



# Repurposing of the $\beta$ -Lactam Antibiotic, Ceftriaxone for Neurological Disorders: A Review

Ebrahim M. Yimer\*, Hailemichael Zeru Hishe and Kald Beshir Tuem

Department of Pharmacology and Toxicology, School of Pharmacy, College of Health Sciences, Mekelle University, Mekelle, Ethiopia

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### \*Correspondence:

Ebrahim M. Yimer  
ebrahim99muhammed@gmail.com  
orcid.org/0000-0003-3140-4967

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To date, there is no cure or disease-modifying agents available for most well-known neurological disorders. Current therapy is typically focused on relieving symptoms and supportive care in improving the quality of life of affected patients. Furthermore, the traditional *de novo* drug discovery technique is more challenging, particularly for neurological disorders. Therefore, the repurposing of existing drugs for these conditions is believed to be an efficient and dynamic approach that can substantially reduce the investments spent on drug development. Currently, there is emerging evidence that suggests the potential effect of a beta-lactam antibiotic, ceftriaxone (CEF), to alleviate the symptoms of different experimentally-induced neurological disorders: Parkinson's disease, Alzheimer's disease, amyotrophic lateral sclerosis, epileptic-seizure, brain ischemia, traumatic brain injuries, and neuropathic pain. CEF also affects the markers of oxidative status and neuroinflammation, glutamatergic systems as well as various aggregated toxic proteins involved in the pathogenesis of different neurological disorders. Moreover, it was found that CEF administration to drug dependent animal models improved the withdrawal symptoms upon drug discontinuation. Thus, this review aimed to describe the effects of CEF against multiple models of neurological illnesses, drug dependency, and withdrawal. It also emphasizes the possible mechanisms of neuroprotective actions of CEF with respective neurological maladies.

**Keywords:** ceftriaxone, drug repurposing, neurodegenerative disorders, Alzheimer's disease, Parkinson's disease, stroke, pain, brain ischemia

## INTRODUCTION

The conventional target-based approach for new drug discovery brings paramount challenges as well as massive economic burden, lengthy processes, and might even expose study participants to unexpected adverse events (Lee and Kim, 2016). One alternative paradigm for novel drug discovery is drug repurposing which has recently emerged as a potential strategy of off-target therapeutic actions of existing drugs (Kim, 2015; Parsons, 2018). Currently, drug repurposing is considered a promising tool for novel drug discovery as it is relatively rapid, less costly, and poses a minimal risk of adverse outcomes to study participants. These advantages would possibly overcome the challenges of the conventional *de novo* discovery of new pharmacological agents (Lee and Kim, 2016; Corsello et al., 2017).

Mental and neurological illnesses are multifarious and have a substantial influence on patients as well as being a significant economic burden for nations (Hurd et al., 2013; Gooch et al., 2017).

The etiological aspects of such disorders are diverse, including pathological protein accumulation causing mostly neurodegeneration and dysregulation of the normal developmental and functional process (Di Luca et al., 2018). In 2015, neurological disorders were placed as the foremost global cause of disability and the 2<sup>nd</sup> leading cause of mortality (~17% of global deaths), while Alzheimer's disease (AD) and other dementias were found to be the 4<sup>th</sup> leading cause of mortality and morbidity worldwide, and is steadily among the top three causes of disability in most nations. During the period of 1990–2015, the overall mortality from neurological illnesses was increased by 37% (Feigin et al., 2017).

Moreover, there is no pharmacological agent currently available for the curative or disease-modifying actions of most of the neurological disorders. The present therapeutic approach therefore provides symptomatic management and supportive care in order to improve the longevity and quality of life of patients (Xie et al., 2014; Cummings, 2017; Dorst et al., 2018).

Thus, the repurposing of existing drugs for central nervous system (CNS) related disorders is an attractive, efficient, and dynamic drug development approach that can substantially reduce the investment spent during drug development in terms of both time and money (Caban et al., 2017; Hernandez et al., 2017).

Ceftriaxone (CEF) is a third generation cephalosporin, under the group of  $\beta$ -lactam antibiotics, and is the most frequently used drug for local (skin and soft tissue infections) as well as systemic community and hospital-acquired infections (Pinto Pereira et al., 2004). Recently, emerging evidence, mainly from preclinical studies, have highlighted the therapeutic efficacy of CEF against various neurological disorders, drug dependency, and withdrawal syndrome as well as its neuroprotective potential against various neurotoxic chemicals (**Tables 1–3**).

Therefore, this review is intended to describe the pharmacological effects of CEF in various CNS related disorders conducted in preclinical and clinical studies along with its respective suggested neuroprotective mechanisms.

## EFFECTS OF CEF AGAINST NEURODEGENERATIVE DISORDERS

Neurodegenerative diseases are a diverse set of disorders characterized by a progressive loss of neuronal structure and function in distinct sections of the central nervous system. The neurological sequel of neurodegeneration results in a devastating outcome on mental as well as physical functioning of patients (Gao and Hong, 2008; Cannon and Greenamyre, 2011). Among many neurodegenerative disorders, Alzheimer's disease (AD) and Parkinson's disease (PD) are the most commonly encountered disorders (Xie et al., 2014). Huntington's disease (HD) and amyotrophic lateral sclerosis (ALS) are still devastating neurological disorders, though less prevalent

(Siddique and Siddique, 2008; Franco-Iborra et al., 2018). To date, there is no cure or disease-modifying agents available for most of these neurodegenerative disorders and therapy is currently focused on symptomatic management and supportive care so as to improve the quality of life of the patients (Xie et al., 2014; Cummings, 2017).

A beta-lactam antibiotic, CEF is currently attracting the scientific community due to its multiple mechanisms to relieve symptoms and modify the natural history of various neurodegenerative diseases. Of the different possible mechanisms, upregulation of GLT-1 expression, attenuation of oxidative stress, and neuroinflammation are among the common suggested mechanisms for its neuroprotective actions (**Table 1** and **Figure 1**). For instance, in animal models of Parkinson's disease, CEF exhibited a recovery of memory deficits (Huang et al., 2015), ameliorated abnormal uncontrolled movements (Chotibut et al., 2017), attenuated oxidative damage and restored the reduced levels of endogenous antioxidant enzymes (Bisht et al., 2014; Kaur and Prakash, 2017). Additionally, CEF modulated the expression of tyrosine hydroxylase (Chotibut et al., 2014),  $\alpha$ -synuclein expression (Ho et al., 2018), and neuroinflammation (Kaur and Prakash, 2017) as well as prevented dopaminergic degeneration (Ho et al., 2014), while upregulating the levels of GLT-1 expression and glutamate uptake (Chotibut et al., 2014).

Alzheimer's disease (AD) is one of the most prevalent forms of dementia (Sadigh-Eteghad et al., 2015). Different animal models of Alzheimer's disease were affected by the administration of CEF (**Table 1**). Despite the most commonly underlining protein implicated in the pathology of AD, beta-amyloid ( $\text{A}\beta$ ) protein was not directly affected (Zumkehr et al., 2015), CEF downregulated the messenger RNA (mRNA) expression of *Bace1* (a gene that encodes  $\beta$ -secretase involved in  $\text{A}\beta$  formation), *Ace2* (a gene that encodes enzymes play a part in  $\text{A}\beta$  metabolism), and the expression of gene of  $\beta$ -actin. Furthermore, CEF amplified the gene expression of *Mme* and *Ide* (genes that encode enzymes involved in  $\text{A}\beta$  degradation) as well as the expression of *EPO* gene (a gene that encodes erythropoietin correlated to endothelial function and removal of  $\text{A}\beta$ ) (Tikhonova et al., 2018). CEF-treated animals also showed improvements in memory impairments and restoration of cognitive function and neuronal density (Tikhonova et al., 2017). CEF also attenuated increased levels of acetylcholine esterase enzyme and oxidative stress (Akina et al., 2013). In addition, CEF administration to animal models of Alzheimer's disease displayed an upregulation of GLT-1 expression, preservation of synaptic proteins and downregulation of *tau* proteins (Zumkehr et al., 2015). Furthermore, CEF showed neuroprotective actions in various models of amyotrophic lateral sclerosis (Lewerenz et al., 2009; Yamada and Jinno, 2011; Cudkowicz et al., 2014) and Huntington's disease (Miller et al., 2008; Sari et al., 2010).

## ANALGESIC EFFECT OF CEF

Analgesic effects of the beta-lactam antibiotic CEF, in different preclinical animal models (nociceptive and neuropathic pain

**Abbreviations:** CEF, Ceftriaxone; AD, Alzheimer's disease; HD, Huntington's disease; PD, Parkinson's disease; CNS, central nervous system; TBI, Traumatic brain injury; GLT 1, Glutamate transporter 1; NAc, Nucleus accumbens; PFC, Prefrontal cortex; EAAT2, excitatory amino acid transporter subtype 2

**TABLE 1 |** Effects of CEF against neurodegenerative (ND) disorders.

Models of ND disorders	Experimental approaches	Study subjects	Interventions	Major outcome (s)	References
Dementia with Lewy bodies	Murine model: dementia with Lewy bodies (DLB) were induced in rats.	Male Wistar rats.	Rats were randomly divided in to the sham + saline and DLB + saline groups received saline (1 ml/kg/day) while the DLB + CEF group received CEF (100 mg/kg/day) for 27 days.	Rats that received CEF suppressed hyperactivity in the subthalamic nucleus (STN). The DLB+CEF treated group showed a significantly higher (corrected) neuronal density of neurons in the Hippocampal CA1. Rats in the DLB + CEF group also displayed a significantly lower density of $\alpha$ -synuclein positive cells compared to the DLB control group.	Ho et al., 2018
Parkinson's disease (PD)	Murine model: 6-hydroxydopamine (6-OHDA) induced lesion and amphetamine induced rotation in rats.	Male Sprague Dawley rats.	Rats were grouped into L-dopa alone, L-dopa + CEF, and CEF alone received groups. All groups administered unilateral 6-OHDA lesions. Saline and CEF (200 mg/kg) were administered on day 7 post-6-OHDA lesion for 7 successive days and every other week until the end of the study (day 39).	CEF meaningfully decreased abnormal uncontrolled movements at five time points inspected in the course of L-dopa treatment. Partial recovery of motor impairment from nigrostriatal lesion by L-DOPA was unaffected by CEF. CEF received L-dopa group showed a substantial increment of striatal GLT-1 expression and glutamate uptake. The loss of striatal tyrosine hydroxylase in this group was not suggestively varied compared to the L-DCPA alone received group.	Chotibut et al., 2017
				Daily administration of CEF (100 mg/kg) significantly enhanced the contralateral forepaw stepping (~44%) (this action continued for 1 month after CEF discontinuation). The daily administration of 50 mg/kg CEF showed comparable efficacy to 10 mg/kg of L-DOPA in amplifying the contralateral forepaw stepping (~40%). Moreover, CEF did not produce dyskinesia instead reduced its development but did not affect the occurrence of L-DOPA induced dyskinesia.	Kelsey and Neville, 2014
	Murine model: 6-OHDA induced PD model.	Male Long Evans rats.	In Experiment 1, a 100 mg/kg of CEF for nine rats and, in Experiment 2, the 50 mg/kg CEF alone and along with L-DOPA for 16 rats were administered. Saline and L-DOPA separately were uses as negative and positive controls, respectively.	The 200 mg/kg CEF injection showed an elevation of striatal glutamate uptake in non-lesioned rats and this effect persisted till 2 weeks post-injection. CEF administration in 6- OHDA lesioned rats showed a substantial attenuation (~57%) of tyrosine hydroxylase (TH) loss compared to corresponding vehicle-received group (~85%). CEF also decreased amphetamine-induced rotation and locomotor behavior alteration induced by 6-OHDA. This reduction of TH loss was accompanied with augmented glutamate uptake and GLT-1 expression.	Chotibut et al., 2014
	Murine model: 6-OHDA lesioned PD model.	Male Sprague-Dawley rats.	For naive (not 6- OHDA-lesioned) rats, CEF was injected (200 mg/kg), while the control group administered saline for 7 subsequent days. In animals underwent 6-OHDA lesion, CEF was administered on the same day of lesion and for subsequent 1 week. Whereas, a control group were administered only a vehicle.	The 200 mg/kg CEF injection showed an elevation of striatal glutamate uptake in non-lesioned rats and this effect persisted till 2 weeks post-injection. CEF administration in 6- OHDA lesioned rats showed a substantial attenuation (~57%) of tyrosine hydroxylase (TH) loss compared to corresponding vehicle-received group (~85%). CEF also decreased amphetamine-induced rotation and locomotor behavior alteration induced by 6-OHDA. This reduction of TH loss was accompanied with augmented glutamate uptake and GLT-1 expression.	Chotibut et al., 2014
				(Continued)	

