



# Advancing Cardiovascular, Neurovascular, and Renal Magnetic Resonance Imaging in Small Rodents Using Cryogenic Radiofrequency Coil Technology

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Research in pathologies of the brain, heart and kidney have gained immensely from the plethora of studies that have helped shape new methods in magnetic resonance (MR) for characterizing preclinical disease models. Methodical probing into preclinical animal models by MR is invaluable since it allows a careful interpretation and extrapolation of data derived from these models to human disease. In this review we will focus on the applications of cryogenic radiofrequency (RF) coils in small animal MR as a means of boosting image quality (e.g., by supporting MR microscopy) and making data acquisition more efficient (e.g., by reducing measuring time); both being important constituents for thorough investigational studies on animal models of disease. This review attempts to make the (bio)medical imaging, molecular medicine, and pharmaceutical communities aware of this productive ferment and its outstanding significance for anatomical and functional MR in small rodents. The goal is to inspire a more intense interdisciplinary collaboration across the fields to further advance and progress non-invasive MR methods that ultimately support thorough (patho)physiological characterization of animal disease models. In this review, current and potential future applications for the RF coil technology in cardiovascular, neurovascular, and renal disease will be discussed.

**Keywords:** magnetic resonance, MRI, cardiovascular imaging, neurovascular imaging; renal imaging, MR technology, radio frequency coils, cryogenic

## INTRODUCTION

For several decades animal models have served a wide span of applications in the life sciences. Transgenic systems have been invaluable for studying molecular signatures and specific cell populations as well as tools for non-invasive reporter gene imaging. Animal models that simulate human pathologies have also been indispensable for uncovering mechanisms behind major diseases as well as the identification of their respective treatments.

A thorough characterization of each animal model remains the crux of the matter. It ensures that the right conclusions are drawn from preclinical studies dealing with questions around pathogenesis and therapy as well as molecular studies that set the groundwork for future therapies and drug design. It is getting increasingly clear that most conditions and diseases, even those with an underlying genetic component, are multi-factorial and complex in nature suggesting that therapy should be equally intricate and versatile. It has been suggested that partial, but multiple, drug actions might be more efficient than a complete drug action at a single target in complex multifactorial disease. This calls for novel drug-design strategies that will depend not only on computational modeling for identifying correct multiple targets but also importantly on more-efficient and high-throughput *in vivo* testing. Further developments in non-invasive *in vivo* imaging in small rodents are necessary to guarantee this, as well as a swift and robust translation into clinical practice. For this to be achieved there is an absolute need for (i) anatomical and function imaging with a superb spatial and temporal resolution, (ii) high reproducibility in results, and (iii) longitudinal studies with sufficient statistical power.

Preclinical Magnetic Resonance Imaging (MRI) is conceptually appealing in the pursuit of basic and translational research as well as for explorations into cardiovascular, neurovascular, and renal disease. MRI has become increasingly important for small animal imaging at multiple levels of pre-clinical research. A growing number of reports manifest the advances for morphological and functional MRI of the heart, large blood vessels, CNS, and kidney. Notwithstanding its success and ubiquity, the relatively low sensitivity of conventional MRI constitutes an impediment for translational research and pre-clinical applications. Constraints common to standard room temperature RF MR detectors include contrast-to-noise-ratio (CNR) and spatial resolution but also acquisition time and signal-to-noise ratio (SNR), which are particular currencies spent for image quality.

In recent years, cryogenic RF coil technology that provides significant improvements in image quality has been made commercially available to small animal researchers. The cryogenic technology substantially increases SNR over standard room temperature RF-coils by considerably reducing thermal noise and signal losses in the RF receiver electronics. This facilitates the acquisition of high spatial resolution images within shorter scan times. The gain in SNR via cryogenic-cooling corresponds to the gain achieved by an equivalent increase in magnetic field strength, but without the extra challenges and costs—which could be prohibitive at extreme ultrahigh magnetic fields.

Recognizing the technical advancement in cryogenic RF coil technology this review attempts to make the (bio)medical imaging, molecular medicine, and pharmaceutical communities aware of this productive ferment and its outstanding significance for anatomical and functional MR in small rodents. The goal is to inspire collaborations across disciplinary boundaries and to attract basic scientists, translational researchers, clinician scientists, and new entrants into the field to advance the capabilities of non-invasive MR imaging through the RF coil

technology. In the sections that follow some of the potential applications for cryogenic RF coil technology are discussed. Neurovascular applications for cryogenic RF coil technology include morphological imaging and functional brain mapping in mice. The benefits of cryogenically-cooled RF coils in supporting MRI microscopy (defined by a spatial resolution  $<100\ \mu\text{m}$ ) *in vivo* are demonstrated; the morphological detail reveals brain pathology in animal models of neuroinflammatory diseases, which opens the opportunity to follow neuroinflammatory processes even during the early stages of disease progression. Examples of MR angiography are presented, especially within the context of neurovascular disease. Early and frontier applications of cryogenic RF coil technology in cardiovascular MRI are surveyed together with the opportunities for high spatial resolution cardiac chamber quantification and parametric mapping; all being facilitated by the traits of cryogenic RF coil technology. Last but not least, the sensitivity gain of cryogenic RF coil technology is put to good use for renal MR microscopy and *in vivo* MRI to support explorations into renal diseases with non-invasive techniques for probing renal perfusion, hypoxia and inflammation. A concluding section ventures a glance beyond the horizon and explores future directions. Of course, MRI of small rodents is an area of vigorous ongoing research, and many potentially valuable developments will receive only brief mention here.

## TECHNICAL CONSIDERATIONS

### State of the Art

To date, cryogenic RF coils have been developed by making use of either copper or high-temperature superconducting (HTS) material. In this review we limit ourselves primarily to copper cryogenic coils (CryoProbe, Bruker Biospin, Erlangen, Germany) since these are available to a broader spectrum of users. Pioneering HTS coils have been developed by specialized research groups and are not commercially available yet. Although complex to operate, these coils can achieve SNR gains of more than 10-fold as demonstrated in small excised samples at high field (Black et al., 1993) or in the living mouse at 1.5 T (Poirier-Quinot et al., 2008).

### Image Quality and Signal-to-noise Ratio

*Image quality* describes the perceived or quantitatively measured degradation of an image in comparison to its “perfect” counterpart. In MRI this translates into how well the reconstructed image represents the characteristics of the RF signals originating from the excited MR nuclei. Image contrasts are the core information in the majority of MRI applications. Contrasts allow the delineation of morphological structures and pathological lesions (spatial contrast) or the detection of signal intensity changes over time in functional MRI (temporal contrast). Detrimental factors for MR image quality include artifacts and noise.

The SNR is a quantitative metric of image quality with regards to noise. From an MR image SNR is commonly calculated by dividing the mean signal intensity of a uniform region of interest covering the target region by the standard deviation of the

noise commonly derived from the background free of signal or artifacts. While high SNRs (>100) allow high confidence in the MR data, low SNRs (<20) mean that the true image characteristics are increasingly masked by noise.

The SNR dependence on MR protocol parameters for a conventional 2D experiment is (McRobbie et al., 2006):

$$\text{SNR} \propto \frac{\text{FOV}_{\text{FE}} \cdot \text{FOV}_{\text{PE}} \cdot \Delta z \cdot F_{\text{sequence}} \cdot \sqrt{NA}}{\sqrt{BW \cdot N_{\text{FE}} \cdot N_{\text{PE}}}} \quad (1)$$

where *FOV* is the field-of-view in frequency encoding (FE) and phase encoding (PE) direction,  $\Delta z$  is the slice thickness,  $F_{\text{sequence}}$  is the appropriate sequence dependent factor [including repetition time *TR* and echo time *TE* in relation to the relaxation times  $T_1$  and  $T_2^{(*)}$ ], *NA* is the number of signal averages, *BW* is the receiver bandwidth across the image and *N* is the acquisition matrix size in FE and PE direction. In practice, temporal signal averaging is the most commonly used approach to balance the competing needs for high spatial resolution, high spatial coverage, temporal resolution, and sufficient signal to noise. Yet, signal averaging comes at the cost of scan time when using standard gradient echo or spin echo imaging techniques, without the implementation of acceleration techniques such as parallel imaging or compressed sensing. The square root relation of SNR with the number of averages very quickly sets a limit at which more averages can no longer be justified with a substantial SNR gain due to the severe scan time penalty governed by the power of 2 of the number of averages.

## Why Cooling the Radiofrequency Hardware Improves Image Quality

SNR of an MR image is closely linked to the quality of the acquired RF signals it is composed of. These RF signal measurements—like all analog electronic measurements—are degraded by noise, which reduces signal quality and hence image quality. RF coil efficiency relies on overcoming thermal noise induced by conductive samples (Hoult and Lauterbur, 1979). The noise in MR signal acquisitions originates predominantly from thermal (Brownian) motion of electrical charge carriers within the passive receiver electronics (e.g., coil, conductors, passive components) and within the sample itself. Particularly susceptible to noise are those parts of the electronics in which the RF signal is very small, i.e., from the RF coil until the preamplifier output. The contribution of a 50 Ohm preamplifier to the noise cannot be so easily described because of the reactive part of its impedance, though it can be evaluated by acquiring and comparing noise images at different temperatures e.g., at 77 and 293 K (Poirier-Quinot et al., 2008; Vaughan and Griffiths, 2012).

The concept of cooling RF coil hardware to reduce thermal noise was originally proposed in the 1970s by D. I. Hoult and R. E. Richards (Hoult and Richards, 1976), although the first actual application of cooled RF coils in room-temperature samples was introduced nearly a decade later (Styles et al., 1984). The temperature dependency of the SNR can be described with (Kovacs et al., 2005; Junge, 2012):

$$\text{SNR} \propto \frac{\omega_0 B_1}{\sqrt{R_S T_S + R_C T_C + (R_S + R_C) T_A}} \quad (2)$$

where  $\omega_0$  is the Larmor angular frequency,  $B_1$  is the effective RF field in the sample volume,  $R_S$  represents the so-called magnetic and electric loss in the sample (sample resistance),  $R_C$  is the resistive loss in the RF coil and  $T_{S/C/A}$  is the temperature of the sample, coil, and the noise temperature of the preamplifier, respectively. Cooling reduces the coil noise  $R_C T_C$  and preamplifier noise  $(R_S + R_C) T_A$  and directly increases SNR. An additional benefit of cooling the receiver electronics is owed to the decrease of electrical resistances in the conductor ( $R_C$ ) with decreasing temperatures. Reducing losses within the RF receive chain enhances the SNR.

However, SNR is also dependent on other factors such as frequency  $\omega_0$ , RF coil geometry/quality (creating the effective  $B_1$ ) and sample noise  $R_S T_S$ . Comparing the impacts of coil noise and sample noise on SNR in the context of RF coil size helps to answer the question whether—and under which circumstances—cooling the RF coil is worthwhile. As shown by Junge (2012), the coil and sample noise contributions to SNR at room temperature are of comparable magnitude for a single loop coil with a diameter of approximately 3.0 cm (at 1.5 T) or 1.4 cm (at 9.4 T). For larger RF coils sizes, as commonly used in human MRI, sample noise contributions dominate SNR. For small animal MRI the volumes of interest and RF coil sizes are typically in the range of 1–2 cm, rendering coil noise contributions comparable or larger than sample noise contributions. Although cooling the RF hardware will always reduce noise, the engineering and cost of cryogenic RF hardware starts to pay off only when the thermal noise originating from the receiver electronics is comparable or larger than the noise originating from the sample (patient, animal, object under investigation). It is for that reason that cryogenic RF coils have been developed primarily for nuclear magnetic resonance spectroscopy (NMR; see review Kovacs et al., 2005) using small samples of few millimeter diameter or few cubic-millimeter volume and more recently for small animal MRI.

The theoretical upper limit for the SNR improvement at a magnetic field strength of 9.4 T ( $f = 400$  MHz) is an estimated factor of 2.8 for a transmission line resonator consisting of two planar split rings (with an inner diameter of 14 mm) that are placed on either side of a dielectric substrate, under the assumption of cooling the RF coil from room temperature down to 0 K (Junge, 2012). Cooling the preamplifier has a similar effect on SNR for small samples (Kovacs et al., 2005), but taking a typical preamplifier noise temperature of 15 K with a dominating sample noise ( $R_S \gg R_C$ ) at 310 K, the SNR equation shows that the noise contribution from the preamplifier is not that critical. In practice however the SNR gain achievable by a cryogenic RF coil is lower than the theoretical estimate. The RF hardware cannot be cooled down to 0 K. RF coil and preamplifier are typically operated at temperatures around  $-253^\circ\text{C}$  (20 K) and  $-193^\circ\text{C}$  (80 K), respectively. Notwithstanding these limitations, an SNR gain as high as 2.5 was reported for *in vivo* mouse brain MRI when comparing cryogenic RF hardware (400 MHz CryoProbe, Bruker Biospin MRI GmbH, Ettlingen, Germany) with a conventional room temperature RF coil setup of similar geometry (Baltes et al., 2009).

According to Hoult and Lauterbur (1979), sample resistance  $R_S$  increases quadratically with the Larmor frequency  $\omega_0$  for

a well-designed RF coil. Thus, it may counterbalance the upper frequency term in the above Equation (2) toward higher frequencies. Higher SNR gains are expected at frequencies lower than 400 MHz and have been indeed reported at 200 MHz using the same cooled copper coil technology (Ratering et al., 2008). The gain becomes even higher when using the HTS coil technology at lower frequencies (Darrasse and Ginefri, 2003): SNR gains larger than 10-fold were achieved at 64 MHz in the living mouse brain (Poirier-Quinot et al., 2008). Thus, the cryogenic technology will be particularly valuable for preclinical studies when used in combination with clinical field strengths.

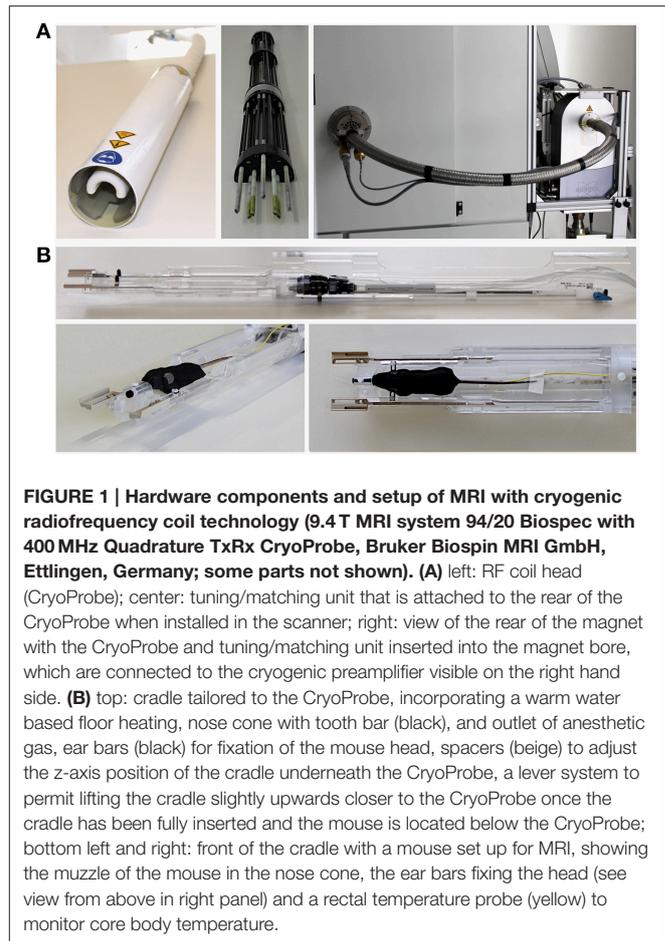
## Cryogenic Radiofrequency Coil Hardware

The first commercial cryogenically cooled RF probe for NMR spectroscopy was installed in 1999 (Kovacs et al., 2005). It took almost 10 years for this technology to mature and to become commercially available for small animal MR scanners. The feasibility and benefit of cryogenically-cooled MRI RF probes was demonstrated for *in vivo* mouse brain imaging at 9.4 T (Baltes et al., 2009). This success built upon the pioneering explorations into cryogenic RF coil designs for biomedical MRI by researchers such as Hurlston et al. (1999), Ginefri et al. (2007), Nouls et al. (2008) and the successful development of a prototype for 4.7 T (Ratering et al., 2008).

Modern cryogenic RF hardware employs high temperature superconducting (HTS) materials to dramatically reduce resistive losses (Black et al., 1993; Hurlston et al., 1999; Darrasse and Ginefri, 2003; Ginefri et al., 2007; Nouls et al., 2008; Junge, 2012; not commercially available yet). Cold helium gas has replaced liquid cryogenics as cooling media. Simply speaking, Helium gas is compressed in one chamber and then chilled through expansion in another chamber in a closed-loop cooling system—comprised of the RF coil, a cryogenic cooling unit and a helium compressor. The waste heat produced during the gas compression is transferred from the helium compressor to a cold water system via a connected water chiller.

For a review of cryogenically cooled RF coil hardware, we will focus here on the range of commercially available cryogenic RF surface coils for preclinical MRI (CryoProbe™, Bruker Biospin MRI GmbH, Ettlingen, Germany). The RF coil head (CryoProbe; **Figure 1A** left) is installed at the center of the magnet bore. In this type, a set of torque rods (**Figure 1A** center) are fitted to the RF probe to allow its tuning and matching from the rear end of the MR system. The preamplifier and the interconnection between preamplifier and CryoProbe are also cryogenically cooled (**Figure 1A** right). Positioning of the mouse underneath the CryoProbe is facilitated by a dedicated animal cradle (**Figure 1B** top), which features a warm water heated floor, nose cone with tooth bar and anesthetic gas outlet, ear bars for head fixation, and spacers to adjust the z-axis position of the cradle, as well as a lever system to permit lifting the cradle closer to the CryoProbe once both are fully inserted and the mouse is located below the RF coil (**Figure 1B** bottom).

