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Editorial: Brain extracellular matrix: Involvement in adult neural functions and disease volume II

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Editorial on the Research Topic

Brain extracellular matrix: Involvement in adult neural functions and disease volume II

In recent years, the brain extracellular matrix (ECM) has come to the forefront of the neuroscience field. Its role as a key player in developmental and adult brain functions as well as in brain disease is being increasingly recognized. The ECM wide-ranging functions span an organism's lifetime, from early prenatal development to adulthood and aging (Ishii and Maeda, 2008; Cabungcal et al., 2013; Senkov et al., 2014; Suttkus et al., 2014; Maeda, 2015; Pomin, 2015; Karus et al., 2016; Carulli and Verhaagen, 2021; Fawcett et al., 2022). Growing interest in the brain ECM is leading to a greater understanding of the mechanisms underlying the ECM's role in synaptic plasticity, learning and memory (Senkov et al., 2014; Tsilibary et al., 2014; Beroun et al., 2019; Carulli and Verhaagen, 2021; Fawcett et al., 2022), regulation of neuronal activity (Wingert and Sorg, 2021), neurodevelopmental processes such as neuronal migration and axon guidance (Pizzorusso et al., 2002; Ishii and Maeda, 2008; Mauney et al., 2013; Maeda, 2015; Karus et al., 2016), neuroprotection (Morawski et al., 2004; Cabungcal et al., 2013; Suttkus et al., 2014), and hypothalamic regulation of metabolism (Dingess et al., 2018; Mirzadeh et al., 2019). Mounting evidence implicates the ECM in a growing number of brain disorders including schizophrenia, bipolar disorder, autism (Eastwood and Harrison, 2006; Ma et al., 2009; Weiss et al., 2009; Abdallah et al., 2012; Poelmans et al., 2013; Pantazopoulos and Berretta, 2016; Velmeshev et al., 2019; Brandenburg and Blatt, 2022), substance use disorders (Slaker et al., 2016; Garcia-Keller et al., 2019; Seney et al., 2021), depression (Riga et al., 2017; Alaiyed et al., 2020; Koskinen et al., 2020; Blanco and Conant, 2021), Alzheimer's disease (Scholefield et al., 2003; Schworer et al., 2013; Morawski et al., 2014; Yang et al., 2017), and stroke (Hobohm et al., 2005; Hartig et al., 2017).

This collection of original and review articles in Volume II of this Special Issue, includes examples of novel ECM structures and the ECM's role in plasticity, axonal regeneration, neural injury, neuroinflammation as well as a broad range of brain disorders, providing a window into a growing but still largely unexplored field of investigations.

For several decades, Wisteria floribunda agglutinin (WFA) lectin labeling, first introduced in the 1990's by Hartig et al. (1992) has been the primary method for studying the ECM, and more specifically a distinct population of perineuronal nets (PNNs). Indeed, many important studies using WFA labeling have provided information on the functional of role of these ECM structured forms, their association with distinct neuronal populations and involvement in brain disorders. In this special issue, Hartig et al., provide an extensive perspective on methodological aspects related to WFA lectin for PNN labeling, based on 30 years of experience. Several methodological factors potentially impacting PNN detection with WFA are discussed, along with technical guidelines and a PNN labeling "toolbox". Notably, applications for in vivo and in vivo labeling as well as electron microscopy are discussed. At the same time, the authors share important considerations related to experimental conditions that may affect WFA ECM labeling and introduce bias. The authors point to intriguing discrepancies between WFA labeling and its putative main target, i.e., aggrecan-associated chondroitin sulfate chains, discuss some of the factors potentially contributing to this discrepancy and emphasize the need for use of multiple PNN markers.

