# Astrocytic modulation of blood brain barrier: perspectives on Parkinson's disease

Ricardo Cabezas¹, Marcos Ávila¹, Janneth Gonzalez¹, Ramon Santos El-Bachá², Eliana Báez¹, Luis Miguel García-Segura³, Juan Camilo Jurado Coronel¹, Francisco Capani⁴, Gloria Patricia Cardona-Gomez⁵ and George E. Barreto¹\*

- Departamento de Nutrición y Bioquímica, Facultad de Ciencias, Pontificia Universidad Javeriana, Bogotá, D.C., Colombia
- <sup>2</sup> Departamento de Biofunção, Universidade Federal da Bahia, Salvador, Brazil
- <sup>3</sup> Instituto Cajal, CSIC, Madrid, Spain
- <sup>4</sup> Laboratorio de Citoarquitectura y Plasticidad Neuronal, Facultad de Medicina, Instituto de Investigaciones cardiológicas Prof. Dr. Alberto C. Taquini (ININCA), UBA-CONICET. Buenos Aires. Argentina
- <sup>5</sup> Cellular and Molecular Neurobiology Area, Group of Neuroscience of Antioquia, Faculty of Medicine, SIU, University of Antioquia UdeA, Medellín, Colombia

#### Edited by:

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#### \*Correspondence:

George E. Barreto, Departamento de Nutrición y Bioquímica, Facultad de Ciencias, Pontificia Universidad Javeriana, Edf. Carlos Ortiz, Oficina 107, Cra 7 40-62, Bogotá, D.C., Colombia

e-mail: gsampaio@javeriana.edu.co

The blood-brain barrier (BBB) is a tightly regulated interface in the Central Nervous System (CNS) that regulates the exchange of molecules in and out from the brain thus maintaining the CNS homeostasis. It is mainly composed of endothelial cells (ECs), pericytes and astrocytes that create a neurovascular unit (NVU) with the adjacent neurons. Astrocytes are essential for the formation and maintenance of the BBB by providing secreted factors that lead to the adequate association between the cells of the BBB and the formation of strong tight junctions. Under neurological disorders, such as chronic cerebral ischemia, brain trauma, Epilepsy, Alzheimer and Parkinson's Diseases, a disruption of the BBB takes place, involving a lost in the permeability of the barrier and phenotypical changes in both the ECs and astrocytes. In this aspect, it has been established that the process of reactive gliosis is a common feature of astrocytes during BBB disruption, which has a detrimental effect on the barrier function and a subsequent damage in neuronal survival. In this review we discuss the implications of astrocyte functions in the protection of the BBB, and in the development of Parkinson's disease (PD) and related disorders. Additionally, we highlight the current and future strategies in astrocyte protection aimed at the development of restorative therapies for the BBB in pathological conditions.

Keywords: BBB, astrocytes, reactive astrogliosis, endothelial cells, Parkinson disease

#### INTRODUCTION

The Blood Brain Barrier (BBB) is an essential regulatory component of the neural interface with the brain vasculature. It exerts a tightly regulation in the movement of ions, molecules and cells between the neural cells and the blood (Daneman, 2012; Wong et al., 2013), thus maintaining the ionic homeostasis, hormonal and transmitter levels and transport of nutrients in the brain (Luissint et al., 2012). In this aspect, BBB is important for the separation of neurotransmitters pools and neuroactive agents that regulate brain microenvironment (Abbott et al., 2006). Furthermore, the BBB supplies the brain with different nutrients, exerts a restriction of ionic substances between the blood and the brain through specific ion transporters, regulates the ISF (interstitial fluid), prevents the formation of additional injuries during diseases and cerebrovascular accidents and is an important barrier for the brain transport and metabolization of drugs (Abbott et al., 2006; Daneman, 2012; Wong et al., 2013).

The BBB is composed by brain capillary endothelial cells (ECs), with a specific phenotype located in a strong association with astrocytic endfeet processes and mesenchymal-like

cells pericytes. Importantly, the BBB is characterized by the presence of tight junctions between ECs, and the expression of specific polarized transport systems (Luissint et al., 2012). On the other hand, astrocytes through their endfeet establish the link between the endothelial blood flux and neurons, and are important regulators in the formation and maintenance of the BBB (Alvarez et al., 2013). BBB dysfunction has been associated with pathological conditions and diseases including cerebral ischemia, brain trauma, glioblastoma, stroke, multiple sclerosis, epilepsy, Alzheimer and Parkinson's Disease (PD; Haseloff et al., 2005; Daneman, 2012; Alvarez et al., 2013).

PD is a progressive neurodegenerative disorder caused by neuronal death in substantia nigra (SN), degeneration of dopaminergic neurotransmission, and the presence of  $\alpha$ -synuclein and protein inclusions in neuronal cells, also known as Lewy bodies (Nutt and Wooten, 2005; Halliday and Stevens, 2011). Main symptoms of Parkinson include asymmetrical bradikinesia, rigidity, resting tremor and postural instability (Fernandez, 2012; Singer, 2012). Initiation and progression of PD is dependent upon cellular events, such as failures in the

protein degradation machinery, oxidative stress, mitochondrial dysfunction, defects in mitochondrial autophagy (mitophagy) and the continuous accumulation of  $\alpha$ -synuclein, driven through cell to cell interactions between glial cells and neurons that ultimately lead to apoptosis (Jenner, 2003; Halliday and Stevens, 2011; Vives-Bauza and Przedborski, 2011). Although there is not a cure for the disease, the most used and cheaper treatment for PD continues to be Levodopa, frequently accompanied by carbidopa or benserazide (Singer, 2012; Ossig and Reichmann, 2013). However, about 40% of patients developed motor fluctuations and dyskinesias after 4 to 6 years of treatment (Ogawa et al., 2005; Fernandez, 2012), demonstrating that further pharmacological research is needed in order to counterbalance these side effects.

Current research suggests that the exact cause of PD remains unknown (Hirsch et al., 2003; Fernandez, 2012; Schwartz and Sabetay, 2012). Mutations in various proteins such as LRRK2, PARK2), phosphatase and tensin homolog (PTEN)-induced putative kinase 1 (PINK1), and (DJ-1) have been observed in familiar cases of Parkinson, which only account for 10-15% of diagnosed cases (Hirsch et al., 2003; Rappold and Tieu, 2010; Pan-Montojo et al., 2010; Wang et al., 2011). Similarly, various environmental factors have been found to induce PD-like symptoms, including vascular insults to the brain, oxidative stress, neuroleptic drugs, heavy metals exposure and the exposure to pesticides like rotenone or paraguat (Betarbet et al., 2000; Brown et al., 2006; Rappold and Tieu, 2010; Tanner et al., 2011). Similarly, there is clinical and in vitro evidence of BBB disruption during PD development (Kortekaas et al., 2005; Hirano et al., 2008; Ohlin et al., 2011; Lee and Pienaar, 2014). In this aspect, previous studies have suggested that α-synuclein deposition has an increase in BBB permeability (Jangula and Murphy, 2013), suggesting the importance of α-synuclein in BBB disruption and PD development. (Braak et al., 2006; Halliday and Stevens, 2011).

A great body of research has shown the importance of astrocytes in the maintenance of BBB properties both during normal and pathological conditions (Ramaswamy and Kordower, 2009; Yasuda and Mochizuki, 2010; Alvarez et al., 2013). Astrocytic secreted molecules are important for the regulation of interactions between BBB components such as ECs and pericytes (Alvarez et al., 2013; Lee and Pienaar, 2014). Furthermore, astrocytes produce antioxidative molecules like GSH, ascorbate and SOD (superoxide dismutase) and a great number of growth factors and neurotrophins, important for brain cell survival during neurodegenerative processes (Dringen, 2000; Ramaswamy and Kordower, 2009; Yasuda and Mochizuki, 2010; Zheng et al., 2010; Barreto et al., 2011).

In the present review we provide a throughout overview of the astrocytic functions in the BBB and its importance during pathophysiological events elicited in PD. Additionally, we highlight the current and future strategies in astrocyte protection aimed at the development of restorative therapies for the BBB in pathological conditions.

# COMPONENTS OF THE BBB

# **ENDOTHELIAL CELLS**

ECs within the brain have a characteristic phenotype that makes them different from EC located elsewhere (Dejana, 2004;

Stamatovic et al., 2008; Nag, 2011; Daneman, 2012). For example, brain ECs have similarities with epithelial cells, as they are polarized cells that express some specific transporters and in that they are connected by circumferential tight junctions that interfere with the paracellular transport of molecules and ions between cells (Nag, 2011; Daneman, 2012). As well, brain EC have an increased density of mitochondria when compared with the peripheral vasculature, suggesting a higher risk of reactive oxygen species (ROS) formation (Nag, 2011; Lee and Pienaar, 2014). Structurally, EC are in contact with astrocytic endfeet and pericyte through the basal lamina, thus forming the neurovascular unit (NVU), with neurons (Hawkins and Davis, 2005; Stanimirovic and Friedman, 2012; Najjar et al., 2013).

Among its functions in BBB maintenance, EC are important in the bidirectional transport across the brain through ion transporters, protein and peptide carriers and active efflux transport (Nag, 2011). Furthermore, EC have highly organized tight and adherent junctions which restrict the passage of polar substances including hexose sugars, amino acids, nucleosides monocarboxylic acids, and vitamins (Grammas et al., 2011; Mokgokong et al., 2014). Importantly, the integrity of tight junctions is essential to prevent the paracellular transport of many molecules and ions, and its disruption is associated with pathological events in the brain such as microbial infection, cancer, inflammatory responses, stroke, Alzheimer disease and PD (Stamatovic et al., 2008; Luissint et al., 2012). Moreover, some studies have shown alterations in endothelial tight junctions during PD development (Kim et al., 2003; Chen et al., 2008; Lee and Pienaar, 2014). For example, Chen et al. (2008) found a decrease in the tight junction proteins occludin and ZO-1 in a MPTP murine model of PD. Similarly, the exposure of murine EC to ROS increased the activity of metalloproteinase-9 (MMP-9), which caused degradation of the basal lamina and BBB disruption. This oxidative damage was reduced by the overexpression of SOD1 and catalase, suggesting the importance of oxidative stress in BBB disruption (Kim et al., 2003). Additionally, there is in vitro and clinical evidence of angiogenic activity in PD development caused by an upregulation in the expression of vascular endothelial growth factor (VEGF; Wada et al., 2006; Lee and Pienaar, 2014). In summary, the cellular and molecular properties of brain ECs are essential for maintaining BBB permeability through an adequate ionic balance, conservation of the junctional structure and an adequate interaction with cells of the NVU.