Since the introduction of the first cryogenic MRI RF coil the range of CryoProbes is steadily expanding. The initial mouse brain quadrature transmit/receive MRI CryoProbes (for



4.7–15.2 T MR systems) are now-a-days complemented by four-channel array receive-only CryoProbes (for 7.0 and 9.4 T MR systems), four-channel array CryoProbes for rats, and mouse X-nuclei CryoProbes (for 9.4 T; e.g., <sup>13</sup>C, Sack et al., 2014). The latter are used in conjunction with a built-in room temperature <sup>1</sup>H RF coil for decoupling and acquisition of anatomical reference images. Finally, the receive-only CryoProbes allow for the use of a room temperature volume resonator for RF excitation, which improves transmission field (B<sub>1</sub><sup>+</sup>) homogeneity.

## Characteristics of Cryogenic RF Surface Coils—The Pros and Cons

Cryogenic cooling of the RF coil and preamplifier more than doubles the SNR compared with an equivalent RT coil setup. *In vivo* mouse studies revealed typical SNR gains of 2.5–2.8 in the brain (**Figure 2**; Baltes et al., 2009, 2011; Junge, 2012), 3.0–5.0 in the heart (Wagenhaus et al., 2012), and 3.0–3.5 for <sup>13</sup>C spectroscopy in the brain (Sack et al., 2014). These examples of sensitivity enhancements obtained for 400 MHz (9.4 T) may only serve as a basic reference because the SNR gain depends on the frequency (see equation for SNR and Ratering et al., 2008) and the choice of RT coil against which the performance of the CryoProbe is benchmarked.

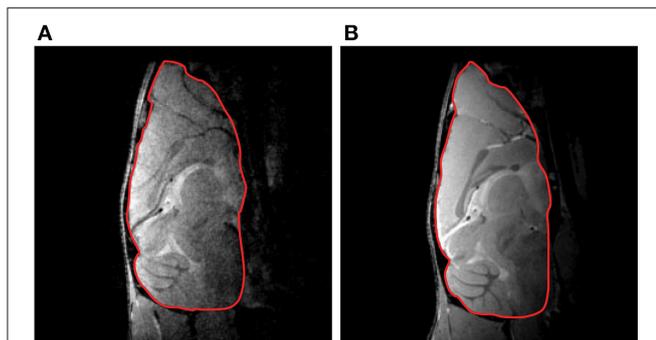
In practical terms, a 100% SNR gain by use of a CryoProbe is substantial enough to facilitate an improvement in spatial resolution from e.g.,  $(100 \times 100) \mu\text{m}^2$  to  $(71 \times 71) \mu\text{m}^2$  by reducing the FOV. Maintaining the same FOV and acquisition time the spatial resolution can be improved to  $(63 \times 63) \mu\text{m}^2$ , as a result of increasing the matrix size (by 58%) combined with a reduction in the number of averages (by 36%). Alternatively an acquisition time reduction by 75% could be attained with this SNR gain via reducing the number of averages.

An SNR gain of 100% is equivalent to significantly increasing the magnet field strength, which is not only more cost intensive, but also comes along with challenges such as increased susceptibility artifacts, disadvantageous relaxation time changes, extra constraints for RF coil design at higher frequencies due to wave length shortening and adverse effects for physiological signals due to the interference with the magnetic field.

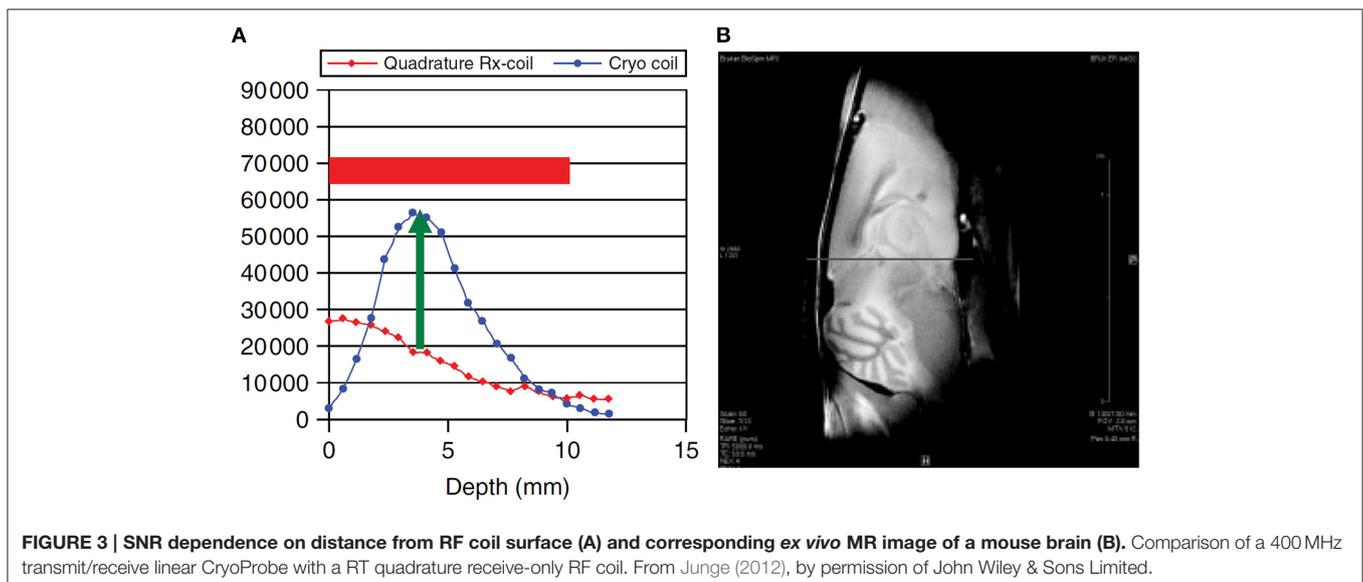
A surface coil design CryoProbe is essential to position the coil as close as possible to the object under investigation, to conform

the coil geometry to the target anatomy, to keep coil size small and remain within the coil noise dominated regime. Although beneficial for signal sensitivity, an inherent limitation of surface coils is an inhomogeneous distribution in both the transmit RF field ( $B_1^+$ ) and receive sensitivity profile ( $B_1^-$ ). **Figure 3** demonstrates the depth dependence of SNR for a spin-echo imaging protocol. SNR is greatly enhanced by the CryoProbe, but only within a certain range of depth, here approximately 2–8 mm from the surface of the coil. The receive sensitivity profile of the surface RF coil reduces the ability to detect RF signals from locations very close to as well as far away from the RF coil. These variations might be a result of a transmit RF field inhomogeneity of the coil which then translates into a variation in flip angle across the field of view, thereby impacting on the signal intensity and  $T_1$  image contrast. This applies to flip angles of excitation pulses as well as any other RF pulses, such as refocussing RF pulses with the exception of adiabatic pulses. Over a typical field of view of 6–8 mm (perpendicular to RF coil surface) the relative  $B_1$  that is proportional to the flip angle can vary by up to a factor of 2 (**Figure 4**; Baltes et al., 2009; Wagenhaus et al., 2012). In gradient-echo images this variability may often go unnoticed. However, in spin-echo images, large deviations from the  $90^\circ$  excitation pulses and the  $180^\circ$  refocusing pulses will eventually lead to inevitable signal losses. Transmit/receive surface coils like most of the CryoProbes require RF power adjustment on a coronal slice (rather than the standard axial slice) which must be carefully positioned to achieve sufficient RF power at larger depths while avoiding signal loss close to the RF coil in spin-echo acquisitions due to too much RF power.  $B_1^+$  inhomogeneity can be largely avoided with the recently available receive-only CryoProbes, which make use of an additional RT volume resonator for RF transmission.

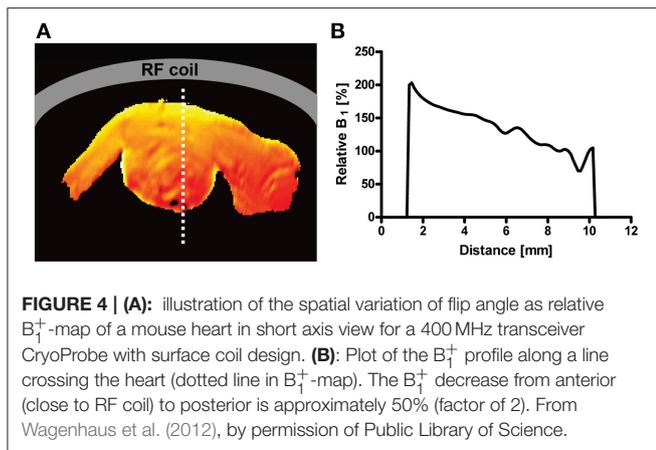
In conclusion, various factors play a role in determining the actual SNR gain, including magnet field strength, MR nucleus, distance to the RF coil, RF power adjustment as well as the MRI acquisition method. The typical improvement in SNR by



**FIGURE 2 |** Mouse brain gradient-echo images acquired at 9.4T with a RT receive-only RF coil (A) and a transmit/receive quadrature CryoProbe (B). The overall SNR gain for the delineated brain region of interest was 2.8. From Junge (2012), by permission of John Wiley & Sons Limited.



**FIGURE 3 |** SNR dependence on distance from RF coil surface (A) and corresponding *ex vivo* MR image of a mouse brain (B). Comparison of a 400 MHz transmit/receive linear CryoProbe with a RT quadrature receive-only RF coil. From Junge (2012), by permission of John Wiley & Sons Limited.



commercially-available CryoProbes can be expected to be a factor of 2–3, which can be translated into more than 60% higher spatial in-plane resolution or 75% shorter acquisition time.

## NEUROVASCULAR APPLICATIONS

### Animal Models of Multiple Sclerosis

The two key benefits of an SNR gain when using cryogenically-cooled coils—namely reduced scan time and/or increased image detail—are fundamental for studying dynamic pathological processes in animal models of disease. In several pathological conditions, especially those related to inflammation and vascular remodeling, an important challenge is to differentiate between hemodynamic alterations, inflammation, and degenerative processes during different stages of disease. Differentiating between pathological processes is essential in chronic neuroinflammatory conditions such as multiple sclerosis (MS) that include an inflammatory, demyelinating and neurodegenerative component. A differentiation between these processes is necessary to make the right therapeutic decision and follow the correct line of treatment for each individual patient (Sinnecker et al., 2012a,b, 2013; Wuerfel et al., 2012; Kuchling et al., 2014).

The histopathological hallmark of MS is the demyelinated plaque, which is associated with perivascular and parenchymal inflammatory cell infiltration and axonal injury (Kuhlmann et al., 2008). MS plaques can occur throughout the CNS; in periventricular and deep white matter, optic nerves and tracts, cerebellar peduncles, brainstem, spinal cord, and also in the gray matter (Sinnecker et al., 2012b). MRI is the most sensitive test to detect and demonstrate MS lesions (Milo and Miller, 2014): while active inflammation is associated with newly appearing, hyperintense MS lesions on  $T_2$ -weighted MRI and enhancement on  $T_1$ -weighted MRI after contrast agent (Barkhof et al., 1997; Brück et al., 1997), neurodegeneration is associated with hypointensities on  $T_1$ -weighted images and indicates severe tissue damage (Van Waesberghe et al., 1999).

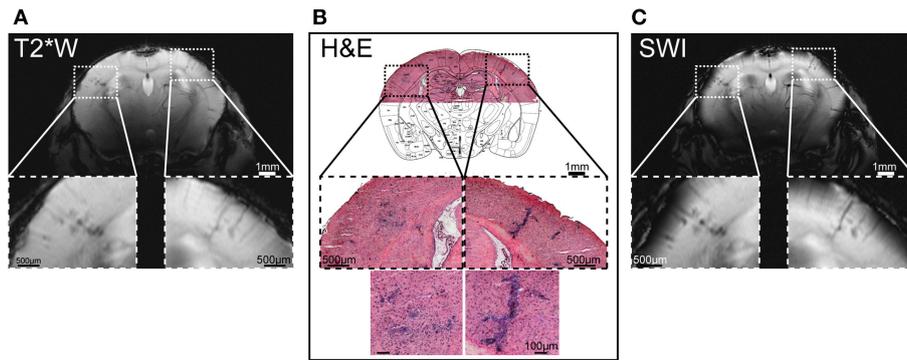
An animal model that resembles MS pathology is the experimental autoimmune encephalomyelitis (EAE). EAE can be induced by immunizing different susceptible species of animals

with specific CNS antigen e.g., proteolipid protein (PLP) in SJL/J mice. The EAE model has been indispensable for understanding neuroinflammatory disease (Ransohoff, 2009) and for evaluating the effectiveness of nascent therapeutic approaches for MS (Ben Nun et al., 1981; Steinman and Zamvil, 2006; Ransohoff, 2012). Similarly to MS, lesions that are hyperintense on  $T_2$ -weighted MRI are present in EAE brains and correspond to inflammation, demyelination and neurodegeneration (Deboy et al., 2007). Also similar to MS, EAE lesions are distributed in time and space (Baxter, 2007), commonly in the brain stem, midbrain, cerebellum and periventricular area (especially in non-human primates) but also in spinal cord, corpus callosum and cerebral gray matter in small rodents (Verhoye et al., 1996; Hart et al., 1998; Boretius et al., 2006; Deboy et al., 2007; Wuerfel et al., 2007; Waiczies et al., 2012).

Brain MR microscopy is an invaluable tool to visualize succinct inflammatory patterns, even prior to neurological disease in the EAE. The main strengths of cryogenically-cooled coils to boost SNR and thereby provide high resolution MR microscopy and/or reduced scan time are best appreciated in animal models such as EAE that undergo highly dynamic disease profiles. Apart from the established hyperintense lesions on  $T_2$ -weighted images (commonly in the cerebellum), focal hypointense lesions were identified in the somatosensory cortex of EAE mice, when using high spatial resolution  $T_2^*$  and  $T_2$  MRI in association with the CryoProbe (Waiczies et al., 2012). With the cryogenic system, an in plane resolution is as good as  $(35 \times 35) \mu\text{m}^2$  and complete coverage of the brain could be achieved on both  $T_2^*$  and  $T_2$ -weighted imaging. With this capacity clear punctate lesions in cerebral gray matter could be revealed in association with intracortical vessels and distributing into the corpus callosum (Waiczies et al., 2012). Thanks to the spatial resolution facilitated by Cryoprobe technology the lesions that were revealed *in vivo* by MR histology could be clearly corroborated as inflammatory infiltrates with hematoxylin and eosin histology (Figure 5).

To gain a comprehensive and longitudinal view of brain inflammation, particularly during the early stages of EAE, MR methods that increase SNR are advantageous not only to increase detail via high resolved brain microscopic imaging but also to reduce scan time. Highly resolved MR images of the whole brain could be achieved in 11 min with the CryoProbe (Figure 5; Waiczies et al., 2012). By reducing scan time, the number of measurements per individual animal can be increased during progression of disease. Repeated MR measurements are fundamental when making assessments regarding structural changes relevant to the pathology; even macroscopic changes (that do not require highly resolved MRI to be revealed) can be easily revealed. As a result of the reduced scan time and repeated measurements achievable by the CryoProbe, ventriculomegaly prior to neurological symptoms was revealed in the EAE (Lepore et al., 2013).

In the past few years animal models that simulate the neurodegenerative and demyelinating components of MS have been studied independently of the inflammatory component (Ransohoff, 2012). For instance transgenic models involving suicide genes that induce oligodendroglialopathy are powerful



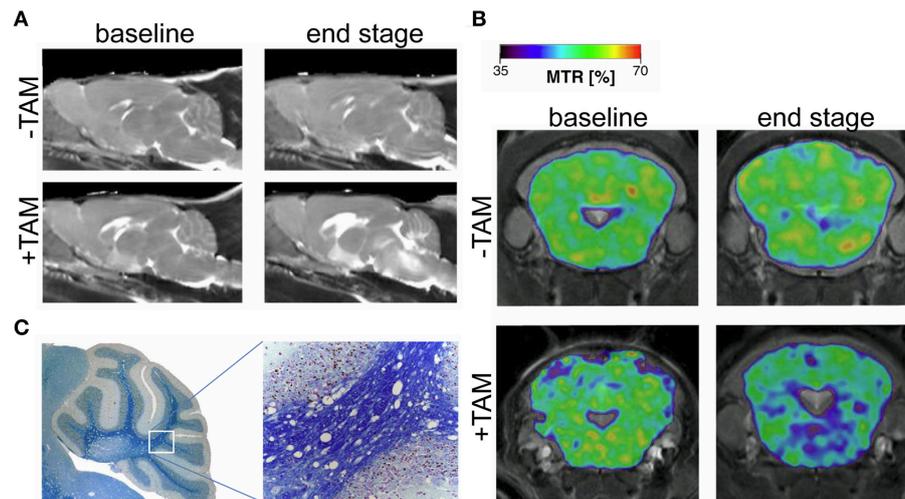
**FIGURE 5 | Hypo-intense regions on microscopic MRI correspond to cellular infiltrates detected by histology. (A)** Coronal  $T_2^*$  weighted imaging ( $T_2^*W$ ) using a multislice fast low angle shot (2D FLASH: TR/TE: 473/18 ms, FA 40°, matrix 512 × 512) sequence with an in plane resolution of  $(35 \times 35) \mu\text{m}^2$ , 22 slices of 500  $\mu\text{m}$ , acquisition time to image the whole mouse brain = 11 min. **(B)** Cellular infiltrates in cerebral cortex; the overview of the H&E histology is overlaid with a coronal slice (plate 41) from Paxinos and Franklin (2013), by permission of Elsevier. **(C)** Susceptibility weighted imaging (SWI) of  $T_2^*W$  scans using fully-automated post-processing by ParaVision 5.1 (Bruker, Ettlingen, Germany). From Waiczies et al. (2012), by permission of Public Library of Science.

tools for studying non-inflammatory demyelination and remyelination (Traka et al., 2010; Pohl et al., 2011). Using a cryogenically-cooled RF coil, pronounced  $T_2$  hyperintensities were revealed in brain stem and cerebellar structures of this model (Figure 6A); these changes were accompanied with a decreased magnetization transfer ratio (MTR; Figure 6B; Mueggler et al., 2012). MTR reflects the exchange of magnetization between pools of differently mobilized protons (Wolff and Balaban, 1989), commonly a rather freely mobile pool and a rather immobile pool associated with macromolecules, such as in axonal membranes or myelin (Alonso-Ortiz et al., 2015). MTR correlates significantly with the degree of myelination as shown in postmortem tissue of MS patients (Schmierer et al., 2004). The decline in MTR that was observed in the oligodendrogliaopathy model with the aid of the CryoProbe (Figure 6B) could be corroborated with evidence of white matter pathology upon histological analysis (Figure 6C; Mueggler et al., 2012).

