Epigenetic therapy in lung cancer – role of microRNAs

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INTRODUCTION

Lung cancer is the most common invasive cancer and cause of cancer death worldwide (Ferlay et al., 2010). Among lung cancers, 80% are classified as non-small cell lung cancer (NSCLC) and 20% are small cell lung cancer (SCLC). While recent developments in computed tomography (CT) screening for NSCLC may lead to detection of tumors at earlier stages (Aberle et al., 2011a), currently over 70% of lung cancers are loco-regionally advanced or metastatic at the time of diagnosis. Despite advances in chemotherapy, radiotherapy, and surgery the death rate from lung cancer has remained largely unchanged. In recent years therapeutic decisions in NSCLC have been more and more based on histological and molecular characteristics. The era of molecular targeted therapy in lung cancer had its origin in 2004, when activating mutations in the epidermal growth factor receptor (EGFR) and their correlation with clinical response to EGFR tyrosine kinase inhibitors (TKIs) were discovered (Lynch et al., 2004; Paez et al., 2004; Pao et al., 2004). Other oncogenic drivers in lung adenocarcinoma include mutations of BRAF, NRAS, KRAS, MET, PIK3CA, HER2, RET, JAK2, ROS1, and ALK (Bronte et al., 2010; Garber, 2010; Janku et al., 2010; Pao and Girard, 2011). Although most of the patients with advanced NSCLC do not harbor one of the mentioned molecular alterations. Therefore, it is important to identify new markers for molecularly targeted therapy.

The most studied epigenetic phenomena include posttranslational modifications in DNA and histone proteins. In the recent years the changes in expression levels of small, non-coding, singlestranded RNAs – so called microRNAs (miRNAs) – have been detected and described. miRNAs are usually 18–25 nucleotides long. Depending on the degree of homology to their 3'UTR target sequence, miRNAs induce translational repression or degradation of mRNAs (**Figure 1**). It is estimated that more than 1,000 miR-NAs are transcribed and that 30% of the human genome is under miRNA regulation, one miRNA being able to modulate posttranscriptionally hundreds of downstream genes. In this regard, miRNAs control a wide range of biological processes including

Lung cancer is the leading cause of cancer deaths worldwide. microRNAs (miRNAs) are a class of small non-coding RNA species that have been implicated in the control of many fundamental cellular and physiological processes such as cellular differentiation, proliferation, apoptosis, and stem cell maintenance. Some miRNAs have been categorized as "oncomiRs" as opposed to "tumor suppressor miRs." This review focuses on the role of miRNAs in the lung cancer carcinogenesis and their potential as diagnostic, prognostic, or predictive markers.

Keywords: microRNA, lung cancer, biomarker, diagnosis, prognosis, therapy

cellular differentiation, proliferation, apoptosis, and stem cell maintenance. More than half of the miRNAs genes are located in cancer-associated genomic regions or in fragile sites. Several miRNAs located in deleted regions have low expression levels in cancer tissues (Calin et al., 2004). miRNAs can act as tumor suppressors when their function loss can initiate or contribute to the malignant transformation of a normal cell. The loss of function of a miRNA could be due to several mechanisms, including genomic deletion, mutation, epigenetic silencing, and/or miRNA processing alterations (Calin et al., 2002, 2005; Saito et al., 2006; Nakamura et al., 2007). On the other hand miRNAs can act as oncogenes.

In this review the focus will be on the state-of-the-art knowledge currently available on the clinical significance of miRNAs in lung carcinogenesis and their role as diagnostic, prognostic, and predictive markers in lung cancer.

MicroRNAs AND LUNG CARCINOGENESIS

Several studies have identified an important role of miRNAs in the regulation of the cell cycle and carcinogenesis in different organs, including the lung (Carleton et al., 2007; Chivukula and Mendell, 2008). The first evidence of miRNA deregulation in lung cancer came from the study by Volinia et al. (2006), who identified a group of miRNAs frequently aberrantly expressed in tumor tissues with respect to the normal tissue counterpart. Takahashi et al. (2009) found decreased expression of miR-107, miR-185, and let-7a, but increased expression of miR-31 in lung carcinoma tissues and cancer cell lines. Overexpression of miR-107 and miR-185 significantly reduced A549 and H1299 cell proliferation. Both miRNAs significantly increased the percentage of cancer cells in the G1 cell cycle phase. Inhibition of miR-107 increased the proliferation of A549 lung cancer cells (Cheng et al., 2005). Overexpression of let-7a, miR-107, and miR-126 suppresses lung cancer cell growth (Zhong et al., 2010).

Let-7 is an important miRNA and is highly expressed in lung tissue, as supported by *in vitro* and *in vivo* studies (Johnson et al., 2005; Yanaihara et al., 2006). The tumor-suppressive let-7 family



II (Pol II) into long primary mIRNA transcripts, which are cleaved in the nucleus by the RNase III enzyme Drosha, resulting in a hairpin precursor form called pre-miRNA. Pre-miRNA is exported from the nucleus to the cytoplasm

transient miRNA duplex. Only one strand of the miRNA duplex (mature miRNA) is incorporated into a large protein complex called RISC (RNA-induced silencing complex).

targets well known oncogenes like Ras (Johnson et al., 2005), c-Myc (Sampson et al., 2007), and HMGA2 (Lee and Dutta, 2007) which are involved in lung cancer carcinogenesis. The let-7 family has a tumor-suppressive role in NSCLC in animal models (Kumar et al., 2008; Trang et al., 2010). It was demonstrated that 40% of lung tumors and 60% of lung cancer cell lines have low expression levels of let-7 (Takamizawa et al., 2004). In another study low let-7 expression levels were confirmed and could be correlated to high RAS expression in lung tumor models (Johnson et al., 2005, 2007). Together with further studies showing different miRNA expression profiles (Fabbri et al., 2007; Navarro et al., 2009; Raponi et al., 2009; Seike et al., 2009) seven miRNAs have frequently been found to be downregulated in NSCLC: let-7, miR-29 family, miR-30a, miR-124a, miR-126*, miR-126, and miR-145 whereas six miRNAs are higher expressed in NSCLC tissue based on these studies: miR-17, miR-21, miR-106a, miR-182, miR-203, and miR-210. Own data confirmed a significant downregulation of miR-29b in lung adenocarcinoma compared to surrounding non-cancerous tissue (Rothschild et al., 2012a).

