



# **Transition Dynamics of a Dentate Gyrus-CA3 Neuronal Network during Temporal Lobe Epilepsy**

Liyuan Zhang<sup>1</sup>, Denggui Fan<sup>2</sup> and Qingyun Wang<sup>1\*</sup>

<sup>1</sup> Department of Dynamics and Control, Beihang University, Beijing, China, <sup>2</sup> Department of Information and Computing Science, School of Mathematics and Physics, University of Science and Technology Beijing, Beijing, China

In temporal lobe epilepsy (TLE), the variation of chemical receptor expression underlies the basis of neural network activity shifts, resulting in neuronal hyperexcitability and epileptiform discharges. However, dynamical mechanisms involved in the transitions of TLE are not fully understood, because of the neuronal diversity and the indeterminacy of network connection. Hence, based on Hodgkin-Huxley (HH) type neurons and Pinsky-Rinzel (PR) type neurons coupling with glutamatergic and GABAergic synaptic connections respectively, we propose a computational framework which contains dentate gyrus (DG) region and CA3 region. By regulating the concentration range of N-methyl-D-aspartate-type glutamate receptor (NMDAR), we demonstrate the pyramidal neuron can generate transitions from interictal to seizure discharges. This suggests that enhanced endogenous activity of NMDAR contributes to excitability in pyramidal neuron. Moreover, we conclude that excitatory discharges in CA3 region vary considerably on account of the excitatory currents produced by the excitatory pyramidal neuron. Interestingly, by changing the backprojection connection, we find that glutamatergic type backprojection can promote the dominant frequency of firings and further motivate excitatory counterpropagation from CA3 region to DG region. However, GABAergic type backprojection can reduce firing rate and block morbid counterpropagation, which may be factored into the terminations of TLE. In addition, neuronal diversity dominated network shows weak correlation with different backprojections. Our modeling and simulation studies provide new insights into the mechanisms of seizures generation and connectionism in local hippocampus, along with the synaptic mechanisms of this disease.

Keywords: temporal lobe epilepsy, dynamical transition, dentate gyrus-CA3 network, glutamatergic receptor, GABAergic receptor

# INTRODUCTION

A great many *in vivo* and *in vitro* studies focus on revealing the mechanism underlying epilepsy, and significant progresses have been made in understanding the causes of temporal lobe epilepsy (TLE). TLE is the most common form of localization-related epilepsy in adults accounting for approximately  $60 \sim 80\%$  of all patients with epilepsy (Tatum, 2012). It is a chronic neurological disorder that is caused by various factors including neuron loss (Margerison and Corsellis, 1966), vulnerability of mossy cells, hippocampal sclerosis, and excessive expression of brain derived

#### OPEN ACCESS

#### Edited by:

Jun Ma, Lanzhou University of Technology, China

#### Reviewed by:

Xiaoli Yang, Shaanxi Normal University, China Yong Xie, Xi'an Jiaotong University, China Li Yuye, Chifeng University, China

> \*Correspondence: Qingyun Wang

nmqingyun@163.com

**Received:** 24 May 2017 **Accepted:** 26 June 2017 **Published:** 11 July 2017

#### Citation:

Zhang L, Fan D and Wang Q (2017) Transition Dynamics of a Dentate Gyrus-CA3 Neuronal Network during Temporal Lobe Epilepsy. Front. Comput. Neurosci. 11:61. doi: 10.3389/fncom.2017.00061 neurotrophic factor (BDNF; McNamara and Scharfman, 2010). One of the most important goals in TLE research today is to address the underlying mechanisms that contribute to seizures (Liotta et al., 2011; Curia et al., 2014). Another important issue is establishing the accurate network to understand the transition from normal state to morbid state. Temporal lobe is the most epileptogenic area of brain (Avoli et al., 2002). Focal seizure may be originated from internal structures such as hippocampus, which consists of dentate gyrus (DG) region, CA3 region, CA1 region, and subiculum (Lazarov and Hollands, 2016). In the field of epilepsy, the relationship between inhibition/excitation balance and epileptogenesis has been largely investigated in experimental studies (Amakhin et al., 2016). Mossy cell has excitatory glutamatergic receptors consisting of N-methyl-Daspartate receptor (NMDAR) and α-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid receptor (AMPAR; Min et al., 1998). Pyramidal neuron (Shi et al., 2001) and granule cell (Ye et al., 2005) are adjusted by AMPAR. Furthermore, inhibitory GABAergic receptor exists in fast spiking interneuron and oriens lacunosum-moleculare (O-LM) cell (Gloveli et al., 2005; Tort et al., 2007). Epilepsy has been shown to be associated with a dysfunction of inhibitory signaling mediated by GABAA receptors (Sloviter, 1987). In particular, GABAA receptors from epileptic tissue become less responsive to repeated activation than those from healthy tissue (Stamboulian-Platel et al., 2016). This suggests that promoting the activation of GABA<sub>A</sub> receptors might be an effective antiepileptic strategy to suppress focal seizures in TLE. The well-known signaling pathways between hippocampal regions are the perforant path from entorhinal cortex (EC) to DG and the mossy fibers from DG to CA3 (Ahn et al., 2016). Excitatory signal from DG granule cell could be transmitted by mossy cell and relayed by CA3 pyramidal neuron (Amaral et al., 2007). In addition, CA3 pyramidal neuron also projects "back" to the dentate gyrus (Scharfman, 2007). Previous studies have shown a pyramidal neuron-mossy cell-interneurongranule cell pathway is more responsible than a simply pyramidal neuron-interneuron-granule cell route (Scharfman, 1994; Kneisler and Dingledine, 1995). Some evidences show that CA3 pyramidal neuron can not only activate but also inhibit granule cell indirectly (Scharfman, 2016). However, the exact backprojection mechanism related to excitatory or inhibitory receptors is not fully revealed.

