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Maternal monosaccharide diets evoke cognitive, locomotor, and emotional disturbances in adolescent and young adult offspring rats

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Anxiety and depression are the most common mental disorders affecting people worldwide. Recent studies have highlighted that a maternal high-sugar diet (HSD) could be a risk factor for neurobehavioural dysregulations, including mood disorders. Increased consumption of added sugar in food such as refined fructose/glucose can increase the risk of metabolic disorders and impact susceptibility to mental disorders. Furthermore, a few papers have reported disabilities in learning and memory among offspring after maternal HSD, thus suggesting a relationship between maternal nutrition and offspring neurogenesis. In this study, we evaluated the impact of maternal monosaccharide consumption based on a glucose (GLU) or fructose (FRU) diet during pregnancy and lactation in adolescent and young adult offspring rats of both sexes on cognitive, locomotor, and emotional disturbances. Locomotor activity, short-term memory, anxiety-like and depressive-like behavior were evaluated in the offspring. We report for the first time that the maternal GLU or FRU diet is sufficient to evoke anxiety-like behavior among adolescent and young adult offspring. Moreover, we found that maternal monosaccharide diets lead to hyperactivity and depressive-like behavior in male adolescent rats. We also noticed that a maternal FRU diet significantly enhanced novelty-seeking behavior only in young adult male rats. Our novel findings indicated that the maternal monosaccharide diet, especially a diet enriched in FRU, resulted in strong behavioral alterations in offspring rats at early life stages. This study also revealed that male rats were more susceptible to hyperactivity and anxiety- and depressive-like phenotypes than female rats. These results suggest that maternal monosaccharide consumption during pregnancy and lactation is an important factor affecting the emotional status of offspring.

KEYWORDS

anxiety, behavior disorders, fructose, glucose, cognition, depression, hyperactivity

1. Introduction

Mental disorders are diseases characterized by a clinically significant disturbance in an individual's cognition, emotional regulation, or behavior that reflects a dysfunction in the psychological, biological, or brain developmental processes (1). Mental disorders are global problems in modern society, and they affect people across genders and ages, imposing significant barriers to a person's ability to function normally. In 2019, one in every eight people, or 970 million people worldwide, lived with a mental disorder, among which anxiety and depressive disorders were the most common (2). On the other hand, it was estimated that in the United States, young adults aged 18–25 years had a higher prevalence of serious mental illness than adults aged 26–49 years and older (3). The development of mental disorders is due to biological, psychological, and social factors, and these risk factors may manifest differently at different ages and stages in life, with some of these risks appearing as early as the prenatal period (4). The Developmental Origins of Health and Disease hypothesis indicates that fetal exposure during a critical period of development and growth to external (environmental) and internal (maternal) factors can significantly impact the short- and long-term health of the offspring (5). Maternal diet during pregnancy and lactation can lead to metabolic or neurological diseases among offspring (6, 7).

Foods that contain sugar such as glucose (GLU) and fructose (FRU) are an important source of energy for cells and crucial for the maintenance of the body's biochemical processes. The brain consumes \sim 20% of GLU-derived energy, making it the main consumer of GLU (8). Both GLU and FRU act as sweeteners and added sugars in many food products, such as FRU in highfructose corn syrup (HFCS) or GLU in sugar-sweetened beverages (SSB). Importantly, youth and adults consume calories from added sugar in processed food (9, 10). Moreover, many adults exceed the daily intake of free sugars, which is limited by the World Health Organization to <10% of one's total energy intake (11). More than 10% of Americans' daily calories come from FRU consumption, which has potentially more harmful effects on the body than other major caloric sweeteners (12). Clinical observations have indicated that sugar-rich foods are desirable during pregnancy and were consumed too often during the study period, suggesting that many pregnant women were exposed to the adverse effects of a high-sugar diet (HSD) (13).

In a recent review, perinatal and postnatal HSD (such as sugar, sucrose, fructose, or glucose) consumption was considered a potential risk factor for neurobehavioural dysregulations such as mental disorders (14). Although several preclinical studies have evaluated the effects of maternal HSD on offspring behavior (15-18), most of these studies were performed using male or female rodents independently. It is also worth adding that most preclinical studies have focused on the relationship between postnatal HSD consumption and behavioral disturbances (19-24). However, maternal diet in the prenatal period also plays a crucial role in offspring neurodevelopmental disorders (25-27). Human observational studies have shown that stress-induced sugar or SSB consumption was correlated with increased anxiety and depression in adolescents and adults (28-31). Furthermore, HSD and sugar itself can be addictive and can predispose an individual to the risk of food addiction (32). Finally, sugar intake includes numerous neural mechanisms within the mesocorticolimbic system, hypothalamic orexigenic and anorexigenic pathways, and hypothalamic-pituitary-adrenal axis, all of which are strongly correlated with mental disorder development (33).

An increase in sugar consumption and several cognitive, locomotor, and emotional disturbances after maternal HSD suggest an interaction between these variables. However, there are no studies that have reported the effect of particular monosaccharides in the maternal diet on behavioral phenotypic changes in both sexes of offspring at later stages of life. Similarly, there are few preclinical studies have specifically investigated the impact of maternal HSD intake on offspring cognition and anxiety-like or depressive-like behavior development, while the impact of a maternal high-fat diet has been widely studied in this context. Hence, this study aimed to examine the effect of a maternal GLU or FRU diet during pregnancy and lactation on behavioral outcomes in adolescent and young adult unrelated offspring rats of both sexes.

2. Materials and methods

2.1. Experimental animal and maternal diets

Wistar rats obtained from the licensed animal Charles River breeder (Sulzfeld, Germany) were housed in standard plastic rodent cages in an animal breeding room at $21 \pm 2^{\circ}$ C and $40 \pm 10\%$ humidity with a 12-h light-dark (LD 12:12) cycle (lights on at 6:00 a.m.). Animals had free access to water and food *ad libitum*. Twenty-four virgin female rats (201–225 g), after the acclimatization period (14 days) and during the proestrus phase (the oestrous cycle phases determined by daily vaginal smears), were mated with six randomly assigned males (226–250 g) until the presence of sperm in the smears was confirmed. Pregnant females were individually housed and randomly assigned to one of three different experimental condition groups: standard control/VRF1 diet (SD), modified glucose (GLU) diet, or modified fructose (FRU) diet (Table 1). All diets were delivered by Special Diets Services (SDS; London, UK).

During pregnancy (21 days) and lactation (21 days), dams were given ad libitum access to water and food, depending on the diet group (Figure 1). One day after birth, the pups were weighed, and the litter sizes were counted and normalized to 8-10 pups. Next, at postnatal day (PND), 21 offspring were weaned and separated according to sex, marked, housed 5 per cage, and switched to SD. At the same time, we took two offspring sets (selected from 7 to 8 different dams and six different males) to minimize the parental genetic influence and reduce individual offspring predisposition by increasing their variability in cohorts (34). For behavioral studies, we used one of two subsets of each set randomly consisting of eight male and eight female rats for each diet. Two independent offspring subsets were used to assess spontaneous behavior in the early life stage periods without exposure to earlier experimental conditions. Female and male rats were tested separately. All experiments were carried out in conformity with the European Union Directive (2010/63/EU), Polish Act on the Protection of Animals (Dz.U. z 2020 r. poz. 638), and with approval of the Local Ethics Commission (Kraków, Poland; approval numbers 18/2021 and 54/2021) with the three Rs rule.

2.2. Offspring behavioral tests

2.2.1. Locomotor activity

Locomotor activity (LMA) was recorded for each nonhabituated rat using the procedure described previously by Wydra et al. (35). Briefly, rats at PND 27 (subset from set I) and PND 59 (subset from set II) were placed individually in Opto-Varimex

Diet		Control/VRF1 (SD)	Glucose (GLU)	Fructose (FRU)
Carbohydrates	Pectin (%)	1.37	0.00	0.00
	Hemicellulose (%)	8.76	0.13	0.13
	Cellulose (%)	3.95	6.40	6.21
	Lignin (%)	1.06	0.00	0.00
	Starch (%)	35.41	0.00	0.00
	Sugar (%)	4.64 (from sucrose)	53.76 (from glucose)	54.35 (from fructose)
Protein (%)		19.11	19.13	18.56
Fat (%)		4.75	5.06	4.91
Total energy (kcal/g)		3.40	3.44	3.43

TABLE 1 Composition of main macronutrients (shown as a percentage of energy) and energy value of maternal experimental diets during pregnancy and lactation periods.

