



Enhanced Late Na and Ca Currents as Effective Antiarrhythmic Drug Targets

Hrayr S. Karagueuzian^{1,2*}, Arash Pezhouman^{1,2}, Marina Angelini³ and Riccardo Olcese^{2,3,4}

¹ From the Translational Arrhythmia Section, David Geffen School of Medicine, University of California, Los Angeles, Los Angeles, CA, USA, ² Cardiovascular Research Laboratory, Departments of Medicine (Cardiology), David Geffen School of Medicine, University of California, Los Angeles, Los Angeles, CA, USA, ³ Department of Anesthesiology and Perioperative Medicine, David Geffen School of Medicine, University of California, Los Angeles, Los Angeles, CA, USA, ⁴ Department of Physiology, David Geffen School of Medicine, University of California, Los Angeles, Los Angeles, CA, USA

OPEN ACCESS

Edited by:

Mohamed Chahine,
Laval University, Canada

Reviewed by:

Adrien Moreau,
Institut NeuroMyoGène-Ecole
Normale Supérieure de Lyon, France
Jordi Heijman,
Maastricht University, Netherlands

*Correspondence:

Hrayr S. Karagueuzian
hkaragueuzian@mednet.ucla.edu

Specialty section:

This article was submitted to
Pharmacology of Ion Channels and
Channelopathies,
a section of the journal
Frontiers in Pharmacology

Received: 14 November 2016

Accepted: 18 January 2017

Published: 06 February 2017

Citation:

Karagueuzian HS, Pezhouman A,
Angelini M and Olcese R (2017)
Enhanced Late Na and Ca Currents
as Effective Antiarrhythmic Drug
Targets. *Front. Pharmacol.* 8:36.
doi: 10.3389/fphar.2017.00036

While recent advances clarified the molecular and cellular modes of action of antiarrhythmic drugs (AADs), their link to suppression of dynamical arrhythmia mechanisms remains only partially understood. The current classifications of AADs (Classes I, III, and IV) rely on blocking peak Na, K and L-type calcium currents ($I_{Ca,L}$), with Class II with dominant beta receptor blocking activity and Class V including drugs with diverse classes of actions. The discovery that the calcium and redox sensor, cardiac Ca/calmodulin-dependent protein kinase II (CaMKII) enhances both the late Na (I_{Na-L}) and the late $I_{Ca,L}$ in patients at high risk of VT/VF provided a new and a rational AAD target. Pathological rise of either or both of I_{Na-L} and late $I_{Ca,L}$ are demonstrated to promote cellular early afterdepolarizations (EADs) and EAD-mediated triggered activity that can initiate VT/VF in remodeled hearts. Selective inhibition of the I_{Na-L} without affecting their peak transients with the highly specific prototype drug, GS-967 suppresses these EAD-mediated VT/VFs. As in the case of I_{Na-L} , selective inhibition of the late $I_{Ca,L}$ without affecting its peak with the prototype drug, roscovitine suppressed oxidative EAD-mediated VT/VF. These findings indicate that specific blockers of the late inward currents without affecting their peaks (gating modifiers), offer a new and effective AAD class action i.e., “Class VI.” The development of safe drugs with selective Class VI actions provides a rational and effective approach to treat VT/VF particularly in cardiac conditions associated with enhanced CaMKII activity such as heart failure.

Keywords: late Na current, late Ca current, ventricular fibrillation, early afterdepolarization, triggered activity, roscovitine, GS-458-967, antiarrhythmic drugs

A BRIEF HISTORICAL PERSPECTIVE OF ANTIARRHYTHMIC DRUG (AAD) THERAPY

It may seem ironic that “antiarrhythmic” drug therapy long antedated the electrocardiographic diagnosis of cardiac arrhythmias when the French physician Jean Baptist de Sénac first described in 1749 the benefits of quinine in patients with “palpitations,” an implicit reference to atrial fibrillation (AF).

“Of all the stomachic remedies the one whose effects appeared to be the most constant and the most prompt in many cases is quinine, mixed with a little rhubarb. Long and rebellious palpitations have ceded to this febrifuge seconded with a light purgative” (Willius and Keys, 1941).

The Paris-based physician, who perhaps was the first to describe in patients the correlation between palpitation (AF) and mitral valve disease, did extensive research on the cinchona extract that contains quinine/quinidine and the plant rhubarb that contains anthraquinones (laxatives). Sénac however, did not know if the fever reducing effect of quinine or the laxative effect of the anthraquinones or their combination were responsible to halt the presumed AF (Karagueuzian, 1995). Sénac's dilemma was apparently solved more than 150 years later thanks to a Dutch merchant who had discovered for himself the value of quinine against AF. The traveling merchant, who regularly took quinine to prevent malaria during his trips to the West Indies suffered from AF and was a patient of no less than the famed cardiologist Wenckebach, who recalled the experience of his patient this way:

"In 1912, a patient presented himself in my office wishing to get rid of his attacks of auricular fibrillation...On my telling him that I could promise nothing, he told me that he knew how to get rid of his attacks, as I did not believe him he promised to come back the next morning with a regular pulse, and he did...I was greatly struck by this fact, and afterward tried this sort of treatment on many cases of auricular fibrillation. My success was disappointing, in that quinine abolished auricular fibrillation in only a few cases, and these cases only when the onset of this form was quite recent, never when it was of several years' duration" (Moe, 1970).

Frey in 1918 reported that the d-isomer of quinine, quinidine, also found in cinchona bark, exerts more powerful effect against AF than the l-isomer quinine (Moe, 1970). The empiricism in AAD therapy continued through the thirties by the collaborating team of anesthesiologists and cardiac surgeons during open heart surgery. They discovered in 1936 that the application of the local anesthetic agent, procaine, either at the epicardial surface of the heart or by intravenous injections in patients during cardiac surgery provided "protection" against VT/VF by reducing "cardiac irritability" (Mautz, 1936). This success led to the ultimate development of the longer and more potent acting analog of procaine, procainamide with lesser central nervous system toxicity. Encouraged by the results of the local anesthetic procaine, intravenous lidocaine was introduced by Hitchcock and associates for the rapid conversion of VT to sinus rhythm in the late fifties (Hitchcock and Keown, 1959). An early attempt to a "mechanistic" approach to combat VT/VF was reported by Harris and Kokernot (1950). These authors suggested the presence of a similarity between the ectopic rapid ventricular discharges in dogs after coronary artery occlusion and the focal epileptogenic discharges in the brain of patients with epilepsy:

"...Drugs that have proved effective in preventing focal seizures might suppress ectopic ventricular discharges which

Abbreviations: APD, action potential duration; AAD, antiarrhythmic drug; AF, atrial fibrillation; CaMKII, calcium/calmodulin-dependent protein kinase II; CPVT, catecholaminergic polymorphic ventricular tachycardia; CCB, calcium channel blocker; EAD, early after depolarization; DAD, delayed afterdepolarization; ECC, excitation-contraction coupling; VT, ventricular tachycardia; VF, ventricular fibrillation; I_{Na-L} , late I_{Na} ; $I_{Ca,L}$, L-type Ca current; VW, Vaughan Williams; LQT, long QT syndrome; RyR2, ryanodine receptor; PLB, phospholamban; SERCA2a, sarcoplasmic-endoplasmic Ca-activated ATPase pump; I_{NCX} , Na-Ca exchanger current.

accompany acute myocardial infarction" (Harris and Kokernot, 1950).

Using dilantin (diphenylhydantoin, or DPH), a drug with demonstrated efficacy against epileptic seizures in humans, these authors recorded a measure of success in dogs with acute ischemic VT/VF by asserting that: "The excitability and automaticity of cardiac tissues are markedly similar to the changes produced in excitability and automaticity of nerve and skeletal muscle" (Harris and Kokernot, 1950). However, others have suggested that DPH's direct actions on cardiac Purkinje fibers (shortening of the APD, reducing spontaneous phase 4 depolarization and firing rate) "were sufficient to suggest an explanation for its anti-arrhythmic effect on ventricular arrhythmias in vivo" (Bigger et al., 1968). It is possible that both central and direct actions of DPH may conspire to bring about a measure of antiarrhythmic effect (Karagueuzian, 1995). Currently DPH has very limited use against VT/VF (Bäckman et al., 1989).

AAD CLASS ACTIONS: FROM HUGGINS TO SINGH-VAUGHAN WILLIAMS TO HARRISON TO SICILIAN GAMBIT AND BACK

The first classification of AADs was made in 1949 by Huggins and associates (Huggins et al., 1949). These authors grouped the drugs into three categories based on the drugs' (1) local anesthetic effect; (2) adrenergic potency and (3) coronary artery relaxing effects (Huggins et al., 1949). This first ever AAD classification, surprisingly not acknowledged in any of the subsequent AAD classifications, was the origin of all subsequent classifications. These pioneering authors tested the efficacy of what they called "three groups" of drugs against epinephrine-induced ventricular fibrillation (VF) in chloroform anesthetized dogs (Huggins et al., 1949). The "first group of agents" included the coronary vasodilators (i.e., sodium nitrite, aminophylline, papaverine, and quinacrine); "the second group" included drugs that decreased myocardial excitability (local anesthetic action) such as procaine, quinidine sulfate; and the "third group of agents" manifested sympatholytic activity (i.e., beta blocking effect) such as prisolol and dibenamine (Huggins et al., 1949). When the vasodilator group was found ineffective it was dropped from the list leaving (1) the myocardial excitability depressants and (2) the beta adrenergic receptor blockers.

With the introduction in the late 1940s of cellular transmembrane action potential recordings with the glass microelectrode technique from atrium and ventricle, Singh and Vaughan Williams introduced a "third class" of AAD action in 1970 when they discovered that d-sotalol's antiarrhythmic efficacy was independent of beta blocking effect. Instead they attributed d-sotalol's antiarrhythmic efficacy to the drug's ability to prolong cardiac cellular action potential duration (APD), thus proposing a "third class" of AAD action: prolongation of the APD (Singh and Vaughan Williams, 1970a). This proposal was based on two additional observations. First, on shortened APD during atrial fibrillation (AF) initiated by thyrotoxicosis

and resolution of the AF with thyroidectomy associated with prolonged APD (Vaughan Williams, 1970). Second, based on the effects of amiodarone, initially introduced as an antianginal agent (Charlier et al., 1968) and later as an AAD (Charlier et al., 1969) by prolonging atrial and ventricular APD (Singh and Vaughan Williams, 1970a,b). The similarities of d-sotalol and amiodarone to thyroidectomy in suppressing arrhythmias by prolonging the APD led these authors to conclude that the “prolongation of the APD may be considered as a third type of action reducing the probability of the arrhythmia” (Vaughan Williams, 1970). However, an apparent dilemma was created when it was discovered that both quinidine and procainamide, two Class I myocardial depressant drugs, also prolong the APD similar to Class III drugs like d-sotalol and amiodarone. These authors however, asserted that the myocardial depressant effect of excitability caused by Class I AADs emerges *prior* to the APD prolongation, therefore attributing quinidine’s antiarrhythmic efficacy solely to its myocardial depressant effect of excitability rather than to its APD prolonging effect (Szekeres and Williams, 1962; Singh and Vaughan Williams, 1970a). However, the later confirmation of significant directional differences of the effects of Class I AADs on cellular APD necessitated their separations into three “sub-classes” on the basis of their ability to prolong (Class 1A action), shorten (Class 1B action), or have no effect (Class 1C action) on the APD (Harrison, 1985; **Table 1**).

There are great deals of overlaps between drugs of different classes (Charlier, 1970) making rigid categorizations of the current AADs a difficult task. Moreover, the classification is based on the effects of AADs on cardiac action potential characteristics rather than on clinical arrhythmias. For example, Class 1C drug effect indicates “reduction of peak Na current (I_{Na}) and no change in APD” and says nothing about its therapeutic value. In short, the classification provides a useful conversational shorthand with no distinct clinical value. In fact, the shortcomings of the AAD classification was recognized back in 1970 by Singh and Vaughan Williams in their original proposal of Class III AAD action:

“The effect of anti-arrhythmic agents on various parameters of cardiac function in vitro and in vivo have made it possible to classify the drugs into groups which do or do not have certain clearly definable pharmacological actions though these actions do not necessarily determine anti-arrhythmic effectiveness” (Singh and Vaughan Williams, 1970a).

Class IV AADs The discovery of verapamil in the seventies, a Ca channel blocker (CCB) (Kohlhardt et al., 1972), that exerts potent AA efficacy against diverse experimental models of arrhythmias including myocardial ischemia, A–V nodal reentrant tachycardia and digitalis toxicity necessitated the introduction of CCB as a fourth class (Class IV) of AAD action (Kohlhardt et al., 1972; Rosen et al., 1975; **Table 1**), replacing the “centrally acting” Class IV AADs (i.e., Dilantin) originally proposed by Vaughan Williams (1970). Unfortunately, the concentrations of Class IV AADs that suppress arrhythmias cause severe negative inotropic effects that greatly diminish their therapeutic value, particularly in patients with compromised cardiac function (Rosen et al., 1975).

TABLE 1 | Updated classification of antiarrhythmic drugs.

Class I		
IA	Peak I_{Na+} current blockers	Quinidine, Procainamide, Disopyramide, Cibenzoline
IB		Lidocaine, Tocainide, Mexiletine, Ethmozine, Ranolazine*
IC		Encainide, Lorcinide, Flecainide, Propafenone
Class II		
	Beta blockers	Propranolol, Esmolol, Timolol, Atenolol, Metoprolol, Carvedilol
Class III		
	K^+ channel blockers	Amiodarone, d-Sotalol, Ibutilide, Dofetilide, Dronaderone
Class IV		
	Ca^{2+} channel blocker	Verapamil, Nifedipine, Diltiazem, Bepridil
Class V		
	I_f blocker, and other agents	Ivabradine, Adenosine, Digitalis, Magnesium Sulfate
Class VI		
	Late I_{Na} and late $I_{Ca,L}$ blockers	GS-458967 Roscovitine

* Given the structural similarity to lidocaine, ranolazine was initially grouped as Class 1B. However, depending on the relative contributions of ranolazine’s block of IKr vs. $INa-L$ the APD may prolong (Class 1A), shorten (Class 1B) or remain unchanged (Class 1C).

