



Reversing the Tumor Target: Establishment of a Tumor Trap

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Despite the tremendous progress made in the field of cancer therapy in recent years, certain solid tumors still cannot be successfully treated. Alongside classical treatments in the form of chemotherapy and/or radiotherapy, targeted treatments such as immunotherapy that cause fewer side effects emerge as new options in the clinics. However, these alternative treatments may not be useful for treating all types of cancers, especially for killing infiltrative and circulating tumor cells (CTCs). Recent advances pursue the trapping of these cancer cells within a confined area to facilitate their removal for therapeutic and diagnostic purposes. A good understanding of the mechanisms behind tumor cell migration may drive the design of traps that mimic natural tumor niches and guide the movement of the cancer cells. To bring this trapping idea into reality, strong efforts are being made to create structured materials that imitate myelinated fibers, blood vessels, or pre-metastatic niches and incorporate chemical cues such as chemoattractants or adhesive proteins. In this review, the different strategies used (or could be used) to trap tumor cells are described, and relevant examples of their performance are analyzed.

Keywords: tumor cell migration, tumor trap, biomimetic trap, cancer therapy, premetastatic niche recruitment

INTRODUCTION

For many decades, surgery, radiotherapy, and chemotherapy have served as the mainstay trident in the fight against cancer (**Figure 1** Scheme I). During this period, the prognosis of many types of cancer has been significantly improved (Brustugun et al., 2018; Zeng et al., 2018; Iacobucci, 2019; Trama et al., 2019). However, the widespread use of these treatments has also uncovered several major limitations. For example, the feasibility of surgery is very much dependent on the localization and the size of the tumor. The procedure is also contraindicated in patients with poor clinical performance. As for radiotherapy and chemotherapy, these treatments are often implicated with serious side effects that, in some cases, may outweigh their potential therapeutic benefits. Moreover, these treatments lack the capacity to prevent metastases, which are responsible for roughly 90% of cancer-associated deaths (Rankin and Giaccia, 2016).

Numerous studies in the quest of improving cancer treatments are driven by the concept of “magic bullet” (**Figure 1** Scheme II-2) put forward by the German scientist Paul Ehrlich (Strebhardt and Ullrich, 2008). If radio- and chemotherapy are considered as weapons of mass destruction, Ehrlich’s strategy can be regarded as the sniper of cancer therapy. This concept is mainly based on the idea of increasing the bioavailability and specificity of vector-associated active agents in the body while limiting their premature degradation and toxicity. In the context of anticancer