Many computational models have been proposed to describe epilepsy. These are divided to models of interictal activity and models of ictal activity (Wendling et al., 2016). Interictal epileptic spikes (IEs) are often observed in human partial epilepsies (Ratnadurai-giridharan et al., 2012). IEs correspond to transient signals presenting with a more or less sharp initial component sometimes followed by a slower, more or less pronounced, component of opposite polarity (Chauvière et al., 2012). El-Hassar et al. employed a pilocarpine model of TLE to change the glutamatergic receptors and GABAergic receptors during epileptogenesis, which has been investigated in a neural mass model (NMM) of CA1 hippocampal region (El-Hassar et al., 2007). The interictal-like activity was reproduced, which was derived from extensive simulations where model parameters were varied at the soma and dendrites of the

pyramidal cell. Ratnadurai-Giridharan et al. generated template IEs using a subset of artificial paroxysmal depolarization shift (PDS) constructs (Ratnadurai-giridharan et al., 2012). NMM was also employed by Wendling et al. to simulate the transition from interictal to fast ictal activity, which further explained the impairment of dendritic inhibition (Wendling et al., 2012). For partial epilepsies, seizures are often characterized by the appearance of fast oscillation in LFPs or EEG signals (Schiff et al., 2000). High fast oscillations (HFOs) are usually associated with lower frequency range such as beta band and low gamma band (Wendling et al., 2002). Traub et al. developed a model based on postulated electrical coupling between pyramidal cell axons and reproduced very fast oscillations (VFOs; Traub and Bibbig, 2000; Scharfman and Buckmaster, 2014). Inspired by Traub's model, Munro et al. showed that an axonal plexus could also exhibit a second, distinctly different kind of VFO in a wide parameter range, and analyzed the role of electrical couplings among neurons (Munro and Börgers, 2010). Following pioneer studies about hippocampal CA3 area, scholars developed detailed models to investigate the network mechanisms underlying focal seizure initiation and propagation in other brain structures. Van et al. established a neural network with 656 neurons and found that weakly coupled cortical network could create synchronized cellular activity and seizure-like bursting (Van et al., 2005). Consequently, research on computational modeling is important for understanding the transitions of TLE, together with regional connection mechanism.

As discussed above, richness and complementarity of the various modeling approaches in the field of epilepsy research have been shown. However, in order to provide deeper understanding about discharge transitions in DG-CA3 area during TLE and investigate the backprojection mechanism behind synaptic connection, a neuronal diversity leading comprehensive network model is required. Therefore, we propose the overall structure of a comprehensive hippocampal network model that includes DG region containing granule cell, mossy cell and GABAergic interneuron, and CA3 region involving pyramidal neuron, GABAergic interneuron and O-LM cell. Cells of each region are global connection and there exists a backprojection path from CA3 to DG. In this paper, we conduct computational modeling based on biological mechanism that can reproduce the epileptiform propagation activities of pyramidal neurons, including interictal, preictal, and seizure onset, by increasing the NMDA junction conductance strength. We then compare the regional characteristics of CA3 and DG. Specifically, we change the backprojection positions and strength to study counterpropagation mechanism contributed by glutamatergic and GABAergic receptor, respectively.

# **DG-CA3 NETWORK MODEL FORMATION**

We use Hodgkin-Huxley (HH) type models to imitate granule cell (Yuen and Durand, 1991), GABAergic interneuron, pyramidal cell, and O-LM cell (Cutsuridis et al., 2010), respectively. The mossy cell model is based on a twocompartment model which was introduced by Pinsky and



(B) The backprojection is from pyramidal cell to GABAergic interneuron of DG.