Members of the miRNA-29 family (miR-29a, miR-29b, and miR-29c) are known to be highly expressed in normal tissues and downregulated in different types of cancer, including neuroblastoma, sarcoma, glioma, high-risk chronic lymphatic leukemia (CLL), invasive breast cancer, cholangiocarcinoma, and lung cancer (Calin et al., 2005; Iorio et al., 2005; Yanaihara et al., 2006; Mott et al., 2007; Wu et al., 2009; Xu et al., 2009). The three miR-29 isoforms are arranged in two clusters: miR-29b1/miR-29a located on chromosome 7q32 and miR-29b2/miR-29c located on chromosome 1q32. miR-29a has been shown to reduce invasiveness and proliferation of human carcinoma cell lines (Muniyappa et al., 2009). In lung cancer the miR-29 family members also target DNA methyltransferases (DNMT3A and DNMT3B), and can thereby restore patterns of DNA methylation and expression of silenced tumor suppressor genes (Fabbri et al., 2007). In our own study we demonstrated direct binding or miR-29b to inhibitor of differentiation 1 (ID1) (Rothschild et al., 2012a). Anti-miR-29b enhanced ID1 mRNA and protein levels, and significantly increased lung cancer cell migration and invasion, a hallmark of the Src-ID1

pathway. miR-29b suppressed the level of ID1 and significantly reduced migration and invasion.

Hypoxia-inducible factor- 1α (HIF- 1α) is considered a key regulator of tumor angiogenesis (Semenza, 2010). The polycistronic cluster miR-17-92 consisting of six miRNAs (miR-17, miR-18a, miR-19a, miR-19b-1, miR-20a, miR-92a-1) is involved in embryonal lung development (Lu et al., 2007). Mice deficient for miR-17-92 die shortly after birth with lung hypoplasia and a ventricular septal defect (Ventura et al., 2008). This miRNA cluster is overexpressed in several lung cancer cell lines, preferentially in SCLC and the overexpression is associated with increased cell proliferation (Hayashita et al., 2005). The inhibition of miR-17-92 by antisense oligonucleotides can induce apoptosis in overexpressing cell lines (Matsubara et al., 2007). miR-17-92 targets HIF-1 α which might be the key mechanism of action in lung cancer cell proliferation (Taguchi et al., 2008). Moreover, miR-519c binds to the HIF-1 α 3'UTR and thereby reduces tumor angiogenesis (Cha et al., 2010).

Retinoblastoma 1 (Rb1) was the first identified tumor suppressor. Overexpression of miR-192 inhibited cell proliferation in A549, H460, and 95D lung cancer cells by decreasing Rb1 mRNA and protein expression (Feng et al., 2011). The same effect was demonstrated in a nude mouse model.

Epidermal growth factor receptor signaling has been shown to play an important role in cell migration and invasion (Prenzel et al., 2001). miR-125a-5p is regulating several downstream genes involved in EGFR signaling (Wang et al., 2009). Inhibition of miR-125a-5p significantly enhanced cell migration and invasion, suggesting that miR-125a-5p is negatively correlated with lung cancer invasion and metastasis.

Further miRNAs that have been described to have tumorsuppressive function in lung cancer are miR-93 (Du et al., 2009), miR-98 (Du et al., 2009), miR-101 (Zhang et al., 2011), miR-182 (Sun et al., 2010), miR-197 (Du et al., 2009), miR-212 (Incoronato et al., 2010), miR-451 (Wang et al., 2011).

Recently, it has been shown that miRNAs are also present in body fluids, shuttled by so called exosomes, which are secreted both by normal and tumor cells. Two miRNAs (miR-23 and miR-225) are expressed at higher levels in the circulating exosomes of NSCLC patients compared to healthy donors (Chen et al., 2008). There seems to be a correlation between the expression of miR-NAs in tumor tissues and in the blood suggesting that the tumor releases circulating miRNAs inside exosome particles (Rabinowits et al., 2009). A recent publication identified specific miRNAs (e.g., miR-21 and miR-29a) in exosome of lung cancer patients able to bind and activate Toll-like receptors (TLR) in cancer surrounding immune cells (Fabbri et al., 2012). By activating TLRs, these miRNAs trigger the NF-kappaB pathway which ultimately leads to an increased production of interleukin-6 (IL-6) and tumor necrosis factor-alpha (TNF-alpha) by the immune cells, leading to increased tumor growth and metastatic potential. This study describes a novel mechanism of action of miRNAs and support the role of cancer-released exosomes in lung cancer carcinogenesis.

In general, the current literature suggests that miRNAs are involved in lung cancer carcinogenesis by different mechanisms. They might act directly by targeting oncogenes or tumor suppressor genes and regulate their expression or by influencing their epigenetic regulation within the tumor. As a novel mechanism, miRNAs also affect lung carcinogenesis by triggering a TLR-mediated oncogenic inflammatory response.

MicroRNA AS DIAGNOSTIC MARKERS OF LUNG CANCER

Since there is no validated population-based screening procedure available, most patients with lung cancer are diagnosed at advanced stages. Recently a large trial showed that low-dose helical CT screening in an older, high-risk population reduced lung cancer mortality by 20% (Aberle et al., 2011a,b). However there is a high number of false positive findings in CT scans (Aberle et al., 2011a; Barba et al., 2011). Therefore, development of a reliable and non-invasive confirmatory test would reduce overdiagnosis. Interestingly, miRNA expression levels show apparent differences following exposure to cigarette smoking (Izzotti et al., 2009). Therefore changes in miRNA expression might be a reliable tool for cancer early detection in smokers. Xie et al. (2010) found that miR-21 expression in the sputum specimens was significantly higher in cancer patients compared with healthy controls. Further studies have shown that increased miR-21 expression resulted in 70% sensitivity and 100% specificity in the diagnosis of lung cancer by sputum cytology (Wang et al., 2012). Therefore, investigating miRNA expression levels in the sputum might be a potential non-invasive approach for early lung cancer detection.

Yanaihara et al. (2006) studied miRNA expression profiles in lung cancer tissues and normal adjacent tissue. They identified a unique miRNA profile allowing discriminating lung cancers from non-cancerous lung tissues. Furthermore they describe a signature that distinguishes histological subtypes of NSCLC. Some of the miRNAs (e.g., miR-21, miR-155, and members of the miR-17-92 cluster) were common to the two studies, and have been found to be upregulated in other types of cancer. Shen et al. (2011) identified 12 miRNAs aberrantly expressed in early-stage NSCLC. Four of them (miR-21, miR-126, miR-210, and miR-486-5p) yielded 86% sensitivity and 97% specificity in distinguishing NSCLC patients from healthy controls. Furthermore, these miRNAs had higher sensitivity (92%) in diagnosing lung adenocarcinoma compared with squamous cell carcinomas (82%) (p < 0.05). In a recent study miRNA expression in tumor and non-cancerous tissue pairs collected from stage I-III lung squamous cell carcinoma patients treated with neoadjuvant chemotherapy or radiotherapy were analyzed (Tan et al., 2011). Five miRNAs (miR-30a, miR-140-3p, miR-182, miR-210, and miR-486-5p) were identified to distinguish invasive lung cancer from non-cancerous lung tissue. The classifier had an accuracy of 94.1% in a training cohort (34 patients) and 96.2% in a test cohort (26 patients). Yu et al. (2010) found that the combined overexpression of miR-21, miR-200b, miR-375, and miR-486 in surgical tissues and sputum were biomarkers in the prediction of lung adenocarcinoma from normal controls with 81% sensitivity and 92% specificity. The results have been validated in 64 lung cancer patients and 58 cancer-free participants. The detection of miR-205, miR-210, and miR-708 in sputum was highly sensitive (73%) and specific (96%) in diagnosing lung cancer (Xing et al., 2010).