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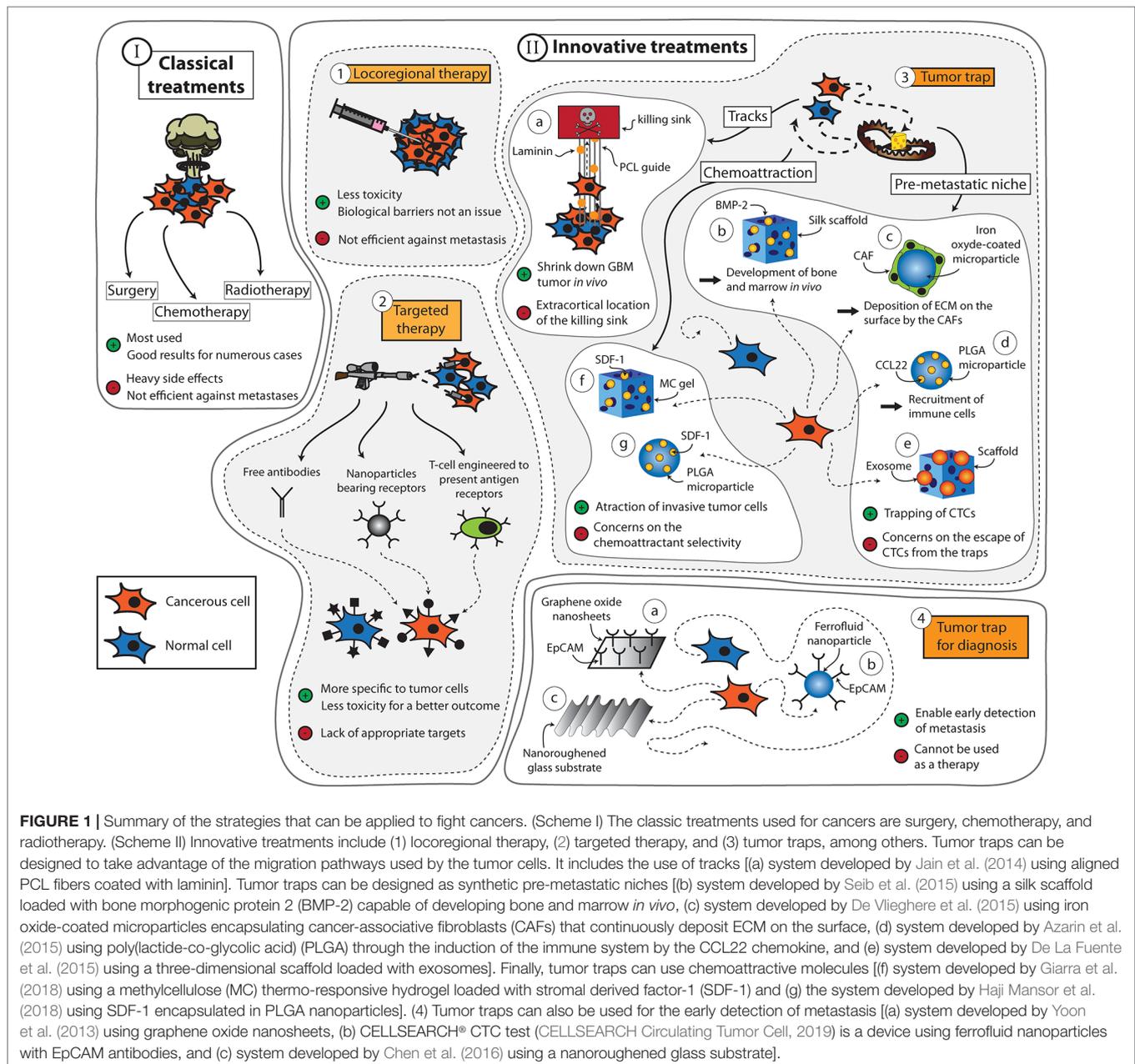
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approaches, the success of selective therapies depends on the discovery of targeting elements that, when coupled with active ingredients and/or diagnostic cues, enable the recognition of well-characterized molecules, cells, or tissues. For example, Adcetris® targets the antigen CD30 in the treatment of Hodgkin's lymphoma, and Kadcyla® targets HER2, which is present in about 20% of breast cancer patients (Kim and Kim, 2015). Nevertheless, the discovery of appropriate targets that are specific to tumor cells remains a challenging task, despite the significant advancements made in the field of genomics and proteomics in recent decades.

Fortunately, a plethora of new therapies are being approved regularly for the treatment of cancer. Among them is the use of locoregional therapies (Figure 1 Scheme II-1) that includes

Nanotherm® (MagForce) that involves injection of magnetic nanoparticles inside the tumor or into the resection cavity. A magnetic field is then applied to generate heat *via* the nanoparticles and kill the cancerous cells locally (Maier-Hauff et al., 2011). It is currently licensed in Europe for the treatment of brain tumors and has received FDA approval in February 2018 to be used in clinical trials involving prostate cancer patients (MagForce, 2018). Another example is Optune® (Novocure Ltd), a tumor-treating field (TTF) device composed of electrodes that can be placed on the patients' scalp and connected to a generator to deliver a low-intensity electric field of 200 kHz (Taphoorn et al., 2018). It is believed to exert anticancer effects by disrupting the division of tumor cells (Giladi et al., 2015). The device has

been approved for the treatment of glioblastoma and shown to increase the median survival from 15 to 21 months when used on top of the standard treatments for this cancer (Stupp et al., 2017). However, many countries and insurance companies do not cover the cost of this treatment, and the clinical adoption of this technology remains limited due to concerns regarding the lack of understanding of the device's exact mechanism of action. Moreover, some skepticism exists toward the legitimacy of the device approval process due to the poor consideration of any placebo effect during the clinical trial phase (Fabian et al., 2019).