## Animal Models of Cerebral Amyloid Angiopathies

Neurodegenerative pathologies that involve cerebrovascular dysfunction include the cerebral amyloid angiopathies (CAA); these form a major part of the pathophysiology of Alzheimer's disease (AD; Biffi and Greenberg, 2011). Also in this field, transgenic systems and high resolution MR imaging are proving extremely valuable for investigating the role of vascular dysfunction and amyloid deposition in the pathophysiology and treatment of CAA and AD (Klohs et al., 2014). An important role for MR methods is for them to capture vascular remodeling as a result of hemodynamic alterations; this is important when studying the outcome of CAA on vascular integrity and function (Salat, 2014). The cerebral vascular tree can be assessed by different magnetic resonance angiography (MRA) techniques. The signal in time-of-flight (TOF)-MRA depends on the blood flow and is useful for determining changes in the architecture of major blood vessels as well as reduced blood flow in mouse

models of AD and CAA. When spatial resolution is limited, the tissue surrounding vessels also contributes to the MR signal. Originally TOF-MRA was limited to large cerebral vessels with high flow rates; small vessels suffer from both spin-saturation artifacts and partial volume effects and their edges are more difficult to visualize (Lin et al., 1997). The application of a cryogenically-cooled coil system to increase the resolution of TOF-MRA is a key to improve the detection of smaller vessel (Figure 7A) in order to help identification of pathologies related to neurovascular disease. The enhanced spatial resolution per unit time introduced by the cryogenic system is key to start overcoming some of the partial volume effects that contribute to loss of small vessel visualization. Using a spatial resolution of  $(150 \times 100 \times 100) \mu\text{m}^3$ , flow disturbances at the level of the circle of Willis could be observed in the major arteries of aged animals containing the amyloid precursor protein (APP)23 transgene using TOF-MRA (Beckmann et al., 2003). Administration of superparamagnetic iron oxide (SPIO) particles in association with signal attenuations on three-dimensional (3D) gradient-echo MRI also revealed CAA-related microvascular lesions in different transgenic mouse models of AD including APP23 even at a spatial resolution of  $(109 \times 133 \times 300) \mu\text{m}^3$  (Beckmann et al., 2011). Using a cryogenically-cooled coil system and a spatial resolution of  $(60 \times 60 \times 61) \mu\text{m}^3$ , contrast-enhanced (CE) micro ( $\mu$ )MRA identified age-dependent and CAA-related remodeling of the cerebral microvasculature in arcA $\beta$  mice (Klohs et al., 2012). These transgenic mice express human APP and are characterized by strong CAA pathology (Merlini et al., 2011). Using longer scan times, CE- $\mu$ MRA was performed at even higher spatial resolutions of  $(31 \times 31 \times 93) \mu\text{m}^3$  and vessels could be tracked far into the periphery (Figueiredo et al., 2012). Contrast enhanced (CE)- $\mu$ MRI is expected to complement TOF-MRA (Klohs et al., 2012); after administration of the iron oxide contrast agent, hypointensities not discernable on the precontrast image become visible; these represent intact blood vessels (Figure 7B). In this study, CE- $\mu$ MRA in association with a CryoProbe revealed a reduction in the density of the



**FIGURE 6 | MRI signature in a novel mouse model of genetically induced adult oligodendrocyte cell death. (A)** Parasagittal, quantitative  $T_2$  maps at baseline (day 2), and end stage (day 41) oligodendrogliaopathy following tamoxifen-induced ablation of oligodendrocytes (+TAM) and controls (–TAM). **(B)** MTR maps from representative animals before (–TAM) and after (+TAM) oligodendrocyte ablation at baseline (day 2) and end stage (day 41). MTR maps are color-coded for percentage and superimposed on  $T_2$ -weighted images (cerebellum and brainstem section at the level of  $-2.3$  mm relative to Bregma). **(C)** Magnifications of Histological cerebellar sections from end stage diseased animal stained with Luxol-Nissl for myelin content. From Mueggler et al. (2012), by permission of Academic Press.

microvasculature in the arcA $\beta$  mouse during advanced disease state (Klohs et al., 2012).

## Tumor Models

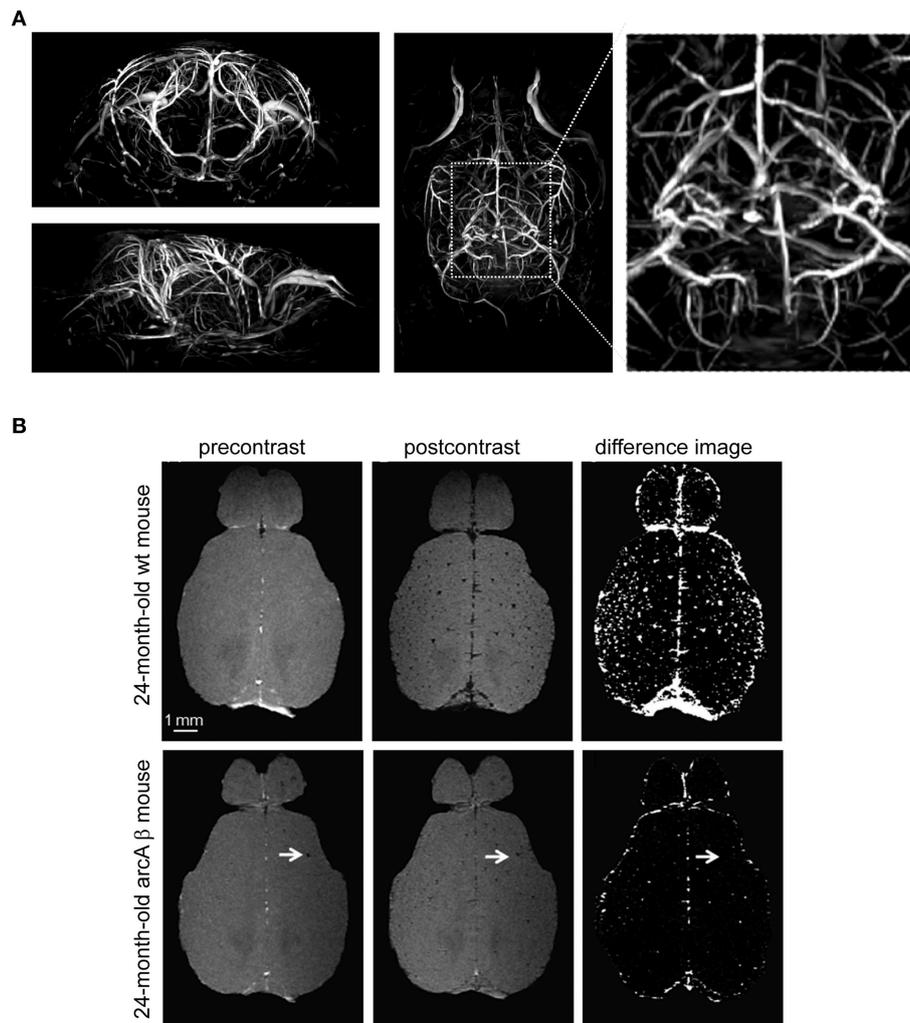
The summed-up advantages of the Cryoprobe are of significant benefit to visualize the pathology of tumor disease as well as assess the outcome of treatment options. One major drawback of standard clinical tumor imaging is associated with limitations in spatial resolution. This major limitation hinders an adequate delineation of tumor tissue from the surrounding healthy tissue prior to surgical tumor resection and results in the reported brain shift during navigated neurosurgical procedures (Reinges et al., 2004). A thorough delineation of tumor borders could be recently achieved in an animal model of high grade glioma by using a Cryoprobe for acquisition of MR images with an in-plane spatial resolution of 51  $\mu$ m (Ku et al., 2013; Vinnakota et al., 2013). In these studies tumor MR microscopy was employed to determine the molecular mechanisms behind tumorigenesis as a precursor to therapeutic studies directed toward identified molecular targets.

Angiogenesis and neovascularization are histopathological hallmarks of cancer (Welti et al., 2013). Molecular determinants that pave these processes need to be targeted to overcome the current limitation of standard treatment regimen in reaching malignant gliomas (Agarwal et al., 2011). The tumor vasculature is functionally and structurally irregular and anti-angiogenic therapies aim to normalize this vascular irregularity (Jain, 2005). MR methods measuring perfusion, permeability, and vessel size has been a necessary tool, commonly requiring application of different contrast agents to monitor tumor neovascularization especially as a means of assessing antiangiogenic therapy response (Zwick et al., 2009; Knutsson et al., 2010). Recently,

vessel architectural imaging (VAI) was introduced and exploited to study tumor vessel caliber, hemodynamics, and relative oxygen saturation and used as a determinant of vessel type and function, especially for therapy response, in glioma patients (Emblem et al., 2013). Vessel size imaging (VSI) and DCE-MRI produce complementary information about the tumor vasculature but need to be performed separately due to the interaction between contrast agents regarding magnetization (Beaumont et al., 2009; Zwick et al., 2009). Recently, a cryogenically-cooled RF coil system was used to combine VSI and DCE-MRI in a glioma mouse model; a single shot gradient echo spin echo (GE-SE) sequence was combined with echo-planar imaging (EPI) and one contrast agent was used (Kording et al., 2014). The superior SNR of the cryogenically-cooled coil that made an increase in the receiver bandwidth possible, in combination with careful shimming routines, ascertained that segmentation and geometry between  $T_2$  maps, ADC maps, GE and SE echo planar images (EPI) was not severely distorted (Kording et al., 2014). In this study the distribution of vessel size was shown to be larger in untreated glioma mice when compared to glioma mice treated with anti-VEGF antibody (bevacizumab) (Kording et al., 2014).

## Blood Oxygenation Level Dependent (BOLD) Functional MRI

With the increase of SNR in small animal MRI it has become increasingly rewarding to advance upon investigations related to central nervous processing. Functional MRI (fMRI) in mice is a crucial tool to characterize transgenic mouse models of human cerebral pathologies and determine the outcome of therapies on the function of large-scale brain circuits. Functional



**FIGURE 7 | Visualization of vascular structures using cryogenically cooled RF coil technology to provide high resolution time-of-flight (TOF)-MRA and spin dephasing following intravascular iron oxide contrast agent. (A)** High-resolution 3D TOF-MRA (axial and horizontal view) of the intracranial and extracranial vasculature of a 2-month-old wild type control mouse using a gradient echo sequence with flow compensation acquired at 9.4 T using the CryoProbe (TR = 30 ms, TE = 5.9 ms, FOV = (30 × 15 × 15) mm<sup>3</sup>, 512 × 256 × 256, isotropic resolution = 59 μm<sup>3</sup>). From Supplementary Data in Waiczies et al. (2012), by permission of Public Library of Science. **(B)** Contrast-enhanced cerebral MRA of a 24-month-old wt mouse and an age-matched arcAβ mouse before and after administration of a superparamagnetic iron oxide contrast agent. Images were acquired using a 3D FLASH sequence [TR = 150 ms, TE = 2.9 ms, FA = 20°, FOV = (15 × 12 × 2.2) mm<sup>3</sup>, spatial resolution of (60 × 60 × 61) mm<sup>3</sup>]. Difference images were obtained by subtraction of postcontrast image from precontrast image. The arrows point to focal hypointense areas that are present before administration of the contrast agent. Scale bar: 1 mm. From Klohs et al. (2012), by permission of Elsevier.

MRI takes advantage of neurovascular coupling as surrogate of neuronal activity. Established measuring techniques track local temporal changes in either blood flow, blood volume or blood oxygenation—all shown as positively correlated with firing rates of adjacent neuronal cell populations (Kim and Ogawa, 2012). Most common techniques in human fMRI make use of the blood oxygenation level-dependent (BOLD) effect. Due to different magnetic properties of oxygenated and deoxygenated hemoglobin temporal changes in local field homogeneity can be detected by  $T_2^*$  weighted MR techniques. Single shot gradient echo based echo planar imaging (GE-EPI) is commonly applied to meet the requirements on BOLD signal sensitivity and high temporal resolution.

To gain sufficient SNR at short acquisition times of 1–8 s comes at the cost of spatial resolution limits in mouse fMRI. At (ultra)high magnetic fields partial brain volumes are commonly acquired with in plane resolutions of about 190–260 microns and a slice thickness of 500 μm (Adamczak et al., 2010; Baltes et al., 2011; Bosshard et al., 2012; Nasrallah et al., 2014; Schroeter et al., 2014). At these voxel sizes GE-EPI is prone to severe image distortions induced by macroscopic magnetic field inhomogeneities, which increase with field strength. Mostly affected regions are air-tissue boundaries (interfaces of bone, tissue, and air). Gaining SNR by using systems with  $B_0 \geq 11.7$  T further increases EPI image distortions (Adamczak et al., 2010). Improving upon SNR by utilizing a CryoProbe permits

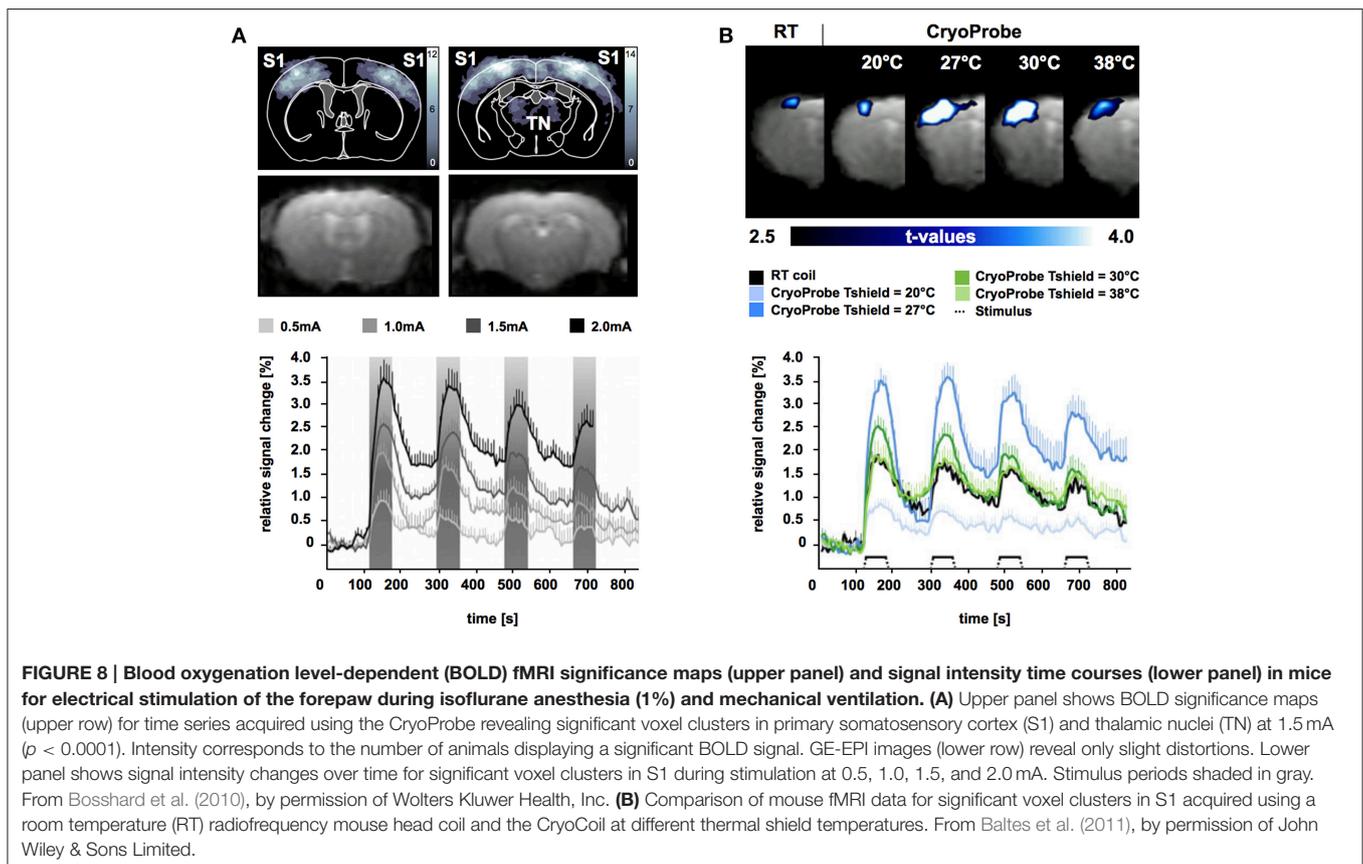
preserving MR signal in imperiled brain regions. The CryoProbe was shown to achieve a three-fold increase in SNR for GE-EPI at 9.4 T compared to a conventional RT surface coil (Baltes et al., 2011). Leveraging on the sensitivity gain of a cryogenically cooled RF coil permits increases in receiver bandwidth, which facilitates inter-echo time shortening in EPI. This results in an increase in the bandwidth along the phase encoding direction which allows for reduction in EPI image distortion (**Figure 8A**).

