

Post-ischemic inflammation regulates neural damage and protection

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Akihiko Yoshimura, Department of Microbiology and Immunology, School of Medicine, Keio University, 35 Shinanomachi, Shinjuku-ku, Tokyo 160-8582, Japan e-mail: yoshimura@a6.keio.jp Post-ischemic inflammation is important in ischemic stroke pathology. However, details of the inflammation process, its resolution after stroke and its effect on pathology and neural damage have not been clarified. Brain swelling, which is often fatal in ischemic stroke patients, occurs at an early stage of stroke due to endothelial cell injury and severe inflammation by infiltrated mononuclear cells including macrophages, neutrophils, and lymphocytes. At early stage of inflammation, macrophages are activated by molecules released from necrotic cells [danger-associated molecular patterns (DAMPs)], and inflammatory cytokines and mediators that increase ischemic brain damage by disruption of the blood–brain barrier are released. After post-ischemic inflammation, macrophages are also involved in tissue repair and neural cell regeneration by producing tropic factors. The mechanisms of inflammation resolution and conversion of inflammation to neuroprotection are largely unknown. In this review, we summarize information accumulated recently about DAMP-induced inflammation and the neuroprotective effects of inflammatory cells, and discuss next generation strategies to treat ischemic stroke.

Keywords: damage-associated molecular patterns (DAMPs), inflammation, cytokines, inflammasome, resolution of inflammation

INTRODUCTION

Inflammation is implicated in almost all of central nervous system (CNS) diseases (Lo, 2010; Moskowitz et al., 2010; Iadecola and Anrather, 2011). Neurodegeneration, infection, trauma, and ischemia stimulate immune responses in the brain, although to varying degrees. The process in neuronal injury involves various intracellular mechanisms (abnormal metabolism and degeneration of protein, dysfunction of organelles, etc.), which cause the activation of microglia and the infiltration of circulating immune cells (Lo, 2010). Inflammation may not be always main process in the pathology of CNS diseases; nonetheless, the distinct characteristics of ischemic stroke are large amount of necrotic neuronal death and extreme infiltration of immune cells (Moskowitz et al., 2010; Iadecola and Anrather, 2011).

Severe inflammation causes cerebral swelling, which is often fatal in ischemic stroke patients. Broad necrotic lesion generates abundant inflammatory mediators and damage-associated molecular patterns (DAMPs), which enhance the chemotaxis of circulating immune cells and make them more efficient participants to promote inflammation (Moskowitz et al., 2010; Iadecola and Anrather, 2011). Cerebral inflammation exaggerates vascular dysfunction and induces further neuronal cell death (Dirnagl et al., 1999). Thus, post-ischemic inflammation is an essential process in the pathophysiology of ischemic stroke and is closely related to the prognosis after stroke (Dirnagl et al., 1999; Lo, 2010; Moskowitz et al., 2010; Iadecola and Anrather, 2011). In addition, inflammation is generally considered useful for the clearance of the large amount of debris caused by brain cell necrotic death (Moskowitz et al., 2010; Iadecola and Anrather, 2011). Inflammation, resolution of inflammation, and repair of neural damage are sequential pivotal events after stroke. To clarify the detailed mechanisms of each step of cerebral inflammation is indispensable to develop next generation therapies for ischemic stroke. The molecular basis of these steps is now being clarified by the recent accumulating evidences. We summarize these findings and discuss the principles of post-ischemic inflammation from beginning to end.

INFLAMMATORY DAMPs

Brain ischemia induces various large metabolic changes in brain cells. Hypoxic stress, nutrients stress, and ER stress will cause cell death and trigger post-ischemic inflammation. Although receptors for pathogens such as Toll-like receptors (TLRs) are thought to be involved in early step of inflammation, brain is a sterile organ. Thus, endogenous molecules, i.e., DAMPs derived from injured brain cells, must trigger the inflammatory response in immune cells (**Table 1**). These DAMPs induce the activation of TLRs and other pattern recognition receptors [receptor for advanced glycation end products (RAGE) and c-type lectin receptors], which promote inflammatory mediator expression and tissue injury (**Figure 1**; Tang et al., 2007; Yanai et al., 2009; Suzuki et al., 2013). Recent scientific advances have suggested the existence of various types of DAMPs in ischemic brain.

NUCLEIC ACIDS AND NUCLEOTIDES

Various intracellular components are released into the extracellular space by necrotic brain cell death. Among these,

Table 1 | List of inflammatory DAMPs.

		DAMPs	Receptor	Reference
Signal 1	Nucleic acid	Mitochondrial DNA	TLR9	Zhang etal. (2010), Sun etal. (2013), Maeda and Fadeel (2014), Walko etal. (2014), Wenceslau etal. (2014)
		Self RNA, DNA	TLR7,9	Hyakkoku etal. (2010), Kawai and Akira (2010), Brea etal. (2011), Stevens etal. (2011), Leung etal. (2012)
	Lipid	Carboxyalkylpyrroles	TLR2	West et al. (2010)
		Oxidized phospholipids	CD36	Cho etal. (2005), Gao etal. (2006), Abe etal. (2010), Haider etal. (2011), Miller etal. (2011), Ho etal. (2012), Matt etal. (2013)
	Protein	HMGB1	TLR2,4, RAGE	Qiu et al. (2008), Zhang et al. (2011)
		Peroxiredoxin	TLR2,4	Shichita et al. (2012), Kuang et al. (2014)
		S100A8, A9	TLR4	Tsai et al. (2014)
		Mrp8, 14	TLR4	Loser et al. (2010)
		CIRP	TLR2,4	Qiang et al. (2013)
Signal 2	Nucleotide	ATP	P2X, P2Y	Martinon et al. (2002), Abulafia et al. (2009), Ceruti et al. (2009), Denes et al. (2013), Fann et al. (2013), Yang et al. (2014),
	Lipid	Phospholipids	?	Clemens et al. (1996), Bonventre et al. (1997), Muralikrishna Adibhatla and Hatcher (2006), Shanta et al. (2012), Iyer et al. (2013), Zhong et al. (2013)
	Protein	ASC specks	?	Baroja-Mazo et al. (2014), Franklin et al. (2014)



FIGURE 1 | Mechanisms of post-ischemic inflammation. DAMPs are released into extracellular compartment and activate infiltrating immune cells by two ways: Signal 1 (via the activation of pattern recognition receptor) and Signal 2 (via the activation of inflammasome). Various inflammatory cytokines promote neuronal injury, and induce further inflammation mediated by T cells in subacute phase. After days and week after stroke onset, the resolution of post-ischemic inflammatory mediators, and the production of anti-inflammatory molecules or neurotrophic factors. In this recover phase, inflammatory immune cells turn into neuroprotective cells.

nucleic acids and nucleotides are major DAMPs that have recently received much attention. Mitochondrial DNA released by cellular injury can be detected as DAMPs by immune cells, because mitochondria are considered to have a symbiotic origin that carries numerous characteristics resembling bacteria. Mitochondrial DNA is a sensor molecule of innate immunity by activating TLR9 and can be detected in cerebrospinal fluid after traumatic brain injury (Zhang et al., 2010; Walko et al., 2014). Vascular permeability is also increased by circulating mitochondrial DNA after injury (Sun et al., 2013; Wenceslau et al., 2014). Recently accumulated data indicates that mitochondrial DAMPs could be an important candidate for the trigger of post-ischemic inflammation, even if there is not yet any direct evidence (Maeda and Fadeel, 2014).