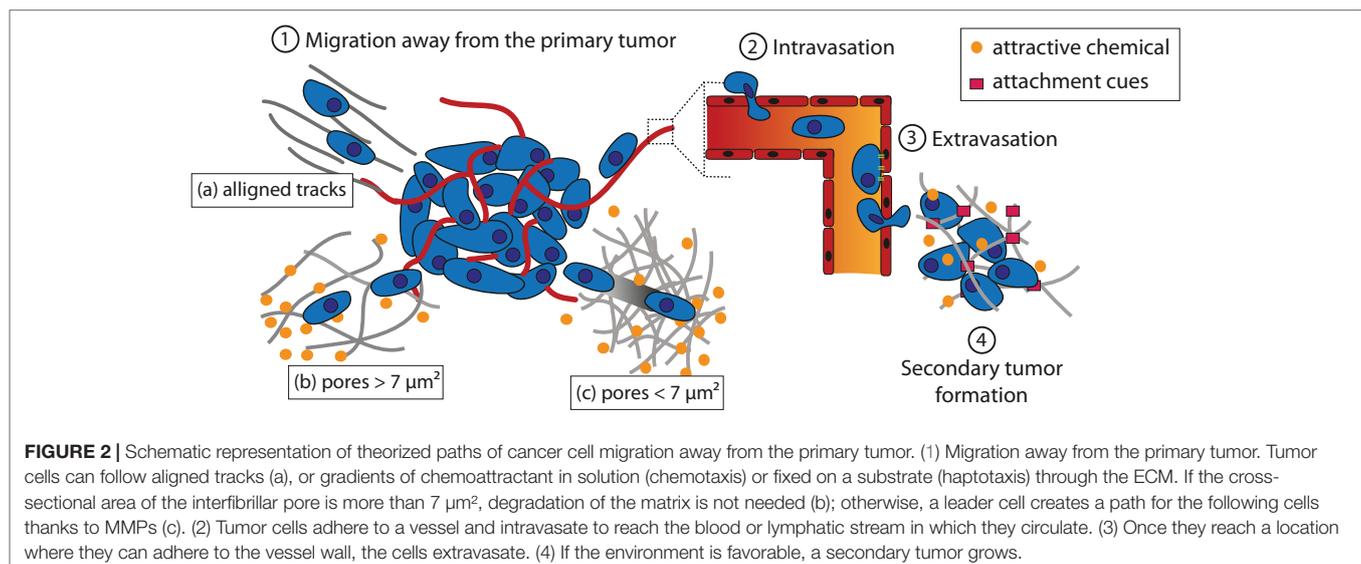
Among the numerous classes of novel anticancer treatments entering the market, cancer immunotherapy is arguably the one that is currently attracting the highest level of attention (Figure 1 Scheme II-2). This class of treatment aims to treat cancer through artificial stimulation of the patient's immune system (Zhang and Chen, 2018). The most cutting-edge subset of this type of treatment is the chimeric antigen receptor (CAR) T-cell immunotherapy, which involves harvesting T cells from a patient and genetically modifying these cells to express a receptor that can bind to a tumor antigen before injecting them back into the patient (Feins et al., 2019). CAR-T cell immunotherapy made its debut in the clinic in August 2017 when Kymriah® (Novartis) was approved by the FDA for the treatment of B-cell acute lymphoblastic leukemia (BCALL) (FDA, 2017a). This was followed by the approval of Yescarta® (Gilead Sciences) in October of the same year for the treatment of diffuse large B-cell lymphoma (FDA, 2017b). Both Kymriah® and Yescarta® exert their effects by targeting CD19 antigen (Zheng et al., 2018). However, there are numerous ongoing clinical studies that explore the feasibility of targeting other antigens including PD-L1 (ClinicalTrials.gov Identifier NCT03672305, NCT03198052, NCT03330834), EpCAM (ClinicalTrials.gov Identifier NCT03013712, NCT03563326, NCT02729493), and CD123 (ClinicalTrials.gov Identifier NCT03796390, NCT02937103, NCT03672851). Many of these trials also attempt to evaluate the efficacy of CAR-T cell immunotherapy against solid tumors to expand its indication beyond certain blood cancers. More comprehensive

reviews on the current status and future directions of CAR T-cell immunotherapy as well as other subsets of cancer immunotherapy such as immune checkpoint inhibitors and cancer vaccines can be found elsewhere (Dougan et al., 2018; Li et al., 2018; Sambhi et al., 2019).