Functional brain mapping surveys voxel-wise signal changes over time series of acquired image volumes. Signal and image consistency can be significantly impaired by physical (thermal noise, gradient heating) and physiological noise (cardiac, respiration, vascular oscillations). For mouse fMRI physiological noise is particularly relevant: due to the small body size inner organs like heart and lungs are in close proximity to the receiving RF head coil. Physiological noise (e.g., respiratory induced periodic changes in  $B_0$  field homogeneity) gets amplified with higher field strengths. Temporal SNR (tSNR) considers physical and physiological fluctuations over time making it an important measure for fMRI data quality. Baltes and colleagues reported a 1 to 3-fold increase in tSNR with the CryoProbe over a conventional RT mouse head coil at 9.4 T (Baltes et al., 2011).

Somatosensory studies using the CryoProbe revealed most significant BOLD signal magnitudes in mouse fMRI with external stimuli (Bosshard et al., 2010, 2012, 2015; Baltes et al., 2011; Mueggler et al., 2011; Schroeter et al., 2014). Signal

intensity changes of up to 3.5% were observed in the primary somatosensory cortex for electro-stimulation (**Figures 8A,B**; Bosshard et al., 2010, 2012; Baltes et al., 2011; Schroeter et al., 2014) and moderate noxious heat stimulation (Bosshard et al., 2015) of the murine paw under isoflurane anesthesia. In murine fMRI studies investigating somatosensation and pain commonly anesthesia is applied for ethical and practical reasons (Borsook and Becerra, 2011). Under these conditions BOLD magnitude strongly depends on the applied anesthetic protocol and control of physiological parameters (Schroeter et al., 2014). This makes it somewhat difficult to compare fMRI data obtained with a CryoProbe setup to other somatosensory mouse fMRI studies using conventional room temperature RF coils.

A direct comparison of maximum BOLD signal changes during electrostimulation of the murine forepaw using the CryoProbe compared to a conventional RF mouse head coil was conducted by Baltes et al. (2011). In this study scalp temperature was found to be an important factor influencing the BOLD signal magnitude. To ensure physiological temperatures at the surface of the CryoProbe it features a thermal shield, which can be adjusted in a certain temperature range. An internal heater generates a thermal gradient along the ceramic surface. The shield heating can be adjusted to result in the desired temperature at the scalp-coil interface. In anesthetized mice reduced scalp temperatures promote vasoconstrictive effects ascertained by reduced baseline perfusion (Baltes et al., 2011). Higher SNR



is expected to aid BOLD signal detection but should not alter the BOLD effect itself. At physiological temperatures ( $T_{\text{scalp}}$  of approx.  $33.5^{\circ}\text{C}$ ) maximum BOLD magnitude was found to be similar for the CryoProbe and conventional RF mouse head coil during electrostimulation of the forepaw (**Figure 8B**). Mild cerebral hypothermia ( $T_{\text{shield}} = 27^{\circ}\text{C}$ ) was shown to significantly boost BOLD signal magnitude whereas severe hypothermia ( $T_{\text{shield}} = 20^{\circ}\text{C}$ ) decreased BOLD signal intensity, most likely due to impairments in physiological processes. The temporal noise for baseline measurements was significantly reduced for the CryoProbe for all thermal shield temperatures up to factor 2 compared to conventional RF mouse head coil (Baltes et al., 2011). Therefore, the CryoProbe provides higher BOLD signal sensitivity and might be capable to detect smaller neurovascular changes. The possibility to adjust its surface temperature permits to sustain more physiological thermal conditions in anesthetized mice.

## CARDIOVASCULAR APPLICATIONS

### *In Vivo* Assessment of Cardiac Morphology and Function

Realizing that cardiovascular disease (CVD) is one of the leading causes of death (Heidenreich et al., 2011), research in pathologies related to heart function remain at the forefront of academia and pharmaceutical industry. For phenotyping of disease models (e.g., myocardial infarction or hypertension-induced myocardial injury/remodeling), as well as assessment of gender effects, evaluation of novel therapeutics and long-term follow-up studies non-invasive *in vivo* imaging of the heart is conceptually appealing. Cardiovascular magnetic resonance (CMR) is widely used in preclinical studies, the workhorse being an assessment of cardiac morphology and function. For a comprehensive and up-to-date review of CMR in small rodents please refer to Bakermans et al. (2015).

A highly reproducible quantitative assessment of cardiac morphology and function by CMR demands excellent spatial and temporal resolution. Both come at the cost of SNR loss, which can only be partially compensated for by increasing measurement time. Together with the small size of the mouse heart and its rapid motion (typical heart rates are 400–600 beats per minute) these SNR limitations pose serious challenges to CMR in mice. For cardiac chamber quantification and left ventricular function assessment an excellent delineation of myocardial borders, high ventricular blood-to-myocardium contrast and full coverage of the cardiac cycle with high temporal resolution are required. Furthermore, a complete coverage of the entire heart with a sufficiently high spatial resolution to facilitate reliable segmentation of the endo- and epicardial borders is crucial.

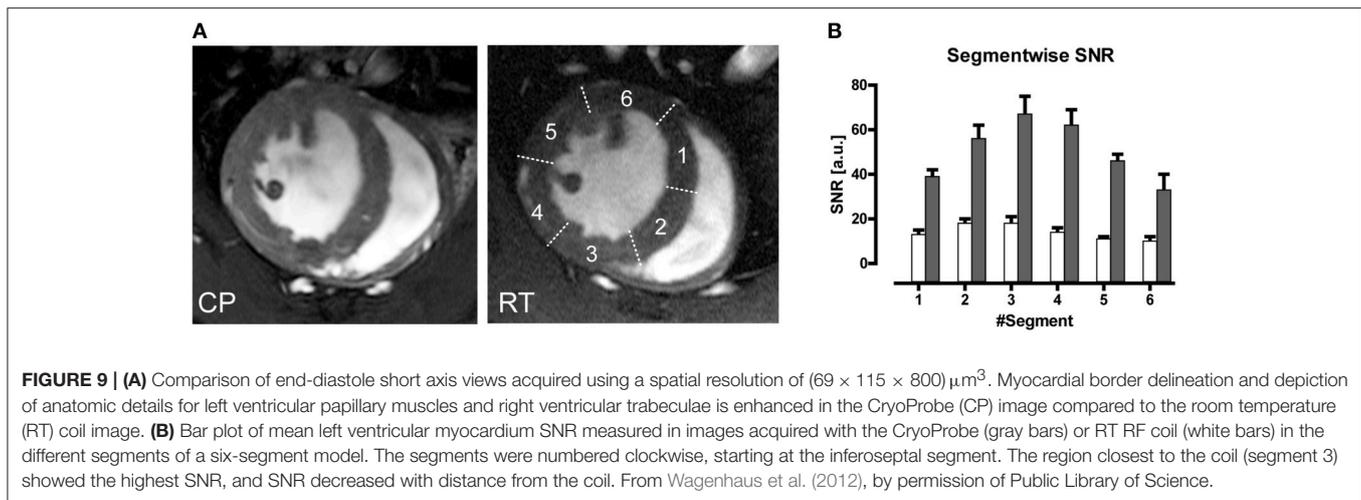
Current experimental CMR of mice uses dedicated birdcage volume RF coils or surface RF coil arrays with the geometry adjusted to the anatomy of the mice (Epstein, 2007; Heijman et al., 2007; Hiba et al., 2007; Young et al., 2009; Ratering et al., 2010; Schneider, 2011; Schneider et al., 2011) and affords images with a typical in-plane spatial resolution of 100–200  $\mu\text{m}$ , heart coverage of 6–13 slices of 1.0 mm thickness and measurement

of 10–20 cardiac phases (Epstein, 2007; Heijman et al., 2007, 2008; Hiba et al., 2007; Sosnovik et al., 2007; Young et al., 2009; Ratering et al., 2010; Bovens et al., 2011; Schneider, 2011; Schneider et al., 2011). Although providing acceptable image quality for the quantitative assessment of the left ventricle (LV), further improvement in image quality is highly desirable, particularly for the assessment of the right ventricle (RV). To meet this goal, enhancements in the spatial resolution are essential, which in turn build upon SNR improvements.

An average gain in SNR of 3.6 in murine cardiac MRI was achieved when comparing a CryoProbe at 9.4 T with a conventional room temperature RF coil setup (receive-only four-element mouse heart surface array RF coil in combination with a volume resonator for transmission (Wagenhaus et al., 2012)). The SNR gain ranged 3.0–5.0 within the myocardium and could be translated into higher spatial resolution imaging. The highly detailed anatomical images derived from the CryoProbe acquisitions provided significantly improved myocardial border sharpness vs. the RT-coil acquisitions as illustrated in **Figure 9A**. Hard evidence for the benefit of the enhanced image quality was the improved reproducibility of the quantitative morphology and function assessment. Using the CryoProbe, intraobserver, and interobserver variability were smaller for almost all cardiac function parameters. For instance intraobserver variability in end-diastolic mass (EDM), end-diastolic volume (EDV), and end-systolic volume (ESV) were reduced on average by 59 and 66% for the left and right ventricle, respectively. The practical advantage is two-fold. An increased sensitivity will allow an earlier detection of pathological changes as well as a more thorough assessment of therapeutic effects. A smaller statistical variability within experimental groups allows for a significant reduction in animal group size.

Currently available cryogenic RF technology comes along with some challenges for performing CMR. The CryoProbe cannot easily be placed onto any anatomical region of interest since it must be installed inside the magnet bore. Instead, the animal is placed underneath the surface RF coil using a dedicated cradle. This set-up is tailored and well-suited for mouse brain MRI. To use it for CMR requires a special supine positioning of the mouse. Training and practice is necessary to position the heart correctly within the field of view of the CryoProbe. The supine position alters the shape of the RV (see short-axis view in **Figure 9A** and long-axis view in Wagenhaus et al., 2012) but does not impact on the functional parameters (Wagenhaus et al., 2012). In some animals the supine position is accompanied by a difference in motion of the septum, which continues throughout diastole and results in blurring of the endocardial borders of the septum.

Inherent to surface coils is a decrease in signal-amplitude with increasing distance (as explained above). Hence the CryoProbe setup—just like the conventional RT surface RF coil setup—displays a spatial variation in SNR across the myocardial segments of a short axis view (**Figure 9B**), which can be explained with the proximity to the surface coil. Inhomogeneity on the RF transmit side ( $B_1^+$ ) is usually of little concern for gradient-echo based protocols such as the CINE FLASH commonly used for assessment of cardiac morphology and function (**Figure 9A**; Wagenhaus et al., 2012). Yet for techniques in which an exact



flip angle is crucial—such as saturation based  $T_1$ -weighted gadolinium-enhanced first pass bolus perfusion studies (Utz et al., 2007, 2008) or inversion recovery prepared  $T_1$ -mapping for detection of fibrosis or characterization of myocardial tissue (Messroghli et al., 2003)— $B_1^+$  inhomogeneity may be an unfavorable characteristic of the CryoProbe. Such cardiac applications can benefit from using a volume resonator for transmission in conjunction with the state-of-the-art receive-only CryoProbe.

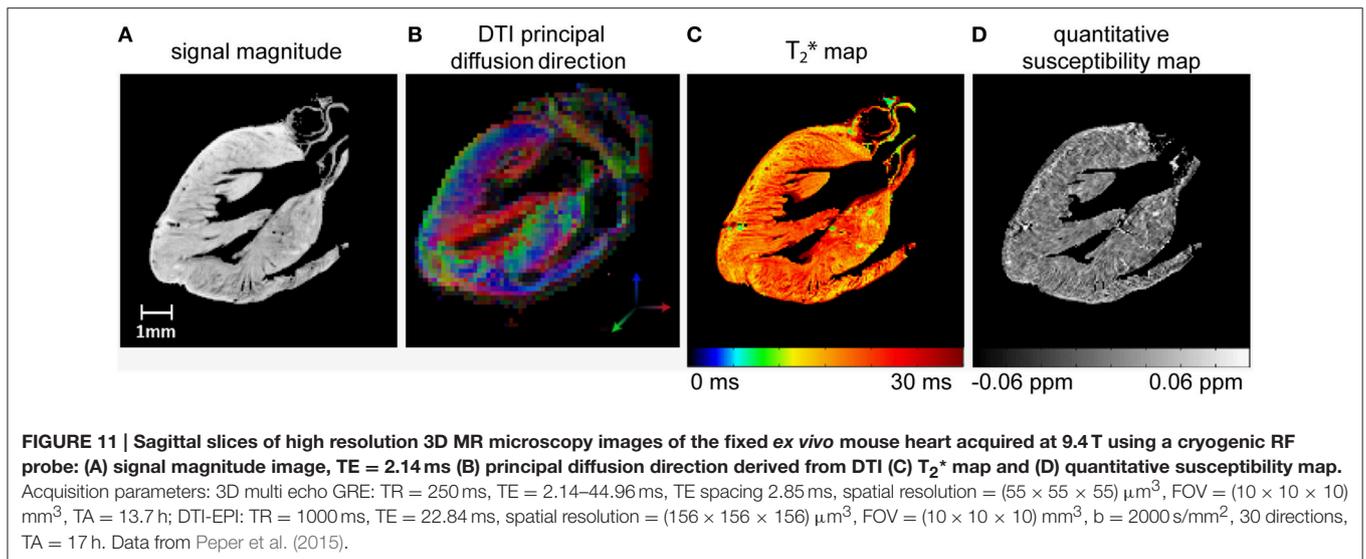
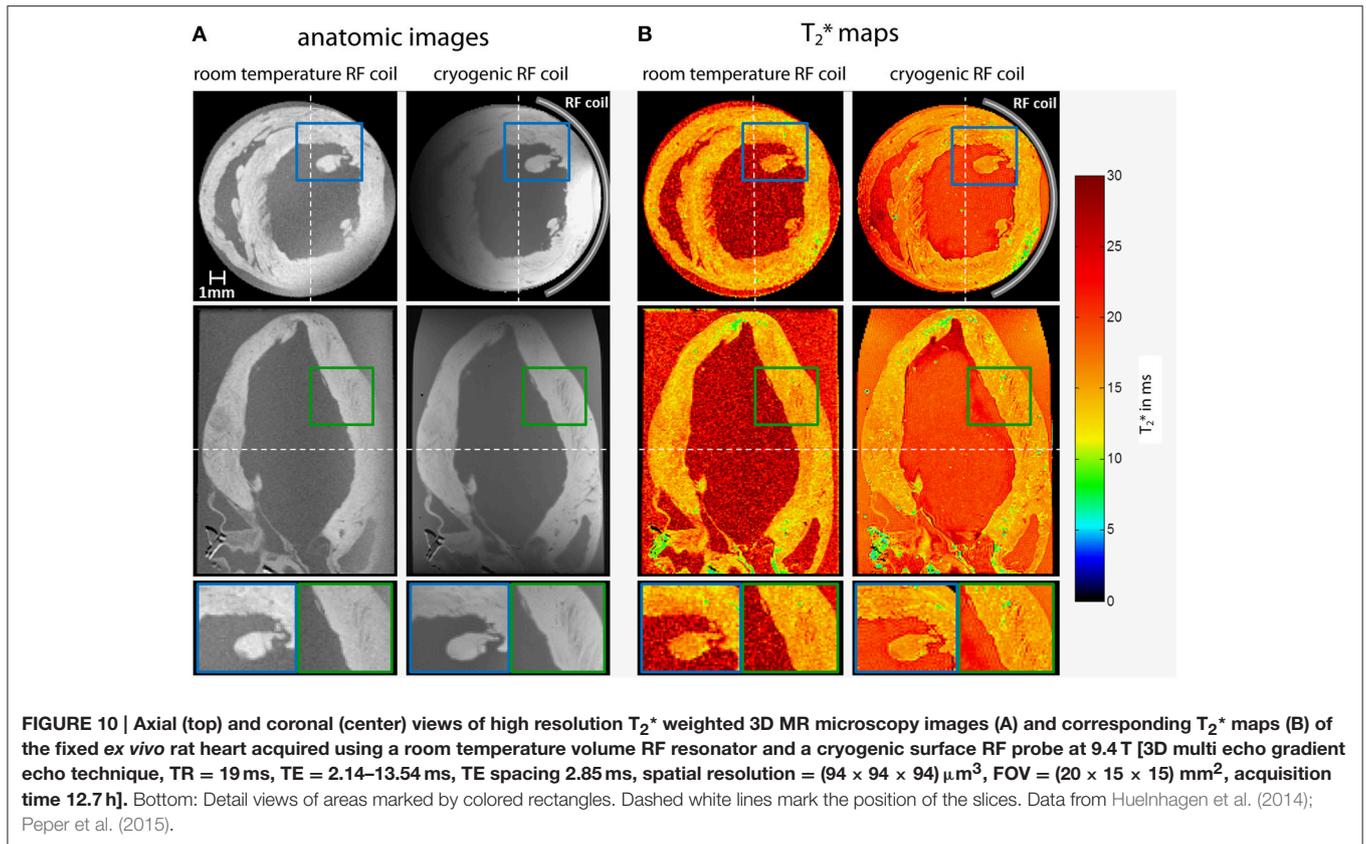
In conclusion, cryogenically cooled RF coils represent a valuable means of enhancing the image quality in *in-vivo* CMR of mice. They permit high spatial resolution CINE imaging of the mouse heart with excellent SNR, which measurably improves the reproducibility of quantitative cardiac morphology and function assessment. This is important since segmentation of the myocardium is challenging and prone to introduce significant data variability.

