



Selenium Transport Mechanism via Selenoprotein P—Its Physiological Role and Related Diseases

Yoshiro Saito*

Laboratory of Molecular Biology and Metabolism, Graduate School of Pharmaceutical Sciences, Tohoku University, Sendai, Japan

Selenoprotein P (SELENOP) is selenium (Se)-containing protein in plasma, which is primarily produced in the liver. The "P" in SELENOP originated from the presence in plasma. SELENOP contains selenocysteine, a cysteine analog containing Se instead of sulfur. SELENOP is a multi-functional protein to reduce phospholipid hydroperoxides and to deliver Se from the liver to other tissues, such as those of the brain and testis, playing a pivotal role in Se metabolism and antioxidative defense. Decrease in SELENOP causes various dysfunctions related to Se deficiency and oxidative stress, while excessive SELENOP causes insulin resistance. This review focuses on the Se transport system of SELENOP, particularly its molecular mechanism and physiological role in Se metabolism. Furthermore, the chemical form of Se and its biological meaning is discussed.

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*Correspondence:

Yoshiro Saito yoshiro.saito.a8@tohoku.ac.jp

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INTRODUCTION

Selenoprotein P (encoded as SELENOP) was first described in 1973, and its character was reported as the major selenium (Se)-containing protein in plasma (1, 2). SELENOP is mainly synthesized in the liver and secreted to the plasma after cleavage of the signal peptide. The "P" in SELENOP denotes its presence in plasma. SELENOP contains the essential trace element Se in the form of selenocysteine (Sec), which is an analog of cysteine that contains Se instead of sulfur (2-4). Twentyfive genes encoding human Sec-containing proteins, i.e., selenoproteins, have been discovered, which play significant roles in several physiological processes such as antioxidant defense and metabolism; five types of glutathione (GSH) peroxidases (GPXs) play significant roles in the removal of several hydroperoxides, three types of thioredoxin reductases (TRXRs) in redox regulation, three types of iodothyronine deiodinases in the regulation of thyroid hormones, and selenophosphate synthetase 2 (SEPHS2) in Sec synthesis (4, 5). Most selenoproteins have a Sec residue, while SELENOP has 10, making it multifunctional (6, 7). SELENOP possesses two different functions: Se transport activity to supply Se to cells and GPX-like activity to reduce phospholipid hydroperoxide (8, 9). SELENOP maintains selenoenzymes in several tissues and plays a crucial role in antioxidative defense and Se metabolism (10, 11). A decrease in SELENOP causes deficiency in selenoproteins and various dysfunctions with oxidative stress, while excess SELENOP induces insulin resistance, which can lead to type 2 diabetes (12, 13). This review focuses on the Se transport system via SELENOP, particularly its molecular mechanism and physiological role in Se metabolism. Furthermore, the chemical form of Se and its biological meaning are discussed.

SELENOPROTEINS IN HUMAN PLASMA

In human plasma, there are two kinds of selenoproteins: SELENOP and extracellular GPX (GPX3), possessing Se as Sec residue (14). SELENOP is primarily secreted from the liver, while GPX3 is synthesized in the kidney. To estimate the Se content derived from each selenoprotein, SELENOP- and GPX3-deficient human plasma were prepared using immobilized specific antibody (9). The absorption of SELENOP resulted in the decrease in Se content to 47% of the total, while the removal of GPX3 decreased it to 81%, indicating that 53 and 19% of plasma Se is derived from SELENOP and GPX3, respectively (Figure 1). Similar results on the contributions of SELENOP and GPX3 to plasma Se have been reported by the laboratories of Burk and Schomburg (10), Olson et al. (15), and Brodin et al. (16). The residual 28% of Se might be derived from selenomethionine (SeMet) in albumin, and/or low molecular Se compounds, which have in part been identified as selenosugars (17).

Sec is encoded by the UGA codon, known as a stop (opal) codon, and is called the 21st amino acid in the genetic code (18, 19). In eukaryotes, the Sec insertion sequence (SECIS), which is a specific hairpin structure located in the 3' untranslated region (3'UTR) of selenoprotein mRNA, is essential for the incorporation of Sec during the biosynthesis of selenoproteins. SECIS binds SECIS-binding protein 2 (SBP2) and forms a complex for Sec incorporation via the recruitment of the Secspecific eukaryotic elongation factor (eEFsec) and Sec-tRNA^{Sec} (an anticodon complementary to the UGA codon) (20).