Much of the focus on brain ECM studies has been on PNNs, specialized structures first described by Camillo Golgi in 1898 (Golgi, 1989). PNNs ensheath distinct populations of neurons and regulate key neural functions such as synaptic plasticity, electrophysiological properties and access to neuroactive molecules (Kalb and Hockfield, 1988; Sugiyama et al., 2008; Gogolla et al., 2009). Emerging evidence shows that PNNs are only one of many structured ECM forms in the brain, including perisynaptic and perinodal ECM (Dours-Zimmermann et al., 2009; Bekku and Oohashi, 2010; Faissner et al., 2010; Frischknecht and Seidenbecher, 2012; Fawcett et al., 2019), chondroitin-6-sulfate clusters (CS6-clusters) and axonal coats (Hayashi et al., 2007; Bruckner et al., 2008; Okuda et al., 2014; Pantazopoulos et al., 2015). Evidence from the original manuscript by Pantazopoulos et al. shows that the chondroitin sulfate proteoglycans (CSPGs) NG2 and brevican form axonal coats, i.e., structured ECM sheaths, surrounding myelinated axons in the human thalamus. Detailed confocal and electron microscope analyses demonstrate intricate interweaving patterns of CSPGs with myelin sheaths and a preferential relationship with large axons. These findings, consistent with a role of brevican and NG2 in the regulation of axonal functions such as saltatory conductance and fasciculation, add to emerging evidence for a variety of ECM

structures in the brain, likely to support specialized functions (see also Ray et al.).

The regulation of axonal functions, and specifically axonal growth, has been intensively studied in the context of axon injuries (Miyata and Kitagawa, 2015; Hussein et al., 2020). During neurodevelopment the ECM guides axons to their targets and regulates fasciculation (Snow et al., 2003; Kwok et al., 2012). In contrast, during adulthood, the ECM inhibits axonal regrowth following injuries (Miyata and Kitagawa, 2015; Hussein et al., 2020). Takiguchi et al. offer an important contribution to this field of investigations. Their elegant studies address an important question, i.e., whether growing axons following an injury reach and synapse with their targets in animals treated with ECM enzymatic degradation. Their results indeed demonstrate that, in experimental animals with complete spinal resection, enzymatic ECM degradation led to axonal growth through the injured site, formation of synapses between the newly grown axons and motor neurons, and improved motor functions.

ECM functions are in large part mediated by CSPGs and heparan sulfate proteoglycans (HSPGs). These large molecules are composed by a protein core to which a variable number of disaccharide chains are attached. The specificity of their biological effects is mediated by the sulfation patterns of the sugar chains, which determine their conformation, charge, and molecular affinities (Miyata et al., 2012; Smith et al., 2015). The elegant review contributed by Fawcett and Kwok focuses on the functional role of disaccharide sulfation on CSPGs and HSPGs. These latter have received less attention in the context of investigations on the brain ECM. Evidence compellingly reviewed by the authors highlights and compares CSPG and HSPGs functional roles in memory processing, neural injury, aging, axonal regeneration, and their modulation encoded by sulfation patterns.

The considerations above, particularly those emphasized by Hartig et al. and Fawcett and Kwok, resonate with those poignantly made by Scarlett et al. in the context of studies on the role of the ECM in brain disorders, and particularly Alzheimer's disease. As the authors eloquently point out, changes in the ECM sulfation "coding," together with technical limitations surrounding traditional methods for PNN analyses, may reside at the root of important discrepancies in literature on the role of the ECM in neurodegenerative disorders. The paper puts forth the intriguing hypothesis that disease state-related chondroitin sulfation changes may impact PNN detection, and in turn interpretations of results showing altered PNN representation (Baig et al., 2005; Crapser et al., 2020). The distinction between altered PNN numbers and altered chondroitin sulfation patterns is significant, as each can affect CSPG functions, such as their ability to restrict or promote synaptic plasticity, in a different manner (Yang et al., 2021).

Inflammation has rapidly taken center stage as a key factor in a growing number of brain disorders, from autism

to dementias, motor, and psychiatric disorders. The bloodbrain barrier functions as a key interface regulating molecular transport between the brain and the peripheral circulatory system. In this special issue, Tabet et al. offer a comprehensive description of the relationship between the ECM and several components of the blood-brain barrier including the basement membrane and the glycocalyx. The authors discuss the role of the ECM in modulating blood-brain barrier permeability and its mediation of blood-brain barrier alterations during brain injury and inflammation.

