



# Adenosine A<sub>1</sub>-A<sub>2A</sub> Receptor Heteromer as a Possible Target for Early-Onset Parkinson's Disease

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**Keywords:** early-onset Parkinson's disease, adenosine A<sub>1</sub> receptor, oligomer

## OPEN ACCESS

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### Specialty section:

This article was submitted to  
Neurodegeneration,  
a section of the journal  
Frontiers in Neuroscience

**Received:** 04 October 2017

**Accepted:** 09 November 2017

**Published:** 22 November 2017

### Citation:

Fernández-Dueñas V,  
Pérez-Arévalo A, Altafaj X, Ferré S and  
Ciruela F (2017) Adenosine A<sub>1</sub>-A<sub>2A</sub>  
Receptor Heteromer as a Possible  
Target for Early-Onset Parkinson's  
Disease. *Front. Neurosci.* 11:652.  
doi: 10.3389/fnins.2017.00652

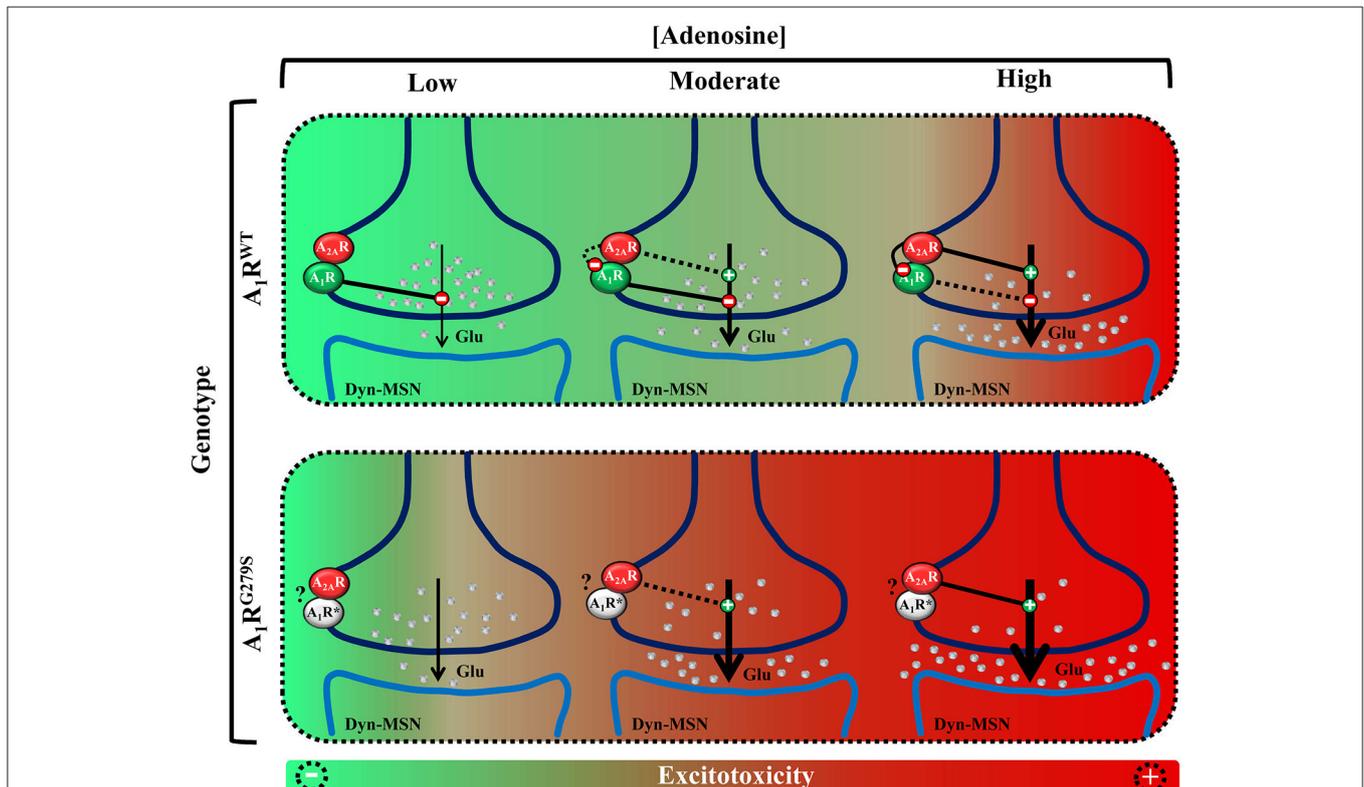
Parkinson's disease (PD) is a progressive, neurodegenerative disorder that affects ~1% of individuals over the age of 60, which turns to 5% in subjects up to 85 years (de Lau and Breteler, 2006). On the other hand, a form of PD, called early-onset PD (EOPD), arises at an earlier age (<45; Bonifati et al., 2005; Ylikotila et al., 2015). EOPD patients generally display a slower progression of the disease and present a better response to dopaminergic treatments; however, they may finally develop a full PD symptomatology (i.e., bradykinesia, resting tremor, muscular rigidity and postural instability, drug-induced dyskinesia; Olgiati et al., 2016). The etiology of both PD and EOPD is still not completely elucidated. Thus, although genetic studies have provided some information about the main genes involved, epidemiological data showed that behavioral and environmental factors play a key role in the pathogenesis and progression of PD (Puschmann, 2013; Ascherio and Schwarzschild, 2016). Importantly, the contribution of genetic causes in EOPD has been extensively studied. For instance, mutations in PD-associated genes, such as *PRKN* (*PARK2*; MIM number 600116), *PINK1* (*PARK5*; MIM number 605909), and *DJ-1* (*PARK7*; MIM number 602533), have often been associated to autosomal-recessive forms of EOPD (Lücking et al., 2000; Bonifati et al., 2005; Olgiati et al., 2016). Recently, an Iranian research group described a new autosomal-recessive mutation in two siblings (30 and 34 years old) with consanguine parents, which was associated to EOPD (Jaberi et al., 2016). Interestingly, while both brothers did not present alterations in the main PD-related genes (i.e., *PRKN*, *PINK1*, and *DJ-1*), a homozygous missense mutation (c.835G > A) in the adenosine A<sub>1</sub> receptor (A<sub>1</sub>R) gene (*ADORA1*) was found (Jaberi et al., 2016). This nucleotide point mutation in *ADORA1* involves the substitution of a highly conserved amino acid (p.Gly279Ser) within the transmembrane 7 (TM7) domain, but the functional consequences remain unknown. In contrast, it was recently determined that mutations affecting *ADORA1* gene and more particularly the missense matution *ADORA1* (p.G279S), are not a common risk factor for PD in the European population, arguing against *ADORA1* as a candidate gene in PD (Blauwendraat et al., 2017). Altogether, these opposing data indicate that additional work must be done toward the elucidation of the potential contribution of *ADORA1* mutations in PD pathogenesis, and the contribution of genetic and environmental factors.