Self RNA and DNA (in complex with LL37 peptide) activate immune cells via TLR7 or TLR9 (Kawai and Akira, 2010). TLR7 is associated with the deterioration in ischemic stroke patients; in contrast, ischemic brain damage was not reduced in TLR9-deficient mice (Hyakkoku et al., 2010; Brea et al., 2011). Several reports demonstrate the implications of TLR7 and TLR9 in ischemic preconditioning. In these articles, the pretreatment using a TLR7 or TLR9 agonist reveals significant neuroprotection after cerebral ischemia by activating interferon regulatory factor 3/7- (IRF3/7)-induced type I interferon (IFN) signaling pathway (Stevens et al., 2011; Leung et al., 2012). Although the interaction between self nucleic acids and TLRs in the ischemic brain remains controversial, ischemic preconditioning via the TLR7 or TLR9 signaling pathway may represent a therapeutic strategy.

Purines (ATP and UTP) released from injured brain cells and their receptors, P2X and P2Y, function as alerting signals in CNS (Ceruti et al., 2009). Importantly, ATP also activates inflamma-somes, which are large multimolecular complexes that control the activity of the proteolytic enzyme caspase-1 that cleaves pro-IL-1 β to an active 17 kDa form (Martinon et al., 2002). The activation of

the NLRP1 or NALP3 inflammasome has been recently reported to promote post-ischemic inflammation and neuronal death (Abulafia et al., 2009; Fann et al., 2013; Yang et al., 2014). Because IL-1 β produced from both infiltrating immune cells and brain cells is important (Denes et al., 2013), it should be clarified how the inflammasome is activated in ischemic brain or hematopoietic cells. Inhibition of the inflammasome activation pathway may be a possible therapeutic strategy for ischemic stroke.

LIPIDS

Various types of lipids are also important regulators of innate immunity. For example, oxidized low density lipoprotein (oxLDL) is a popular inflammatory mediator, which activates TLRs through binding with its receptor, CD36 (Stewart et al., 2010). Although the function of oxLDL in ischemic brain remains unclear, recent research has indicated that end products of lipid oxidation may be implicated in cerebral post-ischemic inflammation (Uchida, 2013). Carboxyalkylpyrroles, which are generated in inflammatory tissue, activate TLR2 and promote angiogenesis in ischemic organs (West et al., 2010). Oxidized phospholipids are also generated during cerebral inflammation and are considered to be DAMPs (Gao et al., 2006; Haider et al., 2011; Ho et al., 2012). Oxidized phospholipids are CD36 ligands that promote inflammation via TLR2 activation in ischemic brain (Cho et al., 2005; Abe et al., 2010). The recognition and endocytosis of oxidized lipids by pattern recognition receptors could regulate post-ischemic inflammation (Miller et al., 2011; Matt et al., 2013).

Phospholipids could also be inflammasome activators. Phospholipid metabolism is drastically altered by cerebral ischemia (Shanta et al., 2012). There are several reports showing the activation of phospholipase A2 (PLA2) in ischemic brain, which results in hydrolysis of membrane phospholipids (Clemens et al., 1996; Bonventre et al., 1997; Muralikrishna Adibhatla and Hatcher, 2006). Phospholipid hydrolysis and mitochondrial dysfunction induced by cerebral ischemia generate reactive oxygen species (ROS). Two recent studies have identified both ROS-dependent and ROS-independent pathways for inflammasome activation. The former is demonstrated by a charged phospholipid liposome that consecutively induces ROSdependent calcium influx and NLRP3 inflammasome activation (Zhong et al., 2013). In the latter case, mitochondrial cardiolipin has been reported to directly bind to and activate the NLRP3 inflammasome (Iver et al., 2013). Thus, the metabolism and modification of lipids during cerebral ischemia may be closely associated with the post-ischemic inflammation start signal.

PROTEINS

High mobility group box 1 (HMGB1) and peroxiredoxin (Prx) family proteins are two major DAMPs in ischemic brain. There is a difference in the functional phase of these two proteins (Shichita et al., 2012). HMGB1, which is included in the nucleus of brain cells, is released extracellularly at the hyperacute phase (several hours after the stroke onset; Qiu et al., 2008). On the other hand, Prx family proteins function at the acute and subacute phases (12–72 h after the onset), especially in the penumbral area (Shichita et al., 2012). This is because Prx family protein expression is induced by an intracellular increase in ROS, which results from ischemic change. HMGB1 directly breaks down the blood–brain barrier and increases vascular permeability (Zhang et al., 2011). However, Prx directly induces the activation of infiltrating immune cells via TLR signaling. Ligustilide has been reported as a therapeutic candidate that suppresses cerebral post-ischemic inflammation by inhibiting the Prx/TLR4 signaling pathway (Kuang et al., 2014).

S100A8, S100A9, Mrp8, Mrp14, and cold-inducible RNA binding protein (CIRP) have also been reported to be protein DAMPs, although their relevance in post-ischemic inflammation has not yet been clarified (Loser et al., 2010; Qiang et al., 2013; Tsai et al., 2014). Inflammatory responses by these protein DAMPs occur through the activation of TLR2, TLR4, and RAGE. TLR2 and TLR4 signaling pathways are essential for sterile inflammation, including ischemic stroke (Chen et al., 2007). TLR2-blocking antibody is neuroprotective against ischemic brain injury (Ziegler et al., 2011). Similarly, resatorvid, which inhibits the TLR4 signaling pathway, attenuates ischemic brain injury and also suppresses Nox4-induced oxidative stress and neuronal apoptosis (Suzuki et al., 2012). It is also possible that DAMP-mediated TLR activation requires other adaptor molecules (Chun and Seong, 2010). CD14, a TLR4 co-receptor, may be implicated in post-ischemic inflammation (Reed-Geaghan et al., 2009). Heat shock protein gp96 is another candidate molecule that functions as an adaptor for both TLR2 and TLR4 (Yang et al., 2007).

It is not known whether protein DAMPs can activate inflammasomes. Recently, aggregated ASC (apoptosis-associated speck-like protein containing a caspase recruitment domain) has been reported to be released into the extracellular space after cell death and it activates inflammasomes in the surrounding immune cells (Baroja-Mazo et al., 2014; Franklin et al., 2014). Inflammasome activation, which occurs through ASC polymerization, results in caspase-1 activation and pyroptotic cell death. Extracellularly released ASC is internalized by surrounding macrophages and induces lysosomal damage and inflammasome activation. These mechanisms of inflammasome activation remain to be elucidated in ischemic brain injury.

OTHER INFLAMMATORY DAMPs

Basic research may neglect the influence of aging and life habits by using healthy young rodents. These are important factors for the generation of DAMPs. Aging and continuous high serum glucose levels increase lipid peroxidation and AGEs in body systems (Basta et al., 2004; Cai et al., 2014). AGEs are proteins that are modified by sugar, through the Amadori and Maillard reaction. AGEs are found in chronic lesions; for example, the amyloid deposits that are surrounded by macrophages in patients with dialysis-related amyloidosis (Miyata et al., 1993). Thus, AGEs usually take a long time (more than a month) to generate; however, AGEs can be generated in a short period of time during inflammation (Weil, 2012). Glyoxal and glyceraldehyde induce AGE formation within 1 week (Takeuchi et al., 2001). In addition, the pivotal role of the RAGE in post-ischemic inflammation has been demonstrated (Muhammad et al., 2008). AGEs can be a potential DAMP, especially in aged human ischemic stroke patients.