The mRNA of SELENOP has a unique property of containing 10 UGA codons in the open reading frame (ORF) and two SECIS in the 3'UTR, while other selenoprotein mRNAs have only one SECIS element (21) (Figure 1A). Multiple Sec residues in SELENOP are important for its function; one N-terminal Sec residue forms an active site of enzyme activity to reduce phospholipid hydroperoxide, while the nine C-terminal Sec residues function as Se transporter (22) (Figure 1A). The first SECIS element, which is located on the 5' side near the stop codon, mainly facilitates the processive Sec incorporation, while the second SECIS functions slow decoding at the first UGA codon (23). Plasma kallikrein cleaves SELENOP by limited proteolysis with Arg-235-Gln-236 and Arg-242-Asp-243, generating Nterminal fragments (residues 1-235) with enzyme activity and C-terminal fragment (residues 243-361) exhibiting Se-supply activity (22). N-terminal, a possible catalytic center of SELENOP, has U(Sec)XXC motif, similar to the active-site of thioredoxin (CXXC motif), which suggests the reactivity of SELENOP against protein thiols. Actually, SELENOP has broad thiol specificity (24), and it uses not only GSH but also other thiols, such as TRX, dithiothreitol, and mercaptoethanol, as reducing agents, while the thiol specificity of cellular GPX (GPX1) is narrow, using only GSH as reductant. In human plasma, the concentration of GSH and TRX is $\sim 5 \,\mu$ M and 2 nM, respectively (25, 26). Based on kinetic analysis, SELENOP uses TRX 500-fold more effectively than GSH (24), but it is still uncertain whether both thiols could contribute to the reduction of SELENOP in plasma.

The mRNA of GPX3 has a UGA codon in the ORF and a SECIS in the 3'UTR, which is a representative feature of selenoproteins, such as the GPX family. GPX3 has catalytic triad composed of Sec, Gln, and Trp, in which the Se of a Sec residue is activated by hydrogen bonding to Trp and Gln residue (27). GPX3, like GPX1, is a homotetramer, while GPX4 (PHGPX) and SELENOP are monomers. GPX3 reduces diverse hydroperoxides, including hydrogen peroxide, tert-butyl hydroperoxide, and phospholipid hydroperoxide (24). GPX3 also possesses broad thiol specificity and uses TRX, dithiothreitol, and mercaptoethanol as reducing agents.

Se incorporated in Sec is specifically regulated via the synthesis of selenoproteins (28, 29). Sec is synthesized on tRNA using inorganic Se (Figure 1B). Servl-tRNA synthetase (SerRS) binds Ser to tRNA^{Sec}, which has an anticodon of UGA, and the hydroxyl residue of Ser undergoes phosphorylation. SEPHS2 produces selenophosphate (H₂SePO₃) from inorganic Se and ATP. SEPHS2 is a selenoprotein, suggesting the self-regulation of the Sec synthesis system. The Sec synthase catalyzes the formation of a selenol residue on tRNA using H₂SePO₃ and phosphorylated Ser-tRNA^{Sec} (Figure 1B). Inorganic Se, such as sodium selenite, is recognized as "Se" in mammals and is reduced, phosphorylated, and incorporated into the synthesis pathway of Sec. Sec derived from the diet is also recognized as "Se" and is converted to inorganic Se by Sec lyase (30) (Figure 1B). In contrast, SeMet, a methionine analog that contains Se instead of sulfur, is recognized as "Met" in mammals and is incorporated into proteins in the same manner as Met (31). SeMet is also metabolized by Met-metabolizing enzymes. Se in SeMet is recognized as "Se" when it is converted to Sec via metabolism by cystathionine β -synthase and cystathionine γ -lyase, which are also known as Cys-persulfide-producing enzymes (Figure 1B). SeMet is considered "masked Se," and the contents of SeMet in the diet influence the concentration of Se in blood. As described above, \sim 70% of "Se" is derived from Sec of SELENOP and GPX3, and the other 28% might be from SeMet and low molecular weight Se. Sec lyase is specific for Sec and provides "Se" to SEPHS2. Only through this pathway, Se is used for the synthesis of Sec on the tRNA^{Sec}.

