



# Baicalein Exerts Neuroprotective Effects in FeCl<sub>3</sub>-Induced Posttraumatic Epileptic Seizures *via* Suppressing Ferroptosis

Qin Li<sup>1,2,3,4</sup>, Qiu-Qi Li<sup>1,2,3,4</sup>, Ji-Ning Jia<sup>1,2,3,4</sup>, Qian-Yi Sun<sup>1,2,3,4</sup>, Hong-Hao Zhou<sup>1,2,3,4</sup>, Wei-Lin Jin<sup>5,6,7\*</sup> and Xiao-Yuan Mao<sup>1,2,3,4\*</sup>

<sup>1</sup> Department of Clinical Pharmacology, Xiangya Hospital, Central South University, Changsha, China, <sup>2</sup> Institute of Clinical Pharmacology, Central South University, Hunan Key Laboratory of Pharmacogenetics, Changsha, China, <sup>3</sup> Engineering Research Center of Applied Technology of Pharmacogenomics, Ministry of Education, Changsha, China, <sup>4</sup> National Clinical Research Center for Geriatric Disorders, Changsha, China, <sup>5</sup> Centers for Translational Medicine, Ruikang Hospital, Guangxi University of Chinese Medicine, Nanning, China, <sup>6</sup> Institute of Nano Biomedicine and Engineering, Department of Instrument Science and Engineering, Key Laboratory for Thin Film and Microfabrication Technology of Ministry of Education, School of Electronic Information and Electronic Engineering, Shanghai Jiao Tong University, Shanghai, China, <sup>7</sup> Shaanxi Key Laboratory of Brain Disorders and Institute of Basic and Translational Medicine, Xi'an Medical University, Xi'an, China

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#### \*Correspondence:

Xiao-Yuan Mao xiaoyuanm@csu.edu.cn maoxiaoyuan2011@163.com Wei-Lin Jin weilinjin@sjtu.edu.cn

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Li Q, Li Q-Q, Jia J-N, Sun Q-Y, Zhou H-H, Jin W-L and Mao X-Y (2019) Baicalein Exerts Neuroprotective Effects in FeCl<sub>3</sub>-Induced Posttraumatic Epileptic Seizures via Suppressing Ferroptosis. Front. Pharmacol. 10:638. doi: 10.3389/fphar.2019.00638 Posttraumatic epilepsy (PTE) is a prevalent type of acquired epilepsy secondary to traumatic brain injury, and is characterized by repeated seizures. Traditional antiepileptic drugs have minimal response in preventing posttraumatic epileptic seizures. It is essential for the development of new therapeutic strategy. Our previous work disclosed a potent neuroprotective role of baicalein, a flavonoid extracted from Scutellaria baicalensis Georgi, against inherited epilepsy in rats. Whether baicalein has protective potential in posttraumatic epileptic seizures and the possible molecular mechanism remain elusive. Additionally, the brain is vulnerable to lipid peroxidation-induced damage due to high consumption of oxygen and abundant polyunsaturated fatty acids in neuronal membranes. Our present investigation aimed to elucidate whether baicalein exerts neuroprotective effects on posttraumatic epileptic seizures by inhibiting ferroptosis, a newly discovered lipid peroxidation-dependent cell death modality. We found that baicalein significantly reduced seizure score, number of seizures, and average seizure duration in an iron chloride (FeCl<sub>3</sub>)-induced PTE mouse model. The neuroprotective effect of baicalein was also validated in a ferric ammonium citrate (FAC)-induced HT22 hippocampal neuron damage model. Moreover, in vitro, baicalein could remarkably decrease ferroptotic indices (lipid reactive oxygen species, 4-hydroxynonenal, and prostaglandin endoperoxide synthase 2) and inhibit the expression of 12/15-lipoxygenase (12/15-LOX) in an iron-induced HT22 cell damage model. These findings were also validated in a mouse PTE model. It was concluded that baicalein exerted neuroprotective effects against posttraumatic epileptic seizures via suppressing ferroptosis and 12/15-LOX was likely to be involved in baicalein's neuroprotection.

Keywords: posttraumatic epileptic seizures, baicalein, 12/15-lipoxygenase, lipid peroxidation, ferroptosis, neuroprotective

## INTRODUCTION

Posttraumatic epilepsy (PTE), a recurrent seizure disorder, is the consequence of traumatic brain injury (TBI) (Agrawal et al., 2006; Kharatishvili and Pitkänen, 2010; Lamar et al., 2014; Pitkanen et al., 2014). It is estimated that nearly 80% of patients suffer from seizures within 1 year post-TBI (Volavka, 2011). Epidemiological studies have also shown that more than 3 million people have PTE globally, comprising approximately 5-6% of all epilepsy patients (Pi et al., 2014; Szaflarski et al., 2014; Singh and Trevick, 2016; Chartrain et al., 2017). Additionally, PTE is the most common cause of new onset epilepsy in young adults (Annegers and Coan, 2000). PTE not only significantly reduces the quality of life of patients physically, cognitively, and emotionally, but also increases the risk of mortality (Englander et al., 2009; Christensen, 2015; Xu et al., 2017). Traditional antiepileptic drugs have minimal response in seizures after TBI (Chartrain et al., 2017). Conceptually, the latency between injury and seizures is usually very short. It is hoped that effective treatment could be administered at some point within this latency to stop the process, so that posttraumatic epileptic seizures will not occur. Currently, neuroprotection is increasingly considered as a promising therapy for preventing and treating epilepsy (Acharya et al., 2008; McKinnon et al., 2018). Identification of potential compounds or methods that provide neuroprotective effects on posttraumatic epileptic seizures is important.

Traditional Chinese medicine (TCM) has a long history of preventing and curing diseases (Wu et al., 2011; Mao et al., 2018). Recently, TCM has demonstrated good prospects for improving neurological diseases, cancer, and cardiovascular disease due to its easy penetration of the blood-brain barrier and substantial existence in nature (Chen et al., 2019; Pan and Ma, 2019). TCM possessed various beneficial biological effects, including antioxidant, anti-inflammatory, antitumor, etc. (Li and Zhang, 2008; Pan et al., 2011; Zhang et al., 2019). For example, baicalein, curcumin, and resveratrol exert neuroprotection against neurological diseases, such as epilepsy, ischemic brain damage, and Alzheimer's disease (AD) (Ahn et al., 2007; Liu et al., 2012; Liu et al., 2015; Lu and Wang, 2015; Gu et al., 2016; Li et al., 2016; Andrade et al., 2018). Particularly, baicalein acted neuroprotective effect in inherited epilepsy (Mao et al., 2014) and in a pilocarpine-induced acute epileptic rats model (Liu et al., 2012) by inhibiting oxidative stress. However, little is known about the neuroprotective effects of baicalein in posttraumatic epileptic seizures. Although it was previously reported that baicalein could protect hippocampal neurons in the PTE model through its antioxidant capacity (Hamada et al., 1993), its specific molecular mechanism remains unclear.

The antioxidant capacity is one of the important potential mechanisms of TCM for neuroprotective effects. For instance, baicalein was reported to protect cerebral ischemia/reperfusion injury *via* inhibiting the function of NF-κB (Liu et al., 2015). And a large body of evidence has demonstrated that higher levels of oxidative stress markers including elevated superoxide dismutase activity, lipid oxidation products, and protein nitrotyrosine exist in the brain of some neurological diseases (Islam, 2017; Poprac et al., 2017; Zhang et al., 2018a). These indicate that oxidative stress is an important target for the neuroprotective effects of many TCM including baicalein. Specifically, due to highly enriched polyunsaturated fatty acids (PUFAs) in the brain, lipid peroxidation is likely to more frequently occur during oxidative damage (Bazinet and Laye, 2014). It has been reported that lipoxygenases (LOXs) serve as a type of key enzymes which involve in the synthesis of lipid hydroperoxide and the facilitation of catalyzing the oxidation of PUFAs (Bazinet and Laye, 2014; Wenzel et al., 2017). As a major subtype of LOX family, 12/15-LOX mediates the oxidation of arachidonic acid (Wenzel et al., 2017) and inhibits the intracellular lipid deposition in foam cells (Belkner et al., 2005), suggesting that it is a key enzyme in lipid peroxidation. Additionally, baicalein was previously found to significantly suppress the expression of 12/15-LOX and protect neuronal cells from death in various neurological diseases such as ischemic brain damage, and AD (van Leyen et al., 2006; Gu et al., 2016). Therefore, our present work aimed to explore whether baicalein could exert neuroprotective effects on (FeCl<sub>3</sub>)-induced posttraumatic epileptic seizures by inhibiting 12/15-LOXmediated lipid peroxidation. Recently, lipid peroxidation has been found to cause a novel type of cell death, ferroptosis, which constitutes a cell death pathway that is genetically, morphologically, and biochemically different from apoptosis and autophagy (Dixon et al., 2012; Yang and Stockwell, 2016; Conrad et al., 2018). Hence, we also explored whether baicalein could abrogate ferroptosis by 12/15-LOX-mediated lipid peroxidation and finally exert neuroprotective effects on posttraumatic epileptic seizures.