INFLAMMATION SUPPRESSION AND RESOLUTION

Activated immune cells and brain cells are the major players after various DAMPs trigger post-ischemic inflammation. These cells produce inflammatory cytokines, chemokines, and other cytotoxic mediators, and this leads to prolonged inflammation and progressive brain edema during several days after the stroke onset (Figure 1). However, post-ischemic inflammation rarely lasts for a long period of time, and the most intense inflammatory phase takes place within 7 days after stroke onset (Dirnagl et al., 1999; Iadecola and Anrather, 2011). In this phase, the number of infiltrating immune cells decreases remarkably, and remaining immune cells in ischemic brain produce anti-inflammatory or neurotrophic factors (Shichita et al., 2009; Smirkin et al., 2010). For example, the detailed mechanisms about the infiltration and the change to antiinflammatory phenotype of neutrophils have been recently clarified (Cuartero et al., 2013; Gorina et al., 2014). The period of cerebral post-ischemic inflammation always ends, and thus, the mechanisms of its resolution must exist in ischemic brain.

Three major points on the resolution of inflammation have been discussed in a recent publication (Buckley et al., 2012). These points are the production of anti-inflammatory mediators, the depletion of inflammatory mediators, and the induction of anti-inflammatory immune cells. After post-ischemic inflammation, infiltrating macrophages turn into anti-inflammatory macrophages, which produce neurotropic factors and clear necrotic debris. Inflammatory DAMPs will also be implicated in the induction of anti-inflammatory macrophages, although its mechanism still remains to be clarified. We introduce recent advantages of the relationship between post-ischemic inflammation and its resolution.

ANTI-INFLAMMATORY MEDIATOR PRODUCTION

Many molecules have been reported to be neuroprotective factors. However, most of these molecules failed to improve neurological deficits in ischemic stroke patients, even if they are effective in animal stroke models. It has been suggested that neuroprotection alone is not sufficient to improve the prognosis of human ischemic stroke. Anti-inflammatory mechanisms in the entire brain and how these mechanisms are triggered needs to be determined. Because most brain cells are dead in the ischemic region several days after the stroke onset, infiltrating immune cells and reactive glial cells could be major players in the tissue repair. In practice, accelerating their effect is a potential next generation therapeutic strategy, and direct *in vivo* reprogramming of reactive glial cells into functional neurons after cerebral injury by retroviral transduction of the NeuroD1 gene was recently reported (Guo et al., 2014).

IL-10 and TGF- β are major anti-inflammatory molecules in various organ injuries. Both are produced by infiltrating immune cells and reactive glial cells after ischemic brain injury. Viral overexpression of IL-10 in ischemic brain is neuroprotective (Ooboshi et al., 2006). One recent report demonstrated the anti-inflammatory effect of TGF- β by inhibiting excessive neuroinflammation during the subacute phase of brain ischemia (Cekanaviciute et al., 2014). Although the anti-inflammatory effects of IL-10 and TGF- β have been pivotal, it remains to be clarified whether these effects last up to 1 week after the stroke onset (Pál et al., 2012). If the mechanisms for stimulating TGF- β and IL-10 expression can be controlled, this may become a strong therapeutic method.

DEPLETION OF INFLAMMATORY MEDIATORS AND CELLS

Infiltrating immune cells and reactive glial cells produce various inflammatory mediators. TNF-a and IL-1B directly induce neuronal cell death. IL-23 and IL-1ß activate T cell-mediated innate immunity and promote secondary ischemic damage during the subacute phase of ischemic brain injury (Shichita et al., 2009; Konoeda et al., 2010). The existence of these inflammatory mediators, including DAMPs, prolongs post-ischemic inflammation and will be a threat to neuronal survival and repair. However, inflammatory mediator degradation mechanisms remain mostly unknown. Inflammatory molecules may be degraded by some enzymes or consumed by receptor-mediated endocytosis. It is expected that nucleotides and lipids are rapidly metabolized in the ischemic brain, and transfer by the blood stream or cerebrospinal fluid (CSF) will help to scavenge inflammatory mediator. Further research should clarify the detailed mechanisms to scavenge inflammatory molecules produced in the ischemic brain, and antibody therapy will be a pivotal therapeutic method targeting this potential mechanism. TNF- α and IL-23 neutralizing antibody have been used clinically for rheumatoid arthritis and psoriasis patients, respectively. Natalizumab is the neutralizing antibody for integrin- $\alpha 4$, which is necessary for T cell infiltration into the inflammatory tissue, and has already been used for multiple sclerosis (Yednock et al., 1992). T cell depletion from ischemic brain has received attention as a potential next generation therapy for ischemic stroke (Meisel and Meisel, 2011; Wei et al., 2011). Thus, antibody therapies may be used to help treat ischemic stroke patients in the near future.

Activation of inflammasomes in immune cells induces the production of IL-1 β in its mature form, and finally results in the rapid cell death of the same cells, which is called pyroptosis. Pyroptosis may be a possible mechanism for the clearance of inflammatory immune cells. This is supported by the fact that dying cells detected using the TdT-mediated dUTP nick end labeling (TUNEL) staining method include macrophages and glial cells in the ischemic brain (Mabuchi et al., 2000).

INDUCTION OF IMMUNE CELL REPAIR

The repair process for damaged brain tissues and regeneration of neural cells takes place during resolution of inflammation. It is difficult to separate this process from the anti-inflammatory mechanism, because they may overlap each other. We will further discuss neuroprotective factors and repairing the damage to immune cells.

NEUROPROTECTIVE FACTORS

Various growth factors are also produced by immune cells and glial cells (Gudi et al., 2011). Among these, IGF-1 and FGF-2 are produced by infiltrating macrophages and microglia during the recovery phase of ischemic brain injury (which occurs

1 week after the stroke onset). IGF-1 and FGF-2 improve the neurological outcome by saving neuron and glial cells from cell death (Ness et al., 2004; Ikeda et al., 2005; Zhu et al., 2008; Hill et al., 2012; Lalancette-Hébert et al., 2012). IGF-1 also enhances repair after ischemic stroke by promoting neural regeneration, remyelination, and synaptogenesis (Lecker et al., 2007; Zhu et al., 2008, 2009; Kooijman et al., 2009). Further investigation should clarify the mechanisms of IGF-1 and FGF-2 induction in the ischemic brain. Recently, transfer of mesenchymal stem cells (MSCs) has been explored as a next generation therapy for ischemic stroke (Kalladka and Muir, 2014). MSCs produce various growth factors and promote neuronal survival and neurogenesis (Calió et al., 2014). By improving the transfer method, cell therapy may become a pivotal therapeutic strategy (Guo et al., 2013).

