



Mechanisms Underlying Overactive Bladder and Interstitial Cystitis/Painful Bladder Syndrome

Luke Grundy^{1,2*}, Ashlee Caldwell^{1,2} and Stuart M. Brierley^{1,2*}

¹ Visceral Pain Research Group, Centre for Neuroscience, College of Medicine and Public Health, Flinders University, Adelaide, SA, Australia, ² Centre for Nutrition and Gastrointestinal Diseases, Discipline of Medicine, South Australian Health and Medical Research Institute, The University of Adelaide, Adelaide, SA, Australia

The bladder is innervated by extrinsic afferents that project into the dorsal horn of the spinal cord, providing sensory input to the micturition centers within the central nervous system. Under normal conditions, the continuous activation of these neurons during bladder distension goes mostly unnoticed. However, for patients with chronic urological disorders such as overactive bladder syndrome (OAB) and interstitial cystitis/painful bladder syndrome (IC/PBS), exaggerated bladder sensation and altered bladder function are common debilitating symptoms. Whilst considered to be separate pathological entities, there is now significant clinical and pre-clinical evidence that both OAB and IC/PBS are related to structural, synaptic, or intrinsic changes in the complex signaling pathways that mediate bladder sensation. This review discusses how urothelial dysfunction, bladder permeability, inflammation, and cross-organ sensitisation between visceral organs can regulate this neuroplasticity. Furthermore, we discuss how the emotional affective component of pain processing, involving dysregulation of the HPA axis and maladaptation to stress, anxiety and depression, can exacerbate aberrant bladder sensation and urological dysfunction. This review reveals the complex nature of urological disorders, highlighting numerous interconnected mechanisms in their pathogenesis. To find appropriate therapeutic treatments for these disorders, it is first essential to understand the mechanisms responsible, incorporating research from every level of the sensory pathway, from bladder to brain.

OPEN ACCESS

Edited by:

Janet R. Keast, The University of Melbourne, Australia

Reviewed by:

Jennifer DeBerry, The University of Alabama at Birmingham, United States Wouter Everaerts, KU Leuven, Belgium

*Correspondence:

Luke Grundy luke.grundy@flinders.edu.au; luke.grundy@adelaide.edu.au Stuart M. Brierley stuart.brierley@flinders.edu.au

Specialty section:

This article was submitted to Autonomic Neuroscience, a section of the journal Frontiers in Neuroscience

Received: 10 September 2018 Accepted: 27 November 2018 Published: 12 December 2018

Citation:

Grundy L, Caldwell A and Brierley SM (2018) Mechanisms Underlying Overactive Bladder and Interstitial Cystitis/Painful Bladder Syndrome. Front. Neurosci. 12:931. doi: 10.3389/fnins.2018.00931 Keywords: bladder, overactive bladder, interstitial cystitis, afferent, peripheral, central, sensitisation

INTRODUCTION

Overactive bladder syndrome (OAB) and interstitial cystitis/painful bladder syndrome (IC/PBS) are common, chronic, pelvic disorders affecting approximately ~16% of the western population (Hanno, 2002; Irwin et al., 2006; McLennan, 2014; Truzzi et al., 2016). Urgency, frequency, and nocturia are common symptoms of both OAB and IC. However, these conditions may be differentiated by the presence of urge urinary incontinence in patients with OAB and pelvic pain in IC patients (MacDiarmid and Sand, 2007; Hanno and Dmochowski, 2009; Haylen et al., 2010; Homma et al., 2016). As both of these disorders are diagnosed in the absence of bacterial infection or obvious pathology, the etiology of OAB and IC/PBS symptoms remain unknown. Accordingly, efficacious therapeutic options are limited, contributing to the significant societal and economic

1

impact of greater than \$70 billion per annum in the United States (Pierce and Christianson, 2015; Durden et al., 2018).

Normal bladder function requires coordination of afferent signals originating from the bladder wall with excitatory and inhibitory signals from the anterior cingulate cortex (ACC), insula, and hypothalamus to provide an overview of the appropriateness to urinate that is ultimately under conscious control by the prefrontal cortex (Figure 1; Griffiths, 2015; Lovick, 2016). Bladder afferents embedded within the detrusor smooth muscle show exquisite sensitivity for mechanical distension but are also found innervating the urothelium (Zagorodnyuk et al., 2006, 2007, 2009; Spencer et al., 2018). This topology provides a secondary level of resolution to the transmission of sensory stimuli, including the detection of bladder infection, urothelial inflammation, or barrier breakdown (Figure 1). Accordingly, bladder sensory afferents express a range of anti- and pronociceptive receptors and ion channels (Erickson et al., 2018; Grundy et al., 2018c) that integrate the input from this complex signaling environment and can induce a range of sensations from fullness through to pain (Fowler et al., 2008). These afferents, whose cell bodies are located within the dorsal root ganglia (DRG), project via the pelvic, hypogastric/splanchnic nerves, synapse within the dorsal horn of the lumbosacral (LS, L5-S1) and thoracolumbar (TL, T10-L2) spinal cord (Figure 1; Fowler et al., 2008; de Groat and Yoshimura, 2015) and terminate within the periaqueductal gray (PAG) (Fowler et al., 2008). The PAG acts as an integration center for afferent signals from the spinal cord and higher brain centers (Griffiths, 2015). A conscious "urge" to urinate is perceived when afferent activity increases beyond a pre-set threshold and, if modulating input from the brain permits, the PAG activates the pontine micturition centre (PMC) to induce efficient voiding (Andersson and Arner, 2004; Fowler et al., 2008; Griffiths, 2015; Lovick, 2016).

Clinical studies reveal that patients with OAB and IC/PBS perceive sensations of bladder fullness, urge to void, and pain at lower cystometric volumes than healthy subjects (Van Brummen et al., 2004; Kim et al., 2009; Parsons and Drake, 2011). These observations thereby implicate the sensitisation of bladder afferent pathways to physiological stimuli as a key component in the mechanisms underlying these disorders (Yamaguchi et al., 2007; Yoshimura et al., 2014; de Groat and Yoshimura, 2015). Furthermore, co-morbidity of depression and anxiety is significantly higher in patients with OAB and IC/PBS, while high psychological stress levels is strongly correlated to exacerbated bladder symptoms (Goldstein et al., 2008; Lai et al., 2015; Golabek et al., 2016; Leue et al., 2017; McKernan et al., 2017). As such, increased intensity of the afferent signal or modulation of the emotional affective state can have profound effects on bladder sensation. A key concept therefore, in the development of OAB and IC/PBS, is chronic sensitisation of the neuronal networks regulating bladder sensation, incorporating structural, synaptic, or intrinsic changes of peripheral or central structures that may drive subsequent changes in downstream sensory pathways (Brierley and Linden, 2014; Grundy et al., 2018c).

In this review, for brevity, we summarize both preclinical and clinical research to highlight how alterations in peripheral

afferent excitability contribute to the symptoms of OAB and IC/PBS to provide insights into the mechanisms that are hypothesized to mediate these distinct disorders which have many overlapping symptoms. For an excellent review on the potential changes in central processes occurring in OAB see Reynolds et al. (2016).

UROTHELIAL PERMEABILITY

Toxic waste metabolites excreted in urine are prevented from accessing the bladder interstitium and embedded afferent endings by a tight urothelial barrier (Spencer et al., 2018; Figure 1). The urothelial barrier is maintained by tight junctions between apical urothelial cells, hydrophobic uroplakin plaques, and a considerable glycosaminoglycan (GAG) mucus layer, that block the movement of small molecules and urine (Birder and Andersson, 2013; Hurst et al., 2015). From a clinical point of view, numerous studies have identified that patients with IC/PBS, but not OAB, have a diminished or damaged urothelium (Elbadawi and Light, 1996; Tomaszewski et al., 2001; Lai et al., 2013; Keay et al., 2014; Hurst et al., 2015), as well as reduced expression of the tight junction proteins zona occludens 1 (ZO-1) and E-cadherin (Liu et al., 2012). Increased bladder permeability due to reduced urothelial integrity is thought to underlie the sensitivity of IC/PBS patients (75%) to the potassium sensitivity test (PST) compared to control patients (4%) (Parsons et al., 1998). A number of clinical studies also show that OAB patients respond to the PST, however, this may be due to overlap and misdiagnosis between IC/PBS and OAB (Minaglia et al., 2005; MacDiarmid and Sand, 2007; Chung et al., 2010). Despite these observations, it remains to be determined if bladder permeability is part of the underlying pathology of bladder hypersensitivity in IC/PBS patients or a downstream consequence of localized inflammation that further exacerbates the condition.

Glycosaminoglycan replacement therapy with pentosane polysulfate (PPS) has been shown to improve the symptoms for some, but not all, IC/PBS patients (Nickel et al., 2012; Lai et al., 2013; Oliveira et al., 2014; Hurst et al., 2015). However, PPS also induces broad anti-inflammatory actions, including the inhibition of mast cell histamine release in the bladder (Chiang et al., 2000; Anderson and Perry, 2006; Wu et al., 2011; Sanden et al., 2017). In addition, pre-clinical studies inducing urothelial permeability with protamine sulfate identified a relatively rapid recovery of the urothelial barrier and structure (Lavelle et al., 2002; Greenwood-Van Meerveld et al., 2015), via injury-induced proliferation of basal urothelial and stromal cells (Shin et al., 2011). A more likely scenario therefore is that bladder permeability in IC/PBS patients is secondary to localized inflammation. Continuous access of toxic urine contents combined with inflammatory mediators, such as cytokines, histamine, and proteases, can sensitize peripheral afferent endings and have the potential to trigger long term changes in neuronal function and neuroplasticity within the entire afferent network (Brierley and Linden, 2014; Grundy et al., 2018c).