Rinzel (1995). For DG region, depending on AMPA synaptic connection, granule cell innervates mossy cell and GABAergic interneuron. Then GABAergic interneuron returns inhibitory synaptic inputs to granule cell and mossy cell, through GABAA synaptic connection. There exist NMDA type and AMPA type receptors in mossy cell, so as to propagate excitatory inputs to granule cell and GABAergic interneuron, respectively. Then DG region delivers excitatory synaptic inputs to CA3 region through mossy fiber path. As for CA3 region, mossy cell excites pyramidal cell relaying on NMDA and AMPA coupling connections. Pyramidal cell couples with GABAergic interneuron and O-LM cell by AMPA coupling connections, respectively. GABAA synaptic connections are employed, by which GABAergic interneuron and O-LM cell could inhibit other neurons. Besides, the backprojection can locate either to soma of mossy cell or to GABAergic interneuron of DG. The DG-CA3 network schematic plot is presented in Figure 1 (See Appendix in Supplementary Material for network modeling methods).

## RESULTS

## Effects of NMDAR from DG to CA3

One of the most disabling aspects of epilepsy is the unpredictable timing of the attacks. And, the study of transition from interictal to ictal processes is important for not only the insights into mechanism of epileptic seizure, but also the potential clinical applications for seizure detection and warning. Thus, we construct a micro-network to elucidate factors might be responsible for state-dependent changes in chemical receptors concentrations that subsequently lead to more seizure activities. In our study, network activity can be modified by

increasing the glutamate releasing in presynaptic neuron, which leads to TLE action potential generation in the postsynaptic neuron. In this section, from pyramidal cell to mossy cell i.e.,  $g_{P-MS}$ , is set as 0.0001 (shown as Figure 1A). All of the neurons are set to the same external stimulus conditions. The simulation work is carried out under the circumstance of MATLAB R2013a. Simulation duration is 10 s, but we select a potential section of 1 or 2s in order to obtain unambiguous variation. Time series, spectrograms, frequency-power figures and interspike interval (ISI) bifurcation diagram (for 10s) are plotted to show firing behavior of pyramidal cell as the parameter changes. From Figure 2A, we can observe rich dynamical transitions as the bifurcation parameter  $g_{NMDA-P}$ increases. First, the slow interictal continuous spiking whose characteristic is 20~50 ms ISI is generated by decreasing inhibition function on pyramidal cell. As shown in Figure 2 (c1), peak valleys at 17.2 Hz own the maximum power compared with peaks. Thereafter, we conduct a moderate increase of  $g_{NMDA-P}$ , pyramidal cell then displays preictal discharges presenting fast spiking with a dominant frequency about 37.4 Hz which is demonstrated in Figure 2 (c2). From Figure 2D, we observe that there exists a bifurcation point near  $g_{NMDA-P} = 0.002$ , which is the boundary of preictal and seizure onset. Following preictal discharge, seizure initiates and generates gamma (30~80 Hz) oscillations in hippocampal network. In the beginning, gamma oscillation demonstrates small bursts with dominant frequency of 51.5 Hz (shown in Figure 2 (c3)) and short ISI. Then oscillation splits into small bursts with long ISI. From Figure 2 (c4-c5) we notice that dominant frequency and amplitude show a descending trend, which implies that excessive excitatory current inhibits the neuron from discharging. In short, due to the increasing



gluta matergic conductance  $\mathbf{g}_{NMDA-P}$ , pyramidal cell could present TLE discharge transitions from interictal to seizure on set. Meanwhile, GABAergic interneuron and O-LM cell receive excitatory inputs from pyramidal cell and provide inhibitory output to CA3 region. Compared **Figures 2,3**,



it is revealed that GABAergic interneuron and O-LM cell become excited, accompanied by an increase of glutamatergic receptor. Figures 3A,B show potential evolutions of GABAergic interneuron as  $g_{NMDA-P} = 0.001$  and  $g_{NMDA-P} = 0.1$ , respectively. In addition, the ISI bifurcation plot is demonstrated in Figure 3C. Discharge pattern initially presents irregular spiking with longer interval, which, whereafter, transfers to fast spiking with shorter interval. Similarly, discharge transition situations of O-LM cell are displayed in Figures 3D-F. Also, potential evolution shows the process from slow spiking to fast spiking. However, the ISI of O-LM cell is obviously longer than that of GABAergic interneuron. This implies that firing rate of O-LM cell is lower than that of GABAergic interneuron, but varies dramatically in contrast. Nevertheless, two types of neurons only receive excitatory inputs from pyramidal cell so as to make the whole CA3 network become inhibitive. GABAergic interneuron and O-LM cell could not demonstrate abundant TLE discharge patterns on account of their intrinsic inhibitory mechanism. In spite of this, the enhancement of excitatory coupling strength between mossy cell and pyramidal cell could promote excitability of CA3 and induce TLE discharge transitions of pyramidal cell.