Compared with normal lung tissues, the top 10 deregulated miRNAs in lung tumors that were appropriate for discriminating CT-detected lung cancer from normal lung tissue were let-7, miR-21, miR-200b, miR-210, miR-219-1, miR-324, which were all upregulated, and miR-30a, miR-126, miR-451, and miR-486, which were all downregulated (Boeri et al., 2011).

The histopathological subclassification of NSCLC into the two large groups adenocarcinoma and squamous cell carcinoma has gained interest during the last years because adenocarcinoma histology in stage IV NSCLC is predictive for treatment with pemetrexed (Scagliotti et al., 2009). Two miRNAs (miR-21, miR-205) were reported to accurately distinguish adenocarcinoma from squamous cell carcinoma subtype (Lebanony et al., 2009).

Moreover, different miRNA expression profiles were also described between primary lung tumors and metastases. Overexpression of miR-182 was found in primary lung cancers, whereas miR-126 was highly expressed in metastatic tumors (Barshack et al., 2010).

In conclusion, these results suggest that miRNA signatures can help to differentiate lung cancer tissue from non-cancerous lung tissue. Furthermore, there are specific miRNAs associated with early lung cancer development which could be measured in sputum specimens and might support early lung cancer diagnosis. Other miRNAs can help to distinguish between histopathological subtypes of NSCLC or between lung primary tumors and metastases.

MicroRNAs AS PROGNOSTIC BIOMARKERS OF LUNG CANCER

At least two studies demonstrated the prognostic role of let-7 in patients with NSCLC. Takamizawa et al. (2004) investigated 159 patients with stage I-III NSCLC and showed that low expression of let-7 was associated with a shorter postoperative survival. This finding was confirmed by Yanaihara et al. (2006) showing that overexpression of the precursor of miR-155 and the downregulation of let-7a-2 correlated with poor survival in NSCLC patients. This result was confirmed in a further study (Chin et al., 2008). In a study by Yu et al. (2008) let-7a was part of a signature of five miRNAs (let-7a, miR-137, miR-182*, miR-221, miR-372) that were prognostic predictors. Reduced expression of miR-146b was found to be associated with a reduced overall survival in patients undergoing radical surgery for squamous cell carcinoma of the lung (Raponi et al., 2009). Another study in stage I-III squamous cell carcinoma showed a negative prognostic impact of high miR-31 expression (Tan et al., 2011). In male smokers with stage I-IIIA squamous cell of the lung, Landi et al. (2010) identified a signature of five downregulated miRNAs (let-7e, miR-25, miR-34a, miR-34c-5p, miR-191) predicting for poor outcome. miR-328 might be a marker for patients at higher risk for brain metastases as this miRNA has been found to be differentially expressed in patients with and without brain metastases of NSCLC (Arora et al., 2011). Lower expression of miR-34a in tumor tissues was associated with a high recurrent risk (Gallardo et al., 2009). High miR-21 and low miR-181 expression levels predicted poor survival independent of TNM staging (Gao et al., 2010). In another study overexpression of miR-429 was associated with shorter disease-free survival and downregulation of miR-486-5p in plasma predicted for poor outcome (Donnem et al., 2011).

The role of adjuvant chemotherapy in surgically resected stage I NSCLC is controversial (Besse and Le Chevalier, 2012). Reliable biomarkers predicting the risk of relapse are needed to identify patients most likely to benefit from adjuvant chemotherapy. Patnaik et al. (2010) demonstrated that miRNAs can predict lung cancer recurrence with an accuracy of 83%. Lu et al. (2012) reported on 527 stage I NSCLC patients investigated for miRNA expression levels. Two miRNA signatures were associated with relapse-free survival. The first contained 34 miRNAs derived from 357 patients whereas the second was based on 27 miRNAs from adenocarcinoma histologies. Both signatures were validated in an independent data set with 170 stage I NSCLC patients.

In the largest cohort of NSCLC patients investigating the prognostic and predictive value of miRNA signatures 639 patients with radically resected stage I-III NSCLC randomized to adjuvant cisplatin-based chemotherapy or follow-up [IALT trial (Arriagada et al., 2004)] were included (Voortman et al., 2010). miRNA expression levels in the tumor tissue were analyzed using quantitative real-time PCR (qRT-PCR). Only low expression of miR-21 was associated with shorter survival time. None of the investigated miRNAs nor any of the tested signatures was of predictive value for adjuvant chemotherapy. Hu et al. (2010) described a signature of circulating miRNAs (miR-1, miR-30d, miR-486, miR-499) associated with the outcome in 303 surgically resected stage I-IIIA NSCLC patients treated with adjuvant chemotherapy.

In our own study, we extracted total RNA from tumors and matched non-cancerous lung tissue from 23 cases with a pathological diagnosis of lung adenocarcinoma (Rothschild et al., 2012a). miRNA expression levels were determined using qRT-PCR. In 19 (83%) of the cases, miR-29b was downregulated in tumor compared with matched lung. Using the median tumor level as cut-off value, tumor miR-29b expression significantly correlated with event-free (p = 0.003) and overall survival (p = 0.039). In another study we extracted total RNA from 18 tissue samples of lung adenocarcinoma and also from non-cancerous tissue from the same patients (Rothschild et al., 2012b). In 15/18 (83%) of analyzed adenocarcinoma samples, miR-381 was downregulated in tumor compared with normal lung tissue. Overall miR-381 expression was significantly lower in adenocarcinomas than in matched normal lung tissue (Mann–Whitney U test, p = 0.003). Using the median tumor level as cut-off, tumor miR-381 expression significantly correlated with event-free survival (p = 0.003)and overall survival (p = 0.02).

In conclusion, several miRNAs show tissue specific expression levels and can be correlated with patient outcome.

MicroRNA AS PREDICTIVE MARKERS IN LUNG CANCER

As discussed in the section on miRNA as prognostic markers, the largest clinical trial evaluating the role of miRNA signatures in NSCLC did not show any of the examined miRNAs to be of predictive value (Voortman et al., 2010). Nevertheless, several preclinical studies as well as miRNA expression analyses from clinical trials showed that certain miRNAs might be of predictive value and do influence the sensitivity of lung cancer cells to chemotherapeutic or targeted agents as well as to irradiation.