The glucose (GLU) and fructose (FRU) diets are synthetic mimics of the control (SD) diet. The type of sugar was carefully modified and formulated to match the VRF1 diet.



 $(43 \times 43 \text{ cm})$ cages (Columbus Instruments, Columbus, USA). Spontaneous locomotor activity, measured as horizontal activity, was recorded for 2 h using the Auto-Track system (Columbus Instruments, USA) every 5-min trial and presented as the distance traveled (cm).

2.2.2. Novel object recognition

To evaluate the ability to recognize a novel object (NOR) in the environment and working short-term memory in rats at PND 29 (subset from set I) and PND 61 (subset from set II), the novel object recognition test described previously by Gawlińska et al. (36) was used. On the first day, after 1 h of acclimatization in the experimental room, rats were placed separately for 10 min in the center of the empty, non-transparent plastic box ($57 \times 37 \times 30$ cm) arena for habituation. Next, for 5 min, the animals were presented with two identical objects—A, as a familiarization phase [A + A]. After a 1 h break in the home cages, rats again were placed in the same plastic box for 5 min and presented two objects, the familiar (F)—A and the first novel (N1)—B object, as a recognition phase I [A + B]. On the second day, 24 h after the familiarization phase, the rats were placed in the arena for 10 min and again presented with two objects: the previous (A) and the second novel (N2) (C) object as a recognition phase II [A + C]. Active object exploration was defined as time spent sniffing, touching, and close approach, not including the time spent sitting on the objects. The percentage of time spent exploring the novel object relative to the total time spent exploring both objects was measured using the recognition index (RI) formula: RI = $TN/(TN + TF) \times 100$ (%), where T was the time spent exploring the novel (N1 or N2) and familiar (F) objects. An RI above 50% indicated novel object preference, below 50% indicated familiar object preference and 50% indicated no preference (37, 38). The following objects were presented during the test: black (F) and orange (N1) metal cans (13.5 cm high, 5 cm in diameter) and a violet glass block (6 cm high, 3–5 cm in diameter) with a green plastic cone (N2; 2 cm high, 3–5 cm in diameter). Objects were placed on two opposite sides of the box, 13 cm from the center of the arena and 22 cm from each object. Plastic boxes and objects were cleaned using 20% ethanol to minimize olfactory cues before the next round. To reduce the stress of a bright lighting room, areas were illuminated with 170–230 lux light (1 m above the box). The test was video recorded by one investigator, while the results were manually extracted from videos by two independent blinded assessors.

2.2.3. Elevated zero maze

Anxiety-like behavior in rats was assessed using the elevated zero maze test (EZM) (39). The annular, black platform maze (105 cm in diameter, 10 cm path wide) consisted of four quadrants: two opposite open (with 1 cm high walls) arms and two opposite closed (with 25 cm high walls) arms, which were elevated to 65 cm above the floor. The rats were placed individually, alternately in one of the two closed arms of the maze, and a 5-min test was recorded. Behavioral measures comprised the time (s) spent in and the number of entries into open areas, the frequency of head dips from the dark to the light zone, beyond the wall in the open zone, and the frequency of stretch postures from closed to open quadrants. The maze was illuminated by a light source (120-150 lux) suspended 50 cm from the maze path. The test was conducted in the experimental, dark room after 1 h of acclimatization, 5 min of habituation to the surface, and color of the maze in the EZM box for PND 31 (subset from set I) and PND 63 (subset from set II) rats. The maze was cleaned using 20% ethanol before the next trial. All tests were performed and manually assessed by two independent investigators who remained at an appropriate distance of 2 m from the maze in the experimental room.

2.2.4. Forced swim test

The modified forced swim test (FST) was used to assess offspring rats' responses to stress at PND 34 (subset from set I) and PND 66 (subset from II) (40, 41). On the first day (pretest), after 1 h of acclimatization, adolescent rats were placed individually in glass-transparent cylinders (41 cm high, 20 cm diameter) containing clean water (25 \pm 1°C) at a depth of 30 cm (preventing touching the bottom) for 15 min. Then, all rats were removed from the water, dried, warmed, and returned to their home cages. Twenty-four hours after the pre-test, the rats were retested for 5 min under the same conditions, where one investigator recorded each animal's activity using a digital camera. Each cylinder was emptied and cleaned before the test for the next rat. Next, two independent blinded assessors manually extracted from the video the time spent swimming, climbing, and immobility. Immobility time was observed when the rat floated in water without struggling and made only the movements necessary to keep its head above water. Swimming time was defined as active swimming motions (displaced body around the cylinder), more than necessary to merely maintain head above water. Climbing time was measured when the rat was climbing, making active movements with forepaws in and out of the water.

2.4. Statistical analysis

All experimental data are expressed as the mean \pm SEM (standard error of the mean). Statistical analyses were performed with Statistica 13.3 (TIBCO Software Inc., Palo Alto, CA, USA) and visualized in GraphPad Prism 9.4.1 (GraphPad Software, Inc., San Diego, CA, USA). The Shapiro-Wilk test and Levene test were used to test the normality and equal homogeneity of variance, respectively. For normally distributed data, twoway analyses of variance (ANOVA: diet + gender + diet × gender; F statistic) were performed. Alternative two-way ANOVA with Welch's correction (W statistic) was used for unequal variances. For non-normally distributed data, a non-parametric Kruskal-Wallis test by ranks (H statistic) was performed. In significant differences between diet (compared to SD), either parametric Dunnett's and Dunnett's T3 or non-parametric Dunn's multiple comparisons post hoc tests were performed. For the sex-specific changes, parametric Šidák's or Dunnett's T3 multiple comparisons post hoc tests were performed. "N" corresponds to the number of individuals. p < 0.05 was considered statistically significant.

3. Results

3.1. Effect of monosaccharide diets on body weight and caloric intake

During the experiment, dams showed a significant main effect of the period $[F_{(1.634,32.67)} = 106.90; p < 0.0001]$ and interaction $[F_{(18,180)} = 2.346; p = 0.0024]$ during 5-day measurements of body weight (Figure 2). During pregnancy (on day 15) and lactation (on day 5), SD dams had 8% and 9% higher body weights, respectively, than FRU dams (Figure 2). Moreover, we observed a main effect of the period $[F_{(1,40)} = 159.50; p < 0.001]$, diet $[F_{(2,40)} = 6.44; p = 0.0038]$ and interaction $[F_{(2,40)} = 7.13;$ p = 0.0023] on total calorie intake (Figure 3A). The GLU and FRU dams showed 23 and 27%, respectively, less total calorie intake than SD dams in the lactation period (Figure 3A). The total calorie intake was similar in the pregnancy period between the groups. Assuming that 1 g of sugar has 4 kcal, and the percentage of sugar in the diet has been described (Table 1), we also assessed the contribution of calories from the sugar in the total calorie intake (Figure 3B). Here, period $[F_{(1,40)} = 164.70;$ p < 0.001] and diet [$F_{(1,40)} = 4.87$; p = 0.0128] effects were observed. Conversely, for total caloric intake, both GLU and FRU dams showed 31% higher calorie intake than SD dams in the pregnancy period (Figure 3B). No changes were observed in the lactation period. Although we observed different litter sizes between dams in each diet, these changes were not statistically significant $[H_{(2,N=23)} = 1.32, p = 0.5146;$ Figure 4A]. In turn, we observed significant differences in pup weight $[H_{(2,N=224)}]$ 30.69, p < 0.0001; Figure 4B] between diet groups. The GLU and FRU pups weighed less by 7 and 11%, respectively than the SD



FIGURE 2

Effects of maternal glucose (GLU) or fructose (FRU) diet intake on dam body weight during pregnancy and lactation. At 15 days of pregnancy and 5 days of lactation, FRU dams showed less body weight than SD dams. N = 8 dams/SD and GLU, 7 dams/FRU. Data are comparable to the SD groups (*p < 0.05, two-way repeated measure ANOVA followed by Dunnett's multiple comparison test).



offspring (Figure 4B). Moreover, adolescent FRU male and GLU female offspring displayed 7 and 8%, respectively, higher body weights than SD rats [$W_{(5,18.66)} = 5.73$, p = 0.0023; Figure 4C]. Both GLU male and female young adult offspring had 6% higher

body weight than SD offspring. Additionally, SD, GLU, and FRU female rats demonstrated 18%, 19%, and 20% lower body weight, respectively, than male rats in the same diet group $[W_{(5,19.14)} = 64.52, p < 0.0001;$ Figure 4D].



