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**Background:** Limited evidence exists on the impact of adding a taxane, using endocrine therapy and carrying a deleterious germline *BRCA* mutation on ovarian reserve measured by anti-müllerian hormone (AMH) levels of young breast cancer patients receiving (neo)adjuvant cyclophosphamide- and anthracycline-based chemotherapy.

**Methods:** This is a biomarker analysis including young ( $\leq$ 40 years) early breast cancer patients with known germline *BRCA* mutational status and available prospectively collected frozen plasma samples before and after chemotherapy. Chemotherapy consisted of either six cycles of FEC (5 fluorouracil 500 mg/m<sup>2</sup>, epirubicin 100 mg/m<sup>2</sup>, cyclophosphamide 500 mg/m<sup>2</sup>) or three cycles of FEC followed by three cycles of docetaxel (D, 100 mg/m<sup>2</sup>). Endocrine therapy consisted of tamoxifen ( $\pm$ GnRH agonists). AMH levels at baseline, 1 and 3 years after diagnosis were compared according to type of chemotherapy (FEC only vs. FEC-D), use of endocrine therapy (yes vs. no) and deleterious germline *BRCA* mutations (mutated vs. negative).

**Results:** Out of 148 included patients, 127 (86%) received D following FEC chemotherapy, 90 (61%) underwent endocrine therapy, and 35 (24%) had deleterious germline *BRCA* mutations. In the whole cohort, AMH levels drastically dropped 1 year after diagnosis (p < 0.0001) with a slight but significant recovery at 3 years (p < 0.0001).

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One year after diagnosis, patients treated with FEC only had higher median AMH levels than those who received FEC-D (0.22 vs. 0.04  $\mu$ g/L, p = 0.0006); no difference was observed at 3 years (0.06 and 0.18  $\mu$ g/L, p = 0.47). Patients under endocrine therapy had significantly higher AMH levels than those who did not receive this treatment 1 year after diagnosis (0.12 vs. 0.02  $\mu$ g/L; p = 0.008), with no difference at 3 years (0.11 and 0.20  $\mu$ g/L, p = 0.22). AMH levels were similar between *BRCA*-mutated and *BRCA*-negative patients at baseline (1.94 vs. 1.66  $\mu$ g/L, p = 0.53), 1 year (0.09 vs. 0.06  $\mu$ g/L, p = 0.39) and 3 years (0.25 vs. 0.16  $\mu$ g/L; p = 0.43) after diagnosis.

**Conclusions:** In breast cancer patients receiving FEC chemotherapy, adding D appeared to negatively impact on their ovarian reserve in the short-term; no further detrimental effect was observed for endocrine therapy use and presence of a deleterious germline *BRCA* mutation.

Keywords: breast cancer, AMH, taxane, endocrine therapy, BRCA mutations

## INTRODUCTION

As a consequence of the higher incidence of aggressive tumor subtypes and the negative prognostic value of young age at diagnosis (1, 2), a significant proportion of young women with early breast cancer are candidates to receive adjuvant or neoadjuvant chemotherapy. In young patients, a major potential drawback associated with the use of systemic cytotoxic therapy is represented by the risk of causing gonadal damage with subsequent premature ovarian insufficiency (POI) and infertility (3). Considering the substantial quality of life implications associated with the development of these side effects, appropriate oncofertility counseling is now considered mandatory with all cancer patients diagnosed during their reproductive years (4–6).

Age and use of cyclophosphamide-based chemotherapy are the two major known determinants influencing the risk of gonadal damage following the use of anticancer treatments in young women with breast cancer (3). On the contrary, the impact of other treatment- or patient-related factors remains controversial. Specifically, while the effect of cyclophosphamide- and anthracycline-based chemotherapy regimens is well-established, it remains unclear if the addition of a taxane can further increase the risk of gonadal damage (7). Similarly, in young patients with hormone receptor-positive disease, the gonadotoxic impact of using endocrine therapy following chemotherapy remains debated (7, 8). Finally, recent evidence suggests that carrying a deleterious germline BRCA mutation may have a negative impact on women ovarian reserve raising the important concern of a potential increased gonadotoxicity with the use of anticancer treatments in this patient population (9, 10).