Figure 4 demonstrates time series and ISI bifurcation diagrams of cells within DG region. As shown in Figure 4, under the condition of weakprojection from pyramidal cell to mossy cell, e.g.,  $g_{P-MS} = 0.0001$ , NMDAR has little influence on discharge transitions of DG cells. Specifically, although the discharge interval of mossy cell is a little longer for the much small  $g_{NMDA-P}$  than that of relatively large  $g_{NMDA-P}$ , it maintains spiking activity no matter how g<sub>NMDA-P</sub> varies. Similarly, both the granule cell and GABAergic interneuron always demonstrate fast spiking activities. However, it can be observed that GABAergic interneuron oscillates with much higher frequency than that of mossy cell. This suggests that the cells within DG region possess weak correlations under the circumstance of weak backprojection which could be factored into the robustness of DG cells to the changes of NMDAR. In addition, it has been proposed that mossy cellmediated excitation of granule cell is increased after dysfunction, causing hyperexcitability. In other aspects, changes of inhibitory









control of granule cell including GABA<sub>A</sub> receptors might block excitatory counterpropagation from CA3 region to DG region. Therefore, in the following section we will explore the effect of backprojection, which could contribute to the hyperexcitability or suppression of DG cells.

# Effects of Backprojection from CA3 to DG

CA3 and DG are the main subfields of hippocampal formation. They receive direct innervations from EC, while previous studies believed that they were generally considered as a series of unidirectionally connected components forming the intrahippocampal "trisynaptic loop" (Avoli et al., 2002; Oliveira et al., 2011). Within the main associational network of the hippocampal formation, axon collaterals of pyramidal cell return to DG where they likely innervate mossy cells, GABAergic interneuron and granule cell (Freund, 1992; Scharfman, 2007). It is hypothesized that neuronal information emerging from the recurrent network of the CA3 region is fed back to its main driving input, the DG (Scharfman, 1994). In addition, previous physiological observation has shown that stimulation of CA3 region typically induces large inhibitory postsynaptic potentials (IPSPs) in granule cells, and it is hypothesized that this inhibition is conveyed by the excitation of GABAergic interneuron (Scharfman, 2016). However, epileptic pyramidal cell could counterpropagate morbid excitatory information to mossy cell or granule cell if GABAA receptor appears inhibitory dysfunction. Thus, it suggests that CA3-DG feed-back pathway can lead to both suppression and excitation of DG region. According to this thinking, the study of effects of backprojection is carried out.

We connect the neural network according to glutamatergic type backprojection plotted in Figure 1A, it means the backprojection is from pyramidal cell to mossy cell. Figure 5 plots potential evolutions and ISI bifurcation graphs of DG with  $g_{P-MS} = 0.0001$  and  $g_{P-MS} = 0.01$ , respectively. Besides,  $g_{NMDA-P} = 0.001$  in this process. As time goes on, mossy cell exhibits transition from normal discharge to epileptic lowvoltage fast activity, which is shown in Figures 5A-FT. What's more, as  $g_{P-MS}$  increasing, amplitude of oscillation decreases to lower values. In particular, in the beginning of Figures 5C,F we can observe distinct ISI reduction of mossy cell thereby increases with  $g_{P-MS}$  increasing until neural system reaches to the stable states. Granule cell also exhibits small-amplitude excitatory oscillation with long latency in contrast with mossy cell. GABAergic interneurons show the damped oscillations into equilibrium states. The firing latency of it can be extended when  $g_{P-MS}$  increases from 0.0001 to 0.01, which also suggests the enhancement in excitability of GABAergic interneuron.

Besides, we analyze the corresponding dominant frequency of DG-CA3 network, which is illustrated in **Figure 6**. Following with the increasing of  $g_{P-MS}$ , the dominant frequencies of mossy cell and granule cell vibrate at first, which then can be elevated to



a higher i.e., 32 Hz, and maintained till the last. It is obvious that enhancing the backprojection coupling strength from pyramidal cell to mossy cell could promote hyperexcitability of mossy cell and granule cell, and produce low gamma oscillation. Meanwhile, GABAergic interneuron rises slowly with lower frequency value in contrast with mossy cell and granule cell. As for CA3 region, dominant frequencies of pyramidal cell and GABAergic interneuron also rise up slightly. They discharge at a relative constant frequency, such as pyramidal cell is about 33 Hz and GABAergic interneuron fires at a frequency about 32 Hz. In addition, the dominant frequency of O-LM cell increases with two jumps from 5.5 Hz to about 12 Hz. It implies that excited mossy cell could motivate CA3 region through mossy fiber even though the feed-forward coupling strength is not enhanced.