Effects of maternal glucose (GLU) or fructose (FRU) diet on litter size and offspring weight. (A) Litter size of each dam. Maternal diet affects litter size individually, but in total, no significant difference was observed, N = 8 dams/SD, 7 dams/GLU and FRU (Kruskal–Wallis tests), (B) Pups weight one day after birth (at PND2). GLU and FRU offspring rats weighed less than SD pups. N = 87 pups/SD, 74 pups/GLU and 63 pups/FRU. Data are comparable to the SD groups (***p < 0.001, ****p < 0.001, Kruskal–Wallis tests followed by Dunn's multiple comparison test). (C) Body weight of adolescent (at PND27) offspring. FRU male and GLU female adolescents had higher body weights than SD offspring. (D) Body weight of a young adult (at PND59) offspring. Both GLU male and female young adults had higher body weights than SD offspring. Young adult females had reduced body weight compared to males. N = 8 rats/group. Data are comparable to the SD groups (*p < 0.05, two-way ANOVA with Welch's correction followed by Dunnett's T3 multiple comparison test) or between sexes in the same group ($^{\wedge\wedge}p < 0.001$, $^{\wedge\wedge\wedge}p < 0.0001$, two-way ANOVA with Welch's correction followed by Dunnett's T3 multiple comparison test).

3.2. Effects of maternal HMD on offspring locomotor activity

First, we assessed spontaneous locomotor activity measured as distance traveled in adolescent (at PND 27) and young adult (at PND 59) offspring (Figure 5). Adolescent rats showed a significant main effect of diet $[F_{(2,42)} = 9.05; p = 0.0005]$ and sex $[F_{(1,42)} =$ 20.19; p < 0.0001], as well as an interaction effect [$F_{(2,42)} = 3.88$; p= 0.0283] in 5-min trial measurements (Figure 5A). Both GLU and FRU male offspring showed 22 and 44% higher locomotor activity, respectively than SD male offspring. Moreover, FRU male rats demonstrated 29% higher activity than female rats. Likewise, after a 120-min trial (Figure 5B), GLU and FRU adolescent male offspring displayed 42 and 48%, respectively, enhancement in spontaneous locomotor activity $[H_{(5,N=48)} = 18.57, p = 0.0023]$ compared with SD male rats. No significant interaction effects by diet and sex were observed in young adult offspring during 5- $[H_{(5,N=48)} = 3.57, p]$ = 0.612; Figure 5C] and 120- $[F_{(2,42)} = 0.81, p = 0.447;$ Figure 5D] min trials.

3.3. Effects of maternal HMD on offspring memory

The effect of maternal diets on short-term memory and the ability to recognize a novel object in adolescent (at PND29) and young adult (at PND61) offspring were evaluated using the NOR test (Figure 6). In adolescent offspring, SD, GLU, and FRU females displayed 24%, 26%, and 28% higher recognition indices, respectively, than male rats $[W_{(5,19,11)}]$ = 9.56, p = 0.0001] 1 h after the familiarization phase (Figure 6A). Moreover, no interaction effect was observed 24 h after the familiarization phase $[F_{(2,42)} = 2.71, p = 0.078;$ Figure 6B].

In young adult rats 1 h after the familiarization phase, only the sex effect was detected $[F_{(1,42)} = 7.39, p = 0.0095]$ without significant differences between male and female rats (Figure 6C). On the other hand, 24h after the familiarization phase, significant main diet $[F_{(2,42)} = 11.35, p = 0.0001]$ and interaction $[F_{(2,42)} = 3.47, p = 0.040]$ effects were



followed by Šidák multiple comparison test).

noticed. Interestingly, only young adult FRU male rats manifested significant (26%) increases (Figure 6D) in the recognition index in comparison to SD male rats. After the 24-h familiarization phase, both adolescent and young adult offspring had RIs above 50%, suggesting a preference for the novel object.

3.4. Effects of maternal HMD on offspring anxiety-like behavior

We assessed anxiety-like behavior in offspring using an EZM test (Figure 7). In adolescent offspring, the time spent in an open area (Figure 7A) in FRU male and female offspring was 77% lower



FIGURE 6

Maternal glucose (GLU) or fructose (FRU) diet changed the ability to recognize the new object in adolescent (at PND29) and young adult (at PND61) offspring. (A) Adolescent SD, GLU, and FRU female but not male rats showed a greater increase in the recognition index (RI) during the novel object recognition (NOR) test after a 1 h familiarization phase. FRU males and females were characterized by a higher time spent with novel objects than SD and GLU offspring. (B) Young adult SD, GLU, and FRU female, but not male, rats showed a decrease in RI after the 1 h familiarization phase. (C) After the 24-h familiarization phase, adolescent offspring showed no difference in RI. (D) Young adult FRU males showed a significantly increased recognition index in NOR after the 24-h familiarization phase. A larger number of offspring had an RI above 50%, which indicated a preference for the new object. N = 8 rats/group. Data are comparable to the SD groups (****p < 0.0001, two-way ANOVA followed by Dunnett's multiple comparison test).

than that in SD rats [$H_{(5,N=48)} = 26.25$, p < 0.0001]. FRU male and female offspring had reduced entrance to the open area (Figure 7B) by 74% [$F_{(2,42)} = 27.83$, p < 0.0001] and had reduced stretched postures (Figure 7C) by 40% and 46% [$F_{(2,42)} = 16.22$, p < 0.0001], respectively, compared to SD offspring. In addition, FRU male, as well as FRU and GLU female offspring, showed significant (36%, 61% and 25%, respectively) decrease in head dips (Figure 7D) compared to SD [$F_{(2,42)} = 28.63$, p < 0.0001]. The average number of head dips in FRU female offspring was 38% lower than that in male offspring [$F_{(1,42)} = 4.89, p = 0.032$].

Furthermore, FRU and GLU young adult male offspring demonstrated significant decreases in time spent in the open area (Figure 7E) by 43 and 59%, respectively, compared to SD male offspring [$F_{(2,42)} = 3.70$, p = 0.033]. Moreover, FRU and GLU male rats entered the open area (Figure 7F) in 30 and 57%, respectively, less time than SD offspring [$F_{(2,42)} = 7.99$, p = 0.001].



Additionally, SD female rats showed 39% fewer entries into the open area (Figure 7F) than male rats [$F_{(1,42)} = 4.58$, p = 0.038]. No significance was observed in young adult stretched postures [$W_{(5,19,20)} = 2.45$; p = 0.070; Figure 7G]. However, FRU female offspring showed 31% lower head dips (Figure 7H) than male offspring [$F_{(2,42)} = 3.48$, p = 0.039].

3.5. Effects of maternal HMD on offspring depressive-like behavior

Finally, we examined offspring rat depressive-like behavior in the forced swim test (FST; Figure 8). In adolescent offspring, GLU and FRU males displayed enhanced (18 and 29%, respectively) time spent immobile [$W_{(5,19,32)} = 6.49$, p = 0.001] compared with SD males (Figure 8A). Additionally, compared to controls, FRU male offspring showed 24% decreased swimming behavior [$H_{(5,N=48)} =$ 16.25, p = 0.006; Figure 8B]. Moreover, GLU male offspring and both FRU male and female offspring demonstrated a reduction (by 24%) in climbing time [$F_{(2,42)} = 6.41$, p = 0.003] compared with the SD groups (Figure 8C).

In young adult offspring, significant diet $[F_{(2,42)} = 5.48, p = 0.007]$ and interaction $[F_{(2,42)} = 4.59, p = 0.015]$ effects were observed for immobility time. Young adult GLU female offspring

showed 16% less immobility than the SD group and a 17% decrease in comparison to GLU male offspring (Figure 8D). SD female rats had a 32% reduction in swimming time compared with SD male rats [$W_{(5,18,11)} = 4.13$, p = 0.011; Figure 8E]. Moreover, only FRU female offspring showed reduced climbing time by 55% [$H_{(5,N=48)} = 21.55$, p = 0.0006; Figure 8F] compared to SD female offspring.

4. Discussion

The present study showed changes in maternal monosaccharide diet intake and energy expenditure during pregnancy and lactation. For the first time, we presented significant associations between maternal FRU or GLU diet and cognitive, locomotor, and emotional dysregulation in two unrelated offspring cohorts.