TABLE 1 | Continued

Models of ND disorders	Experimental approaches	Study subjects	Interventions	Major outcome (s)	References
Murine model: MPTP-induced models of PD in rats.	Murine model: MPTP-induced models of PD in rats.	Male Wistar rats.	Rats were randomly grouped into 7 and all groups except vehicle-treated group injected MPTP repeatedly. Group-I: Sham control group. Group-II: saline, Group-III: CEF (100 mg/kg). Group-IV: CEF (200 mg/kg). Group-V: Ropinirole (1.5 mg/kg), Group-VI: Ropinirole (3 mg/kg), Group-VII: CEF (100 mg/kg) + Ropinirole (1.5 mg/kg).	CEF (100 and 200 mg/kg) received animals meaningfully enhanced the motor impairments. CEF injection diminished the oxidative injury and restored the reduced level of endogenous antioxidant enzymes. CEF also meaning fully attenuated the pro-inflammatory cytokines such as TNF- $\alpha$ and IL- $\beta$ in striatum region. Ropinirole pre-treatment with lower dosage of CEF remarkably enhanced the protective effect of CEF as compare to CEF alone. CEF attenuated the MPTP-induced memory impairments. CEF also averted the MPTP lesion-induced degeneration of DAergic neurons in the nigrostriatal area. It also terminated the microglial activation in the substantia nigra pars compacta (SNc) and cellular loss in the hippocampal CA1 region.	Bisht et al., 2014
Murine model: MPTP-induced PD rat model.	Murine model: MPTP-induced PD rat model.	Male Wistar rats.	The MPTP-injected group received either CEF (200 mg/kg/day) (CEF group; $n = 11$ ) or saline (saline group; $n = 14$ ), while the sham-operated group received saline injections but not MPTP (saline group; $n = 12$ ).	MPTP lesioning-induced impairment of motor function, working memory, and object recognition were prevented by CEF. Administration of CEF also ameliorated behavioral deficits in MPTB-induced PD rat's model. CEF also prevented MPTP-induced neuronal loss and partly promote neurogenesis.	Hsieh et al., 2017
Murine model: MPTP-induced PD rat model.	Murine model: MPTP-induced PD rat model.	Male Wistar rats.	CEF or saline was injected for 15 subsequent days with the regimes of: the sham-operated group injected with saline (1 ml/kg/day, $n = 13$ ), while the MPTP-lesioned rats were subdivided into either saline (1 ml/kg/day, $n = 13$ ), or CEF (5 mg/kg/day, $n = 13$ ) groups.	Rats that received either pre- or post-lesioning with CEF prevented the DAergic degeneration in SNc and striatum and improved working memory and object recognition. Lesioning that could cause neurodegeneration and glutamatergic hyperactivity were markedly suppressed by CEF treatment. CEF received group also displayed an upregulated GLT-1 expression.	Hsu et al., 2015
Murine model: MPTP-induced PD rat model.	Murine model: MPTP-induced PD rat model.	Male Wistar rats.	The sham-operated groups injected with either saline ( $n = 10$ ) or CEF at a dose of 100 ( $n = 11$ ) or 200 mg/kg/day ( $n = 12$ ), while the MPTP-lesioned groups received saline ( $n = 11$ ) or CEF at the dose of 100 or 200 mg/kg daily beginning either on day 5 or day 3.	Memory impairments were prominently decreased or abolished in rats administered either CEF alone or in combination with EPO. The co-administration of the 2 agents (CEF + EPO) showed superior action than individual agents. CEF, EPO, or CEF + EPO also reduced or eliminated MPTP lesioning-induced neurodegeneration.	Huang et al., 2015
Murine model: MPTP-induced Parkinson's disease (PD) rat model.	Murine model: MPTP-induced Parkinson's disease (PD) rat model.	Male Wistar rats.	After MPTP lesioning, animals were injected with daily CEF (5 mg/kg), erythropoietin (100 U/kg), or CEF + erythropoietin (EPO) and undertook the bar-test, T-maze test, and object recognition test. Saline was given both for sham-operated and one of MPTP received groups as control.	The combined administration of these agents also displayed a better outcome in the densities of DAergic terminals. CEF (200 mg/kg) treated rats exhibited substantial betterment of behavioral impairment and oxidative damage. CEF also attenuated the marked upsurge of neuroinflammatory markers including NF $\kappa$ B, TNF- $\alpha$ , and IL-1 $\beta$ in MPTP received groups. CEF significantly recovered the reduced activity of BDNF in MPTP received rats. Pre-treatment of memantine with lower dose of CEF enhanced the protective actions of CEF.	Kaur and Prakash, 2017
Murine model: 6-OHDA lesion-induced PD model.	Sprague-Dawley rats.		The treatment groups were received CEF (200 mg/kg/day) for 1 week prior to lesion surgery while the control groups were administered a 0.9% saline for the same duration.	CEF pre-treatment ameliorated the muscular rigidity and contralateral rotation in 6-OHDA-lesioned rats. CEF treated group also showed a reduction of dopaminergic neuronal deaths. Both protein expression and TH immunoreactivity for GLT-1 were overexpressed in CEF treated rats.	Leung et al., 2012

(Continued)

TABLE 1 | Continued

Models of ND disorders	Experimental approaches	Study subjects	Interventions	Major outcome (s)	References
<i>In vitro</i> study: 6-OHDA exposed PC12 cells.	Neural (PC12) cells.	PC12 cells were treated with varying concentrations of CEF (10–100 µM) alone or along with a 6-OHDA for 24 h. For separate experiment, 100 thousand PC12 cells/cm <sup>2</sup> were plated and treated after 1 day.		CEF interacted with high affinity to α-synuclein and interfered with the <i>in vitro</i> polymerization. PC12 cells pre-treatment with CEF (100 µM) resulted in a down-regulation of α-synuclein expression, but not at 10 µM concentration). CEF at the concentration of 100 µM, showed a substantial recovery of cell apoptosis and restore PC12 viability. Both lower (10 µM) and maximal conc. (~1 mM) failed to recover 6-OHDA-induced apoptosis.	Ruzza et al., 2014
Murine model: MPTP-induced PD rat model.	Male Wistar rats.	Rats were grouped (5 rats/group) into sham-operated and two MPTP-lesioned groups; the sham group received saline, while the MPTP-lesioned groups received either daily saline or CEF (100 mg/kg) for 15 days.		CEF treatment prevented the MPTP-induced decreases in neurogenesis of rats in the dentate gyrus of the hippocampus. CEF treatment also ameliorated the decrease in both neuronal density and activity in the nigrostriatal, hippocampus, and subthalamic nucleus observed in the MPTP-induced rats. Besides, unlike saline received group, CEF treated group showed an intact density of DAergic neurons in the SNc and DAergic terminals in the striatum.	Weng et al., 2016
<i>In vitro</i> study: astrocytes exposed to a neurotoxin, 1-methyl-4- phenyl-pyridinium (MPP <sup>+</sup> ) model.	Cultured primary astrocytes from Sprague-Dawley rat pups.	After cells were cultured, they were randomly divided into (a) control group (received saline); (b) MPP <sup>+</sup> treatment group (received culture medium having MPP <sup>+</sup> ); (c) CEF treatment group (received culture medium having 100 µM CEF); and (d) Co-treatment with MPP <sup>+</sup> and CEF group (received culture medium containing MPP <sup>+</sup> plus 100 µM CEF).		CEF (100 µM) enhanced the expression and uptake of glutamate in astrocytic cells exposed to MPP <sup>+</sup> . CEF also upregulated the expression of GLT-1 and stimulates astrocyte viability after <i>in vitro</i> MPP <sup>+</sup> exposure. CEF Attenuated MPP <sup>+</sup> -induced apoptosis, cytotoxicity and neurotoxicity in primary astrocytes. The cytoprotective actions of CEF is mediated through downregulation of NF-κB and JNK/c-Jun Signaling.	Zhang et al., 2015
Alzheimer's disease (AD).	<i>In vitro</i> and Murine model: triple trans- genic model of AD [3xTg-AD].	3xTg-AD mice; astrocyte and neuron 1 <sup>0</sup> cell culture.	Mice were grouped in to CEF and vehicle treated group: 4 female and 4 male mice received saline (control group) while 3 female and 4 male mice received CEF (200 mg/kg).	Prolonged administration of CEF in aged 3xTg-AD mice showed a substantial upregulation GLT-1 expression, improving cognitive impairments, preserving synaptic proteins and reducing of tau pathology. But both beta amyloid (Aβ) and amyloid precursor protein were not significantly altered in CEF treated group compared to control group.	Zumkehr et al., 2015
Murine model: rat model of accelerated senescence.	Male Wistar and OXYS rats.	Animals were administered daily with either saline (Wistar + saline, n = 10 and OXYS + saline groups, n = 10) or CEF at the doses of 50 (OXYS + CEF50 group, n = 10) or 100 mg/kg (OXYS + CEF100 group, n = 10) for 36 days.		Long-term treatment of CEF in a dose of 100 mg/kg moderately improved movement impairments and recovered the deficit of new object recognition. Both doses of CEF (50 and 100 mg/kg) increased the density of pyramidal neurons in the CA1 area in OXYS rats. A 50 mg/kg doses of CEF also expressively amplified the immunoreactivity of TH in the striatum.	Tikhonova et al., 2017
Genetic murine model: on two different genotypes rats genotype.	Male Wistar rats, OXYS male rats.	Beginning from the age of 14 weeks, animals were administered either daily CEF (100 mg/kg) or saline for 36 days.		CEF treated group showed downregulation of mRNA levels of <i>Bace1</i> (that encodes β-secretase involved in Aβ production) and <i>Ace2</i> (that encodes enzymes involved in Aβ degradation) in the hypothalamus. CEF received group also exhibited diminution of the expression of gene of β-actin ( <i>Aktb</i> ) in the frontal cortex. CEF amplified <i>Mme1, ide</i> (that encode enzymes involved in Aβ degradation). CEF also upregulated the expression of <i>Ebp2</i> mRNA level in the amygdala and the levels of <i>Ece1</i> and <i>Aktb</i> in the striatum.	Tikhonova et al., 2018

(Continued)

**TABLE 1 |** Continued

Models of ND disorders	Experimental approaches	Study subjects	Interventions	Major outcome (s)	References
Murine model: APP/PS1 transgenic AD model.	Swiss albino mice.	Animals were divided in to 6 groups (6 mice/group); group I (vehicle), group II (vehicle + scopolamine), group III (Donepezil + scopolamine), group IV (206 mg/kg of CEF + scopolamine), group V (0.49 mg/kg of selegiline + scopolamine), group VI (206 mg/kg of CEF + 0.49 mg/kg of selegiline + scopolamine).	CEF and selegiline improved scopolamine induced cognitive impairment. Both drugs (CEF and selegiline) showed a substantial memory enhancing ability compared to negative control. Elevated level of acetylcholine esterase (AchE) were attenuated in mice that received either CEF or selegiline. CEF or selegiline received mice also showed improved oxidative status.	CEF and selegiline improved cognitive-in improvement, AchE and antioxidant activities.	Akina et al., 2013
Murine model: APP/PS1 transgenic AD mice model.	APP/PS1 transgenic AD mice.	Animals were assigned: wild type (received normal saline), APP/PS1 (saline) and CEF (100, 200, and 300 mg/Kg) groups. All these groups of the APP/PS1 mice were treated with either the vehicle or CEF once daily for 2 weeks. One additional group was also received 200 mg/Kg CEF after receiving dihydrokainate.	CEF received group substantially enhanced the cognitive impairment in early stage of AD animals. CEF (all the 3 doses) also upregulated GLT-1, glutamine synthetase (GS), and system N-glutamate transporter-1 (SN1) protein expressions in the hippocampus of APP/PS1 AD mice. Dihydrokainate, a selective inhibitor of GLT-1 reversed enhanced memory functioning, GS activity, and SN1 expression of CEF in APP/PS1 AD mice.	CEF received group substantially enhanced the cognitive impairment in early stage of AD animals.	Fan et al., 2018
Murine model: APP/PS1 AD model.	Mice of either APP/PS1 or wild-type mice.	Mice were allocated into CEF treated (APP/PS1 and wild-type mice; 200 mg/Kg CEF for 5 days, $n = 9$ ) and control groups (APP/PS1 and wild-type mice) received vehicle for five subsequent days, $n = 5$ /group).	CEF administered group showed partial restoration of the reduced GLT-1 level in the area around to A $\beta$ plaques. CEF treated mice also displayed a marked decrement of the chronically overexpressed glutamate levels that were identified in the vicinity of the amyloid aggregate. Hence the neurotoxic microenvironment encompassing A $\beta$ aggregates were substantially improved in CEF received group of mice.	The stage 1 investigation showed a linear PK, and CSF trough levels for both low and high dose levels beyond the pre-stated target trough level.	Hafezehi et al., 2016
Amyotrophic lateral sclerosis (ALS).	Sixty-six human subjects having defined ALS.	In Stage 1, participants at 10 clinical sites were randomized into 3 equal study groups receiving either daily placebo or CEF (2 gm or 4 gm in to 2 divided doses). Study groups were continued their allocated treatment in Stage 2.	Tolerability outcomes (Stages 1 and 2) displayed that CEF at dosages up to 4 gm/day was well-tolerated until 20 weeks. Biliary related side effects were more common with CEF, which were dose independent and were overcome with ursodeoxycholic treatment.	The stage 1 investigation showed a linear PK, and CSF trough levels for both low and high dose levels beyond the pre-stated target trough level.	Berry et al., 2013
A multi-phase randomized trial of human-subjects diagnosed with ALS.				In the period of stages 1 and 2, mean ALSFRS-R scores were decreased gradually in participants who administered CEF (4 gm) compared to placebo. Nonetheless in stage 3 both functional decline and survival between groups were not substantially different.	Cuckowicz et al., 2014
Five hundred and fourteen eligible adult patients with ALS.				Handheld dynamometry endpoints in patients who completed the study displayed no noticeable differences between groups. GLT and hepatobiliary associated adverse effects were more common in CEF groups than control group.	(Continued)

**TABLE 1 |** Continued

Models of ND disorders	Experimental approaches	Study subjects	Interventions	Major outcome (s)	References
	<i>in vitro</i> study; from tissue culture.	Mouse embryonic fibroblast, HT22 cell; human motor neurons derived from embryonic stem cells.	Cells were treated either with CEF (30, 100 or 300 $\mu$ M) or vehicles.	Chronic CEF treatment prevent all tested cells against oxidative glutamate toxicity and induces EAAT expression in dose dependent fashion.	Lewerenz et al., 2009
Surgical model; axotomized mice model.		Adult male C57BL/6 mice were used.	Twenty-six axotomized mice were grouped into either drugs (CEF or minocycline) or saline-received groups from the 1st day of hypoglossal axotomy: CEF-administered group (200 mg/kg, $n = 8$ ), minocycline-received group (30 mg/kg, $n = 8$ ), vehicle-administered group (5 ml/kg, $n = 10$ ).	CEF-exhibited a slow upregulation of Nr2 without any pro- or antioxidant activity.	Yamada and Jimmo, 2011
Huntington's disease (HD).	Murine model: R6/2 mice, transgenic model of HD.	Male transgenic R6/2 mice (HD phenotype).	Animals were treated with either CEF (200 mg/kg, or same volume of vehicle (saline) once daily for 5 subsequent days.	Both CEF and minocycline treatment group showed a substantial enhancement of survival rate of lesioned motor neurons. But there are no noticeable differences in the cellular densities of astrocytes in CEF treated and control groups. CEF upregulated the expression of GLT-1 in the hypoglossal nucleus, while it inhibited the reactive increment of the glial protein expression.	Miller et al., 2008
		Male transgenic R6/2 and Wild type mice.	Mice were administered either daily CEF (200 mg/kg) or vehicle for 5 successive days.	CEF treatment in R6/2 mice attenuated various manifestation of HD including a decreased paw clasping and twitching, while motor flexibility and open-field climbing were amplified. Compared to vehicle received group, CEF treated group showed an upregulation of GLT-1 expression in striatum. CEF also restored the glutamate uptake in striatum of R6/2 mice. Upregulation of the functional GLT1 level by CEF attenuated the multiple HD behavioral phenotype.	Sari et al., 2010
				CEF treatment upregulated the expression of cortical and striatal GLT1 level compared to saline treated group . This action is persisted even after GLT1 levels began to decrease when these mice are 13 weeks of age and overtly symptomatic. Therefore, the cellular machinery underlying the CEF- upregulated GLT1 expression possibly working in late-stage HD.	