Breakdown of the urothelial barrier allows access of toxic urine contents into the underlying bladder interstitium which can activate bladder afferents. Bladder afferents project via the pelvic nerve or the splanchnic nerve to the dorsal horn of the thoracolumbar and lumbosacral spinal cord, where they activate second order neurons within the spinal cord synapse in the thalamus or the PAG of the midbrain. Thalamic projections provide input into limbic and cortical structures to provide the emotional affective and conscious component of the voiding reflex pathway. The thalamus relays to the PAG and the PAG feeds into the PMC to signal micturition. TL, Thoracolumbar; LS, lumbosacral; SC, spinal cord; PFC, Prefrontal Cortex; ACc, Anterior cingulate cortex; Hyp, hypothalamus; HGN, hypogastric nerve; PAG, periaqueductal gray; PMC, primary micturition center; DRG, dorsal root ganglion; DH, Dorsal horn; Agd, amygdala; LSN, lumbar splanchnic nerve; SPN, pelvic nerve; IMG, inferior mesenteric ganglion; PP, Pelvic Plexus. Figure modified from Grundy et al. (2018c).

Altered Release of Urothelial Factors

In addition to its role as a physical barrier, the urothelium provides bi-directional communication with underlying primary afferents (Lazzeri, 2006; Birder and Andersson, 2013; Merrill et al., 2016) via the detection and/or release of a range of excitatory and inhibitory neurotransmitters and neuromodulators including ATP, acetylcholine, nitric oxide (NO), NGF, prostaglandin E2 (PGE2), neurokinin A, and inflammatory mediators as described above (and extensively reviewed by Birder et al.) (Everaerts et al., 2010a,b; Birder and Andersson, 2013; de Groat and Yoshimura, 2015; Grundy et al., 2018a). Altered urothelial mediator release has been identified from OAB and IC/PBS patients in a number of studies and may be a compounding mechanism in the development of chronic neuronal hypersensitivity (Kim et al., 2005, 2006; Sun and Chai, 2006; Suh et al., 2017).

Chronic bladder inflammation enhances ATP release from the urothelium and augments purinergic signaling of bladder afferents in rats (Smith et al., 2005), whilst both stretch-mediated ATP release and its receptor P2X3 are increased in the urothelium of OAB and IC/PBS patients (Sun and Chai, 2004, 2006; Contreras-Sanz et al., 2016; Jhang and Kuo, 2016). Preclinically, acetylcholine, acting via muscarinic receptors also triggers release of ATP, as well as NO and prostanoids from urothelial cells (Winder et al., 2014; Michel, 2015). In animal studies, experimentally induced cystitis upregulates muscarinic receptors and acetylcholine release, as well as subsequent cholinergic regulated urothelial NO release (Giglio et al., 2005; Giglio and Tobin, 2009; Andersson, 2011; McDermott et al., 2013). Notably, PGE2 levels are significantly increased in OAB patients (Kim et al., 2005, 2006), whilst NO and PGE2 levels and receptor expression are also altered in IC/PBS patients with Hunner's legions (Wada et al., 2015; Jhang et al., 2016). Bradykinin stimulates urothelial NGF release and enhances stretch-induced ATP release in a human urothelial cell line (Ochodnicky et al., 2013; Winder et al., 2014), whilst bradykinin 1 (B1) receptor expression is upregulated in human IC/PBS patient bladder samples (Arms and Vizzard, 2011) and in the urothelium of a rat CYP-induced cystitis model (Chopra et al., 2005).

Limited causations have been determined for altered urothelial neurotransmitter release or receptor expression in OAB and IC/PBS, but there is accumulating evidence that they are a downstream consequence of inflammation, infection or urothelial breakdown.

INFLAMMATION

By definition, the presence of bladder inflammation precludes the clinical diagnosis of OAB. Only a small population of IC/PBS patients exhibit significant inflammation, which is characterized by the presence of Hunner's ulcers (Leiby et al., 2007). However, it is widely reported that there are increases in the amount of pro-inflammatory mediators within the bladder and urine of both OAB and IC/PBS patients (Kastrup et al., 1983; El-Mansoury et al., 1994; Hauser et al., 2008; Jacobs et al., 2010; Liu et al., 2012; Jhang and Kuo, 2016; Furuta et al., 2018). These pro-inflammatory mediators including histamine, nerve growth factor (NGF), and those released from mast cells, are known to directly sensitize afferent nerve terminals (Davidson et al., 2014). Furthermore, overexpression of pro-inflammatory genes, oedema, tissue granulation and an increase in macrophages, chemokines, cytokines, eosinophils, as well as T and B cell markers have also been identified in IC/PBS patients (Hauser et al., 2008; Abernethy et al., 2017). As the majority of cystitisinduced inflammation is localized to the superficial mucosa, urothelial afferents are ideally placed to detect and respond to these environmental changes.

Inflammation-induced sensitisation of afferents is an essential mechanism for the induction of normal wound healing, however, chronic sensitisation of afferents can occur during prolonged inflammation or following a severe bout of inflammation (Brierley and Linden, 2014; Abraham and Miao, 2015). In support of an inflammatory-mediated pathophysiology in urological disorders, animal models of cystitis have employed a range of chemicals, including acetic acid, acrolein, cyclophosphamide (CYP), zymosan and lipopolysaccharide, that induce both acute and longer lasting bladder hyperactivity (Takezawa et al., 2014; Liu and Dong, 2015; Abdi et al., 2016; Hughes et al., 2016). Whilst there are many limitations to the use of inflammatory animal models in the study of symptom defined disorders such as OAB and IC/PBS (comprehensively discussed by Fry et al., 2010), these animals show an overactive bladder phenotype, with altered cystometry and enhanced visceromotor response during bladder distension, replicating the reduced bladder capacity, plus the allodynia and hyperalgesia to bladder distension observed in humans (Fry et al., 2010; Lai et al., 2011; DeBerry et al., 2014, 2015b). Furthermore, bladder afferents show direct sensitisation to chemical and inflammatory stimuli (de Groat and Yoshimura, 2009), whilst retrogradely traced bladder-innervating DRG neurons from CYP-treated rats or a naturally occurring feline interstitial cystitis model exhibit lower activation thresholds and sensitisation to current injection (Dang et al., 2008; Buffington, 2011). The efficacy of intraluminal therapies to treat OAB and

IC/PBS (Cvach and Rosamilia, 2015; Manriquez et al., 2015), that either (1) block bladder afferent firing (such as lidocaine and neosaxitoxin) or (2) cause peripheral nerve desensitization (with agents such as resiniferatoxin; RTX), highlight the important role of peripheral afferents in mediating bladder hypersensitivity to distension (Apostolidis et al., 2005).

The transient receptor potential (TRP) channel TRPV1 is upregulated in the bladders of patients with OAB and IC/PBS (Liu and Kuo, 2007), whilst both TRPV1 and TRPA1 have consistently been implicated in mediating normal and cystitis-induced mechanical sensitivity in rodents by modulating neuronal activation thresholds and enhancing bladder afferent responses to P2X receptor activation (Daly et al., 2007; Wang et al., 2008; DeBerry et al., 2014, 2015b; Yoshiyama et al., 2015; Grundy et al., 2018b). A host of additional receptors and channels associated with nociception have also been identified upon bladder afferents that regulate neuronal sensitivity and neuronal excitability in animal models of cystitis, including voltage gated sodium (Nav) channels (Erickson et al., 2018; Grundy et al., 2018d), potassium channels (K_V) (Hayashi et al., 2009), P2X receptors (Dang et al., 2008; Chen and Gebhart, 2010), TRPV4 (Merrill et al., 2012) and cannabinoid receptors (Hedlund, 2014; Izzo et al., 2015; Bakali et al., 2016; Hedlund and Gratzke, 2016; Munoz, 2016).

It is possible that a population of patients present without active inflammation or increased bladder permeability but are in fact in remission from a preceding bladder infection or inflammation. Such a scenario could induce a protracted hypersensitive state and correspond to their enhanced sensory symptoms. Indeed, women with a clinical history of recurrent UTI as children are significantly more likely to have a diagnosis of IC/PBS as adults (Peters et al., 2009), and preclinical investigations of neonatal bladder insult in rats suggests this may be due to long term sensitisation of sensory pathways (Randich et al., 2006; DeBerry et al., 2007; Ness and Randich, 2010). Neuroplasticity of peripheral afferent circuitry following the resolution of inflammation or recovery from tissue injury has been well documented in both somatic and visceral pain models through the induction of neurogenic inflammation and neuronal sprouting (de Groat and Yoshimura, 2009; Gregory et al., 2013; Brierley and Linden, 2014). Neonatal bladder inflammation in rats results in hypersensitive responses to inflammatory stimuli as an adult, inducing an overactive bladder phenotype (Randich et al., 2006; DeBerry et al., 2007, 2010), as well as enhanced spontaneous and urinary bladder distension-evoked activity of spinal visceral nociceptive neurons (Ness and Randich, 2010). Alterations in spinal cord circuits responsible for bladder sensation may regulate this phenomenon, as neonatal inflammation induces a downregulation of GABA (Aa-1) receptor microRNA and altered opioid peptide content in the dorsal horn (Sengupta et al., 2013; Shaffer et al., 2013). Furthermore, neonatal zymosan enhances bladder neuropeptide content of CGRP and Substance P compared to sham controls (DeBerry et al., 2010; Shaffer et al., 2011).

Similarly, in pre-clinical models of adult cystitis, bladder overactivity is associated with increases in tyrosine receptor kinase (Trk) A, Trk B, and calcitonin gene-related peptide (CGRP) (Vizzard, 2001; Qiao and Vizzard, 2002), which in turn promote inflammation in the tissue where the afferent terminals reside (Rosa and Fantozzi, 2013). Furthermore, patients with IC/PBS have higher elevated serum and urinary NGF levels than healthy controls (Chen et al., 2016). NGF overexpression in mouse urothelium leads to neuronal hyper-innervation, increased mast cell counts and changes in bladder function (Schnegelsberg et al., 2010). These discoveries may explain the increased sprouting of neuronal terminals identified in the bladders of IC/PBS patients (Christmas et al., 1990; Lundeberg et al., 1993), that has been replicated in rodent models of inflammation *in vivo* and *in vitro* (Dupont et al., 2001; Schnegelsberg et al., 2010; Boudes et al., 2013; Ekman et al., 2017).

MICROBIOME/CHRONIC URINARY TRACT INFECTION

Following the recent identification of a bladder-specific microbiome, and the role of the gut microbiome in chronic functional gastrointestinal diseases (Hughes et al., 2013; Harper et al., 2018), a link between the balance of bacteria in the bladder and the symptoms of OAB and IC/PBS has been postulated and explored (Contreras-Sanz et al., 2016; Angelini, 2017; Drake et al., 2017). Moreover, the traditional colony forming unit thresholds for confirming urinary tract infection (UTI) in clinical practice have been questioned, and a role for chronic UTI in the pathogenesis of OAB has been investigated (Balachandran et al., 2016).