Our data indicated that both baicalein and ferroptosis inhibitors could ameliorate epileptic seizure behavior in FeCl<sub>3</sub>-induced PTE model mice. Furthermore, we found that baicalein could exert neuroprotective effects in FAC-induced HT22 cell damage model and FeCl<sub>3</sub>-induced seizures by suppressing ferroptosis and 12/15-LOX is involved in baicalein's neuroprotection. We believe that targeting ferroptosis could promote the clinical application of baicalein and might contribute substantially to the prevention of posttraumatic epileptic seizures.

#### MATERIALS AND METHODS

#### **Chemicals and Reagents**

Dulbecco's modified Eagle's medium (DMEM) and fetal bovine serum (FBS) were purchased from GIBCO (Grand Island, NY, USA). Ferric ammonium citrate (FAC) and iron chloride (FeCl<sub>3</sub>) were obtained from Sigma-Aldrich (St. Louis, MO, USA). Baicalein, erastin, ferrostatin-1 (Fer-1), and liproxstatin-1 (Lipo-1) were purchased form Selleck Chemicals (Houston, TX, USA).

Abbreviations: PTE, posttraumatic epilepsy; TBI, traumatic brain injury; AD, Alzheimer's disease; PD, Parkinson's disease; TCM, traditional Chinese medicine; PUFAs, polyunsaturated fatty acids; 12/15-LOX, 12/15-lipoxygenase; DMEM, Dulbecco's modified Eagle's medium; FBS, fetal bovine serum; BAC, baicalein; FAC, ferric ammonium citrate; Fecl<sub>3</sub>, iron chloride; Fer-1, ferrostatin-1; Lipo-1, liproxstatin-1; CCK8, cell counting kit-8; PBS, phosphate buffer saline; HBSS, Hanks' balanced salt solution; TBST, tris-buffered saline tween; PTGS2, prostaglandin endoperoxide synthase 2; Lipid ROS, lipid reactive oxygen species; MDA, malondialdehyde; 4-HNE, 4-hydroxynonenal; GPX4, glutathione peroxidase 4.

# Animals and Establishment of FeCl<sub>3</sub>-Induced PTE Model

All adult male C57/BL6 mice weighing 18-22g were obtained from the Experimental Animal Center of Central South University, China. The protocol of animal experiment was approved by the Medical Ethics Committee of Xiangya Hospital and performed in accordance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals. All efforts had been made to minimize the animal's suffering. All animals were housed in cages with a 12/12-h light/dark cycle and other standard laboratory conditions, including a room temperature of  $23 \pm 1^{\circ}$ C and access to food and water *ad libitum*. After anesthetization with 10% chloral hydrate (0.3 ml/100 g, i.p.) and stereotaxic placement, the mice were drilled with burr holes in the surface of the skull at stereotaxically marked sites.  $FeCl_3$  (5 µl containing 50 mM FeCl\_3) dissolved in phosphate buffer saline (PBS) was stereotaxically injected in 5 min through the burr hole (coordinates anteroposterior -1.3 mm; lateral 2.0 mm; and ventral 1.6 mm) in the somatosensory region of the cortex. The needle was kept in place for an additional 10 min after injection to avoid reflux. Animals were randomly divided into six groups as follows: 1) control group (n = 6) was given vehicle intracranial injection (PBS 5  $\mu$ ); 2) FeCl<sub>3</sub> group (n = 6) was given 5 µl 50 mM FeCl<sub>3</sub>; 3) and 4) baicalein groups were pretreated with baicalein 50 mg/kg (n = 6) and 100 mg/kg (n = 6) 30 min prior to FeCl<sub>3</sub> administration, respectively; 5) ferroptosis inhibitor group (n = 6) was administered continuously with 10 mg/kg Lipo-1 3 d prior to FeCl<sub>3</sub> administration; and 6) baicalein administration group (n = 6) was given 100 mg/kg baicalein once by intraperitoneal injection 30 min prior to 5 µl PBS administration.

### **Behavior Tasks**

The behavioral changes of mice were recorded within 6 h after injection of FeCl<sub>3</sub>. The scoring system described by Racine was used for evaluating the seizure severity of the treated mice (Racine, 1972; Sato et al., 1990). Racine stages were described as follows: stage 0, no response; stage 1, ear and facial twitching; stage 2, convulsive twitching axially through the body; stage 3, myoclonic jerks and rearing; stage 4, turning over onto the side, wild running, and wild jumping; stage 5, generalized tonic-clonic seizures; and stage 6, death. If the stage 3 seizure or higher were observed, animals were considered to have seizures. Over an observation time of 6 h after FeCl<sub>3</sub> injection, the number of seizures and the single episode duration of individual seizures were recorded for each mouse simultaneously by two trained observers blinded to the animals' drug treatment designs.

# **Cell Cultures and Drug Treatments**

Immortalized mouse hippocampal HT22 cells were cultured in DMEM (C11995500BT, Gibco, USA) containing 10% FBS and 100 units of penicillin and 100 µg/ml streptomycin. Cells were then incubated at 37°C in a 5% CO<sub>2</sub> atmosphere. For experiments, HT22 cells were pretreated with 12.5 µM Fer-1 (S7243, Selleck, USA) or 1 µM Lipo-1 (S7699, Selleck, USA) for 2 h. Then, 300 µM FAC (F5879, Sigma, USA) was added to the medium for 36 h.

# **Cell Viability Assay**

Cell viability assays were performed using Cell Counting Kit-8 (CCK8) (C0038, Beyotime Institute of Biotechnology, China) following the manufacturer's instructions. Briefly, cells were seeded in 96-well plates in a medium containing 10% FBS. After drug treatment at the indicated time points, 10  $\mu$ l CCK8 solution was added to each well and incubated for 1 h. Cell viability was finally measured using a microplate reader at a wavelength of 450 nm.

# Cell Morphological Observation by Inverted Microscope

HT22 cells were seeded in 24-well plates at a density of 15% and cultured overnight. The plated cells were pretreated with 32  $\mu$ M baicalein (S2268, Selleck, USA) for 2 h. Then, medium containing FAC (F5879, Sigma, USA) or erastin (S7242, Selleck, USA) at the final concentration of 300  $\mu$ M and 500 nM were added and cultured for 36 and 8 h, respectively. Alteration of morphology in treated cells was observed under an inverted microscope.

### Detection of Lipid Reactive Oxygen Species (Lipid ROS) by Flow Cytometry

After treatment at the indicated time, cells were washed with PBS, harvested by trypsinization, resuspended in 500 µl Hanks' balanced salt solution (HBSS) (14065056BT, Gibco, USA) containing 2 µl C11-BODIPY 581/591 (D3861, Thermo Fisher, USA), and incubated for 15 min at 37°C in a tissue culture incubator. Cells were washed with HBSS, centrifuged at 3,000 × *g* for 5 min, and then resuspended in 200 µl PBS. A minimum of 10,000 events per replicate were collected and analyzed using a flow cytometer. Data were collected from the FL1 channel, and subsequently analyzed with FlowJo software.

