



Reversing autism by targeting downstream mTOR signaling

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A commentary on

Autism-related deficits via dysregulated eIF4E-dependent translational control

by Gkogkas, C. G., Khoutorsky, A., Ran, I., Rampakakis, E., Nevarko, T., Weatherill, D. B., et al. (2013). *Nature* 493, 371–377.

Autism spectrum disorders (ASDs) are a group of clinically and genetically heterogeneous neurodevelopmental disorders characterized by impaired social interactions, repetitive behaviors and restricted interests (Baird et al., 2006; Zoghbi and Bear, 2012). The genetic defects in ASDs may interfere with synaptic protein synthesis. Synaptic dysfunction caused by aberrant protein synthesis is a key pathogenic mechanism for ASDs (Kelleher and Bear, 2008; Richter and Klann, 2009; Ebert and Greenberg, 2013). Understanding the details about aberrant synaptic protein synthesis is important to formulate potential treatment for ASDs. The mammalian target of the rapamycin (mTOR) pathway plays central roles in synaptic protein synthesis (Hay and Sonenberg, 2004; Hoeffler and Klann, 2010; Hershey et al., 2012). Recently, Gkogkas and colleagues published exciting data on the role of downstream mTOR pathway in autism (Gkogkas et al., 2013) (Figure 1).

Previous studies have indicated that upstream mTOR signaling is linked to ASDs. Mutations in tuberous sclerosis complex (*TSC*) 1/*TSC2*, neurofibromatosis 1 (*NF1*), and Phosphatase and tensin homolog (*PTEN*) lead to syndromic ASD with tuberous sclerosis, neurofibromatosis, or macrocephaly, respectively (Kelleher and Bear, 2008; Bourgeron, 2009; Hoeffler