In our own study we raised the hypothesis that miR-29b is a potential predictive marker for SRC TKIs due to the fact that inhibition of miR-29b diminished the effects of SRC tyrosine kinase inhibition (Rothschild et al., 2012a).

miR-17-92, miR-221, and miR-222 have been shown to sensitize lung cancer cells to cytotoxic agents (Hayashita et al., 2005; Matsubara et al., 2007; Garofalo et al., 2008). Galluzzi et al. (2010) demonstrated that miR-181a and miR-630 modulate the sensitivity of lung cancer cell lines toward cis-diamminedichloroplatinum (cis-DDP). pre-miR-630 diminished the sensitivity of A549 cells in response to DDP by inducing cancer cell arrest in G0-G1 and increasing the cell cycle inhibitor p27^{kip1}. Ectopic expression of miR-451 on the other hand sensitized A549 cells to DDP possibly by increasing DDP-induced apoptosis (Bian et al., 2011).

The description of oncogenic mutations (e.g., EGFR mutations) and the introduction of new therapeutics directly targeting these molecular changes (e.g., EGFR TKIs) have changed the therapeutic strategies in lung cancer treatment. Therefore, higher interest in understanding sensitivity and resistance mechanisms toward these new therapeutic approaches is a consequence of this development. Higher expression of miR-126 enhanced cytotoxicity induced by gefitinib in lung cancer cells (Zhong et al., 2010). EGFR is directly targeted by miR-128b and miR-128b lossof-heterozygosity (LOH), which is frequently found in NSCLC, is a direct regulator of EGFR, correlates with clinical response and survival following gefitinib therapy (Weiss et al., 2008). Also, miR-21 was shown to enhance the therapeutic effects of EGFR TKIs in lung cancer cell lines (Seike et al., 2009). miR-7 has been described to target EGFR, AKT, and ERK and therefore suppressing EGFR pathway signaling (Webster et al., 2009). Ectopic expression of miR-7 was able to overcome EGFR TKI resistance in lung cancer cell lines (Rai et al., 2011). Another study described that restoration of tumor suppressor miR-145 inhibits cancer cell growth in EGFR mutant lung adenocarcinoma (Cho et al., 2009).

TRAIL (Apo2L/tumor necrosis factor-related apoptosisinducing ligand) is a member of the TNF family that can induce apoptosis in several types of cancer, including lung cancer (Schaefer et al., 2007). miR-221 and miR-222 silence PTEN and TIMP3 tumor suppressors and therefore lead to resistance of lung cancer cells toward TRAIL (Garofalo et al., 2009).

The role of miRNA expression related to irradiation has been investigated by several groups. A549 cells, which contain lower let-7 levels and higher activated RAS had significant changes in miRNAs as early as 2 h after irradiation (Weidhaas et al., 2007). Inhibition of let-7b caused significant protection from radiation (Galluzzi et al., 2010). On the other hand, overexpression of let-7g protected A549 cells from radiation, whereas inhibition of let-7g increased radiosensitization in lung cancer cells (Galluzzi et al., 2010).

REFERENCES

- Aberle, D. R., Adams, A. M., Berg, C. D., Black, W. C., Clapp, J. D., Fagerstrom, R. M., et al. (2011a). Reduced lung-cancer mortality with low-dose computed tomographic screening. *N. Engl. J. Med.* 365, 395–409. doi:10.1056/NEJMoa1102873
- Aberle, D. R., Berg, C. D., Black, W. C., Church, T. R., Fagerstrom, R.

M., Galen, B. I., et al. (2011b). The National Lung Screening Trial: overview and study design. *Radiology* 258, 243–253. doi:10.1148/radiol.10091808

Arora, S., Ranade, A. R., Tran, N. L., Nasser, S., Sridhar, S., Korn, R. L., et al. (2011). MicroRNA-328 is associated with (non-small) cell lung cancer (NSCLC) brain In conclusion, despite a large negative study on the predictive role of miRNAs signatures in NSCLC further studies are warranted on the basis of several preclinical data. As shown in various studies miRNA can modulate the response of lung cancer cells to EGFR TKIs by interacting with the respective signaling pathway. Confirmation of their predictive role in larger clinical trials is needed.

CONCLUSION

Since their discovery in the 1990s miRNAs have increasingly been recognized as key player in carcinogenesis and cancer progression, but also as potential diagnostic markers and biomarkers. In this overview more insights in the involvement of miRNAs in lung carcinogenesis as well as in their role as diagnostic, prognostic, and predictive biomarkers have been summarized. The current evidence on the prognostic and predictive role of miRNAs is at least partially inconsistent. Therefore, prospective trials evaluating the role of miRNAs described to have prognostic or predictive impact in small retrospective analysis should be planned. For this, standardized protocols for the isolation and the analysis of expression levels of miRNAs are needed.

For the future, miRNAs from body fluids, especially the blood plasma might be the center of attention. As discussed, numerous studies have identified aberrant miRNA expression profiles in lung cancer. Therefore, miRNAs can potentially be used as biomarkers in the diagnosis and classification of lung cancer. Moreover, miRNA expression levels in blood plasma may be used as a non-invasive confirmatory screening test complementary to the low-dose helical CT screening in the near future.

Another interesting research field is the use of miRNAs as therapeutic agents. The miRNA replacement or the anti-miRNAs will possibly become a new strategy for lung cancer therapy. At the moment several challenges have to be resolved before miRNAs could be investigated in clinical trials. One of the main problems is to find a suitable non-toxic delivery system capable of selectively transporting miRNA-based therapeutics to the tumor site. As discussed in the introduction, one single miRNA is able to target more than 100 mRNAs. It is therefore highly questionable if miRNAs as therapeutic tools could be of sufficient specificity.

In conclusion, current knowledge on miRNAs in lung cancer has extended our understanding of lung cancer carcinogenesis and provides health care professionals with a new and innovative cancer biomarker. Beside preclinical research, there are several clinical trials evaluating the role of miRNAs as prognostic and/or predictive biomarkers ongoing.

metastasis and mediates NSCLC migration. Int. J. Cancer 129, 2621–2631. doi:10.1002/ijc.2 5939

Arriagada, R., Bergman, B., Dunant, A., Le Chevalier, T., Pignon, J. P., and Vansteenkiste, J. (2004). Cisplatinbased adjuvant chemotherapy in patients with completely resected non-small-cell lung cancer. N. *Engl. J. Med.* 350, 351–360. doi:10.1056/NEJMoa031644

Barba, M., Felsani, A., Rinaldi, M., Giunta, S., Malorni, W., and Paggi, M. G. (2011). Reducing the risk of overdiagnosis in lung cancer: a support from molecular biology. *J. Cell. Physiol.* 226, 2213–2214. doi:10.1002/jcp.2 2558