*MPTP*, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; 6-HODA, 6-Hydroxydopamine; CEF, Ceftriaxone; ALS, Amyotrophic Lateral Sclerosis; ALSFRS-R, Amyotrophic Lateral Sclerosis Functional Rating Scale-Revised; GLT 1, glutamate transporter 1; EAAT2, excitatory amino acid transporter subtype 2; HD, Huntington's disease; CSF, Cerebrospinal fluid; PK, pharmacokinetic; AD, Alzheimer's disease; AchE, Acetylcholine esterase.

models) have also been reported (**Table 2**). In a neuropathic pain model in rats, CEF-treated groups showed a meaningful overexpression of GLT-1 level and glutamate uptake in the spinal dorsal horn (Hu et al., 2010). Another study on a similar model showed that CEF significantly attenuated the production of TNF- $\alpha$  and IL-1 $\beta$  (Amin et al., 2012). CEF mitigated the increased levels of Bax and cleaved forms of caspases 3 and 9, while the expression of Bcl2 was markedly amplified (Amin et al., 2014). CEF inhibited opioid-induced hyperalgesia (OIH) in mice. Further, the beta-lactam antibiotic, CEF reversed the OIH induced downregulation of GLT-1 expression (Chen et al., 2012). Finally, CEF produced marked dose-dependent antihyperalgesic effects in the somatic inflammatory and visceral pain models (Stepanovic-Petrovic' et al., 2014).

## PROTECTIVE EFFECT OF CEF AGAINST BRAIN ISCHEMIA

Various studies reported the protective role of CEF against different models of brain ischemia (**Table 2**). CEF pre-treatment reduced infarct volume and apoptotic index and CEF treatment significantly increased GLT-1 mRNA and protein levels (Mimura et al., 2011). In a rat model of global brain ischemia, it was found that pre-treatment with CEF substantially prevented the delayed neuronal death in the hippocampal CA1 area commonly induced by global brain ischemia (Hu et al., 2015; **Table 2**).

## PROTECTIVE EFFECT OF CEF AGAINST STROKE AND TRAUMATIC BRAIN INJURY

CEF also showed protective activity on animal models of stroke (**Table 2**). CEF strongly reduced infarct size, marked improvement of neuronal survival within the penumbra, and diminished neurological impairment (Thöne-Reineke et al., 2008). Furthermore, CEF was found to produce promising protective effects against different models of brain injury (Cui et al., 2014; **Table 2**).

## EFFECT OF CEF ON ALCOHOL DEPENDENCE AND WITHDRAWAL

Studies evidenced that glutamate transmission is implicated in various features of drug addiction (Sari et al., 2011). The foremost transporter of glutamate, GLT1, in the nucleus accumbens (NAc) and prefrontal cortex (PFC) plays a notable role in alcohol dependence behavior attenuation (Rao and Sari, 2014a). CEF is found to be the most effective  $\beta$ -lactam antibiotic in increasing the expression of GLT1 in the brain (Rothstein et al., 2005). Though CEF attenuated ethanol consumption in P rats, only the relatively higher doses (100 and 200 mg/kg) were associated with marked upregulation of GLT1 expression in the PFC and NAc. This increases in the expression of GLT1 appear to be inversely associated with a post treatment attenuation of ethanol intake (Sari et al., 2011). According to Lee et al. CEF induced indirect upregulation of GLT-1 expression and GSH via the activation of NF- $\kappa$ B, specifically at the NF- $\kappa$ B binding site (272 position)

of the GLT-1 promoter (Lee et al., 2008). Another study has also revealed that CEF treatment improved GSH and xCT (a catalytic subunit of xC $^{-}$ ) levels in PFC, NAc, and associated brain regions (Lewerenz et al., 2009). This CEF-induced increase in the xCT level was found to be correlated with downregulation of extracellular levels of glutamate (Sari et al., 2011), which is the very reason for alcohol dependence. On the other hand, upregulation in the levels of xCT and subsequent upsurges of GSH level may be one possible way to reverse the GLT impairments caused by free radical overproduction. Ethanol withdrawal is related to an increment of oxygen-derived free radicals (Labra Ruiz et al., 2018), which in turn have been shown to inhibit glutamate uptake by oxidation of thiol groups (Volterra et al., 1994).

During CEF treatment in ethanol dependence, an increase in water intake was observed that could be due to the compensatory effect of water to the decrease in ethanol intake (Sari et al., 2011; **Table 3**). On the other side, GLT1 and ENT1 were inversely affected as a consequence of ethanol consumption and this suggests that the neuroadaptative mechanisms are involving these proteins in both NAc core and shell (Sari et al., 2013).

Alcohol withdrawal syndrome is a medical emergency which is related to significant mortality rates (Campos et al., 2011). CEF administration was found to reduce or almost completely abolish all manifestations of ethanol withdrawal in P and Wistar rat variants and prevented withdrawal-induced escalation of alcohol intake. CEF treatment was also associated with long-term upregulation of excitatory amino acid transporter subtype 2 (EAAT2) in the striatum that was downregulated by ethanol withdrawal (Abulseoud et al., 2014). CEF might therefore be used as a potential therapeutic treatment for the attenuation of relapse-like ethanol-drinking behavior (Alajaji et al., 2013; Qrunfleh et al., 2013; **Table 3**).

## EFFECT OF CEF ON NICOTINE WITHDRAWAL AND NICOTINE-INDUCED REINSTATEMENT

Tobacco use is the foremost cause of premature mortality in the United States and around the globe. Nicotine is the primarily responsible component for addiction and related behaviors (Castane et al., 2005). Different studies suggested that nicotine-induced adaptations in glutamatergic neurotransmission play an important role in the development of nicotine dependence (Liechti and Markou, 2008). With this regard, CEF administration reversed nicotine withdrawal manifestations and mitigated nicotine-primed reinstatement of nicotine-conditioned place preference (CPP) without affecting acquisition (Alajaji et al., 2013; **Table 3**).

## EFFECT OF CEF ON ACUTE COCAINE-EVOKED DOPAMINERGIC NEUROTRANSMISSION

CEF exerts its prominent effects on the dopaminergic system in addition to its effects on the glutamatergic system. It decreased

**TABLE 2 |** Effect of CEF against ischemia, pain, traumatic brain injury, and stroke.

Model of disorders	Experimental approach	Study subjects	Interventions	Major treatment outcomes	References
Ischemia	Murine model; focal hypoxic-injury (H-I) induced ischemia in rats.	Male and female Sprague Dawley rats.	Animals were randomly divided and pretreated with CEF (200 mg/kg), minocycline (45 mg/kg), erythromycin (25 mg/kg) and equal volume of saline for 5 subsequent days. Then, rats underwent hypoxic-ischemic (H-I) or sham operated procedure.	CEF treatment meaningfully increased the expression of GLT-1 mRNA and protein levels; CEF pretreated animals showed a reduction of infarct size and apoptotic index. CEF treated animals also showed increment in microtubule-associated protein 2-positive area, while decreased the TUNEL-positive cells.	Minura et al., 2011
	Murine model; focal ischemic cortical lesions in rats.	Male Long-Evans hooded rats.	Animals were allocated to receive CEF at a dose of 200 mg/kg ( $n = 12$ ) or vehicle ( $n = 11$ ) daily for 5 days. Then, the effect of CEF on motor skill learning and rehabilitative training-induced functional enhancement after ischemic lesions was assessed.	In normal animals, CEF failed to influence the skill learning rate or final level of reaching performance. After rats underwent ischemic lesion, CEF exacerbated initial deficits in reaching performance. CEF also affected acquisition of the rehabilitative training task.	Kim and Jones, 2013
	Murine model; cortical vein occlusion by photochemical thrombosis in rats.	Male Wistar rats.	Animals were assigned to receive CEF (100 or 200 mg/kg), vehicle or vehicle or CEF together with GLT-1 inhibitor, Dihydrokainate (DHK) for 5 days prior to venous ischemia.	There is no significant difference in lesion size between CEF and vehicle received groups.	Inui et al., 2013
	Murine model; rat model of global brain ischemia (GB).	Male Wistar rats.	The animals were randomly assigned to sham group, CEF group (50, 100, and 200 mg/kg). The CEF was administered as control, pre- and post-treatment groups.	CEF treated animals showed to decrease infarct volume compared to vehicle received group. The effect of CEF was reduced by administration of the GLT-1 inhibitor, DHK. This suggests that GLT-1 is essential for the observed actions of CEF; the effect of CFT on NTs receptor density did not significantly differ from vehicle treated rats.	Hu et al., 2015
	Murine model; global brain ischemia/ reperfusion (I/R) injury in rats.	Wistar-albino rats.	Animals were allocated in to control ( $n = 10$ ), I/R ( $n = 10$ ), and I/R-CEF ( $n = 10$ , 100 mg/kg CEF 2 h before I/R) groups.	CEF pre-treatment expressively prevented delayed neuronal death of pyramidal neurons in hippocampal CA1 area induced by GBI in dose-dependent fashion. Pre-administration with CEF also up-regulated the expression and glutamate uptake of GLT-1. Inhibitors of GLT-1, DHK significantly reduced CEF-induced up-regulation of GLT-1 and its neuroprotective effect against global ischemia.	Altas et al., 2013
	Murine model; focal cerebral ischemia in rats.	Male Sprague Dawley rats.	Animals were randomly allocated to sham group ( $n = 12$ ), control group ( $n = 30$ ), and CEF (200 mg/kg) pretreated group ( $n = 30$ ) for 5 days.	The levels of MDA was meaningfully decreased, while increased the activity of SOD and GSH in the I/R-CEF group compared with the I/R and control groups. CEF provided morphological improvement of histopathological (microvessel and neuron structure) analysis of brain tissue than the I/R group; there were no significant differences observed in the level of NO among the groups.	Lujia et al., 2014

(Continued)

TABLE 2 | Continued

Model of disorders	Experimental approach	Study subjects	Interventions	Major treatment outcomes	References
Murine model: focal cerebral ischemia in rats.	Male Wistar rats.	Animals were assigned to receive CEF (200 mg/kg), n-acetyl cysteine (NAC) (150 mg/kg), or saline for 5 subsequent days.		CEF meaningfully reduced infarct size and improved neurological deficits induced by a middle artery cerebral occlusion.	Kiryzanowska et al., 2016
Murine model: neonatal rat model of hypoxic-ischemic encephalopathy (HIE).	Neonatal Sprague Dawley (SD) rats.	Neonatal rats were administered either of CEF (50, 100, 200 mg/kg) or saline 2 days before experimental HIE.		CEF treated group showed an enhanced level of GLT-1 expression in the dorsal striatum. In ischemic rats, CEF also upregulated the expression of xC- mRNA in frontal cortex and dorsal striatum. CEF not NAC treated group also upregulate the GLT-1 expression in the frontal cortex and dorsal striatum. CEF treated group also decreased the extent of apoptotic cells in the hippocampus.	Lai et al., 2011
Pain	Male Sprague Dawley rats.	Rats were allocated to receive vehicle or CEF (200 mg/kg) the CEF group were either administered with CEF (200 mg/kg) once daily for 7 days beginning immediately after CCI starting on postoperative day (day 9).		Pre-treatment with CEF (200 mg/kg) substantially attenuated the scores of neonatal rats' brain injury. CEF treated group also decreased the extent of GLT-1 expression in the cortical neurons.	Hu et al., 2010
Murine model: chronic constrictive nerve injury (CCI) in rats.				Both preventive and therapeutic CEF administration showed an up-regulation of GLT-1 expression and glutamate uptake; the action was suppressed by GLT-1 blocker.	
Murine model: opioid-induced hyperalgesia (OIH) in mice.	Male ICR mice.	Animals received morphine sulphate (20 mg/kg) BID for 3 days and two more injections of 40 mg/kg morphine sulfate on 4th day. Mice were received CEF (200 mg/kg) before morphine injections and continued for 1 week.		Pre-treatment of CEF prevented the development of mechanical allodynia and thermal hyperalgesia induced by CCI (anti-nociceptive effects). Therapeutic administration of CEF also significantly increased the latency of thermal withdrawal and the threshold of mechanical withdrawal. The preventive or therapeutic injection of CEF displayed an inhibiting effect on the development and maintenance of mechanical allodynia.	Chen et al., 2012
Murine model: spinal nerve ligation induced neuropathic pain in rats.	Male SD rats.	Animals were allocated to receive control, pioglitazone (5, 10, or 20 mg/kg), CEF (100 or 200 mg/kg) alone or combination for 28 days.		CEF administration exhibited prevention of OIH expression brought by OIH. It also alleviated the mechanical allodynia, and thermal hyperalgesia induced by repeated administration of opioid.	Pottabathini et al., 2016
				CEF also recovered the downregulation of GLT-1 expression brought by OIH.	
				CEF (200 mg/kg) administration substantially improved the paw withdrawal threshold.	
				Treatment with CEF monotherapy (200 mg/kg) expressively prevented the behavioral, biochemical, mitochondrial and cellular alterations.	
				CEF (200 mg/kg) also significantly attenuated the TNF- $\alpha$ , IL-6 and caspase-3 activities.	