# **Real-Time RT-PCR Analysis**

Total RNA was extracted using TRIzol reagent (Invitrogen) according to the manufacturer's procedure. The extracted RNA was reverse transcribed into cDNA by a reverse transcription kit (Perfect Real Time) (RR047A, Takara Bio, Japan). Real-time PCR was performed using double-stranded DNA dye SYBR Green (RR091A, Takara, Japan) and LightCycler 480 Roche system. PCR conditions were as follows: 30s hot-start at 95°C followed by 40 cycles of 5s at 95°C, 30s at 55°C and 30s at 72°C; 30s melting curve at 95°C. All samples were analyzed in triplicate, and gene expression was normalized to  $\beta$ -actin mRNA levels. Primer sequences designed to detect specific genes are as follows: PTGS2 forward: 5'-GGGAGTCTGGAACAT TGTGAA-3', reverse: 5'-GTGCACATTGTAAGTAGGTGGACT-3'; ATCB forward: 5'-GTGACGTTGACATCCGTAAAGA-3', reverse: 5'-GCCGGACTCATCGTACTCC-3'.

#### Immunofluorescence

HT22 cells were seeded in a 35-mm dish at a certain density. After drug treatments, cells were fixed in 4% paraformaldehyde for 30 min, permeabilized in 0.2% Triton X-100 for 10 min, and

blocked in donkey serum (SL034, Solarbio, China) for 30 min, all at room temperature. Cells were then incubated with primary antibody (12/15-LOX, rabbit, BS8547, 1:100 dilution, Bioworld) in blocking buffer at 4°C overnight. PBS replaced the primary antibody in negative controls. After washing three times with PBS, the cells were then incubated with secondary antibody. The secondary antibody was donkey anti-rabbit Alexa Fluor 488 (711-545-152, Jackson ImmunoResearch, USA), and used at 1/250 dilution for 45 min at room temperature. DAPI (S2110, Solarbio, China) was applied to stain the cell nucleus for 5 min. Immunofluorescence-stained cells were examined under a laserscanning confocal microscope (Nikon, Japan).

### Western Blot

Cell lysates were prepared in radio immunoprecipitation assay lysis buffer with fresh protease and phosphatase inhibitors mixture. Protein concentrations were measured by BCA protein assay reagent (P0006, Beyotime Biotechnology, China). In brief, protein (20 µg) was resolved by sodium dodecyl sulfate-polyacrylamide gel electrophoresis and transferred to polyvinylidene fluoride membranes. After blocking in tris-buffered saline tween (TBST) buffer with 5% nonfat milk for 1 h, the membranes were incubated with the following primary antibodies: glutathione peroxidase4 (GPX4) (rabbit, 17 kDa, ab125066, 1:5,000 dilution, Abcam), 4-hydroxynonenal (4-HNE) (rabbit, 17-76 kDa, ab46545, 1:3,000 dilution, Abcam), 12/15-LOX (rabbit, ~75 kDa, BS8547, 1:1,000 dilution, Bioworld), and β-actin (mouse, 43 kDa, A5441, 1:5,000 dilution, Sigma), overnight at 4°C. The next day, after being washed in TBST buffer three times, the membranes were further incubated with secondary antibodies (mouse, A9044, 1:10,000 dilution, Sigma; rabbit, A9169, 1:10,000 dilution, Sigma). Immunoreactive bands were visualized by the ChemiDoc XRS+ imaging system (Bio-Rad, USA). β-Actin was used as an internal control. For quantification of the protein bands, we utilized Image J software (National Institutes of Health, USA) to obtain densitometric values.

#### **Statistical Analysis**

Results were expressed as mean  $\pm$  SD. Statistical significance was determined by analysis of variance (ANOVA) followed by a multiple comparison test with a Bonferroni test. P-values of less than 0.05 were considered statistically significant.

### RESULTS

# Baicalein Attenuates FeCl<sub>3</sub>-Induced Epileptic Seizure Behavior

We first examined whether baicalein could attenuate epileptic seizure behavior in the FeCl<sub>3</sub>-induced PTE model. We used four different concentrations (20, 35, 50, 100 mM) of FeCl<sub>3</sub> to investigate the FeCl<sub>3</sub>-induced PTE model. Mice with grade 4–5 seizures were considered successful. It was found that an administration of a dose of 50 mM FeCl<sub>3</sub> or 100 mM FeCl<sub>3</sub> (data not shown) resulted in increasing convulsive activity, leading to generalized tonic–clonic seizure (**Figure 1A**). And we also observed that the number and duration time of seizure

were significantly elevated with the increase in dose of FeCl<sub>3</sub> (**Figure 1B** and **C**). However, the mortality rate of mice was high in the 100 mM FeCl<sub>3</sub> injection group (data not shown). So, we performed the FeCl<sub>3</sub>-induced PTE model by injecting 5  $\mu$ l of 50 mM FeCl<sub>3</sub> into the sensorimotor cortex of the mice. All of the mice in each group survived without any complications at the end of the experiment period. We then pretreated with different concentrations of baicalein (50 and 100 mg/kg) before administration of a dose of 50 mM FeCl<sub>3</sub>. The data showed that preadministration of 100 mg/kg baicalein significantly reduced the seizure score, number of seizures, and seizure duration compared to the FeCl<sub>3</sub>-induced posttraumatic epileptic seizure group (**Figure 1D**–**F**). Moreover, the 100 mg/kg dose of baicalein had no toxic effect on sham control mice.

# Baicalein Alleviates FAC-Induced Neuronal Damage by Inhibiting Ferroptosis

To further elucidate the potential neuroprotective mechanism of baicalein against posttraumatic epileptic seizures, we then established an in vitro iron-induced neuronal injury model using FeCl<sub>3</sub>. However, there was no significant effect after intervention with different concentrations of FeCl<sub>3</sub> in HT22 cells (Supplementary Figure S1). Alternatively, we used HT22 cells with FAC as a source of iron according to the previous descriptions (Lee et al., 2016; Lee et al., 2018). The toxic effects of FAC on HT22 cells were detected via CCK-8 assay. It was obviously found that FAC at a concentration of  $300 \ \mu\text{M}$  for 36 h exerted the most potent impairment on HT22 hippocampal neurons (nearly 50% reduction in comparison with vehicle control), as indicated in Figure 2A. The cytotoxic effect was attenuated by pretreatment with baicalein for 2 h in a dose-dependent manner (Figure 2B). It was noteworthy that baicalein at a concentration of 32  $\mu$ M exerted the most obvious protective effects. Morphologically, it was also found that 32 µM baicalein improved cell viability the most in HT22 cells under FAC exposure (Figure 2C). Furthermore, in HT22 cells following FAC treatment, baicalein (32 µM) robustly suppressed ferroptotic indices, including lipid ROS, 4-HNE (a common lipid peroxidation by-product) levels, and the relative mRNA expression of PTGS2 (a downstream marker of ferroptosis), as compared with treatment with Fer-1 or Lipo-1 (Figure 2D-F). Additionally, we also found that baicalein could significantly upregulate the expression of GPX4 (a crucial regulator of ferroptosis) (Figure 2G). The above-mentioned results were also validated in HT22 neurons after exposure of erastin (a specific ferroptotic inducer) exposure (Figure 3A-G).

#### 12/15-LOX is Involved in the Inhibition of Ferroptosis in FAC-Induced HT22 Cell Injury Model by Baicalein

Given that baicalein was reported to be a selective inhibitor of 12/15-LOX (Deschamps et al., 2006; Li et al., 2019), we next investigated whether or not 12/15-LOX was involved in ferroptosis in the FAC-induced HT22 cell damage model. To achieve this, we analyzed expression levels of 12/15-LOX Li et al.



presented as seizure score, number of seizures, and average seizure duration (Sec), n = 6. (D–F) Intraperitoneal injection of different doses of baicalein (50 and 100 mg/kg) 30 min prior to the injection of 50 mM FeCl<sub>3</sub>. The seizure score, number of seizures, and average seizure duration (Sec) were reduced significantly at different doses of baicalein, n = 6. \*p < 0.05, \*\*p < 0.01, and \*\*\*p < 0.001.

in the FAC- or erastin-induced cell impairment model by immunofluorescence. Compared with the FAC or erastin group, significantly decreased immunofluorescence signals for 12/15-LOX were present in the baicalein pretreatment group, which was consistent with the effect of PD146176 (a specific 12/15-LOX inhibitor) on significantly reducing the immunofluorescence signals of 12/15-LOX (Figure 4A). Meanwhile, as shown in Figure 4B, 1 µM PD146176 remarkably reversed FAC-induced cell damage. Consistently, we observed that pretreatment with PD146176 could alleviate FAC- and erastin-induced cell damage in morphology (Figure 4C and D). Additionally, we also found that pretreatment with PD146176, Fer-1, and Lipo-1 remarkably suppressed the production of lipid ROS induced by FAC and erastin (Figure 4E and F). The above results indicated that baicalein-mediated inhibition of 12/15-LOX contributed to the inhibition of FAC-induced ferroptosis.