Class V AADs

Class V agents include molecules with diverse pharmacological actions, including digitalis, adenosine, ivabradine, and magnesium chloride. Some agents in this Class like vernakalant, which blocks peak Na current and is effective against AF (European Heart Rhythm et al., 2010), can be placed in several classes, while others like ivabradine (sinus node pacemaker I_f blocker), cannot be listed under any of the four classes (Thireau et al., 2011). Same goes for digitalis that increases vagal activity via its action on the central nervous system, thus decreasing the conduction of electrical impulses through the AV node (van Veldhuisen et al., 1996). As for adenosine, its actions are, for the most part, due to the direct activation of the G protein-gated inward rectifying potassium channels and antagonist of cAMP-stimulated ion currents such as the pacemaker current and the L type calcium current ($I_{Ca,L}$) (Lerman and Belardinelli, 1991; Shryock and Belardinelli, 1997). The cardiac actions of adenosine include slowing of sinus rate, depression of atrioventricular (AV) nodal conduction, suppressing AV nodal reentrant tachycardia (Belhassen et al., 1988; Camm and Garratt, 1994; Prystowsky et al., 2015). Finally, $MgCl_2$ is found effective against EAD-mediated triggered activity in canine Purkinje fibers (Bailie et al., 1988) and is suggested as a potential adjunctive therapy for cardiac arrhythmias in humans (Baker, 2016).

THE SICILIAN GAMBIT “CLASSIFICATION”

The dissatisfaction with the current AAD classification systems and the disappointing results with two major clinical trials, the Cardiac Arrhythmia Suppression Trial (CAST) testing the efficacy of Class I AADs (Echt et al., 1991) and the Survival With Oral D-Sotalol (SWORD) clinical trial testing the efficacy of Class III drug (Waldo et al., 1995) led a group of established cardiac

electrophysiologists to propose supplementing the current AAD class actions by incorporating the broad pharmacodynamic actions of each AAD (Task force of the working group on arrhythmias of the European Society on C, 1991). While informative, this new approach, known as the Sicilian Gambit, was argued to be “essentially similar” to the original classification (Vaughan Williams, 1992). Indeed, and two decades later members of the Group of the Sicilian Gambit Investigators, had this to say:

“The Sicilian Gambit... was not meant in aggregating drugs into categories, instead it was intended to provide background information so to challenge thought and investigation rather than to resolve issues” (Rosen and Janse, 2010).

The AAD therapy remains largely empirical. The link between specific ionic effect(s) of a drug and suppression of a specific arrhythmia mechanism(s) remain(s) incompletely understood (Weiss et al., 2015). All AADs block with differing degrees of potencies, multiple ionic currents that prevent precise understanding as to which specific drug-induced ionic change(s) in the heart is or are responsible for the outcome. The lack of mechanistic understanding of the success or failure with AAD therapy, greatly diminished the enthusiasm in innovative AAD research. At the present there are important gaps in our understanding of the ionic mechanism(s) of the arrhythmia sought for therapy. As articulated by van Hamel:

“Manifestations of an arrhythmogenic substrate and its triggers can be almost completely suppressed for a while -in clinical terms defined as a therapeutic success and therefore satisfying- without understanding in detail its mechanisms, anatomical extent, natural course and potential complications. This attitude toward arrhythmia management offers only short-term solutions and still worse, suppresses our intellectual curiosity” (van Hamel, 2003).

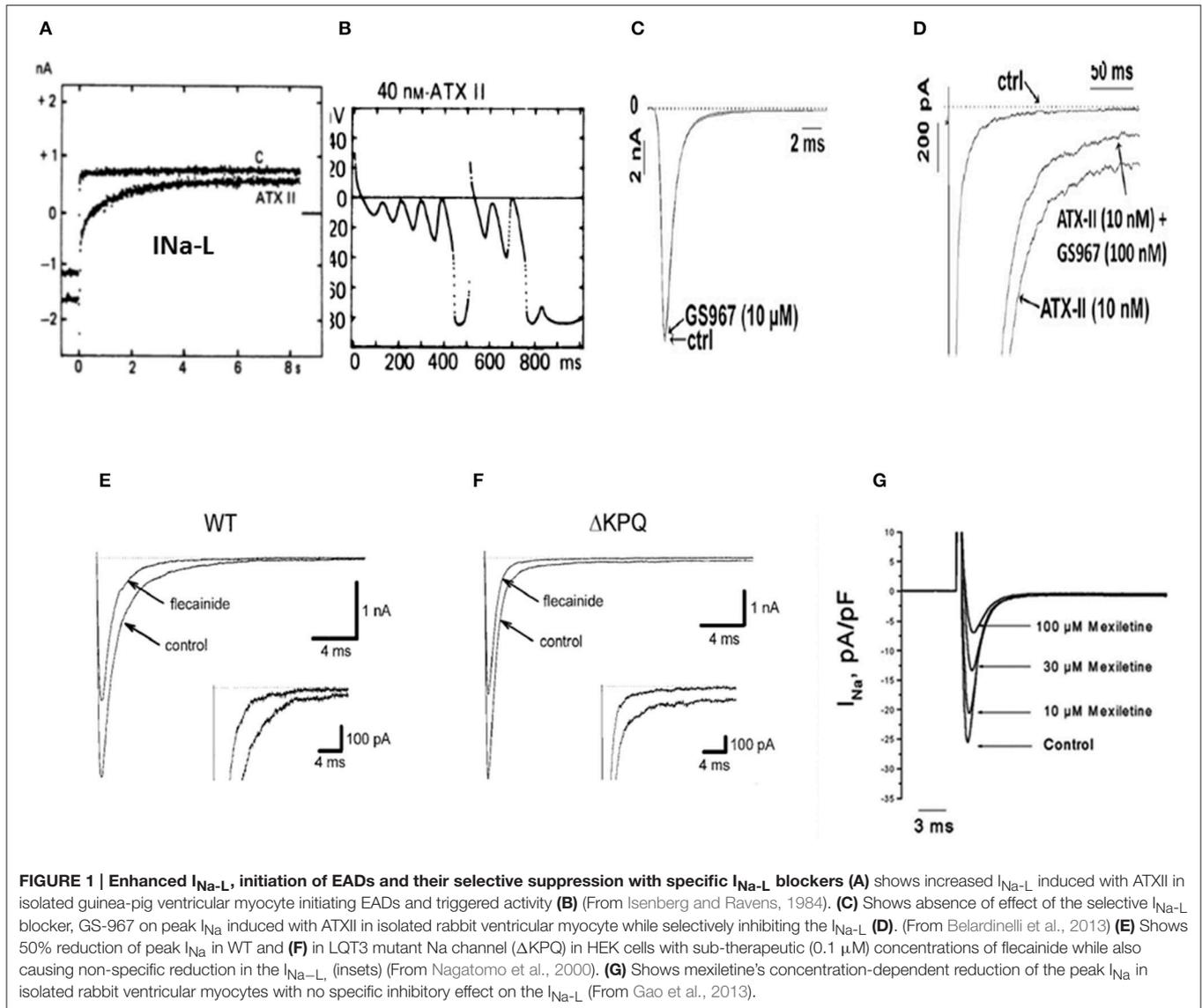
Indeed, it is clear that better understanding of the molecular pathological mechanisms of the arrhythmia would result in a better and more effective management of the arrhythmia because of possibility of specific drug targeting of the pathological mechanisms of the arrhythmia. Unfortunately, at the present there remain important gaps concerning the comprehension of these mechanisms preventing a rational and specifically targeted drug therapy of these arrhythmias.

The recent recognition of the pathological increases in the late Na current (I_{Na-L}) and $I_{Ca,L}$ as “vulnerable parameters” for early afterdepolarizations (EADs)-mediated VT/VF provides an opportunity to specifically target the dynamical arrhythmia mechanism. Importantly, the suppression of these VT/VFs with specific prototype blockers of I_{Na-L} and the late $I_{Ca,L}$ without affecting their peaks, provide a rational and effective AAD therapy that specifically target the arrhythmia mechanism. Under normal conditions, voltage-gated sodium channels (NaV1.5) open suddenly upon depolarization permitting entry of Na^+ that peaks in ≈ 1 ms (I_{Na}) then rapidly decays to baseline as most Na^+ channels quickly close (inactivate) during the plateau phase of the action potential (AP). While a very small

persistent I_{Na} current may be present under normal conditions, however, under diseased conditions I_{Na} inactivation can be greatly slowed, independent of its peak, providing persistent late Na current during the plateau phase of the AP three to tenfold over the level of normal conditions (Belardinelli et al., 2015). It is difficult to have a precise idea in absolute terms the level at which I_{Na-L} increases as this level is profoundly affected by diverse factors including differences in the clamp protocol, species use, age (neonatal vs. adult), temperature (room vs. body temperature), nature of the stress (i.e., heart failure, ATX, hydrogen peroxide, toxins like aconitine). Importantly the level of the I_{Na-L} depends on the timing when the “late” I_{Na} is measured, 50 or 100 ms after the clamp pulse or during the steady state (Ward and Giles, 1997; Song et al., 2006; Maltsev and Undrovinas, 2008; Undrovinas and Maltsev, 2008; Antzelevitch et al., 2014; Belardinelli et al., 2015; Wagner et al., 2015). It is estimated that during the steady-state pathological increases of the I_{Na-L} can reach up to 20-fold (roughly about 100 pA) of control as shown in **Figure 1D**. Similarly as in the case of the I_{Na} , the $I_{Ca,L}$ inactivation rate can also be considerably slowed under diverse cardiac conditions secondary to CaMKII activation generating an enhanced late inward depolarizing current. This $I_{Ca,L}$ component, particularly relevant during the plateau and the repolarization phases of the cardiac AP, became known as the late $I_{Ca,L}$ (Madhvani et al., 2015; Markandeya and Kamp, 2015). Under normal conditions late $I_{Ca,L}$ is minimal however, under pathological conditions its amplitude increases significantly. As in the case of the I_{Na-L} the absolute values of the late I_{Ca-L} varies greatly due to the same factors mentioned for I_{Na-L} plus the rundown phenomenon that is associated with the LCCs (Yue et al., 1990; Xie et al., 2009; Madhvani et al., 2011, 2015; Kim et al., 2016). **Figures 2A,B** shows variations in the absolute amplitude of the late I_{Ca-L} depending on the conditions of the study. Diverse acquired and congenital cardiac conditions at risk of developing VT/VF including heart failure, myocardial infarction and cardiomyopathies manifest pathological rises in these late inward currents, often exacerbated by activation of CaMKII signaling (Maltsev et al., 2007; Mollova et al., 2015). In the ensuing sections, animal and human models of VT/VF in which CaMKII activation plays a key role are presented. Electrophysiological studies scaling from isolated myocytes to intact hearts show that downstream increases in the I_{Na-L} and late $I_{Ca,L}$ play key roles in CaMKII-driven VT/VF providing a rational and effective AAD class action not considered in previous AAD classifications.

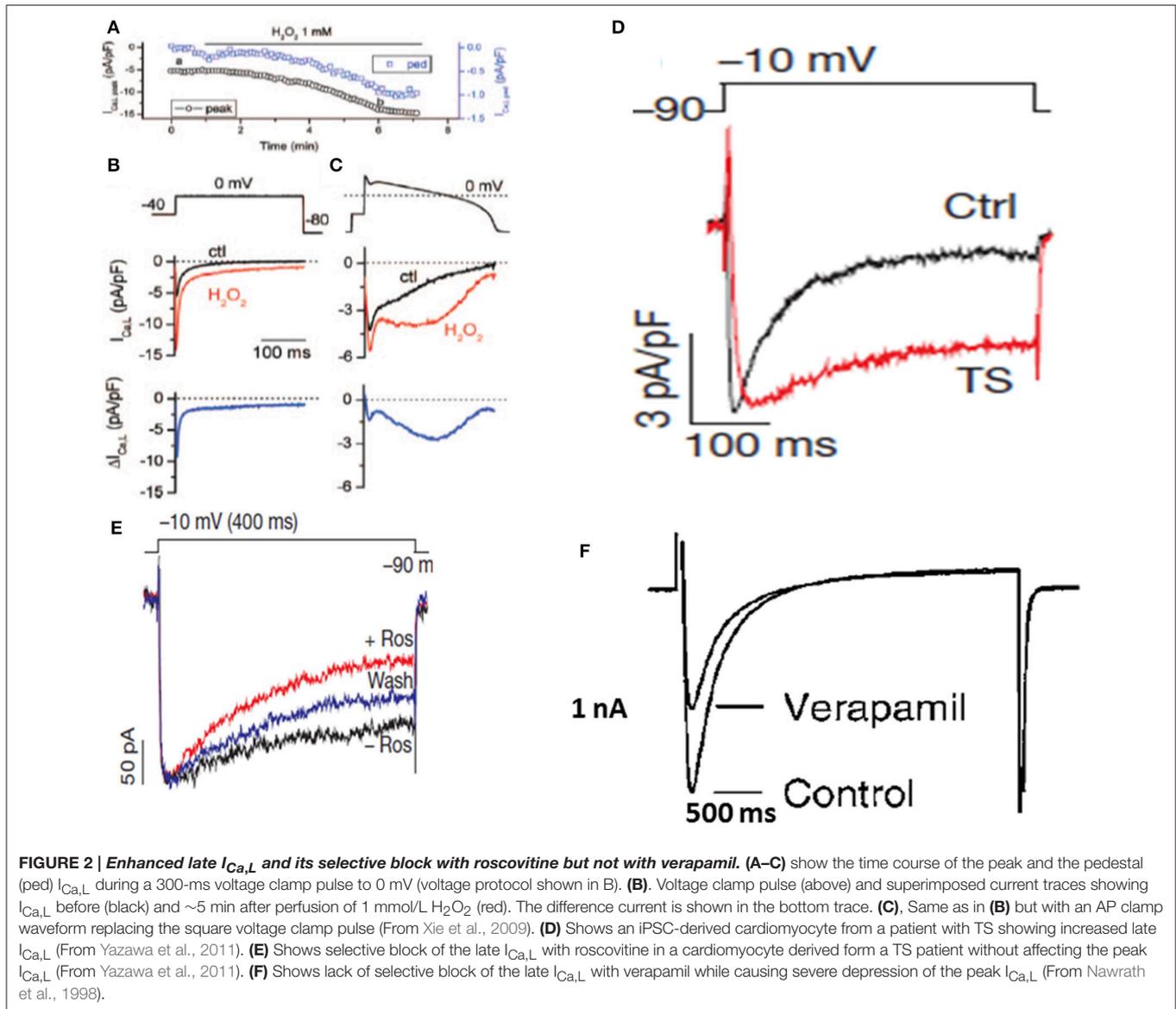
ENHANCEMENT OF I_{Na-L} AND LATE $I_{Ca,L}$ VIA CAMKII SIGNALING