(Continued)

TABLE 2 | Continued

Model of disorders	Experimental approach	Study subjects	Interventions	Major treatment outcomes	References
Murine model; chronic constriction injury model (CCI) in rats.	Male Wistar rats.	Animals were assigned to receive control, CEF (100, 150, and 200 mg/kg) or minocycline (25, 50, and 100 mg/kg) alone or combination of CEF and minocycline for 1 week.	CEF produced a dose-dependent reversal effects of the neuropathic pain behaviors. Administration of CEF (200 mg/kg) significantly attenuated CCI induced production of TNF- $\alpha$ and IL-1 $\beta$ than control group.	The highest dose of CEF (200 mg/kg) attenuated tactile allodynia in CCI rats.	Amin et al., 2012
Murine model; somatic Inflammatory Hyperalgesia in rats and Visceral Nociception in mice.	Male Wistar rats and Swiss Webster mice.	Animals were allocated to control, CEF (10–200 mg/kg) alone and in combinations with various analgesic drugs in carrageenan-induced paw inflammatory hyperalgesia and in the acetic acid-induced writhing test for 7 days.	Pre-treatment with CEF for 1 week showed a significant analgesic action in the somatic inflammatory model in dose-dependent manner. Pre-administration of CEF also exhibited a substantial antinociceptive effect in the visceral pain model in dose-dependent fashion.	Combination of CEF and different analgesics showed a synergistic reduction of hyperalgesia and nociception in both somatic and visceral inflammatory pain than separated drugs used.	Stepanovic-Petrovic et al., 2014
Murine model; chronic constriction injury (CCI) in rats.	Male Wistar rats.	Animals were randomly allocated into saline, sham-operated and CEF (200 mg/kg) for 1 week.	COIs that received CEF showed a remarkable upregulation in the levels of antiapoptotic protein, Bcl2, and decreased the contents of Bax protein. CEF also attenuated the cleaved forms of caspases 3 and 9.	CEF treated groups were also protected from the CCI-brought oxidative stress.	Amin et al., 2014
Murine model; mechanical allodynia and hyperalgesia through STZ induced neuropathic pain.	Male Wistar rats.	Rats were injected either daily CEF (50–200 mg/kg) or control for 1 week.	CEF increased in the paw withdrawal thresholds. The GLT-1 transporter inhibitor, DKA, recovered anti-allodynic and anti-hyperalgesia actions of CEF. The highest dose of CEF (200 mg/kg) significantly reduced both mechanical allodynia and hyperalgesia.	CEF treatment also attenuated both mechanical allodynia and modulated the expression of GLAST and GFAP.	Gunduz et al., 2011a
Murine model; radicular pain through dorsal nerve root compression in rats.	Male Holtzman rats.	Animals were randomly allocated to receive either saline or CEF (40 $\mu$ l of 10 $\mu$ g intrathecal injection).	CEF injection also restored the reduced spinal astrocytic activity, and neuronal hyperexcitation in the spinal dorsal horn.	CEF administration showed a significant upregulation of GLT 1 expression.	Nicholson et al., 2014
Murine model; peripheral pain induced by injections of formalin; neuropathic pain using the spinal nerve ligation (SNL).	Male Sprague Dawley rats.	Animals were received either CEF (200 mg/kg) or same volume of saline.	CEF significantly increased both mechanical and thermal withdrawal threshold in naïve rats. After induction of neuropathic pain by SNL, effects of CEF was substantial in reducing thermal hyperalgesia but failed to show significant effect on mechanical allodynia.	CEF also delayed the intensity of painful behaviors in response to formalin induced inflammatory hyperalgesia.	Elijaja et al., 2011

(Continued)

TABLE 2 | Continued

Model of disorders	Experimental approach	Study subjects	Interventions	Major treatment outcomes	References
Murine model; experimental autoimmune encephalomyelitis (EAE) and chronic constriction nerve injury (CCI) models.	Male Sprague Dawley rats and male Dark Agouti rats.	Animals were received either saline or CEF (150 µg intrathecally).		Pre-administration of CEF attenuated the development of hyperalgesia and allodynia in response to repeated morphine. In experimental EAE, CEF displayed tactile allodynia and arrested the progression of motor weakness and paraparesis. Similarly, CEF recovered tactile allodynia induced by CCI.	Ramos et al., 2010
Murine model; chronic constriction nerve injury (CCI) in rats.	Male Wistar rats.	Rats were allocated to receive CEF (100, 200 or 400 mg/kg), clavulanic acid (CLA) (0.1, 1, or 10 ng/kg) gabapentin (10 mg/kg as positive control) or saline.		Both CEF and CLA produced antiallodynic effects in response to mechanical and cold stimulation in dose dependent fashion. The antiallodytic effect of CEF was antagonized by haloperidol and naloxone, which indicates CEF may be mediate its antiallodynic actions through dopamine and opioid receptors. Moreover, the plasma TNF- $\alpha$ levels were attenuated following the highest dose of CEF (200 mg/kg) and CLA (10 mg/kg) treatment.	Ochoa-Aguilar et al., 2017
Double-blind randomized trials and Murine model.	Forty five human subjects and male C57/ Black mice.	Patients were randomly allocated to receive an IV infusion of saline ( $n = 15$ ), CEF (2 gm) ( $n = 15$ ), or cefazolin (2 g) ( $n = 15$ ) 1 h prior to surgery. Similarly, in the animal model of postoperative pain, mice were assigned to receive a single dose of saline, CEF (200 mg/kg), or cefazolin (200 mg/kg).		A single dose CEF produced a substantial analgesia in all patients. The whole patients responded to CEF and about 10-fold increment in the threshold of nociception. A single injection of CEF exhibited a substantial antinociceptive effects in mouse models of inflammatory or postsurgical pain. CEF also increased the expression of GLT-1 (2-fold) in the spinal cord.	Macaluso et al., 2013
Traumatic Brain Injury (TBI).	Male Sprague Dawley rats.	Animals were randomly assigned to sham group ( $n = 30$ ), TBI group ( $n = 60$ ) and TBI treated CEF group ( $n = 60$ , 200 mg/kg). CEF was injected for 5 days starting just after TBI and both sham and TBI groups administered equal volumes of saline.		Daily administration of CEF attenuated TBI-induced brain edema and cognitive impairments in rats. CEF treated group restored TBI induced downregulation of GLT-1 expression. CEF also revealed a substantial suppression of autophagy marker protein, LC3 II, in hippocampus than the TBI group.	Cui et al., 2014
Murine model; rat lateral fluid percussion injury TBI model.	Male Long-Evans rats.	Animals were divided in to "saline-sham," "saline-TBI," and "CEF-TBI" (CEF, 200 mg/kg) groups and treatment continued daily for 7 days after TBI surgery.		CEF infection reduced the level of regional glial fibrillary acid protein (GFAP) expression (43%) in the lesioned cortex. CEF received group also showed a notable attenuation of cumulative post-traumatic seizure duration. The reduced level of GLT-1 expression after TBI was restored by administration of CEF.	Goodrich et al., 2013

(Continued)

TABLE 2 | Continued

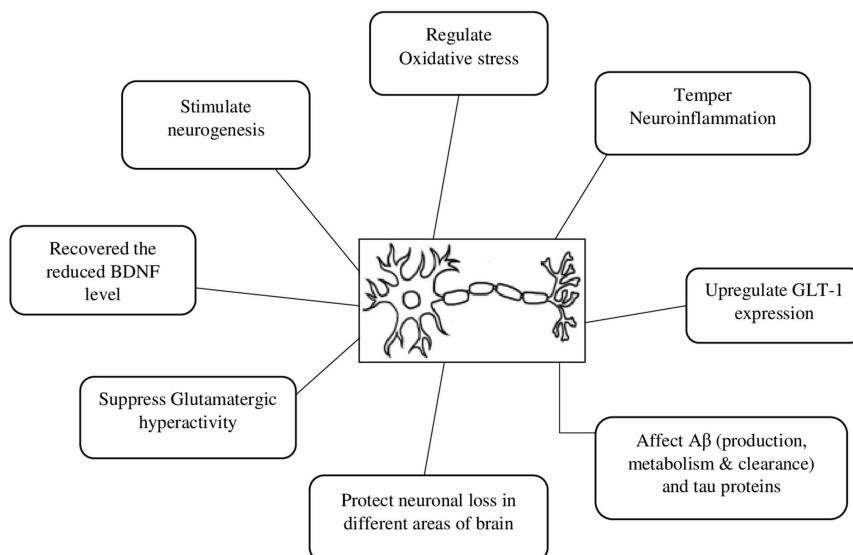
Model of disorders	Experimental approach	Study subjects	Interventions	Major treatment outcomes	References
Murine model: TBI model in rats.	Sprague Dawley rats.	Rats were randomly allocated to sham-operated group ( $n = 18$ ), trauma group (TBI, $n = 27$ ) and trauma + CEF treatment group (TBI + CEF, $n = 27$ ). After TBI, CEF (200 mg/kg) was administered in TBI + CEF group just after trauma, while both sham and TBI groups received only vehicle.		CEF treated group showed an attenuation of TBI-induced cerebral edema and functional cognitive impairments. CEF administration also decreased the levels of proinflammatory cytokines interleukin-1 $\beta$ , interferon- $\gamma$ , and TNF- $\alpha$ . It also up-regulated the expression of GLT-1 level after TBI.	Wei et al., 2012
<i>in vitro</i> and Murine model: a rat model of subarachnoid hemorrhage (SAH).	Male Sprague Dawley (SD) rats and Primary astrocyte cultures.	Animals were assigned randomly to sham, CEF (SAH + 50 or 100 mg/kg) and vehicle (SAH + saline) groups.		CEF meaningfully alleviated the SAH-induced cognitive deficits in spatial learning memory and reference memory. CEF administered rats showed a reduction of hippocampal neuronal apoptosis following SAH. CEF treated animals also reversed the downregulation of EAAT2 expression expressively following SAH. CEF also enhanced the nuclear translocation of p65 and the activation of Akt in hippocampal astrocytes. CEF received group displayed a substantial reduction of early mortality (from 34.5 to 0%). CEF treatment also meaningfully decreased the infarct size and improved neuronal survival within the penumbra. CEF further ameliorated neurological dysfunctions, which led to an overexpression of neurotrophins in the peri-infarct area and also upregulated the expression of GLT-1 level.	Feng et al., 2014
Stroke	Male normotensive Wistar rats.	Rats were randomly allocated to receive vehicle (sham, $n = 8$ ), CEF (sham, $n = 13$ ); CEF (200 mg/kg), vehicle [stroke, $n = 29$ , CEF (stroke, $n = 19$ ; 200 mg/kg)].		Acute hippocampal slices obtained from rats and organotypic hippocampal slices treated with either vehicle or CEF for 5 days (200 mg/kg.) and exposed to oxygen-glucose deprivation.	Lipski et al., 2007 Thöre-Reinke et al., 2008
<i>In vitro</i> model of stroke.	Hippocampal slices obtained from adult rats and organotypic hippocampal cultures.			CEF treated group delayed the occurrence of oxygen-glucose deprivation-induced hypoxic spreading depression (neuroprotective effect). The glutamate-induced NMDA currents from CA1 pyramidal neurons showed a greater enhancement of these currents in CEF-treated groups. But pre-treatment of slice cultures with CEF (10–200 $\mu$ M for 5 days) failed to affect either the 1 <sup>0</sup> (direct) or 2 <sup>0</sup> (delayed) damage of CA1 pyramidal neurons brought by glutamate and CEF exposure also did not increase GLT-1 expression.	