We also investigate the correlations among neurons. We employ correlation coefficient to estimate correlation and synchronism. Linear correlation coefficient is defined as:

$$C_{xy}\left(l\right) = \left\langle \left(\frac{x_i - \bar{x}}{\sigma_x}\right) \left(\frac{y_{i+l} - \bar{y}}{\sigma_y}\right) \right\rangle_i \tag{1}$$

where  $\bar{x}$  and  $\bar{y}$  are mean values, respectively.  $\sigma_x$  and  $\sigma_y$ represent variances separately. *l* denotes time delay (here *l* equals to zero) and  $\langle \cdot \rangle$  expresses average value. Figure 7A reflects statistical overview of correlation coefficient in CA3 region, Figure 7B describes the interrelation in DG region and Figure 7C represents the relationships between CA3 region and DG region. From Figure 7A we can clearly observe that pyramidal cell and GABAergic interneuron show stronger negative relation than that between pyramidal cell and O-LM cell. Whereas the relationship between GABAergic interneuron and O-LM is weakly positive. In the case of DG region, mossy cells are highly positively related, the correlation coefficient of which is larger than 0.6. What's more, mossy cell's soma and other neurons demonstrate weak positive correlation, while mossy cell's dendrite and the rest cells show weak negative correlations. In addition, granule cell and GABAergic interneuron coupled with each other also show a weak correlation. In the Figure 7C, we consider the relationships between regions of CA3 and DG. Firstly, pyramidal cell has positive correlations with three types of neurons within DG except for dendrite of mossy cell. Then, GABAergic interneuron of CA3 weakly and positively interacts with DG region apart from soma of mossy cell.





Besides, O-LM cell has weak correlation with DG region. View the situation as a whole, pyramidal cell shows the maximum fluctuation and susceptibility, while O-LM cell is in a completely opposite situation. Mossy cell has the highest value of correlation coefficient. Good correlation between pyramidal cell and GABAergic interneuron of CA3 could be found. In addition, GABAergic interneuron of DG couples with granule cell with a relative higher correlation coefficient. Judging from **Figures 5**–7, we summarize that enhancing backprojection strength from pyramidal cell to mossy cell's soma could strengthen the degree of epilepsy morbidity in DG region, even though neurons do not fire hyper-synchronously, which is induced by neuronal diversity.

Next, we reassemble the network in accordance with GABAergic type backprojection shown in Figure 1B, where backprojection is from pyramidal cell to GABAergic interneuron of DG region, i.e.,  $g_{P-DI}$ . Besides,  $g_{NMDA-P} = 0.001$  in this process. Figure 8 presents time series and ISI birfucation graphs of DG region with the increasing of  $g_{P-DI}$ . As depicted in Figures 8A,B,D,E, firing interval of mossy cell is prolonged, which means inhibitory current generated by GABAergic interneuron blocks excitatory propagation from pyramidal cell to mossy cell. Figures 8C,F give ISI evolutions about mossy cell. It is shown that mossy cell discharges with small ISI at first, then the ISI of mossy cells can be gradually enlarged when  $g_{P-DI}$ increased beyond the critical point  $g_{P-DI} = 0.02$ . As for granule cell, similar ISI variation can be observed in Figure 8I, but the oscillations seems to be more stochastic compared to mossy cells. As backprojection strength is enhanced, GABAergic interneuron of DG region leads a transition from damped oscillations to fast spiking with low amplitude, shown in **Figures 8J,K**, which suggests that the activity of GABAergic interneuron has been enhanced.

Evolution of dominant frequencies is illustrated in **Figure 9**. Affected by GABAergic interneuron, dominant frequency of mossy cell decreases distinctly, and so does pyramidal cell. Granule cell fluctuates mildly from 21.25 to 18.5 Hz. Dominant frequency of GABAergic interneuron in CA3 basically maintains a constant as a whole, except for the outliers within  $g_{P-DI} \in$ (0.017, 0.03). Similarly, dominant frequency of O-LM cell decays from 10 to 5 Hz after several abrupt jumps. It is noteworthy that GABAergic interneuron of DG has almost the same variation trend with pyramidal cell. Although dominant frequency of GABAergic interneuron of DG declines, it discharges at a frequency of 29 Hz in the beginning and owns higher frequency, by making a comparison between **Figures 6**, **9**, which implies that the excitability of GABAergic interneuron of DG is enhanced.

In order to be consistent with previous study, analysis about correlation in this case is exhibited in **Figure 10**. Firstly, we conclude from **Figure 10A** that correlation between pyramidal cell and GABAergic interneuron is negative which is weaker in comparison with **Figure 7A**. Similarly, pyramidal cell also weakly related to O-LM cell. Besides, GABAergic interneuron has little effects on O-LM cell. In consideration of DG region, mossy cell's soma and dendrite still have the maximum positive correlation, while the whole mossy cell has negative correlation with granule cell and positive correlation with



GABAergic interneuron, respectively. Nevertheless, granule cell and GABAergic interneuron present weak correlation. At last, we study correlation between CA3 and DG. Pyramidal cell demonstrates positive correlation with DG region, which means pyramidal cell could motivate neurons of DG. In addition, we find that O-LM cell weakly correlates with DG region.