# **PERICYTES**

Pericytes are enwrapping cells of blood microvessels, and are located between the EC and astrocytic endfeet and neurons (Wong et al., 2013). They are important regulatory cells for the maintenance of both homeostasis and hemostasis in the BBB (Dore-Duffy and Cleary, 2011). Additionally, pericytes are relevant in functions such as stromal regeneration, angiogenesis and neovascularization, antigen presenting cells under brain pathologies, control of EC proliferation, and promotion of neural stem cell properties (Lange et al., 2013; Elali et al., 2014; Hurtado-Alvarado et al., 2014). In this regard, pericytes have shown to differentiate *in vitro* into chondrocytes, vascular smooth muscles cells (VsMCS), osteoblasts and skeletal muscle, suggesting a

promising clinical use for pericytes in Central Nervous System (CNS) injuries and other pathologies (Armulik et al., 2010; Lange et al., 2013). Both pericytes and EC are enveloped by a basal membrane that is continuous between the two cell types, which separates pericytes from astrocyte endfeet (Sá-Pereira et al., 2012). This association is achieved through the endothelial secretion of PDGF-B and other angiogenic factors such as VEGF, TGF- $\beta$  and angiopoietins (Angs), through the interaction of multiple signaling pathways (Dore-Duffy, 2008; Armulik et al., 2010; Ribatti et al., 2011).

Morphologically, pericytes exhibit an oval cell body with a great number of projections that enwrap ECs in different patterns, along the abluminal surface (Armulik et al., 2010). The two main types of pericytes, granular (95% of total pericytes) and agranular have been described in the brain according to the presence or absence of lysosome granules in the cytoplasm. Interestingly, alterations in granular pericytes have been associated with amyloid deposition, and lipid accumulation in human brain cultures, suggesting the importance of pericyte alterations in Alzheimer disease and other pathologies (Castejón, 2011).

Of greater importance are the interactions between astrocytes and pericytes. In this aspect, it has been shown that both pericytes and astrocytes are essential for brain vasculogenesis and BBB maintenance possibly through the activation of PDGFRB signaling (Dejana, 2004; Bonkowski et al., 2011). Moreover, both pericytes and astrocytes are important in the preservation of EC tight junctions through the regulation of proteins like occludin, claudin and ZO-1 (zona occludens-1, Haseloff et al., 2005; Wolburg et al., 2009; Bonkowski et al., 2011). This result suggests the importance of astrocyte-pericyte communication in brain physiology. However, further research is needed in order to understand the implications of the mentioned interactions during neurodegenerative disorders.

# **ASTROCYTES**

Astrocytes are the most common cell type in the mammalian brain, conforming the glia with oligodendrocytes and microglia (Chen and Swanson, 2003). Among its many functions, astrocytes are essential for many metabolic processes in the brain such as the promotion of neurovascular coupling, the attraction of cells through the release of chemokines,  $K^+$  buffering, release of gliotransmitters, release of glutamate by calcium signaling, control of brain pH, metabolization of dopamine and other substrates by monoamine oxidases, uptake of glutamate and  $\gamma$ -aminobutyricacid (GABA) by specific transporters and production of antioxidant compounds like glutathione (GSH) and enzymes such as superoxide dismutases (SODs; Volterra and Meldolesi, 2005; Chinta and Andersen, 2008; Hamby and Sofroniew, 2010; Kimelberg and Nedergaard, 2010; Parpura et al., 2011).

Globally, astrocytes are characterized by the expression of the intermediate filaments vimentin (Vim) and glial fibrillary acidic protein (GFAP), which are upregulated under CNS insults, in a process known as astrogliosis (Volterra and Meldolesi, 2005; Hamby and Sofroniew, 2010; Céspedes et al., 2013). Morphologically, astrocytes are characterized by a stellate shape with

multiple processes and ramifications (Chen and Swanson, 2003; Volterra and Meldolesi, 2005), and become activated following brain injuries and degenerative diseases (Barreto et al., 2007, 2009, 2011, 2012; Adelson et al., 2012).

Although a great heterogeneity exists among astrocytes, two main types have been described in the CNS: protoplasmic astrocytes of the grey matter which envelope neuronal bodies and synapses, and fibrous astrocytes from the white matter that interact with the nodes of Ranvier and oligodendroglia (Halliday and Stevens, 2011; Oberheim et al., 2012). Current research has suggested that only protoplasmic astrocytes have an increase in the accumulation of  $\alpha$ -synuclein, and these are of importance for PD development (Braak et al., 2006; Halliday and Stevens, 2011). Interestingly, protoplasmic astrocytes are arranged in nonoverlapping domains forming a syncytial network that may contact approximately 160.000 synapses, thus integrating neural activity with the vascular network (Bushong et al., 2002; Barreto et al., 2011). This architecture is altered under pathological events such as Alzheimer and Epilepsia and has been associated with reactive astrogliosis (Oberheim et al., 2012), suggesting the importance of structural alterations during damaging processes.

Astrocytic terminal processes, known as endfeet, contact the brain vasculature surface facing ECs and pericytes and enwrap the neuronal synapses, enabling the modulation of both neuronal activity and cerebral blood flow, following an elevation in intracellular Ca<sup>2+</sup> levels in the endfeet (Zonta et al., 2003; Maragakis and Rothstein, 2006). Importantly, astrocytic endfeet express specialized molecules such as Kir4.1 K<sup>+</sup> channels and aquaporin 4 that regulate BBB ionic concentrations, and protein transporters such as glucose transporter-1 and P-glycoprotein, suggesting the importance of the endfeet in astrocyte polarization (Abbott et al., 2006; Nag, 2011). Additionally, astrocytes communicate between each other through gap junctions forming a functional syncitium with well-coordinated responses (Theis et al., 2005; Alvarez et al., 2013). In this aspect, it has been suggested that the astrocytic mechanisms that regulate vasodilation and vasoconstriction are transmitted through this inter-astroglial gap junctions (Alvarez et al., 2013). Furthermore, astrocytes are important in the development and maintenance of BBB characteristics in ECs through the release of growth factors like VEGF, glial cell line-derived neurotrophic factor (GDNF), basic fibroblast growth factor (bFGF), and ANG-1 (Alvarez et al., 2013; Wong et al., 2013). These growth factors are important in the formation of tight junctions, the promotion of enzymatic systems and the polarization of transporters (Wong et al., 2013). Astrocyte-secreted growth factors are also important for neuronal growth and maintenance, and have survival properties during brain damaging processes like PD (Hamby and Sofroniew, 2010).

# **EXTRACELLULAR MATRIX (ECM)**

In addition to the different cell types which constitute the BBB, the extracellular Matrix (ECM) is an important structural element of the BBB that serves as an anchor for the endothelium through the interaction of endothelial integrin receptors and matrix proteins such as laminin (Hawkins and Davis, 2005). In the brain, the ECM is composed of hyaluronan, hyaluronic acid, lecticans, proteoglicans and tenascins, which are important

for the maintenance of the paracellular diffusion in the BBB (Hawkins and Davis, 2005; Wong et al., 2013). Previous studies have suggested that the disruption of the ECM is strongly associated with an increase in BBB permeability during pathogenic states such as glioblastoma multiforme, ischemia and hemorrhagic necrosis of the brain. For example, during ischemia, the basement membrane suffers a breakdown caused by the increased expression of the matrix metalloproteinases (MMPs) MMP9 and MMP2 which in addition may cause microglial activation (del Zoppo and Milner, 2006; Lau et al., 2013). Furthermore, increased expression of MMP9 and GFAP in astrocytes was observed in a parkinsonian mouse model with MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; Annese et al., 2014). These results suggest the importance of ECM breakdown in glial activation during neurodegeneration and PD.

#### **PARKINSON DISEASE AND BBB**

#### **CAUSES OF DISRUPTION OF THE BBB**

Several processes may affect the integrity of the BBB, including an increase in ROS production, elevated levels of proinflammatory cytokines, inappropriate clearance of Ab peptide and other toxic substances (Minagar and Alexander, 2003; Popescu et al., 2009; van Sorge and Doran, 2012). Previous studies have shown an increase in BBB permeability associated with age that is in part responsible for pathological alterations such as white matter lesions (Simpson et al., 2007; Popescu et al., 2009). In this aspect, it has been reported that elder individuals and senescence mouse models have a higher albumin and IgG concentration than younger individuals caused by a leakage through the BBB (Popescu et al., 2009). Moreover, ageing is also associated with an increased production of ROS and proinflammatory cytokines in vascular ECs, which have been linked with memory and learning impairment in mouse models (Fukui et al., 2001; Popescu et al., 2009; Enciu and Popescu, 2013). Aged people have shown a diminished activity of the P-glycopotein efflux transporter that is associated with a limited removal of toxic substances from the brain (Popescu et al., 2009), demonstrating an important correlation between ageing processes (such as the increased expression of ROS) and BBB dysfunction. Taking into account that PD is associated with both age and ROS production, it is important to explore the cellular and molecular mechanisms that are activated during BBB disruption in this pathology and its protective mechanisms.

