



Human Papilloma Viruses and Breast Cancer – Assessment of Causality

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High risk human papilloma viruses (HPVs) may have a causal role in some breast cancers. Case-control studies, conducted in many different countries, consistently indicate that HPVs are more frequently present in breast cancers as compared to benign breast and normal breast controls (odds ratio 4.02). The assessment of causality of HPVs in breast cancer is difficult because (i) the HPV viral load is extremely low, (ii) HPV infections are common but HPV associated breast cancers are uncommon, and (iii) HPV infections may precede the development of breast and other cancers by years or even decades. Further, HPV oncogenesis can be indirect. Despite these difficulties, the emergence of new evidence has made the assessment of HPV causality, in breast cancer, a practical proposition. With one exception, the evidence meets all the conventional criteria for a causal role of HPVs in breast cancer. The exception is "specificity." HPVs are ubiquitous, which is the exact opposite of specificity. An additional reservation is that the prevalence of breast cancer is not increased in immunocompromised patients as is the case with respect to HPV-associated cervical cancer. This indicates that HPVs may have an indirect causal influence in breast cancer. Based on the overall evidence, high-risk HPVs may have a causal role in some breast cancers.

Keywords: breast cancer, human papilloma virus, HPV, causality

INTRODUCTION

High-risk human papilloma viruses (HPVs) may have a causal role in some breast cancers. However, the assessment of causality of HPVs in breast cancer is extremely difficult. The main difficulty is that HPV infections are common, but HPV-associated breast cancers are uncommon. Further, HPV infections may precede the development of breast and other cancers by years or even decades. Further, HPV oncogenesis can be indirect. Despite these difficulties, the emergence of new evidence has made this assessment a practical proposition.

A good starting point is to review the evidence according to the criteria developed by A. Bradford Hill in 1965 and later modified by others (1–3). The Hill criteria are (i) strength of the association, (ii) consistency, (iii) specificity, (iv) temporality (time sequence), (v) biological gradient, (vi) plausibility, (vii) coherence, (viii) experimental evidence, and (ix) analogy. There is an overlap between several criteria such as strength of the association and specificity. If viruses, such as high risk for cancer HPVs, fulfill all or most of these criteria, there is a likely relationship, but not necessarily a conclusive confirmation of causality. One weakness of the Hill criteria is that each is given similar weight or importance. This is in contrast to the key requirement of zur Hausen that there should be a regular presence of the nucleic acid of the virus in specific malignant tumors (3). Even this criteria may

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be difficult because of the ability of HPVs to act indirectly by causing genetic instability or promoting the influence of other viruses such as Epstein Barr virus (EBV).

Strength of the Association between Identification of HPV and Breast Cancer

The HPV viral load in breast cancer is extremely low as compared to HPV in cervical cancer (approximately 2,000-fold lower, see **Table 1**) (4, 5). As a consequence, identification of HPV in breast tumors is difficult whether by amplification techniques such as polymerase chain reaction (PCR) or massive parallel sequencing. The low viral load is the likely reason that several laboratories have not been able to detect HPV. On the other hand, HPV gene sequences have been identified in breast tumors in over 40 studies conducted in 20 countries (6, 7).

The prevalence of high-risk HPV positive breast cancers varies between 0 and 2% in some Chinese Provinces to 86% in North America (7). HPV types 16 and 18 are the most prevalent, but HPV 33 and 58 are common in China and Japan (8, 9). A recent meta-analysis of 22 case–control studies indicated that HPV infection was associated with an increased risk of breast cancer as compared to benign or normal breast controls (odds ratio 4.02) (7). Increased risk of breast cancer is influenced by geographic region, PCR techniques, and publication period. All of these studies were based on PCR analyses. HPV gene sequences identified by *in situ* PCR are shown in **Figure 1**. High risk HPVs have been identified in invasive breast cancers using Next Generation Sequencing (NGS), but this technique is much less sensitive than PCR (10).

Case-control studies, which compare the prevalence of high-risk HPVs in breast cancer as compared to benign breast or normal breast are shown in **Table 2**. The prevalence of HPV is fourfold higher in breast cancer (21.5%) than controls (5.1%) in studies conducted in different laboratories in a wide range of countries. This parallels the findings of a recent meta-analysis based on selected studies, which are included in this Table (7). It is of interest that high-risk HPVs were identified in only 4.1% of benign or normal (usually from cosmetic surgery) as compared to 17.7% in non-cancerous breast tissues located next to breast cancer tissues. The implication is that HPVs may be have increased prevalence in non-cancerous breast tissues located next to cancerous breast tissues and should not be used as controls.