Investigations on the ECM role in the pathogenesis of opioid use disorder (OUD) are rapidly gathering pace. Ray et al. offer a compelling overview of the ECM functional role within reward circuits and its involvement in OUD, including opioid seeking, craving and relapse. The authors consider the role of several forms of ECM, a recurring and important theme in this special issue. Particularly notable is the emphasis on sex-specificity, proposed to play a role both on the effects of opioid exposure on the ECM and on the contribution of ECM molecular signaling to opioid use disorder.

A novel and intriguing report by Liu et al. provides evidence for a role of the ECM in insomnia. The authors analyzed serum samples from patients with insomnia and demonstrated significant changes impacting several ECM factors, including matrix metalloproteinase 9 (MMP9) in persons suffering from insomnia. These findings bring forth a potential role for the ECM in sleep regulation. The intersection of ECM and immune factors found to be altered in insomnia offers new avenues for investigations on sleep deficits and potential therapeutic targets.

We believe that the articles highlighted in this special issue provide an exciting overview of the current state of research on

the role of the ECM in the regulation of brain processes and involvement in disease states, and current directions regarding PNN labeling novel ECM structures. We hope that they will stimulate further studies the role of ECM in adult brain processes and psychiatric disorders. A deeper understanding of the role the ECM in these processes has the potential to allow us to leverage this system for preventative and therapeutic treatments.

Author contributions

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

Abdallah, M. W., Pearce, B. D., Larsen, N., Greaves-Lord, K., Norgaard-Pedersen, B., Hougaard, D. M., et al. (2012). Amniotic fluid MMP-9 and neurotrophins in autism spectrum disorders: an exploratory study. *Autism Res.* 5, 428–433. doi:10.1002/aur.1254

Alaiyed, S., McCann, M., Mahajan, G., Rajkowska, G., Stockmeier, C. A., Kellar, K. J., et al. (2020). Venlafaxine stimulates an MMP-9-dependent increase in excitatory/inhibitory balance in a stress model of depression. *J. Neurosci.* 40, 4418–4431. doi: 10.1523/JNEUROSCI.2387-19.2020

Baig, S., Wilcock, G. K., and Love, S. (2005). Loss of perineuronal net Nacetylgalactosamine in Alzheimer's disease. *Acta Neuropathol.* 110, 393–401. doi: 10.1007/s00401-005-1060-2

Bekku, Y., and Oohashi, T. (2010). Neurocan contributes to the molecular heterogeneity of the perinodal ECM. *Arch. Histol. Cytol.* 73, 95–102. doi: 10.1679/aohc.73.95

Beroun, A., Mitra, S., Michaluk, P., Pijet, B., Stefaniuk, M., and Kaczmarek, L. (2019). MMPs in learning and memory and neuropsychiatric disorders. *Cell. Mol. Life Sci.* 76, 3207–3228. doi: 10.1007/s00018-019-03180-8

Blanco, I., and Conant, K. (2021). Extracellular matrix remodeling with stress and depression: studies in human, rodent and zebrafish models. *Eur. J. Neurosci.* 53, 3879–3888. doi: 10.1111/ejn.14910

Brandenburg, C., and Blatt, G. J. (2022). Region-specific alterations of perineuronal net expression in postmortem autism brain tissue. *Front. Mol. Neurosci.* 15, 838918. doi: 10.3389/fnmol.2022.838918

Bruckner, G., Morawski, M., and Arendt, T. (2008). Aggrecan-based extracellular matrix is an integral part of the human basal ganglia circuit. *Neuroscience* 151, 489–504. doi: 10.1016/j.neuroscience.2007.10.033