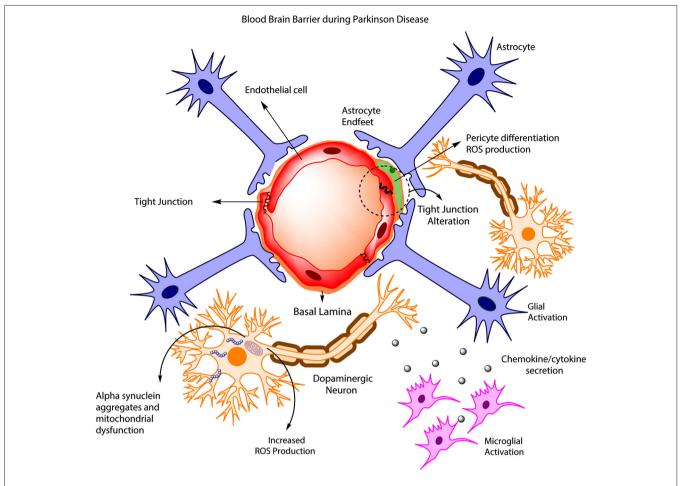
# DISRUPTION OF BBB IN PARKINSON DISEASE

Disruption of BBB in PD has been quite controversial. It was initially assumed that BBB remained unaltered during the development of the pathology, as observed in animal models and permeability studies of PD drugs such as levodopa and benserazide (Kurkowska-Jastrzebska et al., 1999; Haussermann et al., 2001). More recently, clinical studies have presented evidence of BBB disruption in PD patients (Kortekaas et al., 2005; Hirano et al., 2008; Ohlin et al., 2011; Lee and Pienaar, 2014). For example, an early study (Kortekaas et al., 2005) pointed out an increase in the brain uptake of drugs that usually do not cross the BBB including benzerazide and [\$^{11}C\$] verapamil in PD patients and rat models, suggesting a possible BBB breakdown. Additionally, a PET study (positron emission tomography) found deficiencies

in cerebral blow flow in PD patients that were highly associated with dyskinesias and levodopa treatment (Hirano et al., 2008). These changes in cerebral blood flow have been associated with an increased BBB permeability and angiogenesis that are mediated by VEGF (Kortekaas et al., 2005; Ohlin et al., 2011). Similarly, various toxin-induced PD models have shown BBB disruption, including 6-OHDA treated rats and MPTP-treated mice (Carvey et al., 2005; Chen et al., 2008). On the other hand, a growing body of evidence has shown the importance of ABC multidrug transporters such as P-gp in BBB disruption (Kortekaas et al., 2005; Bartels et al., 2008; Bartels, 2011). In this aspect, KO mice for P-glycoprotein have shown an increased accumulation of neurotoxin ivermectin and the carcinostatic drug vinblanstine in the brain, suggesting the importance of P-glycoprotein in the clearance of toxic substances and a possible BBB disruption in PD (Schinkel et al., 1994). Additionally, Kortekaas et al. (2005) has suggested that Parkinson patients have a reduced P-gp (glycoprotein) function in the midbrain, which is associated with a BBB disruption. Interestingly, some PET studies reported a decrease in BBB P-gp function in several brain regions during aging, demonstrating that elder people are more susceptible to the accumulation of toxin compounds in the brain. Taking into account that α-synuclein accumulation is associated with PD pathogenesis, it is possible that a reduction of P-gp could be related with an accumulation of  $\alpha$ -synuclein in the brain (Bartels, 2011). However, further research is needed to assess the importance of P-gp in this process. Finally, the release of proinflammatory cytokines by microglia and astrocytes during PD is associated with both an increased neuronal death and protein rearrangements in tight junctions on EC surface (Figure 1; Desai Bradaric et al., 2012). For example, increased levels of the cytokines IL-6, IL-1B and TNF-A and a decrease in proteins ZO-1 and occludin in tight junctions have been associated with a reduction in the transendothelial electrical resistance, suggesting an alteration in BBB permeability (Wong et al., 2004). Importantly, the loss of signaling interactions between astrocytes and CNS vasculature through changes in protein expression in astrocytic endfeet is associated with morphological changes including hypertrophy, upregulation of GFAP and vimentin and therefore triggering the induction of astrocytes to a more reactive state (Robel et al., 2009; Alvarez et al., 2013). These results highlight the importance of astrocytes in the modulation of BBB properties and the involvement of the reactive astrogliosis during BBB disruption (Figure 1).

# **REACTIVE ASTROGLIOSIS IN PD**

Reactive astrogliosis is the main reaction of astrocytes following brain insults such as infection, inflammatory processes, trauma, α-synuclein accumulation, ischemia and neurodegenerative diseases (Barreto et al., 2007, 2009; Gu et al., 2010; Hamby and Sofroniew, 2010; Xiong et al., 2011; Adelson et al., 2012). This process involves both molecular and morphological changes in astrocytes, which include the hypertrophy of cell bodies and glial processes, increased expression of proteins like GFAP, vimentin, nestin, tenascin-C and chondroitin sulfate proteoglycans (CSPGs; Alvarez et al., 2013). Other characteristics of the process are the increased uptake of glutamate caused by an alteration of vesicular transporters of GABA (vGAT) and glutamate (vGLUT),



**FIGURE 1 | BBB disruption in PD**. During PD development, increased ROS production leads to the accumulation of  $\alpha$ -synuclein in DAneurons, and this is accompanied by mitochondrial dysfunction and increased neuronal death.

Concurrently, astrocyte and microglia became activated, promoting cytokine release, which in turn affects endothelial tight junctions, pericyte phenotype and BBB permeability.

production of cytokines and chemokines that have a modulatory effect on microglia (Croisier and Graeber, 2006), and in some cases the formation of glial scar (Hirsch et al., 2003; Hamby and Sofroniew, 2010; Kang and Hebert, 2011; Colangelo et al., 2014).

Importantly, reactive astrogliosis is a mechanism highly dependent on the cellular and molecular context of the events triggering it, therefore it may have both beneficial and detrimental effects on surrounding neural and non-neural cells (Hamby and Sofroniew, 2010). For example, the glial scar produced after severe astrogliosis may separate necrotic tissue from healthy one, but also has the detrimental effect of impairing axonal regeneration through the expression of molecules like CSPGs, semaphorins and ephrin (Fitch and Silver, 2008; Duffy et al., 2009).

Experimental evidence using cellular and animal models have shown that environmental and biological toxins, like  $\alpha$ -synuclein, LPS (lipopolysaccharides), herbicides and pesticides like rotenone or MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine), can induce both astrogliosis and microgliosis, which is accompanied by altered striatal neuronal morphology, neuronal death, mitochondrial dysfunction and nuclear fragmentation (Langston

et al., 1999; Samantaray et al., 2007; Niranjan et al., 2010). Additionally, injection of LPS in rat brains was followed by an increase in the inducible nitric oxide synthase (iNOS), suggesting that chronic glial activation can cause oxidative stress in the brain, similarly to that in neurodegenerative processes like AD and Parkinson (Sugaya et al., 1998; Hirsch et al., 2003; de Oliveira et al., 2011). Similarly, there is clinical evidence showing that astrogliosis is present in different areas of the brain in PD patients, including the SN, the putamen and the hippocampus (Baxendale et al., 1998; Dickson et al., 2002; Dickson, 2012). Finally, some studies have shown that activated glial cells can participate in the death of dopaminergic neurons, probably via activation of apoptosis by cytokines like TNF-α, IL-1B, IL-6 (Figure 2) and interferon-y and the subsequent production of nitric oxide by the iNOS that may diffuse toward the neurons and induce lipid peroxidation, DNA strands breaks and inhibition of mitochondrial metabolism (Hirsch et al., 2003; Rappold and Tieu, 2010). Released cytokines may bind to TNFR1 and 2, specific receptors in dopaminergic neurons, and activate proapoptotic mechanisms through the activation of caspase 3, caspase 8, and cytochrome

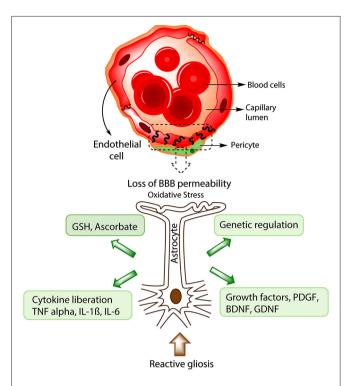


FIGURE 2 | Protective strategies of astrocytes during BBB disruption. In advanced stages of PD, BBB disruption takes place and causes a lost in barrier permeability, entrance of toxic substances and in some instances immune cell infiltration. Processes such as increased ROS production, reactive gliosis and cellular death will inevitably occur. Astrocytic response to BBB disruption includes the production of antioxidative molecules like GSH and ascorbate, generation of growth factors like Brain derived neurotrophic factor (BDNF) and GDNF that could alleviate the cellular death and promote angiogenesis. Furthermore, astrocytes are important in the genetic regulation of endothelial proteins from the tight junction like Occludin and ZO-1. During chronic brain damage, astrocytes also induce the liberation of cytokines like TNF-α, IL-1B, IL-6, important in microglial activation and neuronal death.

C (Hirsch et al., 2003). These results suggest that both the glial reaction and the consequent inflammatory processes could be considered as a promising therapy to reduce neuronal damage during PD (Hirsch et al., 2003).

# PROTECTION STRATEGIES OF ASTROCYTES IN BBB DISRUPTION

Over the last years, much research has focused on specific molecules produced by astrocytes as promising neuroprotective strategies in neuropathologies. These molecules include antioxidant enzymes such as SODs, growth factors, peptide hormones and heat shock proteins (Dringen, 2000; Zheng et al., 2010; Barreto et al., 2011). Many of them have shown protective effects both in dopaminergic neurons and glial cells, and have been used in animal models and clinical trials with remarkable results (Ramaswamy and Kordower, 2009; Yasuda and Mochizuki, 2010). In the last section of our review we discuss the current methods used in neuroprotection based on astrocyte molecules. Additionally, we highlight the future strategies in astrocyte protection

aimed at the development of restorative therapies for the BBB in pathological conditions.