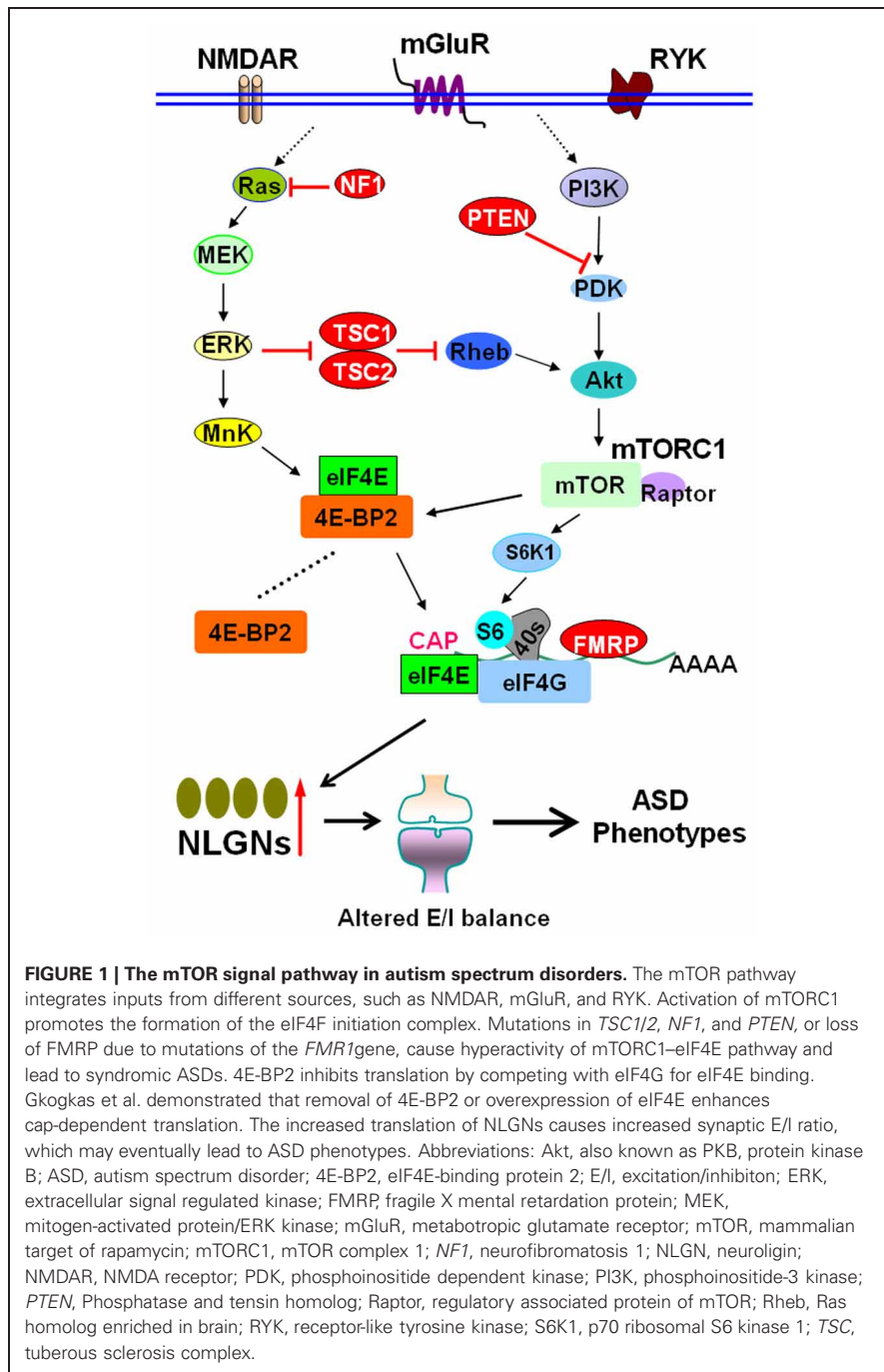
and Klann, 2010; Sawicka and Zukin, 2012). *TSC1/TSC2*, *NF1*, and *PTEN* act as negative regulators of mTOR complex 1 (mTORC1), which is activated by phosphoinositide-3 kinase (PI3K) pathway (Kelleher and Bear, 2008; Auerbach et al., 2011; Sawicka and Zukin, 2012) (Figure 1). Activation of cap-dependent translation is a principal downstream mechanism of mTORC1. The eIF4E recognizes the 5' mRNA cap, recruits eIF4G and the small ribosomal subunit (Richter and Sonenberg, 2005; Hershey et al., 2012). The eIF4E-binding proteins (4E-BPs) bind to eIF4E and inhibit translation initiation. Phosphorylation of 4E-BPs by mTORC1 promotes eIF4E release and initiates cap-dependent translation (Richter and Klann, 2009; Hoeffler and Klann, 2010) (Figure 1). A hyperactivated mTORC1–eIF4E pathway is linked to impaired synaptic plasticity in fragile X syndrome, an autistic disorder caused by lack of fragile X mental retardation protein (FMRP) due to mutation of the *FMR1* gene (Wang et al., 2010; Auerbach et al., 2011; Santoro et al., 2012; Wang et al., 2012), suggesting that downstream mTOR signaling might be causally linked to ASDs. Notably, one pioneering study has identified a mutation in the *EIF4E* promoter in autism families (Neves-Pereira et al., 2009), implying that deregulation of downstream mTOR signaling (eIF4E) could be a novel mechanism for ASDs.

As an eIF4E repressor downstream of mTOR, 4E-BP2 has important roles in synaptic plasticity, learning and memory (Banko et al., 2005; Richter and Klann, 2009). Writing in their *Nature* article, Gkogkas and colleagues reported that deletion of the gene encoding 4E-BP2

(*Eif4ebp2*) leads to autistic-like behaviors in mice. Pharmacological inhibition of eIF4E rectifies social behavior deficits in *Eif4ebp2* knockout mice (Gkogkas et al., 2013). Their study in mouse models has provided direct evidence for the causal link between dysregulated eIF4E and the development of ASDs.