Evidence accumulated over the last two decades has provided novel mechanistic insights into the ionic mechanisms of triggered VT/VF that can be specifically targeted by small drug molecules. The multifunctional and ubiquitous enzyme CaMKII is a well demonstrated sensor of calcium and redox signaling that enhances the I_{Na-L} and late $I_{Ca,L}$ and plays a key role in the pathogenesis of heart failure and arrhythmias both in



humans and in animal disease models (Erickson et al., 2008; Xie et al., 2009; Belardinelli et al., 2015; Hund and Mohler, 2015; Pezhouman et al., 2015; Mustroph et al., 2016). Patients with heart failure, hypertrophy and cardiomyopathies manifest sustained hyperactivity of CaMKII (Erickson et al., 2008, 2011; Mollova et al., 2015) promoting VT/VF (Swaminathan et al., 2012; Luczak and Anderson, 2014). Isolated myocytes studies have shown that CaMKII activation enhances the I_{Na-L} and the late $I_{Ca,L}$ (Xie et al., 2009; Belardinelli et al., 2015) promoting EADs and triggered arrhythmias (Morotti et al., 2014; Foteinou et al., 2015; Pezhouman et al., 2015). Consistent with these findings, transgenic CaMKII overexpression in the mouse leads to development of heart failure, while CaMKII deletion prevents onset of heart failure following transaortic constriction (Maier et al., 2003; Zhang et al., 2003; Backs et al., 2009; Ling et al., 2009). CaMKII-based therapy of cardiac arrhythmias however, is inherently difficult due to its vast signaling network requiring

a careful balance between the therapeutic benefit and the potential off-target effects (e.g., neuronal or metabolic; Hund and Mohler, 2015). An alternative would be specific downstream targets of the CaMKII pathway known to play key role in promoting arrhythmias. The I_{Na-L} and the late $I_{Ca,L}$ are two demonstrated down-stream targets that play a key role in EAD-mediated arrhythmias (Morita et al., 2011a; Madhvari et al., 2015). Indeed, dynamic-clamp and simulation studies have shown that selective enhancement of the late $I_{Ca,L}$ or the I_{Na-L} without changing any other ionic currents is sufficient to promote EADs and triggered activity (Madhvari et al., 2011, 2015). These findings indicate that the other downstream effects of CaMKII activation including, phosphorylation of ryanodine receptor, (RyR2), phospholamban (PLB), sarco/plasmic-endoplasmic Ca-activated ATPase type 2a (SERCA2a), and increased Na-Ca exchanger current (I_{NCX}) (Marx et al., 2000; Hund and Mohler, 2015; Mustroph et al., 2016), while potentially modulating the



genesis of EADs do not appear necessary for oxidative EAD formation in isolated ventricular myocytes (Madhvani et al., 2011, 2015) and EAD-mediated VT/VF in intact hearts (Morita et al., 2011a; Pezhouman et al., 2016). The Ca handling proteins however, could play an important role both in atrial and ventricular tissue in promoting DAD-mediated fibrillation as shown in human and animal studies (Voigt et al., 2012; Luczak and Anderson, 2014).

The Late Na Current (I_{Na-L})

The pathological rise of the late Na current known as late I_{Na} (I_{Na-L}), emerges under cardiac conditions associated with increased risk of developing VT/VF (Belardinelli et al., 2015) secondary to the activation of CaMKII signaling pathway (Erickson et al., 2011) and mutations of Nav1.5 channels (Bennett et al., 1995; Ulbricht, 2005). The cardiac conditions associated with increased risk of VT/VF include human congenital

LQT3 (Bennett et al., 1995; Ulbricht, 2005) and laminopathy (Markandeya et al., 2016) and a host of acquired chronic cardiac diseases including heart failure (Maltsev et al., 2007), myocardial ischemia (Belardinelli et al., 2013), increased pro-oxidant states (Ward and Giles, 1997; Song et al., 2006; Xie et al., 2009), and hypertrophic cardiomyopathy (Coppini et al., 2013; Belardinelli et al., 2015). It however must be emphasized that in the congenital LQT3, the I_{Na-L} is not necessarily CaMKII-dependent. The importance of pathological rise of the I_{Na-L} in the genesis of EADs and EAD-mediated arrhythmia has been systematically studied in isolated cardiac myocytes using potent toxins like sea anemone toxin II (ATX-II) that specifically enhance the I_{Na-L} with minimal effect on other ionic currents (Isenberg and Ravens, 1984; Belardinelli et al., 2015; **Figures 1A,B**). The key role played by the selective increase of the I_{Na-L} current in EAD-mediated arrhythmias is demonstrated using the highly selective and potent (IC₅₀ of 134 nM) blocker of the I_{Na-L} ,

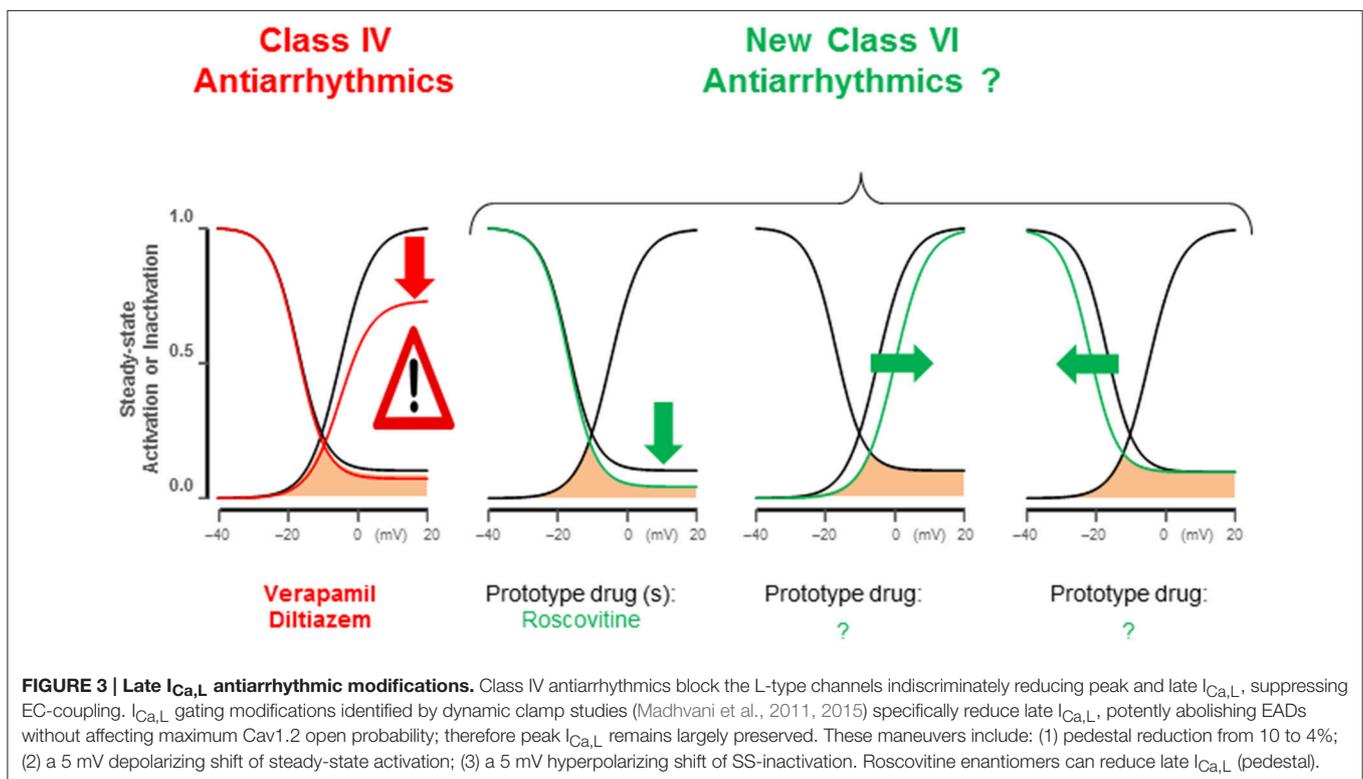
GS-458967 (GS-967) (Belardinelli et al., 2013) in suppressing I_{Na-L} as shown in **Figures 1C,D**). This selective effect of GS-967 on I_{Na-L} is fundamentally different from the traditional Class 1 AADs like flecainide and mexiletine that potently block the peak I_{Na} as shown in **Figures 1E–G**. The demonstrated efficacy of GS-967 to suppress EAD-mediated arrhythmias provided a direct link between a specific molecular action of a drug (i.e., “gating modifier” specific block of I_{Na-L}) and a dynamical arrhythmia mechanism (enhanced I_{Na-L} -mediated EADs) (Qu et al., 2013). The term “gating modifier” has been used to refer to the mode of action of toxins that alter the voltage dependence of voltage gated ion channels by binding to structures that undergo conformational changes during gating, e.g., the voltage sensing domains (Catterall et al., 2007; Kalia et al., 2015). Here, we used the term gating modifier to refer to drug action that alter channel opening and channel closing without directly blocking the pore. Modulation of the gating of brain sodium channels accounts, at least in part, for the ability of several anti-epileptic drugs including lamotrigine to suppress epileptic attacks. Interestingly it is shown that lamotrigine exerts its effect by binding to the two voltage sensors, segments S4, of the domains III and IV of Nav channel that potently suppress EAD-mediated triggered activity (epileptic discharges) without affecting ordinary nerve action potential firing (Rogawski and Loscher, 2004). This finding suggest that the voltage sensor domain of the voltage gated Na channels can be a drugable target for antiepileptic drugs. Indeed, in a transgenic mouse model of epilepsy (*Scn2a*^{Q54}), the I_{Na-L} was found to be increased significantly from 1 to 3 percent of peak I_{Na} and importantly the specific I_{Na-L} blocker prototype GS-967,

mitigated epileptic attacks (Anderson et al., 2014). These findings suggest that the gating modifiers of Nav channels may be effective and target specific anti-epileptics (Anderson et al., 2014; George, 2015).

Unfortunately, the current Classes of AADs are incapable to selectively discriminate between the pathological I_{Na-L} and the normal peak I_{Na} necessary for normal AP upstroke velocity as they equally suppress both the early and late components of the I_{Na} . For example, flecainide (Class 1C AAD) blocks both the peak I_{Na} and I_{Na-L} in wild type and in LQT3 mutant Na channels as shown in **Figures 1E,F**. Similarly, mexiletine, a Class 1B AAD has similar effects as flecainide on peak and I_{Na-L} (**Figure 1G**; Gao et al., 2013). The partially selective I_{Na-L} blocker ranolazine, while suppressing the I_{Na-L} and EAD-mediated VT/VF (Sicouri et al., 2008; Morita et al., 2011a) was tentatively suggested to fit Class 1B, due to its structural similarity to lidocaine (Thireau et al., 2011). However, ranolazine blockade of I_{Kr} which tends to prolong the APD and the QT interval may well place it in the Class 1A category, like quinidine (**Table 1**).

The Late L-Type Ca Current ($I_{Ca,L}$)

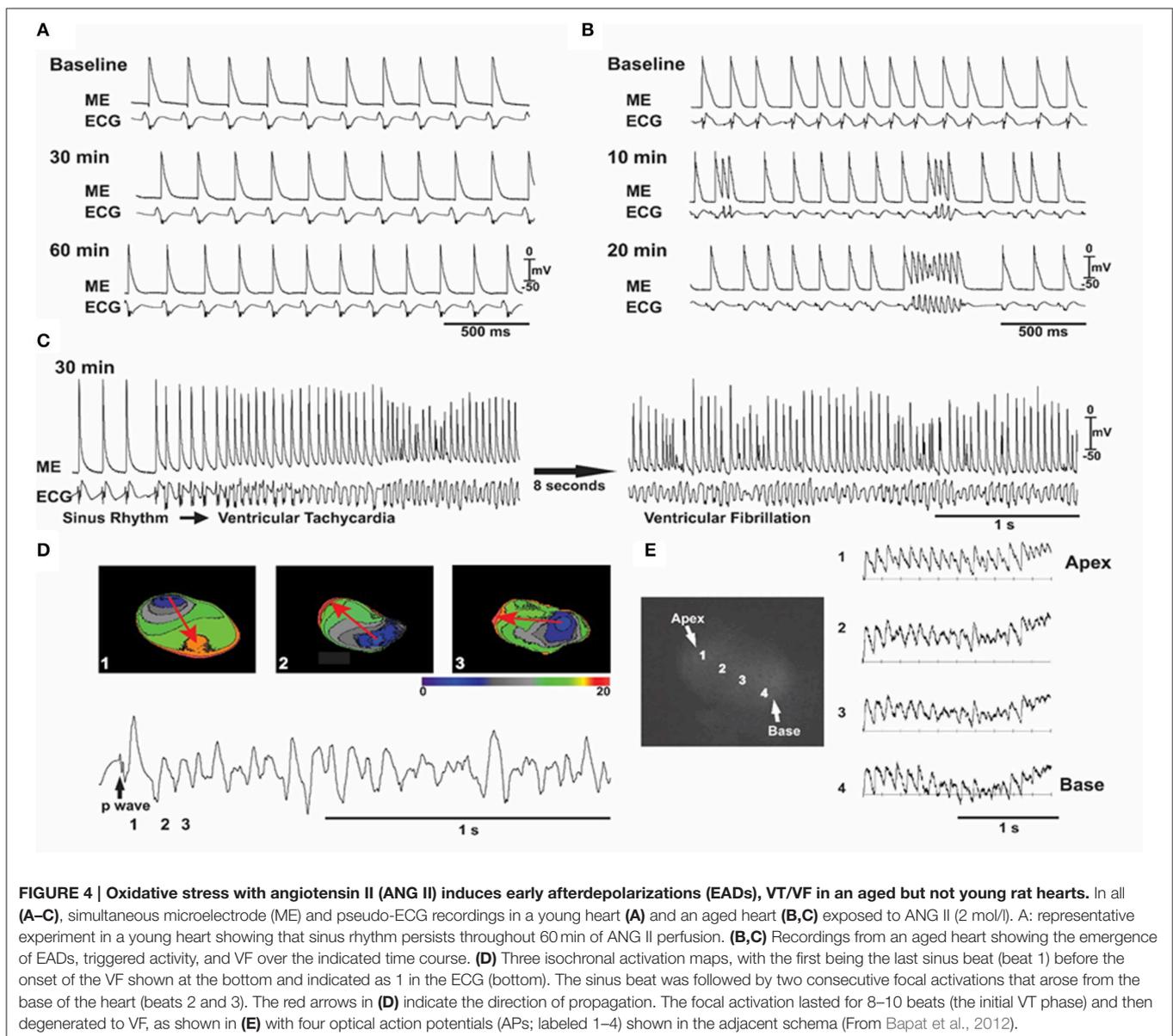
Cardiac conditions that increase the late $I_{Ca,L}$ are invariably associated with increased risk of VT/VF, including heart failure (Mewes and Ravens, 1994; Cooper et al., 2010), congenital LQT8 mutations and Timothy Syndrome (TS) (**Figure 2A**; Limpitikul et al., 2014; Marsman et al., 2014; Dick et al., 2016) increased pro-oxidant states, (**Figure 2B**; Xie et al., 2009) aging, (Zhou et al., 1998; Salameh et al., 2010), and Duchenne muscular dystrophy (Koenig et al., 2011; Viola et al., 2013) and in stem cell derived



cardiomyocytes from patients with Timothy syndrome (Yazawa et al., 2011; **Figure 2D**).