Patients with OAB may have genuine uropathogenic infections, and are therefore misdiagnosed, as large numbers of bacteria are undetected by routine mid-stream urine cultures (Khasriya et al., 2013). Indeed, a significantly greater number of patients with refractory idiopathic detrusor overactivity show low count bacteriuria vs. controls (Walsh et al., 2011). Undiagnosed intracellular bacterial colonization of urothelial cells may also occur in OAB (Scott et al., 2015), as OAB patients exhibit significantly greater infected urothelial cell counts and microscopic pyuria than healthy subjects, which also correlates to urgency symptoms (Gill et al., 2018). Uropathic E. coli infection initiates the release of multiple mediators from the urothelium, including cytokines and interleukins, as well as promoting urothelial barrier defects (Wood et al., 2012), which alert the immune system to impending damage and initiate an immune response (Abraham and Miao, 2015). Immune cell infiltration and the release of pro-inflammatory cytokines are known to sensitize peripheral afferents (Ren and Dubner, 2010), and in this way enhance bladder sensation. In support of these considerations, a recent pilot study revealed that combination antibiotic treatment of both Gram-negative and Gram-positive bacteria significantly improved OAB symptoms as well as the perception of their bladder condition (Vijaya et al., 2013). Furthermore, shifts in the bacterial species that constitute the bladder microbiome have been associated with both the presence and severity of OAB and IC/PBS (Siddiqui et al., 2012; Whiteside et al., 2015; Contreras-Sanz et al., 2016; Lakeman and Roovers, 2016; Curtiss et al., 2017).

For example, women with IC/PBS, but not OAB, have a less diverse microbiota than those without (Hilt et al., 2014; Pearce et al., 2014; Abernethy et al., 2017; Curtiss et al., 2017). Interestingly, despite significant inter-patient variability in bladder microbiome, a decrease in Lactobacillus, which has antimicrobial properties, in both OAB and IC/PBS patients compared to controls is a common finding (Hilt et al., 2014; Pearce et al., 2014; Curtiss et al., 2017). Furthermore, the absence of Lactobacillus acidophilus correlates with higher pain scores and higher scores on the interstitial cystitis symptom index (Abernethy et al., 2017). Proteus, the urinary pathogen, is also identified more commonly in patients with OAB and lower urinary tract symptoms than healthy controls (Khasriya et al., 2013; Curtiss et al., 2017). These data support a line of communication between the urinary microenvironment and underlying afferent nerves that is likely mediated by the urothelium.

Cross-Organ Sensitisation

Considerable clinical evidence suggests that diseases of the colon, such as irritable bowel syndrome (IBS) and inflammatory bowel disease (IBD), can induce subsequent development of pathology in an otherwise unaffected adjacent organ, such as the bladder (Grundy et al., 2018d).

A mouse model of colitis induced by intra-rectal instillation of 2,4,6-trinitrobenzene sulfonic acid (TNBS) induces hyperexcitability of the entire peripheral sensory pathway, from the afferent ending in the colon to the spinal cord (Brierley and Linden, 2014). Importantly, TNBS colitis also prompts consistent changes in bladder voiding parameters that replicate the clinical symptoms of urgency and frequency, as well as increased bladder-afferent sensitivity to bladder distention (Brumovsky and Gebhart, 2010; Ustinova et al., 2010; Greenwood-Van Meerveld et al., 2015; Yoshikawa et al., 2015). These symptoms occur in the absence of any overt inflammation or histological damage to the bladder, highlighting the importance of altered afferent sensitivity in maintaining OAB and IC/PBS symptomology (Yoshikawa et al., 2015).

This "cross-organ sensitisation" is considered to originate within the physiological co-ordination of these pelvic organs, and persistent pathological plasticity of their shared sensory pathways within the thoracolumbar and lumbosacral DRG and spinal cord (Persson et al., 2015; Grundy et al., 2018d). Approximately 15% of colonic innervating TL and LS DRG neurons exhibit dichotomising afferents, simultaneously innervating the bladder (Christianson et al., 2007; Yoshikawa et al., 2015), whilst a similar proportion of spinal dorsal horn neurons also respond to both urinary bladder and colonic distension (Grundy et al., 2018d). As such, sensitisation of colonic afferent pathways has the potential to directly influence the excitability of bladder afferent pathways. Indeed, a very recent pre-clinical study also indicates that chronic sensitisation of colonic afferent pathways results in the subsequent sensitisation of bladder afferent pathways and the triggering of uncontrolled urinary voiding in mice. Intriguingly, these changes in bladder function can be reversed by a therapeutic treatment targeted only to the colon (Grundy et al., 2018e).

Additionally, neurochemical changes occur within colonic afferent pathways following colitis that indicate the development of neurogenic inflammation (Grundy et al., 2018d). Multiple studies have shown that this translates to persistent upregulation of the neuromodulators NGF, BDNF, CGRP, and the high affinity receptor TrkB in bladder, bladder-innervating DRG neurons, and spinal cord (Liang et al., 2007; Qiao and Grider, 2007; Pan et al., 2010; Xia et al., 2015; Kawamorita et al., 2016). These receptors and neuropeptides share an intimate relationship enabling the positive feedback of each other (Malykhina et al., 2006; Christianson et al., 2007; Lei et al., 2013; Yoshikawa et al., 2015), consequently inducing neuronal sensitisation and neurite outgrowth, and likely contributing to cross-organ sensitisation through paracrine actions within the ganglia to increase bladder afferent excitability (Xia et al., 2015; Sorkin et al., 2018).

Cortical Regulation: Stress, Anxiety, and Depression

The sensory signals from bladder afferent converge in the PAG where they are modulated by input from the limbic system (amygdala, hypothalamus, thalamus, cingulate gyrus), insula, and prefrontal cortex (Fowler et al., 2008), which can in turn modulate or be modulated by the hypothalamic pituitary adrenal (HPA) axis. As such, changes to cortical networks or modulation of the emotional affective state can have profound effects on bladder sensation and may be an underlying mechanism in the development and persistence of OAB and IC/PBS symptoms.

Patients with OAB and IC/PBS report psychological stress levels that are significantly higher than healthy controls (Lai et al., 2015), which may be a consequence of HPA axis dysregulation following chronic early life stress (ELS) (Taylor, 2010). Exposure of children to ELS is a significant risk factor for developing HPA abnormalities (Anand, 1998; Pierce and Christianson, 2015), and shows strong correlations with the development of depression and anxiety in later life (Egeland, 2009; Heim and Binder, 2012). To this end, a number of studies report higher incidences of ELS and trauma in IC/PBS patients than healthy controls (Fuentes and Christianson, 2018), whilst clinical studies have demonstrated a strong correlation between stress, anxiety, depression, and the symptoms of bladder overactivity in patients with OAB and IC/PBS (Goldstein et al., 2008; Golabek et al., 2016; Leue et al., 2017). The increased prevalence of depression and anxiety in patients with IC/PBS occurs both following and prior to bladder symptoms, indicating a reciprocity in cause and effect, with no clear way to delineate patient cohorts to provide increased mechanistic understanding (McKernan et al., 2017). In addition to relationships with cognitive disorders, structural abnormalities within the white matter of the brain in women with IC/PBS, which facilitates the communication between and within brain regions, correlates closely to symptom severity (Farmer et al., 2015). It remains unclear whether these white matter properties are causes or consequences of IC/PBS. It is possible that certain white matter architecture may reflect a predisposition to develop disease, but it is equally plausible that these changes are a consequence of IC/PBS disease progression and future longitudinal studies are required to test this hypothesis.

The link between stress and bladder disorders is supported by pre-clinical studies in rodents that consistently induce bladder overactivity or mechanical hyperalgesia following stresstreatments (Black et al., 2009; Merrill et al., 2013; DeBerry et al., 2015a; Lee et al., 2015; Wang et al., 2017). In addition, this hypersensitivity has been found to be dependent on (DeBerry et al., 2015a), or correlate with, significant changes in brain regions associated with emotional processing and bladder control (Wang et al., 2017). Neonatal maternal separation in female mice, as a model of ELS, enhances visceromotor responses to urinary bladder distension accompanied by altered hippocampal input onto the HPA axis (Pierce et al., 2016). If sensitisation of peripheral afferent endings occurs in these models as a consequence of stress, or if sensitisation is arbitrated solely within the CNS has yet to be fully determined.

CONCLUSION

This review highlights the complex nature of both OAB and IC/PBS, incorporating evidence for changes in the dynamic signaling environment between the bladder lumen, urothelium, and afferent nerves, coordinating with adaptations to the HPA axis, and emotional affective components of sensory processing mediated within the limbic system. The bladder microbiome, bacterial infection, inflammation, and urothelial permeability contribute to the development of peripheral afferent hyperexcitability that is fundamental to the development of frequency and urgency in OAB, and pain in IC/PBS. In addition, the higher psychological stress levels, increased prevalence of anxiety and depression, as well as clinical co-morbidities with other visceral pain disorders suggests pathological plasticity within the CNS is an important component in the mechanisms underlying both OAB and IC/PBS. Determining the underlying mechanisms of bladder hypersensitivity is paramount to providing novel targets for the development of safer and more efficacious treatments.

AUTHOR CONTRIBUTIONS

AC and LG performed literature searches. AC, LG, and SB wrote the manuscript. All authors made significant contributions to the formation and correction of the manuscript in preparation for submission.

ACKNOWLEDGMENTS

The authors would like to thank the National Health and Medical Research Council of Australia (NHMRC) for their funding through a NHMRC R.D Wright Biomedical Research Fellow (APP1126378 to SB), NHMRC Australia Project Grants (#1083480, #1139366, and #1140297 to SB), and the Australian Research Council (ARC) Discovery Project (DP180101395 to SB).