## Ex Vivo MR Microscopy of the Rodent Heart

MR microscopy is a powerful tool to get an insight into the micro architecture of myocardial tissue and provides results similar to histology in a non-invasive matter. High spatial resolution and an appropriate organ coverage are both essential. Quantitative techniques like  $T_2^*$  mapping, diffusion tensor imaging (DTI) or quantitative susceptibility mapping (QSM) can complement the information provided from common MR contrasts and allow intersubject assessment. DTI or QSM require 3D datasets with high isotropic resolution. Combining the requirements of high resolution MR microscopy with large volume or whole heart spatial coverage already puts a strain on balancing SNR with scan time.  $T_2^*$  weighting and diffusion weighting, as needed for DTI or QSM, are also factors that inherently reduce SNR: Sufficient initial SNR is needed, to ensure adequate signal even at long TEs or for strong diffusion weighting. Thus, several features of quantitative myocardial MR microscopy constrain SNR and bring about the need for lengthy scan times to compensate for SNR losses. Even in *ex vivo* scans where acquisition time is by far not as limited as in *in vivo* scans, counteracting SNR

losses by averaging can easily result in days of acquisition time. Unfortunately, this condition is not feasible even in an *ex vivo* setting. Using a cryogenic RF probe helps to balance the competing constraints of high spatial resolution, high spatial coverage and sufficient  $T_2^*$  or diffusion weighting in acceptable acquisition times at ample signal to noise.

**Figure 10** shows examples for high resolution  $T_2^*$  weighted 3D MR microscopy of the fixed *ex vivo* rat heart at 9.4T. These short and long axis views of the heart were derived from acquisitions using a conventional room temperature birdcage RF coil and a cryogenic surface RF coil (Huelnhagen et al., 2014; Peper et al., 2015). Signal magnitude and corresponding  $T_2^*$  maps derived from fitting the signal decay over 5 echo times are shown. Sub-millimeter structures, which can clearly be delineated in images obtained with the CryoProbe, are dominated by noise in the images acquired using the room temperature RF coil. With identical sequence and protocol settings and equal acquisition times approximately a five-fold mean SNR enhancement was achieved in this example when using the cryogenic RF probe. Attributing the SNR increase in this case solely to the fact that a cryogenic probe has been used would not be entirely appropriate since moving from a volume to a surface RF resonator usually goes along with an SNR gain at least in regions close to the RF coil. Notwithstanding this limitation it is fair to state that high SNR level images obtained with the cryogenic RF probe cannot be accomplished with the room temperature probe using the same imaging protocol and scan times. The SNR and signal uniformity implications of the limited field of view and  $B_1$  gradient of the cryogenic surface RF coil can be recognized from the axial images. These non-uniformities can be corrected but might constitute a challenge for sample sizes not matching the coil geometry. If the sample size however, is in the range of the field of view specifications of the cryogenic RF surface coil, high resolution MR microscopy provides images that are comparable with histology, earning heart MR microscopy the moniker of cardiac MR histology. **Figure 11** illustrates an example of a fixed *ex-vivo* mouse heart scanned in 3D with an isotropic spatial resolution of  $55 \mu\text{m}^3$  using a cryogenic surface RF coil at 9.4T (Peper et al., 2015).



Making use of the SNR gain of the cryogenic RF probe, despite the very high spatial resolution, an SNR of more than 60 was achieved for the first echo (TE = 2.14 ms). Even for long echo times SNR obtained with the cryogenic RF coil is appropriate to accommodate accurate  $T_2^*$  mapping.

To summarize, MR microscopy of the heart with a high spatial coverage heavily benefits from using cryogenic RF resonators especially when contrast weighting is required, as in parametric

mapping or diffusion weighted applications. The benefit is paramount during the characterization and phenotyping of animal models of cardiac disease as it provides a valuable means of getting a better understanding of myocardial microstructure and myocardial remodeling. The value of such improvements are in positive alignment with human MR studies at ultrahigh magnetic fields (Von Knobelsdorff-Brenkenhoff et al., 2010, 2013; Dieringer et al., 2011; Thalhammer et al., 2012; Winter et al.,

2012; Graessl et al., 2014) which strive for a relative spatial resolution (number of pixel per cardiac anatomy) approaching that of experimental MRI in small rodents with the ultimate goal to provide imaging means for diagnostics and for guiding treatment decisions in cardiovascular and metabolic diseases (Niendorf et al., 2010, 2013, 2015a).

## RENAL APPLICATIONS

### Ex Vivo MR Microscopy of the Rodent Kidney

Hurlston et al. (1999) ventured into an early exploration of *ex vivo* mouse kidney MR microscopy; their prototype HTS Helmholtz RF probe provided a seven-fold gain in SNR compared to a room temperature Helmholtz RF coil. Using the superconducting coil, the feasibility of renal MR microscopy was demonstrated at 9.4 T with  $17\ \mu\text{m}$  in-plane resolution in little more than an hour scan time (Figure 12; 3D spin-echo,  $(17 \times 17 \times 136)\ \mu\text{m}^3$  spatial resolution).

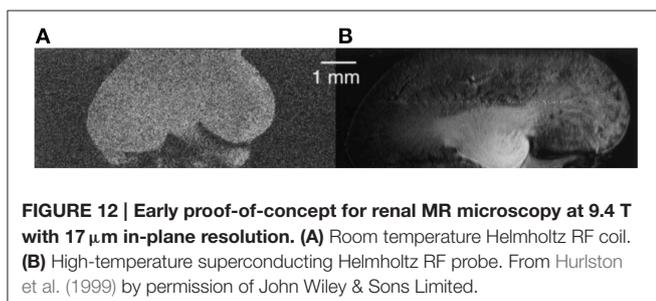
The sensitivity gain of cryogenic RF coil technology can be put to good use for renal MR microscopy of glomeruli, the microscopic filtering units of the kidney. Glomerular size and number is thought to be linked to several renal and cardiovascular diseases (Hoy et al., 2008). While established techniques for measuring/counting glomeruli rely on extrapolations made from measurements/counts within small samples (Beeman et al., 2011), a total and direct quantification can be made by renal MR microscopy (Beeman et al., 2011; Heilmann et al., 2012). By exploiting the sensitivity boost of a CryoProbe, Heilmann et al. (2012) were able to quantify glomerular number and size in ferritin labeled rat kidneys *ex vivo* with a scan of less than 5 h duration. Automatic segmentation of 3D GRE images with  $(35 \times 35 \times 35)\ \mu\text{m}^3$  spatial resolution yielded glomerular diameters (mean  $\pm$  SD =  $109 \pm 4.9\ \mu\text{m}$ ) and counts (mean  $\pm$  SD =  $32,785 \pm 3117$ ) with low intra-subject variability in one-seventh of the time required for traditional stereology.

### In Vivo Structural and Functional Renal MRI

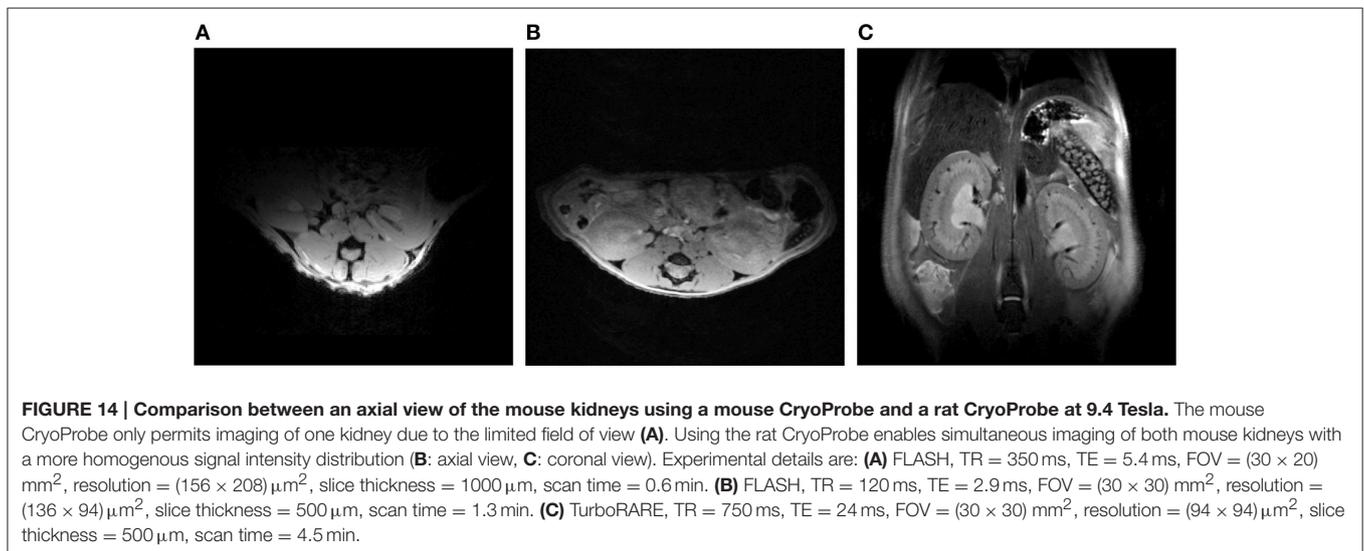
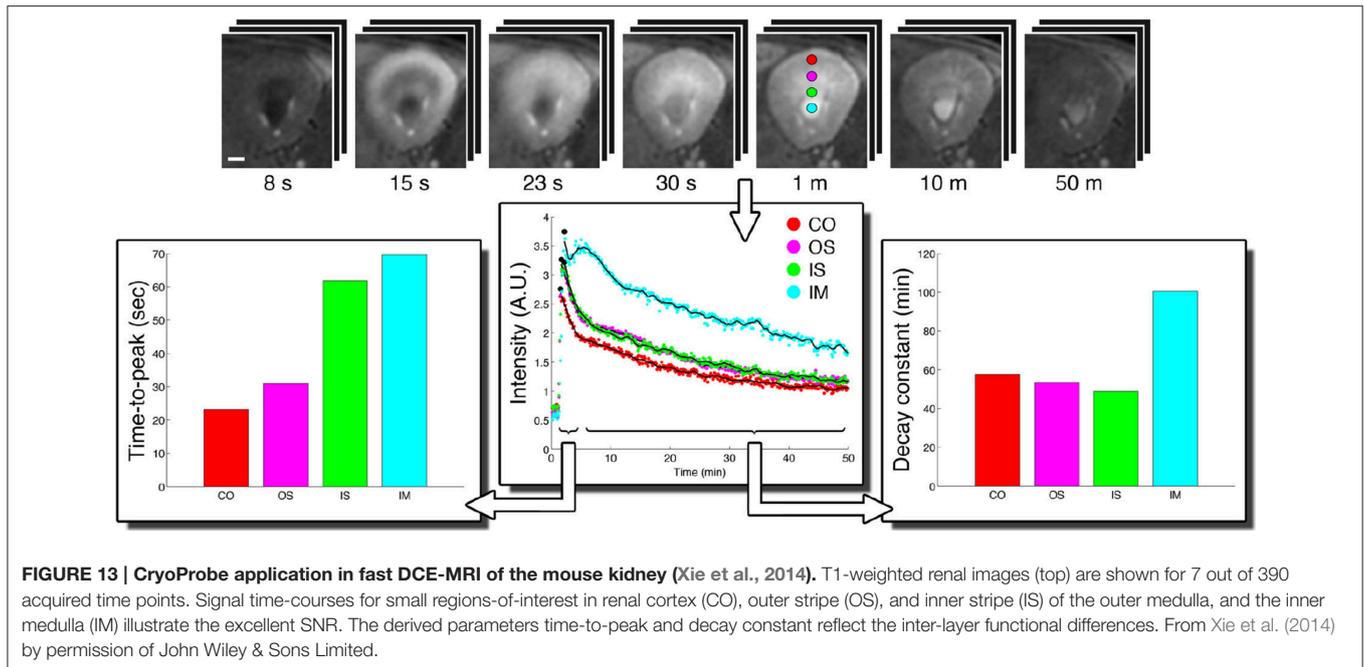
To date the potential for improving spatial resolution and increasing temporal resolution by use of a CryoProbe for *in vivo* renal MRI remains largely untapped. The boost in SNR can be expected to be similar to that reported for cardiac applications,

i.e., approximately three-fold (Wagenhaus et al., 2012). Organ size and depth of location within the thorax/abdomen are comparable. The same room temperature RF coils are typically used for both applications, commonly a four-element saddle-shaped receiver (RX) array in conjunction with a larger transmit (TX) volume resonator. *In vivo* MRI of a mouse kidney is perfectly feasible with a mouse CryoProbe (Xie et al., 2014; Figure 13). However, imaging of both kidneys simultaneously requires a much larger field of view than needed for a single kidney or the heart. The rat CryoProbe may be an attractive alternative as it provides a uniform signal intensity distribution over a larger field of view than the mouse CryoProbe (Figure 14). Objective signal and noise measurements will however still be required since these coils were not originally designed for matching the geometry of the mouse body and for optimized kidney imaging.

The combination of high speed acquisition with almost microscopic spatial resolution is of great interest also in other areas of functional renal MRI, such as  $T_2^*$  monitoring, as surrogate for renal blood oxygenation. Parametric mapping of  $T_2^*$  with sub-minute temporal resolution is essential for capturing the fast dynamic changes in renal hemodynamics and oxygenation during acute ischemic events or physiological test stimuli (Pohlmann et al., 2013b, 2014a). Comparison with renal MR microscopy images (Xie et al., 2012; Niendorf et al., 2015b) suggests that the limited spatial resolution of current *in vivo* protocols used for monitoring  $T_2^*$  masks some of the underlying intra-layer heterogeneity of renal structure and function. Higher spatial resolution for renal parametric MRI, which puts a strain on SNR, is therefore warranted especially in the mouse. Reduced oxygenation of intrarenal blood can lead to an additional loss of signal, which can be substantial in images acquired at larger echo times. Elucidating ischemia/reperfusion (I/R) injury, a consequence of kidney hypoperfusion or temporary interruption of blood flow—a common cause of acute kidney injury (AKI)—requires fast continuous  $T_2^*/T_2$  mapping throughout baseline, ischemia, early reperfusion, and recovery (Pohlmann et al., 2013b) and hence benefits from scan acceleration using parallel imaging strategies. To this end, the SNR gain offered by a multi-channel receive cryogenic RF coil array is instrumental for compensating noise amplification inherent to parallel imaging (Niendorf and Sodickson, 2006a,b, 2008). The speed gain is of benefit in preclinical renal MRI studies where multiple parameters are assessed by multiple modalities including (i) quantitative physiological measurements such as renal perfusion pressure, renal blood flow, local cortical and medullary tissue  $pO_2$  and blood flux and, (ii) comprehensive MRI protocols with tight spatio-temporal resolution constraints dictated by renal (patho)physiology and the interleaving with the quantitative physiological measurements (Pohlmann et al., 2014a). For all these reasons, cryogenic RF coil technology holds great potential for enabling non-invasive *in vivo* investigations into renal hemodynamics and oxygenation with MRI. Conventional methods for assessing renal hemodynamics and oxygenation such as fiber-optical  $pO_2$  probes or ultrasonic flow probes provide quantitative physiological data, but are invasive and the



**FIGURE 12 | Early proof-of-concept for renal MR microscopy at 9.4 T with  $17\ \mu\text{m}$  in-plane resolution. (A)** Room temperature Helmholtz RF coil. **(B)** High-temperature superconducting Helmholtz RF probe. From Hurlston et al. (1999) by permission of John Wiley & Sons Limited.



fiber probes enable measurements in pin-point locations only (Arakelyan et al., 2013; Pohlmann et al., 2013a).

### Assessing Renal Perfusion, Oxygenation, or Inflammatory Cell Infiltration Using Fluorine (<sup>19</sup>F) MRI

Renal tissue hypoxia and inflammatory mechanisms play prominent roles in the pathophysiological chain of events that lead to acute kidney injury and promotes progression from acute injury to chronic kidney disease. Dependent on disease etiology, the contribution and impact of inflammatory mechanisms may vary (Chawla et al., 2014). Inflammatory processes, including

early responses dominated by the innate immune system as well as later responses by the adaptive immune system, can support repair and restoration of renal functions but may also promote renal tissue injury and transition to chronic disease (Bonventre and Yang, 2011; Kinsey and Okusa, 2014; Molitoris, 2014).

Acute kidney injury can also trigger a systemic immune response that may lead to secondary dysfunction of organs such as heart, brain, and lung (Grams and Rabb, 2012). Alternatively, systemic immune diseases can have a crucial impact on the kidney e.g., antineutrophil cytoplasmic antibody (ANCA)-associated vasculitis often affects the kidneys leading to rapid-progressive glomerulonephritis (Schreiber and Choi, 2015).