The neuroprotective effect of prostaglandin E2 (PGE2) and its receptor signaling pathway has received recent attention. PGE2 has an effect via four distinct G protein-coupled EP receptors (E-prostanoid: EP1, EP2, EP3, and EP4). The activation of EP2 signaling has a neuroprotective effect in ischemic brain injury, which was shown in the significant increase in infarct volume in mice lacking the EP2 receptor (McCullough et al., 2004). Similarly, signaling via the EP4 receptor, which is expressed in both neurons and ischemic endothelial cells, has neuroprotective effects against ischemic brain injury (Liang et al., 2011). The administration of an EP4 agonist reduces infarct volume and neurological deficits. In a neonatal hypoxic-ischemic encephalopathy model, the inhibition of EP1 receptor signaling or the activation of EP2, EP3, and EP4 receptor signaling reveals attenuation of the ischemic injury (Taniguchi et al., 2011). PGE2 and EP receptor signaling pathways have various functions, which are dependent on distinct pathology of cerebral diseases (Furuyashiki and Narumiya, 2011). Targeting specific EP receptors in ischemic brain may become a novel therapeutic method.

Similar to PGE2, some lipids have been reported to have antiinflammatory effects and promote neuroregeneration (Serhan, 2014). Cerebral ischemia increases PLA2 activity, which results in the hydrolysis of phospholipids in the cellular membrane (Shanta et al., 2012). Although the PLA2 effect itself is cytotoxic because it disrupts the cellular membrane, PLA2 also generates docosahexaenoic acid (DHA) derivatives and lysophospholipids through phospholipid hydrolysis (Bonventre et al., 1997). Resolvin and Neuroprotectin have been investigated for their anti-inflammatory function in ischemic stroke (Marcheselli et al., 2003; Bazan, 2009). Lysophospholipids also increase in ischemic brain and promote neurite outgrowth (Ikeno et al., 2005; Spohr et al., 2011; Shanta et al., 2012). Regulating the effect of these lipids is expected for the resolution of post-ischemic inflammation.

NEUROPROTECTIVE CELLS

Inflammatory DAMPs activate glial cells and infiltrating immune cells to promote post-ischemic inflammation. Paradoxically, this mechanism results in the infiltrating macrophage cell death and also induces anti-inflammatory and tissue-repairing immune cells.

Immune cell activation also induces anti-inflammatory cells. These cells have been called M2 macrophages, in contrast to the inflammatory M1 macrophages. Many researches have described the M2 macrophage markers; these markers include: arginase-1 (Arg1), chitinase3l3 (Ym), and Relma (Fizz1). These markers are intracellular enzymes that are implicated in collagen synthesis and cell division; therefore, M2 enzymes are considered to promote tissue repair. Arg1 is the only marker that was reported to function as a neuroprotective enzyme (Estévez et al., 2006). However, these M2 markers may not be a good indicator for recovery after ischemic stroke. M2 markers are rapidly expressed in macrophages by TLR activation or other pattern recognition receptors, which also induce inflammatory cytokine expression (Hu et al., 2012). M2 markers appear in ischemic brain mostly during the same phase as the inflammatory mediators, the M1 markers, are expressed. In addition, the transfer of M2 marker positive-macrophages has not been reported to be sufficiently neuroprotective (Desestret et al., 2013).

During post-ischemic inflammation, some populations of macrophages and microglia become neuroprotective (Lalancette-Hébert et al., 2007). Galectin-1 has been suggested to be an inducer of anti-inflammatory macrophage/microglial cells (Starossom et al., 2012; Quintá et al., 2014). Galectin-1 is produced by astrocytes and has a neuroprotective effect against ischemic brain damage (Qu et al., 2011). Thus, the resolution of post-ischemic inflammation can be enhanced by the induction of a specific macrophage/microglial cell population. However, it is not clear whether the M2 markers truly reflect the neuroprotective function of macrophages and microglial cells. Suppressing inflammation alone is not enough to protect the brain from ischemic injury. IGF-1 and FGF-2 production seems to be a good index of repairing function (Lalancette-Hébert et al., 2012).

Further study is required to clarify whether sufficient clearance of inflammatory mediators (including DAMPs) begins neuronal regeneration after ischemic stroke. A recent study has suggested that there is a relationship between TLR activation and neuronal repair (Bohacek et al., 2012). It is possible that DAMPs triggers the secondary signals, which lead to resolution of post-ischemic inflammation, even if the primary signals via pattern recognition receptors promote ischemic damage. What is this mechanism? The role of immune cells, other than macrophages and microglia in part of the repair process, is not fully understood. This understanding may be critical for the establishment of next generation therapies for ischemic stroke.

CONCLUSION

Immunity and various physiological mechanisms are implicated in the triggering, persistence, and resolution of post-ischemic inflammation. Recent accumulating evidences clarify the complexity of these mechanisms to understand the entire mechanisms. They will show promising potential targets to develop therapies for ischemic stroke.

REFERENCES

Abe, T., Shimamura, M., Jackman, K., Kurinami, H., Anrather, J., Zhou, P., et al. (2010). Key role of CD36 in Toll-like receptor 2 signaling in cerebral ischemia. *Stroke* 41, 898–904. doi: 10.1161/STROKEAHA.109.572552