Electrophysiological studies in isolated cardiac myocytes have shown that an increase in the late $I_{Ca,L}$ causes EADs (Xie et al., 2009; **Figure 2C**). Most EADs develop between -40 and 0 mV, a range of membrane potentials commonly referred to as the window current region, where the steady-state activation and steady-state inactivation curves overlap. Within the $I_{Ca,L}$ window region, a fraction of Ca channels are not fully inactivated maintaining a non-zero open probability responsible for the late Ca current. Recent studies based on dynamic clamp data provide compelling evidence that interventions reducing late $I_{Ca,L}$ by either by (1) shifting the $I_{Ca,L}$ steady-state activation in the depolarizing direction by <5 mV, (2) shifting the steady-state inactivation curve in the hyperpolarizing direction by (<5

mV), or (3) reducing the non-inactivating (pedestal) component (Madhvani et al., 2015; **Figure 3**) potently suppresses EADs and therefore carry a strong therapeutic value for suppressing EAD-mediated VT/VF. Traditional CCBs like verapamil or nifedipine can suppress oxidative EADs by blocking $I_{Ca,L}$ (Xie et al., 2009) however, their suppressant effect of EADs comes at a cost of considerable reduction of the peak $I_{Ca,L}$ as shown in **Figure 2E**. The simultaneous block of peak and late $I_{Ca,L}$ by a drug greatly diminishes the therapeutic value particularly in patients with compromised cardiac function (**Figure 2F**; Nawrath et al., 1998). Importantly, the dynamic clamp studies of Madhvani et al. (2015) provide compelling evidence that the blockade of peak $I_{Ca,L}$ is not only undesirable but also not necessary for EADs suppression! These studies made it clear that the selective reduction of the late $I_{Ca,L}$ can preserve Ca transient and



normal EC coupling, while at the same time potently suppressing the EADs. These findings suggest that a new class of drug action, “gating modifiers,” as in the case of I_{Na-L} , that induce subtle changes of the gating properties of the LTCC and Nav channels without blocking their pores suppress EAD-mediated VT/VF without adversely affecting contractility. Interestingly, the identification roscovitine, a purine derivative (anticancer agent) able to selectively reduce the late $I_{Ca,L}$ without affecting the peak $I_{Ca,L}$ represents a turning point in the electropharmacology of AADs (Yarotsky and Elmslie, 2007; Yazawa and Dolmetsch, 2013; **Figure 2D**).

Intact Heart Models of EAD-Mediated VT/VF via CaMKII Signaling

We and others have shown that oxidative activation of CaMKII by hydrogen peroxide (H_2O_2) increases the I_{Na-L} and late $I_{Ca,L}$ and readily promotes EADs and triggered activity in isolated

rat and rabbit ventricular myocytes (Ward and Giles, 1997; Song et al., 2006; Xie et al., 2009; Madhvani et al., 2015). At tissue level however, EADs may be suppressed by source-sink mismatches arising from cell-to-cell coupling. That is, the small inward current which is sufficient to reverse repolarization and cause an EAD at the isolated myocyte level will be “diluted” into adjacent repolarizing myocytes at the tissue level (unless they are also simultaneously primed for an EAD), thereby preventing EAD formation. The isolated single cell vs. tissue level vulnerability to EAD formation was investigated by examining the arrhythmogenic effects of hydrogen peroxide (H_2O_2) in Langendorff rat and rabbit hearts. Oxidative stress was shown to readily induce EADs at the isolated myocyte level in these two species. Consistent with the predicted suppressive effects of well-coupled tissue on EADs, we found that oxidative stress with H_2O_2 failed to induce any ventricular arrhythmias in normal well-coupled young/adult (~4 months old) rat and rabbit hearts

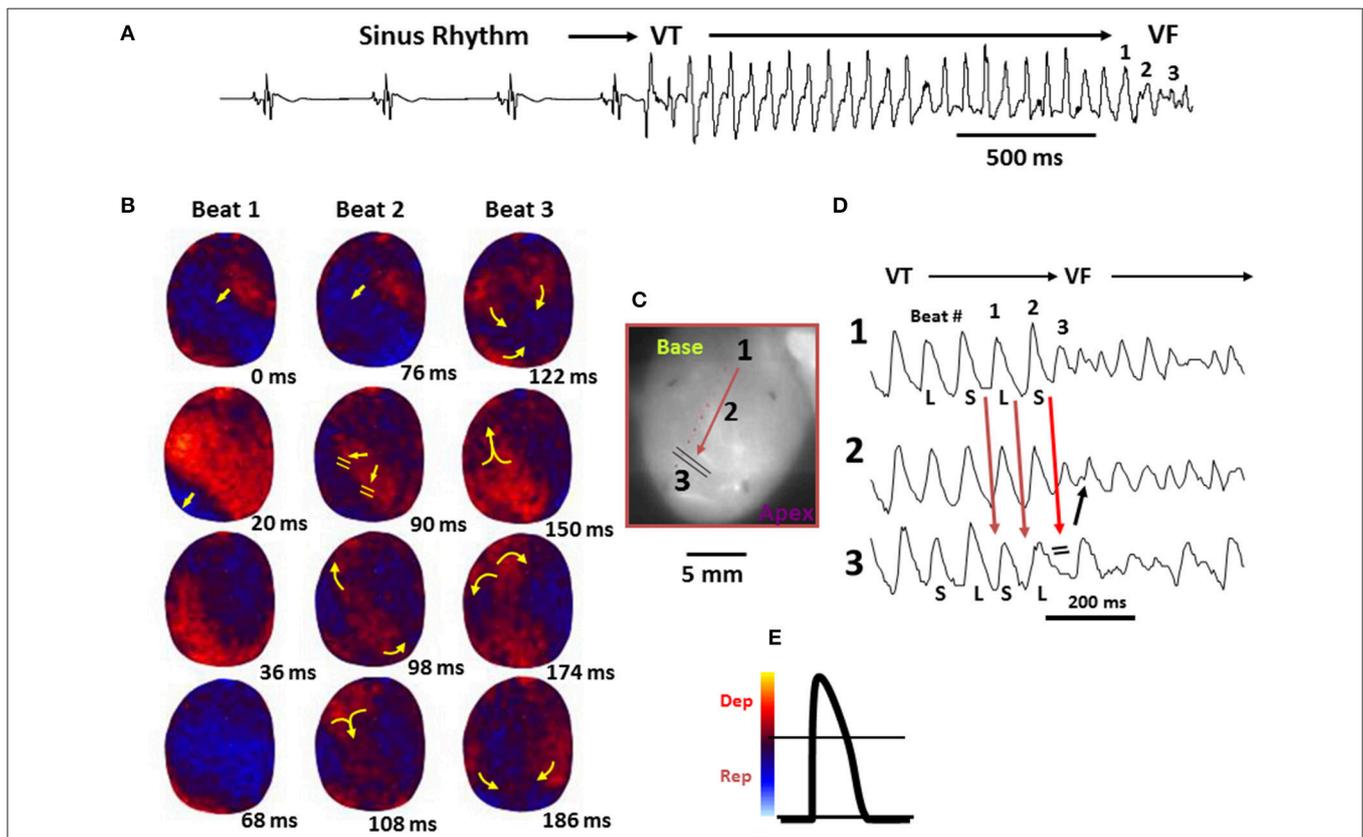


FIGURE 5 | Spontaneous initiation of ventricular tachycardia (VT)/VF in an aged rat heart exposed to 0.1 mM H_2O_2 . (A) ECG showing the last 5 sinus beats before the sudden onset of VT leading to VF. (B) voltage snapshots of the last beat of the VT (beat 1) and of the first 2 beats of the VF (beats 2 and 3). In each snapshot, activation time (in ms) is shown at the bottom right with time 0 (arbitrary) coinciding with the onset of beat 1. The red color in the snapshots represents depolarization (Dep) and the blue repolarization (Rep) as shown in (E). The yellow arrows in the snapshots represent the direction of the wavefront propagation with double horizontal lines denoting the site of conduction block. The VT originates from a focal site at the LV base and propagates as single wavefront toward the apex and undergoes functional conduction block at site 3. The two lateral edges of the front, however, continue to propagate laterally (snapshot, 98 ms) forming figure-eight reentry (snapshot, 108 ms). During the second reentrant wavefront, another wavefront emerges from the apical site of the LV (snapshot, 122 ms), disrupting the activation pattern and signaling the onset of VF. (D) 3 optical action potentials (APs; labeled 1, 2, and 3) recorded from sites identified on the heart silhouette (C). The 2 downward-pointing blue arrows indicate the direction of propagation from site 1 to site 3 with the red downward-pointing arrow showing block at site 3, followed by retrograde activation (upward-pointing arrow). Notice the emergence of spatially discordant AP duration (APD) alternans preceding conduction block at site 3 when the front with short APD (S) at site 1 encroaches a site (site 3) with long APD (L) (From Morita et al., 2009).

(Figure 4A). Even a tenfold increase in the level of peroxide failed to promote EADs in these normal hearts (Morita et al., 2009). However, aged, 24–26 months old rat hearts, manifest considerable heterogeneous increase in interstitial tissue fibrosis (10–90%), with reduced cell-to-cell gap junctional couplings via connexin43 (Cx43), H₂O₂ exposure consistently promoted EADs and triggered activity leading in >90% of the aged fibrotic hearts to VT/VF. (Figures 4B–E) Similarly, middle-aged, 3–5 years old, (normal life span 8–12 years) rabbit hearts, with lesser rise in fibrosis (5–35%) than in aged rat hearts and developed lesser incidence (~50%) of EAD-mediated triggered VF (Morita et al., 2009; Sato et al., 2009). Although other aging-related factors may play a role in EAD-mediated VF, we hypothesized that reduced cell-to-cell gap junctional coupling in fibrotic aged hearts promotes oxidative EAD-related arrhythmias by favorably altering source-sink relationships. Indeed, computer simulation in 1, 2, and 3D cardiac tissue incorporating fibrosis fully supported this interpretation (Xie et al., 2010; Morita et al., 2011a). The involvement of peroxide, as a redox sensor of CaMKII, is validated by demonstrating similar VT/VF outcomes with more proximal oxidative signaling using acute angiotensin II perfusion (Bapat et al., 2012) or by mimicking distal

consequences of oxidative stress, such as glycolytic inhibition (Corretti et al., 1991), all promoting EAD-mediated VT/VF as in the case of H₂O₂ (Morita et al., 2011b). Interestingly both oxidative stress with peroxide (Pezhouman et al., 2014a) or downstream oxidative injury with glycolytic inhibition (Ono et al., 2007) in aged atria promote atrial EADs, tachycardia and fibrillation (AT/AF) in aged rat atria as in aged rat ventricles. However, it remains to be seen if block of the late inward currents would also prevent and suppress AT/AF in other species such as rabbit, goat, sheep and dog models of AT/AF.

ENHANCED LATE INWARD CURRENTS PROMOTE BOTH FOCAL TRIGGERED AND REENTRANT ARRHYTHMIAS

The EADs and the subsequent propagating triggered beats initiate diverse forms of ventricular arrhythmias starting from single PVCs, focal non-reentrant monomorphic or polymorphic VT and torsade de pointes that lead to the highly arrhythmogenic spatially discordant APD alternans causing wavebreak, reentry and transition from VT to VF (Morita et al., 2011a,b; Bapat