Informing young women with newly diagnosed breast cancer about the actual gonadal damage associated with the use of the proposed anticancer treatments is even more complex considering that the majority of the studies that addressed this issue relied only on the presence or absence of menstrual function after the end of chemotherapy (3). However, this cannot be considered a surrogate to determine

treatment-induced gonadotoxicity (11). Being the most accurate indicator of remaining ovarian reserve, anti-müllerian hormone (AMH) is considered a promising biomarker of treatmentinduced gonadotoxicity (12). Nevertheless, to date, there is limited evidence on the actual gonadal damage associated with the use of the proposed anticancer treatments through AMH assessment in young breast cancer patients (12). This is crucial information to be acquired to better inform these patients about the adverse events associated with the proposed therapies as well as on the need of pursuing fertility preservation strategies before treatment initiation. To acquire more insights on this unmet medical issue, we conducted the present study aiming to evaluate the impact of adding a taxane, using endocrine therapy as well as carrying a deleterious germline BRCA mutation on the ovarian reserve measured by AMH levels at baseline and up to 3 years after diagnosis in young early breast cancer patients treated with (neo)adjuvant anthracycline- and cyclophosphamidebased chemotherapy.

## MATERIALS AND METHODS

## **Patients and Treatments**

This is a biomarker analysis conducted within a cohort of consecutive patients with early breast cancer diagnosed at  $\leq$ 40 years that underwent (neo)adjuvant chemotherapy between January 2008 and December 2016 at the Henri Becquerel Cancer Center (Rouen, France). Patients with known germline *BRCA* mutational status and prospectively collected and available frozen plasma samples before and after chemotherapy were eligible for inclusion in the present analysis.

Chemotherapy consisted of either six cycles of FEC (five fluorouracil 500 mg/m<sup>2</sup>, epirubicin 100 mg/m<sup>2</sup>, cyclophosphamide 500 mg/m<sup>2</sup>) or three cycles of FEC followed by three cycles of docetaxel (D, 100 mg/m<sup>2</sup>). Adjuvant endocrine therapy consisted of tamoxifen exclusively or associated with gonadotropin-releasing hormone (GnRH) agonists. None of



the patients received GnRH agonists during chemotherapy for ovarian function and/or fertility preservation.

All patients signed a consent form allowing the conservation and study of their biological samples. The present study was approved by the Institutional Scientific and Ethics Committees of Henri Becquerel Centre (registering order  $N^{\circ}$  1807B).

### **AMH Measurements**

As per routine practice at our center, plasma samples are prospectively collected at fixed timepoints during follow-up in all breast cancer patients and are stored in our plasma bank at  $-20^{\circ}$ C.

For the purpose of the present analysis, plasma samples of eligible patients were used to assess AMH levels at baseline (i.e., before starting chemotherapy),  $\sim$ 1 year and over 3 years after diagnosis.

AMH measurements were centrally performed at Henri Becquerel Cancer Center using fully automated ultra-sensitive Elecsys AMH assay on the Cobas e601 instrument (Roche Diagnostics). The detection and quantification limits were 0.01 and 0.03  $\mu$ g/L, respectively, with an intra-assay imprecision coefficient of variation equal to 1.2% at 1.19  $\mu$ g/L and 0.9% at 5.89  $\mu$ g/L.

## **Study Objectives**

The main objective of this study was to assess the impact of adding a taxane (D), using endocrine therapy and carrying a deleterious germline *BRCA* mutation on the ovarian reserve measured by AMH levels of young breast cancer patients after (neo)adjuvant cyclophosphamide- and anthracycline-based chemotherapy (FEC). AMH levels at baseline, 1 and 3 years after diagnosis were compared according to type of chemotherapy (FEC only vs. FEC-D), use of endocrine therapy (yes vs. no) and deleterious germline *BRCA* mutations (mutated vs. negative).

# **Statistical Analysis**

Baseline characteristics of patients were compared according to type of chemotherapy or endocrine treatment administered and *BRCA* mutational status. Quantitative variables were reported as median with interquartile range (IQR, Q1–Q3). Differences were tested using  $\chi^2$ , Fisher's exact test or two-sample *t*-tests as appropriate.