#### **ASTROCYTIC ANTIOXIDANTS AND PD**

Astrocytes secrete beneficial antioxidant molecules, including GSH, (SODs 1, 2 and 3), and ascorbate, which are important for cell survival during neurodegenerative processes (see Figure 2, Anderson and Swanson, 2000; Dringen, 2000; Lindenau et al., 2000; Sims et al., 2004; Mythri et al., 2011). The tripeptide GSH is the main antioxidant in the brain, which is needed for the conversion of methylglyoxal into d-lactate by glyoxalase 1 (Dringen, 2000; Bambrick et al., 2004). Furthermore, GSH is also important in limiting and repairing the deleterious actions of NO and other ROS in the brain such as nitrations and fibril formations of α-synuclein (Chinta and Andersen, 2008). Interestingly, astrocytes possess a greater concentration of GSH (3.8 mmol/L) than neurons (2.5 mmol) probably due to their higher content of γ-glutamylcysteine-synthetase (Rappold and Tieu, 2010). In this aspect, some studies demonstrated that neurons co-cultured with astrocytes exhibit higher levels of GSH compared to neurons cultured alone, suggesting that astrocytes may provide further antioxidant defenses to neurons and BBB cells (Maier and Chan, 2002; Slemmer et al., 2008; Giordano et al., 2009). Additionally, an increase in GSH peroxidase-containing cells showed to be inversely correlated with the severity of dopaminergic cell loss in cell populations from patients with PD, demonstrating that the quantity of GSH peroxidase in cells might be critical for a protective effect against oxidative stress during PD (Damier et al., 1993). Different murine models have shown the importance of GSH in BBB protection, including maintenance of BBB permeability and oxidative protection of mouse pericytes (Shukla et al., 1993; Agarwal and Shukla, 1999; Price et al., 2012). Similarly, ascorbate was shown to protect BBB integrity in a rat ischemic model by preventing changes in BBB permeability and increased ROS production (Lin et al., 2010). One important problem with the use of GSH as a possible therapeutic agent is that its precursor, N-acetycysteine (NAC), does not cross the BBB in significant amounts, therefore various strategies have been used to improve the GSH transport into the brain, such as the use of liposomes, nanoparticles and L-dopa conjugates (Smeyne and Smeyne, 2013). However, further research is needed to address the use of GSH in BBB disruption.

Previous studies showed that SODs exert neuroprotection in PD and other oxidative-related events (Chen and Swanson, 2003). For example, the overexpression of Cu/Zn SOD (SOD1) was able to rescue dopaminergic neurons and diminishes locomotor disabilities in a *Drosophila* mutant model for  $\alpha$ -synuclein overexpression (Botella et al., 2008). Interestingly, a specific increase in SOD levels in the SN, with no changes in activities of GSH peroxidase, catalase and GSH reductase, is observed in PD patients (Chinta and Andersen, 2008). A similar increase was noted in the mitochondrial isoform of SOD (SOD2) in motor cortex from PD patients (Radunovic et al., 1997), suggesting that SODs have a greater importance than other antioxidant enzymes during PD development. Furthermore, the reduction or induced mutation of SOD1 in astrocytes has been shown to induce neuronal degeneration and injury in ischemic and amyotrophic lateral sclerosis

(ALS) murine models (Kondo et al., 1997; Kim et al., 2001; Blackburn et al., 2009; Papadeas et al., 2011). Finally, it was also reported that the overexpression of SOD1 in a transgenic mouse model attenuated BBB disruption by superoxide anion during ischemia (Kim et al., 2001). Altogether, these results emphasize the importance of antioxidant enzymes for the treatment of PD and BBB disruption.

# **GROWTH FACTORS AND BBB PROTECTION IN PD**

Several neurotrophic and growth factors secreted by astrocytes have been extensively used in animal models of neurodegenerative disorders for exerting protection of dopaminergic neurons and glial cells against toxins and ROS during injury through the activation of specific signaling pathways that are responsible for cell survival, induction of antioxidant enzymes, and axonal sprouting (See Figure 2, Ramaswamy and Kordower, 2009; Yasuda and Mochizuki, 2010; Proschel et al., 2014). Some of them like GDNF and neurturin (NRTN) have been tested in clinical trials for PD and other neurodegenerative diseases (Peterson and Nutt, 2008; Ramaswamy and Kordower, 2009).

BDNF, from the neurotrophin family, has been shown to be critical in the survival of cortical, hippocampal and serotonergic neurons. Reduction in BDNF levels is associated with many pathological conditions such as PD, AD, Huntington Disease, ALS, depression and schizophrenia (Allen et al., 2013). Furthermore, BDNF protects neurons against excitotoxicity through activation of the transcription factor NF-kB, which induces expression of antioxidant enzymes such as Mn-SOD and the anti-apoptotic proteins, Bcl-2 and inhibitor of apoptosis proteins IAPs (Mattson, 2008; Lee et al., 2009). Endogenous administration of BDNF was demonstrated to protect neurons in SN following 6-OHDA and MPTP toxicity in rat and primate PD models (Ramaswamy and Kordower, 2009).

The family of GDNF comprises ligands, such as GDNF, NRTN, artemin (ARTN) and persephin. GDNF, secreted by astrocytes and pericytes, is essential for the survival of dopaminergic neurons, peripheral motor neurons and neurons from the locus coeruleus (Yasuda and Mochizuki, 2010; Allen et al., 2013). In this aspect, GDNF administration by catheter increases dopaminergic neuronal resistance against 6-OHDA toxicity, with preservation of motor functions in rat and rhesus monkey models (Safi et al., 2012). More recently, GDNF was shown to increase the expression of claudin-5 and the transendothelial electrical resistances of brain microvascular ECs, suggesting that it may improve the barrier function of the BBB (Sano et al., 2007). However, clinical trials in patients that were administered GDNF in different regions of the brain have shown mixed results, in part due to the mechanism of administration, and the growth factor inability to cross the BBB, therefore further research is needed in order to surpass this obstacle (Gill et al., 2003; Ramaswamy and Kordower, 2009; Allen et al., 2013).

The family of the fibroblast growth factors (FGF) includes 22 structurally related signaling molecules in humans, such as acid FGF, and bFGF, which are important in processes like angiogenesis, wound healing and embryonic development (Itoh and Ornitz, 2011; Huang et al., 2012). Different studies have shown that bFGF protects hippocampal and cortical neurons against glutamate

toxicity by changing the expression of N-methyl-D-aspartic acid (NMDA) receptors and antioxidant enzymes like SODs and GSH reductase (Timmer et al., 2004; Mattson, 2008). Furthermore, a co-culture of transgenic Schwann cells overexpressing FGF-2 with dopaminergic neurons improved neuronal survival and the behavioral outcome in a parkinsonian rat model lesioned with 6-OHDA (Timmer et al., 2004). Additionally, bFGF preserves BBB endothelial adherens junctions in a mouse model of intracerebral hemorrhage through the inhibition of RhoA protein, suggesting that bFGF maintains BBB integrity (Huang et al., 2012). Finally, there are other neurotrophic factors with potential effects on BBB protection including insulin-like growth factors (IGFs), vascular endothelial growth factor (VEGF-B), hepatocyte growth factor (HGF), mesencephalic astrocyte-derived neurotrophic factor and platelet derived growth factor (PDGF; Aberg et al., 2006; Ramaswamy and Kordower, 2009; Pang et al., 2010; Yasuda and Mochizuki, 2010; Sullivan and Toulouse, 2011). For instance, VEGF has shown to improve cerebral blood flow and the pericyte coverage of brain ECs in a murine ischemic model (Zechariah et al., 2013). Also, PDGF-BB impairment in mice has been associated with a reduced number of pericytes, edema formation and murine embryonic lethality, suggesting its importance in BBB development and maintenance (Bergers and Song, 2005; Bonkowski et al., 2011).

The main obstacle with the use of growth factors as therapeutic agents in neurodegenerative diseases seems to be their inability to cross the BBB thoroughly (Peterson and Nutt, 2008). In this regard, different strategies have been used including injections into the lumbar or ventricular CSF, viral vectors with growth factor genes, the temporal disruption of the BBB with hyperosmotic agent like mannitol, the use of linked peptides or peptidomimetic monoclonal antibodies or nanoparticles (Allen et al., 2013). For example, a recent methodology using magnetic nanocarriers for the transport of BDNF was able to cross the BBB without affecting cell viability seems promising (Pilakka-Kanthikeel et al., 2013). A different approach seems to be the transplantation of dopaminergic neurons or glial precursor cells into the injured regions of the brain, which increases the expression of growth factors like BDNF, GDNF, and IGF (Hauser, 2011; Jankovic and Poewe, 2012; Proschel et al., 2014). In this aspect, a recent study by Proschel et al. (2014) has demonstrated that the transplantation of glial precursor cells in 6-OHDA injured rats causes the recovery of DA neurons of the striatum by an increase in the levels of GSH, GDNF, and BDNF. These results suggest that growth factors are essential in the recovery of BBB injuries and related pathologies.

# **CONCLUSIONS AND FUTURE PERSPECTIVES**

Based on the past studies, it seems to be of greater importance to understand the role of BBB in neurodegenerative diseases. It is likely that the maintenance of the BBB and the NVU will decrease the accumulation of Lewy bodies,  $\alpha$ -synuclein fibrils and ROS that worsen the effects of PD. It is important to determine the extent of BBB disruption in PD, and how this disruption may allow the transport of growth factors and antioxidant molecules to the site of injury. The combination of novel drug therapies, such as the use of growth factors, antioxidant molecules or nanoparticles combined with a better understanding

of the astrocytic functions in the BBB, and the use of other therapies that increase astrocyte survival and its antioxidant function may shed light on a prospective cure of PD in the near future.