CCl, Chronic constriction nerve injury; CEF, Ceftriaxone; GLT-1, Glutamate transporter 1; HI, Hypoxic-injury; I/R, Ischemia/epertusion; SAH, subarachnoid hemorrhage; TBI, Traumatic brain injury; EAE, experimental autoimmune encephalomyelitis; TNF- $\alpha$ , Tumor necrosis factor-alpha; OH, Opioid-induced hyperalgesia; TUNEL, Transferase-mediated deoxy-uridine triphosphate nick end labelling; SNL, Spinal nerve ligation; EAE, experimental autoimmune encephalopathy;

**TABLE 3 |** Effect of CEF on different types drug/alcohol of dependencies and withdrawals.

Type of dependence/ Withdrawal	Model Used	Animals used (species, sample)	Intervention	Main outcome (s) upon CEF administration	References
Ethanol dependence	Murine model: animal model of alcoholism	Alcohol-prefering (P) rats	25, 50, 100, or 200 mg/kg CEF (CEF, i.p.)	Significant reduction in daily ethanol consumption. Dose-dependent increases in water intake. Body Weight did not affect. Not reduction in sucrose. Increases in GLT1 expression levels within the PFC and NAC with the large dose.	Sari et al., 2011
	Murine model: animal model of alcoholism.	Alcohol-prefering (P) rats.	100 mg/kg CEF (i.p.).	Significant reduction in daily ethanol consumption. Dose-dependent increases in water intake. Body Weight did not affect. Increased GLT1 level in the NAC and its shell.	Sari et al., 2013
	Murine model: animal model of alcoholism.	Alcohol-prefering (P) rats.	100 mg/kg CEF (CEF, i.p.).	Significant reduction in daily ethanol consumption. Dose-dependent increases in water intake. Body Weight did not affect. Increases in GLT1 expression levels within the PFC and NAC with the large dose.	Rao and Sari, 2014a,b
Ethanol withdrawal.	Murine model: model of ethanol withdrawal.	Adult male P rats and Wistar rats.	CEF [50 or 200 mg/kg; i.p.].	Completely abolished all manifestations of ethanol withdrawal. Prevented withdrawal-induced escalation of alcohol intake. Upregulation of EAAT 2 in the stratum.	Abulseoud et al., 2014
	Murine model: animal model of alcohol dependence.	Alcohol-prefering (P) rats.	CEF [50 or 100 mg/kg] was administered.	CEF treatment attenuates relapse-like ethanol-drinking behavior. Upregulation of GLT1 level in prefrontal cortex and NAC. CEF has no effect on relapse-like sucrose-drinking behavior.	Qrunfleh et al., 2013
Nicotine withdrawal and nicotine-induced reinstatement.	Murine model: a nicotine-conditioned place preference.	Naïve male 8- to 10-week-old ICR mice.	CEF (200 mg/kg, twice per day) for 4 consecutive days.	CEF did not disrupt the acquisition of nicotine CPP. CEF blocked nicotine-primed reinstatement of nicotine CPP. Physical and affective precipitated nicotine withdrawal signs are attenuated by CEF. Repeated CEF failed to enhance the low dose of nicotine (0.5 mg/kg) in the tail flick or the hot plate assays or body temperature assessments.	Alajaji et al., 2013
Cocaine-seeking relapse.	Murine model: cocaine self-administration.	Rats.	CEF (200 mg/kg IP) for 7 days.	CEF restored GLT-1 and xCT levels and prevented cue- and cocaine-induced reinstatement of drug-seeking behavior.	Knackstedt et al., 2011
Cannabinoid tolerance	Murine model: cannabinoid tolerance.	Male Balb-c albino mice.	CEF, with its higher doses (100–200 mg/kg),	Attenuated the development of tolerance to the analgesic and hypnotic. Had no effect on its cataleptic action. Upregulated the GLT-1 expression.	Gunduz et al., 2011b
Morphine-induced dependence and Tolerance.	Murine model: morphine-induced dependence and Tolerance.	Adult male Albino mice.	CEF [50, 100, and 200 mg/kg].	Attenuated the development of tolerance to the antinociceptive effect. Reduced naloxone-precipitated withdrawal jumping and standing on feet.	Habibi-Asl et al., 2014

CEF, ceftriaxone; GLT 1, glutamate transporter 1; NAC, nucleus accumbens pars compacta; EAAT 2, excitatory amino acid transporter 2; i.p., intraperitoneally.



**FIGURE 1 |** Possible neuroprotective mechanisms of ceftriaxone. A $\beta$ , beta amyloid protein; BDNF, brain-derived neurotrophic factor; GLT 1, Glutamate transporter 1.

the negative effects of cocaine on locomotor and dopamine-activities. This effect was not responsive to a glutamate re-uptake blocker but rather affected some of the principal regulatory components of dopamine transmission in the NAc (Barr et al., 2015; **Table 3**).

## EFFECT OF CEF ON CANNABINOID TOLERANCE

CEF prevented the development of tolerance to the analgesic and hypothermic actions of WIN 55,212-2 (cannabinoid receptor agonist) by increasing glutamate uptake. Even though the mechanism is not clear, stimulation of GLT-1 by CEF is one of the suggested mechanisms in preventing the development of tolerance to cannabinoids (Gunduz et al., 2011b; **Table 3**).

## EFFECT OF CEF ON MORPHINE-INDUCED DEPENDENCE

Morphine-induced tolerance and dependence were diminished when mice were pretreated with amitriptyline and CEF alone or in combination. Moreover, the co-administration of low doses of CEF (50 mg/kg) and amitriptyline (5 mg/kg) significantly attenuated morphine-induced dependence compared to the administration of CEF or amitriptyline alone. This enhanced effect could be due to the additive or synergistic effect of CEF and amitriptyline on morphine-induced drug dependence (Habibi-Asl et al., 2014; **Table 3**).

## ANTIDEPRESSANT EFFECTS OF CEF

For a long time, depressive disorder (DD) was thought to be attributed mainly to biochemical alterations of the monoamines and their receptors (Labra Ruiz et al., 2018). A majority of

the currently available antidepressants were developed based on this hypothesis and act by increasing the availability of the monoamines including norepinephrine, dopamine, and serotonin. This is effected either by preventing the metabolism of these neurotransmitters or by blocking the transporter mediated reuptake of these neurotransmitters (Delgado, 2000; Fiedorowicz and Swartz, 2004).

Moreover, DD is thought to be related to excessive glutamatergic neurotransmission. In such cases increased extracellular concentrations of glutamate were observed in several brain regions (Lowy et al., 2002). In another way, GLT1 expression is elevated as a compensatory mechanism to overcome the increased stress-induced glutamate release (Delgado, 2000). Recent evidence revealed that the  $\beta$ -lactam antibiotic, CEF enhanced the uptake of glutamate via the up-regulation of GLT1, suggesting its antidepressant effect. CEF treated animals also showed a marked decrement of immobility in the forced swim and tail suspension tests. Even though not statistically significant, a similar trend was noted in novelty-suppressed feeding (Mineur et al., 2007; Borah et al., 2018). A more recent study also showed that the antidepressant effect of CEF was comparable to that of fluoxetine in the tail suspension test in a dose-dependent manner (Borah et al., 2018).

## EFFECTS OF CEF IN SEIZURE MODULATION

It was assumed that oxidative stress and reactive oxygen species (ROS) produced an important role in the progression of epileptic-seizures because they gradually disrupt the intracellular calcium homeostasis, which leads to neuronal loss (Chang and Yu, 2010). A study revealed that pre-treatment of animals with CEF provided meaningful protective actions against pentylenetetrazole (PTZ)-induced generalized clonic seizure,

generalized tonic-clonic seizure, and convulsion-associated deaths (Jelenkovic et al., 2008).

A similar study also showed that CEF treatment substantially improved PTZ-induced convulsions and caused a noticeable modulation of oxidative stress indicators and Connexin 43 expression in the CA3 region of the hippocampal area. CEF also prominently reduced tonic-clonic convulsions and duration of these convulsions and prolonged the latency time in the PTZ-kindling model (Hussein et al., 2016).

## THE POSSIBLE MECHANISM OF ACTIONS OF CEF IN NEUROLOGICAL DISORDERS

After induction of different models of neurological illness by lesioning or chemicals, it is known that animals manifest behavioral alterations in early onset and in the long-term they might also exhibit memory deficits (Van Dam and De Deyn, 2011; Savio et al., 2012; More et al., 2016). Numerous studies evidenced that CEF substantially improved such chemical-induced behavioral alteration and memory impairments in different rodent models of neurological conditions (Lai et al., 2011; Wei et al., 2012; Akina et al., 2013; Chotibut et al., 2014; Feng et al., 2014; Hsieh et al., 2017; Kaur and Prakash, 2017; Fan et al., 2018).

It is well-known that an abnormally increased level of glutamate (Glu) in the brain can induce neuronal damage and excitotoxicity that contributes to the pathogenesis of various neurological disorders including epilepsy, ALS, cerebral ischemia, parkinsonism, and AD (Massie et al., 2010; Annweiler et al., 2014; Kleteckova et al., 2014). There are different glutamate transporters involved in terminating glutamatergic transmission and physiological actions (Kanai et al., 2013; Divito and Underhill, 2014). The pre-synaptic glutamate transporter, GLT-1, clears most of the glutamate released in the cortex and hippocampus (Scofield and Kalivas, 2014). Furthermore, GLT-1 is one of the most common transporters and consists of about 80% of the glutamate transporters expressed in the hippocampus (Mookherjee et al., 2011). There is also evolving evidence suggesting a blockade of certain Glu receptors and/ or enhancing the expression of GLT-1 shown to improve neurological outcomes in various experimental models of neurological illnesses (Zlotnik et al., 2008, 2009; Wang et al., 2014; Bai et al., 2016). Among different agents tested, CEF is one of the beta-lactam antibiotics reported to have neuroprotective actions.

Several studies have revealed that the mechanism of action behind the beneficial effect of CEF in neurodegenerative diseases like PD (Leung et al., 2012; Chotibut et al., 2014, 2017; Hsu et al., 2015; Zhang et al., 2015), AD (Zumkehr et al., 2015), HD (Miller et al., 2008; Sari et al., 2010), and ALS (Yamada and Jinno, 2011) is mediated by enhancing the expression of GLT-1 mRNA levels. Similarly, the mechanism behind CEF's analgesic potential has been associated with the upregulation of the pre-synaptic GLT expression (Ramos et al., 2010; Gunduz et al., 2011a; Chen et al., 2012; Macaluso et al., 2013; Nicholson et al., 2014).

The upregulation of GLT-1 expression is associated with the significant attenuation of the neuronal damage caused by brain

ischemia, and the overexpression of GLT-1 in the ischemic cortex is associated with a reduction of the size of the lesion and improves behavioral and cognitive recovery (Harvey et al., 2011). Studies also showed the neuroprotective actions of CEF in different models of ischemia (Lai et al., 2011; Mimura et al., 2011; Inui et al., 2013; Hu et al., 2015; Krzyzanowska et al., 2016), TBI (Goodrich et al., 2013; Cui et al., 2014; Feng et al., 2014), and stroke (Lipski et al., 2007; Thöne-Reineke et al., 2008) supposedly via an increased expression of GLT-1.

Abnormal regulation of glutamatergic neurotransmission, due to an excessive amount of Glu in brain reward circuitry, has been involved in both initiation and expression of addiction to the drug of abuse related behavior (Kalivas et al., 2009; D'Souza, 2015; Liu et al., 2017). It has also been demonstrated that the expression of the catalytic subunit of  $\text{xC}^-$ , xCT, and GLT-1 are reduced in the NAc following substance of abuse (Massie et al., 2015; Roberts-Wolfe and Kalivas, 2015). From various preclinical studies, it has been reported that CEF markedly attenuates alcohol/drug-seeking behavior and drug clued-up recall while restoring the reduced levels of both xCT as well as GLT-1 (Knackstedt et al., 2011; Sari et al., 2011, 2013; Qrunfleh et al., 2013; Abulseoud et al., 2014).

Long-lasting neuroinflammation and oxidative stress are the other pathological processes involved in brain aging and neurodegeneration (Lee et al., 2008; Mishra et al., 2016; Shah et al., 2016). It has been reported that neuroinflammation is a key player in various neurological disorders, including neurodegenerative illnesses and CNS injury (Chen et al., 2016; Yang and Zhou, 2018). Hence, controlling of the neuroinflammation and excitotoxicity are supposed to limit the abnormal alterations and progression of different neurological disorders (Zhou and Hu, 2013; Yimer et al., 2019), which is further supported by evidence reported, where the use of anti-inflammatory agents or targeting inflammatory markers prominently prevented or at least reduced the progression of neurological disorders (Gagne and Power, 2010; Decourt et al., 2016). Fortunately, pre-treatment of CEF showed attenuation of proinflammatory mediators including NF- $\kappa$ B, IL-1 $\beta$ , INF- $\gamma$ , and/ or TNF- $\alpha$  in various models of neurological disorders such as PD (Kaur and Prakash, 2017), TBI (Wei et al., 2012), neuropathic pain (Pottabathini et al., 2016), and cerebral ischemia (Lujia et al., 2014), which might contribute its own share of reported neuroprotective actions of CEF.

In addition to neuroinflammation, oxidative stress, and mitochondrial dysfunction have also been involved in the development and progression of a wide range of neurodegenerative and mental disorders (Tobe, 2013; Rossignol and Frye, 2014; Yimer et al., 2019). There were also some reports suggesting that antioxidant therapy moderated the production of reactive oxygen species and prevented downstream pathologies of certain neurological diseases (Uttara et al., 2009; Du et al., 2017). Similarly, there are reports that animal models of different neurological disorder received CEF, which attenuated oxidative stress by either reducing oxidative markers such as MDA or by improving the endogenous antioxidant actions such as GSH and SOD (Lewerenz et al., 2009; Altaş et al., 2013).

BDNF is a neurotrophic factor which plays a crucial role in neuronal survival, neurogenesis, and plasticity. It has also been reported that abnormal BDNF expression is involved in several neurological illnesses. There are few studies that report CEF caused upregulation and restoration of reduced BDNF levels in certain animal models (Lee et al., 2008; Kaur and Prakash, 2017).

Besides, CEF might exert its neuroprotective actions through other mechanisms such as by affecting A $\beta$  and tau protein metabolism and clearance in an AD model, prevent polymerization of  $\alpha$ -synuclein in DLB (Ho et al., 2018) and PD (Ruzza et al., 2014; Tikhonova et al., 2018) models, which is of course, a call for further research to substantiate that the neuroprotective actions of CEF is also mediated via these important pathological proteins.