Comparisons of the evolution of AMH values over time were restricted to the same patients, using paired tests (Wilcoxon).

A *p*-value of <0.05 was considered statistically significant. Tests were performed using the R<sup>®</sup> statistical software version 3.4.3. Figures were made using the ggplot2 package (R Core Team 2017, https://www.R-project.org/).

# RESULTS

## **Patient Characteristics**

Between January 2008 and December 2016, out of 262 patients diagnosed at  $\leq$ 40 years who underwent (neo)adjuvant chemotherapy for early breast cancer, 148 had plasma samples and *BRCA* mutational status available to be included in the present analysis (**Figure 1**).

Median age was 35 years (IQR 31.5–38; **Table 1**). A total of 127 (86%) patients received D in addition to FEC chemotherapy, 90 (61%) underwent endocrine therapy, and 35 (24%) had deleterious germline *BRCA* mutations.

# AMH Evolution Under Chemotherapy in the Whole Population

At baseline, median AMH level was 1.68  $\mu$ g/L (IQR 1.00–3.30). Higher age was associated with significant lower AMH levels (p = 0.047; Figure S1).

One year after diagnosis (median 387 days, IQR 363–426), AMH levels drastically dropped to a median value of 0.06  $\mu$ g/L (IQR 0–0.25; p < 0.0001). Three years after diagnosis (median 1,132 days, IQR 1,099–1,186), a slight but significant recovery of

#### TABLE 1 | Baseline patients' and tumor characteristics.

Patient characteristics	All patients ( $n = 148$ )
Age at diagnosis, median [IQR]	35.5 [31.5–38]
AMH, median [IQR]	1.68 [1.00–3.30]
AMH, mean [SD]	2.55 [2.81]
Body mass index (kg/m²), median [IQR]	22.7 [21.0-26.2]
Smoker, n (%)	52 (35%)
Genetic mutation, n (%)	35 (24%)
BRCA1	22 (63%)
BRCA2	13 (37%)
Tumor characteristics, n (%)	
Histological grade	
Grade I	6 (4%)
Grade II	54 (36%)
Grade III	87 (59%)
Not available	1 (<1%)
Hormone receptor positivity	92 (62%)
Oestrogen receptor	90 (61%)
Progesteron receptor	65 (44%)
HER2-positive	27 (18%)
Triple-negative	52 (35%)
Pathological nodal status positivity	77 (52%)
Tumor size (T)	
T1	49 (33%)
Τ2	72 (49%)
ТЗ	23 (15%)
Τ4	4 (3%)
Surgical treatment, n (%)	
Conservative	82 (55%)
Mastectomy	66 (45%)
Adjuvant treatment, <i>n</i> (%)	
Radiation therapy	144 (97%)
Endocrine therapy <sup>a</sup>	90 (61%)
Chemotherapy	148 (100%)
Chemotherapy regimen, n (%)	
3 FEC-3 D	127 (86%)
6 FEC	21 (14%)
Endocrine therapy, n (%)	
Tamoxifen	82 (91%)
Tamoxifen + GnRH agonists	8 (9%)
Fertility history, <i>n</i> (%)	
Pregnancy before treatment	129 (84%)
Childbirth before treatment	128 (84%)

<sup>a</sup>2 patients with hormone receptor-positive tumors refused endocrine therapy.

AMH, anti-mullerian hormone; IQR, interquartile range; SD, standard deviation; FEC, fluorouracil, epirubicin, cyclophosphamide; D, docetaxel; GnRH, gonadotropin-releasing hormone.

AMH value was observed with a median level of 0.17  $\mu$ g/L (IQR 0.04–0.41; *p* < 0.0001; **Figure S2**).

#### Impact of Taxanes

Baseline characteristics according to type of chemotherapy (FEC only vs. FEC-D) are reported in **Table S1**.

At baseline, no difference in AMH levels was observed between patients treated with FEC only (1.66  $\mu$ g/L, IQR 1.06–2.85) or FEC-D (1.69  $\mu$ g/L, IQR 0.98–3.33; p = 0.83) (**Figure 2**).