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# **REFERENCES**

- Abbott, N. J., Rönnbäck, L., and Hansson, E. (2006). Astrocyte-endothelial interactions at the blood-brain barrier. Nat. Rev. Neurosci. 7, 41-53. doi: 10. 1038/nrn1824
- Aberg, N. D., Brywe, K. G., and Isgaard, J. (2006). Aspects of growth hormone and insulin-like growthfactor-I related to neuroprotection, regeneration and functional plasticity in the adult brain. Sci. World J. 6, 53-80. doi: 10.1100/tsw. 2006.22
- Adelson, J. D., Barreto, G. E., Xu, L., Kim, T., Brott, B. K., and Ouyang, Y. B. (2012). Neuroprotection from stroke in the absence of MHCI or PirB. Neuron 73, 1100-1107. doi: 10.1016/j.neuron.2012.01.020
- Agarwal, R., and Shukla, G. S. (1999). Potential role of cerebral glutathione in the maintenance of blood-brain barrier integrity in rat. Neurochem. Res. 24, 1507-1514. doi: 10.1023/A:1021191729865
- Allen, S. J., Watson, J. J., Shoemark, D. K., Barua, N. U., and Patel, N. K. (2013). GDNF, NGF, and BDNF as therapeutic options for neurodegeneration. Pharmacol. Ther. 138, 155-175. doi: 10.1016/j.pharmthera.2013.01.004
- Alvarez, J. I., Katayama, T., and Prat, A. (2013). Glial influence on the blood brain barrier. Glia 61, 1939-1958. doi: 10.1002/glia.22575
- Anderson, C. M., and Swanson, R. A. (2000). Astrocyte glutamate transport: review of properties, regulation and physiological functions. Glia 32, 1-14. doi: 10. 1002/1098-1136(200010)32:1<1::aid-glia10>3.3.co;2-n
- Annese, V., Herrero, M. T., Di Pentima, M., Gomez, A., Lombardi, L., Ros, C. M., et al. (2014). Metalloproteinase 9 contributes toinflammatory glia activation and nigro-striatal pathway degeneration in both mouse and monkey models of 1-methyl-4-phenyl-1,2,3,6 tetrahydropyridine (MPTP)-induced Parkinsonism. Brain Struct. Funct. doi: 10.1007/s00429-014-0718-8. [Epub ahead of print].
- Armulik, A., Genové, G., Mäe, M., Nisancioglu, M. H., Wallgard, E., Niaudet, C., et al. (2010). Pericytes regulate the blood-brain barrier. Nature 468, 557-561. doi: 10.1038/nature09522
- Bambrick, L., Kristian, T., and Fiskum, G. (2004). Astrocyte mitochondrial mechanisms of ischemic brain injury and neuroprotection. Neurochem. Res. 29, 601-608. doi: 10.1023/b:nere.0000014830.06376.e6
- Barreto, G. E., Gonzalez, J., Torres, Y., and Morales, L. (2011). Astrocytic-neuronal crosstalk: implications for neuroprotection from brain injury. Neurosci. Res. 71, 107-113. doi: 10.1016/j.neures.2011.06.004
- Barreto, G., Santos-Galindo, M., Diz-Chaves, Y., Pernía, O., Carrero, P., Azcoitia, I., et al. (2009). Selective estrogen receptor modulators decrease reactive astrogliosis in the injured brain: effects of aging and prolonged depletion of ovarian hormones. Endocrinology 150, 5010-5015. doi: 10.1210/en.2009-0352
- Barreto, G. E., White, R. E., Xu, L., Palm, C. J., and Giffard, R. G. (2012). Effects of heat shock protein 72 (Hsp72) on evolution of astrocyte activation following stroke in the mouse. Exp. Neurol. 238, 284-296. doi: 10.1016/j.expneurol.2012. 08.015
- Barreto, G., Veiga, S., Azcoitia, I., Garcia-Segura, L. M., and Garcia-Ovejero, D. (2007). Testosterone decreases reactive astroglia and reactive microglia after brain injury in male rats: role of its metabolites, oestradiol and dihydrotestosterone. Eur. J. Neurosci. 25, 3039-3046. doi: 10.1111/j.1460-9568.2007. 05563.x
- Bartels, A. L. (2011). Blood-brain barrier P-glycoprotein function in neurodegenerative disease. Curr. Pharm. Des. 17, 2771-2777. doi: 10. 2174/138161211797440122
- Bartels, A. L., Willemsen, A. T., Kortekaas, R., de Jong, B. M., de Vries, R., de Klerk, O., et al. (2008). Decreased blood-brain barrier P-glycoprotein function in the progression of Parkinson's disease, PSP and MSA. J. Neural Transm. 115, 1001-1009. doi: 10.1007/s00702-008-0030-y

- Baxendale, S. A., Van Paesschen, W., Thompson, P. J., Duncan, J. S., Harkness, W. F., and Shorvon, S. D. (1998). Hippocampal cell loss and gliosis: relationship to preoperative and postoperative memory function. Neuropsychiatry Neuropsychol. Behav. Neurol. 11, 12-21.
- Bergers, G., and Song, S. (2005). The role of pericytes in blood-vessel formation and maintenance, Neuro Oncol. 7, 452-464, doi: 10.1215/s1152851705000232
- Betarbet, R., Sherer, T. B., MacKenzie, G., Garcia-Osuna, M., Panov, A. V., and Greenamyre, J. T. (2000). Chronic systemic pesticide exposure reproduces features of Parkinson's disease. Nat. Neurosci. 3, 1301-1306. doi: 10.1038/ 81834
- Blackburn, D., Sargsyan, S., Monk, P. N., and Shaw, P. J. (2009). Astrocyte function and role in motor neuron disease: a future therapeutic target? Glia 57, 1251-1264. doi: 10.1002/glia.20848
- Bonkowski, D., Katyshev, V., Balabanov, R. D., Borisov, A., and Dore-Duffy, P. (2011). The CNS microvascular pericyte: pericyte-astrocyte crosstalk in the regulation of tissue survival. Fluids Barriers CNS 8:8. doi: 10.1186/2045-8118-8-8
- Botella, J. A., Bayersdorfe, F., and Schneuwly, S. (2008). Superoxide dismutase overexpression protects dopaminergic neurons in a Drosophila model of Parkinson's disease. Neurobiol. Dis. 30, 65-73. doi: 10.1016/j.nbd.2007.11.013
- Braak, H., de Vos, R. A., Bohl, J., and Del Tredici, K. (2006). Gastric alphasynuclein immunoreactive inclusions in Meissner's and Auerbach's plexuses in cases staged for Parkinson's disease-related brain pathology. Neurosci. Lett. 396, 67-72. doi: 10.1016/j.neulet.2005.11.012
- Brown, T. P., Rumsby, P. C., Capleton, A. C., Rushton, L., and Levy, L. S. (2006). Pesticides and Parkinson's disease—is there a link? Environ. Health Perspect. 114, 156-164. doi: 10.1289/ehp.8095
- Bushong, E. A., Martone, M. E., Jones, Y. Z., and Ellisman, M. H. (2002). Protoplasmic astrocytes in CA1 stratum radiatum occupy separate anatomical domains. J. Neurosci. 22, 183-192.
- Carvey, P. M. C. H., Zhao, B., Hendey, H., Lum, J., Trachtenberg, B. S., Desai, J., et al. (2005). 6-Hydroxydopamine-induced alterations in bloodbrain barrier permeability. Eur. J. Neurosci. 22, 1158-1168. doi: 10.1111/j.1460-9568.2005. 04281.x
- Castejón, O. J. (2011). Ultrastructural pathology of cortical capillary pericytes in human traumatic brain oedema. Folia Neuropathol. 49, 162-173.
- Céspedes, Á. E., Arango, C. A., and Cardona, G. P. (2013). Análisis comparativo de marcadores de lesión en modelos de isquemia cerebral focal y global en ratas. Biomedica 33, 292-305. doi: 10.7705/biomedica.v33i2.830
- Chen, X., Lan, X., Roche, I., Liu, R., and Geiger, J. D. (2008). Caffeine protects against MPTP induced blood-brain barrier dysfunction in mouse striatum. J. Neurochem. 107, 1147-1157. doi: 10.1111/j.1471-4159.2008.05697.x
- Chen, Y., and Swanson, R. A. (2003). Astrocytes and brain injury. J. Cereb. Blood Flow Metab. 23, 137-149. doi: 10.1097/00004647-200302000-00001
- Chinta, S., and Andersen, I. (2008). Redox imbalance in Parkinson. Biochim. Biophys. Acta 1780, 1362-1367. doi: 10.1016/j.bbagen.2008.02.005
- Colangelo, A. M., Alberghina, L., and Papa, M. (2014). Astrogliosis as a therapeutic target for neurodegenerative diseases. Neurosci. Lett. 565, 59-64. doi: 10.1016/j. neulet.2014.01.014
- Croisier, E., and Graeber, M. B. (2006). Glial degeneration and reactive gliosis in alpha-synucleinopathies: the emerging concept of primary gliodegeneration. Acta Neuropathol. 112, 517-530. doi: 10.1007/s00401-006-0119-z
- Damier, P., Hirsch, E. C., Zhang, P., Agid, Y., and Javoy-Agid, F. (1993). Glutathione peroxidase, glial cells and Parkinson's disease. Neuroscience 52, 1-6. doi: 10. 1016/0306-4522(93)90175-f
- Daneman, R. (2012). The blood-brain barrier in health and disease. Ann. Neurol. 72, 648-672. doi: 10.1002/ana.23648
- de Oliveira, D. M., Barreto, G., Galeano, P., Romero, J. I., Holubiec, M. I., Badorrey, M. S., et al. (2011). Paullinia cupana Mart. var. Sorbilis protects human dopaminergic neuroblastoma SH-SY5Y cell line against rotenone-induced cytotoxicity. Hum. Exp. Toxicol. 30, 1382-1391. doi: 10.1177/0960327110389837
- Dejana, E. (2004). Endothelial cell-cell junctions: happy together. Nat. Rev. Mol. Cell Biol. 5, 261-270. doi: 10.1038/nrm1357
- del Zoppo, G. J., and Milner, R. (2006). Integrin-matrix interactions in the cerebral microvasculature. Arterioscler. Thromb. Vasc. Biol. 26, 1966-1975. doi: 10. 1161/01.atv.0000232525.65682.a2
- Desai Bradaric, B., Patel, A., Schneider, J. A., Carvey, P. M., and Hendey, B. (2012). Evidence for angiogenesis in Parkinson's disease, incidental Lewy body disease,

and progressive supranuclear palsy. *J. Neural Transm.* 119, 59–71. doi: 10. 1007/s00702-011-0684-8