4.1. Monosaccharide diet influences body weight and caloric intake

Our study illustrated an alteration in the dam's body weight across a high-GLU or high-FRU diet during pregnancy and lactation. Indeed, FRU female rats had significantly lower body weights than GLU and SD female rats during pregnancy. However, the FRU dam's weights oscillated with the GLU mother's weight



Maternal glucose (GLU) or fructose (FRU) diet display depressive-like behavior in adolescent (at PND34) and young adult (at PND66) offspring. (A) GLU and FRU diets changed adolescent male vulnerability to depressive-like behavior assessed as increased time spent immobile. (B) FRU adolescent male offspring showed decreased swimming time. (C) GLU and FRU adolescent male and FRU adolescent female rats reduced the time spent climbing. (D) Young adult GLU female rats demonstrated a reduction in time spent immobile in comparison to SD females and GLU males. (E) Maternal GLU and FRU diets do not change swimming time in young adult offspring SD female offspring presented a decrease in time spent swimming compared to SD males. (F) Young adult FRU female offspring spent less time climbing. N = 8 rats/group. Data are comparable to the SD groups (*p < 0.05, **p < 0.01, two-way ANOVA with or without Welch's correction or Kruskal–Wallis tests followed by Dunnett's, Dunnett's T3 or Dunn's multiple comparison test, respectively).

during the lactation period. Moreover, neither of these changes correlated with the initial dam's body weight on the first day of pregnancy and lactation. Interestingly, there was no difference in total caloric intake during pregnancy, but as expected, GLU and FRU mothers received more calories from monosaccharides. In contrast to SD dams, GLU and FRU mothers consumed fewer calories during lactation, but no differences in calories from total sugar intake were observed during this period. We found that SD dams, which had a low percentage of monosaccharides in their diet (mainly from polysaccharides and disaccharides), sought to compensate for sugar calories by overeating only during lactation. These changes prove the high energy expenditure of mothers in these two different periods, where sugar is a crucial energy source for the growth and development of the fetus and later for the production of milk (42). Interestingly, GLU and FRU dam's litter sizes did not statistically differ. In turn, the postnatal weights of pups were significantly lower in GLU and FRU dams, indicating disturbances in the fetal stage of the pup's growth and development. Nevertheless, the GLU and FRU adolescent and young adult offspring's body weights during the experiment either balanced or were higher than the body weight of SD offspring. The observed sex-specific differences in body weight between diet groups were typical for any diet experiment (43).

Alternation in offspring body weight during the experiment could be due to maternal metabolic dysregulation during pregnancy and lactation or offspring fetal metabolic reprogramming by maternal GLU and FRU intake. Wistar rat dams fed 10% FRU in the water had increases in blood insulin, triglycerides, and GLU during pregnancy with a reduction in placental GLU transporter type one gene expression (44). Furthermore, FRU intake during pregnancy and lactation increased GLU transporter type five gene expression and glycogen levels with a reduction in fructokinase gene expression in the Wistar rat liver and caused sex-dependent changes in lipid metabolism in offspring (45). Moreover, Sprague-Dawley offspring rats had a higher postnatal body weight after consuming the maternal 13 or 40% water FRU solution (46). Additionally, in C57BL/6J dam mice, a 20% (w/v) FRU solution increased plasma triglycerides and cholesterol levels during pregnancy and predisposed offspring to obesity and metabolic dysfunction by postnatal high-fat diet intake (47). In the same mouse strain, pups after maternal GLU or FRU intake before and during pregnancy had less fetal body weight that was enhanced after weaning and correlated with fat mass gain compared to control offspring (48, 49). The observed sugar-dependent metabolic dysregulation was strictly associated with the different GLU and FRU biodegradation mechanisms. GLU directly enters glycolysis in each body cell, whereas FRU must be converted to glycolysis GLU byproducts in the liver as the energy source mainly for the citric acid cycle. Sugar overconsumption leads to fatty acid and de novo triglyceride biosynthesis, contributing to the development of overweight and obesity. GLU is mainly stored as glycogen in the liver and muscle, while much more fat is formed from FRU and stored peripherally. In contrast to GLU, FRU is the substrate of uric acid production, does not have many intracellular transporters, and is non-regulated by pancreatic hormones (50, 51).

4.2. Monosaccharide diet-induced hyperactivity behavior

In LMA measurements, we demonstrated that a maternal monosaccharide diet significantly disrupted spontaneous locomotor activity in offspring. For the first time, we showed that perinatal FRU and GLU intake increased the distance traveled measured during 5- and 120-min trials in male adolescents compared to male control SD rats. However, in our study, locomotor activity alterations observed in adolescent males were not manifested in another cohort of female and young adult offspring rats. Our study extends the previous observations in another research group using different rat strains. In their study, male Sprague-Dawley offspring rats (at PND50) after maternal 30% FRU-enriched water during pregnancy and lactation had higher vertical activity than females and significantly higher stereotypic behavior than control males in the 5-min open-field test (OFT) (16). Similarly, ICR mouse offspring of both sexes exposed to maternal short-term (E6-E15) dietary manipulations with FRU (30% w/v) supplementation had a specific greater increase in total distance measured in the 20-min OFT compared to the control and sucrose (30% w/v) groups (17). Nevertheless, the effect of maternal HSD on offspring locomotor activity was measured after animal habituation in the experimental cage. Moreover, the dams were fed standard rodent chow with sucrose and FRU supplementation in water. Thus, we measured acute spontaneous locomotor activity in two non-habituated age-independent cohorts at 5- and 120-min trials after maternal FRU- and GLU-enriched diets. On the other hand, it has been shown that chronic postnatal sucrose (25% w/v) or FRU (55% in diet and 10-15% w/v) consumption also increased the distance traveled in the 10-min OFT in male rats (19, 52) and male mice (20).

Hyperactive behavior in rodent studies, characterized by increased or constant movement, is considered a symptom of attention deficit hyperactivity disorder (ADHD) diagnosis in humans. In fact, in cohort and case-control trials, chronic intake of SSBs, soft drinks, and sugar binging enhanced the risk of hyperactivity behavior and ADHD symptoms in children (53). Moreover, hyperactivity and ADHD more frequently occur in boys (54, 55). The mechanism of maternal sugar action on offspring hyperactivity behavior and ADHD development is not yet well-known. Maternal malnutrition or overnutrition during pregnancy and lactation might induce neurodevelopmental effects in offspring. In mouse offspring, maternal sucrose (9 g/kg/day) consumption in pregnancy induces ADHD-like symptoms and leads to a reduction in dopamine receptors (D1, D2, and D4) and increased dopamine active transporters (17). Moreover, the higher duration of the active state has been correlated with higher expression of dopamine receptors in the offspring rat hippocampus after the maternal FRU diet (46).

4.3. Monosaccharide diet-prone memory impairment

In the NOR test, only the FRU diet affected recognition memory associated with short-term memory in response to novel stimuli. In comparison to the SD group, the maternal FRU diet enhanced (by 26%) the memory recognition index in young adult male rats 24 h after familiarization recognition phase II. Moreover, both young adult offspring had RIs above 50% related to a novel but not familiar object preference. Additionally, sex-specific differences were observed in recognition phase I. Female rats had a higher recognition index than male adolescent rats, but these changes were not observed in young adult offspring. In the present study, a higher recognition index observed in FRU young adult male rats could be associated with short-memory impairment or noveltyseeking behavior as a higher ability to perceive novel signals from the environment. These findings are consistent with an earlier report that a maternal HSD (with 40% sucrose) enhanced the recognition index 24 h after the familiarization phase in the twoday NOR test in young adult (at PND65) offspring of both sexes compared to control rats (36). Our results may indicate that recognition memory may not be as sensitive as spatial memory to dietary intervention. Other studies have shown that maternal HSD (with 64% sugar) decreased the preference index in the four-day NOR test in adolescent (at PND28) male Wistar rats. Moreover, a decreased novel location preference index trend was observed in adolescent and young adult (at PND70) HSD male offspring (15). In addition to HSD, a maternal high-FRU (60%) diet resulted in longer escape latency to find the hidden platform of the Morris water maze in adult female offspring, which was reversed by enriched housing (18). Furthermore, maternal FRU supplementation during pregnancy and lactation decreased latency time in FRU Sprague-Dawley offspring rats of both sexes in the passive avoidance learning test (16). A more recent preclinical study indicated that early exposure to a cafeteria diet during pregnancy and lactation impaired the performance of long-term memory tasks in adult (at PND120) male Wistar rats (56). The controversy in the findings of different maternal HSD studies may be explained by the different duration of NOR schedules, different recognition indexes measuring non-identical maternal diet composition, and offspring diversity (age, gender, and rat strain effect).