Cabungcal, J. H., Steullet, P., Morishita, H., Kraftsik, R., Cuenod, M., Hensch, T. K., et al. (2013). Perineuronal nets protect fast-spiking interneurons against oxidative stress. *Proc. Natl. Acad. Sci. U. S. A.* 110, 9130–9135. doi: 10.1073/pnas.1300454110

Carulli, D., and Verhaagen, J. (2021). An extracellular perspective on CNS maturation: perineuronal nets and the control of plasticity. *Int. J. Mol. Sci.* 22, 2434. doi: 10.3390/ijms22052434

Crapser, J. D., Spangenberg, E. E., Barahona, R. A., Arreola, M. A., Hohsfield, L. A., and Green, K. N. (2020). Microglia facilitate loss of perineuronal nets in the Alzheimer's disease brain. *EBioMedicine* 58, 102919. doi: 10.1016/j.ebiom.2020.102919

Dingess, P. M., Harkness, J. H., Slaker, M., Zhang, Z., Wulff, S. S., Sorg, B. A., et al. (2018). Consumption of a high-fat diet alters perineuronal nets in the prefrontal cortex. *Neural Plast.* 2018, 2108373. doi: 10.1155/2018/2108373

Dours-Zimmermann, M. T., Maurer, K., Rauch, U., Stoffel, W., Fassler, R., and Zimmermann, D. R. (2009). Versican V2 assembles the extracellular matrix surrounding the nodes of ranvier in the CNS. *J. Neurosci.* 29, 7731–7742. doi: 10.1523/JNEUROSCI.4158-08.2009

Eastwood,		S. L.,	and	Harrison,	P. J.	(2006).	Cellular
basis	of	reduced	cortical	reelin e	xpression	in schiz	ophrenia.
Am.	J.	Psychiatry.	163,	540-542.	doi:	10.1176/appi.ajp.163.	
3.540							

Faissner, A., Pyka, M., Geissler, M., Sobik, T., Frischknecht, R., Gundelfinger, E. D., et al. (2010). Contributions of astrocytes to synapse formation and maturation - potential functions of the perisynaptic extracellular matrix. *Brain Res. Rev.* 63, 26–38. doi: 10.1016/j.brainresrev.2010.01.001

Fawcett, J. W., Fyhn, M., Jendelova, P., Kwok, J. C. F., Ruzicka, J., and Sorg, B. A. (2022). The extracellular matrix and perineuronal nets in memory. *Mol. Psychiatry*. doi: 10.1038/s41380-022-01634-3. [Epub ahead of print].

Fawcett, J. W., Oohashi, T., and Pizzorusso, T. (2019). The roles of perineuronal nets and the perinodal extracellular matrix in neuronal function. *Nat. Rev. Neurosci.* 20, 451–465. doi: 10.1038/s41583-019-0196-3

Frischknecht, R., and Seidenbecher, C. I. (2012). Brevican: a key proteoglycan in the perisynaptic extracellular matrix of the brain. *Int. J. Biochem. Cell Biol.* 44, 1051–1054. doi: 10.1016/j.biocel.2012.03.022

Garcia-Keller, C., Neuhofer, D., Bobadilla, A. C., Spencer, S., Chioma, V. C., Monforton, C., et al. (2019). Extracellular matrix signaling through beta3 integrin mediates cocaine cue-induced transient synaptic plasticity and relapse. *Biol. Psychiatry*. 86, 377–387. doi: 10.1016/j.biopsych.2019.03.982

Gogolla, N., Caroni, P., Luthi, A., and Herry, C. (2009). Perineuronal nets protect fear memories from erasure. *Science* 325, 1258–1261. doi: 10.1126/science.1174146

Golgi, C. (1989). On the structure of nerve cells. J. Microsc. 155, 3-7. doi: 10.1111/j.1365-2818.1989.tb04294.x

Hartig, W., Brauer, K., and Bruckner, G. (1992). Wisteria floribunda agglutininlabelled nets surround parvalbumin-containing neurons. *Neuroreport* 3, 869–872. doi: 10.1097/00001756-199210000-00012