One year after diagnosis, patients treated with FEC only had higher median AMH levels (0.22  $\mu$ g/L, IQR 0.10–0.36) as compared to those who received FEC-D (0.04  $\mu$ g/L, IQR 0.00–0.21; p = 0.0006), respectively.

Three years after diagnosis, no difference in median AMH levels was observed between patients treated with FEC only (0.06  $\mu$ g/L, IQR 0.04–0.32) or FEC-D (0.18  $\mu$ g/L, IQR 0.04–0.43; p = 0.47). Patients treated with FEC only had no recovery of AMH values between 1 and 3 years (0.22 vs. 0.06  $\mu$ g/L, respectively, p = 0.81) while patients treated by FEC-D had a slight but significant recovery (0.04 vs. 0.18  $\mu$ g/L, p < 0.0001).

#### Impact of Endocrine Therapy

Baseline characteristics according to use of endocrine therapy (yes vs. no) are reported in **Table S2**. Among the 90 (61%) patients who received endocrine therapy, 82 (91%) underwent tamoxifen alone and 8 (9%) tamoxifen combined with GnRH agonists (**Table 1**). At the 3-year timepoint, all but three patients who started endocrine therapy were still under treatment.

At baseline, no difference in AMH values was observed between patients who received endocrine therapy (1.94  $\mu$ g/L, IQR 1.01–3.76) and those who did not (1.50  $\mu$ g/L, IQR 0.96–2.77; p = 0.17) (**Figure 3**).

One year after chemotherapy, patients under endocrine therapy had significantly higher AMH levels (0.12  $\mu$ g/L, IQR 0.02–0.29) than those who did not receive this treatment (0.02  $\mu$ g/L, IQR 0.00–0.12; p = 0.008).

No difference was observed 3 years after diagnosis, with comparable values of AMH for patients undergoing endocrine therapy (0.11  $\mu$ g/L, IQR 0.03–0.43) or not (0.20  $\mu$ g/L, 0.07–0.40; p = 0.22).

# Impact of Carrying a Deleterious Germline BRCA Mutation

Baseline characteristics according to *BRCA* mutational status (mutated vs. negative) are reported in **Table S3**. Among the 35 (24%) *BRCA*-mutated breast cancer patients, 22 and 13 harbored deleterious *BRCA1* or *BRCA2* mutations, respectively. Patients in the *BRCA*-mutated cohort were younger than those without mutation (p = 0.027).

At baseline, no difference in AMH values was observed between *BRCA*-mutated (1.94  $\mu$ g/L, IQR 0.98–3.96) and *BRCA*-negative (1.66  $\mu$ g/L, IQR 1.00–3.02) patients (p = 0.53) (**Figure 4**).

One year after chemotherapy, AMH values drastically dropped in both groups to 0.09  $\mu$ g/L (IQR 0.00–0.30) and 0.06  $\mu$ g/L (IQR 0.00–0.21) in the *BRCA*-mutated and negative cohorts, respectively (p = 0.39).

Recovery at 3 years from diagnosis was observed in a similar extent in both groups with levels of 0.25  $\mu$ g/L (IQR 0.02–0.95) and 0.16  $\mu$ g/L (IQR 0.04–0.39) in the *BRCA*-mutated and negative cohorts, respectively (p = 0.43).

When the analyses were repeated to take into account the different age at diagnosis, similar results were shown with no







difference in AMH levels between the *BRCA*-mutated and the *BRCA*-negative cohorts both at diagnosis and after treatment (**Supplementary Data**). Similarly, within the *BRCA*-mutated cohort, no impact of the type of *BRCA* mutation (*BRCA1* vs. *BRCA2*) was observed (**Figure S3**).

## DISCUSSION

In this study, we assessed the ovarian reserve measured by AMH levels at baseline and up to 3 years after diagnosis in young early breast cancer patients treated with (neo)adjuvant anthracyclineand cyclophosphamide-based chemotherapy. While prior evidence exists on AMH levels following the use of endocrine therapy, this is the first study to our knowledge addressing the impact on patients' ovarian reserve of adding a taxane to anthracycline- and cyclophosphamide-based chemotherapy and the potential influence on treatment gonadotoxictiy of carrying a deleterious germline *BRCA* mutation. In early breast cancer patients receiving FEC chemotherapy, adding D appeared to negatively impact on their ovarian reserve in the short-term; no further detrimental effect was observed for endocrine therapy use and presence of a deleterious germline *BRCA* mutation.