In a recent study, it has been shown that women with HPVassociated cervical pathology are at increased risk from the same HPV type positive breast cancer (31). This observation confirms

TABLE 1 | HPV viral loads in HeLa (cervical cancer cell line), breast cancer tissues, and cervical cancer tissues (4, 5).

	HeLa cells	Breast cancer	Cervical cancer
HPV gene copy/cell	10–50	0.00054-0.0021	13
HPV gene copy/ng	1,510–7,558	0.082-0.32	2,000
HPV gene copy/50 ng	75,500–377,000	4–16	98,500

The viral load is approximately 2,000-fold greater in cervical cancer as compared to breast cancer. ng = nanogram. large epidemiological studies, which show that women with squamous or glandular precancer of the cervix have a significantly higher risk of subsequent breast cancer than the general female population – odds ratios 1.10 and 1.52, respectively (32, 33).

Consistency

The HPV viral load in breast cancer is extremely low (see Table 1). Accordingly, the identification of HPVs by both PCR and NGS is difficult. This may explain the very different findings from the 10 case-control studies conducted in various locations in China. The identification of HPVs in China vary from 0 to 60%. Similarly, the findings in Iran vary from 0 to 34%. There are three relevant studies based on NGS - Tang et al., Khoury et al., and Lawson et al. (10, 34, 35). High risk HPVs were not identified in breast cancers from the The Cancer Genome Atlas (TCGA) series of approximately 800 breast cancers in either the Tang or Khoury studies but were identified in 2.3% of the same series by Lawson et al. (10, 34, 35). Clearly, PCR-based analyses are much more sensitive than current NGS-based analyses. The reasons for this discrepancy have been considered by Vinner et al. who has shown that NGS-based techniques are unlikely to detect viruses with very low loads in cancers as compared to amplification techniques such as PCR (36).

Despite these differences, the outcome of the PCR-based casecontrol studies is consistent despite having been conducted with different techniques, locations, and times by different research groups (**Table 2**). As stated above, the prevalence of high-risk HPVs in breast cancers is fourfold higher than in normal and benign breast controls. This is a substantial difference between benign breast and breast cancers and cannot be attributed to differences in laboratory techniques.



FIGURE 1 | (A) HPV identified by *in situ* PCR in breast ductal carcinoma *in situ*. Koilocytes are present in this specimen. (B,C) HPV E7 protein in a benign breast specimen (B). No HPV E7 protein in invasive breast cancer 4 years later in the same patient (C).

TABLE 2 | Identification of high-risk HPVs in breast cancer and benign or normal breast controls in case-control studies.

Reference Country		HPV-positive breast cancers/total breast cancers	HPV-positive non-cancer breast/total non-cancer breast controls	Main HPV types	
Yu et al. (8)	Japan/China	18/52 (35%)	0/15 (0%)	18, 33	
Damin et al. (11)	Brazil	25/101 (25%)	0/41 (0%)	16, 18	
Tsai et al. (12)	Taiwan	8/62 (13%)	2/42 (5%)	Not available	
Choi et al. (13)	Korea	8/123 (7%)	0/31 (0%)	16, 18, 58	
Gumus et al. (14)	Turkey	37/50 (74%)	9/16 (56%)	18, 33	
He et al. (15)	China	24/40 (60%)	1/20 (5%)	16	
de Leon et al. (16)	Mexico	15/41 (37%)	0/43 (0%)	16, 18	
Heng et al. (17)	Australia	8/26 (31%)	3/28 (11%)	16, 18	
Herrera-Romano et al. (5)	Mexico	6/60 (10%)	7/60 (12%)	16	
Vlou et al. (18)	China	4/62 (6%)	0/46 (0%)	16, 18	
Chang et al. (19)	China	0/48 (0%)	3/30 (10%)	6, 11	
Sigaroodi et al. (20)	Iran	15/43 (35%)	1/40 (3%)	16, 18	
Frega et al. (21)	Italy	9/31 (29%)	0/12 (0%)	16, 18	
Glenn et al. (22)	Australia	25/50 (50%)	8/40 (20%)	16, 18	
Liang et al. (23)	China	48/224 (21%)	6/37 (16%)	16, 18, 33, 58	
Ahangar-Oskouee et al. (24)	Iran	22/65 (34%)	0/65 (0%)	16	
Ali et al. (25)	Iraq	60/129 (47%)	3/41 (7%)	16, 18, 33	
Eslamifar et al. (26)	Iran	0/100 (0%)	0/50 (0%)		
Manzouri et al. (27)	Iran	10/55 (18%)	7/51 (14%)	16	
Peng et al. (28)	China	2/100 (2%)	0/50 (0%)	18	
Fu et al. (9)	China	25/169 (15%)	1/83 (1%)	58	
Li et al. (29)	China	3/187 (2%)	0/92 (0%)	6, 16, 18	
Zhou et al. (6)	China	0/77 (0%)	0/77 (0%)		
Vernet-Tomas et al. (30)	Spain	0/76 (0%)	0/2 (0%)		