Hartig, W., Mages, B., Aleithe, S., Nitzsche, B., Altmann, S., Barthel, H., et al. (2017). Damaged neocortical perineuronal nets due to experimental focal cerebral ischemia in mice, rats and sheep. *Front. Integr. Neurosci.* 11, 15. doi: 10.3389/fnint.2017.00015

Hayashi, N., Tatsumi, K., Okuda, H., Yoshikawa, M., Ishizaka, S., Miyata, S., et al. (2007). DACS, novel matrix structure composed of chondroitin sulfate proteoglycan in the brain. *Biochem. Biophys. Res. Commun.* 364, 410–415. doi: 10.1016/j.bbrc.2007.10.040

Hobohm, C., Gunther, A., Grosche, J., Rossner, S., Schneider, D., and Bruckner, G. (2005). Decomposition and long-lasting downregulation of extracellular matrix in perineuronal nets induced by focal cerebral ischemia in rats. *J. Neurosci. Res.* 80, 539–548. doi: 10.1002/jnr.20459

Hussein, R. K., Mencio, C. P., Katagiri, Y., Brake, A. M., and Geller, H. M. (2020). Role of chondroitin sulfation following spinal cord injury. *Front. Cell. Neurosci.* 14, 208. doi: 10.3389/fncel.2020.00208

Ishii, M., and Maeda, N. (2008). Oversulfated chondroitin sulfate plays critical roles in the neuronal migration in the cerebral cortex. *J. Biol. Chem.* 283, 32610–32620. doi: 10.1074/jbc.M806331200

Kalb, R. G., and Hockfield, S. (1988). Molecular evidence for early activitydependent development of hamster motor neurons. *J. Neurosci.* 8, 2350–2360. doi: 10.1523/JNEUROSCI.08-07-02350.1988

Karus, M., Ulc, A., Ehrlich, M., Czopka, T., Hennen, E., Fischer, J., et al. (2016). Regulation of oligodendrocyte precursor maintenance by chondroitin sulphate glycosaminoglycans. *Glia* 64, 270–286. doi: 10.1002/glia.22928

Koskinen, M. K., van Mourik, Y., Smit, A. B., Riga, D., and Spijker, S. (2020). From stress to depression: development of extracellular matrixdependent cognitive impairment following social stress. *Sci. Rep.* 10, 17308. doi: 10.1038/s41598-020-73173-2

Kwok, J. C., Yuen, Y. L., Lau, W. K., Zhang, F. X., Fawcett, J. W., Chan, Y. S., et al. (2012). Chondroitin sulfates in the developing rat hindbrain confine commissural projections of vestibular nuclear neurons. *Neural Dev.* 7, 6. doi: 10.1186/1749-8104-7-6

Ma, D., Salyakina, D., Jaworski, J. M., Konidari, I., Whitehead, P. L., Andersen, A. N., et al. (2009). A genome-wide association study of autism reveals a common novel risk locus at 5p14.1. *Ann. Hum. Genet.* 73, 263–273. doi: 10.1111/j.1469-1809.2009.00523.x

Maeda, N. (2015). Proteoglycans and neuronal migration in the cerebral cortex during development and disease. *Front. Neurosci.* 9, 98. doi: 10.3389/fnins.2015.00098

Mauney, S. A., Athanas, K. M., Pantazopoulos, H., Shaskan, N., Passeri, E., Berretta, S., et al. (2013). Developmental pattern of perineuronal nets in the human prefrontal cortex and their deficit in schizophrenia. *Biol. Psychiatry* 74, 427–435. doi: 10.1016/j.biopsych.2013.05.007

Mirzadeh, Z., Alonge, K. M., Cabrales, E., Herranz-Perez, V., Scarlett, J. M., Brown, J. M., et al. (2019). Perineuronal net formation during the critical period for neuronal maturation in the hypothalamic arcuate nucleus. *Nat Metab.* 1, 212–221. doi: 10.1038/s42255-018-0029-0