Generally, it appears that the shared mechanism of actions of CEF in the range of various neurologic disorders is through the upregulation of GLT-1 expression and the reduction of proinflammatory mediators and oxidative stress. However, this requires further investigation in order to verify the suggested and possible other newer neuroprotective mechanisms of CEF for each neurological ailment.

## REFERENCES

- Abulseoud, O. A., Camsari, U. M., Ruby, C. L., Kasabeh, A., Choi, S., and Choi, D. S. (2014). Attenuation of ethanol withdrawal by ceftriaxone-induced upregulation of glutamate transporter EAAT2. *Neuropsychopharmacology* 39, 1674–1684. doi: 10.1038/npp.2014.14
- Akina, S., Thati, M., and Puchchakayala, G. (2013). Neuroprotective effect of ceftriaxone and selegiline on scopolamine induced cognitive impairment in mice. *Adv. Biol. Res.* 7, 266–275. doi: 10.5829/idosi.abr.2013.7.6.75119
- Alajaji, M., Bowers, M. S., Knackstedt, L., and Damaj, M. I. (2013). Effects of the beta-lactam antibiotic ceftriaxone on nicotine withdrawal and nicotine-induced reinstatement of preference in mice. *Psychopharmacology* 228, 419–426. doi: 10.1007/s00213-013-3047-3
- Altas, M., Meydan, S., Aras, M., Yilmaz, N., Ulutas, K. T., Okuyan, H. M., et al. (2013). Effects of ceftriaxone on ischemia/reperfusion injury in rat brain. *J. Clin. Neurosci.* 20, 457–461. doi: 10.1016/j.jocn.2012.05.030
- Amin, B., Hajhashemi, V., Abnous, K., and Hosseinzadeh, H. (2014). Ceftriaxone, a Beta-Lactam antibiotic, modulates apoptosis pathways and oxidative stress in a rat model of neuropathic pain. *Biomed. Res. Int.* 2014:937568. doi: 10.1155/2014/937568
- Amin, B., Hajhashemi, V., Hosseinzadeh, H., and Abnous, K. (2012). Antinociceptive evaluation of ceftriaxone and minocycline alone and in combination in a neuropathic pain model in rat. *Neuroscience* 224, 15–25. doi: 10.1016/j.neuroscience.2012.07.058
- Annweiler, C., Brugg, B., Peyrin, J. M., Bartha, R., and Beauchet, O. (2014). Combination of memantine and vitamin D prevents axon degeneration induced by amyloid-beta and glutamate. *Neurobiol. Aging* 35, 331–335. doi: 10.1016/j.neurobiolaging.2013.07.029
- Bai, X., Zhang, C., Chen, A., Liu, W., Li, J., Sun, Q., et al. (2016). Protective effect of edaravone on glutamate-induced neurotoxicity in spiral ganglion neurons. *Neural Plast.* 2016:4034218. doi: 10.1155/2016/4034218
- Barr, J. L., Rasmussen, B. A., Tallarida, C. S., Scholl, J. L., Forster, G. L., Unterwald, E. M., et al. (2015). Ceftriaxone attenuates acute cocaine-evoked dopaminergic neurotransmission in the nucleus accumbens of the rat. *Br. J. Pharmacol.* 172, 5414–5424. doi: 10.1111/bph.13330
- Berry, J. D., Shefner, J. M., Conwit, R., Schoenfeld, D., Keroack, M., Felsenstein, D., et al. (2013). Design and initial results of a multi-phase randomized trial of ceftriaxone in amyotrophic lateral sclerosis. *PLoS ONE* 8:61177. doi: 10.1371/journal.pone.0061177
- Bisht, R., Kaur, B., Gupta, H., and Prakash, A. (2014). Ceftriaxone mediated rescue of nigral oxidative damage and motor deficits in MPTP model of Parkinson's disease in rats. *Neurotoxicology* 44, 71–79. doi: 10.1016/j.neuro.2014.05.009
- Borah, A., Singha, B., and Phukan, S. (2018). Anti-depressant effect of ceftriaxone in forced swimming test and in tail suspension test in mice. *Int. J. Pharm. Sci.* 75, 191–194. doi: 10.22159/ijpps.2016v8i11.14466
- Caban, A., Pisarczyk, K., Kopacz, K., Kapuśniak, A., Toumi, M., Rémuzat, C., et al. (2017). Filling the gap in CNS drug development: evaluation of the role of drug repurposing. *J. Mark. Access Heal. Policy* 5:1299833. doi: 10.1080/20016689.2017.1299833
- Campos, J., Roca, L., and Gude, F., G.-Q. A. (2011). Long-term mortality of patients admitted to the hospital with alcohol withdrawal syndrome. *Alcohol. Clin. Exp. Res.* 35, 1180–1186. doi: 10.1111/j.1530-0277.2011.01451.x
- Cannon, J. R., and Greenamyre, J. T. (2011). The role of environmental exposures in neurodegeneration and neurodegenerative diseases. *Toxicol. Sci.* 124, 225–250. doi: 10.1093/toxsci/kfr239
- Castane, A., Berrendero, F., and Maldonado, R. (2005). The role of the cannabinoid system in nicotine addiction. *Pharm. Biochem Behav* 81, 381–386. doi: 10.1016/j.pbb.2005.01.025
- Chang, S., and Yu, B. (2010). Mitochondrial matters of the brain: mitochondrial dysfunction and oxidative status in epilepsy. *J. Bioenerg. Biomembr.* 41, 457–459. doi: 10.1007/s10863-010-9317-4
- Chen, W. W., Zhang, X., and Huang, W. J. (2016). Role of neuroinflammation in neurodegenerative diseases (Review). *Mol. Med. Rep.* 13, 3391–3396. doi: 10.3892/mmr.2016.4948
- Chen, Z., He, Y., and Wang, Z. J. (2012). The beta-lactam antibiotic, ceftriaxone, inhibits the development of opioid-induced hyperalgesia in mice. *Neurosci. Lett.* 509, 69–71. doi: 10.1016/j.neulet.2011.12.029
- Chotibut, T., Davis, R. W., Arnold, J. C., Frenckel, Z., Gurwara, S., Bondada, V., et al. (2014). Ceftriaxone increases glutamate uptake and reduces striatal tyrosine hydroxylase loss in 6-OHDA Parkinson's model. *Mol. Neurobiol.* 49, 1282–1292. doi: 10.1007/s12035-013-8598-0
- Chotibut, T., Meadows, S., Kasanga, E. A., McInnis, T., Cantu, M. A., Bishop, C., et al. (2017). Ceftriaxone reduces L-dopa-induced dyskinesia severity in 6-hydroxydopamine parkinson's disease model. *Mov. Disord.* 32, 1547–1556. doi: 10.1002/mds.27077
- Corsello, S. M., Bittker, J. A., Liu, Z., Gould, J., McCarren, P., Hirschman, J. E., et al. (2017). The Drug Repurposing Hub: a next-generation drug library and information resource. *Nat. Med.* 23, 405–408. doi: 10.1038/nm.4306

## CONCLUSION

This review revealed that the beta-lactam antibiotic, CEF might have neuroprotective actions, which can affect a wide array of neurological disorders including AD, PD, HD, stroke, brain ischemia, seizure, and drug/alcohol dependency and withdrawal. It is particularly interesting as CEF affects the wider pathological states of CNS disorders including glutamatergic system, oxidative stress, neuroinflammation, apoptotic index, and various toxic protein aggregations. Since most of the studies conducted so far are preclinical studies, clinical studies such as randomized clinical trials and further mechanistic studies are required to assure its neuroprotective actions in real clinical scenarios.

## AUTHOR CONTRIBUTIONS

EY developed the research concept and took initiatives of the work by drafting the manuscript while KT and HH provided greater contributions toward collecting, extracting, and organizing relevant data and also revising the review paper and agreed to be accountable for all aspects of the work.

- Cudkowicz, M. E., Titus, S., Kearney, M., Yu, H., Sherman, A., Schoenfeld, D., et al. (2014). Safety and efficacy of ceftriaxone for amyotrophic lateral sclerosis: a multi-stage, randomised, double-blind, placebo-controlled trial. *Lancet Neurol.* 13, 1083–1091. doi: 10.1016/S1474-4422(14)70222-4
- Cui, C., Cui, Y., Gao, J., Sun, L., Wang, Y., Wang, K., et al. (2014). Neuroprotective effect of ceftriaxone in a rat model of traumatic brain injury. *Neurol. Sci.* 35, 695–700. doi: 10.1007/s10072-013-1585-4
- Cummings, J. (2017). Disease modification and neuroprotection in neurodegenerative disorders. *Transl. Neurodegener.* 6:25. doi: 10.1186/s40035-017-0096-2
- Decourt, B., Lahiri, D., and Sabbagh, M. (2016). Targeting tumor necrosis factor alpha for alzheimer's disease. *Curr. Alzheimer Res.* 13, 1–1. doi: 10.2174/1567205013666160930110551
- Delgado, P. L. (2000). Depression: the case for a monoamine deficiency. *J. Clin. Psychiatry* (61 Suppl. 6), 7–11. doi: 10.1073/pnas.0307294101
- Di Luca, M., Nutt, D., Oertel, W., Boyer, P., Jaarsma, J., Destrebecq, F., et al. (2018). Towards earlier diagnosis and treatment of disorders of the brain. *Bull. World Health Organ.* 96, 298–298A. doi: 10.2471/BLT.17.206599
- Divito, C. B., and Underhill, S. M. (2014). Excitatory amino acid transporters: roles in glutamatergic neurotransmission. *Neurochem. Int.* 73, 172–180. doi: 10.1016/j.neuint.2013.12.008
- Dorst, J., Ludolph, A. C., and Huebers, A. (2018). Disease-modifying and symptomatic treatment of amyotrophic lateral sclerosis. *Ther. Adv. Neurol. Disord.* 11:175628561773473. doi: 10.1177/1756285617734734
- D'Souza, M. S. (2015). Glutamatergic transmission in drug reward: implications for drug addiction. *Front. Neurosci.* 9:404. doi: 10.3389/fnins.2015.00404
- Du, X., West, M. B., Cai, Q., Cheng, W., Ewert, D. L., Li, W., et al. (2017). Antioxidants reduce neurodegeneration and accumulation of pathologic Tau proteins in the auditory system after blast exposure. *Free Radic. Biol. Med.* 108, 627–643. doi: 10.1016/j.freeradbiomed.2017.04.343
- Eljaja, L., Bjerrum, O. J., Honoré, P. H., and Abrahamsen, B. (2011). Effects of the excitatory amino acid transporter subtype 2 (EAAT-2) inducer ceftriaxone on different pain modalities in rat. *Scand. J. Pain* 2, 132–136. doi: 10.1016/j.sjpain.2011.03.003
- Fan, S., Xian, X., Li, L., Yao, X., Hu, Y., Zhang, M., et al. (2018). Ceftriaxone improves cognitive function and upregulates glt-1-related glutamate-glutamine cycle in APP/PS1 mice. *J. Alzheimers Dis.* 66, 1731–1743. doi: 10.3233/JAD-180708
- Feigin, V. L., Abajobir, A. A., Abate, K. H., Abd-Allah, F., Abdulle, A. M., Abera, S. F., et al. (2017). Global, regional, and national burden of neurological disorders during 1990–2015: a systematic analysis for the Global Burden of Disease Study 2015. *Lancet Neurol.* 16, 877–897. doi: 10.1016/S1474-4422(17)30299-5
- Feng, D., Wang, W., Dong, Y., Wu, L., Huang, J., Ma, Y., et al. (2014). Ceftriaxone alleviates early brain injury after subarachnoid hemorrhage by increasing excitatory amino acid transporter 2 expression via the PI3K/Akt/NF-κB signaling pathway. *Neuroscience* 268, 21–32. doi: 10.1016/j.neuroscience.2014.02.053
- Fedorowicz, J. G., and Swartz, K. L. (2004). The role of monoamine oxidase inhibitors in current psychiatric practice. *J. Psychiatr. Pract.* 10, 239–248. doi: 10.1016/j.neuron.2013.04.027
- Franco-Iborra, S., Vila, M., and Perier, C. (2018). Mitochondrial Quality control in neurodegenerative diseases: focus on parkinson's disease and huntington's disease. *Front. Neurosci.* 12:342. doi: 10.3389/fnins.2018.00342
- Gagne, J. J., and Power, M. C. (2010). Anti-inflammatory drugs and risk of parkinson disease: a meta-analysis. *Neurology* 74, 995–1002. doi: 10.1212/WNL.0b013e3181d5a4a3
- Gao, H.-M., and Hong, J.-S. (2008). Why neurodegenerative diseases are progressive: uncontrolled inflammation drives disease progression. *Trends Immunol.* 29, 357–365. doi: 10.1016/j.it.2008.05.002
- Gooch, C. L., Pracht, E., and Borenstein, A. R. (2017). The burden of neurological disease in the United States: a summary report and call to action. *Ann. Neurol.* 81, 479–484. doi: 10.1002/ana.24897
- Goodrich, G. S., Kabakov, A. Y., Hameed, M. Q., Dhamne, S. C., Rosenberg, P. A., and Rotenberg, A. (2013). Ceftriaxone treatment after traumatic brain injury restores expression of the glutamate transporter, GLT-1, reduces regional gliosis, and reduces post-traumatic seizures in the rat. *J. Neurotrauma* 30, 1434–1441. doi: 10.1089/neu.2012.2712
- Gunduz, O., Oltulu, C., Buldum, D., Guven, R., and Ulugol, A. (2011a). Anti-allodynic and anti-hyperalgesic effects of ceftriaxone in streptozocin-induced diabetic rats. *Neurosci. Lett.* 491, 23–25. doi: 10.1016/j.neulet.2010.12.063
- Gunduz, O., Oltulu, C., and Ulugol, A. (2011b). Role of GLT-1 transporter activation in prevention of cannabinoid tolerance by the beta-lactam antibiotic, ceftriaxone, in mice. *Pharmacol. Biochem. Behav.* 99, 100–103. doi: 10.1016/j.pbb.2011.04.012
- Habibi-Asl, B., Vaez, H., Najafi, M., Bidaghi, A., and Ghanbarzadeh, S. (2014). Attenuation of morphine-induced dependence and tolerance by ceftriaxone and amitriptyline in mice. *Acta Anaesthesiol. Taiwanica* 52, 163–168. doi: 10.1016/j.aat.2014.11.001
- Harvey, B. K., Airavaara, M., Hinzman, J., Wires, E. M., Chiocco, M. J., Howard, D. B., et al. (2011). Targeted over-expression of glutamate transporter 1 (GLT-1) reduces ischemic brain injury in a rat model of stroke. *PLoS ONE* 6:22135. doi: 10.1371/journal.pone.0022135
- Hefendehl, J. K., LeDue, J., Ko, R. W. Y., Mahler, J., Murphy, T. H., and MacVicar, B. A. (2016). Mapping synaptic glutamate transporter dysfunction *in vivo* to regions surrounding Aβ plaques by iGluSnFR two-photon imaging. *Nat. Commun.* 7:13441. doi: 10.1038/ncomms13441
- Hernandez, J. J., Pryszlak, M., Smith, L., Yanchus, C., Kurji, N., Shahani, V. M., et al. (2017). Giving drugs a second chance: overcoming regulatory and financial hurdles in repurposing approved drugs as cancer therapeutics. *Front. Oncol.* 7:273. doi: 10.3389/fonc.2017.00273
- Ho, S. C., Hsu, C. C., Pawlak, C. R., Tikhonova, M. A., Lai, T. J., Amstislavskaya, T. G., et al. (2014). Effects of ceftriaxone on the behavioral and neuronal changes in an MPTP-induced Parkinson's disease rat model. *Behav. Brain Res.* 268, 177–184. doi: 10.1016/j.bbr.2014.04.022
- Ho, Y., Weng, J., Lin, C., Shen, M., Li, H., Liao, W., et al. (2018). Ceftriaxone treatment for neuronal deficits: a histological and MEMRI study in a rat model of dementia with lewy bodies. *Behav. Neurol.* 2018:4618716. doi: 10.1155/2018/4618716
- Hsieh, M. H., Meng, W. Y., Liao, W. C., Weng, J. C., Li, H. H., Su, H. L., et al. (2017). Ceftriaxone reverses deficits of behavior and neurogenesis in an MPTP-induced rat model of Parkinson's disease dementia. *Brain Res. Bull.* 132, 129–138. doi: 10.1016/j.brainresbull.2017.05.015
- Hsu, C.-Y., Hung, C.-S., Chang, H.-M., Liao, W.-C., Ho, S.-C., and Ho, Y.-J. (2015). Ceftriaxone prevents and reverses behavioral and neuronal deficits in an MPTP-induced animal model of Parkinson's disease dementia. *Neuropharmacology* 91, 43–56. doi: 10.1016/j.neuropharm.2014.11.023
- Hu, Y., Li, W., Lu, L., Cai, J., Xian, X., Zhang, M., et al. (2010). An anti-nociceptive role for ceftriaxone in chronic neuropathic pain in rats. *Pain* 148, 284–301. doi: 10.1016/j.pain.2009.11.014
- Hu, Y. Y., Xu, J., Zhang, M., Wang, D., Li, L., and Li, W., Bin (2015). Ceftriaxone modulates uptake activity of glial glutamate transporter-1 against global brain ischemia in rats. *J. Neurochem.* 132, 194–205. doi: 10.1111/jnc.12958
- Huang, C. K., Chang, Y. T., Amstislavskaya, T. G., Tikhonova, M. A., Lin, C. L., Hung, C. S., et al. (2015). Synergistic effects of ceftriaxone and erythropoietin on neuronal and behavioral deficits in an MPTP-induced animal model of Parkinson's disease dementia. *Behav. Brain Res.* 294, 198–207. doi: 10.1016/j.bbr.2015.08.011
- Hurd, M. D., Martorell, P., Delavande, A., Mullen, K. J., and Langa, K. M. (2013). Monetary costs of dementia in the united States. *N. Engl. J. Med.* 368, 1326–1334. doi: 10.1056/NEJMsa1204629
- Hussein, A. M., Ghalwash, M., Magdy, K., and Abulseoud, O. A. (2016). Beta lactams antibiotic ceftriaxone modulates seizures, oxidative stress and connexin 43 expression in hippocampus of pentylenetetrazole kindled rats. *J. Epilepsy Res.* 6, 8–15. doi: 10.14581/jer.16002
- Inui, T., Alessandri, B., Heimann, A., Nishimura, F., Frauenknecht, K., Sommer, C., et al. (2013). Neuroprotective effect of ceftriaxone on the penumbra in a rat venous ischemia model. *Neuroscience* 242, 1–10. doi: 10.1016/j.neuroscience.2013.03.018
- Jelenkovic, A. V., Jovanovic, M. D., Stanimirovic, D. D., Bokonjic, D. D., Cicic, G. G., and Boskovic, S. (2008). Beneficial effects of ceftriaxone against pentylenetetrazole-evoked convulsions. *Exp. Biol. Med.* 233, 1389–1394. doi: 10.3181/0803-RM-83
- Kalivas, P. W., LaLumiere, R. T., Knackstedt, L., and Shen, H. (2009). Glutamate transmission in addiction. *Neuropharmacology* 56, 169–173. doi: 10.1016/j.neuropharm.2008.07.011