It has been well-documented that animals exposed to postnatal HSD intake had enhanced novelty-seeking behavior. Male Sprague-Dawley rats after 10% sucrose (w/v) intake for 24 days displayed an increase in exploration time with the novel object but not with a familiar object in comparison to control males (21). Moreover, 7.9% sucrose intake for 12 months significantly prolonged the time spent exploring a novel object in male Sprague-Dawley rats in the object recognition task (57). Furthermore, after 30% (w/v) FRU feeding for 9 weeks, male Swiss mice showed a higher recognition index than females and control males (22). Interestingly, female Wistar rats exposed to oral FRU (15%) supplementation for 24 weeks in a high fat-low protein diet showed increased time spent in the target quadrant and frequency of entries into the target quadrant in the Morris water maze (58). Despite the findings from these studies, several other studies have reported memory impairment after chronic, postnatal HSD consumption. Thus, postnatal sucrose consumption decreased the discrimination ratio measured in NOR (59), demonstrated poorer spatial memory in the Morris water maze (60), and decreased the amount of time exploring a novel object (61, 62) in rats.

Sugar and SSB consumption in humans influence memory and learning (63, 64). For instance, in a cross-sectional analysis of school children, higher consumption of SSB was associated with poorer performance on executive function and a high risk of executive dysfunction in children (65). Moreover, a higher intake of total sugars significantly impacted the lower prevalence of cognitive impairment scores in adult participants (66). Memory and learning processes are controlled by the hippocampus; thus, HSD can modify its neurodevelopment, neuronal plasticity, and metabolism. In a previous study, 12 weeks of maternal sucrose consumption altered episodic and spatial memory, which was correlated with a reduction in glial fibrillary acidic protein- and Nestin-expressing early-phase neurogenesis cells in the male mouse hippocampus (20). Moreover, FRU, but not GLU, consumption evoked memory impairment in rats and was associated with decreased brain-derived neurotrophic factor in the hippocampus (67). Additionally, male Wistar rats that received an FRU diet for 3 weeks showed increases in glial fibrillary acidic protein, FRU transporter (Glut-5), and tumor necrosis factor-alpha and decreases in synaptophysin, synaptotagmin, and postsynaptic density protein-95 protein levels in the frontal cortex associated with short-memory functions (68). Moreover, adolescent, but not adult, male Sprague-Dawley rats who consumed HFCS (11% w/v) for 30 days showed reduced correct hole investigation in the Barnes maze test with an increase in proinflammatory cytokine (IL-1ß and IL-6) protein expression in the dorsal hippocampus (69). A recent paper also linked learning and memory impairment with the downregulation of the Wnt/β-catenin signaling pathway and changes in nervous system development gene expression in the hippocampus of offspring rats after maternal FRU intake (70).

4.4. Monosaccharide diet-induced anxiety-like behavior

For the first time, we demonstrated that maternal monosaccharide diet intake was sufficient to evoke anxietylike behavior symptoms in offspring rats. As observed in our study, maternal FRU diet significantly evoked a decrease (by 77%) in time spent in the EZM with a reduced number of entries into the open area, diminished stretched postures, and head dips in adolescent offspring rats of both sexes. To be precise, the specificity of anxiety-like behavior results from the 5-min EZM test occurring in adolescent FRU offspring, especially in males, was associated with 5-min hyperactivity observed in the same rats in the LMA. Moreover, decreased time spent in the open area and reduced number of entries into the open area also manifested in unrelated young adult FRU male but not female rats. Interestingly, anxietylike behavior appeared after the maternal GLU diet in young adult males, but it was not linked with previous GLU intake in adolescent offspring. Our results indicate that a maternal monosaccharide diet can predispose to anxiety-like disorders and/or manifest anxiety-like symptoms across the offspring's lifespan that were observed in male offspring. Similar to our findings, a few previous studies have shown that offspring exposure to a maternal FRU diet evokes anxiety-like behavior, but only when combined with saline treatment (71) or salt supplementation in the diet (72). Furthermore, maternal supplementation with 30% FRU decreased the exploratory activity of offspring of both sexes compared to control groups in the 5-min OFT (16). A recent study also showed that offspring (males and females together), after maternal 13 and 40% FRU in water intake, had higher active state duration and anxious state duration in a 5-min OFT (46).

As mentioned, our study has provided the first experimental evidence suggesting that the maternal monosaccharide diet induces anxiety-like behavior in the offspring. We also demonstrated ageand sex-dependent differences in behavioral phenotypes following the FRU or GLU diet. Our results complement recent studies that have established the relationship between postnatal sugar intake and anxiety-like behavior occurrence. Thus, male rats supplemented with 2 h of access to 10% sucrose (w/v) for 28 days demonstrated a decrease in time spent in the center area of the elevated plus maze (EPM) (23). In addition to sucrose, consumption of 30%, but not 15%, of FRU solution (w/v) for 9 weeks decreased the time spent in the open arms of the EPM in adult (at PND152) Swiss mice of both sexes (22). Likewise, chronic 18 weeks of FRU (23% w/v) feeding caused reduced entries and time spent in open arms of EPM (24), and preadolescent (at PND92) but not adult (at PND132) male rats fed a 55% high-FRU diet for 8-10 weeks reduced the time spent in the open arms of the EPM (19). Interestingly, 8 weeks of FRU supplementation in HFD (17% w/w) and water (10%) was sufficient to decrease the number of entries in OFT in male Wistar rats (73).

Notably, the maternal monosaccharide diet is sufficient to evoke anxiety-like behavior in offspring. Generalized anxiety disorders are characterized by uncontrollable anxiety, fear, and uncontrolled, exaggerated concern over endeavors. According to large population-based surveys, up to 33.7% of the population is affected by an anxiety disorder during their lifetime (74). Currently, few studies have provided evidence of a clinical or observational trial relationship between higher sugar intake and anxiety disorder events in humans (75, 76). Cohort studies have shown that sugar intake from sweet food/beverages increases the chance of incident mood disorders in men, and there is limited evidence regarding recurrent mood disorders in both sexes (31). A recent study suggested that a maternal FRU diet during gestation and lactation can affect offspring neurodevelopment. Long non-coding RNA interferes with transcript and gene expression crucial for the growth of neuronal cells in the fetal brain (46).

4.5. Monosaccharide diet-induced depression-like behavior

In the FST, we demonstrated that a maternal monosaccharide diet enhanced responsiveness to an acute stressor. In adolescent male rats, both GLU and FRU diets increased immobility and decreased climbing time. Only the FRU diet reduced the swimming time in male and climbing time in female rats. Moreover, in young adult offspring, a specific diet effect was observed in female rats. A GLU diet decreased immobility time, while an FRU diet decreased climbing time. The specificity of the increased depressive-like phenotype observed in adolescent males in the FST confirms the hyperactivity and anxiety-like behavior observed in the same individuals previously tested. To the best of our knowledge, there is a lack of data in the literature showing the effect of maternal HSD on the development of depressive-like behavior in offspring. Thus, our novel results establish behavioral stress-responsiveness after the maternal monosaccharide diet during offspring maturation. Several studies have demonstrated the association between postnatal HSD (mainly fructose) intake and a reduction in struggling and floating time with increasing immobile time. For instance, female Wistar rats treated with 23% FRU for 18 weeks showed increased immobility time (24). Likewise, a 55% high-FRU diet for 10 weeks evoked greater immobility time correlated with an increase in blood corticosterone measured after a 10-min FST in rats (at PND 116-120) of both sexes (77). Similarly, preadolescent (at PND94) male rats supplemented with a 55% high-FRU diet for 8-10 weeks showed an increased immobility time associated with greater corticosterone ejection and decreased struggle (19). A 30% FRU solution (w/v) consumption for 9 weeks increased immobility time in the tail suspension test in adult (at PND151) Swiss mice of both sexes (22). At the molecular level, high-FRU diet intake changed the hypothalamic transcriptome in preadolescent rats, notably corticotropin-releasing factor signaling and pro-opiomelanocortin processing (19).