Miyata, S., and Kitagawa, H. (2015). Mechanisms for modulation of neural plasticity and axon regeneration by chondroitin sulphate. J. Biochem. 157, 13–22. doi: 10.1093/jb/mvu067

Miyata, S., Komatsu, Y., Yoshimura, Y., Taya, C., and Kitagawa, H. (2012). Persistent cortical plasticity by upregulation of chondroitin 6-sulfation. *Nat. Neurosci.* 15, 414–422, S411–S412. doi: 10.1038/nn.3023

Morawski, M., Bruckner, M. K., Riederer, P., Bruckner, G., and Arendt, T. (2004). Perineuronal nets potentially protect against oxidative stress. *Exp. Neurol.* 188, 309–315. doi: 10.1016/j.expneurol.2004.04.017

Morawski, M., Filippov, M., Tzinia, A., Tsilibary, E., and Vargova, L. (2014). ECM in brain aging and dementia. *Prog. Brain Res.* 214, 207–227. doi: 10.1016/B978-0-444-63486-3.00010-4

Okuda, H., Tatsumi, K., Morita, S., Shibukawa, Y., Korekane, H., Horii-Hayashi, N., et al. (2014). Chondroitin sulfate proteoglycan tenascin-R regulates glutamate uptake by adult brain astrocytes. *J. Biol. Chem.* 289, 2620–2631. doi:10.1074/jbc.M113.504787

Pantazopoulos, H., and Berretta, S. (2016). In sickness and in health: perineuronal nets and synaptic plasticity in psychiatric disorders. *Neural Plast.* 2016, 9847696. doi: 10.1155/2016/9847696

Pantazopoulos, H., Markota, M., Jaquet, F., Ghosh, D., Wallin, A., Santos, A., et al. (2015). Aggrecan and chondroitin-6-sulfate abnormalities in schizophrenia and bipolar disorder: a postmortem study on the amygdala. *Transl. Psychiatry* 5, e496. doi: 10.1038/tp.2014.128

Pizzorusso, T., Medini, P., Berardi, N., Chierzi, S., Fawcett, J. W., and Maffei, L. (2002). Reactivation of ocular dominance plasticity in the adult visual cortex. *Science* 298, 1248–1251. doi: 10.1126/science.1072699

Poelmans, G., Franke, B., Pauls, D. L., Glennon, J. C., and Buitelaar, J. K. (2013). AKAPs integrate genetic findings for autism spectrum disorders. *Transl. Psychiatry* 3, e270. doi: 10.1038/tp.2013.48

Pomin, V. H. (2015). Sulfated glycans in inflammation. Eur. J. Med. Chem. 92, 353-369. doi: 10.1016/j.ejmech.2015.01.002

Riga, D., Kramvis, I., Koskinen, M. K., van Bokhoven, P., van der Harst, J. E., Heistek, T. S., et al. (2017). Hippocampal extracellular matrix alterations contribute to cognitive impairment associated with a chronic depressive-like state in rats. *Sci. Transl. Med.* 9, eaai8753. doi: 10.1126/scitranslmed.aai8753

Scholefield, Z., Yates, E. A., Wayne, G., Amour, A., McDowell, W., and Turnbull, J. E. (2003). Heparan sulfate regulates amyloid precursor protein processing by BACE1, the Alzheimer's beta-secretase. *J. Cell Biol.* 163, 97–107. doi: 10.1083/jcb.200303059

Schworer, R., Zubkova, O. V., Turnbull, J. E., and Tyler, P. C. (2013). Synthesis of a targeted library of heparan sulfate hexa- to dodecasaccharides as inhibitors of beta-secretase: potential therapeutics for Alzheimer's disease. *Chemistry* 19, 6817–6823. doi: 10.1002/chem.201204519