After the comprehensive comparisons between Figures 5, 10, it is proved that if pyramidal cell irritates soma of mossy cell, the latter could display hyper-excitatory and release excitatory NMDAR and AMPAR to postsynaptic neuron. Together with mossy cell, granule cell, and GABAergic interneuron of DG also demonstrate an increase in excitability, which behave as low amplitude oscillation. At the same time, CA3 region could also be slightly activated through the mossy fiber. What's more, glutamatergic type backprojection could promote dominant frequency of DG-CA3 network and then strengthen its excitability. Besides, granule cell couples with GABAergic interneuron of DG in a relative high positive correlation, when compared with pyramidal cell which synchronize negatively with GABAergic interneurons. When pyramidal cell connects DG region with GABAergic type backprojection, except GABAergic interneuron of DG, morbid excitability of DG-CA3 network decreases because of the excessive inhibitory currents. In addition, GABAergic type backprojection could decrease dominant frequency of the whole network. Thus, the activation GABAergic interneuron of DG could block counterpropagation of pathological discharges from CA3 to DG, which may prevent propagation of hypersynchronous discharge from another perspective. At last, we can conclude that dendrite of mossy cell discharges with a negative synchronous coefficient and pyramidal cell couples with GABAergic interneuron of DG in positive correlation. Phenomenon of correlation is not obvious due to the neuronal diversity.

# CONCLUSION

Using a combination of TLE theory and numerical advanced models, we constructed a DG-CA3 network to explore the mechanism underlying TLE transitions and the network synaptic connection. This network model offers novel opportunities to model and study epilepsy behavior from dynamics perspective. Action potentials of different cells within the DG-CA3 network are used to describe the network activity state.

Firing characteristics of action potentials are mainly reflected by the spectrograms and interspike interval (ISI). Firstly, by enhancing the coupling strength of DG-CA3 pathway, we have demonstrated complex TLE transitions including interictal, preictal, and seizure onset in pyramidal cell, meanwhile, excitability of CA3 region increases. In particular, in Figure 2 we employed the time-frequency graph to more evidently observe the changes of activity frequencies during TLE transition process. In addition, taking advantage of the connectionism of hippocampal, we have studied the effects of backprojetion during TLE transitions. On the one hand, the detailed dynamics and qualitative analysis have shown that the glutamatergic type backprojetion from pyramidal cell to mossy cell contributes to excitatory morbid discharges of DG region. On the other hand, simulation results also have indicated that the GABAergic type backprojection from pyramidal cell to GABAergic interneuron could block excitatory counterpropagation. Whereas the network acts less hypersynchronous, which is due to the neuronal diversity playing the dominant role in synchronization. Our approach may have validity only within a small range of physiological structure, nevertheless, they can give insight about

## REFERENCES

- Ahn, S., Jun, S. B., Lee, H. W., and Lee, S. (2016). Computational modeling of epileptiform activities in medial temporal lobe epilepsy combined with *in vitro* experiments. J. Comput. Neurosci. 41, 1–17. doi: 10.1007/s10827-016-0614-8
- Amakhin, D. V., Ergina, J. L., Chizhov, A. V., and Zaitsev, A. V. (2016). Synaptic conductances during interictal discharges in pyramidal neurons of rat entorhinal cortex. *Front. Cell. Neurosci.* 10:233. doi: 10.3389/fncel.2016.00233
- Amaral, D. G., Scharfman, H. E., and Lavenex, P. (2007). The dentate gyrus: fundamental neuroanatomical organization (dentate gyrus for dummies). *Prog. Brain Res.* 163, 3–22. doi: 10.1016/S0079-6123(07)63001-5
- Avoli, M., D'Antuono, M., Louvel, J., Köhling, R., Biagini, G., and Pumain, R., et al. (2002). Network and pharmacological mechanisms leading to epileptiform synchronization in the limbic system *in vitro*. *Prog. Neurobiol.* 68, 167–207. doi: 10.1016/S0301-0082(02)00077-1
- Chauvière, L., Doublet, T., Ghestem, A., Siyoucef, S. S., Wendling, F., and Huys, R., et al. (2012). Changes in interictal spike features precede the onset of temporal lobe epilepsy. *Ann. Neurol.* 71, 805–814. doi: 10.1002/ana.23549
- Curia, G., Lucchi, C., Vinet, J., Gualtieri, F., Marinelli, C., and Torsello, A., et al. (2014). Pathophysiogenesis of mesial temporal lobe epilepsy: is prevention of damage antiepileptogenic? *Curr. Med. Chem.* 21, 663–688. doi: 10.2174/0929867320666131119152201
- Cutsuridis, V., Graham, B., Cobb, S., and Vida, I. (2010). *Hippocampal Microcircuits*. New York, NY: Springer Press.
- El-Hassar, L., Milh, M., Wendling, F., Ferrand, N., Esclapez, M., and Bernard, C. (2007). Cell domain-dependent changes in the glutamatergic and GABAergic drives during epileptogenesis in the rat cal region. *J. Physiol.* 578, 193–211. doi: 10.1113/jphysiol.2006.119297
- Ermentrout, G. B., and Kopell, N. (1998). Fine structure of neural spiking and synchronization in the presence of conduction delay. *Proc. Natl. Acad. Sci.* U.S.A. 95, 1259–1264. doi: 10.1073/pnas.95.3.1259
- Freund, T. F. (1992). GABAergic septal and serotonergic median raphe afferents preferentially innervate inhibitory interneurons in the hippocampus and dentate gyrus. *Epilepsy Res. Suppl.* 7, 79–91.
- Gloveli, T., Dugladze, T., Rotstein, H. G., Traub, R. D., Monyer, H., and Heinemann, U., et al. (2005). Orthogonal arrangement of rhythm-generating microcircuits in the hippocampus. *Proc. Natl. Acad. Sci. U.S.A.* 102, 13295–13300. doi: 10.1073/pnas.0506259102