Renal blood volume, blood oxygenation, and inflammatory cell migration can be assessed and monitored using cutting-edge  $^{19}\text{F}$  MR techniques (Ruiz-Cabello et al., 2011; Hu et al., 2014). MRI of x-nuclei suffers from an inherently low SNR due to limited MR sensitivity ( $^{19}\text{F} \approx ^1\text{H}$ ) but also a low abundance of nuclei in the tissue of interest. Indeed,  $^{19}\text{F}$  is virtually absent in the living tissues of rodents and the  $^{19}\text{F}$  signal is created by injection of exogenous  $^{19}\text{F}$  contrast agents. A commonly used variant of such  $^{19}\text{F}$  agents is perfluorocarbon (PFC) (Ruiz-Cabello et al., 2011), which can be used to label inflammatory cells *in vivo* after systemic application (Waiczies et al., 2013) or track specific immune cells *in vivo* after *in vitro* labeling (Ahrens et al., 2005; Waiczies et al., 2011).

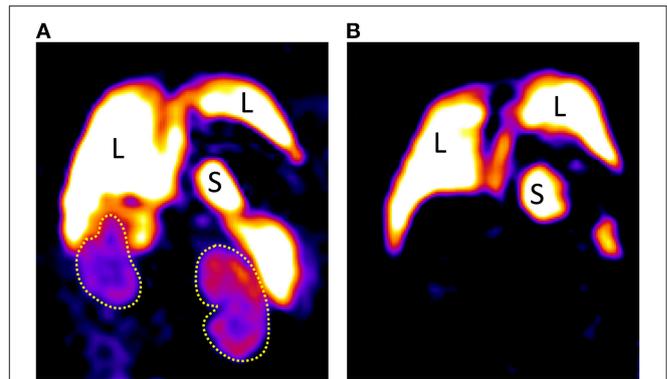
Systemic i.v. administration of  $^{19}\text{F}$  nanoparticles (droplet emulsion) results in uptake and self-labeling by phagocytic immune cells e.g., macrophages, neutrophils or dendritic cells. Application of these nanoparticles to an ANCA-induced glomerulonephritis model demonstrated a significant  $^{19}\text{F}$  MRI signal in the kidney (in contrast to kidneys of control animals that showed negligible signal). This method provides an *in vivo* depiction of renal inflammatory cell dynamics (Figure 15; Pohlmann et al., 2014b).

Yet, the rather low spatial resolution of ( $0.94 \times 0.94 \times 1.88$ )  $\text{mm}^3$  and required acquisition time of almost an hour leave a lot to be desired. Here the low SNR is a fundamental factor that is restricting the capabilities of  $^{19}\text{F}$  MR in rodents. SNR negatively correlates to the detection limit for  $^{19}\text{F}$  labeled cells. Applications in which harvested immune cells are labeled *in vitro* and re-applied to an animal have an even greater need for high sensitivity because in this case the number of labeled cells in the region of interest is much smaller (Waiczies et al., 2013).

Noteworthy for  $^{19}\text{F}$  MRI, a gain in SNR of 3.0–3.5 for the  $^{13}\text{C}$  CryoProbe (Sack et al., 2014) and 2.5 for the  $^1\text{H}$  CryoProbe (Baltes et al., 2009) were reported at 9.4 T. Considering the close proximity of  $^{19}\text{F}$  and  $^1\text{H}$  frequency, an SNR gain of at least 2.5 will be expected for a  $^{19}\text{F}$  CryoProbe at 9.4 T, a gain that would permit an enhanced spatio-temporal resolution of preclinical  $^{19}\text{F}$  MRI and pave the way for better detection and more efficient tracking of cellular therapies such as dendritic cells that are used in a variety of cancers. With a technology that provides better detection of cells, even in far reaching organs, the best cell therapy solutions for pathologies such as recalcitrant cancers can be determined more efficiently and swiftly. These therapeutic solutions in combination with advanced clinical imaging technologies can ultimately be translated to the clinic and incorporated into personalized medicine.

## OUTLOOK

The cryogenic RF coil technology has introduced significant advancements to small animal MR image acquisition in different areas of disease research. With the ever increasing clinical needs and research explorations looming over the MR horizon, further innovations in association with this technology are nevertheless likely to be expected. The opportunity of increasing sensitivity as well as spectral, spatial, and temporal resolution as a result of increased SNR with the cryogenic RF coil technology opens



**FIGURE 15 | (A)** Coronal  $^{19}\text{F}$  images of a mouse with ANCA-induced glomerulonephritis; 8 weeks after bone marrow transplantation showing extensive renal inflammatory cell infiltration, particularly in the cortex in comparison with a naive mouse **(B)** showing a high signal in liver and spleen, but none in the kidneys. The low SNR of room temperature RF technology severely limits the spatial resolution achievable even at extended scan durations of almost 1 h. L indicates liver, S indicates spleen, and yellow dot lines depict kidneys. Data from Schreiber et al. (2013).

up more prospects for advancing evolving MR techniques in spectroscopy (MRS), imaging (MRI), or spectroscopic imaging (MRSI). MR methods for studying the detection and distribution of metabolites containing proton ( $^1\text{H}$ ) and x-nuclei e.g., carbon ( $^{13}\text{C}$ ), oxygen ( $^{17}\text{O}$ ), sodium ( $^{23}\text{Na}$ ), and phosphorous ( $^{31}\text{P}$ ) will benefit immensely from the introduction of cryogenically-cooled coils. In addition to the increased SNR, which is particularly favorable for nuclei with a relatively low biological abundance (e.g.,  $^{13}\text{C}$ ,  $^{23}\text{Na}$ ,  $^{31}\text{P}$ ), better spectral resolutions are expected to reveal metabolites that have until now remained undetected, keeping in mind that resolution comes at a cost in SNR. Another evolving field that will benefit from cryogenically cooled RF coils is functional MRS (fMRS), which when used in conjunction with BOLD fMRI will be invaluable to study changes in resting and activation states in the brain. MRSI employing high-speed echo-planar encoding will also benefit from sensitivity gains to spatially map multiple tissue metabolite signals *in vivo*.

A gain in sensitivity and resolution that can be delivered by cryogenically cooled RF coils can be attained by increasing magnetic field ( $B_0$ ) strengths, but the side effects and costs associated with higher  $B_0$  are in such a case non-existent. Challenging adverse effects of higher  $B_0$  (that can be eliminated with cryogenic RF coil cooling at lower fields) include saturation-related signal losses due to longer T1 relaxation times, shorter echo time acquisitions to compensate for sensitivity losses due to shorter T2 relaxation times and line broadening or off-resonant effects due to  $B_0$  susceptibility effects. For small animal imaging, cryogenically cooled RF coils would benefit from small cryogenic-liquid free magnets systems with variable field operation and automatic field ramping. These MR systems can be operated at magnetic field strengths of 1.5, 3.0, or 7.0 T and hence provide opportunities for harmonizing basic with translational research. The installation and maintenance of such MR systems is affordable and thus ideal for preclinical research. Equipped with one cryogenically cooled RF coil such MR systems with cryogenic

liquid-free magnets will be an invaluable technology for disease phenotyping, multi-modal imaging as well as for spanning of an assortment of x-nuclei for metabolic, molecular, and cellular imaging.

The benefits of the cryogenically-cooled RF coil technologies are in positive alignment with our human MR studies at ultrahigh  $B_0$  fields, which strive for the spatial resolution used in experimental small animal MRI, but necessitate complex techniques (such as volume-selective higher-order  $B_0$  shimming and phase correction strategies) to counterbalance the  $B_0$  adverse effects. Considering the advantages of cryogenically cooled RF coils and the expected immense application for human studies a swift clinical translation is warranted. For starters, tremendous sensitivity gains will be expected when using smaller cryogenically cooled coils in association with higher  $B_0$ ; the stronger magnetic coupling between small coil and sample is attributed to higher signal sensitivities and the smaller volume of tissue seen by smaller coils is attributed to lower noise. MR applications that will particularly benefit from cryogenically-cooled RF coil systems, particularly at UHF strengths, are diffusion weighted imaging (DWI) methods employing stronger diffusion-weighting ( $b$ -values  $> 2000 \text{ s/mm}^{-2}$ ) in order to demonstrate the biexponential decay of brain diffusion, and better differentiate between different water compartments, especially during ischaemic and other brain pathology (Norris and Niendorf, 1995; Dijkhuizen et al., 1996; Niendorf et al., 1996). Although higher  $b$ -values yield higher diffusion contrast, lower SNR is expected on heavily diffusion-weighted images, such that cryogenically-cooled coils will surely be invaluable for this application. Alternatively, the SNR gain inherent to cryogenically-cooled RF coils could be transferred into using a larger range of diffusion weighting, which would be of great benefit for *in vivo* explorations into the biophysics of water diffusion in tissue (Niendorf et al., 1994; Norris et al., 1994; Pyatigorskaya et al., 2014; Le Bihan and Iima, 2015) but also for the study of intracellular compartmentation of metabolites, with the implication feeding into neuroradiology, neurology and related clinical disciplines (Najac et al., 2014). Other foreseen human applications for cryogenically-cooled RF coils will include anatomic micro-imaging of the skin. Pioneering work undertaken with a HTS coil at 1.5 T recently demonstrated a subnanoliter spatial resolution ( $80 \mu\text{m}^3$ ) for human skin

MR microscopy (Laistler et al., 2015). Skin-sodium storage, as a physiologically important regulatory mechanism for blood pressure, volume regulation, and indeed survival, has recently been rediscovered. This prompted the development of MRI methods to assess sodium storage in humans with  $^{23}\text{Na}$ -MRI at 3.0 Tesla (Kopp et al., 2012, 2013) and at 7.0 T; the latter facilitating a spatial resolution across the skin of as good as 1 mm in-plane (Linz et al., 2015). To this end, cryogenically cooled RF technology indeed represents a powerful research tool that can potentially help to unlock questions regarding  $\text{Na}^+$  balance and  $\text{Na}^+$  storage functions of skin with the ultimate goal to provide imaging means for diagnostics and for guiding treatment decisions in cardiovascular and metabolic diseases. It is however envisioned that the benefits of cryogenically-cooled coils for human application at ultrahigh fields will also go beyond skin microimaging of sub-nanoliter voxels. The significance of the cryogenic technology to advance upon current x-nuclei MR methods for metabolic probing (Linz et al., 2015) and pharmacological studies (Ji et al., 2015) is an area that will definitely receive considerable attention in the coming years.

## AUTHOR CONTRIBUTIONS

TN, AP, and SW wrote the manuscript with help from HR, TH, and MK. HW, EP, ES, AS, RK, and KS contributed to data and intellectual feedback.

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## REFERENCES

- Adamczak, J. M., Farr, T. D., Seehafer, J. U., Kalthoff, D., and Hoehn, M. (2010). High field BOLD response to forepaw stimulation in the mouse. *Neuroimage* 51, 704–712. doi: 10.1016/j.neuroimage.2010.02.083
- Agarwal, S., Sane, R., Oberoi, R., Ohlfest, J. R., and Elmquist, W. F. (2011). Delivery of molecularly targeted therapy to malignant glioma, a disease of the whole brain. *Expert Rev. Mol. Med.* 13:e17. doi: 10.1017/S1462399411001888
- Ahrens, E. T., Flores, R., Xu, H., and Morel, P. A. (2005). *In vivo* imaging platform for tracking immunotherapeutic cells. *Nat. Biotechnol.* 23, 983–987. doi: 10.1038/nbt1121
- Alonso-Ortiz, E., Levesque, I. R., and Pike, G. B. (2015). MRI-based myelin water imaging: a technical review. *Magn. Reson. Med.* 73, 70–81. doi: 10.1002/mrm.25198
- Arakelyan, K., Cantow, K., Hentschel, J., Flemming, B., Pohlmann, A., Ladwig, M., et al. (2013). Early effects of an x-ray contrast medium on renal  $T(2)$   $^*/T(2)$  MRI as compared to short-term hyperoxia, hypoxia and aortic occlusion in rats. *Acta Physiol.* 208, 202–213. doi: 10.1111/apha.12094
- Bakermans, A. J., Abdurrachim, D., Moonen, R. P., Motaal, A. G., Prompers, J. J., Strijkers, G. J., et al. (2015). Small animal cardiovascular MR imaging and spectroscopy. *Prog. Nucl. Magn. Reson. Spectrosc.* 88–89, 1–47. doi: 10.1016/j.pnmrs.2015.03.001
- Baltes, C., Bosshard, S., Mueggler, T., Ratering, D., and Rudin, M. (2011). Increased blood oxygen level-dependent (BOLD) sensitivity in the mouse somatosensory cortex during electrical forepaw stimulation using a cryogenic radiofrequency probe. *NMR Biomed.* 24, 439–446. doi: 10.1002/nbm.1613