- Kanai, Y., Clémenton, B., Simonin, A., Leuenberger, M., Lochner, M., Weissenbacher, M., et al. (2013). The SLC1 high-affinity glutamate and neutral amino acid transporter family. *Mol. Aspects Med.* 34, 108–120. doi: 10.1016/j.mam.2013.01.001
- Kaur, B., and Prakash, A. (2017). Ceftriaxone attenuates glutamate-mediated neuro-inflammation and restores BDNF in MPTP model of Parkinson's disease in rats. *Pathophysiology* 24, 71–79. doi: 10.1016/j.pathophys.2017.02.001
- Kelsey, J. E., and Neville, C. (2014). The effects of the  $\beta$ -lactam antibiotic, ceftriaxone, on forepaw stepping and L-DOPA-induced dyskinesia in a rodent model of Parkinson's disease. *Psychopharmacology* 231, 2405–2415. doi: 10.1007/s00213-013-3400-6
- Kim, S. Y., and Jones, T. A. (2013). The effects of ceftriaxone on skill learning and motor functional outcome after ischemic cortical damage in rats. *Restor. Neurol. Neurosci.* 31, 87–97. doi: 10.3233/RNN-2012-120245
- Kim, T.-W. (2015). Drug repositioning approaches for the discovery of new therapeutics for Alzheimer's disease. *Neurotherapeutics* 12, 132–142. doi: 10.1007/s13311-014-0325-7
- Kleteckova, L., Tsenov, G., Kubova, H., Stuchlik, A., and Vales, K. (2014). Neuroprotective effect of the  $3\alpha5\beta$ -pregnanolone glutamate treatment in the model of focal cerebral ischemia in immature rats. *Neurosci. Lett.* 564, 11–15. doi: 10.1016/j.neulet.2014.01.057
- Knackstedt, A. L., Melendez, R. I., and Kalivas, P. W. (2011). Ceftriaxone restores glutamate homeostasis and prevents relapse to cocaine-seeking. *Biol. Psychiatry* 67, 81–84. doi: 10.1016/j.biopsych.2009.07.018.Ceftriaxone
- Krzyzanowska, W., Pomierny, B., Budziszewska, B., Filip, M., and Pera, J. (2016). N-acetylcysteine and ceftriaxone as preconditioning strategies in focal brain ischemia: influence on glutamate transporters expression. *Neurotox. Res.* 29, 539–550. doi: 10.1007/s12640-016-9602-z
- Labra Ruiz, N. A., Santamaría del Ángel, D., Juárez Olgún, H., and Lindoro Silva, M. (2018). Neuroprogression: the hidden mechanism of depression. *Neuropsychiatr. Dis. Treat.* 14, 2837–2845. Available online at: <http://www.ijponline.com/text.asp?2010/42/5/332/70404>
- Lai, P., Huang, Y., Wu, C., Lai, C. J., Wang, P., and Chiu, T. H. (2011). Ceftriaxone attenuates hypoxic-ischemic brain injury in neonatal rats. *J. Biomed. Sci.* 18:69. doi: 10.1186/1423-0127-18-69
- Lee, H.-M., and Kim, Y. (2016). Drug repurposing is a new opportunity for developing drugs against neuropsychiatric disorders. *Schizophr. Res. Treat.* 2016, 1–12. doi: 10.1155/2016/6378137
- Lee, S.-G., Su, Z.-Z., Emdad, L., Gupta, P., Sarkar, D., Borjabad, A., et al. (2008). Mechanism of ceftriaxone induction of excitatory amino acid transporter-2 expression and glutamate uptake in primary human astrocytes. *J. Biol. Chem.* 283, 13116–13123. doi: 10.1074/jbc.M707697200
- Leung, T. C. H., Lui, C. N. P., Chen, L. W., Yung, W. H., Chan, Y. S., and Yung, K. K. L. (2012). Ceftriaxone ameliorates motor deficits and protects dopaminergic neurons in 6-hydroxydopamine-lesioned rats. *ACS Chem. Neurosci.* 3, 22–30. doi: 10.1021/cn200072h
- Lewerenz, J., Albrecht, P., Tien, M. L. T., Henke, N., Karumbayaram, S., Kornblum, H. I., et al. (2009). Induction of Nrf2 and xCT are involved in the action of the neuroprotective antibiotic ceftriaxone *in vitro*. *J. Neurochem.* 111, 332–343. doi: 10.1111/j.1471-4159.2009.06347.x
- Liechti, M., and Markou, A. (2008). Role of the glutamatergic system in nicotine dependence: implications for the discovery and development of new pharmacological smoking cessation therapies. *CNS Drugs* 22, 705–724. doi: 10.2165/00023210-200822090-00001
- Lipski, J., Wan, C. K., Bai, J. Z., Pi, R., Li, D., and Donnelly, D. (2007). Neuroprotective potential of ceftriaxone in *in vitro* models of stroke. *Neuroscience* 146, 617–629. doi: 10.1016/j.neuroscience.2007.02.003
- Liu, X.-L., Li, L., Li, J.-N., Tang, J.-H., Rong, J.-H., Liu, B., et al. (2017). Quantifying absolute glutamate concentrations in nucleus accumbens of prescription opioid addicts by using  $^1\text{H}$  MRS. *Brain Behav.* 7:e00769. doi: 10.1002/brb3.769
- Lowy, M. T., Wittenberg, L., and Yamamoto, B. K. (2002). Effect of acute stress on hippocampal glutamate levels and spectrin proteolysis in young and aged rats. *J. Neurochem.* 65, 268–274. doi: 10.1046/j.1471-4159.1995.6501.0268.x
- Lujia, Y., Xin, L., Shiquan, W., Yu, C., Shuzhuo, Z., and Hong, Z. (2014). Ceftriaxone pretreatment protects rats against cerebral ischemic injury by attenuating microglial activation-induced IL-1 $\beta$  expression. *Int. J. Neurosci.* 124, 657–665. doi: 10.3109/00207454.2013.856009
- Macaluso, A., Bernabucci, M., Trabucco, A., Ciolfi, L., Troisi, F., Baldini, R., et al. (2013). Analgesic effect of a single preoperative dose of the antibiotic ceftriaxone in humans. *J. Pain* 14, 604–612. doi: 10.1016/j.jpain.2013.01.774
- Massie, A., Boillée, S., Hewett, S., Knackstedt, L., and Lewerenz, J. (2015). Main path and byways: non-vesicular glutamate release by system x c – as an important modifier of glutamatergic neurotransmission. *J. Neurochem.* 135, 1062–1079. doi: 10.1111/jnc.13348
- Massie, A., Goursaud, S., Schallier, A., Vermoesen, K., Meshul, C. K., Hermans, E., et al. (2010). Time-dependent changes in GLT-1 functioning in striatum of hemi-Parkinson rats. *Neurochem. Int.* 57, 572–578. doi: 10.1016/j.neuint.2010.07.004
- Miller, B. R., Dorner, J. L., Shou, M., Sari, Y., Barton, S. J., Sengelaub, D. R., et al. (2008). Up-regulation of GLT1 expression increases glutamate uptake and attenuates the Huntington's disease phenotype in the R6/2 mouse. *Neuroscience* 153, 329–337. doi: 10.1016/j.neuroscience.2008.02.004
- Mimura, K., Tomimatsu, T., Minato, K., Jugder, O., Kinugasa-Taniguchi, Y., Kanagawa, T., et al. (2011). Ceftriaxone preconditioning confers neuroprotection in neonatal rats through glutamate transporter 1 upregulation. *Reprod. Sci.* 18, 1193–1201. doi: 10.1177/1933719111410710
- Mineur, Y. S., Picciotto, M. R., and Sanacora, G. (2007). Antidepressant-like effects of ceftriaxone in male. *Biol. Psychiatry* 61, 205–252. doi: 10.1016/j.biopsych.2006.04.037
- Mishra, V., Shuai, B., Kodali, M., Shetty, G. A., Hattiangady, B., Rao, X., et al. (2016). Resveratrol treatment after status epilepticus restrains neurodegeneration and abnormal neurogenesis with suppression of oxidative stress and inflammation. *Sci. Rep.* 5:17807. doi: 10.1038/srep17807
- Mookherjee, P., Green, P. S., Watson, G. S., Marques, M. A., Tanaka, K., Meeker, K. D., et al. (2011). GLT-1 loss accelerates cognitive deficit onset in an Alzheimer's disease animal model. *J. Alzheimers Dis.* 26, 447–455. doi: 10.3233/JAD-2011-110503
- More, S., Kumar, H., Cho, D.-Y., Yun, Y.-S., and Choi, D.-K. (2016). Toxin-induced experimental models of learning and memory impairment. *Int. J. Mol. Sci.* 17:E1447. doi: 10.3390/ijms17091447
- Nicholson, K. J., Gilliland, T. M., and Winkelstein, B. A. (2014). Upregulation of GLT-1 by treatment with ceftriaxone alleviates radicular pain by reducing spinal astrocyte activation and neuronal hyperexcitability. *J. Neurosci. Res.* 92, 116–129. doi: 10.1002/jnr.23295
- Ochoa-Aguilar, A., Sotomayor-Sobrino, M. A., Jaimez, R., Rodríguez, R., Plancarte-Sánchez, R., and Ventura-Martínez, R. (2017). Antialloodynic activity of ceftriaxone and clavulanic acid in acute administration is associated with serum tnf- $\alpha$  modulation and activation of dopaminergic and opioidergic systems. *Drug Dev. Res.* 78, 105–115. doi: 10.1002/ddr.21381
- Parsons, C. G. (2018). CNS repurposing—potential new uses for old drugs: examples of screens for Alzheimer's disease, parkinson's disease and spasticity. *Neuropharmacology* 147, 4–10. doi: 10.1016/j.neuropharm.2018.08.027
- Pinto Pereira, L. M., Phillips, M., Ramlal, H., Teemul, K., and Prabhakar, P. (2004). Third generation cephalosporin use in a tertiary hospital in Port of Spain, trinidad: need for an antibiotic policy. *BMC Infect. Dis.* 4:59. doi: 10.1186/1471-2334-4-59
- Pottabathini, R., Kumar, A., Bhatnagar, A., Garg, S., and Ekaivali (2016). Ameliorative potential of pioglitazone and ceftriaxone alone and in combination in rat model of neuropathic pain: targeting PPAR $\gamma$  and GLT-1 pathways. *Pharmacol. Reports* 68, 85–94. doi: 10.1016/j.pharep.2015.06.010
- Qrunfleh, A. M., Alazizi, A., and Sari, Y. (2013). Ceftriaxone, a beta-lactam antibiotic, attenuates relapse-like ethanol-drinking behavior in alcohol-prefering rats. *J. Psychopharmacology* 27, 541–549. doi: 10.1177/0269881113482529
- Ramos, K. M., Lewis, M. T., Morgan, K. N., Crysdale, N. Y., Kroll, J. L., Taylor, F. R., et al. (2010). Spinal upregulation of glutamate transporter GLT-1 by ceftriaxone: therapeutic efficacy in a range of experimental nervous system disorders. *Neuroscience* 169, 1888–1900. doi: 10.1016/j.neuroscience.2010.06.014
- Rao, P. S. S., and Sari, Y. (2014a). Effectiveness of Ceftriaxone Treatment in preventing relapse-like drinking behavior following long-term ethanol dependence in p rats addiction research and therapy. *Addict. Res. Ther.* 5:1000183. doi: 10.4172/2155-6105.1000183
- Rao, P. S. S., and Sari, Y. (2014b). Effects of ceftriaxone on chronic ethanol consumption: a potential role for xCT and GLT1 modulation of glutamate levels in male P rats. *J. Mol. Neurosci.* 54, 71–77. doi: 10.1007/s12031-014-0251-5