REFERENCES

- Abdi, S. A., Najmi, A. K., and Raisuddin, S. (2016). Cyclophsopahmide-induced down-regulation of uroplakin II in the mouse urinary bladder epithelium is prevented by S-Allyl cysteine. *Basic Clin. Pharmacol. Toxicol.* 119, 598–603. doi: 10.1111/bcpt.12627
- Abernethy, M. G., Rosenfeld, A., White, J. R., Mueller, M. G., Lewicky-Gaupp, C., and Kenton, K. (2017). urinary microbiome and cytokine levels in women with interstitial cystitis. *Obstet. Gynecol.* 129, 500–506. doi: 10.1097/AOG. 000000000001892
- Abraham, S. N., and Miao, Y. (2015). The nature of immune responses to urinary tract infections. *Nat. Rev. Immunol.* 15, 655–663. doi: 10.1038/nri3887
- Anand, K. J. (1998). Clinical importance of pain and stress in preterm neonates. *Biol. Neonate* 73, 1–9. doi: 10.1159/000013953
- Anderson, V. R., and Perry, C. M. (2006). Pentosan polysulfate. Drugs 66, 821–835. doi: 10.2165/00003495-200666060-00006
- Andersson, K. E. (2011). "Muscarinic acetylcholine receptors in the urinary tract," in Urinary Tract. Handbook of Experimental Pharmacology, eds K. E. Andersson, and M. Michel (Berlin: Springer), 319–344. doi: 10.1007/978-3-642-16499-6 16
- Andersson, K. E., and Arner, A. (2004). Urinary bladder contraction and relaxation: physiology and pathophysiology. *Physiol. Rev.* 84, 935–986. doi: 10.1152/ physrev.00038.2003
- Angelini, K. J. (2017). An integrative review of current research on the role of the female urinary microbiota in overactive bladder symptoms. Urol. Nurs. 37, 94–100. doi: 10.7257/1053-816X.2017.37.2.94
- Apostolidis, A., Brady, C. M., Yiangou, Y., Davis, J., Fowler, C. J., and Anand, P. (2005). Capsaicin receptor TRPV1 in urothelium of neurogenic human bladders and effect of intravesical resiniferatoxin. *Urology* 65, 400–405. doi: 10.1016/j.urology.2004.10.007
- Arms, L., and Vizzard, M. A. (2011). Neuropeptides in lower urinary tract function. Handb. Exp. Pharmacol. 202, 395–423. doi: 10.1007/978-3-642-16499-6_19
- Bakali, E., Mcdonald, J., Elliott, R. A., Lambert, D. G., and Tincello, D. G. (2016). Cannabinoid receptor expression in the bladder is altered in detrusor overactivity. *Int. Urogynecol. J.* 27, 129–139. doi: 10.1007/s00192-015-2802-x
- Balachandran, A. A., Wildman, S. S., Strutt, M., and Duckett, J. (2016). Is chronic urinary infection a cause of overactive bladder? *Eur. J. Obstet. Gynecol. Reprod. Biol.* 201, 108–112. doi: 10.1016/j.ejogrb.2016.03.024
- Birder, L., and Andersson, K. E. (2013). Urothelial signaling. *Physiol. Rev.* 93, 653-680. doi: 10.1152/physrev.00030.2012
- Black, L. V., Ness, T. J., and Robbins, M. T. (2009). Effects of oxytocin and prolactin on stress-induced bladder hypersensitivity in female rats. J. Pain 10, 1065–1072. doi: 10.1016/j.jpain.2009.04.007
- Boudes, M., Uvin, P., Pinto, S., Freichel, M., Birnbaumer, L., Voets, T., et al. (2013). Crucial role of TRPC1 and TRPC4 in cystitis-induced neuronal sprouting and bladder overactivity. *PLoS One* 8:e69550. doi: 10.1371/journal.pone.0069550
- Brierley, S. M., and Linden, D. R. (2014). Neuroplasticity and dysfunction after gastrointestinal inflammation. *Nat. Rev. Gastroenterol. Hepatol.* 11, 611–627. doi: 10.1038/nrgastro.2014.103
- Brumovsky, P. R., and Gebhart, G. F. (2010). Visceral organ cross-sensitization an integrated perspective. *Auton. Neurosci.* 153, 106–115. doi: 10.1016/j.autneu. 2009.07.006
- Buffington, C. A. T. (2011). Idiopathic cystitis in domestic cats—beyond the lower urinary tract. J. Vet. Intern. Med. 25, 784–796. doi: 10.1111/j.1939-1676.2011. 0732.x
- Chen, W., Ye, D.-Y., Han, D.-J., Fu, G.-Q., Zeng, X., Lin, W., et al. (2016). Elevated level of nerve growth factor in the bladder pain syndrome/interstitial cystitis: a meta-analysis. *Springerplus* 5:1072. doi: 10.1186/s40064-016-2719-y
- Chen, X., and Gebhart, G. F. (2010). Differential purinergic signaling in bladder sensory neurons of naïve and bladder inflamed mice. *Pain* 148, 462–472. doi: 10.1016/j.pain.2009.12.006
- Chiang, G., Patra, P., Letourneau, R., Jeudy, S., Boucher, W., Green, M., et al. (2000). Pentosanpolysulfate inhibits mast cell histamine secretion and intracellular calcium ion levels: an alternative explanation of its beneficial effect in interstitial cystitis. *J. Urol.* 164, 2119–2125. doi: 10.1016/S0022-5347(05) 66981-9
- Chopra, B., Barrick, S. R., Meyers, S., Beckel, J. M., Zeidel, M. L., Ford, A. P., et al. (2005). Expression and function of bradykinin B1 and B2 receptors in

normal and inflamed rat urinary bladder urothelium. J. Physiol. 562, 859-871. doi: 10.1113/jphysiol.2004.071159

- Christianson, J. A., Liang, R., Ustinova, E. E., Davis, B. M., Fraser, M. O., and Pezzone, M. A. (2007). Convergence of bladder and colon sensory innervation occurs at the primary afferent level. *Pain* 128, 235–243. doi: 10.1016/j.pain.2006. 09.023
- Christmas, T. J., Rode, J., Chapple, C. R., Milroy, E. J., and Turner-Warwick, R. T. (1990). Nerve fibre proliferation in interstitial cystitis. *Virchows Arch. A Pathol. Anat. Histopathol.* 416, 447–451. doi: 10.1007/BF01605152
- Chung, M. K., Butrick, C. W., and Chung, C. W. (2010). The overlap of interstitial cystitis/painful bladder syndrome and overactive bladder. *JSLS* 14, 83–90. doi: 10.4293/108680810X12674612014743
- Contreras-Sanz, A., Krska, L., Balachandran, A. A., Curtiss, N. L., Khasriya, R., Kelley, S., et al. (2016). Altered urothelial ATP signaling in a major subset of human overactive bladder patients with pyuria. *Am. J. Physiol. Renal Physiol.* 311, F805–F816. doi: 10.1152/ajprenal.00339.2015
- Curtiss, N., Balachandran, A., Krska, L., Peppiatt-Wildman, C., Wildman, S., and Duckett, J. (2017). A case controlled study examining the bladder microbiome in women with Overactive Bladder (OAB) and healthy controls. *Eur. J. Obstet. Gynecol. Reprod. Biol.* 214, 31–35. doi: 10.1016/j.ejogrb.2017.04.040
- Cvach, K., and Rosamilia, A. (2015). Review of intravesical therapies for bladder pain syndrome/interstitial cystitis. *Transl. Androl. Urol.* 4, 629–637. doi: 10. 3978/j.issn.2223-4683.2015.10.07
- Daly, D., Rong, W., Chess-Williams, R., Chapple, C., and Grundy, D. (2007). Bladder afferent sensitivity in wild-type and TRPV1 knockout mice. *J. Physiol.* 583, 663–674. doi: 10.1113/jphysiol.2007.139147
- Dang, K., Lamb, K., Cohen, M., Bielefeldt, K., and Gebhart, G. F. (2008). Cyclophosphamide-induced bladder inflammation sensitizes and enhances P2X receptor function in rat bladder sensory neurons. *J. Neurophysiol.* 99, 49–59. doi: 10.1152/jn.00211.2007
- Davidson, S., Copits, B. A., Zhang, J., Page, G., Ghetti, A., and Gereau, R. W. (2014). Human sensory neurons: membrane properties and sensitization by inflammatory mediators. *Pain* 155, 1861–1870. doi: 10.1016/j.pain.2014.06.017
- de Groat, W. C., and Yoshimura, N. (2009). Afferent nerve regulation of bladder function in health and disease. *Handb. Exp. Pharmacol.* 194, 91–138. doi: 10. 1007/978-3-540-79090-7_4
- de Groat, W. C., and Yoshimura, N. (2015). Anatomy and physiology of the lower urinary tract. *Handb. Clin. Neurol.* 130, 61–108. doi: 10.1016/B978-0-444-63247-0.00005-5
- DeBerry, J., Ness, T. J., Robbins, M., and Randich, A. (2007). Inflammationinduced enhancement of the visceromotor reflex to urinary bladder distention: modulation by endogenous opioids and the effects of early-in-life experience with bladder inflammation. J. Pain 8, 914–923. doi: 10.1016/j.jpain.2007.06.011
- DeBerry, J., Randich, A., Shaffer, A. D., Robbins, M. T., and Ness, T. J. (2010). Neonatal bladder inflammation produces functional changes and alters neuropeptide content in bladders of adult female rats. *J. Pain* 11, 247–255. doi: 10.1016/j.jpain.2009.07.010
- DeBerry, J. J., Robbins, M. T., and Ness, T. J. (2015a). The amygdala central nucleus is required for acute stress-induced bladder hyperalgesia in a rat visceral pain model. *Brain Res.* 1606, 77–85. doi: 10.1016/j.brainres.2015.01.008
- DeBerry, J. J., Saloman, J. L., Dragoo, B. K., Albers, K. M., and Davis, B. M. (2015b). Artemin immunotherapy is effective in preventing and reversing cystitis-induced bladder hyperalgesia via TRPA1 regulation. *J. Pain* 16, 628–636. doi: 10.1016/j.jpain.2015.03.014
- DeBerry, J. J., Schwartz, E. S., and Davis, B. M. (2014). TRPA1 mediates bladder hyperalgesia in a mouse model of cystitis. *Pain* 155, 1280–1287. doi: 10.1016/j. pain.2014.03.023
- Drake, M. J., Morris, N., Apostolidis, A., Rahnama'i, M. S., and Marchesi, J. R. (2017). The urinary microbiome and its contribution to lower urinary tract symptoms; ICI-RS 2015. *Neurourol. Urodyn.* 36, 850–853. doi: 10.1002/nau. 23006
- Dupont, M. C., Spitsbergen, J. M., Kim, K. B., Tuttle, J. B., and Steers, W. D. (2001). Histological and neurotrophic changes triggered by varying models of bladder inflammation. J. Urol. 166, 1111–1118. doi: 10.1016/S0022-5347(05)65931-9
- Durden, E., Walker, D., Gray, S., Fowler, R., Juneau, P., and Gooch, K. (2018). The economic burden of overactive bladder (OAB) and its effects on the costs associated with other chronic, age-related comorbidities in the United States. *Neurourol. Urodyn.* 37, 1641–1649. doi: 10.1002/nau.23513