These data indicate that high-risk HPVs are fourfold more prevalent in breast tumors as compared to non-cancer controls. This parallels the findings of a recent meta-analysis based on selected studies included in this Table (7).

Epidemiology

If HPV infections were major contributors to breast cancer, it would be anticipated that countries with high rates of HPV associated cervical cancer would also have high rates of HPV associated breast cancer. The opposite appears likely, namely that countries with low rates of cervical cancer appear to have high rates of breast cancer (37). This is shown in **Table 3**. While these data must be considered only as broad trends, the implication is that HPVs are unlikely to be major contributors to breast cancer. However, it remains possible that HPVs may be involved in some but not all breast cancers.

Epidemiological combined with experimental data have demonstrated that (i) the prevalence of high-risk HPVs is high in early as compared to late breast cancer, (ii) this early influence of HPVs is indirect and operates by the destabilizing effects of HPV on the human genome *via* APOBEC3B enzyme (38). This influence of HPV on APOBEC enzymes has been confirmed (39). It has also been demonstrated that high expression of HPV E7 protein in benign breast tissues as compared to low HPV E7 expression in subsequent HPV positive breast cancer occurs in the same patients (40). This is additional evidence that the influence of HPVs occurs early in breast cancer.

This indirect influence of HPVs in breast cancer needs to be considered in the context of the epidemiological observations by Grulich and Vajdic (41) that the prevalence of breast cancer is not increased in immunocompromised patients (patients with HIV infections or organ transplant recipients). This compares to the fourfold to sixfold increase in cervical cancer in these patients. Again the implication is that the influences of HPV are indirect and occur early in breast oncogenesis.

There is also evidence that HPVs may collaborate with EBV and which together may have oncogenic influences. The co-prevalence of HPVs and EBV in nasopharyngeal and breast cancers has been demonstrated (22, 42, 43).

Specificity

Specificity is a very weak causative criterion in both communicable and non-communicable diseases and probably should not be included in the Hill list of causal criteria. Any associations between HPVs and breast cancer are obviously not specific. HPV is well established as a causal factor in other cancers such as cervical and head and neck cancers. In addition, high-risk HPVs have been identified in a wide range of other cancers and non-cancerous tissues (44). In the past, HPVs have been regarded as being specific to epithelial cells including breast epithelial cells. However, HPV gene sequences have been identified in non-epithelial cell brain tumors and in coronary and aorta artery smooth muscle cells (45).

Temporality (Time Sequence)

High-risk HPVs have been identified in benign breast tissues of the same type as in breast tumors, which subsequently developed in the same patients (10). The identification of HPVs preceded

TABLE 3 | Incidence and mortality per 100,000 women (age adjusted) of cervical and breast cancers.

	Incidence	Incidence	Mortality	Mortality	
	cervical	breast	cervical	breast	
	cancer	cancer	cancer	cancer	
France	8	92	2	17	
England	7	90	2	18	
United States	5	84	4	14	
Australia	5	82	2	14	
Canada	5	76	3	15	
China	6	44	5	5	
India	16	32	10	10	
Japan	7	32	4	8	
Thailand	23	25	10	10	

Selected high and low cervical cancer country data (17).

Low cervical cancer correlates with higher breast cancer incidence and mortality (incidence p = 0.052, mortality p = 0.118). As HPV is almost universally associated with cervical cancer, these data suggest that HPV is not a major cause of breast cancer. These data demonstrate trends only. Incidence is influenced by screening, and mortality is influenced by accuracy of diagnosis and effectiveness of treatment.

the development of breast cancer in the same patients by between 1 and 11 years. As this study was retrospective, the time of the original HPV infection cannot be determined. Similarly, HPV-associated cervical infections and abnormal cervical pathology can precede the development of same type HPV positive breast cancer in the same patient (38, 46). Again, however, the time of the original HPV infection cannot be determined.

These observations confirm that the HPV infections precede the development of HPV-positive breast cancer.