- Baltes, C., Radzwill, N., Bosshard, S., Marek, D., and Rudin, M. (2009). Micro MRI of the mouse brain using a novel 400 MHz cryogenic quadrature RF probe. *NMR Biomed.* 22, 834–842. doi: 10.1002/nbm.1396
- Barkhof, F., Filippi, M., Miller, D. H., Scheltens, P., Campi, A., Polman, C. H., et al. (1997). Comparison of MRI criteria at first presentation to predict conversion to clinically definite multiple sclerosis. *Brain* 120, 2059–2069. doi: 10.1093/brain/120.11.2059
- Baxter, A. G. (2007). The origin and application of experimental autoimmune encephalomyelitis. *Nat. Rev. Immunol.* 7, 904–912. doi: 10.1038/nri2190
- Beaumont, M., Lemasson, B., Farion, R., Segebarth, C., Rémy, C., and Barbier, E. L. (2009). Characterization of tumor angiogenesis in rat brain using iron-based vessel size index MRI in combination with gadolinium-based dynamic contrast-enhanced MRI. *J. Cereb. Blood Flow Metab.* 29, 1714–1726. doi: 10.1038/jcbfm.2009.86
- Beckmann, N., Gérard, C., Abramowski, D., Cannet, C., and Staufenbiel, M. (2011). Noninvasive magnetic resonance imaging detection of cerebral amyloid angiopathy-related microvascular alterations using superparamagnetic iron oxide particles in APP transgenic mouse models of Alzheimer's disease: application to passive A $\beta$  immunotherapy. *J. Neurosci.* 31, 1023–1031. doi: 10.1523/JNEUROSCI.4936-10.2011
- Beckmann, N., Schuler, A., Mueggler, T., Meyer, E. P., Wiederhold, K.-H., Staufenbiel, M., et al. (2003). Age-dependent cerebrovascular abnormalities and blood flow disturbances in APP23 mice modeling Alzheimer's disease. *J. Neurosci.* 23, 8453–8459. doi: 10.5167/uzh-182
- Beeman, S. C., Zhang, M., Gubhaju, L., Wu, T., Bertram, J. F., Frakes, D. H., et al. (2011). Measuring glomerular number and size in perfused kidneys using MRI. *Am. J. Physiol. Renal Physiol.* 300, F1454–F1457. doi: 10.1152/ajprenal.00044.2011
- Ben Nun, A., Wekerle, H., and Cohen, I. R. (1981). Vaccination against autoimmune encephalomyelitis with T-lymphocyte line cells reactive against myelin basic protein. *Nature* 292, 60–61. doi: 10.1038/292060a0
- Biffi, A., and Greenberg, S. M. (2011). Cerebral amyloid angiopathy: a systematic review. *J. Clin. Neurol.* 7, 1–9. doi: 10.3988/jcn.2011.7.1.1
- Black, R., Early, T., Roemer, P., Mueller, O., Mogro-Campero, A., Turner, L., et al. (1993). A high-temperature superconducting receiver for nuclear magnetic resonance microscopy. *Science* 259, 793–795. doi: 10.1126/science.8430331
- Bonventre, J. V., and Yang, L. (2011). Cellular pathophysiology of ischemic acute kidney injury. *J. Clin. Invest.* 121, 4210. doi: 10.1172/JCI45161
- Boretius, S., Schmelting, B., Watanabe, T., Merkler, D., Tammer, R., Czéh, B., et al. (2006). Monitoring of EAE onset and progression in the common marmoset monkey by sequential high-resolution 3D MRI. *NMR Biomed.* 19, 41–49. doi: 10.1002/nbm.999
- Borsook, D., and Becerra, L. (2011). CNS animal fMRI in pain and analgesia. *Neurosci. Biobehav. Rev.* 35, 1125–1143. doi: 10.1016/j.neubiorev.2010.11.005
- Bosshard, S. C., Baltes, C., Wyss, M. T., Mueggler, T., Weber, B., and Rudin, M. (2010). Assessment of brain responses to innocuous and noxious electrical forepaw stimulation in mice using BOLD fMRI. *Pain* 151, 655–663. doi: 10.1016/j.pain.2010.08.025
- Bosshard, S. C., Grandjean, J., Schroeter, A., Baltes, C., Zeilhofer, H. U., and Rudin, M. (2012). Hyperalgesia by low doses of the local anesthetic lidocaine involves cannabinoid signaling: an fMRI study in mice. *Pain* 153, 1450–1458. doi: 10.1016/j.pain.2012.04.001
- Bosshard, S. C., Stuker, F., Von Deuster, C., Schroeter, A., and Rudin, M. (2015). BOLD fMRI of C-fiber mediated nociceptive processing in mouse brain in response to thermal stimulation of the forepaws. *PLoS ONE* 10:e0126513. doi: 10.1371/journal.pone.0126513
- Bovens, S. M., Te Boekhorst, B., Ouden, K. D., Van De Kolk, K. W., Nauwerth, A., Nederhoff, M. G., et al. (2011). Evaluation of infarcted murine heart function: comparison of prospectively triggered with self-gated MRI. *NMR Biomed.* 24, 307–315. doi: 10.1002/nbm.1593
- Brück, W., Bitsch, A., Kolenda, H., Brück, Y., Stiefel, M., and Lassmann, H. (1997). Inflammatory central nervous system demyelination: correlation of magnetic resonance imaging findings with lesion pathology. *Ann. Neurol.* 42, 783–793. doi: 10.1002/ana.410420515
- Chawla, L. S., Eggers, P. W., Star, R. A., and Kimmel, P. L. (2014). Acute kidney injury and chronic kidney disease as interconnected syndromes. *N Engl. J. Med.* 371, 58–66. doi: 10.1056/NEJMra1214243
- Darrasse, L., and Ginefri, J.-C. (2003). Perspectives with cryogenic RF probes in biomedical MRI. *Biochimie* 85, 915–937. doi: 10.1016/j.biochi.2003.09.016
- Deboy, C. A., Zhang, J., Dike, S., Shats, L., Jones, M., Reich, D. S., et al. (2007). High resolution diffusion tensor imaging of axonal damage in focal inflammatory and demyelinating lesions in rat spinal cord. *Brain* 130, 2199–2210. doi: 10.1093/brain/awm122
- Dieringer, M. A., Renz, W., Lindel, T., Seifert, F., Frauenrath, T., Von Knobelsdorff-Brenkenhoff, F., et al. (2011). Design and application of a four-channel transmit/receive surface coil for functional cardiac imaging at 7T. *J. Magn. Reson. Imaging* 33, 736–741. doi: 10.1002/jmri.22451
- Dijkhuizen, R. M., Van Lookeren Campagne, M., Niendorf, T., Dreher, W., Van Der Toorn, A., Hoehn-Berlage, M., et al. (1996). Status of the neonatal rat brain after NMDA-induced excitotoxic injury as measured by MRI, MRS and metabolic imaging. *NMR Biomed.* 9, 84–92.
- Emblem, K. E., Mouridsen, K., Bjornerud, A., Farrar, C. T., Jennings, D., Borra, R. J. H., et al. (2013). Vessel architectural imaging identifies cancer patient responders to anti-angiogenic therapy. *Nat. Med.* 19, 1178–1183. doi: 10.1038/nm.3289
- Epstein, F. H. (2007). MR in mouse models of cardiac disease. *NMR Biomed.* 20, 238–255. doi: 10.1002/nbm.1152
- Figueiredo, G., Brockmann, C., Boll, H., Heilmann, M., Schambach, S. J., Fiebig, T., et al. (2012). Comparison of digital subtraction angiography, micro-computed tomography angiography and magnetic resonance angiography in the assessment of the cerebrovascular system in live mice. *Clin. Neuroradiol.* 22, 21–28. doi: 10.1007/s00062-011-0113-2
- Ginefri, J. C., Poirier-Quinot, M., Girard, O., and Darrasse, L. (2007). Technical aspects: development, manufacture and installation of a cryo-cooled HTS coil system for high-resolution *in-vivo* imaging of the mouse at 1.5 T. *Methods* 43, 54–67. doi: 10.1016/j.ymeth.2007.03.011
- Graessl, A., Renz, W., Hezel, F., Dieringer, M. A., Winter, L., Oezerdem, C., et al. (2014). Modular 32-channel transceiver coil array for cardiac MRI at 7.0T. *Magn. Reson. Med.* 72, 276–290. doi: 10.1002/mrm.24903
- Grams, M. E., and Rabb, H. (2012). The distant organ effects of acute kidney injury. *Kidney Int.* 81, 942–948. doi: 10.1038/ki.2011.241
- Hart, B. A., Bauer, J., Muller, H. J., Melchers, B., Nicolay, K., Brok, H., et al. (1998). Histopathological characterization of magnetic resonance imaging-detectable brain white matter lesions in a primate model of multiple sclerosis: a correlative study in the experimental autoimmune encephalomyelitis model in common marmosets (*Callithrix jacchus*). *Am. J. Pathol.* 153, 649–663. doi: 10.1016/S0002-9440(10)65606-4
- Heidenreich, P. A., Trogdon, J. G., Khavjou, O. A., Butler, J., Dracup, K., Ezekowitz, M. D., et al. (2011). Forecasting the future of cardiovascular disease in the United States a policy statement from the American heart association. *Circulation* 123, 933–944. doi: 10.1161/CIR.0b013e31820a55f5
- Heijman, E., Aben, J. P., Penners, C., Niessen, P., Guillaume, R., Van Eys, G., et al. (2008). Evaluation of manual and automatic segmentation of the mouse heart from CINE MR images. *J. Magn. Reson. Imaging* 27, 86–93. doi: 10.1002/jmri.21236
- Heijman, E., De Graaf, W., Niessen, P., Nauwerth, A., Van Eys, G., De Graaf, L., et al. (2007). Comparison between prospective and retrospective triggering for mouse cardiac MRI. *NMR Biomed.* 20, 439–447. doi: 10.1002/nbm.1110
- Heilmann, M., Neudecker, S., Wolf, I., Gubhaju, L., Sticht, C., Schock-Kusch, D., et al. (2012). Quantification of glomerular number and size distribution in normal rat kidneys using magnetic resonance imaging. *Nephrol. Dial. Transplant.* 27, 100–107. doi: 10.1093/ndt/gfr273
- Hiba, B., Richard, N., Thibault, H., and Janier, M. (2007). Cardiac and respiratory self-gated cine MRI in the mouse: comparison between radial and rectilinear techniques at 7T. *Magn. Reson. Med.* 58, 745–753. doi: 10.1002/mrm.21355
- Hoult, D. I., and Richards, R. (1976). The signal-to-noise ratio of the nuclear magnetic resonance experiment. *J. Magn. Reson.* 24, 71–85. doi: 10.1016/0022-2364(76)90233-X
- Hoult, D., and Lauterbur, P. C. (1979). The sensitivity of the zeugmatographic experiment involving human samples. *J. Magn. Reson.* 34, 425–433. doi: 10.1016/0022-2364(79)90019-2
- Hoy, W. E., Bertram, J. F., Denton, R. D., Zimanyi, M., Samuel, T., and Hughson, M. D. (2008). Nephron number, glomerular volume, renal

- disease and hypertension. *Curr. Opin. Nephrol. Hypertens.* 17, 258–265. doi: 10.1097/MNH.0b013e3282f9b1a5
- Hu, L., Chen, J., Yang, X., Senpan, A., Allen, J. S., Yanaba, N., et al. (2014). Assessing intrarenal nonperfusion and vascular leakage in acute kidney injury with multinuclear <sup>1</sup>H/<sup>19</sup>F MRI and perfluorocarbon nanoparticles. *Magn. Reson. Med.* 71, 2186–2196. doi: 10.1002/mrm.24851
- Huelnhagen, T., Pohlmann, A., Hezel, F., Peper, E., Ku, M.-C., and Niendorf, T. (2014). Detailing myocardial microstructure in the *ex vivo* rat heart using high isotropic spatial resolution susceptibility weighted MRI and quantitative susceptibility mapping. *Proc. Intl. Soc. Magn. Reson. Med.* 22, 2437. doi: 10.13140/RG.2.1.1358.1522
- Hurlston, S. E., Brey, W. W., Suddarth, S. A., and Johnson, G. A. (1999). A high-temperature superconducting Helmholtz probe for microscopy at 9.4 T. *Magn. Reson. Med.* 41, 1032–1038.
- Jain, R. K. (2005). Normalization of tumor vasculature: an emerging concept in antiangiogenic therapy. *Science* 307, 58–62. doi: 10.1126/science.1104819
- Ji, Y., Waiczies, H., Winter, L., Neumanova, P., Hofmann, D., Rieger, J., et al. (2015). Eight-channel transceiver RF coil array tailored for (1) H/(19) F MR of the human knee and fluorinated drugs at 7.0 T. *NMR Biomed.* 28, 726–737. doi: 10.1002/nbm.3300
- Junge, S. (2012). “Cryogenic and superconducting coils for MRI,” in *eMagRes - Encyclopedia of Magnetic Resonance*, eds J. T. Vaughan and J. R. Griffiths (Hoboken, NJ: John Wiley & Sons, Ltd.), 505–514. doi: 10.1002/9780470034590.emrstm1162
- Kim, S.-G., and Ogawa, S. (2012). Biophysical and physiological origins of blood oxygenation level-dependent fMRI signals. *J. Cereb. Blood Flow Metab.* 32, 1188–1206. doi: 10.1038/jcbfm.2012.23
- Kinsey, G. R., and Okusa, M. D. (2014). Expanding role of T cells in acute kidney injury. *Curr. Opin. Nephrol. Hypertens.* 23, 9. doi: 10.1097/01.mnh.0000436695.29173.de
- Klohs, J., Baltes, C., Prinz-Kranz, F., Ratering, D., Nitsch, R. M., Knuesel, I., et al. (2012). Contrast-enhanced magnetic resonance microangiography reveals remodeling of the cerebral microvasculature in transgenic ArcA $\beta$  mice. *J. Neurosci.* 32, 1705–1713. doi: 10.1523/JNEUROSCI.5626-11.2012
- Klohs, J., Rudin, M., Shimshek, D. R., and Beckmann, N. (2014). Imaging of cerebrovascular pathology in animal models of Alzheimer's disease. *Front. Aging Neurosci.* 6:32. doi: 10.3389/fnagi.2014.00032
- Knutsson, L., Ståhlberg, F., and Wirestam, R. (2010). Absolute quantification of perfusion using dynamic susceptibility contrast MRI: pitfalls and possibilities. *Magn. Reson. Mater. Physics Biol. Med.* 23, 1–21. doi: 10.1007/s10334-009-0190-2
- Kopp, C., Linz, P., Dahlmann, A., Hammon, M., Jantsch, J., Müller, D. N., et al. (2013). <sup>23</sup>Na magnetic resonance imaging-determined tissue sodium in healthy subjects and hypertensive patients. *Hypertension* 61, 635–640. doi: 10.1161/HYPERTENSIONAHA.111.00566
- Kopp, C., Linz, P., Wachsmuth, L., Dahlmann, A., Horbach, T., Schofl, C., et al. (2012). <sup>23</sup>Na magnetic resonance imaging of tissue sodium. *Hypertension* 59, 167–172. doi: 10.1161/HYPERTENSIONAHA.111.183517
- Kording, F., Weidensteiner, C., Zwick, S., Osterberg, N., Weyerbrock, A., Staszewski, O., et al. (2014). Simultaneous assessment of vessel size index, relative blood volume, and vessel permeability in a mouse brain tumor model using a combined spin echo gradient echo echo-planar imaging sequence and viable tumor analysis. *J. Magn. Reson. Imaging* 40, 1310–1318. doi: 10.1002/jmri.24513
- Kovacs, H., Moskau, D., and Spraul, M. (2005). Cryogenically cooled probes—a leap in NMR technology. *Prog. Nucl. Magn. Reson. Spectrosc.* 46, 131–155. doi: 10.1016/j.pnmrs.2005.03.001
- Ku, M.-C., Wolf, S. A., Respondek, D., Matyash, V., Pohlmann, A., Waiczies, S., et al. (2013). GDNF mediates glioblastoma-induced microglia attraction but not astrogliosis. *Acta Neuropathol.* 125, 609–620. doi: 10.1007/s00401-013-1079-8
- Kuchling, J., Ramien, C., Bozin, I., Dörr, J., Harms, L., Rosche, B., et al. (2014). Identical lesion morphology in primary progressive and relapsing-remitting MS -an ultrahigh field MRI study. *Mult. Scler.* 20, 1866–1871. doi: 10.1177/1352458514531084
- Kuhlmann, T., Lassmann, H., and Brück, W. (2008). Diagnosis of inflammatory demyelination in biopsy specimens: a practical approach. *Acta Neuropathol.* 115, 275–287. doi: 10.1007/s00401-007-0320-8
- Laisler, E., Poirier-Quinot, M., Lambert, S. A., Dubuisson, R. M., Girard, O. M., Moser, E., et al. (2015). *In vivo* MR imaging of the human skin at subnanoliter resolution using a superconducting surface coil at 1.5 Tesla. *J. Magn. Reson. Imaging* 41, 496–504. doi: 10.1002/jmri.24549
- Le Bihan, D., and Iima, M. (2015). Diffusion magnetic resonance imaging: what water tells us about biological tissues. *PLoS Biol.* 13:e1002203. doi: 10.1371/journal.pbio.1002246
- Lepore, S., Waiczies, H., Hentschel, J., Ji, Y., Skodowski, J., Pohlmann, A., et al. (2013). Enlargement of cerebral ventricles as an early indicator of encephalomyelitis. *PLoS ONE* 8:e72841. doi: 10.1371/journal.pone.0072841
- Lin, W., Abendschein, D. R., Celik, A., Dolan, R. P., Lauffer, R. B., Walovitch, R. C., et al. (1997). Intravascular contrast agent improves magnetic resonance angiography of carotid arteries in minipigs. *J. Magn. Reson. Imaging* 7, 963–971. doi: 10.1002/jmri.1880070605
- Linz, P., Santoro, D., Renz, W., Rieger, J., Ruehle, A., Ruff, J., et al. (2015). Skin sodium measured with (2)(3)Na MRI at 7.0 T. *NMR Biomed.* 28, 54–62. doi: 10.1002/nbm.3224
- McRobbie, D. W., Moore, E. A., Graves, M. J., and Prince, M. R. (2006). *MRI from Picture to Proton*. Cambridge: Cambridge University Press.
- Merlini, M., Meyer, E., Ulmann-Schuler, A., and Nitsch, R. (2011). Vascular  $\beta$ -amyloid and early astrocyte alterations impair cerebrovascular function and cerebral metabolism in transgenic arcA $\beta$  mice. *Acta Neuropathol.* 122, 293–311. doi: 10.1007/s00401-011-0834-y
- Messroghli, D. R., Niendorf, T., Schulz-Menger, J., Dietz, R., and Friedrich, M. G. (2003). T1 mapping in patients with acute myocardial infarction. *J. Cardiovasc. Magn. Reson.* 5, 353–359. doi: 10.1081/JCMR-120019418
- Milo, R., and Miller, A. (2014). Revised diagnostic criteria of multiple sclerosis. *Autoimmun. Rev.* 13, 518–524. doi: 10.1016/j.autrev.2014.01.012
- Molitoris, B. A. (2014). Therapeutic translation in acute kidney injury: the epithelial/endothelial axis. *J. Clin. Invest.* 124, 2355–2363. doi: 10.1172/JCI72269
- Mueggler, T., Pohl, H., Baltes, C., Riethmacher, D., Suter, U., and Rudin, M. (2012). MRI signature in a novel mouse model of genetically induced adult oligodendrocyte cell death. *Neuroimage* 59, 1028–1036. doi: 10.1016/j.neuroimage.2011.09.001
- Mueggler, T., Razoux, F., Russig, H., Buehler, A., Franklin, T. B., Baltes, C., et al. (2011). Mapping of CBV changes in 5-HT 1A terminal fields by functional MRI in the mouse brain. *Eur. Neuropsychopharmacol.* 21, 344–353. doi: 10.1016/j.euroneuro.2010.06.010
- Najac, C., Branzoli, F., Ronen, I., and Valette, J. (2014). Brain intracellular metabolites are freely diffusing along cell fibers in grey and white matter, as measured by diffusion-weighted MR spectroscopy in the human brain at 7 T. *Brain Struct. Funct.* doi: 10.1007/s00429-014-0968-5. [Epub ahead of print].
- Nasrallah, F. A., Tay, H.-C., and Chuang, K.-H. (2014). Detection of functional connectivity in the resting mouse brain. *Neuroimage* 86, 417–424. doi: 10.1016/j.neuroimage.2013.10.025
- Niendorf, T., Dijkhuizen, R. M., Norris, D. G., Van Lookeren Campagne, M., and Nicolay, K. (1996). Biexponential diffusion attenuation in various states of brain tissue: implications for diffusion-weighted imaging. *Magn. Reson. Med.* 36, 847–857. doi: 10.1002/mrm.1910360607
- Niendorf, T., Graessl, A., Thalhammer, C., Dieringer, M. A., Kraus, O., Santoro, D., et al. (2013). Progress and promises of human cardiac magnetic resonance at ultrahigh fields: a physics perspective. *J. Magn. Reson.* 229, 208–222. doi: 10.1016/j.jmri.2012.11.015
- Niendorf, T., Norris, D. G., and Leibfritz, D. (1994). Detection of apparent restricted diffusion in healthy rat brain at short diffusion times. *Magn. Reson. Med.* 32, 672–677. doi: 10.1002/mrm.1910320520
- Niendorf, T., Paul, K., Oezderem, C., Graessl, A., Klux, S., Huelnhagen, T., et al. (2015a). W(h)ither human cardiac and body magnetic resonance at ultrahigh fields? Technical advances, practical considerations, applications, and clinical opportunities. *NMR Biomed.* doi: 10.1002/nbm.3268. [Epub ahead of print].
- Niendorf, T., Pohlmann, A., Arakelyan, K., Flemming, B., Cantow, K., Hentschel, J., et al. (2015b). How bold is blood oxygenation level-dependent (BOLD) magnetic resonance imaging of the kidney? Opportunities, challenges and future directions. *Acta Physiol.* 213, 19–38. doi: 10.1111/apha.12393