- Abulafia, D. P., de Rivero Vaccari, J. P., Lozano, J. D., Lotocki, G., Keane, R. W., and Dietrich, W. D. (2009). Inhibition of the inflammasome complex reduces the inflammatory response after thromboembolic stroke in mice. *J. Cereb. Blood Flow Metab.* 29, 534–544. doi: 10.1038/jcbfm.2008.143
- Baroja-Mazo, A., Martín-Sánchez, F., Gomez, A., Martínez, C. M., Amores-Iniesta, J., Compan, V., et al. (2014). The NLRP3 inflammasome is released as a particulate danger signal that amplifies the inflammatory response. *Nat. Immunol.* 15, 738– 748. doi: 10.1038/ni.2919
- Basta, G., Schmidt, A. M., and De Caterina, R. (2004). Advanced glycation end products and vascular inflammation: implications for accelerated atherosclerosis in diabetes. *Cardiovasc. Res.* 63, 582–592. doi: 10.1016/j.cardiores.2004.05.001
- Bazan, N. G. (2009). Neuroprotectin D1-mediated anti-inflammatory and survival signaling in stroke, retinal degenerations, and Alzheimer's disease. J. Lipid Res. 50, S400–S405. doi: 10.1194/jlr.R800068-JLR200
- Bohacek, I., Cordeau, P., Lalancette-Hébert, M., Gorup, D., Weng, Y. C., Gajovic, S., et al. (2012). Toll-like receptor 2 deficiency leads to delayed exacerbation of ischemic injury. *J. Neuroinflammation* 9, 191. doi: 10.1186/1742-2094-9-191
- Bonventre, J. V., Huang, Z., Taheri, M. R., O'Leary, E., Li, E., Moskowitz, M. A., et al. (1997). Reduced fertility and postischaemic brain injury in mice deficient in cytosolic phospholipase A2. *Nature* 390, 622–625. doi: 10.1038/37635
- Brea, D., Sobrino, T., Rodríguez-Yáñez, M., Ramos-Cabrer, P., Agulla, J., Rodríguez-González, R., et al. (2011). Toll-like receptors 7 and 8 expression is associated with poor outcome and greater inflammatory response in acute ischemic stroke. *Clin. Imunol.* 139, 193–198. doi: 10.1016/j.clim.2011.02.001
- Buckley, C. D., Gilroy, D. W., Serhan, C. N., Stockinger, B., and Tak, P. P. (2012). The resolution of inflammation. Nat. Rev. Immunol. 13, 59–66. doi: 10.1038/nri3362
- Cai, W., Uribarri, J., Zhu, L., Chen, X., Swamy, S., Zhao, Z., et al. (2014). Oral glycotoxins are a modifiable cause of dementia and the metabolic syndrome in mice and humans. *Proc. Natl. Acad. Sci. U.S.A.* 111, 4940–4945. doi: 10.1073/pnas.1316013111
- Calió, M. L., Marinho, D. S., Ko, G. M., Ribeiro, R. R., Carbonel, A. F., Oyama, L. M., et al. (2014). Transplantation of bone marrow mesenchymal stem cells decreases oxidative stress, apoptosis, and hippocampal damage in brain of a spontaneous stroke model. *Free Radic. Biol. Med.* 70, 141–154. doi: 10.1016/j.freeradbiomed.2014.01.024
- Cekanaviciute, E., Fathali, N., Doyle, K. P., Williams, A. M., Han, J., and Buckwalter, M. S. (2014). Astrocytic transforming growth factor-beta signaling reduces subacute neuroinflammation after stroke in mice. *Glia* 62, 1227–1240. doi: 10.1002/glia.22675
- Ceruti, S., Villa, G., Genovese, T., Mazzon, E., Longhi, R., Rosa, P., et al. (2009). The P2Y-like receptor GPR17 as a sensor of damage and a new potential target in spinal cord injury. *Brain* 132(Pt 8), 2206–2218. doi: 10.1093/brain/ awp147
- Chen, C. J., Kono, H., Golenbock, D., Reed, G., Akira, S., and Rock, K. L. (2007). Identification of a key pathway required for the sterile inflammatory response triggered by dying cells. *Nat. Med.* 13, 851–856. doi: 10.1038/nm1603
- Cho, S., Park, E. M., Febbraio, M., Anrather, J., Park, L., Racchumi, G., et al. (2005). The class B scavenger receptor CD36 mediates free radical production and tissue injury in cerebral ischemia. *J. Neurosci.* 25, 2504–2512. doi: 10.1523/JNEUROSCI.0035-05.2005
- Chun, K. H., and Seong, S. Y. (2010). CD14 but not MD2 transmit signals from DAMP. *Int. Immunopharmaol.* 10, 98–106. doi: 10.1016/j.intimp.2009.10.002
- Clemens, J. A., Stephenson, D. T., Smalstig, E. B., Roberts, E. F., Johnstone, E. M., Sharp, J. D., et al. (1996). Reactive glia express cytosolic phospholipase A2 after transient global forebrain ischemia in the rat. *Stroke* 27, 527–535. doi: 10.1161/01.STR.27.3.527
- Cuartero, M. I., Ballesteros, I., Moraga, A., Nombela, F., Vivancos, J., Hamilton, J. A., et al. (2013). N2 neutrophils, novel players in brain inflammation after stroke: modulation by the PPARg agonist rosiglitazone. *Stroke* 44, 3498–3508. doi: 10.1161/STROKEAHA.113.002470
- Denes, A., Wilkinson, F., Bigger, B., Chu, M., Rothwell, N. J., and Allan, S. M. (2013). Central and haematopoietic interleukin-1 both contribute to ischaemic brain injury. *Dis. Model. Mech.* 6, 1043–1048. doi: 10.1242/dmm.011601
- Desestret, V., Riou, A., Chauveau, F., Cho, T. H., Devillard, E., Marinescu, M., et al. (2013). *In vitro* and *in vivo* models of cerebral ischemia show discrepancy in therapeutics effects of M2 macrophages. *PLoS ONE.* 8:e67063. doi: 10.1371/journal.pone.0067063

- Dirnagl, U., Iadecola, C., and Moskowitz, M. A. (1999). Pathobiology of ischaemic stroke: an integrated view. *Trends Neurosci.* 22, 391–397. doi: 10.1016/S0166-2236(99)01401-0
- Estévez, A. G., Sahawneh, M. A., Lange, P. S., Bae, N., Egea, M., and Ratan, R. R. (2006). Arginase 1 regulation of nitric oxide production is key to survival of trophic factor-deprived motor neurons. *J. Neurosci.* 26, 8512–8516. doi: 10.1523/JNEUROSCI.0728-06.2006
- Fann, D. Y., Lee, S. Y., Manzanero, S., Tang, S. C., Gelderblom, M., Chunduri, P., et al. (2013). Intravenous immunoglobulin suppresses NLRP1 and NLRP3 inflammasome-mediated neuronal death in ischemic stroke. *Cell Death Dis.* 4, e790. doi: 10.1038/cddis.2013.326
- Franklin, B. S., Bossaller, L., De Nardo, D., Ratter, J. M., Stutz, A., Engels, G., et al. (2014). The adaptor ASC has extracellular and 'prionoid' activities that propagate inflammation. *Nat. Immunol.* 15, 727–737. doi: 10.1038/ ni.2913
- Furuyashiki, T., and Narumiya, S. (2011). Stress responses: the contribution of prostaglandin E₂ and its receptors. *Nat. Rev. Endocrinol.* 7, 163–175. doi: 10.1038/nrendo.2010.194
- Gao, S., Zhang, R., Greenberg, M. E., Sun, M., Chen, X., Levison, B. S., et al. (2006). Phospholipid hydroxyalkenals, a subset of recently discovered endogenous CD36 ligands, spontaneously generate novel furan-containing phospholipids lacking CD36 binding activity *in vivo. J. Biol. Chem.* 281, 31298–31308. doi: 10.1074/jbc.M604039200
- Gorina, R., Lyck, R., Vestweber, D., and Engelhardt, B. (2014). β2 integrin-mediated crawling on endothelial ICAM-1 and ICAM-2 is a prerequisite for transcellular neutrophil diapedesis across the inflamed blood–brain barrier. *J. Immunol.* 192, 324–337. doi: 10.4049/jimmunol.1300858
- Gudi, V., Škuljec, J., Yildiz, Ö., Frichert, K., Skripuletz, T., Moharregh-Khiabani, D., et al. (2011). Spatial and temporal profiles of growth factor expression during CNS demyelination reveal the dynamics of repair priming. *PLoS ONE* 6:e22623. doi: 10.1371/journal.pone.0022623
- Guo, L., Ge, J., Wang, S., Zhou, Y., Wang, X., and Wu, Y. (2013). A novel method for efficient delivery of stem cells to the ischemic brain. *Stem Cell Res. Ther.* 4, 116. doi: 10.1186/scrt327
- Guo, Z., Zhang, L., Wu, Z., Chen, Y., Wang, F., and Chen, G. (2014). In Vivo direct reprogramming of reactive glial cells into functional neurons after brain injury and in an Alzheimer's disease model. *Cell Stem Cell* 14, 188–202. doi: 10.1016/j.stem.2013.12.001
- Haider, L., Fischer, M. T., Frishcer, J. M., Bauer, J., Höftberger, R., Botond, G., et al. (2011). Oxidative damage in multiple sclerosis lesions. *Brain* 134(Pt 7), 1914–1924. doi: 10.1093/brain/awr128
- Hill, J. J., Jin, K., Mao, X. O., Xie, L., and Greenberg, D. A. (2012). Intracerebral chondroitinase ABC and heparan sulfate proteoglycan glypican improve outcome from chronic stroke in rats. *Proc. Natl. Acad. Sci. U.S.A.* 109, 9155–9160. doi: 10.1073/pnas.1205697109
- Ho, P. P., Kanter, J. L., Johnson, A. M., Srinagesh, H. K., Chang, E. J., Purdy, T. M., et al. (2012). Identification of naturally occurring fatty acids of the myelin sheath that resolve neuroinflammation. *Sci. Transl. Med.* 4, 137ra73. doi: 10.1126/scitranslmed.3003831
- Hu, X., Li, P., Guo, Y., Wang, H., Leak, R. K., Chen, S., et al. (2012). Microglia/macrophage polarization dynamics reveal novel mechanism of injury expansion after focal cerebral ischemia. *Stroke* 43, 3063–3070. doi: 10.1161/STROKEAHA.112.659656
- Hyakkoku, K., Hamanaka, J., Tsuruma, K., Shimazawa, M., Tanaka, H., Uematsu, S., et al. (2010). Toll-like receptor 4 (TLR4), but not TLR3 or TLR9, knock-out mice have neuroprotective effects against focal cerebral ischemia. *Neuroscience* 171, 258–267. doi: 10.1016/j.neuroscience.2010.08.054
- Iadecola, C., and Anrather, J. (2011). The immunology of stroke: from mechanisms to translation. *Nat. Med.* 17, 796–808. doi: 10.1038/nm.2399
- Ikeda, N., Nonoguchi, N., Zhao, M. Z., Watanabe, T., Kajimoto, Y., Furutama, D., et al. (2005). Bone marrow stromal cells that enhanced fibroblast growth factor-2 secretion by herpes simplex virus vector improve neurological outcome after transient focal cerebral ischemia in rats. *Stroke* 36, 2725–2730. doi: 10.1161/01.STR.0000190006.88896.d3
- Ikeno, Y., Konno, N., Cheon, S. H., Bolchi, A., Ottonello, S., Kitamoto, K., et al. (2005). Secretory phospholipases A2 induce neurite outgrowth in PC12 cells through lysophosphatidylcholine generation and activation of G2A receptor. J. Biol. Chem. 280, 28044–28052. doi: 10.1074/jbc.M503343200