Biological Gradient, Plausibility, and Coherence

These criteria overlap. The biological gradient or response differs greatly with respect to the HPV viral load in breast as compared to cervical cancer. The HPV viral load in breast cancer is 2,000-fold less than in cervical cancer, as shown in Table 1. This viral load is so low that any direct HPV oncogenic influence is likely to be small. It is possible that HPV oncogenic influences in breast cancer are indirect. This is because there is no increase in the prevalence of breast cancer in patients with induced immunosuppression or with immunity suppressed by human immunodeficiency virus (HIV) (41). This is in contrast to the up to sixfold increase in prevalence of HPV associated cervical cancer in similar patients. Transcription from HPV DNA to RNA in HPV-associated breast cancer has not been identified by some investigators (47). However, a very low prevalence of HPV transcripts has been identified in the TCGA series of breast tumors (10). An indication of some HPV viral transcription in some breast cancers is the identification of HPV E7 protein expression (10).

The oncogenic capacity of high-risk HPVs is well established (3). Accordingly, it is plausible that high-risk HPVs could have a role in breast cancer. The transmission of HPVs during sexual activities involving surface to surface contact between epithelial cells is also well documented (48). Not so well known is the identification of high-risk HPVs in white blood cells, which suggests a means of HPV transmission throughout the body (49).

Band et al. have repeatedly demonstrated that HPV DNAs can immortalize normal human mammary epithelial cells in experimental conditions (50, 51). This evidence demonstrates that HPVs have the capacity to have a causal role in breast cancer. However, it is not conclusive evidence of causality because the influence of HPVs and their oncogenic proteins may differ in experimental conditions than in normal women.

Mechanisms of Oncogenesis

The oncogenic mechanisms by which HPV induces cervical cancer have been intensively studied (3). High-risk HPV encodes a series of proteins, designated as early (E1–E7) or late (L1 and L2). Although all of the viral proteins have a role in viral replication, only a small number of the viral early proteins have a role in cellular transformation. Transformation is due to the HPV E6 and E7 oncoproteins, which work in concert to disrupt cell-cycle regulation, inhibit apoptosis, and stimulate cell cycle progression by binding/inhibiting the p53 and p110RB tumor suppressor genes, respectively.

p53 protein expression has been shown to be significantly inhibited in HPV-positive breast cancer as compared to HPVnegative breast cancer (46). This indicates that HPVs in breast cancer may be biologically active.

In addition, HPV E5 and E6 act early in transformation (before integration) and can disrupt cytokeratin causing perinuclear cytoplasmic clearing and nuclear enlargement, which leads to the appearance of a koilocyte (52). Koilocytes are shown in **Figure 1**. HPV associated koilocytes have been identified in breast tumors, which is an indication of early influences of HPVs on normal breast epithelial cells (53). Oncogenic protein HPV E7 expression has been observed in both benign and subsequent breast cancers in the same patients (10). This is an indication of HPV oncogenic activity. HPV type 16 E6 and E7 oncoproteins have been shown to convert non-invasive breast cancer cells into an invasive form both *in vitro* and in nude mice (54).

HPV E7 is more frequently expressed in benign breast biopsy specimens than in subsequent breast cancers that developed several years after the biopsy (40). This is shown in **Figure 1**. This suggests that HPVs may have an early role in breast cancer – the "hit and run" phenomenon. These direct influences are probably limited because, as indicated above, there is no increase in the prevalence of breast cancer in immunocompromised patients (41). If HPVs had a major direct influence in breast cancer, it would be anticipated that the prevalence would be increased in these patients.

Recent evidence suggests that high-risk HPVs may have an indirect oncogenic influence in the development of breast cancer. HPVs can promote the activity or influences of a DNAediting enzyme APOBEC3B, which is a protein encoded by the APOBEC3B gene (38, 39). Upregulated APOBEC3B protein expression was first identified by Burns et al. as an enzymatic source of mutations in breast cancer (55). Upregulation of APOBEC3B can occur in many different cancers (55). It has also been shown that high expression of APOBEC3B can be associated with aggressive breast cancers (56). Finally, HPVs may collaborate with other oncogenic viruses such as EBV to cause breast cancer. The evidence for such a collaboration is very limited and mainly consists as an association (22, 43, 57). There is unpublished experimental evidence, which supports this notion (58).