Despite the continuous increase in the number of novel anticancer treatments entering the clinic, local recurrence in previously healthy tissues seen in many cases of solid tumors remains an unsolved conundrum among clinicians and researchers alike. Development of new therapies for *in situ* control of the disease, while avoiding the problems of biological barriers and systemic toxicity, still proves to be a formidable task. Thus, in parallel to the innovative approaches mentioned previously, the idea of trapping infiltrative or circulating tumor cells (CTCs) within a confined area to facilitate their removal for therapeutic or diagnostic purposes has risen (Figure 1 Scheme II-3,4). Over the last years, this concept has developed progressively. The aim is twofold: a) to avoid the uncontrolled dissemination of tumor cells and b) to efficiently prevent the phenomenon of epithelial-mesenchymal transition (EMT) or development of metastases. The concept is largely inspired by the "ecological trap" theory (van der Sanden et al., 2013). By considering cancers as ecosystems, it is possible to develop tumor traps not only for the infiltrative tumor cells, but also for the CTCs that are responsible for metastasis. However, imitating the traditional features of a natural habitat or niche for tumor cells and directing their migration pathways present numerous physical and biological challenges. The focus of this review will be on understanding the mechanisms of tumor cell migration and how this knowledge can be used to capture them, keeping in mind that different tumors are likely to utilize different mechanisms.

MIGRATION OF TUMOR CELLS

Tumor cells must cover a great distance on their journey to form metastases (Figure 2). The first step of the process is to



migrate away from the primary tumor. Tumor cells can follow aligned tracks a), or gradients of chemoattractant in solution (chemotaxis) or fixed on a substrate (haptotaxis) through the extracellular matrix (ECM) b). If the cross-sectional area of interfibrillar pores is more than $7 \mu\text{m}^2$, degradation of the matrix is not needed for cell movement. Alternatively, a leader cell can open a path for the following cells by virtue of the matrix metalloproteinase (MMP) activity c). The second step is to intravasate into the bloodstream or the lymphatic system in which the tumor cells will transit through the circulation. Third, cells extravasate to secondary tissues once they reach a location where they can adhere to the walls of the vessel. The fourth and final step deals with the formation of a secondary tumor. This only occurs if the environment is favorable to tumor growth.

The different strategies implemented to mislead these cells into a trap are described in next sections. These strategies exploit the current knowledge on cancer cell migration and metastasis and the specificities of each type of tumor.

Migration Away From the Primary Tumor

The physical interactions between the ECM and cancer cells play a key role in allowing the cells to start migrating. Cancer cells may undergo an EMT to acquire a motile phenotype (Polyak and Weinberg, 2009). This translates into the loss of intracellular adhesion molecules such as E-cadherin and cytokeratins, resulting in detachment of the cells from the primary tumor, and an overexpression of MMP on their surface that allows the cells to digest laminin and collagen IV to progress in the dense ECM (Polyak and Weinberg, 2009). These changes are thought to be related to the stiffness of the matrix around the tumor, which is of higher values than that of normal tissues (Wozniak et al., 2003; Paszek et al., 2005; Kumar and Weaver, 2009; Levental et al., 2009). For example, the stiffness of GBM tissues is of ~ 25 kPa, while normal brain tissues have a stiffness of 0.1 to 1 kPa (Wang et al., 2014). Wang et al. investigated the effect of matrix stiffness on GBM cells and found that an increase in matrix rigidity could induce an upregulation of MMP-1, Hras, RhoA, and ROCK1 (Wang et al., 2014), which are involved in increasing cell motility (Pulukuri and Rao, 2008; Liu et al., 2014; Wu et al., 2016; Li et al., 2017). Another physical factor that governs the dissemination of cancer cells is the architecture of the extracellular environment, which includes pores of a diameter ranging from less than 1 to $20 \mu\text{m}$ (Wolf et al., 2009). Matrix degradation is usually required for cancer cell migration to occur when the cross-sectional area of the interfibrillar pore is less than $7 \mu\text{m}^2$, which corresponds to about 10% of the nuclear cross-section of cancer cells (Wolf et al., 2013). Above this value, cells can undergo deformation to migrate through the ECM.