experimental paradigms via simulations and analyses bridging the gap neuronal excitability, network and TLE evolution across large temporal scales. Hence, we hope that our work will be useful for further epilepsy disease analysis and be helpful for other nerve system disease.

#### **AUTHOR CONTRIBUTIONS**

LZ, DF, and QW designed and performed the research as well as wrote the paper.

## ACKNOWLEDGMENTS

This research was supported by the National Science Foundation of China (Grants 11325208, 11572015, 11172017).

#### SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: http://journal.frontiersin.org/article/10.3389/fncom. 2017.00061/full#supplementary-material

- Kneisler, T. B., and Dingledine, R. (1995). Synaptic input from CA3 pyramidal cells to dentate basket cells in rat hippocampus. J. Physiol. 487, 125–146. doi: 10.1113/jphysiol.1995.sp020866
- Lazarov, O., and Hollands, C. (2016). Hippocampal neurogenesis: learning to remember. Prog Neurobiol. 1, 138–140. doi: 10.1016/j.pneurobio.2015.12.006
- Liotta, A., Caliskan, G., Ul, H. R., Hollnagel, J. O., Rösler, A., and Heinemann, U., et al. (2011). Partial disinhibition is required for transition of stimulus-induced sharp wave-ripple complexes into recurrent epileptiform discharges in rat hippocampal slices. J. Neurophysiol. 105, 172–187. doi: 10.1152/jn.00186.2010
- Margerison, J. H., and Corsellis, J. A. (1966). Epilepsy and the temporal lobes. A clinical, electroencephalographic and neuropathological study of the brain in epilepsy, with particular reference to the temporal lobes. *Brain* 89, 499–530. doi: 10.1093/brain/89.3.499
- McNamara, J. O., and Scharfman, H. E. (2010). Temporal lobe epilepsy and the BDNF receptor, TrkB. *Epilepsia* 51(Suppl. 5), 46–46. doi: 10.1111/j.1528-1167.2010.02832.x
- Min, M. Y., Rusakov, D. A., and Kullmann, D. M. (1998). Activation of AMPA, kainate, and metabotropic receptors at hippocampal mossy fiber synapses. *Neuron* 21, 561–570. doi: 10.1016/S0896-6273(00)80566-8
- Munro, E., and Börgers, C. (2010). Mechanisms of very fast oscillations in networks of axons coupled by gap junctions. J. Comput. Neurosci. 28, 539–555. doi: 10.1007/s10827-010-0235-6
- Oliveira, M. S., Pacheco, L. F., Mello, C. F., Cavalheiro, E. A., and Garrido-Sanabria, E. R. (2011). Epileptiform activity in the limbic system. *Front. Biosci.* 3:565. doi: 10.2741/s172
- Olufsen, M., Whittington, M., Camperi, M., and Kopell, N. (2003). New functions for the gamma rhythm: population tuning and preprocessing for the beta rhythm. J. Comput. Neurosci. 14, 33–54. doi: 10.1023/A:1021124317706
- Pinsky, P. F., and Rinzel, J. (1995). Intrinsic and network rhythmogenesis in a reduced Traub model for CA3 neurons. J. Comput. Neurosci. 2, 275–275. doi: 10.1007/BF00961439
- Ratnadurai-giridharan, S., Stefanescu, R. A., Khargonekar, P. P., Carney, P. R., and Talathi, S. S. (2012). Genesis of interictal spikes in the CA1: a computational investigation. *Front. Neural Circuits* 13:2. doi: 10.3389/fncir.2014.00002
- Scharfman, H. E. (1994). Evidence from simultaneous intracellular recordings in rat hippocampal slices that area CA3 pyramidal cells innervate dentate hilar mossy cells. J. Neurophysiol. 72, 2167–2180.
- Scharfman, H. E. (2007). The CA3 "backprojection" to the dentate gyrus. Prog. Brain Res. 163, 627–637. doi: 10.1016/S0079-6123(07)63034-9