#### Baicalein Counteracts FeCl<sub>3</sub>-Induced Epileptic Seizures by Inhibiting Ferroptosis

Next, we validated the results of the *in vitro* experiments in the FeCl<sub>3</sub>-induced seizure model. The results of reduced

production of 4-HNE and lipid ROS after pretreatment of baicalein and ferroptosis inhibitors suggested that ferroptosis was involved in the neuroprotective effects of baicalein. Lipo-1, a fat-soluble small molecule compound, is a specific inhibitor of ferroptosis. So, we evaluated the effects of Lipo-1 in the FeCl<sub>3</sub>-induced PTE seizure model. Lipo-1 was administered to mice at a dose of 10 mg/kg daily for 3 days prior to administration of FeCl<sub>3</sub>. Similar to the previous results, we found that Lipo-1 could significantly reduce the seizure score of mice in the FeCl<sub>3</sub>-induced PTE seizure model (Figure 5A). We next collected hippocampus tissue from those mice and compared the levels of the production of 4-HNE and the mRNA expression of PTGS2. As Figure 5B and C shows, in the FeCl<sub>3</sub>-induced PTE seizure model, the production of 4-HNE and the mRNA expression of PTGS2 was significantly diminished after pretreatment with baicalein and Lipo-1 compared to the group treatment with FeCl<sub>3</sub> alone. In addition, we also examined the changes of GPX4. It was found that the expression of GPX4 was significantly down-regulated, but this phenomenon was reversed when pretreated with baicalein and Lipo-1 (Figure 5D). Consistent with the in vitro results, expression of 12/15-LOX was increased in hippocampus



**FIGURE 2** Baicalein attenuates FAC-induced neuronal damage by inhibiting ferroptosis. (A) Cell viability was measured with CCK8 assay, in which HT22 cells were incubated with various concentrations (37.5, 75, 150, and 300  $\mu$ M) of FAC for different periods of time (12–36 h). (B) HT22 cells were incubated with different concentrations (1, 2, 4, 8, 16, and 32  $\mu$ M) of baicalein for 2 h, prior to incubation with 300  $\mu$ M of FAC for 36 h. (C) FAC exposure caused cell shrinkage and nuclear condensation. Baicalein pretreatment reduced cell death. Scale bar: 200  $\mu$ m (D) after 2 h pretreatment with 32  $\mu$ M baicalein, 12.5  $\mu$ M Fer-1, and 1  $\mu$ M Lipo-1, lipid ROS in HT22 cells was assessed by FACS analysis of C11-BODIPY fluorescence after treatment with FAC for 36 h. (E) Representative Western blots of 4-HNE with 32  $\mu$ M baicalein and ferroptosis inhibitors pretreatment for 2 h in the FAC-induced HT22 cell injury model. (F) The relative mRNA expression of PTGS2 was reduced after baicalein and ferroptosis inhibitors pretreatment. (G) Representative Western blots of GPX4 with 32  $\mu$ M baicalein and ferroptosis inhibitors pretreatment (for 2 h in the FAC-induced HT22 cell might baicalein and ferroptosis inhibitors pretreatment. (G) Representative of three independent experiments with similar results, \*p < 0.05, \*\*p < 0.01, and \*\*\*p < 0.001.

from the FeCl<sub>3</sub>-induced PTE seizure model, and pretreatment with baicalein remarkably inhibited the increase of 12/15-LOX (**Figure 5E**). Overall, the results that pretreatment with baicalein and Lipo-1 could reduce the production of 4-HNE, the mRNA expression of PTGS2, and the expression of 12/15-LOX and up-regulate the expression of GPX4 in FeCl<sub>3</sub>-induced posttraumatic epileptic seizures further support that baicalein could exert neuroprotective effects in FeCl<sub>3</sub>-induced epileptic seizures by inhibiting 12/15-LOX-related ferroptosis.

## DISCUSSION

Our study showed that pretreatment with baicalein in FeCl<sub>3</sub>induced epileptic seizure mice significantly improved the epileptic seizure behavior and exhibited great neuroprotection in PTE mice. In particular, baicalein exerted neuroprotective effects in PTE mice by inhibiting ferroptosis, and inhibition of 12/15-LOX was related to its antiferroptosis activity. More importantly, these results provide a new therapeutic drug for the treatment of PTE.



**FIGURE 3** Baicalein attenuates erastin-induced neuronal damage by inhibiting ferroptosis. (A) Cell viability was measured with CCK8 assay, in which HT22 cells were incubated with various concentrations (125, 250, 500, and 1,000 nM) of erastin for 8 h. (B) HT22 cells were incubated with different concentrations of baicalein for 2 h, prior to incubation with 500 nM erastin for 8 h. (C) Baicalein (32  $\mu$ M) pretreatment could reduce cell death induced by 500 nM erastin. Scale bar: 200  $\mu$ m (D) after 2 h pretreatment with 32  $\mu$ M baicalein, lipid ROS in HT22 cells was assessed by FACS analysis of C11-BODIPY fluorescence after treatment with erastin for 8 h. (E) Representative Western blots of 4-HNE with 32  $\mu$ M baicalein pretreatment for 2 h in the erastin-induced HT22 cell injury model. (F) The relative mRNA expression of PTGS2 was reduced after baicalein pretreatment. (G) Western blots illustrated that GPX4 was significantly diminished in the erastin-induced HT22 cell injury model and baicalein treatment reversed this phenomenon. Data are representative of three independent experiments with similar results, \*p < 0.05, \*\*p < 0.01, and \*\*\*p < 0.001.

With the wide application of neuroprotective strategies in neurological disease (Szwajgier et al., 2017; Mendes et al., 2019), traditional Chinese medicine has attracted increasing attention due to its advantages of easy penetration of blood-brain barrier and abundance in nature (Zeng, 2017; Chen and Lin, 2018). Baicalein (5,6,7-trihydroxyflavone) is a type of flavonoid compound, which can be acquired from the roots of *Scutellaria baicalensis*, has possessed remarkable neuroprotective effects in neurological diseases, such as AD and PD (Li et al., 2011; Gao et al., 2015; Gu et al., 2016; Zhou et al., 2016; Dinda et al., 2017; Liu et al., 2017). In addition, it has recently been reported that baicalein could exert significant neuroprotective effects in the treatment of temporal lobe epilepsy by reducing oxidative stress and inflammation (Qian et al., 2019). And our previous study suggested that baicalein could reduce the total time and number of epileptic discharges in the inherited epilepsy model *via* suppressing oxidative stress (Mao et al., 2014). Our present extensive work further disclosed the obvious neuroprotection against FeCl<sub>3</sub>-induced posttraumatic epileptic seizures, a kind of important acquired epileptic seizure and this neuroprotective mechanism may be related to suppressing 12/15-LOX-involved ferroptosis. Posttraumatic epileptic seizures were previously reported to be successfully established by FeCl<sub>3</sub> (Pitkanen and McIntosh, 2006). Currently, our present work firstly prepared the mouse model of posttraumatic epileptic seizures using different doses of FeCl<sub>3</sub>, and we found that FeCl<sub>3</sub> by the dose



FIGURE 4 [12/15-LOX is involved in the inhibition of terroptosis in FAC-induced F122 cell injury model by balcalein. (A) immunolitorescence analysis indicated that FAC and erastin significantly increased the expression of 12/15-LOX in HT22 cells, which was reversed by balcalein and PD146176. Scale bar: 50 µm (B) CCK8 analysis showed that pretreatment with PD146176 could reverse cell damage induced by FAC. (C-D) Exposure to FAC and erastin caused HT22 cells to shrink, and 1 µM PD146176 reversed this phenomenon. Scale bar: 200 µm (E-F) pretreatment with 1 µM PD14617, 12.5 µM Fer-1, and 1 µM Lipo-1 could reverse increased lipid ROS in HT22 cells induced by FAC and erastin. Data are representative of three independent experiments with similar results, \*p < 0.05, \*\*p < 0.01, and \*\*\*p < 0.001.

of 50 mM was sufficient for inducing posttraumatic epileptic seizures in mice.