SE TRANSPORT SYSTEM VIA SELENOPROTEIN P AND OTHERS

Serum free-culture of neurons and several cells requires Se addition, because SELENOP functions as a Se carrier in serum-containing culture (32, 33). SELENOP was identified as a survival-promoting factor for cultured neurons in serumfree medium in 1998, suggesting the Se transport system via SELENOP (32). The Se transport system of SELENOP has been described in *in vitro* experiments using human T lymphoma

Abbreviations: ApoER2, apolipoprotein E receptor 2; GPX, glutathione peroxidase; GSH, glutathione; LRP1, low-density lipoprotein receptor-related protein 1; PAH, pulmonary arterial hypertension; PASMC, pulmonary artery smooth muscle cell; PHGPX, phospholipid hydroperoxide GPX; SBP2, SECIS-binding protein 2; Se, selenium; Sec, selenocysteine; SECIS, Sec insertion sequence; SeMet, selenomethionine; SELENOP, selenoprotein P; SEPHS2, selenophosphate synthetase 2; TRX, thioredoxin; TRXR, thioredoxin reductase; 3'UTR, 3' untranslated region.



Jurkat cells and selenoprotein-deficient human serum prepared by the immobilized antibody for SELENOP and GPX3 (9). When cultured with SELENOP-depleted serum, and not GPX3-depleted or the control serum, the activity of GPX1 decreased to 17% compared with that of the control (**Figure 2A**). The activity of two other selenoproteins, GPX4 and TRXR, also decreased to 16 and 38%, respectively (9). When cultured with SELENOPdepleted serum, the whole-cell Se content also decreased to 19% compared with that of control cells. Time-dependent analysis revealed that the GPX1 activity of cells cultured with SELENOP-depleted serum was almost undetectable after 4 days (**Figure 2B**). The addition of 270 ng/ml purified SELENOP (SELENOP concentration of 5% human serum) resulted in the complete recovery of GPX1 activity. Thus, SELENOP functions as a major Se transporter in this culture system.

The comparison of SELENOP with other Se-containing materials as a Se supplier demonstrated that SELENOP was the most effective with a 50% effective dose (ED_{50}) of 5 nM (Se equivalent), followed by GPX3, sodium selenite, selenocystine,

SeMet, and albumin (9) (Figure 2C). The ED_{50} of the former three reagents was 25 nM, and that of the latter two was 300 and 500 nM, respectively. Ebselen had no effect up to 500 nM. These results suggest the effective Se transport system via SELENOP (9). Next to SELENOP, the inorganic Se and Sec group, which is recognized as "Se," is also a good source of Se. Third group SeMet and albumin, which is classified as "masked Se," is not effective, but it could function as a Se source in high concentration. The Se concentrations of human plasma used in the previous study was $1.5 \,\mu$ M, and the estimated Se concentration of SELENOP, GPX3, and the others were 750, 300, and 450 nM, respectively. Thus, it is considered that the effective Se transport system of SELENOP might be less functional in cells that have direct contact with plasma, and that it will be effective in the interstitial fluid where the plasma is diluted. It needs further characterization in the SELENOP concentration of the interstitial fluid. The biological significance of the effective Se transport system via SELENOP has been demonstrated by SELENOP KO mice (35, 36). The decrease in tissue Se in SELENOP KO mice has been reported in