Apart from the porosity of the ECM, the spatial arrangement of the matrix fibers near the primary tumor sites can also influence the motility of tumor cells; aligned fibers offer tracks that are more conducive to migration (Provenzano et al., 2008; Paul et al., 2017). These tracks are found along the ECM fibers in the interstitial space, between the muscle and nerve fibers, and along or within the vasculature of organs, among others (Gritsenko et al., 2012). Moreover, it has been observed that leader tumor cells are able to

align collagen fibers to assist the migration of the following cells (Provenzano et al., 2006). In addition to creating the required physical space, these tracks also facilitate cancer cell migration by providing relevant molecular guidance. For example, cancer cells can be guided toward laminin and hyaluronan molecules in the ECM by their integrins and CD44 receptors, respectively, and also *via* haptotaxis by chemokines and growth factors immobilized along the tracks (Aznavorian et al., 1990; Gritsenko et al., 2012). Jain et al. took inspirations from these biological phenomena and designed a scaffold to guide GBM cells toward a killing sink in an extracortical location (**Figure 1** Scheme II-3-a) (Jain et al., 2014). They utilized aligned poly-L-lysine and laminin-coated polycaprolactone (PCL) nanofibers ($10 \mu\text{m}$ thick) encased in a PCL/polyurethane support (2.4 mm diameter) to imitate the white matter tracts (Bernstein and Woodard, 1995; Giese et al., 2003). The killing sink was composed of a collagen-based hydrogel conjugated to the chemotherapeutic agent cyclophosphamide. With this approach, the tumor mass of induced GBM in mice could be reduced. However, despite the positive results, this strategy as itself has limited clinical appeals, as the establishment of an extracortical sink in human patients may invite numerous technical difficulties. Instead, exploiting the local (intracortical) migration of the cancer cells may be a more translatable strategy to develop an efficient tumor trap for this cancer.

Intravasation and Tumor Cell Circulation

Tumor cells can circulate through the blood and lymphatic vessels on their journey to form a secondary tumor distant from the primary site (Chiang et al., 2016). This requires the cells to intravasate by passing through the endothelial cell junctions. Intravasation into the blood vessels occurs frequently due to the leaky nature of tumor vasculature. In addition, it has been observed *in vivo* that metastatic cells are able to polarize toward blood vessels. A possible explanation to this phenomenon is that these cells have an increased expression of epidermal growth factor (EGF) and/or colony-stimulating factor 1 (CSF-1) receptors. Thus, they migrate toward a gradient of EGF or CSF-1 released by the macrophages lining the blood vessels (Wyckoff et al., 2000; Wyckoff et al., 2004). However, it is still easier for tumor cells to enter the lymphatic system, as the surrounding ECM network is easier to penetrate and that the endothelial junctions are looser (Wong and Hynes, 2006). Either route can lead to blood vessel dissemination since the lymphatic circulation drains into the blood. As the lymphatic fluid is filtered by the lymph nodes, tumor cells are invariably invading them, starting with the nearest (Nathanson, 2003).

Once in the blood circulation, the trajectory of the tumor cells is influenced by the blood flow, the diameter of the blood vessels, and the intercellular adhesion (Wirtz et al., 2011). Two mechanisms can lead to the arrest of a CTC: physical occlusion and cell adhesion (**Figure 3**). Physical occlusion occurs when the diameter of the blood vessel is smaller or equal to the one of the CTC (usually around $10 \mu\text{m}$). This has been observed in the brain by real-time imaging in a mouse model (Kienast et al., 2010). Adhesion of CTCs to the vessel walls occurs when there is a balance between the adhesion force and the shear force

