developed faster than tetrodotoxin-induced inhibition of Na channels in the rabbit GCs (Lopez-Lopez and Gonzalez, 1992). Although not studied in detail, the speed of BK channel inhibition was comparable between the hypoxic superfusion and the charybdotoxin superfusion (Wyatt and Peers, 1995). The data indicates that a small drop of O₂ tension acts in the order of seconds to inhibit BK channels as charybdotoxin does. We observed K current inhibition at 1 min after switching the superfusate from normoxia to hypoxia (Figure 4B), indicating that BK channel inhibition by lowering O₂ develops quickly.

Our previous study (Balbir et al., 2007) showed that the gene expression of TASK-like K channels were indifferent between the DBA/2J and the A/J strains. Therefore, in our experimental model using these mice, we did not focus on these channels. Our current data clearly indicate that mild hypoxia mainly inhibits BK channels in GCs of the DBA/2J mouse: with application of iberiotoxin, K current was inhibited, and mild hypoxia did not further influence K current. However, our results do not mean that other oxygen-sensitive K channels are not important in hypoxic chemotransduction processes. Several types of oxygen-sensitive K channels in the CB have been identified in several species. These include BK channels and background TASK-like K channels in the rat (Williams and Buckler, 2004), Kv4.1 and Kv4.3 in the rabbit (Sanchez et al., 2002), and Kv3 in the C57Bl/6J mouse (Perez-Garcia et al., 2004). Most of these studies used severe hypoxia such as PO₂ ~ 10 mmHg. The relationship between PO₂ and K current inhibition was investigated in some studies: oxygen-sensitive K current decreased lineally in the rabbit (Montoro et al., 1996) and in the cat (Shirahata and Sham, 1999), or maximal inhibition occurred at PO₂ of 80 mmHg in the rabbit (Lopez-Lopez et al., 1989). These data suggest a possibility that distinct K channels may be inhibited at different PO₂ levels.

It has been long known that hypoxic sensitivity among individuals is variable (Hirshman et al., 1975). Human studies in twins and in individuals over time suggest that hypoxic sensitivity of the CB is genetically controlled (Collins et al., 1978; Kawakami et al., 1982; Nishimura et al., 1991; Thomas et al., 1993). Inbred strains of mice are useful tools to investigate genetic background of physiological phenomena, and several studies have shown differential hypoxic responses between the A/J (lower sensitivity) and the DBA/2J (higher sensitivity) mice (Tankersley et al., 1994; Rubin et al., 2003; Campen et al., 2004). The current study provided some molecular mechanisms of the differential hypoxic sensitivity of the CB in these mice.

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