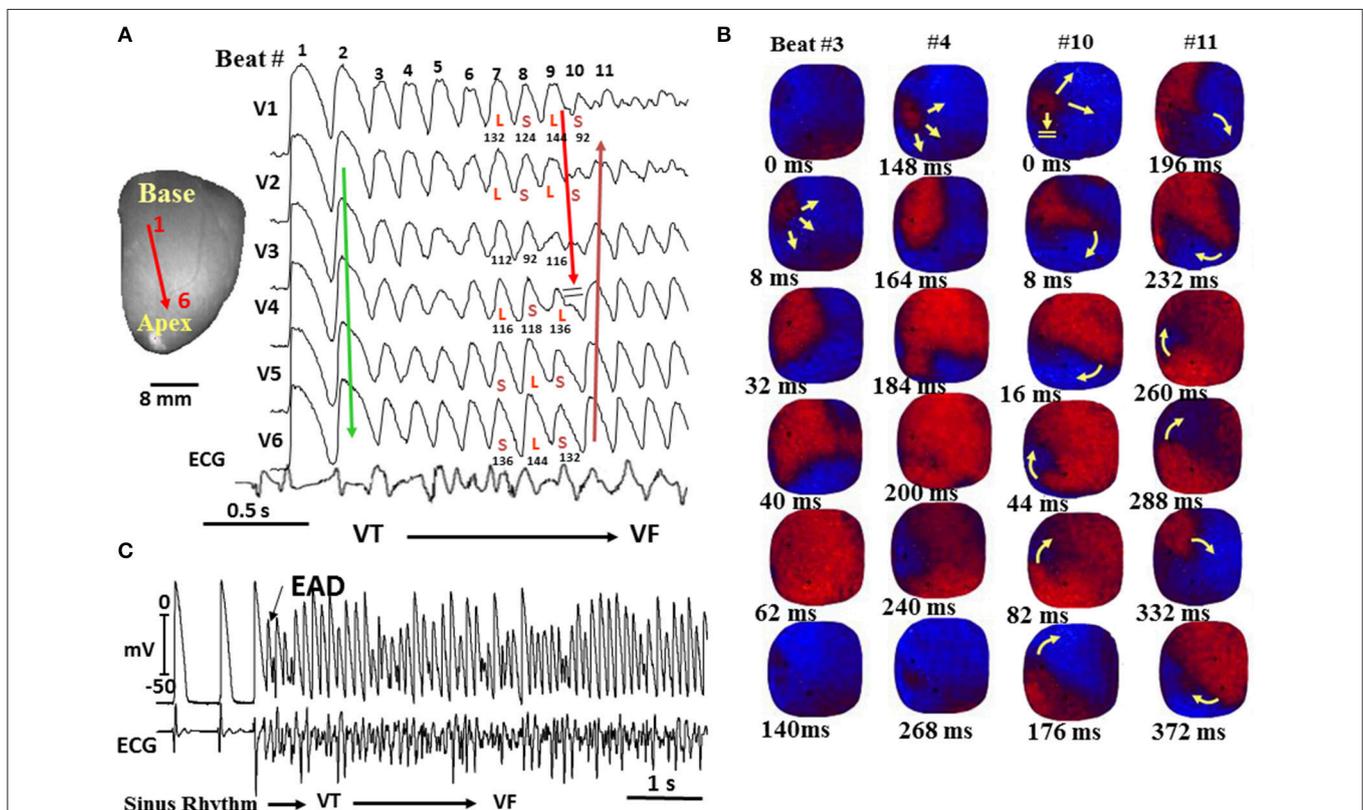
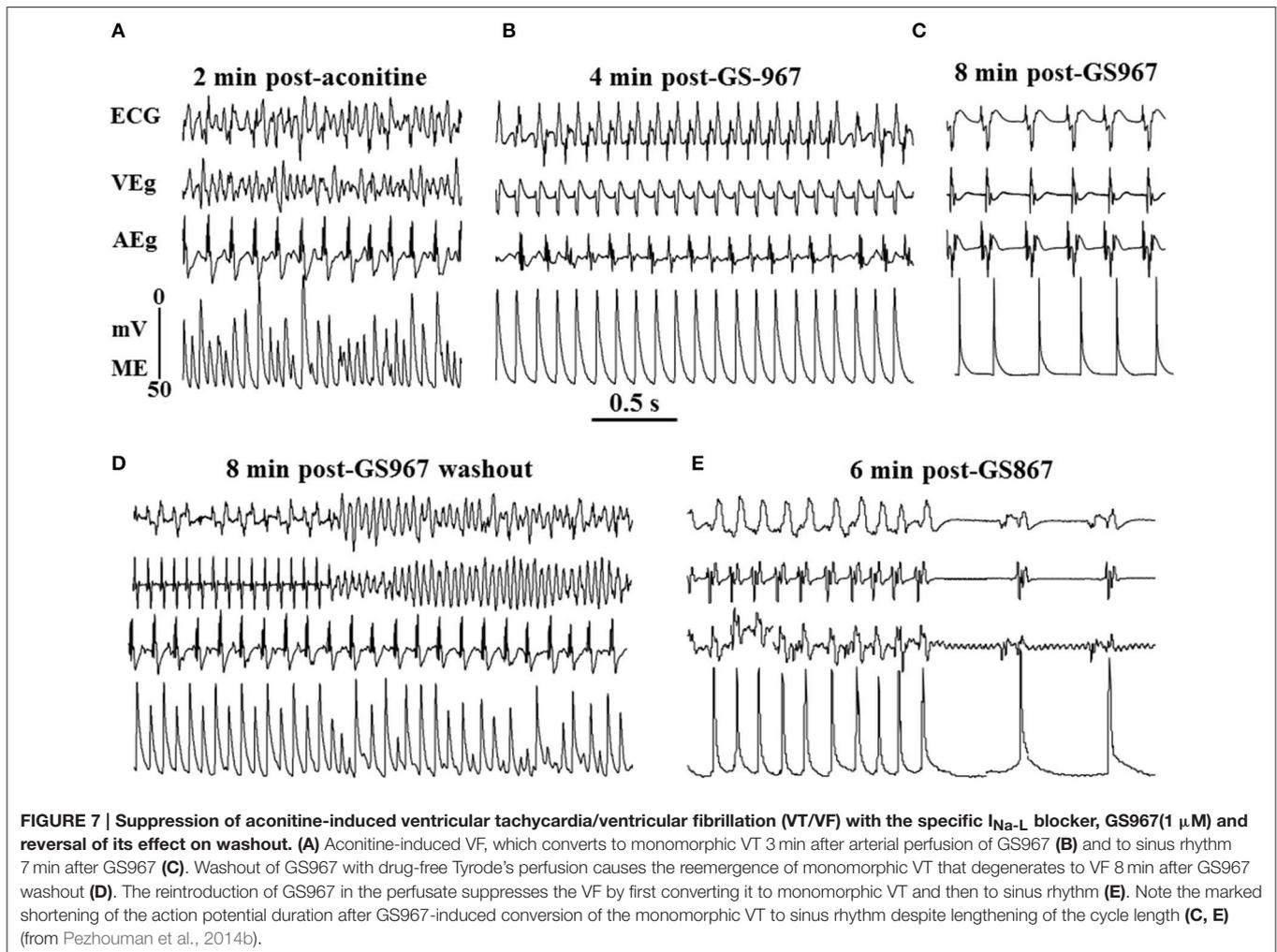
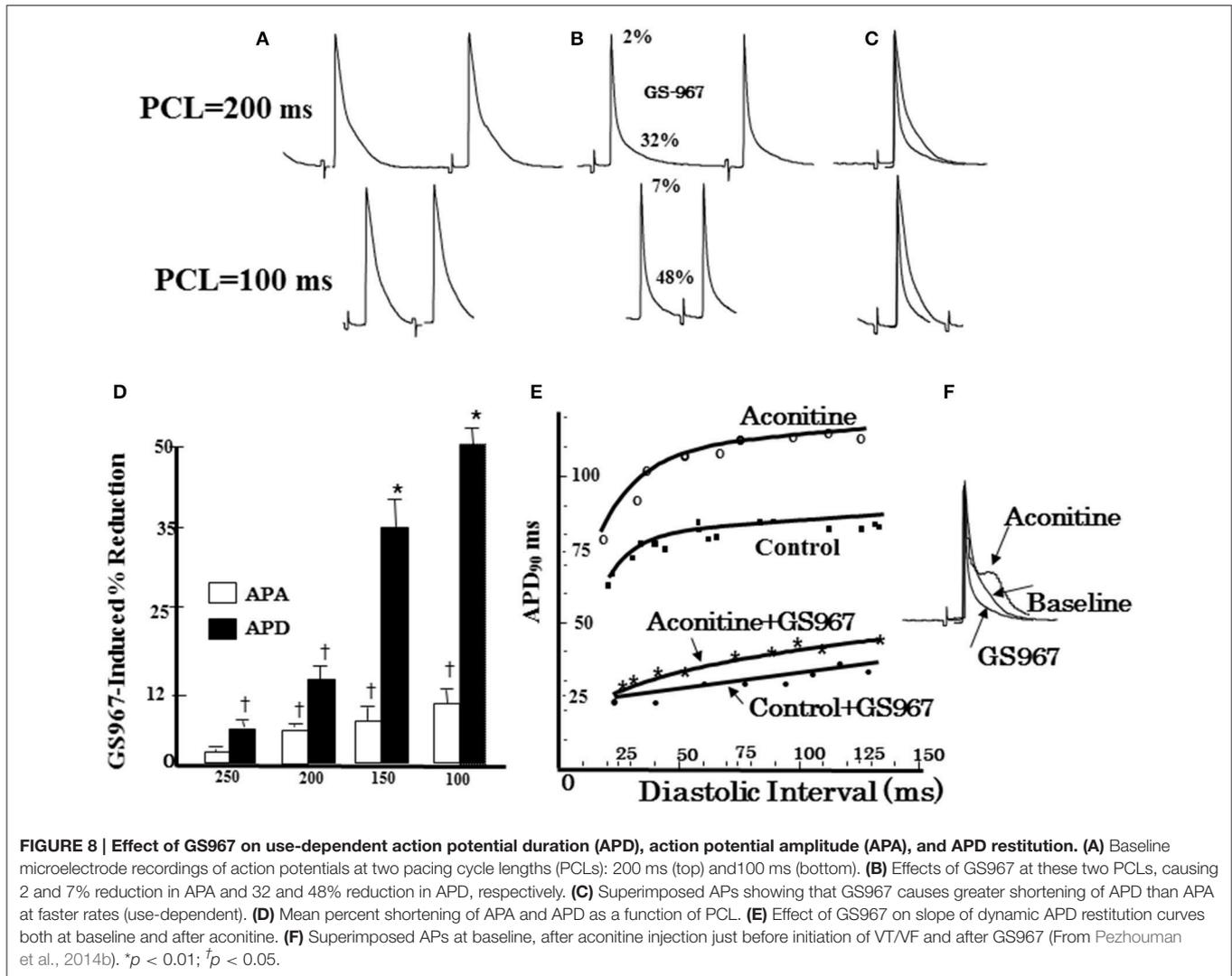


FIGURE 6 | Spontaneous initiation of VF in a middle-aged rabbit exposed to 0.1 mM H₂O₂. (A) Six epicardial optical APs (V1–V6) recorded from sites shown on the left silhouette of the heart. After nine focal activations arising from the base of the heart (B), the wavefront undergoes block at mid-LV anterior wall after spatially discordant APD alternans emerges (A). The wavefront, however, continues to propagate lateral to the site of block in a clockwise direction causing a reentrant excitation as shown in the snapshots in (B). The numbers under each snapshot is activation time starting with 0 ms (arbitrary) for beats 3 and 4 and then again for beats 10 and 11. (C) microelectrode recording of another VF episode recorded in the same heart showing spontaneous initiation of VF by a mechanism compatible with early afterdepolarization (EAD)-mediated triggered activity (TA) as in the aged rat hearts shown in Figure 6 (From Morita et al., 2009).

et al., 2012; Chang et al., 2012; Pezhouman et al., 2015) as shown in **Figures 5, 6**. In addition to triggered foci, the oxidative stress may also initiate reentry by the phenomenon of “repolarization failure” during which the AP remains in the plateau region for a few seconds before undergoing full repolarization (Bapat et al., 2012). Localized myocardial regions with failed repolarization creates a considerable increase in spatial dispersion of repolarization promoting wavebreak and reentry (Weiss et al., 2000; Antzelevitch, 2008; Sato et al., 2009; Bapat et al., 2012; Karagueuzian et al., 2013). Heterogeneous increase in the late inward currents can also promote APD dispersion via their actions on the kinetics of APD restitution. Enhanced I_{Na-L} and late $I_{Ca,L}$ by prolonging the APD increase the slope of the APD restitution curve (Qu et al., 2000; Guo et al., 2011; Morita et al., 2011a; Pezhouman et al., 2014b; **Figure 7**). Increasing the slope of the APD restitution curve promotes voltage- but not Ca-driven APD alternans (Weiss et al., 2006) which with a critical increase in the rate of activation converts the concordant APD alternans to the highly arrhythmogenic spatially discordant APD alternans (out-of-phase APD alternans) promoting unidirectional conduction block and reentry (Weiss

et al., 2006; Morita et al., 2009; Karagueuzian et al., 2013). Selective block of the I_{Na-L} with GS-967 or selective block of the late $I_{Ca,L}$ with roscovitine flatten the slope of the APD restitution curve preventing wavefront breakup and reentry by preventing the emergence of spatially discordant APD alternans (Pezhouman et al., 2014b, 2016; Angelini et al., 2016; **Figures 7, 8**). Interestingly flattening of the slope of the APD restitution with drugs that do not block the I_{Na-L} also prevent wavebreak and reentry but do not prevent EAD-mediated triggered activity (I_{Na-L} remains elevated) causing the maintenance of VT with no transition to VF. This dynamic scenario is confirmed with the studies with diacetyl monoxime (DAM), a drug that flattens the slope of the APD restitution curve without affecting the late currents (Coulombe et al., 1990; Riccio et al., 1999). DAM decreases in a concentration-dependent manner the amplitude of both the slow inward calcium current and the transient outward current, accelerating their inactivation and shifting their steady-state inactivation-voltage relationships toward negative potentials (Coulombe et al., 1990). In isolated-perfused swine ventricles in which EAD-mediated triggered VT/VF is initiated by myocardial



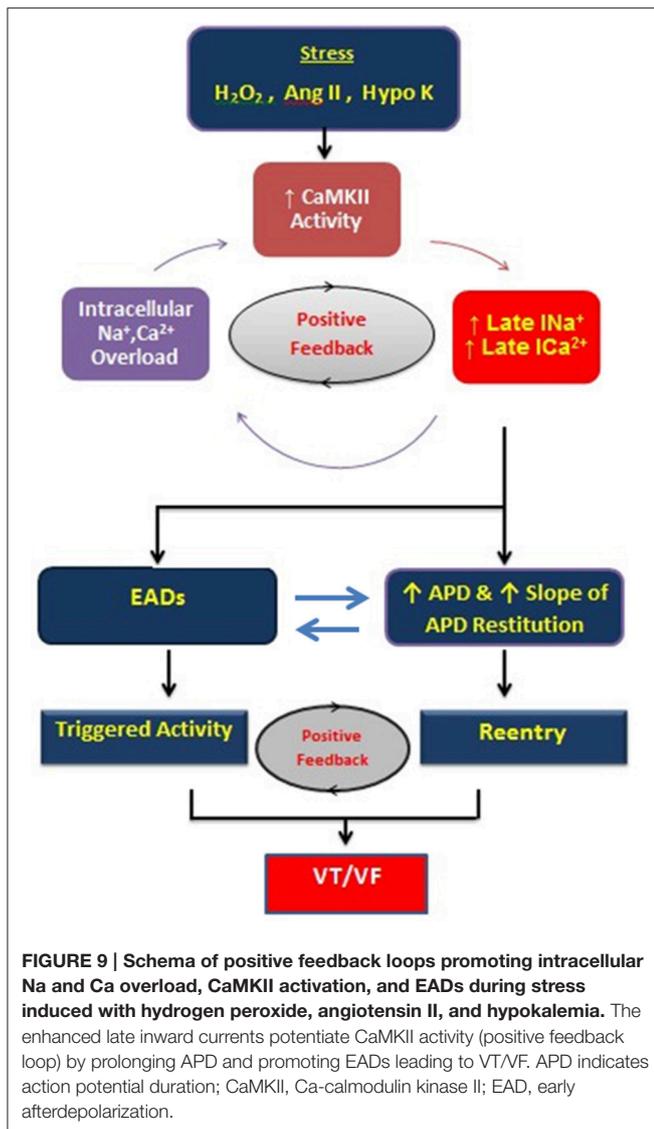


local injection of aconitine (Swissa et al., 2002) an agent that selectively enhances the I_{Na-L} considerably (Peper and Trautwein, 1967), the administration of DAM prevented transition of aconitine-induced EAD-mediated triggered VT to VF which remained maintained by a single triggered focus maintaining a monomorphic VT (Swissa et al., 2002). The flattening of the slope of the APD restitution curve by DAM differs from that of GS-967 in that the latter unlike DAM by selectively blocking the I_{Na-L} suppresses both EAD-mediated triggered activity and reentry caused by wavebreak due to the flattening the slope of the APD restitution resulting in the conversion aconitine-induced VF to sinus rhythm (Pezhouman et al., 2014b; Figures 7, 8).