- Egeland, B. (2009). Taking stock: childhood emotional maltreatment and developmental psychopathology. *Child Abuse Negl.* 33, 22–26. doi: 10.1016/j. chiabu.2008.12.004
- Ekman, M., Zhu, B., Sward, K., and Uvelius, B. (2017). Neurite outgrowth in cultured mouse pelvic ganglia Effects of neurotrophins and bladder tissue. *Auton. Neurosci.* 205, 41–49. doi: 10.1016/j.autneu.2017.03.004
- Elbadawi, A. E., and Light, J. K. (1996). Distinctive ultrastructural pathology of nonulcerative interstitial cystitis: new observations and their potential significance in pathogenesis. *Urol. Int.* 56, 137–162. doi: 10.1159/000282832
- El-Mansoury, M., Boucher, W., Sant, G. R., and Theoharides, T. C. (1994). Increased urine histamine and methylhistamine in interstitial cystitis. J. Urol 152, 350–353. doi: 10.1016/S0022-5347(17)32737-4
- Erickson, A., Deiteren, A., Harrington, A. M., Garcia-Caraballo, S., Castro, J., Caldwell, A., et al. (2018). Voltage-gated sodium channels: (NaV)igating the field to determine their contribution to visceral nociception. *J. Physiol* 596, 785–807. doi: 10.1113/JP273461
- Everaerts, W., Vriens, J., Owsianik, G., Appendino, G., Voets, T., De Ridder, D., et al. (2010a). Functional characterization of transient receptor potential channels in mouse urothelial cells. *Am. J. Physiol. Renal Physiol.* 298, F692– F701. doi: 10.1152/ajprenal.00599.2009
- Everaerts, W., Zhen, X., Ghosh, D., Vriens, J., Gevaert, T., Gilbert, J. P., et al. (2010b). Inhibition of the cation channel TRPV4 improves bladder function in mice and rats with cyclophosphamide-induced cystitis. *Proc. Natl. Acad. Sci.* U.S.A. 107, 19084–19089. doi: 10.1073/pnas.1005333107
- Farmer, M. A., Huang, L., Martucci, K., Yang, C. C., Maravilla, K. R., Harris, R. E., et al. (2015). Brain white matter abnormalities in female interstitial cystitis/bladder pain syndrome: a MAPP network neuroimaging study. *J. Urol.* 194, 118–126. doi: 10.1016/j.juro.2015.02.082
- Fowler, C. J., Griffiths, D., and De Groat, W. C. (2008). The neural control of micturition. *Nat. Rev. Neurosci.* 9, 453–466. doi: 10.1038/nrn2401
- Fry, C. H., Daneshgari, F., Thor, K., Drake, M., Eccles, R., Kanai, A. J., et al. (2010). Animal models and their use in understanding lower urinary tract dysfunction. *Neurourol. Urodyn.* 29, 603–608. doi: 10.1002/nau.20903
- Fuentes, I. M., and Christianson, J. A. (2018). The influence of early life experience on visceral pain. *Front. Syst. Neurosci.* 12:2. doi: 10.3389/fnsys.2018.00002
- Furuta, A., Yamamoto, T., Suzuki, Y., Gotoh, M., Egawa, S., and Yoshimura, N. (2018). Comparison of inflammatory urine markers in patients with interstitial cystitis and overactive bladder. *Int. Urogynecol. J.* 29, 961–966. doi: 10.1007/ s00192-017-3547-5
- Giglio, D., Ryberg, A. T., To, K., Delbro, D. S., and Tobin, G. (2005). Altered muscarinic receptor subtype expression and functional responses in cyclophosphamide induced cystitis in rats. *Auton. Neurosci.* 122, 9–20. doi: 10.1016/j.autneu.2005.07.005
- Giglio, D., and Tobin, G. (2009). Muscarinic receptor subtypes in the lower urinary tract. *Pharmacology* 83, 259–269. doi: 10.1159/000209255
- Gill, K., Kang, R., Sathiananthamoorthy, S., Khasriya, R., and Malone-Lee, J. (2018). A blinded observational cohort study of the microbiological ecology associated with pyuria and overactive bladder symptoms. *Int. Urogynecol. J.* 29, 1493–1500. doi: 10.1007/s00192-018-3558-x
- Golabek, T., Skalski, M., Przydacz, M., Swierkosz, A., Siwek, M., Golabek, K., et al. (2016). Lower urinary tract symptoms, nocturia and overactive bladder in patients with depression and anxiety. *Psychiatr. Pol.* 50, 417–430. doi: 10.12740/ PP/OnlineFirst/59162
- Goldstein, H. B., Safaeian, P., Garrod, K., Finamore, P. S., Kellogg-Spadt, S., and Whitmore, K. E. (2008). Depression, abuse and its relationship to interstitial cystitis. *Int. Urogynecol. J. Pelvic. Floor Dysfunct.* 19, 1683–1686. doi: 10.1007/ s00192-008-0712-x
- Greenwood-Van Meerveld, B., Mohammadi, E., Tyler, K., Van Gordon, S., Parker, A., Towner, R., et al. (2015). Mechanisms of visceral organ crosstalk: importance of alterations in permeability in rodent models. *J. Urol.* 194, 804–811. doi: 10.1016/j.juro.2015.02.2944
- Gregory, N., Harris, A. L., Robinson, C. R., Dougherty, P. M., Fuchs, P. N., and Sluka, K. A. (2013). An overview of animal models of pain: disease models and outcome measures. J. Pain 14, 1255–1269. doi: 10.1016/j.jpain.2013.1006.1008
- Griffiths, D. (2015). Neural control of micturition in humans: a working model. Nat. Rev. Urol. 12, 695-705. doi: 10.1038/nrurol.2015.266
- Grundy, L., Chess-Williams, R., Brierley, S. M., Mills, K., Moore, K. H., Mansfield, K., et al. (2018a). NKA enhances bladder afferent mechanosensitivity

via urothelial and detrusor activation. Am. J. Physiol. Renal Physiol. 315, F1174–F1185. doi: 10.1152/ajprenal.00106.2018

- Grundy, L., Daly, D. M., Chapple, C., Grundy, D., and Chess-Williams, R. (2018b). TRPV1 enhances the afferent response to P2X receptor activation in the mouse urinary bladder. *Sci. Rep.* 8:197. doi: 10.1038/s41598-017-18136-w
- Grundy, L., Erickson, A., and Brierley, S. M. (2018c). Visceral pain. Annu. Rev. Physiol. [Epub ahead of print].
- Grundy, L., Erickson, A., Caldwell, A., Garcia-Caraballo, S., Rychkov, G., Harrington, A., et al. (2018d). Tetrodotoxin-sensitive voltage-gated sodium channels regulate bladder afferent responses to distension. *Pain* 159, 2573–2584. doi: 10.1097/j.pain.00000000001368
- Grundy, L., Harrington, A., Castro, J., Garcia-Caraballo, S., Dieteren, A., Maddern, J., et al. (2018e). Chronic linaclotide treatment reduces colitisinduced neuroplasticity and reverses persistent bladder dysfunction. *JCI Insight* doi: 10.1172/jci.insight.121841 [Epub ahead of print].
- Hanno, P., and Dmochowski, R. (2009). Status of international consensus on interstitial cystitis/bladder pain syndrome/painful bladder syndrome: 2008 snapshot. *Neurourol. Urodyn.* 28, 274–286. doi: 10.1002/nau.20687
- Hanno, P. M. (2002). Interstitial cystitis-epidemiology, diagnostic criteria, clinical markers. *Rev. Urol.* 4(Suppl. 1), S3–S8.
- Harper, A., Naghibi, M. M., and Garcha, D. (2018). The role of bacteria, probiotics and diet in irritable bowel syndrome. *Foods* 7:E13. doi: 10.3390/foods702 0013
- Hauser, P. J., Dozmorov, M. G., Bane, B. L., Slobodov, G., Culkin, D. J., and Hurst, R. E. (2008). Abnormal expression of differentiation related proteins and proteoglycan core proteins in the urothelium of patients with interstitial cystitis. *J. Urol.* 179, 764–769. doi: 10.1016/j.juro.2007.09.022
- Hayashi, Y., Takimoto, K., Chancellor, M. B., Erickson, K. A., Erickson, V. L., Kirimoto, T., et al. (2009). Bladder hyperactivity and increased excitability of bladder afferent neurons associated with reduced expression of Kv1.4 alphasubunit in rats with cystitis. Am. J. Physiol. Regul. Integr. Comp. Physiol. 296, R1661–R1670. doi: 10.1152/ajpregu.91054.2008
- Haylen, B. T., De Ridder, D., Freeman, R. M., Swift, S. E., Berghmans, B., Lee, J., et al. (2010). An International Urogynecological Association (IUGA)/International Continence Society (ICS) joint report on the terminology for female pelvic floor dysfunction. *Neurourol. Urodyn.* 29, 4–20. doi: 10.1002/ nau.20798
- Hedlund, P. (2014). Cannabinoids and the endocannabinoid system in lower urinary tract function and dysfunction. *Neurourol. Urodyn.* 33, 46–53. doi: 10.1002/nau.22442
- Hedlund, P., and Gratzke, C. (2016). The endocannabinoid system a target for the treatment of LUTS? *Nat. Rev. Urol.* 13, 463–470. doi: 10.1038/nrurol.2016.110
- Heim, C., and Binder, E. B. (2012). Current research trends in early life stress and depression: review of human studies on sensitive periods, gene– environment interactions, and epigenetics. *Exp. Neurol.* 233, 102–111. doi: 10.1016/j.expneurol.2011.10.032
- Hilt, E. E., Mckinley, K., Pearce, M. M., Rosenfeld, A. B., Zilliox, M. J., Mueller, E. R., et al. (2014). Urine is not sterile: use of enhanced urine culture techniques to detect resident bacterial flora in the adult female bladder. *J. Clin. Microbiol.* 52, 871–876. doi: 10.1128/JCM.02876-13
- Homma, Y., Ueda, T., Tomoe, H., Lin, A. T., Kuo, H. C., Lee, M. H., et al. (2016). Clinical guidelines for interstitial cystitis and hypersensitive bladder updated in 2015. *Int. J. Urol.* 23, 542–549. doi: 10.1111/iju.13118
- Hughes, F. M. Jr., Kennis, J. G., Youssef, M. N., Lowe, D. W., Shaner, B. E., and Purves, J. T. (2016). The NACHT, LRR and PYD Domains-Containing Protein 3 (NLRP3) inflammasome mediates inflammation and voiding dysfunction in a lipopolysaccharide-induced rat model of cystitis. *J. Clin. Cell Immunol.* 7:396. doi: 10.4172/2155-9899.1000396
- Hughes, P. A., Zola, H., Penttila, I. A., Blackshaw, L. A., Andrews, J. M., and Krumbiegel, D. (2013). Immune activation in irritable bowel syndrome: can neuroimmune interactions explain symptoms? *Am. J. Gastroenterol.* 108, 1066– 1074. doi: 10.1038/ajg.2013.120
- Hurst, R. E., Meerveld, B. G., Wisniewski, A. B., Vangordon, S., Lin, H., Kropp, B. P., et al. (2015). Increased bladder permeability in interstitial cystitis/painful bladder syndrome. *Transl. Androl. Urol.* 4, 563–571.
- Irwin, D. E., Milsom, I., Hunskaar, S., Reilly, K., Kopp, Z., Herschorn, S., et al. (2006). Population-based survey of urinary incontinence, overactive bladder, and other lower urinary tract symptoms in five countries: results of the EPIC