- Barshack, I., Lithwick-Yanai, G., Afek, A., Rosenblatt, K., Tabibian-Keissar, H., Zepeniuk, M., et al. (2010). MicroRNA expression differentiates between primary lung tumors and metastases to the lung. *Pathol. Res. Pract.* 206, 578–584. doi:10.1016/j.prp.2010.03.005
- Besse, B., and Le Chevalier, T. (2012). Developments in the treatment of early NSCLC: when to use chemotherapy. *Ann. Oncol.* 23(Suppl. 10), x52–x59. doi:10.1093/annonc/mds347
- Bian, H. B., Pan, X., Yang, J. S., Wang, Z. X., and De, W. (2011). Upregulation of microRNA-451 increases cisplatin sensitivity of non-small cell lung cancer cell line (A549). *J. Exp. Clin. Cancer Res.* 30, 20. doi:10.1186/1756-9966-30-20
- Boeri, M., Verri, C., Conte, D., Roz, L., Modena, P., Facchinetti, F., et al. (2011). MicroRNA signatures in tissues and plasma predict development and prognosis of computed tomography detected lung cancer. *Proc. Natl. Acad. Sci. U.S.A.* 108, 3713–3718. doi:10.1073/pnas.110004 8108
- Bronte, G., Rizzo, S., La Paglia, L., Adamo, V., Siragusa, S., Ficorella, C., et al. (2010). Driver mutations and differential sensitivity to targeted therapies: a new approach to the treatment of lung adenocarcinoma. *Cancer Treat. Rev.* 36(Suppl. 3), S21–S29. doi:10.1016/S0305-7372(10)70016-5
- Calin, G. A., Dumitru, C. D., Shimizu, M., Bichi, R., Zupo, S., Noch, E., et al. (2002). Frequent deletions and down-regulation of micro-RNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. *Proc. Natl. Acad. Sci. U.S.A.* 99, 15524–15529. doi:10.1073/pnas.242606799
- Calin, G. A., Ferracin, M., Cimmino, A., Di Leva, G., Shimizu, M., Wojcik, S. E., et al. (2005). A microRNA signature associated with prognosis and progression in chronic lymphocytic leukemia. *N. Engl. J. Med.* 353, 1793–1801. doi:10.1056/NEJMoa050995
- Calin, G. A., Sevignani, C., Dumitru, C. D., Hyslop, T., Noch, E., Yendamuri, S., et al. (2004). Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. *Proc. Natl. Acad. Sci. U.S.A.* 101, 2999–3004. doi:10.1073/pnas.0307323101
- Carleton, M., Cleary, M. A., and Linsley, P. S. (2007). MicroR-NAs and cell cycle regulation.

Cell Cycle 6, 2127–2132. doi:10.4161/cc.6.17.4641

- Cha, S. T., Chen, P. S., Johansson, G., Chu, C. Y., Wang, M. Y., Jeng, Y. M., et al. (2010). MicroRNA-519c suppresses hypoxia-inducible factor-1alpha expression and tumor angiogenesis. *Cancer Res.* 70, 2675–2685. doi:10.1158/0008-5472.CAN-09-2448
- Chen, X., Ba, Y., Ma, L., Cai, X., Yin, Y., Wang, K., et al. (2008). Characterization of microRNAs in serum: a novel class of biomarkers for diagnosis of cancer and other diseases. *Cell Res.* 18, 997–1006. doi:10.1038/cr.2008.282
- Cheng, A. M., Byrom, M. W., Shelton, J., and Ford, L. P. (2005). Antisense inhibition of human miRNAs and indications for an involvement of miRNA in cell growth and apoptosis. *Nucleic Acids Res.* 33, 1290–1297. doi:10.1093/nar/gki200
- Chin, L. J., Ratner, E., Leng, S., Zhai, R., Nallur, S., Babar, I., et al. (2008). A SNP in a let-7 microRNA complementary site in the KRAS 3' untranslated region increases non-small cell lung cancer risk. *Cancer Res.* 68, 8535–8540. doi:10.1158/0008-5472.CAN-08-2129
- Chivukula, R. R., and Mendell, J. T. (2008). Circular reasoning: microRNAs and cell-cycle control. *Trends Biochem. Sci.* 33, 474–481. doi:10.1016/j.tibs.2008.06.008
- Cho, W. C., Chow, A. S., and Au, J. S. (2009). Restoration of tumour suppressor hsa-miR-145 inhibits cancer cell growth in lung adenocarcinoma patients with epidermal growth factor receptor mutation. *Eur. J. Cancer* 45, 2197–2206. doi:10.1016/j.ejca.2009.04.039
- Donnem, T., Eklo, K., Berg, T., Sorbye, S. W., Lonvik, K., Al-Saad, S., et al. (2011). Prognostic impact of MiR-155 in non-small cell lung cancer evaluated by in situ hybridization. *J. Transl. Med.* 9, 6. doi:10.1186/1479-5876-9-6
- Du, L., Schageman, J. J., Subauste, M. C., Saber, B., Hammond, S. M., Prudkin, L., et al. (2009). miR-93, miR-98, and miR-197 regulate expression of tumor suppressor gene FUS1. *Mol. Cancer Res.* 7, 1234–1243. doi:10.1158/1541-7786.MCR-08-0507
- Fabbri, M., Garzon, R., Cimmino, A., Liu, Z., Zanesi, N., Callegari, E., et al. (2007). MicroRNA-29 family reverts aberrant methylation in lung cancer by targeting DNA methyltransferases 3A and 3B. *Proc. Natl. Acad. Sci. U.S.A.* 104, 15805–15810. doi:10.1073/pnas.0707628104