- Roberts-Wolfe, D., and Kalivas, P. (2015). Glutamate transporter GLT-1 as a therapeutic target for substance use disorders. *CNS Neurol. Disord. Drug Targets* 14, 745–756. doi: 10.2174/1871527314666150529144655
- Rossignol, D. A., and Frye, R. E. (2014). Evidence linking oxidative stress, mitochondrial dysfunction, and inflammation in the brain of individuals with autism. *Front. Physiol.* 5:150. doi: 10.3389/fphys.2014.00150
- Rothstein, J. D., Patel, S., Regan, M. R., Haenggeli, C., Huang, Y. H., Bergles, D. E., et al. (2005).  $\beta$ -Lactam antibiotics offer neuroprotection by increasing glutamate transporter expression. *Nature* 433, 73–77. doi: 10.1038/nature03180
- Ruzza, P., Siligardi, G., Hussain, R., Marchiani, A., Islami, M., Bubacco, L., et al. (2014). Ceftriaxone blocks the polymerization of  $\alpha$ -synuclein and exerts neuroprotective effects *in vitro*. *ACS Chem. Neurosci.* 5, 30–38. doi: 10.1021/cn400149k
- Sadigh-Eteghad, S., Sabermarouf, B., Majdi, A., Talebi, M., Farhoudi, M., and Mahmoudi, J. (2015). Amyloid-beta: a crucial factor in Alzheimer's disease. *Med. Princ. Pract.* 24, 1–10. doi: 10.1159/000369101
- Sari, Y., Prieto, A. L., Barton, S. J., Miller, B. R., and Rebec, G. V. (2010). Ceftriaxone-induced up-regulation of cortical and striatal GLT1 in the R6/2 model of Huntington's disease. *J. Biomed. Sci.* 17:62. doi: 10.1186/1423-0127-17-62
- Sari, Y., Sakai, M., Weedman, J. M., Rebec, G. V., and Bell, R. L. (2011). Ceftriaxone, a  $\beta$ -lactam antibiotic, reduces ethanol consumption in alcohol-prefering rats. *Alcohol.* 46, 239–246. doi: 10.1093/alcalc/agl023
- Sari, Y., Sreemantula, S. N., Lee, M. R., and Choi, D. S. (2013). Ceftriaxone treatment affects the levels of GLT1 and ENT1 as well as ethanol intake in alcohol-prefering rats. *J. Mol. Neurosci.* 51, 779–787. doi: 10.1007/s12031-013-0064-y
- Savio, L. E. B., Vuaden, F. C., Piatto, A. L., Bonan, C. D., and Wyse, A. T. S. (2012). Behavioral changes induced by long-term proline exposure are reversed by antipsychotics in zebrafish. *Prog. Neuro Psychopharmacol. Biol. Psychiatry* 36, 258–263. doi: 10.1016/j.pnpbp.2011.10.002
- Scofield, M. D., and Kalivas, P. W. (2014). Astrocytic dysfunction and addiction: consequences of impaired glutamate homeostasis. *Neuroscientist* 20, 610–622. doi: 10.1177/1073858413520347
- Shah, S. A., Amin, F. U., Khan, M., Abid, M. N., Rehman, S. U., Kim, T. H., et al. (2016). Anthocyanins abrogate glutamate-induced AMPK activation, oxidative stress, neuroinflammation, and neurodegeneration in postnatal rat brain. *J. Neuroinflammation* 13:286. doi: 10.1186/s12974-016-0752-y
- Siddique, N., and Siddique, T. (2008). Genetics of amyotrophic lateral sclerosis. *Phys. Med. Rehabil. Clin. N. Am.* 19, 429–439. doi: 10.1016/j.pmr.2008.05.001
- Stepanovic-Petrovic, R. M., Micov, A. M., Tomic, M. A., Kovacevic, J. M., and Bošković, B. D. (2014). Antihyperalgesic/Antinociceptive effects of ceftriaxone and its synergistic interactions with different analgesics in inflammatory pain in rodents. *Anesthesiology* 120, 737–750. doi: 10.1097/ALN.0000435833.33515.b
- Thöne-Reineke, C., Neumann, C., Namolleck, P., Schmerbach, K., Krikov, M., Scheife, J. H., et al. (2008). The  $\beta$ -lactam antibiotic, ceftriaxone, dramatically improves survival, increases glutamate uptake and induces neurotrophins in stroke. *J. Hypertens.* 26, 2426–2435. doi: 10.1097/HJH.0b013e328313e403
- Tikhonova, M. A., Amstislavskaya, T. G., Belichenko, V. M., Fedoseeva, L. A., Kovalenko, S. P., Pisareva, E. E., et al. (2018). Modulation of the expression of genes related to the system of amyloid-beta metabolism in the brain as a novel mechanism of ceftriaxone neuroprotective properties. *BMC Neurosci.* 19(Suppl. 1):13. doi: 10.1186/s12868-018-0412-5
- Tikhonova, M. A., Ho, S. C., Akopyan, A. A., Kolosova, N. G., Weng, J. C., Meng, W. Y., et al. (2017). Neuroprotective effects of ceftriaxone treatment on cognitive and neuronal deficits in a rat model of accelerated senescence. *Behav. Brain Res.* 330, 8–16. doi: 10.1016/j.bbr.2017.05.002
- Tobe, E. (2013). Mitochondrial dysfunction, oxidative stress, and major depressive disorder. *Neuropsychiatr. Dis. Treat.* 9, 567–573. doi: 10.2147/NDT.S44282
- Uttara, B., Singh, A., Zamboni, P., and Mahajan, R. (2009). Oxidative stress and neurodegenerative diseases: a review of upstream and downstream antioxidant therapeutic options. *Curr. Neuropharmacol.* 7, 65–74. doi: 10.2174/157015909787602823
- Van Dam, D., and De Deyn, P. P. (2011). Animal models in the drug discovery pipeline for Alzheimer's disease. *Br. J. Pharmacol.* 164, 1285–1300. doi: 10.1111/j.1476-5381.2011.01299.x
- Volterra, A., Trott, D., Tromba, C., Floridi, S., and Racagni, G. (1994). Glutamate uptake inhibition by oxygen free radicals in rat cortical astrocytes. *J. Neurosci.* 14, 2924–2932. doi: 10.1523/JNEUROSCI.14-05-02924.1994
- Wang, J., Pang, T., Hafko, R., Benicky, J., Sanchez-Lemus, E., and Saavedra, J. M. (2014). Telmisartan ameliorates glutamate-induced neurotoxicity: roles of AT1 receptor blockade and PPARY activation. *Neuropharmacology* 79, 249–261. doi: 10.1016/j.neuropharm.2013.11.022
- Wei, J., Pan, X., Pei, Z., Wang, W., Qiu, W., Shi, Z., et al. (2012). The beta-lactam antibiotic, ceftriaxone, provides neuroprotective potential via anti-excitotoxicity and anti-inflammation response in a rat model of traumatic brain injury. *J. Trauma Acute Care Surg.* 73, 654–660. doi: 10.1097/TA.0b013e31825133c0
- Weng, J. C., Tikhonova, M. A., Chen, J. H., Shen, M. S., Meng, W. Y., Chang, Y. T., et al. (2016). Ceftriaxone prevents the neurodegeneration and decreased neurogenesis seen in a Parkinson's disease rat model: an immunohistochemical and MRI study. *Behav. Brain Res.* 305, 126–139. doi: 10.1016/j.bbr.2016.02.034
- Xie, A., Gao, J., Xu, L., and Meng, D. (2014). Shared mechanisms of neurodegeneration in Alzheimer's disease and parkinson's disease. *Biomed. Res. Int.* 2014:648740. doi: 10.1155/2014/648740
- Yamada, J., and Jinno, S. (2011). Alterations in neuronal survival and glial reactions after axotomy by ceftriaxone and minocycline in the mouse hypoglossal nucleus. *Neurosci. Lett.* 504, 295–300. doi: 10.1016/j.neulet.2011.09.051
- Yang, Q., and Zhou, J. (2018). Neuroinflammation in the central nervous system: symphony of glial cells. *Glia.* doi: 10.1002/glia.23571. [Epub ahead of print].
- Yimer, E. M., Surur, A., Wondafrash, D. Z., and Gebre, A. K. (2019). The effect of metformin in experimentally induced animal models of epileptic seizure. *Behav. Neurol.* 2019:6234758. doi: 10.1155/2019/6234758
- Zhang, Y., Zhang, X., and Qu, S. (2015). Ceftriaxone protects astrocytes from MPP<sup>+</sup>-via suppression of NF- $\kappa$ B/INK/c-jun signaling. *Mol. Neurobiol.* 52, 78–92. doi: 10.1007/s12035-014-8845-z
- Zhou, W., and Hu, W. (2013). Anti-neuroinflammatory agents for the treatment of Alzheimer's disease. *Future Med. Chem.* 5, 1559–1571. doi: 10.4155/fmc.13.125
- Zlotnik, A., Gruenbaum, S. E., Artru, A. A., Rozet, I., Dubilet, M., Tkachov, S., et al. (2009). The neuroprotective effects of oxaloacetate in closed head injury in rats is mediated by its blood glutamate scavenging activity. *J. Neurosurg. Anesthesiol.* 21, 235–241. doi: 10.1097/ANA.0b013e3181a2bf0b
- Zlotnik, A., Gurevich, B., Cherniavsky, E., Tkachov, S., Matuzani-Ruban, A., Leon, A., et al. (2008). The contribution of the blood glutamate scavenging activity of pyruvate to its neuroprotective properties in a rat model of closed head injury. *Neurochem. Res.* 33, 1044–1050. doi: 10.1007/s11064-007-9548-x
- Zumkehr, J., Rodriguez-Ortiz, C. J., Cheng, D., Kieu, Z., Wai, T., Hawkins, C., et al. (2015). Ceftriaxone ameliorates tau pathology and cognitive decline via restoration of glial glutamate transporter in a mouse model of Alzheimer's disease. *Neurobiol. Aging* 36, 2260–2271. doi: 10.1016/j.neurobiolaging.2015.04.005

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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