study. Eur. Urol. 50, 1306–1314; discussion 1314–1315. doi: 10.1016/j.eururo. 2006.09.019

- Izzo, A. A., Muccioli, G. G., Ruggieri, M. R., and Schicho, R. (2015). Endocannabinoids and the digestive tract and bladder in health and disease. *Handb. Exp. Pharmacol.* 231, 423–447. doi: 10.1007/978-3-319-20825-1_15
- Jacobs, B. L., Smaldone, M. C., Tyagi, V., Philips, B. J., Jackman, S. V., Leng, W. W., et al. (2010). Increased nerve growth factor in neurogenic overactive bladder and interstitial cystitis patients. *Can. J. Urol.* 17, 4989–4994.
- Jhang, J. F., Hsu, Y. H., and Kuo, H. C. (2016). Urothelial functional protein and sensory receptors in patients with interstitial cystitis/bladder pain syndrome with and without Hunner's lesion. Urology 98, 44–49. doi: 10.1016/j.urology. 2016.08.029
- Jhang, J. F., and Kuo, H. C. (2016). Pathomechanism of interstitial cystitis/bladder pain syndrome and mapping the heterogeneity of disease. *Int. Neurourol. J.* 20, S95–S104. doi: 10.5213/inj.1632712.356
- Kastrup, J., Hald, T., Larsen, S., and Nielsen, V. G. (1983). Histamine content and mast cell count of detrusor muscle in patients with interstitial cystitis and other types of chronic cystitis. *Br. J. Urol.* 55, 495–500. doi: 10.1111/j.1464-410X.1983. tb03356.x
- Kawamorita, N., Yoshikawa, S., Kashyap, M., Tyagi, P., Arai, Y., Chancellor, M. B., et al. (2016). Liposome-based Intravesical Therapy targeting Nerve Growth Factor (NGF) ameliorates bladder hypersensitivity in rats with experimental colitis. J. Urol. 195, 1920–1926. doi: 10.1016/j.juro.2015.12.090
- Keay, S. K., Birder, L. A., and Chai, T. C. (2014). Evidence for bladder urothelial pathophysiology in functional bladder disorders. *BioMed. Res. Int.* 2014:865463. doi: 10.1155/2014/865463
- Khasriya, R., Sathiananthamoorthy, S., Ismail, S., Kelsey, M., Wilson, M., Rohn, J. L., et al. (2013). Spectrum of bacterial colonization associated with urothelial cells from patients with chronic lower urinary tract symptoms. *J. Clin. Microbiol.* 51, 2054–2062. doi: 10.1128/JCM.03314-12
- Kim, J. C., Park, E. Y., Hong, S. H., Seo, S. I., Park, Y. H., and Hwang, T. K. (2005). Changes of urinary nerve growth factor and prostaglandins in male patients with overactive bladder symptom. *Int. J. Urol.* 12, 875–880. doi: 10.1111/j.1442-2042.2005.01140.x
- Kim, J. C., Park, E. Y., Seo, S. I., Park, Y. H., and Hwang, T. K. (2006). Nerve growth factor and prostaglandins in the urine of female patients with overactive bladder. *J. Urol.* 175, 1773–1776; discussion 1776. doi: 10.1016/S0022-5347(05) 00992-4
- Kim, S. H., Kim, T. B., Kim, S. W., and Oh, S. J. (2009). Urodynamic findings of the painful bladder syndrome/interstitial cystitis: a comparison with idiopathic overactive bladder. J. Urol. 181, 2550–2554. doi: 10.1016/j.juro.2009.01.106
- Lai, H., Gardner, V., Vetter, J., and Andriole, G. L. (2015). Correlation between psychological stress levels and the severity of overactive bladder symptoms. *BMC Urol.* 15:14. doi: 10.1186/s12894-015-0009-6
- Lai, H. H., Qiu, C. S., Crock, L. W., Morales, M. E., Ness, T. J., and Gereau, R. W. T. (2011). Activation of spinal extracellular signal-regulated kinases (ERK) 1/2 is associated with the development of visceral hyperalgesia of the bladder. *Pain* 152, 2117–2124. doi: 10.1016/j.pain.2011.05.017
- Lai, M. C., Kuo, Y. C., and Kuo, H. C. (2013). Intravesical hyaluronic acid for interstitial cystitis/painful bladder syndrome: a comparative randomized assessment of different regimens. *Int. J. Urol.* 20, 203–207. doi: 10.1111/j.1442-2042.2012.03135.x
- Lakeman, M. M., and Roovers, J. P. (2016). Urinary tract infections in women with urogynaecological symptoms. *Curr. Opin. Infect. Dis.* 29, 92–97. doi: 10.1097/ QCO.00000000000229
- Lavelle, J., Meyers, S., Ramage, R., Bastacky, S., Doty, D., Apodaca, G., et al. (2002). Bladder permeability barrier: recovery from selective injury of surface epithelial cells. *Am. J. Physiol. Renal Physiol.* 283, F242–F253. doi: 10.1152/ajprenal.00307. 2001
- Lazzeri, M. (2006). The physiological function of the urothelium-more than a simple barrier. *Urol. Int.* 76, 289–295. doi: 10.1159/000092049
- Lee, U. J., Ackerman, A. L., Wu, A., Zhang, R., Leung, J., Bradesi, S., et al. (2015). Chronic psychological stress in high-anxiety rats induces sustained bladder hyperalgesia. *Physiol. Behav.* 139, 541–548. doi: 10.1016/j.physbeh.2014.11.045
- Lei, Q., Pan, X. Q., Villamor, A. N., Asfaw, T. S., Chang, S., Zderic, S. A., et al. (2013). Lack of transient receptor potential vanilloid 1 channel modulates the development of neurogenic bladder dysfunction induced by cross-sensitization in afferent pathways. J. Neuroinflammation 10:3. doi: 10.1186/1742-2094-10-3