Prior studies investigating treatment impact on patients' ovarian reserve showed that AMH decreases rapidly during the



first cycles of chemotherapy reaching the lowest values at the end of systemic cytotoxic therapy (12-16). However, limited data on AMH values beyond 1 year following chemotherapy have been reported so far. Median AMH value at baseline in our study (1.68 µg/L) was in line with published data among non-cancer patients of similar age (median 2.3  $\mu$ g/L) (17). With an estimated AMH value decline of 5.6%/year among non-cancer patients  $\leq 40$  years (17), the physiological decline in our population would have led to a median AMH value 3 years after diagnosis of 1.4 µg/L, which is far more elevated than the value observed (0.17  $\mu$ g/L). Thus, our findings confirmed a deep and persistent impact of anthracycline- and cyclophosphamide-based chemotherapy on AMH levels up to 3 years after diagnosis. In addition, the specific features of our homogenous patient cohort provided a unique opportunity to investigate the influence on AMH levels of adding a taxane, using endocrine therapy as well as carrying a deleterious germline BRCA mutation.

sequential А treatment with anthracyclineand cyclophosphamide-based chemotherapy followed by a taxane is the current standard (neo)adjuvant chemotherapy regimen in young breast cancer patients (5). Despite being a widely used regimen since many years, the gonadotoxicity of such treatment remains controversial. While a prior meta-analysis showed no statistically significant increased risk of amenorrhea with the addition of a taxane (7), larger studies have recently shown a potential negative effect (18, 19). However, limited evidence exists on the actual and potential damage induced by taxanes on patients' ovarian reserve measured by AMH levels (20). Our findings, for the first time with a clear comparison between FEC only and FEC-D, provide some evidence on the potential increased gonadotoxic burden of administering a taxane following anthracycline- and cyclophosphamide-based chemotherapy. However, partial

AMH recovery between 1 and 3 years after diagnosis was observed for FEC-D while a further decrease was shown for FEC only: hence, there was no difference between the two treatment options at 3 years. Notably, the dose of cyclophosphamide for patients treated with FEC only was approximately twice as compared to the one received by women who underwent FEC-D (6 vs. 3 cycles, respectively). As shown in animal models, taxanes appear to damage specifically the growing follicles with no apparent direct effect on primordial follicles (21). This specific gonadotoxic mechanism may explain our observation on the early (within 1 year) negative impact of taxanes on AMH levels. This is in contrast with cyclophosphamide that causes massive atresia of both primordial and growing follicles, with subsequent longer impact on women ovarian reserve reflected by poorer long-term AMH recovery (22).

All young women with hormone receptor-positive breast cancer are candidates to receive adjuvant endocrine therapy (5). For those who are previously exposed to chemotherapy, it is crucial to counsel them on the potential impact of this additional treatment on their ovarian reserve. Several studies have shown an increased risk of post-treatment amenorrhea when tamoxifen was administered after chemotherapy (7, 18, 19). On the contrary, the limited data on endocrine therapy gonadotoxicity measured by AMH levels did not show any difference between patients who received or not tamoxifen following chemotherapy (8, 13, 16, 23). Our findings confirm the lack of detrimental effect of endocrine therapy on patients' ovarian reserve. Interestingly, patients treated with tamoxifen had significantly higher AMH levels at 1 year as compared to those who received chemotherapy alone. Similarly, higher AMH values in patients treated with tamoxifen following chemotherapy has been also observed in two recent studies (8, 23), with one of them showing a faster AMH recovery between 3 and 6 months after the

end of systemic cytotoxic therapy for women treated with endocrine therapy (23). Notably, when given concurrently with chemotherapy, preclinical studies have suggested a potential protective effect of tamoxifen against anticancer treatment gonadotoxicity, including of cyclophosphamide-based therapy (24). Taken together, although tamoxifen may cause perturbation in menstrual function after chemotherapy, the available evidence including our findings suggests the lack of detrimental effect on patients' ovarian reserve. Nevertheless, a proper oncofertility counseling in these women is particularly important and should take into account also the need to prolong such treatment up to 10 years after diagnosis (5), with their subsequent ovarian aging. Indeed, patients with hormone receptor-positive breast cancer have lower chances of post-treatment pregnancies as compared to women who are not candidates to endocrine therapy (8, 25). An international study is currently ongoing to investigate the safety of a temporary interruption of endocrine therapy after 18-30 months of treatment to allow a pregnancy (26).