SPECIFIC VS. NON-SPECIFIC BLOCK OF DRUGS AND VT/VF

Many of the Class I AADs like mexiletine, flecainide, and propafenone, while blocking the peak I_{Na} (Figure 1)

also manifest variable degree of block of the I_{Na-L} as well (Antzelevitch et al., 2014). However, the potential of proarrhythmia with these agents led to the development of more selective blockers of I_{Na-L} , with no appreciable effect on the peak I_{Na} (Belardinelli et al., 2013). Furthermore, Class IC drugs like flecainide and propafenone in addition of being potent peak I_{Na} inhibitors also inhibit ryanodine 2 (RyR2) channels in the open-state, suppressing Ca^{2+} waves and preventing VT in the congenital catecholaminergic polymorphic ventricular tachycardia (CPVT) both in mice and humans (Hilliard et al., 2010). Clinical studies have shown that CaMKII activation in heart failure (Swaminathan et al., 2012) with its downstream increases of both I_{Na-L} and late I_{Ca-L} may require the block of both late inward currents to be effective against VT/VF (Figure 9). Support to this contention comes from simulation (Foteinou et al., 2015) and clinical studies that showed the need to combine ranolazine with verapamil to effectively suppress VT/VF in a patient with TS (LQT8) (Shah et al., 2012). While triggered activity either by EAD and subsequent emergent of



DADs (Pezhouman et al., 2014b) remains a viable mechanism of VT/VF, the prevailing concept of AAD therapy in patients with healed myocardial infarction and other cardiomyopathies however, is largely based on how to disrupt reentrant excitation to terminate VT/VF (de Bakker et al., 1991; Pogwizd et al., 1992; Wu et al., 1998; Nash et al., 2006). Accordingly, the strategy of AAD is largely based (save for rare channelopathies, like CPVT) on a drug's ability to block ionic currents to disrupt reentrant arrhythmias. In fact, by directly altering cardiac wavefront conduction and refractoriness reentry may be prevented (Rosen and Janse, 2010). With this mindset it is proposed that Na channel blockers (i.e., Class I AAD) suppress reentry by decreasing conduction velocity and excitability converting unidirectional conduction block necessary for reentry formation to bidirectional conduction block preventing reentry and VT/VF (Karagueuzian and Chen, 2001). These expectations proved to be not only disappointing with respect to clinical therapy of VT/VF but also carried significant proarrhythmic risks. For example, the

outcome of CAST clinical trial indicated that inhibition of peak Na current (Class 1C) was ineffective and even carried greater risk of mortality (Echt et al., 1991). Similarly, the SWORD clinical trials with d-sotalol (Class III) (Waldo et al., 1995) were also based on the premise of preventing reentry formation this time by increasing the cardiac refractoriness, an effect that will eliminate (close) the excitable gap necessary for reentry causing termination of the reentry. The results of CAST and SWORD showed drugs with Class I and Class III actions are ineffective in suppressing the arrhythmia following myocardial infarction (Echt et al., 1991; Waldo et al., 1995).

New AAD Class Action (Class VI)

The current AAD classification, classically known as the Vaughan Williams classification, needs to be refined and updated with recent findings that attribute key arrhythmogenic role to the pathological rises of the late inward currents. As stated by Thireau and associates: "The Vaughan Williams classification may no longer be best suited to the realities of new AA drug design" (Thireau et al., 2011).

The key role played by the sustained elevation of the I_{Na-L} and late $I_{Ca,L}$ in the genesis of focal and reentrant VT/VF qualify these pathological changes in ionic current kinetics *bone fide* "vulnerable parameters" of arrhythmias that are selectively targetable by small molecule drugs. In fact, the emergences of potent and prototype specific blockers of these late currents offers a new and effective AAD class action not considered previously. Since a fifth class of AAD actions is already assigned to diverse other agents (Table 1) it is suggested that the specific modifiers of the I_{Na-L} and late $I_{Ca,L}$ (gating modifiers) be assigned a sixth AAD class action (Class VI). Prototype drugs with these class of actions are steadily becoming available including GS-967, the mexiletine analogs, HBRI21 & HBRI23 and roscovitine, that manifest high selectivity against the late I_{Na-L} and the $I_{Ca,L}$ respectively without affecting their peaks (McKeithan et al., 2016). To distinguish between the two inward currents, the notations of Class VI_{Na} and Class VI_{Ca} may be added to indicate specific I_{Na-L} and late $I_{Ca,L}$ blockers respectively. The concept of AAD therapy based on changes in ion channel gating rather than peak current block (i.e., gating modifiers) represents a turning point in AAD therapy that would provide a stimulus for future research in the field of AAD discovery that remained idle for decades (Kamath and Mittal, 2008; Thireau et al., 2011). Here, it is fitting to re-state the Editorial comments entitled "Rational strategy to stop arrhythmias..." made our group's findings (Madhvani et al., 2015) on the role of the late $I_{Ca,L}$ in the genesis of EADs:

"...Existing strategies for developing antiarrhythmic drugs have largely failed, and so new, innovative approaches as described by Madhvani et al. 2015 need to be aggressively pursued and tested" (Markandeya and Kamp, 2015).

AUTHOR CONTRIBUTIONS

HK conceived and wrote the paper and RO discussed and added his recently published data and critically read the

manuscript with suggestions and additions. AP and MA performed the experiments on the novel prototype drugs (Class VI), analyzed the data and prepared the figures for the manuscript.

REFERENCES

- Anderson, L. L., Thompson, C. H., Hawkins, N. A., Nath, R. D., Petersohn, A. A., Rajamani, S., et al. (2014). Antiepileptic activity of preferential inhibitors of persistent sodium current. *Epilepsia* 55, 1274–1283. doi: 10.1111/epi.12657
- Angelini, M. P. A., Savalli, N., Pantazis, A., Melkonian, A., Weiss, J. N., Olcese, R. et al. (2016). Roscovitine as the archetypal member of a novel class of antiarrhythmics targeting late I_{Ca-L}. *Biophys. J.* 110:272a. doi: 10.1016/j.bpj.2015.11.1478
- Antzelevitch, C. (2008). Drug-induced spatial dispersion of repolarization. *Cardiol. J.* 15, 100–121.
- Antzelevitch, C., Nesterenko, V., Shryock, J. C., Rajamani, S., Song, Y., and Belardinelli, L. (2014). The role of late I_{Na} in development of cardiac arrhythmias. *Handb. Exp. Pharmacol.* 221, 137–168. doi: 10.1007/978-3-642-41588-3_7
- Bäckman, N., Holm, A. K., Hånström, L., Blomquist, H. K., Heijbel, J., and Säfström, G. (1989). Folate treatment of diphenylhydantoin-induced gingival hyperplasia. *Scand. J. Dent. Res.* 97, 222–232. doi: 10.1111/j.1600-0722.1989.tb01606.x
- Backs, J., Backs, T., Neef, S., Kreusser, M. M., Lehmann, L. H., Patrick, D. M., et al. (2009). The delta isoform of CaM kinase II is required for pathological cardiac hypertrophy and remodeling after pressure overload. *Proc. Natl. Acad. Sci. U.S.A.* 106, 2342–2347. doi: 10.1073/pnas.0813013106
- Baillie, D. S., Inoue, H., Kaseda, S., Ben-David, J., and Zipes, D. P. (1988). Magnesium suppression of early afterdepolarizations and ventricular tachyarrhythmias induced by cesium in dogs. *Circulation* 77, 1395–1402. doi: 10.1161/01.CIR.77.6.1395
- Baker, W. L. (2016). Treating arrhythmias with adjunctive magnesium: identifying future research directions. *Eur. Heart J. Cardiovasc. Pharmacother.* doi: 10.1093/ehjcvp/pvw028. [Epub ahead of print].
- Bapat, A., Nguyen, T. P., Lee, J. H., Sovari, A. A., Fishbein, M. C., Weiss, J. N., et al. (2012). Enhanced sensitivity of aged fibrotic hearts to Angiotensin II- & hypokalemia-induced early afterdepolarizations-mediated ventricular arrhythmias. *Am. J. Physiol. Heart Circ. Physiol.* 302, H2331–H2340. doi: 10.1152/ajpheart.00094.2012
- Belardinelli, L., Giles, W. R., Rajamani, S., Karagueuzian, H. S., and Shryock, J. C. (2015). Cardiac late Na current: Proarrhythmic effects, roles in long QT syndromes, and pathologic relationship to CaMKII and oxidative stress. *Heart Rhythm* 12, 440–448. doi: 10.1016/j.hrthm.2014.11.009
- Belardinelli, L., Liu, G., Smith-Maxwell, C., Wang, W. Q., El-Bizri, N., Hirakawa, R., et al. (2013). A novel, potent, and selective inhibitor of cardiac late sodium current suppresses experimental arrhythmias. *J. Pharmacol. Exp. Ther.* 344, 23–32. doi: 10.1124/jpet.112.198887
- Belhassen, B., Glick, A., and Landiadi, S. (1988). Comparative clinical and electrophysiologic effects of adenosine triphosphate and verapamil on paroxysmal reciprocating junctional tachycardia. *Circulation* 77, 795–805. doi: 10.1161/01.CIR.77.4.795
- Bennett, P. B., Yazawa, K., Makita, N., and George, A. L. Jr. (1995). Molecular mechanism for an inherited cardiac arrhythmia. *Nature* 376, 683–685. doi: 10.1038/376683a0
- Bigger, J. T. Jr., Bassett, A. L., and Hoffman, B. F. (1968). Electrophysiological effects of diphenylhydantoin on canine purkinje fibers. *Circ. Res.* 22, 221–236. doi: 10.1161/01.RES.22.2.221
- Camm, A. J., and Garratt, C. J. (1994). Adenosine and supraventricular tachycardia. *N. Engl. J. Med.* 325, 1621–1629.
- Catterall, W. A., Cèstele, S., Yarov-Yarovoy, V., Yu, F. H., Konoki, K., and Scheuer, T. (2007). Voltage-gated ion channels and gating modifier toxins. *Toxicol.* 49, 124–141. doi: 10.1016/j.toxicol.2006.09.022
- Chang, M. G., Sato, D., de Lange, E., Lee, J. H., Karagueuzian, H. S., Garfinkel, A., et al. (2012). Bi-stable wave propagation and early afterdepolarization-mediated cardiac arrhythmias. *Heart Rhythm.* 9, 115–122. doi: 10.1016/j.hrthm.2011.08.014
- Charlier, R. (1970). Cardiac actions in the dog of a new antagonist of adrenergic excitation which does not produce competitive blockade of adrenoceptors. *Br. J. Pharmacol.* 39, 668–674. doi: 10.1111/j.1476-5381.1970.tb09892.x
- Charlier, R., Delaunoy, G., Bauthier, J., and Deltour, G. (1969). Research on the benzofuran series. XL. Anti-arrhythmic properties of amiodarone. *Cardiologia* 54, 83–90. doi: 10.1159/000166244
- Charlier, R., Deltour, G., Baudine, A., and Chaillet, F. (1968). Pharmacology of amiodarone, and anti-anginal drug with a new biological profile. *Arzneimittelforschung* 18, 1408–1417.
- Cooper, P. J., Soeller, C., and Cannell, M. B. (2010). Excitation-contraction coupling in human heart failure examined by action potential clamp in rat cardiac myocytes. *J. Mol. Cell. Cardiol.* 49, 911–917. doi: 10.1016/j.yjmcc.2010.04.012
- Coppini, R., Ferrantini, C., Yao, L., Fan, P., Del Lungo, M., Stillitano, F., et al. (2013). Late sodium current inhibition reverses electromechanical dysfunction in human hypertrophic cardiomyopathy. *Circulation* 127, 575–584. doi: 10.1161/CIRCULATIONAHA.112.134932
- Corretti, M. C., Koretsune, Y., Kusuoka, H., Chacko, V. P., Zweier, J. L., and Marban, E. (1991). Glycolytic inhibition and calcium overload as consequences of exogenously generated free radicals in rabbit hearts. *J. Clin. Invest.* 88, 1014–1025. doi: 10.1172/jci115361
- Coulombe, A., Lefevre, I. A., Deroubaix, E., Thuringer, D., and Coraboeuf, E. (1990). Effect of 2,3-butanedione 2-monoxime on slow inward and transient outward currents in rat ventricular myocytes. *J. Mol. Cell. Cardiol.* 22, 921–932. doi: 10.1016/0022-2828(90)90123-J
- de Bakker, J. M. T., van Capelle, F. J. L., Janse, M. J., van Hemel, N., Hauer, R. N. W., Defauw, J. J. A. M., et al. (1991). Macroreentry in the infarcted human heart: The mechanism of ventricular tachycardias with a “Focal” activation pattern. *J. Am. Coll. Cardiol.* 18, 1005–1014. doi: 10.1016/0735-1097(91)90760-7
- Dick, I. E., Joshi-Mukherjee, R., Yang, W., and Yue, D. T. (2016). Arrhythmogenesis in timothy syndrome is associated with defects in Ca²⁺-dependent inactivation. *Nat. Commun.* 7:10370. doi: 10.1038/ncomms10370
- Echt, D. S., Liebson, P. R., Mitchell, L. B., Peters, R. W., Obias-Manno, D., Barker, A. H., et al. (1991). Mortality and morbidity in patients receiving encainide, flecainide, or placebo. The cardiac arrhythmia suppression trial. *N. Engl. J. Med.* 324, 781–788. doi: 10.1056/NEJM199103213241201
- Erickson, J. R., He, B. J., Grumbach, I. M., and Anderson, M. E. (2011). CaMKII in the cardiovascular system: sensing redox states. *Physiol. Rev.* 91, 889–915. doi: 10.1152/physrev.00018.2010
- Erickson, J. R., Joiner, M. L., Guan, X., Kutschke, W., Yang, J., Oddis, C. V., et al. (2008). A dynamic pathway for calcium-independent activation of CaMKII by methionine oxidation. *Cell.* 133, 462–474. doi: 10.1016/j.cell.2008.02.048
- European Heart Rhythm, A., European Association for Cardio-Thoracic, S., Camm, A. J., Kirchhof, P., Lip, G. Y., Schotten, U., et al. (2010). Guidelines for the management of atrial fibrillation: the task force for the management of atrial fibrillation of the European Society of Cardiology (ESC). *Europace* 12, 1360–1420. doi: 10.1093/europace/euq350
- Foteinou, P. T., Greenstein, J. L., and Winslow, R. L. (2015). Mechanistic investigation of the arrhythmogenic role of oxidized CaMKII in the Heart. *Biophys. J.* 109, 838–849. doi: 10.1016/j.bpj.2015.06.064
- Gao, Y., Xue, X., Hu, D., Liu, W., Yuan, Y., Sun, H., et al. (2013). Inhibition of late sodium current by mexiletine: a novel pharmacotherapeutic approach in timothy syndrome. *Circ. Arrhythm. Electrophysiol.* 6, 614–622. doi: 10.1161/CIRCEP.113.000092

FUNDING

NIH, Supported in part by NIH/NHLBI grant P01 HL078931 and by UCLA Theodore Laubisch Endowment Fund.

