



Corrigendum: A Novel Microtubule-Tau Association Enhancer and Neuroprotective Drug Candidate: Ac-SKIP

Yanina Ivashko-Pachima and Illana Gozes*

Dr. Diana and Zelman Elton (Elbaum) Laboratory for Molecular Neuroendocrinology, Department of Human Molecular Genetics and Biochemistry, Sackler Faculty of Medicine, Sagol School of Neuroscience, Adams Super Center for Brain Studies, Tel Aviv University, Tel Aviv, Israel

Keywords: tau, microtubules, EBs, ADNP, SKIP

A Corrigendum on

OPEN ACCESS

Edited and reviewed by: Dirk M. Hermann, University of Duisburg-Essen, Germany

*Correspondence:

Illana Gozes igozes@tauex.tau.ac.il; igozes@post.tau.ac.il

Specialty section:

This article was submitted to Cellular Neuropathology, a section of the journal Frontiers in Cellular Neuroscience

> Received: 29 March 2021 Accepted: 28 April 2021 Published: 26 May 2021

Citation:

Ivashko-Pachima Y and Gozes I (2021) Corrigendum: A Novel Microtubule-Tau Association Enhancer and Neuroprotective Drug Candidate: Ac-SKIP. Front. Cell. Neurosci. 15:687301. doi: 10.3389/fncel.2021.687301 Α Novel Microtubule-Tau Association Enhancer and Neuroprotective Drug **Candidate: Ac-SKIP** bv Ivashko-Pachima, Y., and Gozes, Ι. (2019).Front. Cell. Neurosci. 13:435.

by Ivashko-Pachima, Y., and Gozes, I. (2019). Front. Cell. Neurosci. doi: 10.3389/fncel.2019.00435

In the original article, there were inconsistencies when comparing one by one the panels of **Figure 4A** in the main text and the representative extended immunoblot results in **Supplementary Figure S3**, as published. Thus, some of the representative images in the main text were unfortunately represented by extended mirrored, duplicated and rotated blots as explained below. It should be emphasized that for scientific rigor and data corroboration, each experiment was performed in triplicates and repeated three times and each immunoblot was exposed at least three times (three films), hence, the blot shown as a full image in the original supplementary **Figure S3** legend. To maintain scientific rigor, there were 36 blots and almost 100 films. Our mistakes, corrected below, stem from an unfortunate mix-up coupled with an extremely high similarity between all repetitive experiments, we apologize for the mistakes.

Corrections have been made to **Figure 4**, panel A, and **Supplementary Figure S3**. The corrected figures are shown below.

The authors apologize for these errors and state that they do not change the scientific conclusions of the article in any way. The original article has been updated.

Copyright © 2021 Ivashko-Pachima and Gozes. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



FIGURE 4 | quantification of soluble tubulin ratios. The intensity of each band was quantified by densitometry and the soluble protein ratio was calculated by dividing the densitometric value of soluble proteins by the total protein content (S/[S + P]). Statistical analysis was performed by One Way ANOVA with Tukey HSD. P < 0.05, P < 0.01, P < 0.01; P < 0.001; Control, n = 15; Zn, n = 18; Zn + Ac-SKIP 10^{-12} M, n = 15; $Zn + SKIP <math>10^{-12}$ M, n = 9; Zn + Ac-SKIP 10^{-9} M, n = 18; $Zn + SKIP <math>10^{-9}$ M, n = 11. **(C,D)** A Co-IP assay was performed with EB1 or Tau antibodies, linked to agarose beads. SKIP (2 µg/sample) or Ac-SKIP (2 µg/sample), diluted into Pierce lysis buffer (see section "Materials and Methods") or the equal volume of lysis buffer w/o peptides (IP: EB1 Cont; IP: Tau Cont) were added to cell lysate of differentiated SH-SY5Y cells, 15 min before EB1 or Tau column application (IP: EB1; IP: Tau). Sequential IP elution fractions (E1, E2) were further analyzed by immunobloting with EB1, Tau, and Tubulin antibodies (IB: EB1; IB: Tau; IB: Tubulin). In addition, columns with free agarose beads were used as negative controls (IP: Neg cont). The intensity of each band was quantified by densitometry. The bar graph shows the ratio of band intensities (densitometric value \pm SEM) obtained upon SKIP and Ac-SKIP treatments as compared with non-treated cells (Cont). Statistical analysis was performed by One Way ANOVA with LSD HSD. Experiments were independently repeated three times. P < 0.05, P < 0.01, P < 0.001. **(E)** Cell lysates without column exposure were used as positive controls (Input). IP – immunoprecipitation, IB – immunoblot.