- Niendorf, T., and Sodickson, D. (2006a). Acceleration of cardiovascular MRI using parallel imaging: basic principles, practical considerations, clinical applications and future directions. *Rofo* 178, 15–30. doi: 10.1055/s-2005-858686
- Niendorf, T., and Sodickson, D. K. (2006b). Parallel imaging in cardiovascular MRI: methods and applications. *NMR Biomed.* 19, 325–341. doi: 10.1002/nbm.1051
- Niendorf, T., and Sodickson, D. K. (2008). Highly accelerated cardiovascular MR imaging using many channel technology: concepts and clinical applications. *Eur. Radiol.* 18, 87–102. doi: 10.1007/s00330-007-0692-0
- Niendorf, T., Sodickson, D. K., Krombach, G. A., and Schulz-Menger, J. (2010). Toward cardiovascular MRI at 7 T: clinical needs, technical solutions and research promises. *Eur. Radiol.* 20, 2806–2816. doi: 10.1007/s00330-010-1902-8
- Norris, D. G., and Niendorf, T. (1995). Interpretation of DW-NMR data: dependence on experimental conditions. *NMR Biomed.* 8, 280–288. doi: 10.1002/nbm.1940080703
- Norris, D. G., Niendorf, T., and Leibfritz, D. (1994). Health and infarcted brain tissues studied at short diffusion times: the origins of apparent restriction and the reduction in apparent diffusion coefficient. *NMR Biomed.* 7, 304–310. doi: 10.1002/nbm.1940070703
- Nouls, J. C., Izenson, M. G., Greeley, H. P., and Johnson, G. A. (2008). Design of a superconducting volume coil for magnetic resonance microscopy of the mouse brain. *J. Magn. Reson.* 191, 231–238. doi: 10.1016/j.jmr.2007.12.018
- Paxinos, G., and Franklin, K. B. (2013). *Paxinos and Franklin's the Mouse Brain in Stereotaxic Coordinates*. Amsterdam: Elsevier.
- Peper, E., Huelnhagen, T., Pohlmann, A., Ku, M.-C., and Niendorf, T. (2015). Comparison of high resolution T2\* mapping and quantitative susceptibility mapping to investigate myocardial microstructure in the *ex vivo* rodent heart. *Proc. Intl. Soc. Mag. Reson. Med.* 23, 2609. doi: 10.13140/RG.2.1.1882.4400
- Pohl, H. B., Porcheri, C., Mueggler, T., Bachmann, L. C., Martino, G., Riethmacher, D., et al. (2011). Genetically induced adult oligodendrocyte cell death is associated with poor myelin clearance, reduced remyelination, and axonal damage. *J. Neurosci.* 31, 1069–1080. doi: 10.1523/JNEUROSCI.5035-10.2011
- Pohlmann, A., Arakelyan, K., Hentschel, J., Cantow, K., Flemming, B., Ladwig, M., et al. (2014a). Detailing the relation between renal T2\* and renal tissue pO2 using an integrated approach of parametric magnetic resonance imaging and invasive physiological measurements. *Invest. Radiol.* 49, 547–560. doi: 10.1097/RLI.0000000000000054
- Pohlmann, A., Cantow, K., Hentschel, J., Arakelyan, K., Ladwig, M., Flemming, B., et al. (2013a). Linking non-invasive parametric MRI with invasive physiological measurements (MR-PHYSIOL): towards a hybrid and integrated approach for investigation of acute kidney injury in rats. *Acta Physiol.* 207, 673–689. doi: 10.1111/apha.12065
- Pohlmann, A., Hentschel, J., Fechner, M., Hoff, U., Bubalo, G., Arakelyan, K., et al. (2013b). High temporal resolution parametric MRI monitoring of the initial ischemia/reperfusion phase in experimental acute kidney injury. *PLoS ONE* 8:e57411. doi: 10.1371/journal.pone.0057411
- Pohlmann, A., Schreiber, A., Ku, M.-C., Waiczies, H., Kox, S., Kettritz, R., et al. (2014b). Assessment of renal inflammatory cell infiltration in a murine ANCA-induced glomerulonephritis model by 19F-MRI. *Proc. Intl. Soc. Mag. Reson. Med.* 22, 2207. doi: 10.13140/RG.2.1.4143.4326
- Poirier-Quinot, M., Ginefri, J.-C., Girard, O., Robert, P., and Darrasse, L. (2008). Performance of a miniature high-temperature superconducting (HTS) surface coil for *in vivo* microimaging of the mouse in a standard 1.5T clinical whole-body scanner. *Magnetic Resonance in Medicine* 60, 917–927. doi: 10.1002/mrm.21605
- Pyatigorskaya, N., Le Bihan, D., Reynaud, O., and Ciobanu, L. (2014). Relationship between the diffusion time and the diffusion MRI signal observed at 17.2 Tesla in the healthy rat brain cortex. *Magn. Reson. Med.* 72, 492–500. doi: 10.1002/mrm.24921
- Ransohoff, R. M. (2009). Immunology: in the beginning. *Nature* 462, 41–42. doi: 10.1038/462041a
- Ransohoff, R. M. (2012). Animal models of multiple sclerosis: the good, the bad and the bottom line. *Nat. Neurosci.* 15, 1074–1077. doi: 10.1038/nn.3168
- Ratering, D., Baltes, C., Dörries, C., and Rudin, M. (2010). Accelerated cardiovascular magnetic resonance of the mouse heart using self-gated parallel imaging strategies does not compromise accuracy of structural and functional measures. *J. Cardiovasc. Magn. Reson.* 12:43. doi: 10.1186/1532-429X-12-43
- Ratering, D., Baltes, C., Nordmeyer—Massner, J., Marek, D., and Rudin, M. (2008). Performance of a 200-MHz cryogenic RF probe designed for MRI and MRS of the murine brain. *Magn. Reson. Med.* 59, 1440–1447. doi: 10.1002/mrm.21629
- Reinges, M. H. T., Nguyen, H. H., Krings, T., Hütter, B. O., Rohde, V., and Gilsbach, J. M. (2004). Course of brain shift during microsurgical resection of supratentorial cerebral lesions: limits of conventional neuronavigation. *Acta Neurochir.* 146, 369–377. doi: 10.1007/s00701-003-0204-1
- Ruiz-Cabello, J., Barnett, B. P., Bottomley, P. A., and Bulte, J. W. (2011). Fluorine (19F) MRS and MRI in biomedicine. *NMR Biomed.* 24, 114–129. doi: 10.1002/nbm.1570
- Sack, M., Wetterling, F., Sartorius, A., Ende, G., and Weber—Fahr, W. (2014). Signal-to-noise ratio of a mouse brain 13C CryoProbe™ system in comparison with room temperature coils: spectroscopic phantom and *in vivo* results. *NMR Biomed.* 27, 709–715. doi: 10.1002/nbm.3110
- Salat, D. H. (2014). Imaging small vessel-associated white matter changes in aging. *Neuroscience* 276, 174–186. doi: 10.1016/j.neuroscience.2013.11.041
- Schmierer, K., Scaravilli, F., Altmann, D. R., Barker, G. J., and Miller, D. H. (2004). Magnetization transfer ratio and myelin in postmortem multiple sclerosis brain. *Ann. Neurol.* 56, 407–415. doi: 10.1002/ana.20202
- Schneider, J. E. (2011). “Assessment of global cardiac function,” in *In Vivo NMR Imaging*, eds L. Schröder and C. Faber (Heidelberg: Springer), 387–405. doi: 10.1007/978-1-61779-219-9\_20
- Schneider, J. E., Lanz, T., Barnes, H., Stork, L. A., Bohl, S., Lygate, C. A., et al. (2011). Accelerated cardiac magnetic resonance imaging in the mouse using an eight-channel array at 9.4 Tesla. *Magn. Reson. Med.* 65, 60–70. doi: 10.1002/mrm.22605
- Schreiber, A., and Choi, M. (2015). The role of neutrophils in causing antineutrophil cytoplasmic autoantibody-associated vasculitis. *Curr. Opin. Hematol.* 22, 60–66. doi: 10.1097/MOH.0000000000000098
- Schreiber, A., Kettritz, R., Ku, M.-C., Waiczies, H., Waiczies, S., Niendorf, T., et al. (2013). 19F-MRI for noninvasive visualization of renal inflammation in a murine ANCA-induced glomerulonephritis model. *J. Am. Soc. Nephrol.* 24:308A. doi: 10.13140/RG.2.1.1265.9923
- Schroeter, A., Schlegel, F., Seuwen, A., Grandjean, J., and Rudin, M. (2014). Specificity of stimulus-evoked fMRI responses in the mouse: the influence of systemic physiological changes associated with innocuous stimulation under four different anesthetics. *Neuroimage* 94, 372–384. doi: 10.1016/j.neuroimage.2014.01.046
- Sinnecker, T., Bozin, I., Dörr, J., Pfueller, C. F., Harms, L., Niendorf, T., et al. (2013). Periventricular venous density in multiple sclerosis is inversely associated with T2 lesion count: a 7 Tesla MRI study. *Mult. Scler.* 19, 316–325. doi: 10.1177/1352458512451941
- Sinnecker, T., Dörr, J., Pfueller, C. F., Harms, L., Ruprecht, K., Jarius, S., et al. (2012a). Distinct lesion morphology at 7-T MRI differentiates neuromyelitis optica from multiple sclerosis. *Neurology* 79, 708–714. doi: 10.1212/WNL.0b013e3182648bc8
- Sinnecker, T., Mittelstaedt, P., Dörr, J., Pfueller, C. F., Harms, L., Niendorf, T., et al. (2012b). Multiple sclerosis lesions and irreversible brain tissue damage: a comparative ultrahigh-field strength magnetic resonance imaging study. *Arch. Neurol.* 69, 739–745. doi: 10.1001/archneurol.2011.2450
- Sosnovik, D. E., Dai, G., Nahrendorf, M., Rosen, B. R., and Seethamraju, R. (2007). Cardiac MRI in mice at 9.4 Tesla with a transmit—receive surface coil and a cardiac—tailored intensity—correction algorithm. *J. Magn. Reson. Imaging* 26, 279–287. doi: 10.1002/jmri.20966
- Steinman, L., and Zamvil, S. S. (2006). How to successfully apply animal studies in experimental allergic encephalomyelitis to research on multiple sclerosis. *Ann. Neurol.* 60, 12–21. doi: 10.1002/ana.20913
- Styles, P., Soffe, N. F., Scott, C. A., Cragg, D. A., Row, F., White, D. J., et al. (1984). A high-resolution NMR probe in which the coil and preamplifier are cooled with liquid helium. *J. Magn. Reson.* 213, 347–354. doi: 10.1016/j.jmr.2011.09.002
- Thalhammer, C., Renz, W., Winter, L., Hezel, F., Rieger, J., Pfeiffer, H., et al. (2012). Two-dimensional sixteen channel transmit/receive coil array for cardiac MRI at 7.0 T: design, evaluation, and application. *J. Magn. Reson. Imaging* 36, 847–857. doi: 10.1002/jmri.23724
- Traka, M., Arasi, K., Avila, R. L., Podojil, J. R., Christakos, A., Miller, S. D., et al. (2010). A genetic mouse model of adult-onset, pervasive central nervous system demyelination with robust remyelination. *Brain* 133, 3017–3029. doi: 10.1093/brain/awq247

- Utz, W., Greiser, A., Niendorf, T., Dietz, R., and Schulz-Menger, J. (2008). Single- or dual-bolus approach for the assessment of myocardial perfusion reserve in quantitative MR perfusion imaging. *Magn. Reson. Med.* 59, 1373–1377. doi: 10.1002/mrm.21611
- Utz, W., Niendorf, T., Wassmuth, R., Messroghli, D., Dietz, R., and Schulz-Menger, J. (2007). Contrast-dose relation in first-pass myocardial MR perfusion imaging. *J. Magn. Reson. Imaging* 25, 1131–1135. doi: 10.1002/jmri.20910
- Van Waesberghe, J., Kamphorst, W., De Groot, C. J., Van Walderveen, M. A., Castelijns, J. A., Ravid, R., et al. (1999). Axonal loss in multiple sclerosis lesions: magnetic resonance imaging insights into substrates of disability. *Ann. Neurol.* 46, 747–754.
- Vaughan, J. T., and Griffiths, J. R. (2012). *RF Coils for MRI*. Hoboken, NJ: John Wiley & Sons.
- Verhoye, M. R., Gravenmade, E. J., Raman, E. R., Van Reempts, J., and Van Der Linden, A. (1996). *In vivo* noninvasive determination of abnormal water diffusion in the rat brain studied in an animal model for multiple sclerosis by diffusion-weighted NMR imaging. *Magn. Reson. Imaging* 14, 521–532. doi: 10.1016/0730-725X(96)00047-1
- Vinnakota, K., Hu, F., Ku, M.-C., Georgieva, P. B., Szulzewsky, F., Pohlmann, A., et al. (2013). Toll-like receptor 2 mediates microglia/brain macrophage MT1-MMP expression and glioma expansion. *Neuro Oncol.* 15, 1457–1468. doi: 10.1093/neuonc/not115
- Von Knobelsdorff-Brenkenhoff, F., Frauenrath, T., Prothmann, M., Dieringer, M. A., Hezel, F., Renz, W., et al. (2010). Cardiac chamber quantification using magnetic resonance imaging at 7 Tesla—a pilot study. *Eur. Radiol.* 20, 2844–2852. doi: 10.1007/s00330-010-1888-2
- Von Knobelsdorff-Brenkenhoff, F., Tkachenko, V., Winter, L., Rieger, J., Thalhammer, C., Hezel, F., et al. (2013). Assessment of the right ventricle with cardiovascular magnetic resonance at 7 Tesla. *J. Cardiovasc. Magn. Reson.* 15:23. doi: 10.1186/1532-429x-15-23
- Wagenhaus, B., Pohlmann, A., Dieringer, M. A., Els, A., Waiczies, H., Waiczies, S., et al. (2012). Functional and morphological cardiac magnetic resonance imaging of mice using a cryogenic quadrature radiofrequency coil. *PLoS ONE* 7:e42383. doi: 10.1371/journal.pone.0042383
- Waiczies, H., Lepore, S., Drechsler, S., Qadri, F., Purfürst, B., Sydow, K., et al. (2013). Visualizing brain inflammation with a shingled-leg radio-frequency head probe for 19F/1H MRI. *Sci. Rep.* 3:1280. doi: 10.1038/srep01280
- Waiczies, H., Lepore, S., Janitzek, N., Hagen, U., Seifert, F., Ittermann, B., et al. (2011). Perfluorocarbon particle size influences magnetic resonance signal and immunological properties of dendritic cells. *PLoS ONE* 6:e21981. doi: 10.1371/journal.pone.0021981
- Waiczies, H., Millward, J. M., Lepore, S., Infante-Duarte, C., Pohlmann, A., Niendorf, T., et al. (2012). Identification of cellular infiltrates during early stages of brain inflammation with magnetic resonance microscopy. *PLoS ONE* 7:e32796. doi: 10.1371/journal.pone.0032796
- Welti, J., Loges, S., Dimmeler, S., and Carmeliet, P. (2013). Recent molecular discoveries in angiogenesis and antiangiogenic therapies in cancer. *J. Clin. Invest.* 123, 3190–3200. doi: 10.1172/JCI70212
- Winter, L., Kellman, P., Renz, W., Gräßl, A., Hezel, F., Thalhammer, C., et al. (2012). Comparison of three multichannel transmit/receive radiofrequency coil configurations for anatomic and functional cardiac MRI at 7.0T: implications for clinical imaging. *Eur. Radiol.* 22, 2211–2220. doi: 10.1007/s00330-012-2487-1
- Wolff, S. D., and Balaban, R. S. (1989). Magnetization transfer contrast (MTC) and tissue water proton relaxation *in vivo*. *Magn. Reson. Med.* 10, 135–144. doi: 10.1002/mrm.1910100113
- Wuerfel, J., Sinnecker, T., Ringelstein, E. B., Jarius, S., Schwindt, W., Niendorf, T., et al. (2012). Lesion morphology at 7 Tesla MRI differentiates Susac syndrome from multiple sclerosis. *Mult. Scler.* 18, 1592–1599. doi: 10.1177/1352458512441270
- Wuerfel, J., Tysiak, E., Prozorovski, T., Smyth, M., Mueller, S., Schnorr, J., et al. (2007). Mouse model mimics multiple sclerosis in the clinico-radiological paradox. *Eur. J. Neurosci.* 26, 190–198. doi: 10.1111/j.1460-9568.2007.05644.x
- Xie, L., Cianciolo, R. E., Hulette, B., Lee, H. W., Qi, Y., Cofer, G., et al. (2012). Magnetic resonance histology of age-related nephropathy in the Sprague Dawley rat. *Toxicol. Pathol.* 40, 764–778. doi: 10.1177/0192623312441408
- Xie, L., Subashi, E., Qi, Y., Knepper, M. A., and Johnson, G. A. (2014). Four-dimensional MRI of renal function in the developing mouse. *NMR Biomed.* 27, 1094–1102. doi: 10.1002/nbm.3162
- Young, A. A., Barnes, H., Davison, D., Neubauer, S., and Schneider, J. E. (2009). Fast left ventricular mass and volume assessment in mice with three-dimensional guide-point modeling. *J. Magn. Reson. Imaging* 30, 514–520. doi: 10.1002/jmri.21873
- Zwick, S., Strecker, R., Kiselev, V., Gall, P., Huppert, J., Palmowski, M., et al. (2009). Assessment of vascular remodeling under antiangiogenic therapy using DCE-MRI and vessel size imaging. *J. Magn. Reson. Imaging* 29, 1125–1133. doi: 10.1002/jmri.21710

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