- George, A. L. Jr. (2015). Misplaced brain sodium channels in heart kindle sudden death in epilepsy. *Circ. Arrhythm. Electrophysiol.* 8, 769–771. doi: 10.1161/CIRCEP.115.003261
- Guo, D., Lian, J., Liu, T., Cox, R., Margulies, K. B., Kowey, P. R., et al. (2011). Contribution of late sodium current (I_{Na-L}) to rate adaptation of ventricular repolarization and reverse use-dependence of QT-prolonging agents. *Heart Rhythm.* 8, 762–769. doi: 10.1016/j.hrthm.2010.12.026
- Harris, A. S., and Kokernot, R. H. (1950). Effects of diphenylhydantoin sodium (dilantin sodium) and phenobarbital sodium upon ectopic ventricular tachycardia in acute myocardial infarction. *Am. J. Physiol.* 163, 505–516.
- Harrison, D. C. (1985). Antiarrhythmic drug classification: new science and practical applications. *Am. J. Cardiol.* 56, 185–187. doi: 10.1016/0002-9149(85)90591-0
- Hilliard, F. A., Steele, D. S., Laver, D., Yang, Z., Le Marchand, S. J., Chopra, N., et al. (2010). Flecainide inhibits arrhythmogenic Ca²⁺ waves by open state block of ryanodine receptor Ca²⁺ release channels and reduction of Ca²⁺ spark mass. *J. Mol. Cell. Cardiol.* 48, 293–301. doi: 10.1016/j.yjmcc.2009.10.005
- Hitchcock, P., and Keown, K. K. (1959). The management of cardiac arrhythmias during cardiac surgery. *South Med. J.* 52, 702–706. doi: 10.1097/00007611-195906000-00015
- Huggins, R. A., Morse, R. A., Handley, C. A., and La Forge, M. (1949). The protective action of various agents against chloroform-epinephrine ventricular fibrillation. *J. Pharmacol. Exp. Ther.* 95, 312–317.
- Hund, T. J., and Mohler, P. J. (2015). Role of CaMKII in cardiac arrhythmias. *Trends Cardiovasc. Med.* 25, 392–397. doi: 10.1016/j.tcm.2014.12.001
- Isenberg, G., and Ravens, U. (1984). The effects of the Anemonia sulcata toxin (ATX II) on membrane currents of isolated mammalian myocytes. *J. Physiol.* 357, 127–149. doi: 10.1113/jphysiol.1984.sp015493
- Kalia, J., Milesco, M., Salvatierra, J., Wagner, J., Klint, J. K., King, G. F., et al. (2015). From foe to friend: using animal toxins to investigate ion channel function. *J. Mol. Biol.* 427, 158–175. doi: 10.1016/j.jmb.2014.07.027
- Kamath, G. S., and Mittal, S. (2008). The role of antiarrhythmic drug therapy for the prevention of sudden cardiac death. *Prog. Cardiovasc. Dis.* 50, 439–448. doi: 10.1016/j.pcad.2007.12.001
- Karagueuzian, H. S. (1995). *Antiarrhythmic Drugs: Mode of Action, Pharmacokinetic Properties, and Therapeutic Uses, 3rd Edn.* Philadelphia, PA: J.B. Lippincott Company.
- Karagueuzian, H. S., and Chen, P. S. (2001). Cellular mechanism of reentry induced by a strong electrical stimulus: implications for fibrillation and defibrillation. *Cardiovasc. Res.* 50, 251–262. doi: 10.1016/S0008-6363(00)0298-4
- Karagueuzian, H. S., Stepanyan, H., and Mandel, W. J. (2013). Bifurcation theory and cardiac arrhythmias. *Am. J. Cardiovasc. Dis.* 3, 1–16.
- Kim, J. G., Sung, D. J., Kim, H. J., Park, S. W., Won, K. J., Kim, B., et al. (2016). Impaired inactivation of L-Type Ca²⁺ current as a potential mechanism for variable arrhythmogenic liability of HERG K⁺ channel blocking drugs. *PLoS ONE* 11:e0149198. doi: 10.1371/journal.pone.0149198
- Koenig, X., Dysek, S., Kimbacher, S., Mike, A. K., Cervenka, R., Lukacs, P., et al. (2011). Voltage-gated ion channel dysfunction precedes cardiomyopathy development in the dystrophic heart. *PLoS ONE* 6:e20300. doi: 10.1371/journal.pone.0020300
- Kohlhardt, M., Bauer, B., Krause, H., and Fleckenstein, A. (1972). New selective inhibitors of the transmembrane Ca conductivity in mammalian myocardial fibres. Studies with the voltage clamp technique. *Experientia* 28, 288–289. doi: 10.1007/BF01928693
- Lerman, B. B., and Belardinelli, L. (1991). Cardiac electrophysiology of adenosine. Basic and clinical concepts. *Circulation* 83, 1499–1509. doi: 10.1161/01.CIR.83.5.1499
- Limpitkul, W. B., Dick, I. E., Joshi-Mukherjee, R., Overgaard, M. T., George, A. L. Jr., and Yue, D. T. (2014). Calmodulin mutations associated with long QT syndrome prevent inactivation of cardiac L-type Ca²⁺ currents and promote proarrhythmic behavior in ventricular myocytes. *J. Mol. Cell. Cardiol.* 74, 115–124. doi: 10.1016/j.yjmcc.2014.04.022
- Ling, H., Zhang, T., Pereira, L., Means, C. K., Cheng, H., Gu, Y., et al. (2009). Requirement for Ca²⁺/calmodulin-dependent kinase II in the transition from pressure overload-induced cardiac hypertrophy to heart failure in mice. *J. Clin. Invest.* 119, 1230–1240. doi: 10.1172/JCI38022
- Luczak, E. D., and Anderson, M. E. (2014). CaMKII oxidative activation and the pathogenesis of cardiac disease. *J. Mol. Cell. Cardiol.* 73, 112–116. doi: 10.1016/j.yjmcc.2014.02.004
- Madhvani, R. V., Angelini, M., Xie, Y., Pantazis, A., Suriany, S., Borgstrom, N. P., et al. (2015). Targeting the late component of the cardiac L-type Ca²⁺ current to suppress early afterdepolarizations. *J. Gen. Physiol.* 145, 395–404. doi: 10.1085/jgp.201411288
- Madhvani, R. V., Xie, Y., Pantazis, A., Garfinkel, A., Qu, Z., Weiss, J. N., et al. (2011). Shaping a new Ca²⁺ conductance to suppress early afterdepolarizations in cardiac myocytes. *J. Physiol.* 589, 6081–6092. doi: 10.1113/jphysiol.2011.219600
- Maier, L. S., Zhang, T., Chen, L., DeSantiago, J., Brown, J. H., and Bers, D. M. (2003). Transgenic CaMKII δ overexpression uniquely alters cardiac myocyte Ca²⁺ handling: reduced SR Ca²⁺ load and activated SR Ca²⁺ release. *Circ. Res.* 92, 904–911. doi: 10.1161/01.RES.0000069685.20258.F1
- Maltsev, V. A., Silverman, N., Sabbah, H. N., and Undrovinas, A. I. (2007). Chronic heart failure slows late sodium current in human and canine ventricular myocytes: implications for repolarization variability. *Eur. J. Heart Fail.* 9, 219–227. doi: 10.1016/j.ejheart.2006.08.007
- Maltsev, V. A., and Undrovinas, A. (2008). Late sodium current in failing heart: friend or foe? *Prog. Biophys. Mol. Biol.* 96, 421–451. doi: 10.1016/j.pbiomolbio.2007.07.010
- Markandeya, Y. S., and Kamp, T. J. (2015). Rational strategy to stop arrhythmias: early afterdepolarizations and L-type Ca²⁺ current. *J. Gen. Physiol.* 145, 475–479. doi: 10.1085/jgp.201511429
- Markandeya, Y. S., Tsubouchi, T., Hacker, T. A., Wolff, M. R., Belardinelli, L., and Balijepalli, R. C. (2016). Inhibition of late sodium current attenuates ionic arrhythmia mechanism in ventricular myocytes expressing LaminA-N195K mutation. *Heart Rhythm.* 13, 2228–2236. doi: 10.1016/j.hrthm.2016.08.007
- Marsman, R. F., Barc, J., Beekman, L., Alders, M., Dooijes, D., van den Wijngaard, A., et al. (2014). A mutation in CALM1 encoding calmodulin in familial idiopathic ventricular fibrillation in childhood and adolescence. *J. Am. Coll. Cardiol.* 63, 259–266. doi: 10.1016/j.jacc.2013.07.091
- Marx, S. O., Reiken, S., Hisamatsu, Y., Jayaraman, T., Burkhoff, D., Roseblit, N., et al. (2000). PKA phosphorylation dissociates FKBP12.6 from the calcium release channel (ryanodine receptor): defective regulation in failing hearts. *Cell* 101, 365–376. doi: 10.1016/S0092-8674(00)80847-8
- Mautz, F. R. (1936). The reduction of cardiac irritability by the epicardial and systemic administration of drugs as a protection in cardiac surgery. *J. Thoracic Surg.* 5, 612–628.
- McKeithan, W. L. R. D., Okolotowicz, K. J., Savtchenko, A., Kass, R. S., Cashman, J. R., and Mercola, M. (2016). “Development of a new antiarrhythmic therapeutic for long QT syndrome type 3,” *Paper Presented at: Stanford Drug Discovery Conference* (Palo Alto, CA: Stanford University).
- Mewes, T., and Ravens, U. (1994). L-type calcium currents of human myocytes from ventricle of non-failing and failing hearts and from atrium. *J. Mol. Cell. Cardiol.* 26, 1307–1320. doi: 10.1006/jmcc.1994.1149
- Moe, G. K. A. (1970). *Antiarrhythmic Drugs The Pharmacological Basis of Therapeutics*, Vol. 711. London: The Macmillan Co.
- Mollova, M. Y., Katus, H. A., and Backs, J. (2015). Regulation of CaMKII signaling in cardiovascular disease. *Front. Pharmacol.* 6:178. doi: 10.3389/fphar.2015.00178
- Morita, N., Lee, J. H., Bapat, A., Fishbein, M. C., Mandel, W. J., Chen, P. S., et al. (2011b). Glycolytic inhibition causes spontaneous ventricular fibrillation in aged hearts. *Am. J. Physiol. Heart Circ. Physiol.* 301, H180–H191. doi: 10.1152/ajpheart.00128.2011
- Morita, N., Lee, J. H., Xie, Y., Sovari, A., Qu, Z., Weiss, J. N., et al. (2011a). Suppression of re-entrant and multifocal ventricular fibrillation by the late sodium current blocker ranolazine. *J. Am. Coll. Cardiol.* 57, 366–375. doi: 10.1016/j.jacc.2010.07.045
- Morita, N., Sovari, A. A., Xie, Y., Fishbein, M. C., Mandel, W. J., Garfinkel, A., et al. (2009). Increased susceptibility of aged hearts to ventricular fibrillation during oxidative stress. *Am. J. Physiol. Heart Circ. Physiol.* 297, H1594–H1605. doi: 10.1152/ajpheart.00579.2009
- Morotti, S., Edwards, A. G., McCulloch, A. D., Bers, D. M., and Grandi, E. (2014). A novel computational model of mouse myocyte electrophysiology to assess the synergy between Na⁺ loading and CaMKII. *J. Physiol.* 592, 1181–1197. doi: 10.1113/jphysiol.2013.266676