- Iyer, S. S., He, Q., Janczy, J. R., Elliott, E. I., Zhong, Z., Olivier, A. K., et al. (2013). Mitochondrial cardiolipin is required for Nlrp3 inflammasome activation. *Immunity* 39, 311–323. doi: 10.1016/j.immuni.2013.08.001
- Kalladka, D., and Muir, K. W. (2014). Brain repair: cell therapy in stroke. Stem Cells Cloning 7, 31–44.
- Kawai, T., and Akira, S. (2010). The role of pattern-recognition receptors in innate immunity: update on Toll-like receptors. *Nat. Immunol.* 11, 373–384. doi: 10.1038/ni.1863
- Konoeda, F., Shichita, T., Yoshida, H., Sugiyama, Y., Muto, G., Hasegawa, E., et al. (2010). Therapeutic effect of IL-12/23 and their signaling pathway blockade on brain ischemia model. *Biochem. Biophys. Res. Commun.* 402, 500–506. doi: 10.1016/j.bbrc.2010.10.058
- Kooijman, R., Sarre, S., Michotte, Y., and De Keyser, J. (2009). Insulin-like growth factor I: a potential neuroprotective compound for the treatment of acute ischemic stroke? *Stroke* 40, e83–e88. doi: 10.1161/STROKEAHA.108.528356
- Kuang, X., Wang, L. F., Yu, L., Li, Y. J., Wang, Y. N., He, Q., et al. (2014). Ligustilide ameliorates neuroinflammation and brain injury in focal cerebral ischemia/reperfusion rats: involvement of inhibition of TLR4/peroxiredoxin 6 signaling. *Free Radic. Biol. Med.* 71, 165–175. doi: 10.1016/j.freeradbiomed.2014.03.028
- Lalancette-Hébert, M., Gowing, G., Simard, A., Weng, Y. C., and Kriz, J. (2007). Selective ablation of proliferating microglial cells exacerbates ischemic injury in the brain. *J. Neurosci.* 27, 2596–2605. doi: 10.1523/JNEUROSCI.5360 -06.2007
- Lalancette-Hébert, M., Swarup, V., Beaulieu, J. M., Bohacek, I., Abdelhamid, E., Weng, Y. C., et al. (2012). Galectin-3 is required for resident microglia activation and proliferation in response to ischemic injury. *J. Neurosci.* 32, 10383–10395. doi: 10.1523/JNEUROSCI.1498-12.2012
- Lecker, R. R., Soldner, F., Velasco, I., Gavin, D. K., Androutsellis-Theotokis, A., and McKay, R. D. (2007). Long-lasting regeneration after ischemia in the cerebral cortex. *Stroke* 38, 153–161. doi: 10.1161/01.STR.0000252156.65953.a9
- Leung, P. Y., Stevens, S. L., Packard, A. E., Lessov, N. S., Yang, T., Conrad, V. K., et al. (2012). Toll-like receptor 7 preconditioning induces robust neuroprotection against stroke by a novel type I interferon-mediated mechanism. *Stroke* 43, 1383– 1389. doi: 10.1161/STROKEAHA.111.641522
- Liang, X., Lin, L., Woodling, N. S., Wang, Q., Anacker, C., Pan, T., et al. (2011). Signaling via the prostaglandin E2 receptor EP4 exerts neuronal and vascular protection in a mouse model of cerebral ischemia. *J. Clin. Invest.* 121, 4362–4371. doi: 10.1172/JCI46279
- Lo, E. H. (2010). Degeneration and repair in central nervous system disease. Nat. Med. 16, 1205–1209. doi: 10.1038/nm.2226
- Loser, K., Vogl, T., Voskort, M., Leuken, A., Kupas, V., Nacken, W., et al. (2010). The Toll-like receptor 4 ligands Mrp8 and Mrp14 are crucial in the development of autoreactive CD8⁺ T cells. *Nat. Med.* 16, 713–717. doi: 10.1038/nm. 2150
- Mabuchi, T., Kitagawa, K., Ohtsuki, T., Kuwabara, K., Yagita, Y., Yanagihara, T., et al. (2000). Contribution of microglia/macrophages to expansion of infarction and response of oligodendrocytes after focal cerebral ischemia in rats. *Stroke* 31, 1735–1743. doi: 10.1161/01.STR.31.7.1735
- Maeda, A., and Fadeel, B. (2014). Mitochondria released by cells undergoing TNF-α-induced necroptosis act as danger signals. *Cell Death Dis.* 5, e1312. doi: 10.1038/cddis.2014.277
- Marcheselli, V. L., Hong, S., Lukiw, W. J., Tian, X. H., Gronert, K., Musto, A., et al. (2003). Novel docosanoids inhibit brain ischemia-reperfusion-mediated leukocyte infiltration and pro-inflammatory gene expression. *J. Biol. Chem.* 278, 43807–43817. doi: 10.1074/jbc.M305841200
- Martinon, F., Burns, K., and Tschopp, J. (2002). The inflammasome: a molecular platform triggering activation of inflammatory caspases and processing of proILβ. *Mol. Cell* 10, 417–426. doi: 10.1016/S1097-2765(02)00599-3
- Matt, U., Sharif, O., Martins, R., Furtner, T., Langeberg, L., Gawish, R., et al. (2013). WAVE1 mediates suppression of phagocytosis by phospholipid-derived DAMPs. *J. Clin. Invest.* 123, 3014–3024. doi: 10.1172/JCI60681
- McCullough, L., Wu, L., Haughey, N., Liang, X., Hand, T., Wang, Q., et al. (2004). Neuroprotective function of the PGE₂ EP2 receptor in cerebral ischemia. J. Neurosci. 24, 257–268. doi: 10.1523/JNEUROSCI.4485-03.2004
- Meisel, C., and Meisel, A. (2011). Suppressing immunosuppression after stroke. N. Engl. J. Med. 365, 2134–2136. doi: 10.1056/NEJMcibr11 12454