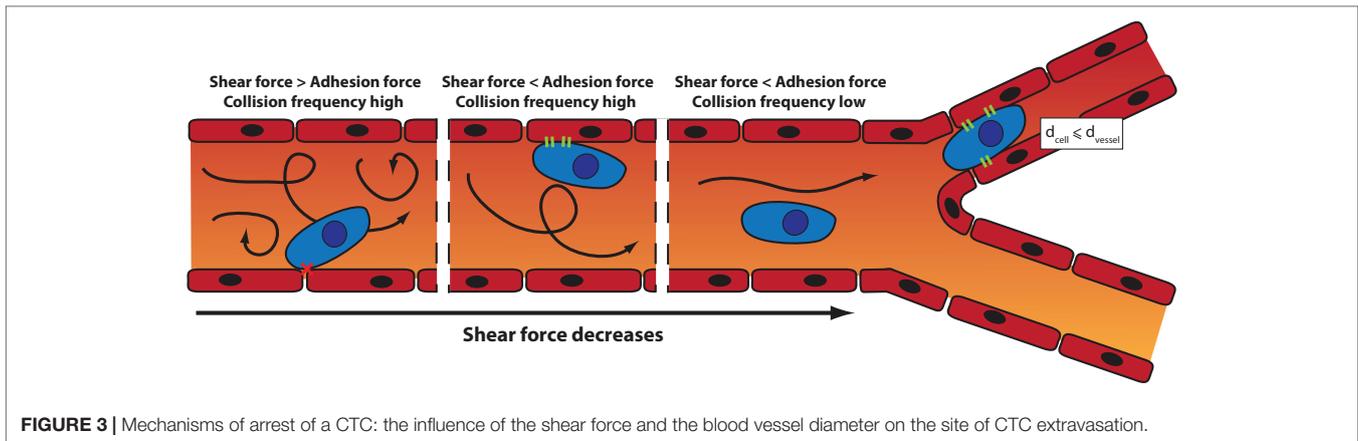


FIGURE 3 | Mechanisms of arrest of a CTC: the influence of the shear force and the blood vessel diameter on the site of CTC extravasation.

exerted inside the blood vessel (Zhu et al., 2008). When the shear force increases, the collisions between cells and the vessel wall increase, which in turn enhances the likelihood of cell adhesion. However, if the shear force is too high, turbulences may prevent the adhesion.

It is therefore possible to capture CTCs by designing a device with strong adhesive cues. Yoon et al. (2013) designed a microfluidic device consisting of graphene oxide nanosheets fixed onto a patterned gold surface to capture CTCs in early-stage cancer for analytical purposes (Figure 1 Scheme II-4-a). The nanosheets were functionalized with epithelial cell adhesion molecule (EpCAM) antibody to capture CTCs. Blood samples were retrieved from healthy donors and mixed with labeled human breast cancer cell lines MCF-7 and Hs-578T and the human prostate cancer cell line PC-3. This microfluidic device captured more than 70% of the cancer cells in the prepared blood sample with high specificity. A similar principle was also implemented in the design of CELLSEARCH® CTC Test, the first and only clinically validated and FDA-approved blood test for enumerating CTCs (CELLSEARCH Circulating Tumor Cell, 2019). It allows for early assessment of patient prognosis as well as follow-up of the patient. The test constitutes the use of ferrofluid nanoparticles with EpCAM antibodies that bind to CTCs (Figure 1 Scheme II-4-b). Once magnetically separated from the rest of the blood sample, cells are stained to discriminate CTCs from leukocytes that can copresent in the sample. Working within the same domain of research, Chen et al. (2016) designed a nanoroughened glass substrate to capture CTCs based on their stronger adhesion capacity compared with normal blood cells (Figure 1 Scheme II-4-c). Such a working principle makes this device useful for capturing CTCs regardless of their surface marker expression profile, which is known to vary according to the type of cancer, patient demographics, and the state of the disease. It is indeed well discussed in the literature that the EMT process may lead to reduction in the epithelial cell adhesion molecule (EpCAM) expression in CTC (Hyun et al., 2016). The capture of CTC using EpCAM antibody alone may lead to an underestimation of the CTC number in the blood. With this device, more than 80% of cancer cells in whole blood samples from mice with induced breast cancer or lung cancer were

captured independently of their EpCAM expression. Based on these findings, it is evident that a number of approaches can be adopted to capture CTCs to enable early detection of metastasis, although most of them are still far from translation to the clinic.