A<sub>1</sub>R has a widespread distribution in the brain, with the highest levels detected in the cortex, hippocampus, and cerebellum (Sebastião and Ribeiro, 2009). In addition, A<sub>1</sub>R is markedly expressed in the basal ganglia. Thus, A<sub>1</sub>R can be found in the major striatal neuronal population, the GABAergic medium-sized spiny neurons (MSNs; Ferré et al., 1996), together with the expression in the cortico-thalamic glutamatergic afferent fibers. These fibers, together with the dopaminergic projections from the substantia nigra pars compacta control the striatal circuitry that are critical in the control of the motor function (Sebastião and Ribeiro, 2009). The selective

death of dopaminergic fibers is the primary cause and a hallmark of PD; however, the dysregulation of cortico-thalamic glutamatergic signaling is also involved in the progression of the disease (Fredholm et al., 2005; Gomes et al., 2011). Under physiological conditions, GABAergic MSNs are continuously activated by cortico-thalamic glutamatergic terminals, but a complex array of presynaptic receptors, which include  $A_1R$ , adenosine  $A_{2A}$  receptor ( $A_{2A}R$ ), cannabinoid  $CB_1$  receptor ( $CB_1R$ ) and dopamine  $D_2$  and  $D_4$  receptors ( $D_2R$  and  $D_4R$ , respectively) modulate this tonic stimulation (Ciruela et al., 2006a; González et al., 2012; Mathur and Lovinger, 2012; Ferreira et al., 2015; Bonaventura et al., 2017). Potentially, the dysregulation of these presynaptic modulatory receptors can lead to abnormal glutamate release in the synaptic cleft, which may over activate postsynaptic glutamate receptors, trigger excitotoxicity and, ultimately, lead to neurodegenerative processes affecting brain circuits involved in the control of motor function (Gomes et al., 2011).

Interestingly,  $A_1R$  colocalizes and interacts with  $A_{2A}R$  at the presynaptic membrane of cortico-thalamic glutamatergic terminals, forming functional receptor heteromers in the striatum (Figure 1; Ciruela et al., 2006a). Importantly, the

striatal  $A_1R/A_{2A}R$  heteromer plays a pivotal role controlling glutamate release, thus acting as an adenosine concentration-dependent switch (Ciruela et al., 2006b; Figure 1). Hence, low to moderate extracellular adenosine concentrations (homeostatic basal levels) mostly stimulate  $A_1R$ , since it displays higher affinity for adenosine compared to  $A_{2A}R$ , and a net inhibition of glutamate release is achieved (Figure 1). Conversely, moderate to high concentrations of striatal adenosine, which should theoretically trigger, in theory, both  $A_1R$  and  $A_{2A}R$  activation, ultimately lead to a predominant  $A_{2A}R$  activation. In such way,  $A_{2A}R$  may block heteromeric  $A_1R$  through a receptor-receptor allosteric trans-inhibition, thus leading to a predominant facilitation of glutamate release (Figure 1; Ciruela et al., 2006b). At this point, the question consists of whether the *ADORA1* (p.G279S) mutation abolishes  $A_1R$  function and whether this alteration depends on its heteromerization with  $A_{2A}R$  receptor, specifically disrupting the function of the adenosine concentration-dependent switch. In the absence of experimental data, we can speculate that the mutation can be affecting the  $A_1R/A_{2A}R$  heteromer, resulting in a potential alteration of the fine-tuning modulation of striatal glutamatergic neurotransmission. Indeed, in such scenario,