More than 10% of breast cancer cases arising in young women are hereditary tumors related to the presence of a deleterious germline BRCA mutation (27, 28). Considering the high mutation rate in this patient population, current guidelines strongly support a genetic testing in all women diagnosed at a young age irrespectively of their family history (5). Preclinical and clinical evidence has suggested a potential negative impact of these mutations on female reproductive potential (29, 30). In breast cancer patients, three small studies investigated AMH levels in BRCA-mutated breast cancer patients but only at the time of diagnosis without data after chemotherapy (9, 10, 31). Therefore, a major unanswered concern in this setting is the potential increased gonadotoxicity risk of BRCA-mutated patients (29, 30). The only study that assessed chemotherapy-induced amenorrhea in this setting did not show any difference between BRCAmutated and BRCA-negative patients (32). Our study is on the same line suggesting for the first time the lack of detrimental effect for carrying a deleterious germline BRCA mutation on chemotherapy-induced gonadal damage. Further studies in breast cancer patients with germline mutations in BRCA or other susceptibility genes are warranted to improve their oncofertility counseling in terms of both estimating the risk of gonadotoxicity with the proposed anticancer treatments as well as the efficacy and safety of fertility preservation procedures in this setting (10, 33).

Some limitations should be considered in the interpretation of our results. This analysis was conducted in a single center and included a relatively small cohort of patients who received FEC only chemotherapy or carried a deleterious germline *BRCA* mutation. A few post-chemotherapy plasma samples were missing and could not be assessed in the whole study cohort. Data on post-treatment pregnancies and menstrual function were not collected so that no correlation with AMH values could be performed. Nevertheless, despite these limitations, our study represents one of the largest AMH analyses and with the longest follow-up conducted so far among breast cancer patients; plasma samples were prospectively collected at predefined timepoints during treatment and oncologic follow-up up to 3 years after diagnosis in a homogenous cohort of women. In addition, our study provides answers on three relevant questions that are of crucial importance in everyday clinical practice to properly counsel these women on the gonadotoxicity of the proposed anticancer treatments. Future larger prospective studies are needed to address the clinical utility of this biomarker before its incorporation in routine clinical practice including its potential role as predictor of treatment-induced POI and infertility in young cancer patients.

# CONCLUSIONS

Our study confirmed the deep and prolonged adverse effect of chemotherapy on the ovarian reserve of young breast cancer patients with only partial recovery 3 years after treatment. We showed that adding D following anthracycline- and cyclophosphamide-based chemotherapy (FEC regimen) appeared to cause an early negative impact on their ovarian reserve. Endocrine therapy with tamoxifen and the presence of a deleterious germline *BRCA* mutation did not appear to further worsen chemotherapy gonadotoxicity. Altogether, although future larger collaborative efforts are needed to validate our results, we provide important data for improving the oncofertility counseling of young breast cancer patients.

# DATA AVAILABILITY

The datasets generated for this study are available on reasonable request to the corresponding author.

# ETHICS STATEMENT

The present study was approved by the local Institutional Review Board (registering order  $N^{\circ}1807B$ ). All patients signed a consent form allowing the conservation and study of their biological samples.

# **AUTHOR CONTRIBUTIONS**

MLa, AP, and FC contributed to the conception and design of the study. CC and AP performed the analysis of all plasma samples. JL performed the statistical analysis. The results were interpreted by MLa, NO, JL, AP, and FC who also drafted the initial manuscript that was then revised critically for important intellectual content and approved by all the authors. All the authors contributed to collection and assembly of data.

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# SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc. 2019.00575/full#supplementary-material

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The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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