- Mustroph, J., Neef, S., and Maier, L. S. (2016). CaMKII as a target for arrhythmia suppression. *Pharmacol Ther.* doi: 10.1016/j.pharmthera.2016.10.006. [Epub ahead of print].
- Nagatomo, T., January, C. T., and Makielski, J. C. (2000). Preferential block of late sodium current in the LQT3 DeltaKQP mutant by the class I(C) antiarrhythmic flecainide. *Mol. Pharmacol.* 57, 101–107.
- Nash, M. P., Mourad, A., Clayton, R. H., Sutton, P. M., Bradley, C. P., Hayward, M., et al. (2006). Evidence for multiple mechanisms in human ventricular fibrillation. *Circulation* 114, 536–542. doi: 10.1161/CIRCULATIONAHA.105.602870
- Nawrath, H., Klein, G., Rupp, J., Wegener, J. W., and Shainberg, A. (1998). Open state block by fendiline of L-type Ca^{++} channels in ventricular myocytes from rat heart. *J. Pharmacol. Exp. Ther.* 285, 546–552.
- Ono, N., Hayashi, H., Kawase, A., Lin, S. F., Li, H., Weiss, J. N., et al. (2007). Spontaneous atrial fibrillation initiated by triggered activity near the pulmonary veins in aged rats subjected to glycolytic inhibition. *Am. J. Physiol. Heart Circ. Physiol.* 292, 639–648. doi: 10.1152/ajpheart.00445.2006
- Peper, K., and Trautwein, W. (1967). The effect of aconitine on the membrane current in cardiac muscle. *Pflugers Arch. Gesamte Physiol. Menschen Tiere* 296, 328–336. doi: 10.1007/BF00362532
- Pezhouman, A. L. J. H., Cao, H., Belardinelli, L., Weiss, J. N., and Karagueuzian, H. S. (2014a). Superiority of the selective late INa blocker, GS967 over Ranolazine in suppressing oxidative atrial fibrillation in fibrotic aged hearts. *Circulation*. 130:A16247.
- Pezhouman, A. M. A., Angelini, M., Weiss, J. N., Olcese, R., and Karagueuzian, H. S. (2016). Roscovitine, a selective inhibitor of the Late L-Type calcium current suppresses early afterdepolarization-mediated triggered ventricular fibrillation: a new class of antiarrhythmic drug action. *Heart Rhythm*. 13:S5544.
- Pezhouman, A., Madadian, S., Stepanyan, H., Ghukasyan, H., Qu, Z., Belardinelli, L., et al. (2014b). Selective inhibition of late sodium current suppresses ventricular tachycardia and fibrillation in intact rat hearts. *Heart Rhythm*. 11, 492–501. doi: 10.1016/j.hrthm.2013.11.026
- Pezhouman, A., Singh, N., Song, Z., Nivala, M., Eskandari, A., Cao, H., et al. (2015). Molecular basis of hypokalemia-induced ventricular fibrillation. *Circulation* 132, 1528–1537. doi: 10.1161/CIRCULATIONAHA.115.016217
- Pogwizd, S. M., Hoyt, R. H., Saffitz, J. E., Corr, P. B., Cox, J. L., and Cain, M. E. (1992). Reentrant and focal mechanisms underlying ventricular tachycardia in the human heart. *Circulation* 86, 1872–1887. doi: 10.1161/01.CIR.86.6.1872
- Prystowsky, E. N., Padanilam, B. J., and Fogel, R. I. (2015). Treatment of atrial fibrillation. *JAMA* 314, 278–288. doi: 10.1001/jama.2015.7505
- Qu, Z., Xie, F., Garfinkel, A., and Weiss, J. N. (2000). Origins of spiral wave meander and breakup in a two-dimensional cardiac tissue model. *Ann. Biomed. Eng.* 28, 755–771. doi: 10.1114/1.1289474
- Qu, Z., Xie, L. H., Olcese, R., Karagueuzian, H. S., Chen, P. S., Garfinkel, A., et al. (2013). Early afterdepolarizations in cardiac myocytes: beyond reduced repolarization reserve. *Cardiovasc. Res.* 99, 6–15. doi: 10.1093/cvr/cvt104
- Riccio, M. L., Koller, M. L., and Gilmour, R. F. Jr. (1999). Electrical restitution and spatiotemporal organization during ventricular fibrillation. *Circ. Res.* 84, 955–963. doi: 10.1161/01.RES.84.8.955
- Rogawski, M. A., and Loscher, W. (2004). The neurobiology of antiepileptic drugs. *Nat. Rev. Neurosci.* 5, 553–564. doi: 10.1038/nrn1430
- Rosen, M. R., and Janse, M. J. (2010). Concept of the vulnerable parameter: the Sicilian Gambit revisited. *J. Cardiovasc. Pharmacol.* 55, 428–437. doi: 10.1097/fjc.0b013e3181bfaddc
- Rosen, M. R., Wit, A. L., and Hoffman, B. F. (1975). Electrophysiology and pharmacology of cardiac arrhythmias. VI. Cardiac effects of verapamil. *Am Heart J.* 89, 665–673. doi: 10.1016/0002-8703(75)90514-1
- Salameh, A., Dhein, S., Fleischmann, B., Grohe, C., Hescheler, J., Linz, K. W., et al. (2010). The aging heart: changes in the pharmacodynamic electrophysiological response to verapamil in aged rabbit hearts. *J. Physiol. Pharmacol.* 61, 141–151.
- Sato, D., Xie, L. H., Sovari, A. A., Tran, D. X., Morita, N., Xie, F., et al. (2009). Synchronization of chaotic early afterdepolarizations in the genesis of cardiac arrhythmias. *Proc. Natl. Acad. Sci. U. S. A.* 106, 2983–2988. doi: 10.1073/pnas.0809148106
- Shah, D. P., Baez-Escudero, J. L., Weisberg, I. L., Beshai, J. F., and Burke, M. C. (2012). Ranolazine safely decreases ventricular and atrial fibrillation in Timothy syndrome (LQT8). *Pacing Clin. Electrophysiol.* 35, e62–e64. doi: 10.1111/j.1540-8159.2010.02913.x
- Shryock, J. C., and Belardinelli, L. (1997). Adenosine and adenosine receptors in the cardiovascular system: biochemistry, physiology, and pharmacology. *Am. J. Cardiol.* 79, 2–10. doi: 10.1016/S0002-9149(97)00256-7
- Sicouri, S., Glass, A., Bellardinelli, L., and Antzelevitch, C. (2008). Antiarrhythmic effects of ranolazine in canine pulmonary vein sleeve preparations. *Heart Rhythm*. 5, 1019–1026. doi: 10.1016/j.hrthm.2008.03.018
- Singh, B. N., and Vaughan Williams, E. M. (1970a). A third class of anti-arrhythmic action. Effects on atrial and ventricular intracellular potentials, and other pharmacological actions on cardiac muscle, of MJ 1999 and AH 3474. *Br. J. Pharmacol.* 39, 675–687.
- Singh, B. N., and Vaughan Williams, E. M. (1970b). The effect of amiodarone, a new anti-anginal drug, on cardiac muscle. *Br. J. Pharmacol.* 39, 657–667. doi: 10.1111/j.1476-5381.1970.tb09891.x
- Song, Y., Shryock, J. C., Wagner, S., Maier, L. S., and Belardinelli, L. (2006). Blocking late sodium current reduces hydrogen peroxide-induced arrhythmogenic activity and contractile dysfunction. *J. Pharmacol. Exp. Ther.* 318, 214–222. doi: 10.1124/jpet.106.101832
- Swaminathan, P. D., Purohit, A., Hund, T. J., and Anderson, M. E. (2012). Calmodulin-Dependent protein kinase II: linking heart failure and arrhythmias. *Circ. Res.* 110, 1661–1677. doi: 10.1161/CIRCRESAHA.111.243956
- Swissa, M., Qu, Z., Ohara, T., Lee, M. H., Lin, S. F., Garfinkel, A., et al. (2002). Action potential duration restitution and ventricular fibrillation due to rapid focal excitation. *Am. J. Physiol. Heart Circ. Physiol.* 282, H1915–H1923. doi: 10.1152/ajpheart.00867.2001
- Szekeres, L., and Williams, E. M. (1962). Antifibrillatory action. *J. Physiol.* 160, 470–482. doi: 10.1113/jphysiol.1962.sp006860
- Task force of the working group on arrhythmias of the European Society on C (1991). The Sicilian Gambit. A new approach to the classification of antiarrhythmic drugs based on their actions on arrhythmogenic mechanisms. *Circulation* 84, 1831–1851.
- Thireau, J., Pasquié, J. L., Martel, E., Le Guennec, J. Y., and Richard, S. (2011). New drugs vs. old concepts: a fresh look at antiarrhythmics. *Pharmacol. Ther.* 132, 125–145. doi: 10.1016/j.pharmthera.2011.03.003
- Ulbricht, W. (2005). Sodium channel inactivation: molecular determinants and modulation. *Physiol. Rev.* 85, 1271–1301. doi: 10.1152/physrev.00024.2004
- Undrovinas, A., and Maltsev, V. A. (2008). Late sodium current is a new therapeutic target to improve contractility and rhythm in failing heart. *Cardiovasc. Hematol. Agents Med. Chem.* 6, 348–359. doi: 10.2174/187152508785909447
- van Hamel, K. N. (2003). Do arrhythmias still deserve our intellectual efforts? The 2002 wenckebach lecture of the netherlands society of cardiology. *Neth. Heart J.* 11, 301–308.
- van Veldhuisen, D. J., de Graeff, P. A., Remme, W. J., and Lie, K. I. (1996). Value of digoxin in heart failure and sinus rhythm: new features of an old drug? *J. Am. Coll. Cardiol.* 28, 813–819.
- Vaughan Williams, E. M. (1970). “Classification of antiarrhythmic drugs,” in *Cardiac Arrhythmias*, eds F.-J. E. Sandoe, E. Olesen, and K. H. Södertälje (Elsinore: AB Astra), 449–472.
- Vaughan Williams, E. M. (1992). Classifying antiarrhythmic actions: by facts or speculation. *J. Clin. Pharmacol.* 32, 964–977. doi: 10.1002/j.1552-4604.1992.tb03797.x
- Viola, H. M., Davies, S. M., Filipovska, A., and Hool, L. C. (2013). L-type Ca^{2+} channel contributes to alterations in mitochondrial calcium handling in the mdx ventricular myocyte. *Am. J. Physiol. Heart Circ. Physiol.* 304, H767–H775. doi: 10.1152/ajpheart.00700.2012
- Voigt, N., Li, N., Wang, Q., Wang, W., Trafford, A. W., Abu-Taha, I., et al. (2012). Enhanced sarcoplasmic reticulum Ca^{2+} leak and increased Na^{+} - Ca^{2+} exchanger function underlie delayed afterdepolarizations in patients with chronic atrial fibrillation. *Circulation* 125, 2059–2070. doi: 10.1161/CIRCULATIONAHA.111.067306
- Wagner, S., Maier, L. S., and Bers, D. M. (2015). Role of sodium and calcium dysregulation in tachyarrhythmias in sudden cardiac death. *Circ. Res.* 116, 1956–1970. doi: 10.1161/CIRCRESAHA.116.304678
- Waldo, A. L., Camm, A. J., deRuyter, H., Freidman, P. L., MacNeil, D. J., Pitt, B., et al. (1995). Survival with oral d-sotalol in patients with left ventricular dysfunction after myocardial infarction: rationale,

- design, and methods (the SWORD trial). *Am. J. Cardiol.* 75, 1023–1027. doi: 10.1016/S0002-9149(99)80717-6
- Ward, C. A., and Giles, W. R. (1997). Ionic mechanism of the effects of hydrogen peroxide in rat ventricular myocytes. *J. Physiol.* 500(Pt 3), 631–642. doi: 10.1113/jphysiol.1997.sp022048
- Weiss, J. N., Chen, P. S., Qu, Z., Karagueuzian, H. S., and Garfinkel, A. (2000). Ventricular fibrillation: how do we stop the waves from breaking? *Circ. Res.* 87, 1103–1107. doi: 10.1161/01.RES.87.12.1103
- Weiss, J. N., Garfinkel, A., Karagueuzian, H. S., Nguyen, T. P., Olcese, R., Chen, P. S., et al. (2015). Perspective: a dynamics-based classification of ventricular arrhythmias. *J. Mol. Cell. Cardiol.* 82, 136–152. doi: 10.1016/j.yjmcc.2015.02.017
- Weiss, J. N., Karma, A., Shiferaw, Y., Chen, P. S., Garfinkel, A., and Qu, Z. (2006). From pulsus to pulseless: the saga of cardiac alternans. *Circ. Res.* 98, 1244–1253. doi: 10.1161/01.RES.0000224540.97431.f0
- Willius, F. A., and Keys, T. E. (1941). *Jean-Baptiste de Senac Classics of Cardiology*, New York, NY: Dover Publications, Inc (One).
- Wu, T. J., Ong, J. J., Hwang, C., Lee, J. J., Fishbein, M. C., Czer, L., et al. (1998). Characteristics of wave fronts during ventricular fibrillation in human hearts with dilated cardiomyopathy: role of increased fibrosis in the generation of reentry. *J. Am. Coll. Cardiol.* 32, 187–196. doi: 10.1016/S0735-1097(98)00184-3
- Xie, L. H., Chen, F., Karagueuzian, H. S., and Weiss, J. N. (2009). Oxidative stress-induced afterdepolarizations and calmodulin kinase II signaling. *Circ. Res.* 104, 79–86. doi: 10.1161/CIRCRESAHA.108.183475
- Xie, Y., Sato, D., Garfinkel, A., Qu, Z., and Weiss, J. N. (2010). So little source, so much sink: requirements for afterdepolarizations to propagate in tissue. *Biophys. J.* 99, 1408–1415. doi: 10.1016/j.bpj.2010.06.042
- Yarotsky, V., and Elmslie, K. S. (2007). Roscovitine, a cyclin-dependent kinase inhibitor, affects several gating mechanisms to inhibit cardiac L-type ($\text{Ca}_v1.2$) calcium channels. *Br. J. Pharmacol.* 152, 386–395. doi: 10.1038/sj.bjp.0707414
- Yazawa, M., and Dolmetsch, R. E. (2013). Modeling Timothy syndrome with iPSC cells. *J. Cardiovasc. Transl. Res.* 6, 1–9. doi: 10.1007/s12265-012-9444-x
- Yazawa, M., Hsueh, B., Jia, X., Pasca, A. M., Bernstein, J. A., Hallmayer, J., et al. (2011). Using induced pluripotent stem cells to investigate cardiac phenotypes in Timothy syndrome. *Nature* 471, 230–234. doi: 10.1038/nature09855
- Yue, D. T., Backx, P. H., and Imredy, J. P. (1990). Calcium-sensitive inactivation in the gating of single calcium channels. *Science* 250, 1735–1738. doi: 10.1126/science.2176745
- Zhang, T., Maier, L. S., Dalton, N. D., Miyamoto, S., Ross, J. Jr., Bers, D. M., et al. (2003). The deltaC isoform of CaMKII is activated in cardiac hypertrophy and induces dilated cardiomyopathy and heart failure. *Circ. Res.* 92, 912–919. doi: 10.1161/01.RES.0000069686.31472.C5
- Zhou, Y. Y., Lakatta, E. G., and Xiao, R. P. (1998). Age-associated alterations in calcium current and its modulation in cardiac myocytes. *Drugs Aging.* 13, 159–171. doi: 10.2165/00002512-199813020-00007

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Karagueuzian, Pezhouman, Angelini and Olcese. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.