DISCUSSION AND CONCLUSION

The fourfold higher prevalence of high-risk HPVs in breast cancer as compared to normal and benign breast tissues is a consistent finding among over 25 studies conducted in different laboratories in many contrasting populations. This phenomenon is strongly suggestive of a role of high-risk HPVs in breast cancer. However, the nature of this role is far from clear. HPVs do not appear to have a direct oncogenic influence in breast cancer as demonstrated by the fact that the prevalence of breast cancer is not increased in immunocompromised patients (as compared to the fourfold to sixfold increase in cervical cancer in these patients). HPV E7 oncprotein is highly expressed in benign breast tissues but lowly expressed in breast cancer tissues in the same patients. This is an indication of early HPV influences. The implication is that the influences of HPV in breast cancer are probably early and indirect and without major influences of the immune system.

It can be argued that ubiquitous infections such as with HPVs cannot be associated with cancer. However, this is not a correct argument as shown by the example of EBV. Over 90% of Western populations become infected to EBV at some time in their lives. Rarely, these ubiquitous EBV infections cause serious cancers such as Burkitt's lymphoma, nasopharyngeal cancer, and lymphomas. Interestingly, EBV infections that lead to cancer frequently do so in collaboration with other infectious agents, such as malaria, in the case of Burkitt's lymphoma and HPVs in the case of nasopharyngeal cancers.

REFERENCES

- 1. Hill AB. The environment and disease:association or causation? *Proc R Soc Med* (1965) 58:295–300.
- 2. Vonka V. Causality in medicine: the case of tumours and viruses. *Philos Trans* R Soc Lond B Biol Sci (2000) 355:1831–41. doi:10.1098/rstb.2000.0738
- Zur Hausen H. Papillomaviruses and cancer: from basic studies to clinical application. Nat Rev Cancer (2002) 2:342–50. doi:10.1038/nrc798
- Khan NA, Castillo A, Koriyama C, Kijima Y, Umekita Y, Ohi Y, et al. Human papillomavirus detected in female breast carcinomas in Japan. *Br J Cancer* (2008) 99:408–14. doi:10.1038/sj.bjc.6604502
- Herrera-Romano L, Fernández-Tamayo N, Gómez-Conde E, Reyes-Cardoso JM, Ortiz-Gutierrez F, Ceballos G, et al. Absence of human papillomavirus sequences in epithelial breast cancer in a Mexican female population. *Med Oncol* (2012) 29:1515–7. doi:10.1007/s12032-011-0059-x
- Zhou Y, Li J, Ji Y, Ren M, Pang B, Chu M, et al. Inconclusive role of human papillomavirus infection in breast cancer. *Infect Agent Cancer* (2015) 10:36. doi:10.1186/s13027-015-0029-6
- Bae JM, Kim EH. Human papillomavirus infection and risk of breast cancer: a meta-analysis of case-control studies. *Infect Agent Cancer* (2016) 11:14. doi:10.1186/s13027-016-0058-9
- Yu Y, Morimoto T, Sasa M, Okazaki K, Harada Y, Fujiwara T, et al. HPV33 DNA in premalignant and malignant breast lesions in Chinese and Japanese populations. *Anticancer Res* (1999) 19:5057–61.

With the exception of the unimportant criteria of specificity, all the criteria for a causal role for high-risk HPVs in breast cancer have been met. High-risk HPVs have a well-documented oncogenic capacity (3). HPVs have been consistently identified in breast cancers by different workers in different laboratories in a wide range of countries (7). The odds ratio of HPV positive breast cancers is high as compared to benign breast biopsy and normal breast controls (7). There is evidence of HPV breast infection prior to the development of HPV positive breast cancer (10, 31, 38). HPV-associated cervical pathology can precede same HPV type positive breast cancer, and women with cervical pathology are at greater than expected risk of developing breast cancer (31, 46). There are plausible mechanisms, which indicate that HPVs may have an oncogenic influence on the breast (38).

However, the extremely low HPV viral load in breast cancers and the unchanged prevalence of breast cancer in immunocompromised women indicate a need for cautious conclusions.

Also, because multiple viruses (mouse mammary tumor virus, EBV, and bovine leukemia virus) may have roles in human breast cancer, it will be difficult to identify the influence of a vaccine aimed at a single virus. On the other hand, given the strong possibility that high-risk HPVs do have a role in some breast cancers, it is reasonable to promote the use of HPV vaccines, such as Gardasil, as they are very effective in reducing the risk of cervical cancer.

Based on this evidence, high-risk HPVs may have a causal role in a subset of breast cancers.

AUTHOR CONTRIBUTIONS

JL, WL, and NW - concepts and preparation of manuscript.