Are these ASD-like phenotypes of the *Eif4ebp2* knockout mice caused by altered translation of a subset mRNAs due to the release of eIF4E? To test this, Gkogkas et al. measured translation initiation rates and protein levels of candidate genes known to be associated with ASDs in hippocampi from *Eif4ebp2* knockout and eIF4E-overexpressing mice. They found that the translation of neuroigin (*NLGN*) mRNAs is enhanced in both lines of transgenic mice. Removal of 4E-BP2 or overexpression of eIF4E increases protein amounts of *NLGN*s in the hippocampus, whereas mRNA levels are not affected, thus excluding transcriptional effects (Gkogkas et al., 2013). In contrast, the authors did not observe any changes in the translation of mRNAs coding for other synaptic scaffolding proteins. Interestingly, treatment of *Eif4ebp2* knockout mice with selective eIF4E inhibitor reduces *NLGN* protein levels to wild-type levels (Gkogkas et al., 2013). These data thus indicate that relief of translational suppression by loss of 4E-BP2 or by the overexpression of eIF4E selectively enhances the *NLGN* synthesis. However, it cannot be ruled out that other proteins (synaptic or non-synaptic) may be affected and contribute to animal autistic phenotypes.

Aberrant information processing due to altered ratio of synaptic excitation to inhibition (E/I) may contribute to



ASDs (Rubenstein and Merzenich, 2003; Bourgeron, 2007; Uhlhaas and Singer, 2012). The increased or decreased E/I ratio has been observed in ASD animal models (Chao et al., 2010; Bateup et al., 2011; Luikart et al., 2011; Schmeisser et al., 2012). In relation to these E/I shifts, Gkogkas et al. then examined the synaptic transmission in hippocampal slices of *Eif4ebp2* knockout mice. They found that 4E-BP2 de-repression results

in an increased E/I ratio, which can be explained by the increase of vesicular glutamate transporter and spine density in hippocampal pyramidal neurons. As expected, application of eIF4E inhibitor restores the E/I balance (Gkogkas et al., 2013).

Finally, in view of the facts that genetic manipulation of NLGNs results in ASD-like phenotypes with altered E/I balance in mouse models (Chubykin et al.,

2007; Tabuchi et al., 2007; Etherton et al., 2011) and NLGN mRNA translation is enhanced concomitant with increased E/I ratio in *Eif4ebp2* knockout mice, Gkogkas et al. tested the effect of NLGN knockdown on synaptic plasticity and behaviour in these mice (Gkogkas et al., 2013). NLGN1 is predominantly postsynaptic at excitatory synapses and promotes excitatory synaptic transmission (Varoqueaux et al., 2006; Kwon et al., 2012). The authors found that NLGN1 knockdown reverses changes at excitatory synapses and partially rescues the social interaction deficits in *Eif4ebp2* knockout mice (Gkogkas et al., 2013). These findings thus established a strong link between eIF4E-dependent translational control of NLGNs, E/I balance and the development of ASD-like animal behaviors (Figure 1).

In summary, Gkogkas et al. have provided a model for mTORC1/eIF4E-dependent autism-like phenotypes due to dysregulated translational control (Gkogkas et al., 2013). This novel regulatory mechanism will prompt investigation of downstream mTOR signaling in ASDs, as well as expand our knowledge of how mTOR functions in human learning and cognition. It may narrow down therapeutic targets for autism since targeting downstream mTOR signaling reverses autism. Pharmacological manipulation of downstream effectors of mTOR (eIF4E, 4E-BP2, and NLGNs) may eventually provide therapeutic benefits for patients with ASDs.

ACKNOWLEDGMENTS

Hansen Wang was supported by the National Natural Science Foundation of China (NSFC, No.30200152) for Rett syndrome studies and a postdoctoral fellowship from the Fragile X Research Foundation of Canada. Laurie Doering was supported by the Natural Sciences and Engineering Research Council of Canada (NSERC) and the Fragile X Research Foundation of Canada.

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Received: 17 January 2013; accepted: 05 March 2013; published online: 26 March 2013.

Citation: Wang H and Doering LC (2013) Reversing autism by targeting downstream mTOR signaling. *Front. Cell. Neurosci.* 7:28. doi: 10.3389/fncel.2013.00028

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