Behavioral despair is a visible predictor of depression disorder development in humans. Depression is a common mental disorder accompanied by low mood and aversion to activity. Approximately 280 million people, including 5% of adults in the world, suffer from depression symptoms. Depression results from a complex interaction of social, psychological, and biological factors. Human research has revealed that high consumption of foods rich in added sugars and sweetened drinks increased the risk of depression in subsequent years (78), resulting in a high incidence and recurrence of mood disorders (79). More recent studies have confirmed that excessive consumption of sugar-sweetened soft drinks was associated with an increased risk of depression in adults and adolescents in Asia (80). In overweight individuals, increased consumption of SSBs was associated with an increased incidence of a diagnosis of depression (81). A recent cross-sectional study indicated that more than medium SSBs or fast-food consumption may lead to increased stress, depressive symptoms, and suicidal ideation in Korean adolescents (82).

5. Conclusion

In conclusion, our findings highlight the important role of maternal nutrition during pregnancy and lactation in proper offspring development. The results from behavioral screening using LAM, NOR, EZM, and FST tests showed that a maternal monosaccharide diet is a sufficient factor for changes in the emotional status of offspring. Herein, we demonstrated for the first time that maternal GLU- and FRU-enriched diets evoke diet-specific and sex-dependent behavioral alterations in unrelated offspring rats. Since the effects of monosaccharide consumption have not been adequately studied in humans, we suggest that the maternal metabolic dysregulation caused by overconsumption of GLU or FRU during pregnancy and lactation can both reprogram offspring metabolism and/or predispose them to behavioral disorders associated with sugar-dependent mechanisms. Based on the behavioral data, future research focusing on maternal monosaccharide diets may provide important clues to mental disorder development and prevention mechanisms.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding authors.

Ethics statement

The animal study was reviewed and approved by 2nd Local Institutional Animal Care and Use Committee (IACUC) in Kraków.

Author contributions

MF: conceived, designed, coordinated the study, and contributed to writing the manuscript. KWi: designed and performed the study, analyzed the data, and wrote the manuscript. KWy: performed the study and analyzed the data. AS: participated in rat generation and behavioral assays. All authors read, revised, and accepted the final version of this manuscript.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

1. Kalin NH. The critical relationship between anxiety and depression. Am J Psychiatry. (2020) 177:365–7. doi: 10.1176/appi.ajp.2020.20030305

2. Moitra M, Santomauro D, Collins PY, Vos T, Whiteford H, Saxena S, et al. The global gap in treatment coverage for major depressive disorder in 84 countries from 2000–2019: a systematic review and Bayesian meta-regression analysis. *PLoS Med.* (2022) 19:e1003901. doi: 10.1371/journal.pmed.1003901

3. Substance Abuse and Mental Health Services Administration. (2021). Key Substance Use and Mental Health Indicators in the United States: Results from the 2020 National Survey on Drug Use and Health (HHS Publication No. PEP21-07-01-003, NSDUH Series H-56). Rockville, MD: Center for Behavioral Health Statistics and Quality, Substance Abuse and Mental Health Services Administration. Available online at: https://www.samhsa.gov/data/

 Blount AJ, Adams CR, Anderson-Berry AL, Hanson C, Schneider K, Pendyala G. Biopsychosocial factors during the perinatal period: risks, preventative factors, and implications for healthcare professionals. *Int J Environ Res Public Health*. (2021) 18:8206. doi: 10.3390/ijerph18158206

5. Mandy M, Nyirenda M. Developmental origins of health and disease: the relevance to developing nations. *Int Health.* (2018) 10:66-70. doi: 10.1093/inthealth/ihy006

6. Besson AA, Lagisz M, Senior AM, Hector KL, Nakagawa S. Effect of maternal diet on offspring coping styles in rodents: a systematic review and meta-analysis. *Biol Rev.* (2016) 91:1065–80. doi: 10.1111/brv.12210

7. Gawlińska K, Gawliński D, Filip M, Przegaliński E. Relationship of maternal high-fat diet during pregnancy and lactation to offspring health. *Nutr Rev.* (2021) 79:709–25. doi: 10.1093/nutrit/nuaa020

8. Mergenthaler P, Lindauer U, Dienel GA, Meisel A. Sugar for the brain: the role of glucose in physiological and pathological brain function. *Trends Neurosci.* (2013) 36:587–97. doi: 10.1016/j.tins.2013.07.001

9. Rosinger A, Herrick K, Gahche J, Park S. Sugar-sweetened beverage consumption among U. S. youth, 2011-2014. NCHS Data Brief. (2017) (271):1–8.

10. Lundeen EA, Park S, Pan L, Blanck HM. Daily intake of sugar-sweetened beverages among US adults in 9 States, by state and sociodemographic and behavioral characteristics, 2016. *Prev Chronic Dis.* (2018) 15:E154. doi: 10.5888/pcd15.180335

11. Lee SH, Zhao L, Park S, Moore LV, Hamner HC, Galuska DA, et al. High added sugars intake among US Adults: characteristics, eating occasions, and top sources, 2015-2018. *Nutrients*. (2023) 15:265. doi: 10.3390/nu15020265

12. Vos MB, Kimmons JE, Gillespie C, Welsh J, Blanck HM. Dietary fructose consumption among US children and adults: the Third National Health and Nutrition Examination Survey. *Medscape J Med.* (2008) 10:160.

13. Casas R, Barquero SC, Estruch R. Impact of sugary food consumption on pregnancy: a review. *Nutrients*. (2020) 12:1–22. doi: 10.3390/nu12113574

14. Witek K, Wydra K, Filip M. A high-sugar diet consumption, metabolism and health impacts with a focus on the development of substance use disorder: a narrative review. *Nutrients*. (2022) 14:2940. doi: 10.3390/nu14142940

15. Mizera J, Kazek G, Niedzielska-Andres E, Pomierny-Chamiolo L. Maternal highsugar diet results in NMDA receptors abnormalities and cognitive impairment in rat offspring. *FASEB J.* (2021) 35:e21547. doi: 10.1096/fj.202002691R

16. Erbas O, Erdogan MA, Khalilnezhad A, Gürkan FT, Yigittürk G, Meral A, et al. Neurobehavioral effects of long-term maternal fructose intake in rat offspring. *Int J Dev Neurosci.* (2018) 69:68–79. doi: 10.1016/j.ijdevneu.2018.07.001

17. Choi CS, Kim P, Park JH, Gonzales ELT, Kim KC, Cho KS, et al. High sucrose consumption during pregnancy induced ADHD-like behavioral phenotypes in mice offspring. *J Nutr Biochem*. (2015) 26:1520–6. doi: 10.1016/j.jnutbio.2015.07.018

18. Wu KLH, Wu CW, Tain YL, Huang LT, Chao YM, Hung CY, et al. Environmental stimulation rescues maternal high fructose intake-impaired learning and memory in female offspring: its correlation with redistribution of histone

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deacetylase 4. Neurobiol Learn Mem. (2016) 130:105-17. doi: 10.1016/j.nlm.2016. 02.004

19. Harrell CS, Burgado J, Kelly SD, Johnson ZP, Neigh GN. High-fructose diet during periadolescent development increases depressive-like behavior and remodels the hypothalamic transcriptome in male rats. *Psychoneuroendocrinology.* (2015) 62:252–64. doi: 10.1016/j.psyneuen.2015.08.025

20. Beecher K, Alvarez Cooper I, Wang J, Walters SB, Chehrehasa F, Bartlett SE, et al. Long-term overconsumption of sugar starting at adolescence produces persistent hyperactivity and neurocognitive deficits in adulthood. *Front Neurosci.* (2021) 15:670430. doi: 10.3389/fnins.2021.670430

21. Wong A, Dogra VR, Reichelt AC. High-sucrose diets in male rats disrupt aspects of decision making tasks, motivation and spatial memory, but not impulsivity measured by operant delay-discounting. *Behav Brain Res.* (2017) 327:144–54. doi: 10.1016/j.bbr.2017.03.029

22. de Souza L, Barros W de M, de Souza RM, Delanogare E, Machado AE, Braga SP, et al. Impact of different fructose concentrations on metabolic and behavioral parameters of male and female mice. *Physiol Behav.* (2021) 228:113187. doi: 10.1016/j.physbeh.2020.113187

23. Xu TJ, Reichelt AC. Sucrose or sucrose and caffeine differentially impact memory and anxiety-like behaviours, and alter hippocampal parvalbumin and doublecortin. *Neuropharmacology*. (2018) 137:24-32. doi: 10.1016/j.neuropharm.2018.04.012