- Fabbri, M., Paone, A., Calore, F., Galli, R., Gaudio, E., Santhanam, R., et al. (2012). MicroRNAs bind to Toll-like receptors to induce prometastatic inflammatory response. *Proc. Natl. Acad. Sci. U.S.A.* 109, E2110–E2116. doi:10.1073/pnas.1209414109
- Feng, S., Cong, S., Zhang, X., Bao, X., Wang, W., Li, H., et al. (2011). MicroRNA-192 targeting retinoblastoma 1 inhibits cell proliferation and induces cell apoptosis in lung cancer cells. *Nucleic Acids Res.* 39, 6669–6678. doi:10.1093/nar/ gkr232
- Ferlay, J., Parkin, D. M., and Steliarova-Foucher, E. (2010). Estimates of cancer incidence and mortality in Europe in 2008. *Eur. J. Cancer* 46, 765–781. doi:10.1016/j.ejca.2009.12.014
- Gallardo, E., Navarro, A., Vinolas, N., Marrades, R. M., Diaz, T., Gel, B., et al. (2009). miR-34a as a prognostic marker of relapse in surgically resected non-small-cell lung cancer. *Carcinogenesis* 30, 1903–1909. doi:10.1093/carcin/bgp219
- Galluzzi, L., Morselli, E., Vitale, I., Kepp, O., Senovilla, L., Criollo, A., et al. (2010). miR-181a and miR-630 regulate cisplatin-induced cancer cell death. *Cancer Res.* 70, 1793–1803. doi:10.1158/0008-5472.CAN-09-3112
- Gao, W., Yu, Y., Cao, H., Shen, H., Li, X., Pan, S., et al. (2010). Deregulated expression of miR-21, miR-143 and miR-181a in non small cell lung cancer is related to clinicopathologic characteristics or patient prognosis. *Biomed. Pharmacother.* 64, 399–408. doi:10.1016/j.biopha.2010.01.018
- Garber, K. (2010). ALK, lung cancer, and personalized therapy: portent of the future? J. Natl. Cancer Inst. 102, 672–675. doi:10.1093/jnci/djq184
- Garofalo, M., Di Leva, G., Romano, G., Nuovo, G., Suh, S. S., Ngankeu, A., et al. (2009). miR-221&222 regulate TRAIL resistance and enhance tumorigenicity through PTEN and TIMP3 downregulation. *Cancer Cell* 16, 498–509.
- Garofalo, M., Quintavalle, C., Di Leva, G., Zanca, C., Romano, G., Taccioli, C., et al. (2008). MicroRNA signatures of TRAIL resistance in human non-small cell lung cancer. Oncogene 27, 3845–3855. doi:10.1038/onc.2008.6
- Hayashita, Y., Osada, H., Tatematsu, Y., Yamada, H., Yanagisawa, K., Tomida, S., et al. (2005). A polycistronic microRNA cluster, miR-17-92, is overexpressed in human lung cancers and enhances cell proliferation. *Cancer Res.* 65, 9628–9632.

doi:10.1158/0008-5472.CAN-05-

- Hu, Z., Chen, X., Zhao, Y., Tian, T., Jin, G., Shu, Y., et al. (2010). Serum microRNA signatures identified in a genome-wide serum microRNA expression profiling predict survival of non-small-cell lung cancer. J. Clin. Oncol. 28, 1721–1726. doi:10.1200/JCO.2009.24. 9342
- Incoronato, M., Garofalo, M., Urso, L., Romano, G., Quintavalle, C., Zanca, C., et al. (2010). miR-212 increases tumor necrosis factorrelated apoptosis-inducing ligand sensitivity in non-small cell lung cancer by targeting the antiapoptotic protein PED. *Cancer Res.* 70, 3638–3646. doi:10.1158/0008-5472.CAN-09-3341
- Iorio, M. V., Ferracin, M., Liu, C. G., Veronese, A., Spizzo, R., Sabbioni, S., et al. (2005). MicroRNA gene expression deregulation in human breast cancer. *Cancer Res.* 65, 7065–7070. doi:10.1158/0008-5472.CAN-05-1783
- Izzotti, A., Calin, G. A., Arrigo, P., Steele, V. E., Croce, C. M., and De Flora, S. (2009). Downregulation of microRNA expression in the lungs of rats exposed to cigarette smoke. *FASEB J.* 23, 806–812. doi:10.1096/fj.08-121384
- Janku, F., Stewart, D. J., and Kurzrock, R. (2010). Targeted therapy in non-small-cell lung cancer – is it becoming a reality? *Nat. Rev. Clin. Oncol.* 7, 401–414. doi:10.1038/nrclinonc.2010.64
- Johnson, C. D., Esquela-Kerscher, A., Stefani, G., Byrom, M., Kelnar, K., Ovcharenko, D., et al. (2007). The let-7 microRNA represses cell proliferation pathways in human cells. *Cancer Res.* 67, 7713–7722. doi:10.1158/0008-5472.CAN-07-1083
- Johnson, S. M., Grosshans, H., Shingara, J., Byrom, M., Jarvis, R., Cheng, A., et al. (2005). RAS is regulated by the let-7 microRNA family. *Cell* 120, 635–647. doi:10.1016/j.cell.2005.01.014
- Kumar, M. S., Erkeland, S. J., Pester, R. E., Chen, C. Y., Ebert, M. S., Sharp, P. A., et al. (2008). Suppression of non-small cell lung tumor development by the let-7 microRNA family. *Proc. Natl. Acad. Sci. U.S.A.* 105, 3903–3908. doi:10.1073/pnas.0712321105
- Landi, M. T., Zhao, Y., Rotunno, M., Koshiol, J., Liu, H., Bergen, A. W., et al. (2010). MicroRNA expression differentiates histology and predicts survival of lung cancer. *Clin. Cancer*

Res. 16, 430-441. doi:10.1158/1078-0432.CCR-09-1736

- Lebanony, D., Benjamin, H., Gilad, S., Ezagouri, M., Dov, A., Ashkenazi, K., et al. (2009). Diagnostic assay based on hsa-miR-205 expression distinguishes squamous from nonsquamous non-small-cell lung carcinoma. J. Clin. Oncol. 27, 2030–2037. doi:10.1200/JCO.2008.19.4134
- Lee, Y. S., and Dutta, A. (2007). The tumor suppressor microRNA let-7 represses the HMGA2 oncogene. *Genes Dev.* 21, 1025–1030. doi:10.1101/gad.1540407
- Lu, Y., Govindan, R., Wang, L., Liu, P. Y., Goodgame, B., Wen, W., et al. (2012). MicroRNA profiling and prediction of recurrence/relapsefree survival in stage I lung cancer. *Carcinogenesis* 33, 1046–1054. doi:10.1093/carcin/bgs100
- Lu, Y., Thomson, J. M., Wong, H. Y., Hammond, S. M., and Hogan, B. L. (2007). Transgenic over-expression of the microRNA miR-17-92 cluster promotes proliferation and inhibits differentiation of lung epithelial progenitor cells. *Dev. Biol.* 310, 442–453. doi:10.1016/j.ydbio.2007.08.007
- Lynch, T. J., Bell, D. W., Sordella, R., Gurubhagavatula, S., Okimoto, R. A., Brannigan, B. W., et al. (2004). Activating mutations in the epidermal growth factor receptor underlying responsiveness of nonsmall-cell lung cancer to gefitinib. *N. Engl. J. Med.* 350, 2129–2139. doi:10.1056/NEJMoa040938
- Matsubara, H., Takeuchi, T., Nishikawa, E., Yanagisawa, K., Hayashita, Y., Ebi, H., et al. (2007). Apoptosis induction by antisense oligonucleotides against miR-17-5p and miR-20a in lung cancers overexpressing miR-17-92. Oncogene 26, 6099–6105. doi:10.1038/sj.onc.1210425
- Mott, J. L., Kobayashi, S., Bronk, S. F., and Gores, G. J. (2007). Mir-29 regulates Mcl-1 protein expression and apoptosis. *Oncogene* 26, 6133–6140. doi:10.1038/sj.onc.1210436
- Muniyappa, M. K., Dowling, P., Henry, M., Meleady, P., Doolan, P., Gammell, P., et al. (2009). MiRNA-29a regulates the expression of numerous proteins and reduces the invasiveness and proliferation of human carcinoma cell lines. *Eur. J. Cancer* 45, 3104–3118. doi:10.1016/j.ejca.2009.09.014
- Nakamura, T., Canaani, E., and Croce, C. M. (2007). Oncogenic All1 fusion proteins target Drosha-mediated microRNA processing. *Proc. Natl. Acad. Sci. U.S.A.* 104, 10980–10985. doi:10.1073/pnas.0704559104