- Miller, Y. I., Choi, S. H., Wiesner, P., Fang, L., Harkewicz, R., Hartvigsen, K., et al. (2011). Oxidation-specific epitopes are danger-associated molecular patterns recognized by pattern recognition receptors of innate immunity. *Circ. Res.* 108, 235–248. doi: 10.1161/CIRCRESAHA.110.223875
- Miyata, T., Oda, O., Inagi, R., Iida, Y., Araki, N., Yamada, N., et al. (1993). beta 2-Microglobulin modified with advanced glycation end products is a major component of hemodialysis-associated amyloidosis. *J. Clin. Invest.* 92, 1243–1252. doi: 10.1172/JCI116696
- Moskowitz, M. A., Lo, E. H., and Iadecola, C. (2010). The science of stroke: mechanisms in search of treatments. *Neuron* 67, 181–198. doi: 10.1016/j.neuron.2010.07.002
- Muhammad, S., Barakat, W., Stoyanov, S., Murikinati, S., Yang, H., Tracey, K. J., et al. (2008). The HMGB1 receptor RAGE mediates ischemic brain damage. J. Neurosci. 28, 12023–12031. doi: 10.1523/JNEUROSCI.2435-08.2008
- Muralikrishna Adibhatla, R., and Hatcher, J. F. (2006). Phospholipase A2, reactive oxygen species, and lipid peroxidation in cerebral ischemia. *Free Radic. Biol. Med.* 40, 376–387. doi: 10.1016/j.freeradbiomed.2005.08.044
- Ness, J. K., Scaduto, R. C. Jr., and Wood, T. L. (2004). IGF-I prevents glutamatemediated bax translocation and cytochrome C release in O4⁺ oligodendrocyte progenitors. *Glia* 46, 183–194. doi: 10.1002/glia.10360
- Ooboshi, H., Ibayashi, S., Shichita, T., Kumai, Y., Takada, J., Ago, T., et al. (2006). Postischemic gene transfer of interleukin-10 protects against both focal and global brain ischemia. *Circulation* 111, 913–919. doi: 10.1161/01.CIR.0000155622.68580.DC
- Pál, G., Vincze, C., Renner, É., Wappler, E. A., Nagy, Z., Lovas, G., et al. (2012). Time course, distribution and cell types of induction of transforming growth factor betas following middle cerebral artery occlusion in the rat brain. *PLoS ONE* 7:e46731. doi: 10.1371/journal.pone.0046731
- Qiang, X., Yang, W. L., Wu, R., Zhou, M., Jacob, A., Dong, W., et al. (2013). Cold-inducible RNA-binding protein (CIRP) triggers inflammatory responses in hemorrhagic shock and sepsis. *Nat. Med.* 19, 1489–1495. doi: 10.1038/nm.3368
- Qiu, J., Nishimura, M., Wang, Y., Sims, J. R., Qiu, S., Savitz, S. I., et al. (2008). Early release of HMGB-1 from neurons after the onset of brain ischemia. *J. Cereb. Blood Flow Metab.* 28, 927–938. doi: 10.1038/sj.jcbfm.9600582
- Qu, W. S., Wang, Y. H., Ma, J. F., Tian, D. S., Zhang, Q., Pan, D. J., et al. (2011). Galectin-1 attenuates astrogliosis-associated injuries and improves recovery of rats following focal cerebral ischemia. *J. Neurochem.* 116, 217–226. doi: 10.1111/j.1471-4159.2010.07095.x
- Quintá, H. R., Pasquini, J. M., Rabinovich, G. A., and Pasquini, L. A. (2014). Glycandependent binding of galectin-1 to neuropilin-1 promotes axonal regeneration after spinal cord injury. *Cell Death. Differ.* 21, 941–955. doi: 10.1038/cdd.2014.14
- Reed-Geaghan, E. G., Savage, J. C., Hise, A. G., and Landreth, G. E. (2009). CD14 and toll-like receptors 2 and 4 are required for fibrillar Aβ-stimulated microglial activation. J. Neurosci. 29, 11982–11992. doi: 10.1523/JNEUROSCI.3158-09.2009
- Serhan, C. N. (2014). Pro-resolving lipid mediators are leads for resolution physiology. *Nature* 510, 92–101. doi: 10.1038/nature13479
- Shanta, S. R., Choi, C. S., Lee, J. H., Shin, C. Y., Kim, Y. J., Kim, K. H., et al. (2012). Global changes in phospholipids identified by MALDI MS in rats with focal cerebral ischemia. *J. Lipid Res.* 53, 1823–1831. doi: 10.1194/jlr. M022558
- Shichita, T., Hasegawa, E., Kimura, A., Morita, R., Sakaguchi, R., Takada, I., et al. (2012). Peroxiredoxin family proteins are key initiators of post-ischemic inflammation in the brain. *Nat. Med.* 18, 911–917. doi: 10.1038/nm.2749
- Shichita, T., Sugiyama, Y., Ooboshi, H., Sugimori, H., Nakagawa, R., Takada, I., et al. (2009). Pivotal role of cerebral interleukin-17-producing γδT cells in the delayed phase of ischemic brain injury. *Nat. Med.* 215, 946–950. doi: 10.1038/nm.1999
- Smirkin, A., Matsumoto, H., Takahashi, H., Inoue, A., Tagawa, M., Ohue, S., et al. (2010). Iba1⁺/NG2⁺ macrophage-like cells expressing a variety of neuroprotective factors ameliorate ischemic damage of the brain. *J. Cereb. Blood Flow Metab.* 30, 603–615. doi: 10.1038/jcbfm.2009.233
- Spohr, T. C., Dezonne, R. S., Rehen, S. K., and Gomes, F. C. (2011). Astrocytes treated by lysophosphatidic acid induce axonal outgrowth of cortical progenitors through extracellular matrix protein and epidermal growth factor signaling pathway. J. Neurochem. 119, 113–123. doi: 10.1111/j.1471-4159.2011.07421.x
- Starossom, S. C., Mascanfroni, I. D., Imitola, J., Cao, L., Raddassi, K., Hernandez, S. F., et al. (2012). Galectin-1 deactivates classically activated microglia and protects from inflammation-induced neurodegeneration. *Immunity* 37, 249–263. doi: 10.1016/j.immuni.2012.05.023