24. Coulibaly SM, Mesfioui A, Berkiks I, Ennaciri A, Chahirou Y, Diagana Y, et al. Effects of the methyl donors supplementation on hippocampal oxidative stress, depression and anxiety in chronically high fructose-treated rats. *Neuroscience.* (2021) 476:1–11. doi: 10.1016/j.neuroscience.2021.09.008

25. Martin-Gronert MS, Ozanne SE. Maternal nutrition during pregnancy and health of the offspring. *Biochem Soc Trans.* (2006) 34(Pt 5):779-82. doi: 10.1042/BST0340779

26. Edlow AG. Maternal obesity and neurodevelopmental and psychiatric disorders in offspring. *Prenat Diagn*. (2017) 37:95–110. doi: 10.1002/pd.4932

27. Bordeleau M, Fernández de Cossío L, Chakravarty MM, Tremblay MÈ. From maternal diet to neurodevelopmental disorders: a story of neuroinflammation. *Front Cell Neurosci.* (2021) 14:612705. doi: 10.3389/fncel.2020.612705

28. Gueye AB, Vendruscolo LF, de Ávila C, le Moine C, Darnaudéry M, Cador M. Unlimited sucrose consumption during adolescence generates a depressive-like phenotype in adulthood. *Neuropsychopharmacology.* (2018) 43:2627–35. doi: 10.1038/s41386-018-0025-9

29. Francis HM, Stevenson RJ. Higher reported saturated fat and refined sugar intake is associated with reduced hippocampal-dependent memory and sensitivity to interoceptive signals. *Behav Neurosci.* (2011) 125:943–55. doi: 10.1037/a00 25998

30. Liu J, Chen T, Chen M, Ma Y, Ma T, Gao D, et al. Sugarsweetened beverages and depressive and social anxiety symptoms among children and adolescents aged 7-17 years, stratified by body composition. *Front Nutr.* (2022) 9:888671. doi: 10.3389/fnut.2022.8 88671

31. Knüppel A, Shipley MJ, Llewellyn CH, Brunner EJ. Sugar intake from sweet food and beverages, common mental disorder and depression: prospective findings from the Whitehall II study. *Sci Rep.* (2017) 7:6287. doi: 10.1038/s41598-017-05649-7

32. DiNicolantonio JJ, O'Keefe JH, Wilson WL. Sugar addiction: is it real? A narrative review. Br J Sports Med. (2018) 52:910–3. doi: 10.1136/bjsports-2017-097971

33. Jacques A, Chaaya N, Beecher K, Ali SA, Belmer A, Bartlett S. The impact of sugar consumption on stress driven, emotional and addictive behaviors. *Neurosci Biobehav Rev.* (2019) 103:178–99. doi: 10.1016/j.neubiorev.2019.05.021

34. Jiménez JA, Zylka MJ. Controlling litter effects to enhance rigor and reproducibility with rodent models of neurodevelopmental disorders. J Neurodev Disord. (2021) 13:2. doi: 10.1186/s11689-020-09353-y

35. Wydra K, Suder A, Frankowska M, Borroto Escuela DO, Fuxe K, Filip M. Effects of intra-accumbal or intra-prefrontal cortex microinjections of adenosine 2A receptor ligands on responses to cocaine reward and seeking in rats. *Psychopharmacology.* (2018) 235:3509. doi: 10.1007/s00213-018-5072-8

36. Gawlińska K, Gawliński D, Korostyński M, Borczyk M, Frankowska M, Piechota M, et al. Maternal dietary patterns are associated with susceptibility to a depressive-like phenotype in rat offspring. *Dev Cogn Neurosci.* (2021) 47:100879. doi: 10.1016/j.dcn.2020.100879

37. Hammond RS, Tull LE, Stackman RW. On the delay-dependent involvement of the hippocampus in object recognition memory. *Neurobiol Learn Mem.* (2004) 82:26–34. doi: 10.1016/j.nlm.2004.03.005

38. Wang D, Noda Y, Zhou Y, Mouri A, Mizoguchi H, Nitta A, et al. The allosteric potentiation of nicotinic acetylcholine receptors by galantamine ameliorates the cognitive dysfunction in beta amyloid25–35 I.c.v.-Injected Mice: involvement of dopaminergic systems. *Neuropsychopharmacology*. (2007) 32:1261–71. doi: 10.1038/sj.npp.1301256

39. Shepherd JK, Grewal SS, Fletcher A, Bill DJ, Dourish CT. Behavioural and pharmacological characterisation of the elevated "zero-maze" as an animal model of anxiety. *Psychopharmacology*. (1994) 116:56–64. doi: 10.1007/BF02244871

40. Nishimura H, Tsuda A, Ida Y, Tanaka M. The modified forced-swim test in rats: influence of rope- or straw-suspension on climbing behavior. *Physiol Behav.* (1988) 43:665–8. doi: 10.1016/0031-9384(88)90223-5

41. Detke MJ, Rickels M, Lucki I. Active behaviors in the rat forced swimming test differentially produced by serotonergic and noradrenergic antidepressants. *Psychopharmacology.* (1995) 121:66–72. doi: 10.1007/BF02245592

42. Ladyman SR, Brooks VL. Central actions of insulin during pregnancy and lactation. J Neuroendocrinol. (2021) 33:e12946. doi: 10.1111/jne.12946

43. Maric I, Krieger JP, van der Velden P, Börchers S, Asker M, Vujicic M, et al. Sex and species differences in the development of diet-induced obesity and metabolic disturbances in rodents. *Front Nutr.* (2022) 9:141. doi: 10.3389/fnut.2022.828522

44. Lineker C, Kerr PM, Nguyen P, Bloor I, Astbury S, Patel N, et al. High fructose consumption in pregnancy alters the perinatal environment without increasing metabolic disease in the offspring. *Reprod Fertil Dev.* (2016) 28:2007–15. doi: 10.1071/RD15119

45. Clayton ZE, Vickers MH, Bernal A, Yap C, Sloboda DM. Early life exposure to fructose alters maternal, fetal and neonatal hepatic gene expression and leads to sex-dependent changes in lipid metabolism in rat offspring. *PLoS ONE.* (2015) 10:e0141962. doi: 10.1371/journal.pone.0141962

46. Zou Y, Guo Q, Chang Y, Zhong Y, Cheng L, Wei W. Effects of maternal high-fructose diet on long non-coding RNAs and anxiety-like behaviors in offspring. *Int J Mol Sci.* (2023) 24:4460. doi: 10.3390/ijms24054460

47. Wang P, Wu T, Fu Q, Liao Q, Li Y, Huang T, et al. Maternal high-fructose intake activates myogenic program in fetal brown fat and predisposes offspring to diet-induced metabolic dysfunctions in adulthood. *Front Nutr.* (2022) 9:848983. doi: 10.3389/fnut.2022.848983

48. Campbell GJ, Lucic Fisher SG, Brandon AE, Senior AM, Bell-Anderson KS. Sex-specific effects of maternal dietary carbohydrate quality on fetal development and offspring metabolic phenotype in mice. *Front Nutr.* (2022) 9:1593. doi: 10.3389/fnut.2022.917880

49. Office FP. Erratum: sex-specific effects of maternal dietary carbohydrate quality on fetal development and offspring metabolic phenotype in mice (Front. Nutr., (2022), 9, (917880), 10.3389/fnut.2022.917880). *Front Nutr.* (2022) 9:2905. doi: 10.3389/fnut.2022.1094120

50. Merino B, Fernández-Díaz CM, Cózar-Castellano I, Perdomo G. Intestinal fructose and glucose metabolism in health and disease. *Nutrients.* (2019) 12:94. doi: 10.3390/nu12010094

51. Hannou SA, Haslam DE, McKeown NM, Herman MA. Fructose metabolism and metabolic disease. *J Clin Invest*, (2018) 128:545–55. doi: 10.1172/JCI96702

52. Franco-Pérez J, Manjarrez-Marmolejo J, Ballesteros-Zebadúa P, Neri-Santos A, Montes S, Suarez-Rivera N, et al. Chronic consumption of fructose induces behavioral alterations by increasing orexin and dopamine levels in the rat brain. *Nutrients*. (2018) 10:1722. doi: 10.3390/nu10111722