- Fu L, Wang D, Shah W, Wang Y, Zhang G, He J. Association of human papillomavirus type 58 with breast cancer in Shaanxi province of China. *J Med Virol* (2015) 87:1034–40. doi:10.1002/jmv.24142
- Lawson JS, Glenn WK, Salyakina D, Delprado W, Clay R, Antonsson A, et al. Human papilloma viruses and breast cancer. *Front Oncol* (2015) 5:277. doi:10.3389/fonc.2015.00277
- Damin AP, Karam R, Zettler CG, Caleffi M, Alexandre CO. Evidence for an association of human papillomavirus and breast carcinomas. *Breast Cancer Res Treat* (2004) 84:131–7. doi:10.1023/B:BREA.0000018411.89667.0d
- Tsai JH, Tsai CH, Cheng MH, Lin SJ, Xu FL, Yang CC. Association of viral factors with non-familial breast cancer in Taiwan by comparison with non-cancerous, fibroadenoma, and thyroid tumor tissues. J Med Virol (2005) 75:276–81. doi:10.1002/jmv.20267
- Choi YL, Cho EY, Kim JH, Nam SJ, Oh YL, Song SY, et al. Detection of human papillomavirus DNA by DNA chip in breast carcinomas of Korean women. *Tumour Biol* (2007) 28:327–32. doi:10.1159/000124238
- Gumus M, Yumuk PF, Salepci T, Aliustaoglu M, Dane F, Ekenel M, et al. HPV DNA frequency and subset analysis in human breast cancer patients' normal and tumoral tissue samples. *J Exp Clin Cancer Res* (2006) 25:515–21.
- He Q, Zhang SQ, Chu YL, Jia XL, Wang XL. The correlations between HPV16 infection and expressions of c-erbB-2 and bcl-2 in breast carcinoma. *Mol Biol Rep* (2009) 36:807–12. doi:10.1007/s11033-008-9249-9
- de León DC, Montiel DP, Nemcova J, Mykyskova I, Turcios E, Villavicencio V, et al. Human papillomavirus (HPV) in breast tumors: prevalence in a