- Stevens, S. L., Leung, P. Y., Vartanian, K. B., Gopalan, B., Yang, T., Simon, R. P., et al. (2011). Multiple preconditioning paradigms converge on interferon regulatory factor-dependent signaling to promote tolerance to ischemic brain injury. J. Neurosci. 31, 8456–8463. doi: 10.1523/JNEUROSCI.0821-11.2011
- Stewart, C. R., Stuart, L. M., Wilkinson, K., van Gils, J. M., Deng, J., Halle, A., et al. (2010). CD36 ligands promote sterile inflammation through assembly of a Toll-like receptor 4 and 6 heterodimer. *Nat. Immunol.* 11, 155–161. doi: 10.1038/ni.1836
- Sun, S., Sursal, T., Adibnia, Y., Zhao, C., Zheng, Y., Li, H., et al. (2013). Mitochondrial DAMPs increase endothelial permeability through neutrophil dependent and independent pathways. *PLoS ONE* 8:e59989. doi: 10.1371/journal.pone. 0059989
- Suzuki, Y., Hattori, K., Hamanaka, J., Murase, T., Egashira, Y., Mishiro, K., et al. (2012). Pharmacological inhibition of TLR4-NOX4 signal protects against neuronal death in transient focal ischemia. *Sci. Rep.* 2, 896. doi: 10.1038/srep00896
- Suzuki, Y., Nakano, Y., Mishiro, K., Takagi, T., Tsuruma, K., Nakamura, M., et al. (2013). Involvement of Mincle and Syk in the changes to innate immunity after ischemic stroke. *Sci. Rep.* 3, 3177. doi: 10.1038/srep03177
- Takeuchi, M., Yanase, Y., Matsuura, N., Yamagishi Si, S., Kameda, Y., Bucala, R., et al. (2001). Immunological detection of a novel advanced glycation end-product. *Mol. Med.* 7, 783–791.
- Tang, S. C., Arumugam, T. V., Xu, X., Cheng, A., Mughal, M. R., Jo, D. G., et al. (2007). Pivotal role for neuronal Toll-like receptors in ischemic brain injury and functional deficits. *Proc. Natl. Acad. Sci. U.S.A.* 104, 13798–13803. doi: 10.1073/pnas.0702553104
- Taniguchi, H., Anacker, C., Suarez-Mier, G. B., Wang, Q., and Andreasson, K. (2011). Function of prostaglandin E2 EP receptors in the acute outcome of rodent hypoxic ischemic encephalopathy. *Neurosci. Lett.* 504, 185–190. doi: 10.1016/j.neulet.2011.09.005
- Tsai, S. Y., Segovia, J. A., Chang, T. H., Morris, I. R., Berton, M. T., Tessier, P. A., et al. (2014). DAMP molecule S100A9 acts as a molecular pattern to enhance inflammation during influenza A virus infection: role of DDX21-TRIF-TLR4-MyD88 pathway. *PLoS Pathog.* 10:e1003848. doi: 10.1371/journal.ppat.1003848
- Uchida, K. (2013). Redox-derived damage-associated molecular patterns: ligand function of lipid peroxidation adducts. *Redox Biol.* 1, 94–96. doi: 10.1016/j.redox.2012.12.005
- Walko, T. D. 3rd, Bola, R. A., Hong, J. D., Au, A. K., Bell, M. J., Kochanek, P. M., et al. (2014). Cerebrospinal fluid mitochondrial DNA: a novel DAMP in pediatric traumatic brain injury. *Shock* 41, 499–503. doi: 10.1097/SHK.00000000 00000160
- Wei, Y., Yemisci, M., Kim, H. H., Yung, L. M., Shin, H. K., Hwang, S. K., et al. (2011). Fingolimod provides long-term protection in rodent models of cerebral ischemia. *Ann. Neurol.* 69, 119–129. doi: 10.1002/ana.22186
- Weil, Z. M. (2012). Ischemia-induced hyperglycemia: consequences, neuroendocrine regulation, and a role for RAGE. *Horm. Behav.* 62, 280–285. doi: 10.1016/j.yhbeh.2012.04.001
- Wenceslau, C. F., McCarthy, C. G., Szasz, T., Spitler, K., Goulopoulou, S., Webb, R. C., et al. (2014). Mitochondrial damage-associated molecular patterns and vascular function. *Eur. Heart J.* 35, 1172–1177. doi: 10.1093/eurheartj/ehu047
- West, X. Z., Malinin, N. L., Merkulova, A. A., Tischenko, M., Kerr, B. A., Borden, E. C., et al. (2010). Oxidative stress induces angiogenesis by activating TLR2 with novel endogenous ligands. *Nature* 467, 972–976. doi: 10.1038/nature09421

- Yanai, H., Ban, T., Wang, Z., Choi, M. K., Kawamura, T., Negishi, H., et al. (2009). HMGB proteins function as universal sentinels for nucleicacid-mediated innate immune responses. *Nature* 462, 99–103. doi: 10.1038/ nature08512
- Yang, F., Wang, Z., Wei, X., Han, H., Meng, X., Zhang, Y., et al. (2014). NLRP3 deficiency ameliorates neurovascular damage in experimental ischemic stroke. J. Cereb. Blood Flow Metab. 34, 660–667. doi: 10.1038/jcbfm.2013.242
- Yang, Y., Liu, B., Dai, J., Srivastava, P. K., Zammit, D. J., Lefrançois, L., et al. (2007). Heat shock protein gp96 is a master chaperone for Toll-like receptors and is important in the innate function of macrophages. *Immunity* 26, 215–226. doi: 10.1016/j.immuni.2006.12.005
- Yednock, T. A., Cannon, C., Fritz, L. C., Sanchez-Madrid, F., Steinman, L., and Karin, N. (1992). Prevention of experimental autoimmune encephalomyelitis by antibodies against α4β1 integrin. *Nature* 356, 63–66. doi: 10.1038/ 356063a0
- Zhang, J., Takahashi, H. K., Liu, K., Wake, H., Liu, R., Maruo, T., et al. (2011). Anti-high mobility group box-1 monoclonal antibody protects the blood–brain barrier from ischemia-induced disruption in rats. *Stroke* 42, 1420–1428. doi: 10.1161/STROKEAHA.110.598334
- Zhang, Q., Raoof, M., Chen, Y., Sumi, Y., Sursal, T., Junger, W., et al. (2010). Circulating mitochondrial DAMPs cause inflammatory responses to injury. *Nature* 464, 104–107. doi: 10.1038/nature08780
- Zhong, Z., Zhai, Y., Liang, S., Mori, Y., Han, R., Sutterwala, F. S., et al. (2013). TRPM2 links oxidative stress to NLRP3 inflammasome activation. *Nat. Commun.* 4, 1611. doi: 10.1038/ncomms2608
- Zhu, W., Fan, Y., Frenzel, T., Gasmi, M., Bartus, R. T., Young, W. L., et al. (2008). Insulin growth factor-1 gene transfer enhances neurovascular remodeling and improves long-term stroke outcome in mice. *Stroke* 39, 1254–1261. doi: 10.1161/STROKEAHA.107.500801
- Zhu, W., Fan, Y., Hao, Q., Shen, F., Hashimoto, T., Yang, G. Y., et al. (2009). Postischemic IGF-1 gene transfer promotes neurovascular regeneration after experimental stroke. *J. Cereb. Blood Flow Metab.* 29, 1528–1537. doi: 10.1038/jcbfm.2009.75
- Ziegler, G., Freyer, D., Harhausen, D., Khojasteh, U., Nietfeld, W., and Trendelenburg, G. (2011). Blocking TLR2 in vivo protects against accumulation of inflammatory cells and neuronal injury in experimental stroke. J. Cereb. Blood Flow Metab. 31, 757–766. doi: 10.1038/jcbfm.2010.161

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