- Dickson, D. W. (2012). Parkinson's disease and parkinsonism: neuropathology. Cold Spring Harb. Perspect. Med. 2:a009258. doi: 10.1101/cshperspect.a0 09258
- Dickson, D. W., Bergeron, C., Chin, S. S., Duyckaerts, C., Horoupian, D., Ikeda, K., et al. (2002). Office of rare diseases neuropathologiccriteria for corticobasal degeneration. J. Neuropathol. Exp. Neurol. 61, 935–946.
- Dore-Duffy, P. (2008). Pericytes: pluripotent cells of the blood brain barrier. Curr. Pharm. Des. 14, 1581–1593. doi: 10.2174/138161208784705469
- Dore-Duffy, P., and Cleary, K. (2011). Morphology and properties of pericytes. *Methods Mol. Biol.* 686, 49–68. doi: 10.1007/978-1-60761-938-3\_2
- Dringen, R. (2000). Metabolism and functions of glutathione in brain. Prog. Neurobiol. 62, 649–671. doi: 10.1016/s0301-0082(99)00060-x
- Duffy, P., Schmandke, A., Schmandke, A. C., Sigworth, J., Shuh, N., Cafferty, W. B., et al. (2009). Rho-associated kinase II (ROCKII) limits axonal growth after trauma within the adult mouse spinal cord. J. Neurosci. 29, 15266–15276. doi: 10.1523/JNEUROSCI.4650-09.2009
- Elali, A., Thériault, P., and Rivest, S. (2014). The role of pericytes in neurovascular unit remodeling in brain disorders. *Int. J. Mol. Sci.* 15, 6453–6474. doi: 10. 3390/ijms15046453
- Enciu, A. M., and Popescu, B. O. (2013). Is there a causal link between inflammation and dementia? Biomed Res. Int. 2013:316495. doi: 10.1155/2013/316495
- Fernandez, H. H. (2012). Updates in the medical management of Parkinson disease. Cleve. Clin. J. Med. 79, 28–35. doi: 10.3949/ccjm.78gr.11005
- Fitch, M. T., and Silver, J. (2008). CNS injury, glial scars and inflammation: inhibitory extracellular matrices and regeneration failure. *Exp. Neurol.* 209, 294–301. doi: 10.1016/j.expneurol.2007.05.014
- Fukui, K., Onodera, K., Shinkai, T., Suzuki, S., and Urano, S. (2001). Impairment of learning and memory in rats caused by oxidative stress and aging and changes in antioxidative defense systems. *Ann. N Y Acad. Sci.* 928, 168–175. doi: 10.1111/j. 1749-6632.2001.tb05646.x
- Gill, S. S., Patel, N. K., Hotton, G. R., O'Sullivan, K., McCarter, R., Bunnage, M., et al. (2003). Direct brain infusion of glial cell line-derived neurotrophic factor in Parkinson disease. *Nat. Med.* 9, 589–595. doi: 10.1038/nm850
- Giordano, G., Kavanagh, T. J., and Costa, L. G. (2009). Mouse cerebellar astrocytes protect cerebellar granule neurons against toxicity of the polybrominateddiphenyl ether (PBDE) mixture DE-71. Neurotoxicology 30, 326–329. doi: 10. 1016/j.neuro.2008.12.009
- Grammas, P., Martinez, J., and Miller, B. (2011). Cerebral microvascular endothelium and the pathogenesis of neurodegenerative diseases. *Expert Rev. Mol. Med.* 13:e19. doi: 10.1017/s1462399411001918
- Gu, X. L., Long, C. X., Sun, L., Xie, C., Lin, X., and Cai, H. (2010). Astrocytic expression of Parkinson's disease-related A53T α-synuclein causes neurodegeneration in mice. *Mol. Brain* 3:12. doi: 10.1186/1756-6606-3-12
- Halliday, G. M., and Stevens, C. H. (2011). Glia: initiators and progressors of pathology in Parkinson's disease. Mov. Disord. 26, 6–17. doi: 10.1002/mds. 23455
- Hamby, M. E., and Sofroniew, M. V. (2010). Reactive astrocytes as therapeutic targets for CNS disorders. *Neurotherapeutics* 7, 494–506. doi: 10.1016/j.nurt. 2010.07.003
- Haseloff, R. F., Blasig, I. E., Bauer, H. C., and Bauer, H. (2005). In search of the astrocytic factor(s) modulating blood-brain barrier functions in brain capillary endothelial cells in vitro. Cell. Mol. Neurobiol. 25, 25–39. doi: 10.1007/s10571-004-1375-x
- Hauser, R. A. (2011). Future treatments for Parkinsons disease: surfing the PD pipeline. *Int. J. Neurosci.* 121(Suppl. 2), 53–62. doi: 10.3109/00207454.2011. 620195
- Haussermann, P., Kuhn, W., Przuntek, H., and Muller, T. (2001). Integrity of the blood–cerebrospinal fluid barrier in early Parkinson's disease. *Neurosci. Lett.* 300, 182–184. doi: 10.1016/s0304-3940(01)01574-9
- Hawkins, B. T., and Davis, T. P. (2005). The blood-brain barrier/neurovascular unit in health and disease. *Pharmacol. Rev.* 57, 173–185. doi: 10.1124/pr. 57.2.4
- Hirano, S., Asanuma, K., Ma, Y., Tang, C., Feigin, A., Dhawan, V., et al. (2008). Dissociation of metabolic and neurovascular responses to levodopa in the treatment of Parkinson's disease. *J. Neurosci.* 28, 4201–4209. doi: 10.1523/JNEUROSCI. 0582-08.2008

- Hirsch, E. C., Breidert, T., Rousselet, E., Hunot, S., Hartmann, A., and Michel, P. P. (2003). The role of glial reaction and inflammation in Parkinson's disease. *Ann. N Y Acad. Sci.* 991, 214–228. doi: 10.1111/j.1749-6632.2003.tb07478.x
- Huang, B., Krafft, P. R., Ma, Q., Rolland, W. B., Caner, B., Lekic, T., et al. (2012).
  Fibroblast growth factors preserve blood-brain barrier integrity through RhoA inhibition after intracerebral hemorrhage in mice. *Neurobiol. Dis.* 46, 204–214. doi: 10.1016/j.nbd.2012.01.008
- Hurtado-Alvarado, G., Cabañas-Morales, A. M., and Gómez-González, B. (2014).
  Pericytes: brain immune interface modulators. Front. Integr. Neurosci. 7:80.
  doi: 10.3389/fnint.2013.00080
- Itoh, N., and Ornitz, D. M. (2011). Fibroblast growth factors: from molecular evolution to roles in development, metabolism and disease. *J. Biochem.* 149, 121–130. doi: 10.1093/jb/mvQ181
- Jangula, A., and Murphy, E. J. (2013). Lipopolysaccharide-induced blood brain barrier permeability is enhanced by alpha-synuclein expression. *Neurosci. Lett.* 551, 23–27. doi: 10.1016/j.neulet.2013.06.058
- Jankovic, J., and Poewe, W. (2012). Therapies in Parkinson's disease. Curr. Opin. Neurol. 25, 433–447. doi: 10.1097/WCO.0b013e3283542fc2
- Jenner, P. (2003). Oxidative stress in Parkinson's disease. Ann. Neurol. 53(Suppl. 3), S26–S36; discussion S36–S38. doi: 10.1002/ana.10483
- Kang, W., and Hebert, J. M. (2011). Signaling pathways in reactive astrocytes, a genetic perspective. Mol. Neurobiol. 43, 147–154. doi: 10.1007/s12035-011-8163-7
- Kim, G. W., Lewén, A., Copin, J., Watson, B. D., and Chan, P. H. (2001). The cytosolic antioxidant, copper/zinc superoxide dismutase, attenuates blood-brain barrier disruption and oxidative cellular injury after photothrombotic cortical ischemia in mice. *Neuroscience* 105, 1007–1018. doi: 10.1016/s0306-4522(01)00237-8
- Kim, G. W., Gasche, Y., Grzeschik, S., Copin, J. C., Maier, C. M., and Chan, P. H. (2003). Neurodegeneration in striatum induced by the mitochondrial toxin 3nitropropionic acid: role of matrix metalloproteinase-9 in early blood-brain barrier disruption? *J. Neurosci.* 23, 8733–8742.
- Kimelberg, H. K., and Nedergaard, M. (2010). Functions of astrocytes and their potential as therapeutic targets. *Neurotherapeutics* 7, 338–353. doi: 10.1016/j. purt 2010 07 006
- Kondo, T., Reaume, A. G., Huang, T. T., Carlson, E., Murakami, K., Chen, S. F., et al. (1997). Reduction of CuZn superoxide dismutase activity exacerbates neuronal cell injury and edema formation aftertransient focal cerebral ischemia. J. Neurosci. 17, 4180–41899.
- Kortekaas, R., Leenders, K. L., van Oostrom, J. C., Vaalburg, W., Bart, J., Willemsen, A. T., et al. (2005). Blood-brain barrier dysfunction in parkinsonian midbrain in vivo. Ann. Neurol. 57, 176–179. doi: 10.1002/ana.20369
- Kurkowska-Jastrzebska, I., Wrónska, A., Kohutnicka, M., Czlonkowski, A., and Czlonkowski, A. (1999). The inflammatory reaction following 1-methyl-4phenyl-1,2,3,6-tetrahydropyridine intoxication in mouse. *Exp. Neurol.* 156, 50– 61. doi: 10.1006/exnr.1998.6993
- Lange, S., Trost, A., Tempfer, H., Bauer, H. C., Bauer, H., Rohde, E., et al. (2013).
  Brain pericyte plasticity as a potential drug target in CNS repair. *Drug Discov. Today* 18, 456–463. doi: 10.1016/j.drudis.2012.12.007
- Langston, J. W., Forno, L. S., Tetrud, J., Reeves, A. G., Kaplan, J. A., and Karluk, D. (1999). Evidence of active nerve cell degeneration in the substantia nigra of humans years after 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine exposure. *Ann. Neurol.* 46, 598–605. doi: 10.1002/1531-8249(199910)46:4<598::aid-ana7>3.0.co;2-f
- Lau, L. W., Cua, R., Keough, M. B., Haylock-Jacobs, S., and Yong, V. W. (2013).
  Pathophysiology of the brain extracellular matrix: a new target for remyelination. *Nat. Rev. Neurosci.* 14, 722–729. doi: 10.1038/nrn3550
- Lee, H., and Pienaar, I. S. (2014). Disruption of the blood-brain barrier in Parkinson's disease: curse or route to a cure? Front. Biosci. (Landmark Ed.) 19, 272–280. doi: 10.2741/4206
- Lee, S. J., Kim, J. S., Lee, K. S., An, J. Y., Kim, W., Kim, Y. I., et al. (2009). The severity of leukoaraiosis correlates with the clinical phenotype of Parkinson's disease. *Arch. Gerontol. Geriatr.* 49, 255–259. doi: 10.1016/j.archger.2008. 09.005
- Lin, J. L., Huang, Y. H., Shen, Y. C., Huang, H. C., and Liu, P. H. (2010). Ascorbic acidprevents bloodbrain barrier disruption and sernsory deficit caused by sustained compressio ofprimary somatosensory cortex. *J. Cereb. Blood Flow Metab.* 30, 1121–1136. doi: 10.1038/jcbfm.2009.277