Extravasation and Secondary Tumor Formation

At the end of their time in blood circulation, the CTCs that survived and adhered to the blood vessel walls extravasate, and a fraction of these proceeds to form a secondary tumor. It has been shown experimentally that only about 0.01% of the cells in the circulation system eventually contribute to metastatic colonization (Fidler, 1970). The location at which they stop and grow into a secondary tumor is not believed to be randomly determined, and the reasons driving the selection of a particular site are still being investigated. In 1889, Paget et al. hypothesized that metastasizing cancer cells are like seeds that can only grow in the proper soil (Paget, 1889). Indeed, it has been observed that invasive cancer cells tend to migrate toward certain preferred sites of metastasis, a phenomenon that has been coined as “tissue tropism” (Seib et al., 2015). More recent studies revealed that the formation of certain microenvironments termed as pre-metastatic niches is crucial to the subsequent formation of metastatic tumors. These microenvironments consist of inflammatory immune cells, stromal cells, ECM proteins, tumor-secreted exosomes, and homing factors (Aguado et al., 2017). Tumor-secreted exosomes are sent to prime the niche at a target organ (often lungs, liver, brain, bone, and lymph nodes) by attracting bone marrow-derived cells (BMDCs) as well as increasing the proliferation of fibroblast-like stromal cells (Peinado et al., 2012). BMDCs include CD11b⁺ myeloid cells, myeloid-derived suppressor cells, neutrophils, tumor-associated macrophages, and regulatory T cells. They are known to create an attractive site for metastasizing cells, and the presence of VEGFR1-positive BMDCs can serve as a predictor for the arrival of tumor cells (Kaplan et al., 2005). Moreover, the establishment of a pre-metastatic niche is associated with an increased secretion of inflammatory cytokines and chemokines (Hiratsuka et al., 2006; Darash-Yahana et al., 2009; Brennecke et al., 2014). The increasing understanding of

the pre-metastatic niches and their roles in welcoming metastatic dissemination has inspired scientists to create synthetic niches as a means to trap migrating cancer cells.

Creation of a Synthetic Pre-metastatic Niche to Trap CTCs

Many different strategies have been explored to engineer pre-metastatic niches. For example, Seib et al. (2015) developed a tumor trap for the metastasizing cells of breast and prostate cancers by imitating the red bone marrow microenvironment (**Figure 1** Scheme II-3-b). The strategy was adopted based on the knowledge that the bone was the preferred site of colonization in more than 60% of cases of metastasis for primary breast cancer and 73% for primary prostate cancer (Weiss, 1992). Evidence shows that red bone marrow attracts migrating cancer cells *via* chemotaxis with stromal derived factor-1 (SDF-1) (Taichman et al., 2002) and CXCL16 (Lee et al., 2013). It also provides adhesion sites that interact with tumor cell surface molecules such as annexin2 (Shiozawa et al., 2008), growth arrest-specific 6 (GASP-6) (Shiozawa et al., 2010), CD44 (Hill et al., 2006), integrins (such as VLA-4, VLA-5, and LFA-1), and cadherins. Moreover, the bone marrow microenvironment is composed of osteoblasts, osteoclasts, stromal cells, stem cells, and mineralized bone marrow surrounded by a rich vascular bed, making it a perfect site for tumor growth (Mishra et al., 2011). To imitate the red bone marrow, Seib et al. designed a silk fibroin scaffold loaded with bone morphogenetic protein 2 (BMP-2) that is capable of developing bone and marrow *in vivo*. After implantation into the mammary fat pads of mice with induced breast or prostate tumor, no effect on the primary tumor growth was observed. However, metastatic growth could be seen taking place in the functionalized scaffolds, suggesting that it is possible to lure metastasizing cells into a trap by imitating the bone marrow microenvironment. A similar strategy was adopted by Bersani et al. (2014). They utilized a polyacrylamide hydrogel coated with bone marrow stromal cells (BMSCs), which was able to capture metastasizing cells of prostate cancer.

De Vlieghere et al. (2015) took a slightly different approach to mimic a pre-metastatic niche by developing traps made of iron oxide-coated microparticles, encapsulating metabolically active cancer-associated fibroblasts (CAFs) (**Figure 1** Scheme II-3-c). The CAFs continuously deposited ECM composed of type I collagen and tenascin C, among others, creating an adhesive environment for disseminated cancer cells. The microparticles were implanted into the peritoneal cavities of mice with induced ovarian cancer. Twenty-four hours after the implantation, the microparticles were magnetically removed, and the adhesion of cancer cells on the microparticles was assessed. The treatment led to a delay in peritoneal metastasis and prolonged the animal survival.