We next focused on the neuroprotective mechanisms of baicalein in posttraumatic epileptic seizures. According to previous reports, the neuroprotective effects of baicalein are closely correlated with its antioxidant properties (Gao et al., 2015; Gu et al., 2016; Zhou et al., 2016). Oxidative stress is intricately involved in the pathogenesis of a variety of neurological diseases, including epilepsy, AD, PD, cerebral ischemia-reperfusion injury, etc. (Butterfield and Lauderback, 2002; Wang et al., 2011; Carvalho et al., 2017; Pauletti et al., 2017; Pearson-Smith and Patel, 2017; Su et al., 2017; Cheignon et al., 2018; Deng et al., 2018; He et al., 2018; Xie et al., 2018b; Xiao et al., 2019). Previous studies indicated that the brain was particularly sensitive to lipid peroxidation-induced damage due to high consumption of oxygen and abundant PUFAs (Salim, 2017). Therefore, lipid peroxidation is the main form of neuronal oxidative damage. Indeed, lipid peroxidation is thought to be involved in the pathogenesis of PTE (Srivastava et al., 2019). Ferroptosis is a lipid peroxidation-dependent new form of oxidative cell death (Dixon et al., 2012; Mao et al., 2018), and can be inhibited by iron chelators and lipid



**FIGURE 5** | Baicalein counteracts  $FeCl_3$ -induced epileptic seizure behavior by inhibiting ferroptosis. (A) Decreased seizure score after treatment with Lipo-1 (a specific inhibitor of ferroptosis) in the  $FeCl_3$ -induced PTE model. (B) Western blots showing expression of 4-HNE in hippocampus tissues of  $FeCl_3$ -induced PTE mice with different doses of baicalein and Lipo-1 pretreatment, n = 6. (C) The relative mRNA expression of PTGS2 was reduced after baicalein and Lipo-1 pretreatment, n = 6. (D) Western blots illustrated that GPX4 was significantly diminished in  $FeCl_3$ -induced PTE mice, and baicalein and Lipo-1 treatment reversed this phenomenon (n = 6 for both groups). (E) Baicalein inhibited the increase of 12/15-LOX in hippocampus tissues from  $FeCl_3$ -induced PTE mice, n = 6. \*p < 0.05, \*\*p < 0.01, and \*\*\*p < 0.001.

peroxidation inhibitors (e.g., Fer-1 and Lipo-1). And we found that ferroptosis was involved in the neuroprotective effects of baicalein in FeCl<sub>3</sub>-induced PTE seizures. To investigate the underlying neuroprotective mechanisms of baicalein in vitro, we established an iron-overloaded HT22 cell damage model by using FAC as a source of iron, but not FeCl<sub>3</sub>. This was because FeCl<sub>3</sub> failed to induce HT22 cell damage (Supplementary Figure S1). FAC is a trivalent organic iron salt used as an iron fortifier for food in the treatment of iron deficiency anemia. It can be absorbed after being reduced to ferrous iron in the body. Moreover, the absorption effect is superior to that of inorganic iron (FeCl<sub>3</sub>) (Lee et al., 2016; Lee et al., 2018). In the FAC-induced HT22 cell damage model, baicalein could reduce the increase of 4-HNE and lipid ROS induced by FAC. Consistent results were obtained in the erastin-induced HT22 cell injury model. This indicated that ferroptosis was involved in FAC-induced HT22 cell damage, and baicalein might act as an inhibitor

of lipid peroxidation to suppress ferroptosis. In addition to antioxidant actions, baicalein was reported to restrain the function of 12/15-LOX (Deschamps et al., 2006; Li et al., 2019). 12/15-LOX is an enzyme that oxidizes PUFAs to produce many bioactive lipid metabolites, and is also an important regulator of ferroptosis (Probst et al., 2017; Kenny et al., 2019). Numerous studies have demonstrated its involvement in the pathogenesis of human diseases (Singh and Rao, 2019). In addition, mice lacking leukocyte-type 12/15-LOX were resistant to PTZinduced epilepsy, which implied that 12/15-LOX might be involved in the process of epileptic seizures (Kanzler et al., 2018). In our study, the expression of 12/15-LOX was increased in the FAC- and erastin-treated HT22 cells and FeCl<sub>3</sub>-induced mice PTE model. Pretreatment with baicalein dramatically inhibited the increase of 12/15-LOX. Intervention of PD146176 (a specific inhibitor of 12/15-LOX) also significantly reduced the production of lipid ROS induced by FAC and erastin. These



findings indicated that baicalein-mediated inhibition of 12/15-LOX contributed to its antiferroptosis activity. Ferroptosis was also reported to take part in glutamate-induced death in a rat organotypic hippocampal slice culture model. The findings revealed that ferroptosis was indeed involved in brain diseases (Dixon et al., 2012). Additionally, ferroptosis was closely associated with neurodegenerative diseases, and could affect the occurrence and development of PD and AD by regulating lipid peroxidation (Xie et al., 2016; Yang and Stockwell, 2016). Moreover, the potent neuroprotective effects of ferroptosis inhibitors in various brain diseases, such as some forms of traumatic and hemorrhagic brain damage, implied that ferroptosis might constitute a potential target for brain disorder therapy (Dixon et al., 2012; Skouta et al., 2014; Do Van et al., 2016; Stockwell et al., 2017; Zille et al., 2017; Anthonymuthu et al., 2018; Xie et al., 2018a; Zhang et al., 2018b). In fact, in the kainic acid-induced rat epilepsy model, it was found that ferroptosis was involved in the pathogenesis of cognitive dysfunction in temporal lobe epilepsy and Fer-1 could improve cognitive dysfunction and hippocampal neuron loss in epileptic rats (Ye et al., 2019). In our present investigation, we found that ferroptosis was involved in the development of posttraumatic epileptic seizures and Lipo-1 could remarkably suppress behavioral seizures in FeCl3-induced seizures in mice. Our studies further demonstrated that baicalein protected neurons against ferroptotic cell death in seizures induced by FeCl<sub>3</sub> and 12/15-LOX was involved in baicalein's neuroprotection (Figure 6).

In summary, our study revealed that ferroptosis was involved in the pathogenesis of PTE. Baicalein, as a naturel bioactive compound, could ameliorate behavioral seizures and play a key neuroprotective role in FeCl<sub>3</sub>-induced posttraumatic epileptic seizures through inhibiting ferroptosis and its neuroprotection might be related to suppression of 12/15-LOX. Our work further extended the neuroprotective action of baicalein in posttraumatic epileptic seizures and provided a theoretical basis for its potential clinical translational applications in PTE.

## DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this manuscript will be made available by the authors, without undue reservation, to any qualified researcher.

## ETHICS STATEMENT

The protocol of animal experiment was approved by the Medical Ethics Committee of Xiangya Hospital and performed in accordance with the National Institute of Health Guide for the Care and Use of Laboratory Animals. All efforts had been made to minimize the animal's suffering.

# **AUTHOR CONTRIBUTIONS**

X-YM conceived and designed the study. QL, Q-QL, J-NJ, and Q-YS performed the experiments. X-YM and H-HZ provided experimental support. QL wrote the paper. X-YM and W-LJ revised the manuscript.