Lindenau, J., Noack, H., Possel, H., Asayama, K., and Wolf, G. (2000). Cellular distribution of superoxide dismutases in the rat CNS. Glia 29, 25-34. doi: 10. 1002/(sici)1098-1136(20000101)29:1<25::aid-glia3>3.0.co;2-g

- Luissint, A. C., Artus, C., Glacial, F., Ganeshamoorthy, K., and Couraud, P. O. (2012). Tight junctions at the blood brain barrier: physiological architecture and disease-associated dysregulation. Fluids Barriers CNS 9:23. doi: 10.1186/2045-
- Maier, C. M., and Chan, P. H. (2002). Role of superoxide dismutases in oxidative damage and neurodegenerative disorders. Neuroscientist 8, 323-334. doi: 10. 1177/107385840200800408
- Maragakis, N. J., and Rothstein, J. D. (2006). Mechanisms of disease: astrocytes in neurode generative disease. Nat. Clin. Pract. Neurol. 2, 679-689. doi: 10. 1038/ncpneuro0355
- Mattson, M. P. (2008). Glutamate and neurotrophic factors in neuronal plasticity and disease. Ann. NY Acad. Sci. 1144, 97-112. doi: 10.1196/annals.1418.005
- Minagar, A., and Alexander, J. S. (2003). Blood brain barrier disruption in multiple sclerosis, Mult. Scler. 9, 540-549, doi: 10.1191/1352458503ms965oa
- Mokgokong, R., Wang, S., Taylor, C. J., Barrand, M. A., and Hladky, S. B. (2014). Ion transporters in brain endothelial cells that contribute to formation of brain interstitial fluid. Pflugers Arch. 466, 887-901. doi: 10.1007/s00424-013-1342-9
- Mythri, R. B., Venkateshappa, C., Harish, G., Mahadevan, A., Muthane, U. B., Yasha, T. C., et al. (2011). Evaluation of markers of oxidative stress, antioxidant function and astrocytic proliferation in the striatum and frontal cortex of Parkinson's disease brains. Neurochem. Res. 36, 1452-1463. doi: 10.1007/s11064-011-0471-9
- Nag, S. (2011). Morphology and properties of brain endothelial cells. Methods Mol. Biol. 686, 3-47. doi: 10.1007/978-1-60761-938-3\_1
- Najjar, S., Pearlman, D. M., Devinsky, O., Najjar, A., and Zagzag, D. (2013). Neurovascular unit dysfunction with blood brain barrier hyperpermeability contributes to major depressive disorder: a review of clinical and experimental evidence, I. Neuroinflammation 10:142, doi: 10.1186/1742-2094-10-142
- Niranjan, R., Nath, C., and Shukla, R. (2010). The mechanism of action of MPTP-induced neuroinflammation and its modulation by melatonin in rat astrocytoma cells, C6. Free Radic. Res. 44, 1304-1316. doi: 10.3109/10715762.
- Nutt, J. G., and Wooten, G. F. (2005). Clinical practice. Diagnosis and initial management of Parkinson's disease. N. Engl. J. Med. 353, 1021-1027. doi: 10. 1056/NEJMcp043908
- Oberheim, N. A., Goldman, S. A., and Nedergaard, M. (2012). Heterogeneity of astrocytic form and function. Methods Mol. Biol. 814, 23-45. doi: 10.1007/978-1-61779-452-0\_3
- Ogawa, N., Asanuma, M., Miyazaki, I., Diaz-Corrales, F. J., and Miyoshi, K. (2005). L-DOPA treatment from the viewpoint of neuroprotection. Possible mechanism of specific and progressive dopaminergic neuronal death in Parkinson's disease. J. Neurol. 252(Suppl. 4), IV23-IV31. doi: 10.1007/s00415-005-4006-7
- Ohlin, K. E., Francardo, V., Lindgren, H. S., Sillivan, S. E., O'Sullivan, S. S., Luksik, A. S., et al. (2011). Vascular endothelial growth factor is upregulated by L-dopa in the parkinsonian brain: implications for the development of dyskinesia. Brain 134, 2339-2357. doi: 10.1093/brain/awr165
- Ossig, C., and Reichmann, H. (2013). Treatment of Parkinson's disease in the advanced stage. J. Neural Transm. 120, 523-529. doi: 10.1007/s00702-013-
- Pang, Y., Zheng, B., Campbell, L. R., Fan, L. W., Cai, Z., and Rhodes, P. G. (2010). IGF-1 can either protect against or increase LPS-induced damage in the developing rat brain. Pediatr. Res. 67, 579-584. doi: 10.1203/PDR.0b013e3181dc240f
- Pan-Montojo, F., Anichtchik, O., Dening, Y., Knels, L., Pursche, S., Jung, R., et al. (2010). Progression of Parkinson's disease pathology is reproduced by intragastric administration of rotenone in mice. PLoS One 5:e8762. doi: 10. 1371/journal.pone.0008762
- Papadeas, S. T., Kraig, S. E., O'Banion, C., Lepore, A. C., and Maragakis, N. J. (2011). Astrocytes carrying the superoxide dismutase 1 (SOD1G93A) mutation induce wild-type motor neurodegeneration in vivo. Proc. Natl. Acad. Sci. U S A 108, 17803-17808. doi: 10.1073/pnas.1103141108
- Parpura, V., Grubisic, V., and Verkhratsky, A. (2011). Ca(2+) sources for the exocytotic release of glutamate from astrocytes. Biochim. Biophys. Acta 1813, 984-991. doi: 10.1016/j.bbamcr.2010.11.006
- Peterson, A. L., and Nutt, J. G. (2008). Treatment of Parkinson's disease with trophic factors. Neurotherapeutics 5, 270-280. doi: 10.1016/j.nurt.2008.02.003