FIGURE 2 [Se transport activity of SELENOP: (A) Effect of serum selenoprotein depletion on cellular selenoenzyme activities. The Jurkat cells were cultured for 3 days in a medium containing each human serum. (B) Effect of SELENOP on CGPX activity. The Jurkat cells were cultured in a medium containing SELENOP-depleted human serum, and cGPX activity was measured. After 6 days, purified SELENOP was added. (C) Effect of the addition of Se-containing materials on cGPX activity. In the presence of variable amounts of Se-containing materials, the Jurkat cells were cultured with SELENOP-depleted serum. (D) Immunohistochemical analysis of incorporated human SELENOP. Left panel. SELENOP-neutralizing monoclonal antibody (mAb) AE2 improved pancreatic β -cells and anti-human SELENOP Ab. Cell nuclei were stained with DAPI (blue). The distribution of human SELENOP in pancreatic β -cells was decreased in mice administered with AE2. Right panel. MIN6 cells were incubated with 10 µg/ml human SELENOP in the presence of AE2 and control IgG (500 µg/ml) for 48 h. Treated MIN6 cells were examined immunohistochemically using anti-human SELENOP Ab. [A–C, (9) with permission and modifications: 5051220115395, D, reference (34) with permission:CC-BY]. the brain, kidney, testis, and bone (35–38). The severe phenotype of SELENOP is spermatogenesis disorder, which could not be recovered by Se supplementation, while the other disorders might be partly recovered by Se supplementation, suggesting the compensatory role of the Se transport system other than SELENOP. It is also notable that SELENOP is contained in mothers' milk and has the function to supply Se to offspring (37). The effective Se transport system of SELENOP is mediated by its receptors, such as apolipoprotein E receptor 2 (ApoER2), megalin, and low-density lipoprotein receptor-related protein 1 (LRP1), which belong to lipoprotein receptors. In addition, Se transport of SELENOP has been observed in a receptor-independent manner (pinocytosis) (39).

THE RECEPTORS FOR SELENOPROTEIN P

Three kinds of SELENOP receptors have been identified, namely, ApoER2 (LRP8), megalin (LRP2), and LRP1. ApoER2 and megalin were first discovered by immobilized SELENOP affinity column chromatography (40, 41), while LRP1 was identified by the siRNA experiment on C2C12 myocytes based on the expression of LRPs (42). SELENOP transports Se to several tissues via these receptors. Based on the phenotype of each receptor KO mice, ApoER2 is associated with SELENOP uptake in the brain, testis, and bone; megalin with the kidney and brain; and LRP1 with the skeletal muscle. It has been reported that megalin mediates brain Se uptake and that Apoer2 mediates neuronal SELENOP uptake (43).

Similar phenotypes of SELENOP KO mice have been reported in the brain and testis of ApoER2 KO mice, indicating the biological significance of receptor-mediated uptake of Se in these tissues (35-37, 44). Similar phenotypes in the brain and testis have been reported in SELENOP^{$\Delta 240-361$} mice, in which the Secrich C-terminal domain of SELENOP had been deleted (34). The interaction between the C-terminal domain of SELENOP and the YWTD β -propeller domain of ApoER2 has been reported, and the importance of this interaction, particularly in maintaining Se levels in the brain and testis, has been manifested in the phenotypes of these KO mice (45). In the normal diet that contains enough Se (0.4 mg Se/kg diet), Se levels in other tissues such as the intestine and lung of SELENOP KO mice, except for the brain, testis, and bone, do not differ greatly from those in WT mice. Selenite in mouse chow is not a normal constituent of diet, but can be imported directly or after metabolism into SELENOP- and ApoER2-deficient cells. In the case of the brain of SELENOP KO mice, Se content was greatly affected by diet; 0.1 mg Se/kg diet induced a significant decrease in brain Se (\sim 50% of WT) and caused severe motor dysfunction, which needed humane endpoint (44). These observations suggest the role of the Se transport system via not only SELENOP but also SELENOP-independent systems in vivo. These observations indicate the role of SELENOP in the Se transport system, which is particularly effective to maintain homeostasis under the Sedeficient condition. This system might be significant to survive in the evolutionary process.

Megalin plays a crucial role in the reabsorption of SELENOP in the kidney, and increase in urine SELENOP has been shown in megalin KO mice (40, 43). Megalin is a large glycoprotein (~600 kDa), which possesses four large clusters of ligand-binding repeats stabilized with a disulfide bond. In the kidney, proximal tubule epithelial cells highly express megalin, indicating the physiological role of megalin in the reabsorption of SELENOP. LRP1 is identified as SELENOP receptor in the skeletal muscle (42). LRP1 is a super-macromolecule with a size of about 600 kDa and has a diverse set of ligands, such as amyloid β. Skeletal muscle has relatively low affinity for SELENOP, and the Se content does not change in SELENOP KO mice. However, the increase in SELENOP in the case of type 2 diabetes enhances the uptake of SELENOP via LRP1, which is related to the increase in insulin and exercise resistance (42). Thus, the SELENOP-LRP1 axis is an important therapeutic target for the cure of type 2 diabetes. Human embryo rhabdomyosarcoma RD cells express both ApoER2 and LRP1, which show low affinity for SELENOP uptake, and high amount of SELENOP is necessary for the uptake and use of Se in SELENOP (46). Treatment of RD cells with siRNA for either ApoER2 or LRP1 resulted in significant decrease in SELENOP uptake and increase in GPX1, suggesting both receptors coordinately work for SELENOP uptake. Diverse variants of ApoER2 have been known, while the relationship between ApoER2 variants and SELENOP uptake is unknown.