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#### REFERENCES

- Acharya, M. M., Hattiangady, B., and Shetty, A. K. (2008). Progress in neuroprotective strategies for preventing epilepsy. *Prog. Neurobiol.* 84 (2), 363– 404. doi: 10.1016/j.pneurobio.2007.10.010
- Agrawal, A., Timothy, J., Pandit, L., and Manju, M. (2006). Post-traumatic epilepsy: an overview. *Clin. Neurol. Neurosurg.* 108, 433–439. doi: 10.1016/j. clineuro.2005.09.001
- Ahn, J. S., Lee, J. H., Kim, J. H., and Paik, S. R. (2007). Novel method for quantitative determination of amyloid fibrils of alpha-synuclein and amyloid beta/A4 protein by using resveratrol. *Anal. Biochem.* 367, 259–265. doi: 10.1016/j.ab.2007.05.023
- Andrade, S., Ramalho, M. J., Pereira, M. D. C., and Loureiro, J. A. (2018). Resveratrol brain delivery for neurological disorders prevention and treatment. *Front. Pharmacol.* 9, 1261. doi: 10.3389/fphar.2018.01261
- Annegers, J. F., and Coan, S. P. (2000). The risks of epilepsy after traumatic brain injury. Seizure 9, 453–457. doi: 10.1053/seiz.2000.0458
- Anthonymuthu, T. S., Kenny, E. M., Lamade, A. M., Kagan, V. E., and Bayir, H. (2018). Oxidized phospholipid signaling in traumatic brain injury. *Free Radic. Biol. Med.* 124, 493–503. doi: 10.1016/j.freeradbiomed.2018.06.031
- Bazinet, R. P., and Laye, S. (2014). Polyunsaturated fatty acids and their metabolites in brain function and disease. *Nat. Rev. Neurosci.* 15, 771–785. doi: 10.1038/nrn3820
- Belkner, J., Chaitidis, P., Stender, H., Gerth, C., Kuban, R. J., Yoshimoto, T., et al. (2005). Expression of 12/15-lipoxygenase attenuates intracellular lipid deposition during *in vitro* foam cell formation. *Arterioscler. Thromb. Vasc. Biol.* 25, 797–802. doi: 10.1161/01.ATV.0000157580.26858.2d
- Butterfield, D. A., and Lauderback, C. M. (2002). Lipid peroxidation and protein oxidation in Alzheimer's disease brain: potential causes and consequences involving amyloid beta-peptide-associated free radical oxidative stress. *Free Radic. Biol. Med.* 32, 1050–1060. doi: 10.1016/S0891-5849(02)00794-3
- Carvalho, A. N., Firuzi, O., Gama, M. J., Horssen, J. V., and Saso, L. (2017). Oxidative stress and antioxidants in neurological diseases: is there still hope? *Curr. Drug Targets* 18, 705–718. doi: 10.2174/1389450117666160401120514
- Chartrain, A. G., Yaeger, K., Feng, R., Themistocleous, M. S., Dangayach, N. S., Margetis, K., et al. (2017). Antiepileptics for post-traumatic seizure prophylaxis after traumatic brain injury. *Curr. Pharm. Des.* 23 (42), 6428–6441. doi: 10.2174/ 1381612823666171031100139
- Cheignon, C., Tomas, M., Bonnefont-Rousselot, D., Faller, P., Hureau, C., and Collin, F. (2018). Oxidative stress and the amyloid beta peptide in Alzheimer's disease. *Redox Biol.* 14, 450–464. doi: 10.1016/j.redox.2017.10.014
- Chen, I. C., and Lin, T. H. (2018). Formulated Chinese Medicine Shaoyao Gancao Tang reduces Tau aggregation and exerts neuroprotection through antioxidation and anti-inflammation. Oxid. Med. Cell Longev. 2018, 9595741. doi: 10.1155/2018/9595741
- Chen, Z. X., Xu, Q. Q., Shan, C. S., Shi, Y. H., Wang, Y., Chang, R. C., et al. (2019). Borneol for regulating the permeability of the blood-brain barrier in experimental ischemic stroke: preclinical evidence and possible mechanism. *Oxid. Med. Cell Longev.* 2019, 2936737. doi: 10.1155/2019/2936737
- Christensen, J. (2015). The epidemiology of posttraumatic epilepsy. *Semin. Neurol.* 35, 218–222. doi: 10.1055/s-0035-1552923
- Conrad, M., Kagan, V. E., Bayir, H., Pagnussat, G. C., Head, B., Traber, M. G., et al. (2018). Regulation of lipid peroxidation and ferroptosis in diverse species. *Genes Dev.* 32, 602–619. doi: 10.1101/gad.314674.118
- Deng, S., Ai, Y., Gong, H., Feng, Q., Li, X., Chen, C., et al. (2018). Mitochondrial dynamics and protective effects of a mitochondrial division inhibitor, Mdivi-1,

### SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fphar.2019.00638/ full#supplementary-material

**FIGURE S1** | Iron overload induces neuronal death in HT22 cells. HT22 cells were incubated with different concentrations of FAC and FeCl<sub>3</sub> (37.5  $\mu$ M, 75  $\mu$ M, 150  $\mu$ M, 300  $\mu$ M, 600  $\mu$ M) for 36 h. Cell morphology was observed by an inverted microscope.

in lipopolysaccharide-induced brain damage. *Biochem. Biophys. Res. Commun.* 496, 865–871. doi: 10.1016/j.bbrc.2018.01.136

- Deschamps, J. D., Kenyon, V. A., and Holman, T. R. (2006). Baicalein is a potent *in vitro* inhibitor against both reticulocyte 15-human and platelet 12-human lipoxygenases. *Bioorg. Med. Chem.* 14, 4295–4301. doi: 10.1016/j. bmc.2006.01.057
- Dinda, B., Dinda, S., DasSharma, S., Banik, R., Chakraborty, A., and Dinda, M. (2017). Therapeutic potentials of baicalin and its aglycone, baicalein against inflammatory disorders. *Eur. J. Med. Chem.* 131, 68–80. doi: 10.1016/j. ejmech.2017.03.004
- Dixon, S. J., Lemberg, K. M., Lamprecht, M. R., Skouta, R., Zaitsev, E. M., Gleason, C. E., et al. (2012). Ferroptosis: an iron-dependent form of nonapoptotic cell death. *Cell* 149, 1060–1072. doi: 10.1016/j.cell.2012.03.042
- Do Van, B., Gouel, F., Jonneaux, A., Timmerman, K., Gele, P., Petrault, M., et al. (2016). Ferroptosis, a newly characterized form of cell death in Parkinson's disease that is regulated by PKC. *Neurobiol. Dis.* 94, 169–178. doi: 10.1016/j. nbd.2016.05.011
- Englander, J., Bushnik, T., Wright, J. M., Jamison, L., and Duong, T. T. (2009). Mortality in late post-traumatic seizures. J. Neurotrauma 26, 1471–1477. doi: 10.1089/neu.2008.0729
- Gao, L., Li, C., Yang, R. Y., Lian, W. W., Fang, J. S., Pang, X. C., et al. (2015). Ameliorative effects of baicalein in MPTP-induced mouse model of Parkinson's disease: a microarray study. *Pharmacol. Biochem. Behav.* 133, 155–163. doi: 10.1016/j.pbb.2015.04.004
- Gu, X. H., Xu, L. J., Liu, Z. Q., Wei, B., Yang, Y. J., Xu, G. G., et al. (2016). The flavonoid baicalein rescues synaptic plasticity and memory deficits in a mouse model of Alzheimer's disease. *Behav. Brain Res.* 311, 309–321. doi: 10.1016/j. bbr.2016.05.052
- Hamada, H., Hiramatsu, M., Edamatsu, R., and Mori, A. (1993). Free radical scavenging action of baicalein. Arch. Biochem. Biophys. 306, 261–266. doi: 10.1006/abbi.1993.1509
- He, C., Zhang, W., Li, S., Ruan, W., Xu, J., and Xiao, F. (2018). Edaravone improves septic cardiac function by inducing an HIF-1alpha/HO-1 pathway. Oxid. Med. Cell. Longev. 2018, 5216383. doi: 10.1155/2018/5216383
- Islam, M. T. (2017). Oxidative stress and mitochondrial dysfunction-linked neurodegenerative disorders. *Neurol. Res.* 39, 73-82. doi: 10.1080/ 01616412.2016.1251711
- Kanzler, M. A., Van Dyke, A. M., He, Y., Hewett, J. A., and Hewett, S. J. (2018). Mice lacking L-12/15-lipoxygenase show increased mortality during kindling despite demonstrating resistance to epileptogenesis. *Epilepsia Open* 3, 255–263. doi: 10.1002/epi4.12221
- Kenny, E. M., Fidan, E., Yang, Q., Anthonymuthu, T. S., New, L. A., Meyer, E. A., et al. (2019). Ferroptosis contributes to neuronal death and functional outcome after traumatic brain injury. *Crit. Care Med.* 47, 410–418. doi: 10.1097/ CCM.00000000003555
- Kharatishvili, I., and Pitkänen, A. (2010). Posttraumatic epilepsy. Curr. Opin. Neurol. 23 (2), 183–188. doi: 10.1097/WCO.0b013e32833749e4
- Lamar, C. D., Hurley, R. A., Rowland, J. A., and Taber, K. H. (2014). Posttraumatic epilepsy: review of risks, pathophysiology, and potential biomarkers. *J. Neuropsychiatry Clin. Neurosci.* 26, iv–113. doi: 10.1176/appi.neuropsych.260201
- Lee, D. G., Kam, M. K., Kim, K. M., Kim, H. S., Kwon, O. S., Lee, H. S., et al. (2018). Peroxiredoxin 5 prevents iron overload-induced neuronal death by inhibiting mitochondrial fragmentation and endoplasmic reticulum stress in mouse hippocampal HT-22 cells. *Int. J. Biochem. Cell Biol.* 102, 10–19. doi: 10.1016/j. biocel.2018.06.005