group of Mexican patients. *BMC Cancer* (2009) 9:26. doi:10.1186/1471-2407-9-26

- Heng B, Glenn WK, Ye Y, Tran B, Delprado W, Lutze-Mann L, et al. Human papilloma virus is associated with breast cancer. *Br J Cancer* (2009) 101:1345–50. doi:10.1038/sj.bjc.6605282
- Mou X, Chen L, Liu F, Shen Y, Wang H, Li Y, et al. Low prevalence of human papillomavirus (HPV) in Chinese patients with breast cancer. *J Int Med Res* (2011) 39:1636–44. doi:10.1177/147323001103900506
- Chang P, Wang T, Yao Q, Lv Y, Zhang J, Guo W, et al. Absence of human papillomavirus in patients with breast cancer in north-west China. *Med Oncol* (2012) 29:521–5. doi:10.1007/s12032-011-9945-5
- Sigaroodi A, Nadji SA, Naghshvar F, Nategh R, Emami H, Velayati AA. Human papillomavirus is associated with breast cancer in the north part of Iran. *ScientificWorldJournal* (2012) 2012:837191. doi:10.1100/2012/837191
- Frega A, Lorenzon L, Bononi M, De Cesare A, Ciardi A, Lombardi D, et al. Evaluation of E6 and E7 mRNA expression in HPV DNA positive breast cancer. *Eur J Gynaecol Oncol* (2012) 33:164–7.
- Glenn WK, Heng B, Delprado W, Iacopetta B, Whitaker NJ, Lawson JS. Epstein-Barr virus, human papillomavirus and mouse mammary tumour virus as multiple viruses in breast cancer. *PLoS One* (2012) 7:e48788. doi:10.1371/ journal.pone.0048788
- Liang W, Wang J, Wang C, Lv Y, Gao H, Zhang K, et al. Detection of highrisk human papillomaviruses in fresh breast cancer samples using the hybrid capture 2 assay. J Med Virol (2013) 85:2087–92. doi:10.1002/jmv.23703
- Ahangar-oskouee M, Shahmahmoodi S, Jalilvand S, Ziaee AA, Esmaeli HA. No detection of "high risk" human papillomavirus in a group of Iranian women with breast cancer. *Asian Pac J Cancer Prev* (2014) 15:4061–5. doi:10.7314/ APJCP.2014.15.9.4061
- Ali SH, Al-Alwan NA, Al-Alwany SH. Detection and genotyping of human papillomavirus in breast cancer tissues from Iraqi patients. *East Mediterr Health J* (2014) 20:372–7.
- Eslamifar A, Ramezani A, Azadmanesh K, Bidari-Zerehpoosh F, Banifazl M, Aghakhani A. Assessment of the association between human papillomavirus infection and breast carcinoma. *Iran J Pathol* (2015) 10:41–6.
- 27. Manzouri L, Salehi R, Shariatpanahi S, Rezaie P. Prevalence of human papilloma virus among women with breast cancer since 2005-2009 in Isfahan. *Adv Biomed Res* (2014) 3:75. doi:10.4103/2277-9175.125873
- Peng J, Wang T, Zhu H, Guo J, Li K, Yao Q, et al. Multiplex PCR/mass spectrometry screening of biological carcinogenic agents in human mammary tumors. J Clin Virol (2014) 61:255–9. doi:10.1016/j.jcv.2014.07.010
- Li J, Ding J, Zhai K. Detection of human papilloma virus DNA in patients with breast tumor in China. *PLoS One* (2015) 10:e0136050. doi:10.1371/journal. pone.0136050
- Vernet-Tomas M, Mena M, Alemany L, Bravo I, De Sanjosé S, Nicolau P, et al. Human papillomavirus and breast cancer: no evidence of association in a Spanish set of cases. *Anticancer Res* (2015) 35:851–6.
- Lawson JS, Glenn WK, Salyakina D, Clay R, Delprado W, Cheerala B, et al. Human papilloma virus identification in breast cancer patients with previous cervical neoplasia. *Front Oncol* (2016) 5:298. doi:10.3389/fonc.2015.00298
- Hansen BT, Nygård M, Falk RS, Hofvind S. Breast cancer and ductal carcinoma in situ among women with prior squamous or glandular precancer in the cervix: a register-based study. *Br J Cancer* (2012) 107:1451–3. doi:10.1038/bjc.2012.438
- Søgaard M, Farkas DK, Ording AG, Sørensen HT, Cronin-Fenton D. Conisation as a marker of persistent human papilloma virus infection and risk of breast cancer. *Br J Cancer* (2016) 115:588–91. doi:10.1038/bjc.2016.150
- Tang KW, Alaei-Mahabadi B, Samuelsson T, Lindh M, Larsson E. The landscape of viral expression and host gene fusion and adaptation in human cancer. *Nat Commun* (2013) 4:2513. doi:10.1038/ncomms3513
- Khoury JD, Tannir NM, Williams MD, Chen Y, Yao H, Zhang J, et al. Landscape of DNA virus associations across human malignant cancers: analysis of 3,775 cases using RNA-Seq. J Virol (2013) 87:8916–26. doi:10.1128/JVI.00340-13
- Vinner L, Mourier T, Friis-Nielsen J, Gniadecki R, Dybkaer K, Rosenberg J, et al. Investigation of human cancers for retrovirus by low-stringency target enrichment and high-throughput sequencing. *Sci Rep* (2015) 5:13201. doi:10.1038/srep13201