- Navarro, A., Marrades, R. M., Vinolas, N., Quera, A., Agusti, C., Huerta, A., et al. (2009). MicroRNAs expressed during lung cancer development are expressed in human pseudoglandular lung embryogenesis. *Oncology* 76, 162–169. doi:10.1159/000201569
- Paez, J. G., Janne, P. A., Lee, J. C., Tracy, S., Greulich, H., Gabriel, S., et al. (2004). EGFR mutations in lung cancer: correlation with clinical response to gefitinib therapy. *Science* 304, 1497–1500. doi:10.1126/science.1099314
- Pao, W., and Girard, N. (2011). New driver mutations in non-smallcell lung cancer. *Lancet Oncol.* 12, 175–180. doi:10.1016/S1470-2045(10)70087-5
- Pao, W., Miller, V., Zakowski, M., Doherty, J., Politi, K., Sarkaria, I., et al. (2004). EGF receptor gene mutations are common in lung cancers from "never smokers" and are associated with sensitivity of tumors to gefitinib and erlotinib. *Proc. Natl. Acad. Sci. U.S.A.* 101, 13306–13311. doi:10.1073/pnas.0405220101
- Patnaik, S. K., Kannisto, E., Knudsen, S., and Yendamuri, S. (2010). Evaluation of microRNA expression profiles that may predict recurrence of localized stage I non-small cell lung cancer after surgical resection. *Cancer Res.* 70, 36–45. doi:10.1158/0008-5472.CAN-09-3153
- Prenzel, N., Fischer, O. M., Streit, S., Hart, S., and Ullrich, A. (2001). The epidermal growth factor receptor family as a central element for cellular signal transduction and diversification. *Endocr. Relat. Cancer* 8, 11–31. doi:10.1677/erc.0.0080011
- Rabinowits, G., Gercel-Taylor, C., Day, J. M., Taylor, D. D., and Kloecker, G. H. (2009). Exosomal microRNA: a diagnostic marker for lung cancer. *Clin. Lung Cancer* 10, 42–46. doi:10.3816/CLC.2009.n.006
- Rai, K., Takigawa, N., Ito, S., Kashihara, H., Ichihara, E., Yasuda, T., et al. (2011). Liposomal delivery of microRNA-7-expressing plasmid overcomes epidermal growth factor receptor tyrosine kinase inhibitorresistance in lung cancer cells. *Mol. Cancer Ther.* 10, 1720–1727. doi:10.1158/1535-7163.MCT-11-0220
- Raponi, M., Dossey, L., Jatkoe, T., Wu, X., Chen, G., Fan, H., et al. (2009). MicroRNA classifiers for predicting prognosis of squamous cell lung cancer. *Cancer Res.* 69, 5776–5783. doi:10.1158/0008-5472.CAN-09-0587
- Rothschild, S. I., Tschan, M. P., Federzoni, E. A., Jaggi, R.,

Fey, M. F., Gugger, M., et al. (2012a). MicroRNA-29b is involved in the Src-ID1 signaling pathway and is dysregulated in human lung adenocarcinoma. *Oncogene* 31, 4221–4232. doi:10.1038/onc.2011.578

- Rothschild, S. I., Tschan, M. P., Jaggi, R., Fey, M. F., Gugger, M., and Gautschi, O. (2012b). MicroRNA-381 represses ID1 and is deregulated in lung adenocarcinoma. *J. Thorac. Oncol.* 7, 1069–1077. doi:10.1097/JTO.0b013e31824fe976
- Saito, Y., Liang, G., Egger, G., Friedman, J. M., Chuang, J. C., Coetzee, G. A., et al. (2006). Specific activation of microRNA-127 with downregulation of the protooncogene BCL6 by chromatinmodifying drugs in human cancer cells. *Cancer Cell* 9, 435–443. doi:10.1016/j.ccr.2006.04.020
- Sampson, V. B., Rong, N. H., Han, J., Yang, Q., Aris, V., Soteropoulos, P., et al. (2007). MicroRNA let-7a down-regulates MYC and reverts MYC-induced growth in Burkitt lymphoma cells. *Cancer Res.* 6, 9762–9770. doi:10.1158/0008-5472.CAN-07-2462
- Scagliotti, G., Hanna, N., Fossella, F., Sugarman, K., Blatter, J., Peterson, P., et al. (2009). The differential efficacy of pemetrexed according to NSCLC histology: a review of two Phase III studies. Oncologist 14, 253–263. doi:10.1634/theoncologist.2008-0232
- Schaefer, U., Voloshanenko, O., Willen, D., and Walczak, H. (2007). TRAIL: a multifunctional cytokine. *Front. Biosci.* 12:3813–3824. doi:10.2741/2354
- Seike, M., Goto, A., Okano, T., Bowman, E. D., Schetter, A. J., Horikawa, I., et al. (2009). MiR-21 is an EGFR-regulated antiapoptotic factor in lung cancer in never-smokers. *Proc. Natl. Acad. Sci. U.S.A.* 106, 12085–12090. doi:10.1073/pnas.0905234106
- Semenza, G. L. (2010). Defining the role of hypoxia-inducible factor 1 in cancer biology and therapeutics. Oncogene 29, 625–634. doi:10.1038/onc.2009.441
- Shen, J., Todd, N. W., Zhang, H., Yu, L., Lingxiao, X., Mei, Y., et al. (2011). Plasma microRNAs as potential biomarkers for non-small-cell lung cancer. *Lab. Invest.* 91, 579–587. doi:10.1038/labinvest.2010.194
- Sun, Y., Fang, R., Li, C., Li, L., Li, F., Ye, X., et al. (2010). Hsa-mir-182 suppresses lung tumorigenesis through down regulation of RGS17 expression in vitro. *Biochem. Biophys.*