- Leiby, B. E., Landis, J. R., Propert, K. J., and Tomaszewski, J. E. (2007). Discovery of morphological subgroups that correlate with severity of symptoms in interstitial cystitis: a proposed biopsy classification system. J. Urol. 177, 142–148. doi: 10.1016/j.juro.2006.08.096
- Leue, C., Kruimel, J., Vrijens, D., Masclee, A., Van Os, J., and Van Koeveringe, G. (2017). Functional urological disorders: a sensitized defence response in the bladder-gut-brain axis. *Nat. Rev. Urol.* 14, 153–163. doi: 10.1038/nrurol.2016. 227
- Liang, R., Ustinova, E. E., Patnam, R., Fraser, M. O., Gutkin, D. W., and Pezzone, M. A. (2007). Enhanced expression of mast cell growth factor and mast cell activation in the bladder following the resolution of trinitrobenzenesulfonic acid (TNBS) colitis in female rats. *Neurourol. Urodyn.* 26, 887–893. doi: 10. 1002/nau.20410
- Liu, H. T., and Kuo, H. C. (2007). Increased expression of transient receptor potential vanilloid subfamily 1 in the bladder predicts the response to intravesical instillations of resiniferatoxin in patients with refractory idiopathic detrusor overactivity. *BJU Int.* 100, 1086–1090. doi: 10.1111/j.1464-410X.2007. 07151.x
- Liu, H. T., Shie, J. H., Chen, S. H., Wang, Y. S., and Kuo, H. C. (2012). Differences in mast cell infiltration, E-cadherin, and zonula occludens-1 expression between patients with overactive bladder and interstitial cystitis/bladder pain syndrome. Urology 80, 225.e13–225.e18. doi: 10.1016/j.urology.2012. 01.047
- Liu, Q., and Dong, X. (2015). The role of the Mrgpr receptor family in itch. *Handb. Exp. Pharmacol.* 226, 71–88. doi: 10.1007/978-3-662-44605-8_5
- Lovick, T. A. (2016). Central control of visceral pain and urinary tract function. *Auton. Neurosci.* 200, 35–42. doi: 10.1016/j.autneu.2016.02.001
- Lundeberg, T., Liedberg, H., Nordling, L., Theodorsson, E., Owzarski, A., and Ekman, P. (1993). Interstitial cystitis: correlation with nerve fibres, mast cells and histamine. *Br. J. Urol.* 71, 427–429. doi: 10.1111/j.1464-410X.1993.tb15 986.x
- MacDiarmid, S. A., and Sand, P. K. (2007). Diagnosis of interstitial cystitis/ painful bladder syndrome in patients with overactive bladder symptoms. *Rev. Urol.* 9, 9–16.
- Malykhina, A. P., Qin, C., Greenwood-Van Meerveld, B., Foreman, R. D., Lupu, F., and Akbarali, H. I. (2006). Hyperexcitability of convergent colon and bladder dorsal root ganglion neurons after colonic inflammation: mechanism for pelvic organ cross-talk. *Neurogastroenterol. Motil.* 18, 936–948. doi: 10.1111/j.1365-2982.2006.00807.x
- Manriquez, V., Castro Caperan, D., Guzman, R., Naser, M., Iglesia, V., and Lagos, N. (2015). First evidence of neosaxitoxin as a long-acting pain blocker in bladder pain syndrome. *Int. Urogynecol. J.* 26, 853–858. doi: 10.1007/s00192-014-2608-2
- McDermott, C., Chess-Williams, R., Mills, K. A., Kang, S. H., Farr, S. E., Grant, G. D., et al. (2013). Alterations in acetylcholine, PGE2 and IL6 release from urothelial cells following treatment with pyocyanin and lipopolysaccharide. *Toxicol. In Vitro* 27, 1693–1698. doi: 10.1016/j.tiv.2013.04.015
- McKernan, L. C., Walsh, C. G., Reynolds, W. S., Crofford, L. J., Dmochowski, R. R., and Williams, D. A. (2017). Psychosocial co-morbidities in Interstitial Cystitis/Bladder Pain syndrome (IC/BPS): a systematic review. *Neurourol. Urodyn.* 37, 926–941. doi: 10.1002/nau.23421
- McLennan, M. T. (2014). Interstitial cystitis: epidemiology, pathophysiology, and clinical presentation. Obstet. Gynecol. Clin. North Am. 41, 385–395. doi: 10. 1016/j.ogc.2014.05.004
- Merrill, L., Girard, B. M., May, V., and Vizzard, M. A. (2012). Transcriptional and translational plasticity in rodent urinary bladder TRP channels with urinary bladder inflammation, bladder dysfunction, or postnatal maturation. J. Mol. Neurosci. 48, 744–756. doi: 10.1007/s12031-012-9867-5
- Merrill, L., Gonzalez, E. J., Girard, B. M., and Vizzard, M. A. (2016). Receptors, channels, and signalling in the urothelial sensory system in the bladder. *Nat. Rev. Urol.* 13, 193–204. doi: 10.1038/nrurol.2016.13
- Merrill, L., Malley, S., and Vizzard, M. A. (2013). Repeated variate stress in male rats induces increased voiding frequency, somatic sensitivity, and urinary bladder nerve growth factor expression. Am. J. Physiol. Regul. Integr. Comp. Physiol. 305, R147–R156. doi: 10.1152/ajpregu.00089.2013
- Michel, M. C. (2015). Therapeutic modulation of urinary bladder function: multiple targets at multiple levels. *Annu. Rev. Pharmacol. Toxicol.* 55, 269–287. doi: 10.1146/annurev-pharmtox-010814-124536

- Minaglia, S., Özel, B., Bizhang, R., and Mishell, D. R. (2005). Increased prevalence of interstitial cystitis in women with detrusor overactivity refractory to anticholinergic therapy. *Urology* 66, 702–706. doi: 10.1016/j.urology.2005.04. 042
- Munoz, A. (2016). Unmasking roles of the peripheral endocannabinoid system associated with bladder overactivity. *BJU Int.* 117, 713–714. doi: 10.1111/bju. 13336
- Ness, T. J., and Randich, A. (2010). Neonatal bladder inflammation alters activity of adult rat spinal visceral nociceptive neurons. *Neurosci. Lett.* 472, 210–214. doi: 10.1016/j.neulet.2010.02.007
- Nickel, J. C., Hanno, P., Kumar, K., and Thomas, H. (2012). Second multicenter, randomized, double-blind, parallel-group evaluation of effectiveness and safety of intravesical sodium chondroitin sulfate compared with inactive vehicle control in subjects with interstitial cystitis/bladder pain syndrome. Urology 79, 1220–1224. doi: 10.1016/j.urology.2012.01.059
- Ochodnicky, P., Michel, M. B., Butter, J. J., Seth, J., Panicker, J. N., and Michel, M. C. (2013). Bradykinin modulates spontaneous nerve growth factor production and stretch-induced ATP release in human urothelium. *Pharmacol. Res.* 70, 147–154. doi: 10.1016/j.phrs.2013.01.010
- Oliveira, P. D., Tarricone, R., and Guzman, S. A. (2014). Glycosaminoglycanreplenishment therapy: Rationale for use and current evidence. *EMJ Urol.* 1, 41–47.
- Pan, X. Q., Gonzalez, J. A., Chang, S., Chacko, S., Wein, A. J., and Malykhina, A. P. (2010). Experimental colitis triggers the release of substance P and calcitonin gene-related peptide in the urinary bladder via TRPV1 signaling pathways. *Exp. Neurol.* 225, 262–273. doi: 10.1016/j.expneurol.2010.05.012
- Parsons, B. A., and Drake, M. J. (2011). "Animal Models in Overactive Bladder Research," in *Urinary Tract*, eds K.-E. Andersson and M. C. Michel (Berlin: Springer), 15–43.
- Parsons, C. L., Greenberger, M., Gabal, L., Bidair, M., and Barme, G. (1998). The role of urinary potassium in the pathogenesis and diagnosis of interstitial cystitis. J. Urol. 159, 1862–1866; discussion 1866–1867.
- Pearce, M. M., Hilt, E. E., Rosenfeld, A. B., Zilliox, M. J., Thomas-White, K., Fok, C., et al. (2014). The female urinary microbiome: a comparison of women with and without urgency urinary incontinence. *mBio* 5:e01283-14. doi: 10.1128/mBio. 01283-14
- Persson, R., Wensaas, K.-A., Hanevik, K., Eide, G. E., Langeland, N., and Rortveit, G. (2015). The relationship between irritable bowel syndrome, functional dyspepsia, chronic fatigue and overactive bladder syndrome: a controlled study 6 years after acute gastrointestinal infection. *BMC Gastroenterol*. 15:66. doi: 10.1186/s12876-015-0296-0
- Peters, K. M., Killinger, K. A., and Ibrahim, I. A. (2009). Childhood symptoms and events in women with interstitial cystitis/painful bladder syndrome. Urology 73, 258–262. doi: 10.1016/j.urology.2008.09.014
- Pierce, A. N., and Christianson, J. A. (2015). "Chapter seventeen stress and chronic pelvic pain," in *Progress in Molecular Biology and Translational Science*, eds T. J. Price and G. Dussor (Cambridge, MA: Academic Press), 509–535.
- Pierce, A. N., Di Silvestro, E. R., Eller, O. C., Wang, R., Ryals, J. M., and Christianson, J. A. (2016). Urinary bladder hypersensitivity and dysfunction in female mice following early life and adult stress. *Brain Res.* 1639, 58–73. doi: 10.1016/j.brainres.2016.02.039
- Qiao, L. Y., and Grider, J. R. (2007). Up-regulation of calcitonin gene-related peptide and receptor tyrosine kinase TrkB in rat bladder afferent neurons following TNBS colitis. *Exp. Neurol.* 204, 667–679. doi: 10.1016/j.expneurol. 2006.12.024
- Qiao, L.-Y., and Vizzard, M. A. (2002). Cystitis-induced upregulation of tyrosine kinase (TrkA, TrkB) receptor expression and phosphorylation in rat micturition pathways. J. Comp. Neurol. 454, 200–211. doi: 10.1002/cne.10447
- Randich, A., Uzzell, T., DeBerry, J. J., and Ness, T. J. (2006). Neonatal urinary bladder inflammation produces adult bladder hypersensitivity. J. Pain 7, 469– 479. doi: 10.1016/j.jpain.2006.01.450
- Ren, K., and Dubner, R. (2010). Interactions between the immune and nervous systems in pain. *Nat. Med.* 16, 1267–1276. doi: 10.1038/nm.2234
- Reynolds, W. S., Dmochowski, R., Wein, A., and Bruehl, S. (2016). Does central sensitization help explain idiopathic overactive bladder? *Nat. Rev. Urol.* 13, 481–491. doi: 10.1038/nrurol.2016.95
- Rosa, A. C., and Fantozzi, R. (2013). The role of histamine in neurogenic inflammation. *Br. J. Pharmacol.* 170, 38–45. doi: 10.1111/bph.12266