- World Health Organisation. GLOBOCAN 2012. Lyon, France: World Health Organisation (2015).
- Ohba K, Ichiyama K, Yajima M, Gemma N, Nikaido M, Wu Q, et al. In vivo and in vitro studies suggest a possible involvement of HPV infection in the early stage of breast carcinogenesis via APOBEC3B induction. *PLoS One* (2014) 9:e97787. doi:10.1371/journal.pone.0097787
- Vieira VC, Leonard B, White EA, Starrett GJ, Temiz NA, Lorenz LD, et al. Human papillomavirus E6 triggers upregulation of the antiviral and cancer genomic DNA deaminase APOBEC3B. *MBio* (2014) 5:e2234–2214. doi:10.1128/mBio.02234-14
- Ngan C, Lawson JS, Clay R, Delprado W, Whitaker NJ, Glenn WK. Early human papilloma virus (HPV) oncogenic influences in breast cancer. *Breast Cancer (Auckl)* (2015) 9:93–7. doi:10.4137/BCBCR.S35692
- Grulich AE, Vajdic CM. The epidemiology of cancers in human immunodeficiency virus infection and after organ transplantation. *Semin Oncol* (2015) 42:247–57. doi:10.1053/j.seminoncol.2014.12.029
- Mirzamani N, Salehian P, Farhadi M, Tehran EA. Detection of EBV and HPV in nasopharyngeal carcinoma by in situ hybridization. *Exp Mol Pathol* (2006) 81:231–4. doi:10.1016/j.yexmp.2006.04.006
- Al Moustafa AE, Al-Antary N, Aboulkassim T, Akil N, Batist G, Yasmeen A. Co-prevalence of Epstein-Barr virus and high-risk human papillomaviruses in Syrian women with breast cancer. *Hum Vaccin Immunother* (2016) 12(7):1936–9. doi:10.1080/21645515.2016.1139255
- Badoual C, Tartour E, Roussel H, Bats AS, Pavie J, Pernot S, et al. HPV (human papilloma virus) implication in other cancers than gynaecological. *Rev Med Interne* (2015) 36:540–7. doi:10.1016/j.revmed.2015.01.003
- Lawson JS, Glenn WK, Tran DD, Ngan CC, Duflou JA, Whitaker NJ. Identification of human papilloma viruses in atheromatous coronary artery disease. Front Cardiovasc Med (2015) 2:17. doi:10.3389/fcvm.2015.00017
- Hennig EM, Kvinnsland S, Holm R, Nesland JM. Significant difference in p53 and p21 protein immunoreactivity in HPV 16 positive and HPV negative breast carcinomas. *Acta Oncol* (1999) 38:931–8. doi:10.1080/028418699432617
- Gannon OM, Antonsson A, Milevskiy M, Brown MA, Saunders NA, Bennett IC. No association between HPV positive breast cancer and expression of human papilloma viral transcripts. *Sci Rep* (2015) 5:18081. doi:10.1038/ srep18081
- Koutsky LA. Epidemiology of genital human papillomavirus infection. Am J Med (1997) 102:3–8. doi:10.1016/S0002-9343(97)00177-0
- Foresta C, Bertoldo A, Garolla A, Pizzol D, Mason S, Lenzi A, et al. Human papillomavirus proteins are found in peripheral blood and semen Cd20+ and Cd56+ cells during Hpv-16 semen infection. *BMC Infect Dis* (2013) 13:593. doi:10.1186/1471-2334-13-593
- Band V, Zajchowski D, Kulesa V, Sager R. Human papilloma virus DNAs immortalize normal human mammary epithelial cells and reduce their growth factor requirements. *Proc Natl Acad Sci U S A* (1990) 87:463–7. doi:10.1073/ pnas.87.1.463
- Wazer DE, Liu XL, Chu Q, Gao Q, Band V. Immortalization of distinct human mammary epithelial cell types by human papilloma virus 16 E6 or E7. Proc Natl Acad Sci U S A (1995) 92:3687. doi:10.1073/pnas.92.9.3687
- Krawczyk E, Suprynowicz FA, Liu X, Dai Y, Hartmann DP, Hanover J, et al. Koilocytosis: a cooperative interaction between the human papillomavirus E5 and E6 oncoproteins. *Am J Pathol* (2008) 173:682–8. doi:10.2353/ ajpath.2008.080280
- Lawson JS, Glenn WK, Heng B, Ye Y, Tran B, Lutze-Mann L, et al. Koilocytes indicate a role for human papilloma virus in breast cancer. *Br J Cancer* (2009) 101:1351–6. doi:10.1038/sj.bjc.6605328
- Yasmeen A, Bismar TA, Kandouz M, Foulkes WD, Desprez PY, Al Moustafa AE. E6/E7 of HPV type 16 promotes cell invasion and metastasis of human breast cancer cells. *Cell Cycle* (2007) 6:2038–42. doi:10.4161/cc.6.16.4555
- Burns MB, Lackey L, Carpenter MA, Rathore A, Land AM, Leonard B, et al. APOBEC3B is an enzymatic source of mutation in breast cancer. *Nature* (2013) 494:366–70. doi:10.1038/nature11881
- 56. Tsuboi M, Yamane A, Horiguchi J, Yokobori T, Kawabata-Iwakawa R, Yoshiyama S, et al. APOBEC3B high expression status is associated with aggressive phenotype in Japanese breast cancers. *Breast Cancer* (2016) 23:780–8. doi:10.1007/s12282-015-0641-8

- Aguayo F, Khan N, Koriyama C, González C, Ampuero S, Padilla O, et al. Human papillomavirus and Epstein-Barr virus infections in breast cancer from Chile. *Infect Agent Cancer* (2011) 6:7. doi:10.1186/1750-9378-6-7
- Cameron J. HPV and EBV collaboration. International Centre for Genetic Engineering and Biotechnology. DNA Tumour Virus Meeting Abstracts. Trieste (2011).

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