SE TRANSPORT SYSTEM VIA SELENOPROTEIN P

SELENOP possesses Se as Sec, which is covalently bonded to the polypeptide chain. To use the Se in SELENOP for selenoprotein synthesis, several biochemical steps are necessary. Based on previous reports, it has considered that SELENOP is incorporated into the cell, and then is degraded to amino acids in the lysosome (45). It is notable that the life span of incorporated SELENOP is long, which makes it possible to observe incorporated cellular SELENOP by *in vivo* and *in vitro* immunostaining (46) (Figure 2D). SELENOP is a glycoprotein, and the carbohydrate chains in SELENOP might function to prevent proteolysis in lysosomes. The generated Sec from SELENOP is cleaved with Sec lyase, converting to hydrogen selenide H₂Se, which is further phosphorylated to H₂SePO₃ with SEPHS2, as described above. The toxicity level of H₂Se is high, and the phosphorylation of H₂Se is important to prevent its toxicity. Notably, the necessity for Sec lyase in Se transport of SELENOP is not clear, because mice lacking Sec lyase do not have selenium deficient phenotypes, such as male sterility (47).

SELENOP is synthesized in peripheral tissues and SELENOPexpressing cells are present in several tissues (48). Based on serum SELENOP concentrations of liver-specific SELENOP KO mice, 60% of SELENOP is estimated to be derived from the liver and the remaining 40% from other tissues (49). SELENOP expression in the brain is significant for maintaining Se and selenoprotein levels, and Se concentrations in the brain are preserved in liverspecific SELENOP KO mice. SELENOP expression has been reported in several cell types of the brain, such as neurons and ependymal cells, that are responsible for cerebrospinal fluid production (50). SELENOP synthesized in the brain is incorporated into other brain cells and used to synthesize selenoproteins, which help maintain Se concentrations in the brain. This system is called the SELENOP cycle, and it retains selenoproteins in several tissues and cells (51).

ApoER2 is a significant mediator of the SELENOP cycle. The similar phenotype between SELENOP and ApoER2 KO mice implies the role of ApoER2 in this cycle. Details are not fully elucidated, but the role of ApoER2 as a mediator of signal transduction has been known, which is realized by pulmonary arterial hypertension (PAH) where the increased expression of SELENOP in pulmonary artery smooth muscle cells (PASMCs) forms lesions (52). PAH-PASMCs are proliferative, and the pulmonary artery is constricted/occluded by the abnormal proliferation of PAH-PASMCs. The decrease in SELENOP expression by SELENOP-siRNA treatment inhibits the proliferation of PAH-PASMCs, and these effects are mediated by ApoER2 (52). Interestingly, the proliferative effects of SELENOP were not explained by Se transport activity; namely, the addition of selenocystine did not reproduce this effect of SELENOP, and proliferation-promoting effects were observed by the overexpression of the mutant in which all Secs were substituted with Cys. The proliferative effects of increased SELENOP on PAH-PASMCs are considered to be mediated by the cell signal from ApoER2 and HIF-1. Se-independent biological effects of SELENOP-ApoER2 axis have been described in a study on PAH,

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and it is interesting to speculate about the possibilities to relate to other physiological and/or pathological conditions.

CONCLUSION

This review focuses on the Se transport system via SELENOP, particularly its molecular mechanism and role in Se metabolism. SELENOP is not a mere Se transporter. It plays the role of multifunctional protein to maintain cellular selenoproteins and regulate cellular redox homeostasis. Furthermore, the Se-independent role of SELENOP suggests the diverse biological and pathophysiological significance of this protein. Further research is necessary to understand the various roles of SELENOP.

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The author confirms being the sole contributor of this work and has approved it for publication.

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Conflict of Interest: The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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