- Sanden, C., Mori, M., Jogdand, P., Jönsson, J., Krishnan, R., Wang, X., et al. (2017). Broad Th2 neutralization and anti-inflammatory action of pentosan polysulfate sodium in experimental allergic rhinitis. *Immun. Inflamm. Dis.* 5, 300–309. doi: 10.1002/iid3.164
- Schnegelsberg, B., Sun, T. T., Cain, G., Bhattacharya, A., Nunn, P. A., Ford, A. P., et al. (2010). Overexpression of NGF in mouse urothelium leads to neuronal hyperinnervation, pelvic sensitivity, and changes in urinary bladder function. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 298, R534–R547. doi: 10.1152/ ajpregu.00367.2009
- Scott, V. C. S., Haake, D. A., Churchill, B. M., Justice, S. S., and Kim, J.-H. (2015). Intracellular bacterial communities: a potential etiology for chronic lower urinary tract symptoms. *Urology* 86, 425–431. doi: 10.1016/j.urology. 2015.04.002
- Sengupta, J. N., Pochiraju, S., Kannampalli, P., Bruckert, M., Addya, S., Yadav, P., et al. (2013). MicroRNA-mediated GABA(Aα-1) receptor subunit downregulation in adult spinal cord following neonatal cystitis-induced chronic visceral pain in rats. *Pain* 154, 59–70. doi: 10.1016/j.pain.2012.09.002
- Shaffer, A. D., Ball, C. L., Robbins, M. T., Ness, T. J., and Randich, A. (2011). Effects of acute adult and early-in-life bladder inflammation on bladder neuropeptides in adult female rats. *BMC Urol.* 11:18. doi: 10.1186/1471-2490-11-18
- Shaffer, A. D., Ness, T. J., Robbins, M. T., and Randich, A. (2013). Early-in-life bladder inflammation alters opioid peptide content in the spinal cord and bladder of adult female rats. J. Urol. 189, 352–358. doi: 10.1016/j.juro.2012.08. 190
- Shin, K., Lee, J., Guo, N., Kim, J., Lim, A., Qu, L., et al. (2011). Hedgehog/Wnt feedback supports regenerative proliferation of epithelial stem cells in bladder. *Nature* 472, 110–114. doi: 10.1038/nature09851
- Siddiqui, H., Lagesen, K., Nederbragt, A. J., Jeansson, S. L., and Jakobsen, K. S. (2012). Alterations of microbiota in urine from women with interstitial cystitis. *BMC Microbiol*. 12:205. doi: 10.1186/1471-2180-12-205
- Smith, C. P., Vemulakonda, V. M., Kiss, S., Boone, T. B., and Somogyi, G. T. (2005). Enhanced ATP release from rat bladder urothelium during chronic bladder inflammation: effect of botulinum toxin A. *Neurochem. Int.* 47, 291–297. doi: 10.1016/j.neuint.2005.04.021
- Sorkin, L. S., Eddinger, K. A., Woller, S. A., and Yaksh, T. L. (2018). Origins of antidromic activity in sensory afferent fibers and neurogenic inflammation. *Semin. Immunopathol.* 40, 237–247. doi: 10.1007/s00281-017-0669-2
- Spencer, N. J., Greenheigh, S., Kyloh, M., Hibberd, T. J., Sharma, H., Grundy, L., et al. (2018). Identifying unique subtypes of spinal afferent nerve endings within the urinary bladder of mice. *J. Comp. Neurol.* 526, 707–720. doi: 10.1002/cne. 24362
- Suh, Y. S., Ko, K. J., Kim, T. H., Lee, H. S., Sung, H. H., Cho, W. J., et al. (2017). Potential biomarkers for diagnosis of overactive bladder patients: urinary nerve growth factor, prostaglandin e2, and adenosine triphosphate. *Int. Neurourol. J.* 21, 171–177. doi: 10.5213/inj.1732728.364
- Sun, Y., and Chai, T. C. (2004). Up-regulation of P2X3 receptor during stretch of bladder urothelial cells from patients with interstitial cystitis. J. Urol. 171, 448–452. doi: 10.1097/01.ju.0000099660.46774.3c
- Sun, Y., and Chai, T. C. (2006). Augmented extracellular ATP signaling in bladder urothelial cells from patients with interstitial cystitis. *Am. J. Physiol. Cell Physiol.* 290, C27–C34. doi: 10.1152/ajpcell.00552.2004
- Takezawa, K., Kondo, M., Kiuchi, H., Soda, T., Takao, T., Miyagawa, Y., et al. (2014). Combination of bladder ultrasonography and novel cystometry method in mice reveals rapid decrease in bladder capacity and compliance in LPSinduced cystitis. *Am. J. Physiol. Renal Physiol.* 307, F234–F241. doi: 10.1152/ ajprenal.00043.2014
- Taylor, S. E. (2010). Mechanisms linking early life stress to adult health outcomes. Proc. Natl. Acad. Sci. U.S.A. 107, 8507–8512. doi: 10.1073/pnas.1003890107
- Tomaszewski, J. E., Landis, J. R., Russack, V., Williams, T. M., Wang, L. P., Hardy, C., et al. (2001). Biopsy features are associated with primary symptoms in interstitial cystitis: results from the interstitial cystitis database study. Urology 57, 67–81. doi: 10.1016/S0090-4295(01)01166-9
- Truzzi, J. C., Gomes, C. M., Bezerra, C. A., Plata, I. M., Campos, J., Garrido, G. L., et al. (2016). Overactive bladder - 18 years - Part I. *Int. Braz. J. Urol.* 42, 188–198. doi: 10.1590/S1677-5538.IBJU.2015.0365
- Ustinova, E. E., Fraser, M. O., and Pezzone, M. A. (2010). Cross-talk and sensitization of bladder afferent nerves. *Neurourol. Urodyn.* 29, 77–81. doi: 10.1002/nau.20817

- Van Brummen, H. J., Heintz, A. P., and Van Der Vaart, C. H. (2004). The association between overactive bladder symptoms and objective parameters from bladder diary and filling cystometry. *Neurourol. Urodyn.* 23, 38–42. doi: 10.1002/nau.10162
- Vijaya, G., Cartwright, R., Derpapas, A., Gallo, P., Fernando, R., and Khullar, V. (2013). Changes in nerve growth factor level and symptom severity following antibiotic treatment for refractory overactive bladder. *Int. Urogynecol. J.* 24, 1523–1528. doi: 10.1007/s00192-012-2038-y
- Vizzard, M. A. (2001). Alterations in neuropeptide expression in lumbosacral bladder pathways following chronic cystitis. J. Chem. Neuroanat. 21, 125–138. doi: 10.1016/S0891-0618(00)00115-0
- Wada, N., Ameda, K., Furuno, T., Okada, H., Date, I., and Kakizaki, H. (2015). Evaluation of prostaglandin E2 and E-series prostaglandin receptor in patients with interstitial cystitis. *J. Urol.* 193, 1987–1993. doi: 10.1016/j.juro.2015. 01.010
- Walsh, C., Allen, W., Parkin, K., Mukerjee, C., and Moore, K. (2011). Lowcount bacteriuria in refractory idiopathic detrusor overactivity versus controls. *Urogynaecologia* 25, 11–15. doi: 10.4081/uij.2011.e4
- Wang, Z., Chang, H. H., Gao, Y., Zhang, R., Guo, Y., Holschneider, D. P., et al. (2017). Effects of water avoidance stress on peripheral and central responses during bladder filling in the rat: a multidisciplinary approach to the study of urologic chronic pelvic pain syndrome (MAPP) research network study. *PLoS One* 12:e0182976. doi: 10.1371/journal.pone.0182976
- Wang, Z. Y., Wang, P., Merriam, F. V., and Bjorling, D. E. (2008). Lack of TRPV1 inhibits cystitis-induced increased mechanical sensitivity in mice. *Pain* 139, 158–167. doi: 10.1016/j.pain.2008.03.020
- Whiteside, S. A., Razvi, H., Dave, S., Reid, G., and Burton, J. P. (2015). The microbiome of the urinary tract-a role beyond infection. *Nat. Rev. Urol.* 12, 81–90. doi: 10.1038/nrurol.2014.361
- Winder, M., Tobin, G., Zupancic, D., and Romih, R. (2014). Signalling molecules in the urothelium. *Biomed. Res. Int.* 2014:297295. doi: 10.1155/2014/29 7295
- Wood, M. W., Breitschwerdt, E. B., Nordone, S. K., Linder, K. E., and Gookin, J. L. (2012). Uropathogenic *E. coli* promote a paracellular urothelial barrier defect characterized by altered tight junction integrity, epithelial cell sloughing and cytokine release. *J. Comp. Pathol.* 147, 11–19. doi: 10.1016/j.jcpa.2011. 09.005
- Wu, J., Guan, T. J., Zheng, S., Grosjean, F., Liu, W., Xiong, H., et al. (2011). Inhibition of inflammation by pentosan polysulfate impedes the development and progression of severe diabetic nephropathy in aging C57B6 mice. *Lab. Invest.* 91, 1459–1471. doi: 10.1038/labinvest.2011.93

- Xia, C., Shen, S., Hashmi, F., and Qiao, L. Y. (2015). Colitis-induced bladder afferent neuronal activation is regulated by BDNF through PLCgamma pathway. *Exp. Neurol.* 285(Pt B), 126–135. doi: 10.1016/j.expneurol.2015.12.006
- Yamaguchi, O., Honda, K., Nomiya, M., Shishido, K., Kakizaki, H., Tanaka, H., et al. (2007). Defining overactive bladder as hypersensitivity. *Neurourol. Urodyn.* 26, 904–907. doi: 10.1002/nau.20482
- Yoshikawa, S., Kawamorita, N., Oguchi, T., Funahashi, Y., Tyagi, P., Chancellor, M. B., et al. (2015). Pelvic organ cross-sensitization to enhance bladder and urethral pain behaviors in rats with experimental colitis. *Neuroscience* 284, 422–429. doi: 10.1016/j.neuroscience.2014.08.064
- Yoshimura, N., Oguchi, T., Yokoyama, H., Funahashi, Y., Yoshikawa, S., Sugino, Y., et al. (2014). Bladder afferent hyperexcitability in bladder pain syndrome/interstitial cystitis. *Int. J. Urol.* 21(Suppl. 1), 18–25. doi: 10.1111/iju. 12308
- Yoshiyama, M., Mochizuki, T., Nakagomi, H., Miyamoto, T., Kira, S., Mizumachi, R., et al. (2015). Functional roles of TRPV1 and TRPV4 in control of lower urinary tract activity: dual analysis of behavior and reflex during the micturition cycle. *Am. J. Physiol. Renal Physiol.* 308, F1128–F1134. doi: 10.1152/ ajprenal.00016.2015
- Zagorodnyuk, V. P., Brookes, S. J., Spencer, N. J., and Gregory, S. (2009). Mechanotransduction and chemosensitivity of two major classes of bladder afferents with endings in the vicinity to the urothelium. J. Physiol. 587, 3523– 3538. doi: 10.1113/jphysiol.2009.172577
- Zagorodnyuk, V. P., Costa, M., and Brookes, S. J. (2006). Major classes of sensory neurons to the urinary bladder. *Auton. Neurosci.* 12, 390–397. doi: 10.1016/j. autneu.2006.02.007
- Zagorodnyuk, V. P., Gibbins, I. L., Costa, M., Brookes, S. J., and Gregory, S. J. (2007). Properties of the major classes of mechanoreceptors in the guinea pig bladder. J. Physiol. 585, 147–163. doi: 10.1113/jphysiol.2007.140244

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Grundy, Caldwell and Brierley. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.