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- Pilakka-Kanthikeel, S., Atluri, V. S., Sagar, V., Saxena, S. K., and Nair, M. (2013). Targeted brain derived neurotropic factors (BDNF) delivery across the bloodbrain barrier for neuro-protection using magnetic nano carriers: an in-vitro study. PLoS One 8:e62241. doi: 10.1371/journal.pone.0062241
- Popescu, B. O., Toescu, E. C., Popescu, L. M., Bajenaru, O., Muresanu, D. F., Schultzberg, M., et al. (2009). Blood-brain barrier alterations in ageing and dementia. J. Neurol. Sci. 283, 99-106. doi: 10.1016/j.jns.2009. 02.321
- Price, T. O., Eranki, V., Banks, W. A., Ercal, N., and Shah, G. N. (2012). Topiramate treatment protects blood-brain barrier pericytes from hyperglycemia induced oxidative damage in diabetic mice. Endocrinology 153, 362–372. doi: 10.1210/en. 2011-1638
- Proschel, C., Stripay, J. L., Shih, C. H., Munger, J. C., and Noble, M. D. (2014). Delayed transplantation of precursor cell-derived astrocytes provides multiple benefits in a rat model of Parkinsons. EMBO Mol. Med. 6, 504-518. doi: 10. 1002/emmm.201302878
- Radunovic, A., Porto, W. G., Zeman, S., and Leigh, P. N. (1997). Increased mitochondrial superoxide dismutase activity in Parkinson's disease but not amyotrophic lateral sclerosis motor cortex. Neurosci. Lett. 239, 105-108. doi: 10. 1016/s0304-3940(97)00905-1
- Ramaswamy, S., and Kordower, J. H. (2009). Are growth factors the answer? Parkinsonism Relat. Disord. 15(Suppl. 3), S176-S180. doi: 10.1016/S1353-8020(09)70809-0
- Rappold, P. M., and Tieu, K. (2010). Astrocytes and therapeutics for Parkinson's disease. Neurotherapeutics 7, 413-423. doi: 10.1016/j.nurt.2010.
- Ribatti, D., Nico, B., and Crivellato, E. (2011). The role of pericytes in angiogenesis. Int. I. Dev. Biol. 55, 261-268, doi: 10.1387/jidb.103167dr
- Robel, S., Mori, T., Zoubaa, S., Schlegel, J., Sirko, S., Faissner, A., et al. (2009). Conditional deletion of beta1-integrin in astroglia causes partial reactive gliosis. Glia 57, 1630-1647. doi: 10.1002/glia.20876
- Safi, R., Gardaneh, M., Panahi, Y., Maghsoudi, N., Zaefizadeh, M., and Gharib, E. (2012). Optimized quantities of GDNF overexpressed by engineered astrocytes are critical for protection of neuroblastoma cells against 6-OHDA toxicity. J. Mol. Neurosci. 46, 654-665. doi: 10.1007/s12031-011-9654-8
- Samantaray, S., Knaryan, V. H., Guyton, M. K., Matzelle, D. D., Ray, S. K., and Banik, N. L. (2007). The parkinsonian neurotoxin rotenone activates calpain and caspase-3 leading to motoneuron degeneration in spinal cord of Lewis rats. Neuroscience 146, 741-755. doi: 10.1016/j.neuroscience.2007.
- Sano, Y., Shimizu, F., Nakayama, H., Abe, M., Maeda, T., Ohtsuki, S., et al. (2007). Endothelial cells constituting blood nerve barrier have highly specialized characteristics as barrier-forming cells. Cell Struct. Funct. 32, 139-147. doi: 10. 1247/csf.07015
- Sá-Pereira, I., Brites, D., and Brito, M. A. (2012). Neurovascular unit: a focus on pericytes. Mol. Neurobiol. 45, 327-347. doi: 10.1007/s12035-012-8244-2
- Schinkel, A. H., Smit, J. J., van Tellingen, O., Beijnen, J. H., Wagenaar, E., van Deemter, L., et al. (1994). Disruption of the mouse mdr1a P-glycoprotein gene leads to a deficiency in the blood-brain barrier and to increased sensitivity to drugs. Cell 77, 491-502. doi: 10.1016/0092-8674(94)90212-7
- Schwartz, M. D., and Sabetay, M. D. (2012). An approach to the continuous dopaminergic stimulation in Parkinson's disease. Isr. Med. Assoc. J. 14, 175–179.
- Shukla, A., Shukla, R., Dikshit, M., and Srimal, R. C. (1993). Alterations in free radical scavenging mechanisms following blood-brain barrier disruption. Free Radic. Biol. Med. 15, 97-100. doi: 10.1016/0891-5849(93)90128-h
- Simpson, I. A., Carruthers, A., and Vannucci, S. J. (2007). Supply and demand in cerebral energy metabolism: the role of nutrient transporters. J. Cereb. Blood Flow Metab. 27, 1766-1791. doi: 10.1038/sj.jcbfm.9600521
- Sims, N. R., Nilsson, M., and Muyderman, H. (2004). Mitochondrial glutathione: a modulator of brain cell death. J. Bioenerg. Biomembr. 36, 329-333. doi: 10. 1023/b:jobb.0000041763.63958.e7
- Singer, C. (2012). Managing the patient with newly diagnosed Parkinson disease. Cleve. Clin. J. Med. 79(Suppl. 2), S3-S7. doi: 10.3949/ccjm.79.s2a.01
- Slemmer, J. E., Shacka, J. J., Sweeney, M. I., and Weber, J. T. (2008). Antioxidants and free radical scavengers for the treatment of stroke, traumatic brain injury and aging. Curr. Med. Chem. 15, 404-414. doi: 10.2174/092986708783 497337

Smeyne, M., and Smeyne, R. J. (2013). Glutathione metabolism and Parkinson's disease. Free Radic. Biol. Med. 62, 13–25. doi: 10.1016/j.freeradbiomed.2013. 05 001

- Stamatovic, S. M., Keep, R. F., and Andjelkovic, A. V. (2008). Brain endothelial cell-cell junctions: how to "open" the blood brain barrier. *Curr. Neuropharmacol.* 6, 179–192. doi: 10.2174/157015908785777210
- Stanimirovic, D. B., and Friedman, A. (2012). Pathophysiology of the neurovascular unit disease cause or consequence? J. Cereb. Blood Flow Metab. 32, 1207–1221. doi: 10.1038/jcbfm.2012.25
- Sugaya, K., Chou, S., Xu, S. J., and McKinney, M. (1998). Indicators of glial activation and brain oxidative stress after intraventricular infusion of endotoxin. *Brain Res. Mol. Brain Res.* 58, 1–9. doi: 10.1016/s0169-328x(97) 00365-3
- Sullivan, A. M., and Toulouse, A. (2011). Neurotrophic factors for the treatment of Parkinson's disease. Cytokine Growth Factor Rev. 22, 157–165. doi: 10.1016/j. cytogfr.2011.05.001
- Tanner, C. M., Kamel, F., Ross, G. W., Hoppin, J. A., Goldman, S. M., Korell, M., et al. (2011). Rotenone, paraquat and Parkinson's disease. *Environ. Health Perspect.* 119, 866–872. doi: 10.1289/ehp.1002839
- Theis, M., Söhl, G., Eiberger, J., and Willecke, K. (2005). Emerging complexities in identity and function of glial connexins. *Trends Neurosci.* 28, 188–195. doi: 10. 1016/i.tins.2005.02.006
- Timmer, M., Muller-Ostermeyer, F., Kloth, V., Winkler, C., Grothe, C., and Nikkhah, G. (2004). Enhanced survival, reinnervation, and functional recovery of intrastriatal dopamine grafts co-transplanted with Schwann cells overexpressing high molecular weight FGF-2 isoforms. *Exp. Neurol.* 187, 118–136. doi: 10. 1016/j.expneurol.2004.01.013
- van Sorge, N. M., and Doran, K. S. (2012). Defense at the border: the bloodbrain barrier versus bacterial foreigners. *Future Microbiol.* 7, 383–394. doi: 10. 2217/fmb.12.1
- Vives-Bauza, C., and Przedborski, S. (2011). Mitophagy: the latest problem for Parkinson's disease. Trends Mol. Med. 17, 158–165. doi: 10.1016/j.molmed.2010. 11.002
- Volterra, A., and Meldolesi, J. (2005). Astrocytes, from brain glue to communication elements: the revolution continues. *Nat. Rev. Neurosci.* 6, 626–640. doi: 10.1038/nrn1722
- Wada, K., Arai, H., Takanashi, M., Fukae, J., Oizumi, H., Yasuda, T., et al. (2006). Expression levels of vascular endothelial growth factor and its receptors in Parkinson's disease. *Neuroreport* 7, 705–709. doi: 10.1097/01.wnr.0000215769. 71657.65
- Wang, H. L., Chou, A. H., Wu, A. S., Chen, S. Y., Weng, Y. H., Kao, Y. C., et al. (2011). PARK6 PINK1 mutants are defective in maintaining mitochondrial membrane potential and inhibiting ROS formation of substantia nigra dopaminergic neurons. *Biochim. Biophys. Acta* 1812, 674–684. doi: 10.1016/j. bbadis.2011.03.007

- Wolburg, H., Noell, S., Mack, A., Wolburg-Buchholz, K., and Fallier-Becker, P. (2009). Brain endothelial cells and the glio-vascular complex. Cell Tissue Res. 335, 75–96. doi: 10.1007/s00441-008-0658-9
- Wong, A. D., Ye, M., Levy, A. F., Rothstein, J. D., Bergles, D. E., and Searson, P. C. (2013). The blood-brain barrier: an engineering perspective. *Front. Neuroeng*. 6:7. doi: 10.3389/fneng.2013.00007
- Wong, D., Dorovini-Zis, K., and Vincent, S. R. (2004). Cytokines, nitric oxide and cGMP modulate the permeability of an in vitro model of the human bloodbrain barrier. Exp. Neurol. 190, 446–455. doi: 10.1016/j.expneurol.2004.08.008
- Xiong, X., Barreto, G. E., Xu, L., Ouyang, Y. B., Xie, X., and Giffard, R. G. (2011). Increased brain injury and worsened neurological outcome in interleukin-4 knockout mice after transient focal cerebral ischemia. Stroke 42, 2026–2032. doi: 10.1161/STROKEAHA.110.593772
- Yasuda, T., and Mochizuki, H. (2010). Use of growth factors for the treatment of Parkinson's disease. Expert Rev. Neurother. 10, 915–924. doi: 10.1586/ern.10.55
- Zechariah, A., Elali, A., Doeppner, T. R., Jin, F., Hasan, M. R., Helfrich, I., et al. (2013). Vascular endothelial growth factor promotes pericyte coverage of brain capillaries, improves cerebral blood flow during subsequent focal cerebral ischemia and preserves the metabolic penumbra. Stroke 44, 1690–1697. doi: 10. 1161/STROKEAHA.111.000240
- Zheng, L., Ishii, Y., Tokunaga, A., Hamashima, T., Shen, J., Zhao, Q. L., et al. (2010). Neuroprotective effects of PDGF against oxidative stress and the signaling pathway involved. J. Neurosci. Res. 88, 1273–1284. doi: 10.1002/jnr.22302
- Zonta, M., Angulo, M. C., Gobbo, S., Rosengarten, B., Hossmann, K. A., Pozzan, T., et al. (2003). Neuron-to-astrocyte signaling is central to the dynamic control of brain microcirculation. *Nat. Neurosci.* 6, 43–50. doi: 10.1038/nn980

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