**FIGURE 1** | Schematic representation of the potential impact of  $A_1R$  mutation in the fine-tuning modulation of striatal glutamatergic neurotransmission. **(Up)** Model of glutamate release control by the  $A_1R/A_{2A}R$  heteromer adenosine concentration-dependent switch. Low to moderate concentrations of adenosine activate predominantly  $A_1R$ , inhibiting glutamate release. Moderate to high concentrations of adenosine also activate  $A_{2A}R$  which, by means of the  $A_1R$ - $A_{2A}R$  intramembrane interaction, antagonizes  $A_1R$  function, therefore facilitating glutamate release. **(Bottom)** Model of *ADORA1*(p.G279S) mutation pathogenic impact ( $A_1R^{G279S}$  or  $A_1R^*$ ) in the striatal glutamatergic neurotransmission. The proposed  $A_1R$  mutant loss-of-function would implicate a dysregulation of the adenosinergic presynaptic control of striatal glutamate release, which may ultimately lead to a higher risk of inducing excitotoxicity and neurodegeneration.

we can hypothesize that moderate concentrations of striatal adenosine would facilitate glutamate release and reduce the excitotoxicity threshold (**Figure 1**).

The dysregulation of this presynaptic module may lead to uncontrolled glutamate release which, in addition, might be potentiated by low dopamine innervation, which would not act upon inhibitory presynaptic D<sub>2</sub>R and D<sub>4</sub>R. Consequently, managing the disturbance of the adenosine switch mechanism regulating glutamatergic striatal innervation (caused either by a direct *ADORA1* mutation or mutations affecting A<sub>1</sub>R/A<sub>2A</sub>R heteromers function), may help to restore the normal functioning of the basal ganglia. In this sense, the A<sub>1</sub>R/A<sub>2A</sub>R heteromer could be considered as a potential therapeutic target for EOPD. Alternatively, the glutamatergic component of these forms of EOPD would represent an initial or master pathogenic event to dopamine denervation, as proposed in Huntington's disease pathophysiology (Gomes et al., 2011). In such way, the predominant role of an aberrant glutamatergic signaling could explain at the molecular level the high effectiveness of PD dopamine-based therapies, either in terms of higher or long-lasting efficacy. Nevertheless, in order to restore physiological neurotransmission it would be necessary to focus not exclusively on dopamine availability, but also in the control of glutamate release which is partially modulated by the A<sub>1</sub>R/A<sub>2A</sub>R oligomer (i.e., A<sub>1</sub>R activation and A<sub>2A</sub>R inhibition). In this sense, the use of A<sub>2A</sub>R antagonists has been assessed for the treatment of PD (Vallano et al., 2011). Regarding the potential use of A<sub>1</sub>R-based therapies, there is a major hurdle related to the A<sub>1</sub>R ubiquitous

expression pattern that might lead to deleterious side-effects. In order to bypass these limitations, novel approaches based on (i) local A<sub>1</sub>R activation or (ii) pharmacological increase of the adenosine tone (below the threshold of A<sub>2A</sub>R activation) using adenosine transporters blockers and/or metabolizing enzymes, are expected to reach an effective treatment for EOPD.

Overall, the discovery of a novel mutation in *ADORA1* presumably leading to EOPD supports the potential beneficial use of a multimodal approach for the pharmacological treatment of this neurodegenerative condition. This approach, based on the combination of pharmacological therapies (i.e., dopaminergic compounds and drugs targeting the A<sub>1</sub>R/A<sub>2A</sub>R oligomer) could be potentially extended to all forms of PD.

## AUTHOR CONTRIBUTIONS

VF-D, XA, SF: wrote the paper; AP-A: conceived the idea; FC: conceived the idea and wrote the paper.

## ACKNOWLEDGMENTS

This work was supported by MINECO/ISCIII (SAF2014-55700-P, PIE14/00034, and PS16/00851), IWT (SBO-140028), and Fundació la Marató de TV3 (Grant 20152031 and Grant 20140210). FC, XA, AP-A, and VF-D belong to the "Neuropharmacology and Pain" accredited research group (Generalitat de Catalunya, 2014 SGR 1251), and by the intramural funds of the National Institute on Drug Abuse to SF.

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**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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