- Scharfman, H. E. (2016). The enigmatic mossy cell of the dentate gyrus. Nat. Rev. Neurosci. 17, 562–575. doi: 10.1038/nrn.2016.87
- Scharfman, H. E., and Buckmaster, P. S. (2014). Issues in Clinical Epileptology: A View from the Bench. Dordrecht: Springer Press.
- Schiff, S. J., Colella, D., Jacyna, G. M., Hughes, E., Creekmore, J. W., and Marshall, A., et al. (2000). Brain chirps: spectrographic signatures of epileptic seizures. *Clin. Neurophysiol.* 111, 953–958. doi: 10.1016/S1388-2457(00)00259-5
- Shi, S. H., Hayashi, Y., Esteban, J. A., and Malinow, R. (2001). Subunit-specific rules governing AMPA receptor trafficking to synapses in hippocampal pyramidal neurons. *Cell* 105, 331–343. doi: 10.1016/S0092-8674(01)00321-X
- Sloviter, R. S. (1987). Decreased hippocampal inhibition and a selective loss of interneurons in experimental epilepsy. *Science* 235, 73–76. doi: 10.1126/science.2879352
- Stamboulian-Platel, S., Legendre, A., Chabrol, T., Platel, J. C., Pernot, F., and Duveau, V., et al. (2016). Activation of gaba a, receptors controls mesiotemporal lobe epilepsy despite changes in chloride transporters expression: *in vivo*, and *in silico*, approach. *Exp. Neurol.* 284, 11–28. doi: 10.1016/j.expneurol.2016.07.009
- Tatum, W. O. (2012). Mesial temporal lobe epilepsy. J. Clin. Neurophysiol. 29, 356–365. doi: 10.1097/WNP.0b013e31826b3ab7
- Tort, A. B. L., Kramer, M. A., Thorn, C., Gibson, D. J., Kubota, Y., Graybiel, A. M., et al. (2008). Dynamic cross-frequency couplings of local field potential oscillations in rat striatum and hippocampus during performance of a T-maze task. *Proc. Natl. Acad. Sci. U.S.A.* 105, 20517–20522. doi: 10.1073/pnas.0810524105
- Tort, A. B., Rotstein, H. G., Dugladze, T., Gloveli, T., and Kopell, N. J. (2007). On the formation of gamma-coherent cell assemblies by oriens lacunosummoleculare interneurons in the hippocampus. *Proc. Natl. Acad. Sci. U.S.A.* 104, 13490–13495. doi: 10.1073/pnas.0705708104
- Traub, R., and Bibbig, A. (2000). A model of high-frequency ripples in the hippocampus based on synaptic coupling plus axon-axon gap junctions between pyramidal neurons. *J. Neurosci.* 20, 2086–2093.
- Van, D. W., Lee, H. C., Hereld, M., Chen, Z., Elsen, F. P., and Stevens, R. L. (2005). Emergent epileptiform activity in neural networks

with weak excitatory synapses. IEEE T. Neur. Sys. Reh. 13, 236–241. doi: 10.1109/TNSRE.2005.847387

- Wang, X. J., and Buzsáki, G. (1996). Gamma oscillation by synaptic inhibition in a hippocampal interneuronal network model. J. Neurosci. 16, 6042–6413.
- Wendling, F., Bartolomei, F., Bellanger, J. J., and Chauvel, P. (2002). Epileptic fast activity can be explained by a model of impaired GABAergic dendritic inhibition. *Eur. J. Neurosci.* 15, 1499–1508. doi: 10.1046/j.1460-9568.2002.01985.x
- Wendling, F., Bartolomei, F., Mina, F., Huneau, C., and Benquet, P. (2012). Interictal spikes, fast ripples and seizures in partial epilepsies – combining multi-level computational models with experimental data. *Eur. J. Neurosci.* 36, 2164–2177. doi: 10.1111/j.1460-9568.2012.08039.x
- Wendling, F., Benquet, P., Bartolomei, F., and Jirsa, V. (2016). Computational models of epileptiform activity. J. Neurosci. Methods 260, 233–251. doi: 10.1016/j.jneumeth.2015.03.027
- Ye, G. L., Yi, S., Gamkrelidze, G., Pasternak, J. F., and Trommer, B. L. (2005). AMPA and NMDA receptor-mediated currents in developing dentate gyrus granule cells. *Brain Res. Dev. Brain Res.* 155, 26–32. doi: 10.1016/j.devbrainres.2004.12.002
- Yuen, G. L., and Durand, D. (1991). Reconstruction of hippocampal granule cell electrophysiology by computer simulation. *Neuroscience* 41, 411–423. doi: 10.1016/0306-4522(91)90337-N

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Zhang, Fan and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.