- Lee, D. G., Park, J., Lee, H. S., Lee, S. R., and Lee, D. S. (2016). Iron overload-induced calcium signals modulate mitochondrial fragmentation in HT-22 hippocampal neuron cells. *Toxicology* 365, 17–24. doi: 10.1016/j.tox.2016.07.022
- Li, C., Lin, G., and Zuo, Z. (2011). Pharmacological effects and pharmacokinetics properties of Radix Scutellariae and its bioactive flavones. *Biopharm. Drug Dispos.* 32, 427–445. doi: 10.1002/bdd.771
- Li, W., Suwanwela, N. C., and Patumraj, S. (2016). Curcumin by down-regulating NF-kB and elevating Nrf2, reduces brain edema and neurological dysfunction after cerebral I/R. *Microvasc. Res.* 106, 117–127. doi: 10.1016/j.mvr.2015.12.008
- Li, X. J., and Zhang, H. Y. (2008). Western-medicine-validated anti-tumor agents and traditional Chinese medicine. *Trends Mol. Med.* 14, 1–2. doi: 10.1016/j. molmed.2007.11.002
- Li, Y., Chen, Q., Ran, D., Wang, H., Du, W., Luo, Y., et al. (2019). Changes in the levels of 12/15-lipoxygenase, apoptosis-related proteins and inflammatory factors in the cortex of diabetic rats and the neuroprotection of baicalein. *Free Radic. Biol. Med.* 134, 239–247. doi: 10.1016/j.freeradbiomed.2019.01.019
- Liu, A., Huang, L., Fan, H., Fang, H., Yang, Y., Liu, S., et al. (2015). Baicalein pretreatment protects against liver ischemia/reperfusion injury via inhibition of NF-kappaB pathway in mice. *Int. Immunopharmacol.* 24, 72–79. doi: 10.1016/j.intimp.2014.11.014
- Liu, R. X., Song, G. H., Wu, P. G., Zhang, X. W., Hu, H. J., Liu, J., et al. (2017). Distribution patterns of the contents of five biologically activate ingredients in the root of *Scutellaria baicalensis. Chin. J. Nat. Med.* 15, 152–160. doi: 10.1016/ S1875-5364(17)30030-4
- Liu, Y. F., Gao, F., Li, X. W., Jia, R. H., Meng, X. D., Zhao, R., et al. (2012). The anticonvulsant and neuroprotective effects of baicalin on pilocarpineinduced epileptic model in rats. *Neurochem. Res.* 37, 1670–1680. doi: 10.1007/ s11064-012-0771-8
- Lu, S., and Wang, X. (2015). The role and potential mechanism of resveratrol in the prevention and control of epilepsy. *Future Med. Chem.* 7, 2005–2018. doi: 10.4155/fmc.15.130
- Mao, C., Wang, X., Liu, Y., Wang, M., Yan, B., Jiang, Y., et al. (2018). A G3BP1interacting lncRNA promotes ferroptosis and apoptosis in cancer *via* nuclear sequestration of p53. *Cancer Res.* 78, 3484–3496. doi: 10.1158/0008-5472. CAN-17-3454
- Mao, X., Cao, Y., Li, X., Yin, J., Wang, Z., Zhang, Y., et al. (2014). Baicalein ameliorates cognitive deficits in epilepsy-like tremor rat. *Neurol. Sci.* 35, 1261– 1268. doi: 10.1007/s10072-014-1695-7
- Mao, X., Jin, M., Chen J., Zhou H., and Jin W. (2018). Live or let die: neuroprotective and anti-cancer effects of nutraceutical antioxidants. *Pharmacol. Ther.* 183, 137–151. doi: 10.1016/j.pharmthera.2017.10.012
- McKinnon, C., Gros, P., Lee, D. J., Hamani, C., Lozano, A. M., Kalia, L. V., et al. (2018). Deep brain stimulation: potential for neuroprotection. *Ann. Clin. Transl. Neurol.* 6 (1), 174–185. doi: 10.1002/acn3.682
- Mendes, M. O., Rosa, A. I., Carvalho, A. N., Nunes, M. J., Dionisio, P., Rodrigues, E., et al. (2019). Neurotoxic effects of MPTP on mouse cerebral cortex: modulation of neuroinflammation as a neuroprotective strategy. *Mol. Cell. Neurosci.* doi: 10.1016/j.mcn.2019.01.003. [Epub ahead of print].
- Pan, R. Y., and Ma, J. (2019). Sodium rutin ameliorates Alzheimer's diseaselike pathology by enhancing microglial amyloid-beta clearance. *Sci. Adv.* 5, eaau6328. doi: 10.1126/sciadv.aau6328
- Pan, W., Kwak, S., Liu, Y., Sun, Y., Fang, Z., Qin, B., et al. (2011). Traditional chinese medicine improves activities of daily living in Parkinson's disease. *Parkinsons Dis.* 2011, 789506. doi: 10.4061/2011/789506
- Pauletti, A., Terrone, G., Shekh-Ahmad, T., Salamone, A., Ravizza, T., Rizzi, M., et al. (2017). Targeting oxidative stress improves disease outcomes in a rat model of acquired epilepsy. *Brain* 140, 1885–1899. doi: 10.1093/brain/ awx117
- Pearson-Smith, J. N., and Patel, M. (2017). Metabolic dysfunction and oxidative stress in epilepsy. Int. J. Mol. Sci. 18, E2365. doi: 10.3390/ijms18112365
- Pi, X., Zhou, L., Cui, L., Liu, A., Zhang, J., Ma, Y., et al. (2014). Prevalence and clinical characteristics of active epilepsy in southern Han Chinese. *Seizure* 23, 636–640. doi: 10.1016/j.seizure.2014.05.002
- Pitkanen, A., and McIntosh, T. K. (2006). Animal models of post-traumatic epilepsy. J. Neurotrauma 23, 241–261. doi: 10.1089/neu.2006.23.241
- Pitkanen, A., Kemppainen, S., Ndode-Ekane, X. E., Huusko, N., Huttunen, J. K., Grohn, O., et al. (2014). Posttraumatic epilepsy-disease or comorbidity? *Epilepsy Behav.* 38, 19–24. doi: 10.1016/j.yebeh.2014.01.013