53. Farsad-Naeimi A, Asjodi F, Omidian M, Askari M, Nouri M, Pizarro AB, et al. Sugar consumption, sugar sweetened beverages and attention deficit hyperactivity disorder: a systematic review and meta-analysis. *Complement Ther Med.* (2020) 53:102512. doi: 10.1016/j.ctim.2020.102512

54. Lien L, Lien N, Heyerdahl S, Thoresen M, Bjertness E. Consumption of soft drinks and hyperactivity, mental distress, and conduct problems among adolescents in Oslo, Norway. *Am J Public Health.* (2006) 96:1815–20. doi: 10.2105/AJPH.2004.059477

55. Del-Ponte B, Anselmi L, Assunção MCF, Tovo-Rodrigues L, Munhoz TN, Matijasevich A, et al. Sugar consumption and attention-deficit/hyperactivity disorder (ADHD): a birth cohort study. *J Affect Disord.* (2019) 243:290–6. doi: 10.1016/j.jad.2018.09.051

56. Mucellini AB, Laureano DP, Silveira PP, Sanvitto GL. Maternal and postnatal obesity alters long-term memory and hippocampal molecular signaling of male rat. *Brain Res.* (2019) 1708:138–45. doi: 10.1016/j.brainres.2018. 12.021 57. Chepulis LM, Starkey NJ, Waas JR, Molan PC. The effects of long-term honey, sucrose or sugar-free diets on memory and anxiety in rats. *Physiol Behav.* (2009) 97:359–68. doi: 10.1016/j.physbeh.2009.03.001

58. Dharavath RN, Arora S, Bishnoi M, Kondepudi KK, Chopra K. High fat-low protein diet induces metabolic alterations and cognitive dysfunction in female rats. *Metab Brain Dis.* (2019) 34:1531-46. doi: 10.1007/s11011-019-00459-4

59. Buyukata C, Vukalo M, Xu TJ, Khore MA, Reichelt AC. Impact of high sucrose diets on the discrimination of spatial and object memories with overlapping features. *Physiol Behav.* (2018) 192:127–33. doi: 10.1016/j.physbeh.2018. 02.027

60. Kendig MD, Boakes RA, Rooney KB, Corbit LH. Chronic restricted access to 10% sucrose solution in adolescent and young adult rats impairs spatial memory and alters sensitivity to outcome devaluation. *Physiol Behav.* (2013) 120:164–72. doi: 10.1016/j.physbeh.2013.08.012

61. Jurdak N, Kanarek RB. Sucrose-induced obesity impairs novel object recognition learning in young rats. *Physiol Behav.* (2009) 96:1–5. doi: 10.1016/j.physbeh.2008.07.023

62. Lemos C, Rial D, Gonçalves FQ, Pires J, Silva HB, Matheus FC, et al. High sucrose consumption induces memory impairment in rats associated with electrophysiological modifications but not with metabolic changes in the hippocampus. *Neuroscience.* (2016) 315:196–205. doi: 10.1016/j.neuroscience.2015. 12.018

63. Cohen JFW, Rifas-Shiman SL, Young J, Oken E. Associations of prenatal and child sugar intake with child cognition. *Am J Prev Med.* (2018) 54:727-35. doi: 10.1016/j.amepre.2018.02.020

64. Shearrer GE, Stice E, Burger KS. Adolescents at high risk of obesity show greater striatal response to increased sugar content in milkshakes. *Am J Clin Nutr.* (2018) 107:859–66. doi: 10.1093/ajcn/nqy050

65. Gui Z, Huang S, Chen Y, Zhao Y, Jiang N, Zhang S, et al. Association between sugar-sweetened beverage consumption and executive function in children. *Nutrients*. (2021) 13:4563. doi: 10.3390/nu13124563

66. Chong CP, Shahar S, Haron H, Din NC. Habitual sugar intake and cognitive impairment among multi-ethnic Malaysian older adults. *Clin Interv Aging.* (2019) 14:1331–42. doi: 10.2147/CIA.S211534

67. Sangüesa G, Cascales M, Griñán C, Sánchez RM, Roglans N, Pallàs M, et al. Impairment of novel object recognition memory and brain insulin signaling in fructose- but not glucose-drinking female rats. *Mol Neurobiol.* (2018) 55:6984– 99. doi: 10.1007/s12035-017-0863-1

68. Spagnuolo MS, Mazzoli A, Nazzaro M, Troise AD, Gatto C, Tonini C, et al. Long-lasting impact of sugar intake on neurotrophins and neurotransmitters from adolescence to young adulthood in rat frontal cortex. *Mol Neurobiol*. (2023) 60:1004– 20. doi: 10.21203/rs.3.rs-1622302/v1

69. Hsu TM, Konanur VR, Taing L, Usui R, Kayser BD, Goran MI, et al. Effects of sucrose and high fructose corn syrup consumption on spatial memory function and hippocampal neuroinflammation in adolescent rats. *Hippocampus*. (2015) 25:227–39. doi: 10.1002/hipo.22368

70. Zou Y, Guo Q, Chang Y, Jia L, Zhai L, Bai Y, et al. Learning and memory impairment and transcriptomic profile in hippocampus of offspring after maternal fructose exposure during gestation and lactation. *Food Chem Toxicol.* (2022) 169:113394. doi: 10.1016/j.fct.2022.113394

71. Bukhari SHF, Clark OE, Williamson LL. Maternal high fructose diet and neonatal immune challenge alter offspring anxiety-like behavior and inflammation across the lifespan. *Life Sci.* (2018) 197:114–21. doi: 10.1016/j.lfs.2018. 02.010

72. Gray C, Gardiner SM, Elmes M, Gardner DS. Excess maternal salt or fructose intake programmes sex-specific, stress- and fructose-sensitive hypertension in the offspring. *Br J Nutr.* (2016) 115:594–604. doi: 10.1017/S00071145150 04936

73. Gancheva S, Galunska B, Zhelyazkova-Savova M. Diets rich in saturated fat and fructose induce anxiety and depression-like behaviours in the rat: is there a role for lipid peroxidation? *Int J Exp Pathol.* (2017) 98:296–306. doi: 10.1111/iep. 12254

74. Bandelow B, Michaelis S. Epidemiology of anxiety disorders in the 21st century. *Dialogues Clin Neurosci.* (2015) 17:327–35. doi: 10.31887/DCNS.2015.17.3/bbandelow

75. Aucoin M, Lachance L, Naidoo U, Remy D, Shekdar T, Sayar N, et al. Diet and anxiety: a scoping review. *Nutrients.* (2021) 13:4418. doi: 10.3390/nu131 24418

76. Monteiro S, Nejad YS, Aucoin M. Perinatal diet and offspring anxiety: a scoping review. *Transl Neurosci.* (2022) 13:275–90. doi: 10.1515/tnsci-2022-0242

77. Kloster A, Hyer MM, Dyer S, Salome-Sanchez C, Neigh GN. High fructose diet induces sex-specific modifications in synaptic respiration and affective-like behaviors in rats. *Neuroscience*. (2021) 454:40–50. doi: 10.1016/j.neuroscience.2019.11.039

78. Reis DJ, Ilardi SS, Namekata MS, Wing EK, Fowler CH. The depressogenic potential of added dietary sugars. *Med Hypotheses*. (2020) 134:109421. doi: 10.1016/j.mehy.2019.109421

79. Xu Y, Zeng L, Zou K, Shan S, Wang X, Xiong J, et al. Role of dietary factors in the prevention and treatment for depression: an umbrella review of meta-analyses of prospective studies. *Transl Psychiatry*. (2021) 11:1-13. doi: 10.1038/s41398-021-01590-6

80. Huang Y, Wang Y, Wang H, Liu Z, Yu X, Yan J, et al. Prevalence of mental disorders in China: a cross-sectional epidemiological study. *Lancet Psychiatry.* (2019) 6:211–24. doi: 10.1016/S2215-0366(18)30511-X

81. Ángeles Pérez-Ara M, Gili M, Visser M, Penninx BWJH, Brouwer IA, Watkins E, et al. Associations of non-alcoholic beverages with major depressive disorder history and depressive symptoms clusters in a sample of overweight adults. *Nutrients*. (2020) 12:1–15. doi: 10.3390/nu12103202

82. Ra JS. Consumption of sugar-sweetened beverages and fast foods deteriorates adolescents' mental health. *Front Nutr.* (2022) 9:1058190. doi: 10.3389/fnut.2022.1058190