Seney, M. L., Kim, S. M., Glausier, J. R., Hildebrand, M. A., Xue, X., Zong, W., et al. (2021). Transcriptional alterations in dorsolateral prefrontal cortex and nucleus accumbens implicate neuroinflammation and synaptic remodeling in opioid use disorder. *Biol. Psychiatry.* 90, 550–562. doi: 10.1016/j.biopsych.2021.06.007

Senkov, O., Andjus, P., Radenovic, L., Soriano, E., and Dityatev, A. (2014). Neural ECM molecules in synaptic plasticity, learning, and memory. *Prog. Brain Res.* 214, 53–80. doi: 10.1016/B978-0-444-63486-3.00003-7

Slaker, M., Blacktop, J. M., and Sorg, B. A. (2016). Caught in the net: perineuronal nets and addiction. *Neural Plast.* 2016, 7538208. doi: 10.1155/2016/7538208

Smith, P. D., Coulson-Thomas, V. J., Foscarin, S., Kwok, J. C., and Fawcett, J. W. (2015). "GAG-ing with the neuron": the role of glycosaminoglycan patterning in the central nervous system. *Exp. Neurol.* 274, 100–114. doi: 10.1016/j.expneurol.2015.08.004

Snow, D. M., Smith, J. D., Cunningham, A. T., McFarlin, J., and Goshorn, E. C. (2003). Neurite elongation on chondroitin sulfate proteoglycans is characterized by axonal fasciculation. *Exp. Neurol.* 182, 310–321. doi: 10.1016/S0014-4886(03)00034-7

Sugiyama, S., Di Nardo, A. A., Aizawa, S., Matsuo, I., Volovitch, M., Prochiantz, A., et al. (2008). Experience-dependent transfer of Otx2 homeoprotein

into the visual cortex activates postnatal plasticity. *Cell* 134, 508–520. doi: 10.1016/j.cell.2008.05.054

Suttkus, A., Rohn, S., Weigel, S., Glockner, P., Arendt, T., and Morawski, M. (2014). Aggrecan, link protein and tenascin-R are essential components of the perineuronal net to protect neurons against iron-induced oxidative stress. *Cell Death Dis.* 5, e1119. doi: 10.1038/cddis.2014.25

Tsilibary, E., Tzinia, A., Radenovic, L., Stamenkovic, V., Lebitko, T., Mucha, M., et al. (2014). Neural ECM proteases in learning and synaptic plasticity. *Prog. Brain Res.* 214, 135–157. doi: 10.1016/B978-0-444-63486-3.00006-2

Velmeshev, D., Schirmer, L., Jung, D., Haeussler, M., Perez, Y., Mayer, S., et al. (2019). Single-cell genomics identifies cell type-specific molecular changes in autism. *Science* 364, 685–689. doi: 10.1126/science.aav8130

Weiss, L. A., Arking, D. E., Gene Discovery Project of Johns Hopkins & the Autism Consortium, Daly, M. J., and Chakravarti, A. (2009). A genome-wide

linkage and association scan reveals novel loci for autism. Nature. 461, 802–808. doi: 10.1038/nature08490

Wingert, J. C., and Sorg, B. A. (2021). Impact of perineuronal nets on electrophysiology of parvalbumin interneurons, principal neurons, and brain oscillations: a review. *Front. Synaptic Neurosci.* 13, 673210. doi: 10.3389/fnsyn.2021.673210

Yang, S., Gigout, S., Molinaro, A., Naito-Matsui, Y., Hilton, S., Foscarin, S., et al. (2021). Chondroitin 6-sulphate is required for neuroplasticity and memory in ageing. *Mol. Psychiatry* 26, 5658–5668. doi: 10.1038/s41380-021-01208-9

Yang, S., Hilton, S., Alves, J. N., Saksida, L. M., Bussey, T., Matthews, R. T., et al. (2017). Antibody recognizing 4-sulfated chondroitin sulfate proteoglycans restores memory in tauopathy-induced neurodegeneration. *Neurobiol. Aging.* 59, 197–209. doi: 10.1016/j.neurobiolaging.2017. 08.002