- Poprac, P., Jomova, K., Simunkova, M., Kollar, V., Rhodes, C. J., and Valko, M. (2017). Targeting free radicals in oxidative stress-related human diseases. *Trends Pharmacol. Sci.* 38, 592–607. doi: 10.1016/j.tips.2017.04.005
- Probst, L., Dachert, J., Schenk, B., and Fulda, S. (2017). Lipoxygenase inhibitors protect acute lymphoblastic leukemia cells from ferroptotic cell death. *Biochem. Pharmacol.* 140, 41–52. doi: 10.1016/j.bcp.2017.06.112
- Qian, X., Wang, Z. R., Zheng, J. J., Ding, J. Q., Zhong, J. G., Zhang, T. Y., et al. (2019). Baicalein improves cognitive deficits and hippocampus impairments in temporal lobe epilepsy rats. *Brain Res.* 1714, 111–118. doi: 10.1016/j. brainres.2019.02.028
- Racine, R. J. (1972). Modification of seizure activity by electrical stimulation. II. Motor seizure. *Electroencephalogr. Clin. Neurophysiol.* 32, 281–294. doi: 10.1016/0013-4694(72)90177-0
- Salim, S. (2017). Oxidative stress and the central nervous system. J. Pharmacol. Exp. Ther. 360, 201–205. doi: 10.1124/jpet.116.237503
- Sato, M., Racine, R. J., and McIntyre, D. C. (1990). Kindling: basic mechanisms and clinical validity. *Electroencephalogr. Clin. Neurophysiol.* 76, 459–472. doi: 10.1016/0013-4694(90)90099-6
- Singh, A., and Trevick, S. (2016). The epidemiology of global epilepsy. Neurol. Clin. 34, 837–847. doi: 10.1016/j.ncl.2016.06.015
- Singh, N. K., and Rao, G. N. (2019). Emerging role of 12/15-lipoxygenase (ALOX15) in human pathologies. *Prog. Lipid Res.* 73, 28–45. doi: 10.1016/j. plipres.2018.11.001
- Skouta, R., Dixon, S. J., Wang, J., Dunn, D. E., Orman, M., Shimada, K., et al. (2014). Ferrostatins inhibit oxidative lipid damage and cell death in diverse disease models. J. Am. Chem. Soc. 136, 4551–4556. doi: 10.1021/ ja411006a
- Srivastava, N. K., Mukherjee, S., Sharma, R., Das, J., Sharma, R., Kumar, V., et al. (2019). Altered lipid metabolism in post-traumatic epileptic rat model: one proposed pathway. *Mol. Biol. Rep.* 46, 1757–1773. doi: 10.1007/ s11033-019-04626-9
- Stockwell, B. R., Friedmann Angeli, J. P., Bayir, H., Bush, A. I., Conrad, M., Dixon, S. J., et al. (2017). Ferroptosis: a regulated cell death nexus linking metabolism, redox biology, and disease. *Cell* 171, 273–285. doi: 10.1016/j. cell.2017.09.021
- Su, J., Liu, J., Yan, X. Y., Zhang, Y., Zhang, J. J., Zhang, L. C., et al. (2017). Cytoprotective effect of the UCP2–SIRT3 signaling pathway by decreasing mitochondrial oxidative stress on cerebral ischemia–reperfusion injury. *Int. J. Mol. Sci.* 18, E1599. doi: 10.3390/ijms18071599
- Szaflarski, J. P., Nazzal, Y., and Dreer, L. E. (2014). Post-traumatic epilepsy: current and emerging treatment options. *Neuropsychiatr. Dis. Treat.* 10, 1469–1477. doi: 10.2147/NDT.S50421
- Szwajgier, D., Borowiec, K., and Pustelniak, K. (2017). The neuroprotective effects of phenolic acids: molecular mechanism of action. *Nutrients* 9, E477. doi: 10.3390/nu9050477
- van Leyen, K., Kim, H. Y., Lee, S. R., Jin, G., Arai, K., and Lo, E. H. (2006). Baicalein and 12/15-lipoxygenase in the ischemic brain. *Stroke* 37, 3014–3018. doi: 10.1161/01.STR.0000249004.25444.a5
- Volavka, J. (2011). Violent crime, epilepsy, and traumatic brain injury. *PLoS Med* 8, e1001148. doi: 10.1371/journal.pmed.1001148
- Wang, G., Yang, Q., Li, G., Wang, L., Hu, W., Tang, Q., et al. (2011). Time course of heme oxygenase-1 and oxidative stress after experimental intracerebral hemorrhage. *Acta Neurochir. (Wien)* 153, 319–325. doi: 10.1007/ s00701-010-0750-2
- Wenzel, S. E., Tyurina, Y. Y., Zhao, J., St Croix, C. M., Dar, H. H., Mao, G., et al. (2017). PEBP1 wardens ferroptosis by enabling lipoxygenase generation of lipid death signals. *Cell* 171, 628–641.e626. doi: 10.1016/j.cell.2017.09.044
- Wu, T. Y., Chen, C. P., and Jinn, T. R. (2011). Traditional Chinese medicines and Alzheimer's disease. *Taiwan J. Obstet. Gynecol.* 50, 131–135. doi: 10.1016/j. tjog.2011.04.004
- Xiao, X., Yuan, Q., Chen, Y., Huang, Z., Fang, X., Zhang, H., et al. (2019). LncRNA ENST00000453774.1 contributes to oxidative stress defense dependent on autophagy mediation to reduce extracellular matrix and alleviate renal fibrosis. J. Cell. Physiol. 234, 9130–9143. doi: 10.1002/jcp.27590
- Xie, B. S., Wang, Y. Q., Lin, Y., Mao, Q., Feng, J. F., Gao, G. Y., et al. (2018a). Inhibition of ferroptosis attenuates tissue damage and improves long-term outcomes after traumatic brain injury in mice. CNS Neurosci. Ther. 25, 465–475. doi: 10.1111/cns.13069

- Xie, Y., Hou, W., Song, X., Yu, Y., Huang, J., Sun, X., et al. (2016). Ferroptosis: process and function. Cell Death Differ. 23, 369–379. doi: 10.1038/cdd.2015.158
- Xie, Y., Wanggou, S., Liu, Q., Li, X., Liu, J., and Wu, M. (2018b). A brainspecific isoform of apoptosis-inducing factor 2 attenuates ischemia-induced oxidative stress in HT22 cells. *Neurochem. Int.* 112, 179–186. doi: 10.1016/j. neuint.2017.07.006
- Xu, T., Yu, X., Ou, S., Liu, X., Yuan, J., Huang, H., et al. (2017). Risk factors for posttraumatic epilepsy: a systematic review and meta-analysis. *Epilepsy Behav.* 67, 1–6. doi: 10.1016/j.yebeh.2016.10.026
- Yang, W. S., and Stockwell, B. R. (2016). Ferroptosis: death by lipid peroxidation. Trends Cell Biol. 26, 165–176. doi: 10.1016/j.tcb.2015.10.014
- Ye, Q., Zeng, C., Dong, L., Wu, Y., Huang, Q., and Wu, Y. (2019). Inhibition of ferroptosis processes ameliorates cognitive impairment in kainic acid-induced temporal lobe epilepsy in rats. *Am. J. Transl. Res.* 11, 875–884.
- Zeng, B. Y. (2017). Effect and mechanism of Chinese herbal medicine on Parkinson's disease. *Int. Rev. Neurobiol.* 135, 57–76. doi: 10.1016/bs.irn.2017.02.004
- Zhang, M. R., Jiang, K., Yang, J. L., and Shi, Y. P. (2019). Flavonoids as key bioactive components of *Oxytropis falcata* Bunge, a traditional antiinflammatory and analgesic Tibetan medicine. *Nat. Prod. Res.*, 1–18. doi: 10.1080/14786419.2019.1574786. [Epub ahead of print].
- Zhang, X., Wang, X., Hu, X., Chu, X., Li, X., and Han, F. (2018a). Neuroprotective effects of a *Rhodiola crenulata* extract on amyloid-beta peptides (Abeta1-42) -induced cognitive deficits in rat models of Alzheimer's disease. *Phytomedicine* 57, 331–338. doi: 10.1016/j.phymed.2018.12.042

- Zhang, Y. H., Wang, D. W., Xu, S. F., Zhang, S., Fan, Y. G., Yang, Y. Y., et al. (2018b). Alpha-lipoic acid improves abnormal behavior by mitigation of oxidative stress, inflammation, ferroptosis, and tauopathy in P301S Tau transgenic mice. *Redox Biol.* 14, 535–548. doi: 10.1016/j.redox.2017.11.001
- Zhou, L., Tan, S., Shan, Y. L., Wang, Y. G., Cai, W., Huang, X. H., et al. (2016). Baicalein improves behavioral dysfunction induced by Alzheimer's disease in rats. *Neuropsychiatr. Dis. Treat.* 12, 3145–3152. doi: 10.2147/NDT. S117469
- Zille, M., Karuppagounder, S. S., Chen, Y., Gough, P. J., Bertin, J., Finger, J., et al. (2017). Neuronal death after hemorrhagic stroke *in vitro* and *in vivo* shares features of ferroptosis and necroptosis. *Stroke* 48, 1033–1043. doi: 10.1161/ STROKEAHA.116.015609

**Conflicts of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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