Res. Commun. 396, 501–507. doi:10.1016/j.bbrc.2010.04.127

- Taguchi, A., Yanagisawa, K., Tanaka, M., Cao, K., Matsuyama, Y., Goto, H., et al. (2008). Identification of hypoxia-inducible factor-1 alpha as a novel target for miR-17-92 microRNA cluster. *Cancer Res.* 68, 5540–5545. doi:10.1158/0008-5472.CAN-07-6460
- Takahashi, Y., Forrest, A. R., Maeno, E., Hashimoto, T., Daub, C. O., and Yasuda, J. (2009). MiR-107 and MiR-185 can induce cell cycle arrest in human non small cell lung cancer cell lines. *PLoS ONE* 4:e6677. doi:10.1371/journal.pone.0006677
- Takamizawa, J., Konishi, H., Yanagisawa, K., Tomida, S., Osada, H., Endoh, H., et al. (2004). Reduced expression of the let-7 microR-NAs in human lung cancers in association with shortened postoperative survival. *Cancer Res.* 64, 3753–3756. doi:10.1158/0008-5472.CAN-04-0637
- Tan, X., Qin, W., Zhang, L., Hang, J., Li, B., Zhang, C., et al. (2011). A 5-microRNA signature for lung squamous cell carcinoma diagnosis and hsa-miR-31 for prognosis. *Clin. Cancer Res.* 17, 6802–6811. doi:10.1158/1078-0432.CCR-11-0419
- Trang, P., Medina, P. P., Wiggins, J. F., Ruffino, L., Kelnar, K., Omotola, M., et al. (2010). Regression of murine lung tumors by the let-7 microRNA. *Oncogene* 29, 1580–1587. doi:10.1038/onc.2009.445
- Ventura, A., Young, A. G., Winslow, M. M., Lintault, L., Meissner, A., Erkeland, S. J., et al. (2008). Targeted deletion reveals essential and overlapping functions of the miR-17 through 92 family of miRNA clusters. *Cell* 132, 875–886. doi:10.1016/j.cell.2008.02.019
- Volinia, S., Calin, G. A., Liu, C. G., Ambs, S., Cimmino, A., Petrocca, F., et al. (2006). A microRNA expression signature of human solid tumors defines cancer gene targets. *Proc. Natl. Acad. Sci. U.S.A.* 103, 2257–2261. doi:10.1073/pnas.0510565103
- Voortman, J., Goto, A., Mendiboure, J., Sohn, J. J., Schetter, A. J., Saito, M., et al. (2010). MicroRNA expression and clinical outcomes in patients treated with adjuvant chemotherapy after complete resection of non-small cell lung carcinoma. *Cancer Res.* 70, 8288–8298. doi:10.1158/0008-5472.CAN-10-1348
- Wang, G., Mao, W., Zheng, S., and Ye, J. (2009). Epidermal growth

factor receptor-regulated miR-125a-5p – a metastatic inhibitor of lung cancer. *FEBS J.* 276, 5571–5578. doi:10.1111/j.1742-4658.2009.07238.x

- Wang, Q., Wang, S., Wang, H., Li, P., and Ma, Z. (2012). MicroRNAs: novel biomarkers for lung cancer diagnosis, prediction and treatment. *Exp. Biol. Med. (Maywood)* 237, 227–235. doi:10.1258/ebm.2011.01 1192
- Wang, R., Wang, Z. X., Yang, J. S., Pan, X., De, W., and Chen, L. B. (2011). MicroRNA-451 functions as a tumor suppressor in human non-small cell lung cancer by targeting ras-related protein 14 (RAB14). Oncogene 30, 2644–2658. doi:10.1038/onc.2010.642
- Webster, R. J., Giles, K. M., Price, K. J., Zhang, P. M., Mattick, J. S., and Leedman, P. J. (2009). Regulation of epidermal growth factor receptor signaling in human cancer cells by microRNA-7. J. Biol. Chem. 284, 5731–5741. doi:10.1074/jbc.M804280200
- Weidhaas, J. B., Babar, I., Nallur, S. M., Trang, P., Roush, S., Boehm, M., et al. (2007). MicroRNAs as potential agents to alter resistance to cytotoxic anticancer therapy. *Cancer Res.*

67, 11111–11116. doi:10.1158/0008-5472.CAN-07-2858

- Weiss, G. J., Bemis, L. T., Nakajima, E., Sugita, M., Birks, D. K., Robinson, W. A., et al. (2008). EGFR regulation by microRNA in lung cancer: correlation with clinical response and survival to gefitinib and EGFR expression in cell lines. *Ann. Oncol.* 19, 1053–1059. doi:10.1093/annonc/mdn006
- Wu, X., Piper-Hunter, M. G., Crawford, M., Nuovo, G. J., Marsh, C. B., Otterson, G. A., et al. (2009). MicroR-NAs in the pathogenesis of lung cancer. J. Thorac. Oncol. 4, 1028–1034. doi:10.1097/JTO.0b013e3181a99c77
- Xie, Y., Todd, N. W., Liu, Z., Zhan, M., Fang, H., Peng, H., et al. (2010). Altered miRNA expression in sputum for diagnosis of non-small cell lung cancer. *Lung Cancer* 67, 170–176. doi:10.1016/j.lungcan.2009.04.004
- Xing, L., Todd, N. W., Yu, L., Fang, H., and Jiang, F. (2010). Early detection of squamous cell lung cancer in sputum by a panel of microRNA markers. *Mod. Pathol.* 23, 1157–1164. doi:10.1038/modpathol.2010.111
- Xu, H. I., Cheung, Y., Guo, H. F., and Cheung, N. K. (2009). MicroRNA miR-29 modulates expression

of immunoinhibitory molecule B7-H3: potential implications for immune based therapy of human solid tumors. *Cancer Res.* 69, 6275–6281. doi:10.1158/0008-5472.CAN-08-4517

- Yanaihara, N., Caplen, N., Bowman, E., Seike, M., Kumamoto, K., Yi, M., et al. (2006). Unique microRNA molecular profiles in lung cancer diagnosis and prognosis. *Cancer Cell* 9, 189–198.
- Yu, L., Todd, N. W., Xing, L., Xie, Y., Zhang, H., Liu, Z., et al. (2010). Early detection of lung adenocarcinoma in sputum by a panel of microRNA markers. *Int. J. Cancer* 127, 2870–2878. doi:10.1002/ijc.25289
- Yu, S. L., Chen, H. Y., Chang, G. C., Chen, C. Y., Chen, H. W., Singh, S., et al. (2008). MicroRNA signature predicts survival and relapse in lung cancer. *Cancer Cell* 13, 48–57. doi:10.1016/j.ccr.2007.12.008
- Zhang, J. G., Guo, J. F., Liu, D. L., Liu, Q., and Wang, J. J. (2011). MicroRNA-101 exerts tumorsuppressive functions in non-small cell lung cancer through directly targeting enhancer of zeste homolog 2. J. Thorac. Oncol. 6, 671–678. doi:10.1097/JTO.0b013e318208eb35

Zhong, M., Ma, X., Sun, C., and Chen, L. (2010). MicroRNAs reduce tumor growth and contribute to enhance cytotoxicity induced by gefitinib in non-small cell lung cancer. *Chem. Biol. Interact.* 184, 431–438. doi:10.1016/j.cbi.2010.01.025

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