Another variation in the strategy for recruiting metastasizing cancer cells was presented by Azarin et al. (2015). They developed a microporous scaffold from poly(lactide-co-glycolic acid) (PLGA) scaffold for *in vivo* capture of metastasizing breast cancer cells through the induction of a local immune response (**Figure 1** Scheme II-3-d). Indeed, it has been shown that immune cells are implicated in tumor cell recruitment (Qian et al., 2009; Qian

et al., 2011). Here, they have either recruited immune cells into the scaffold by grafting the chemokine CCL22, which is known to induce migration of immune cells but not tumor cells, or incorporated the Gr1^{hi}CD11b⁺ immune cells directly into the PLGA scaffold. By doing this, they were able to reduce the number of breast cancer cells that metastasized to the lung by 88%. Similarly, Rao et al. (2016) designed a poly(ϵ -caprolactone) (PCL)-based device with a slower degradation rate than PLGA scaffolds and investigated the immune response induced at the implantation site, the ability of the device to recruit metastatic cells for detection prior to colonization of organs as well as its influence on the survival of mice with induced breast cancer. Pelaez et al. (2018) further developed the strategy to enable the elimination of the attracted metastatic cells by noninvasive focal hyperthermia. To do so, they coupled metal discs to PCL microparticles to allow heat generation through electromagnetic induction using an oscillating magnetic field. The heat generation could be modulated conveniently by changing the size of the disc or the type of metal.

It has been shown that exosomes, which are vesicles involved in the transfer of information between cells, play a role in homing CTCs in the pre-metastatic niche (Peinado et al., 2012). De La Fuente et al. (2015) harnessed the potential of this knowledge and designed a three-dimensional scaffold with embedded exosomes extracted from the ascitic fluid of ovarian cancer patients (**Figure 1** Scheme II-3-e). The scaffold, called M-Trap, was implanted in the inner wall of the peritoneum of mice with a xenograft of human ovarian cancer in the peritoneal cavity. They showed that the scaffold could serve as the preferred site of metastasis, while a peritoneal carcinomatosis was observed in the absence of the M-Trap. Moreover, an increase in the mean survival was observed in the presence of the M-Trap (from 117.5 to 198.8 days), which was further improved by the removal of the scaffold (mean of survival of 309.4 days). The safety and performance of the M-Trap is currently being evaluated in a clinical trial involving female patients with stage IIIC ovarian cancer (ClinicalTrials.gov Identifier NCT03085238).

Chemoattraction of Tumor Cells

Migrating cells can make directional choices when presented with different migration pathways. *In vitro*, it has been shown that neutrophil-like cells can navigate through a microfabricated maze by following a chemical gradient (Skoge et al., 2016). Chemokines and their receptors are particularly involved in this navigation process. They are indeed responsible for the chemoattraction of various cells and could therefore be used to attract migrating tumor cells into a trap. Several receptors/chemokines have been identified to facilitate cancer cell migration. The most investigated one is SDF-1, also called CXCL12, which binds with high affinity to the CXCR4 and CXCR7 receptors. This chemokine is a pro-inflammatory mediator and is known to play a role in the recruitment of T cells, monocytes, and lympho-hemopoietic progenitor cells (Crump et al., 1997). Its overexpression has been linked to an increase in the invasiveness of ovarian cancer (Kajiyama et al., 2008), breast cancer (Bachelder et al., 2002; Zhan et al., 2016), and GBM (Barbero et al., 2003; Hira et al., 2017), among others [further details can be found elsewhere (Kryczek

et al., 2007)]. In addition, despite being less well studied, CXCL16 and its receptor CXCR6 are also suspected to play a role in the migration of tumor cells. Wang et al. (2008) have shown that the expression of CXCR6 increases with the grade of prostate cancer. These results were supported by Lu et al. (2008) who observed that metastatic cells from prostate cancer overexpress the CXCR6 receptor. Moreover, CXCL16 have been shown to induce the migration and enhance the proliferation of CXCR6-expressing cancer cells *in vitro* (Darash-Yahana et al., 2009).

With this knowledge, several groups have tried to stop the migration of tumor cells by inhibiting chemokine receptors, particularly the CXCR4 receptor. A reduction in the migration of cancer cells has been observed *in vitro* (Brennecke et al., 2014; Chittasupho et al., 2017; Zheng et al., 2017). However, this has not been successfully replicated *in vivo*. Brennecke et al. found that the use of CXCR4 antibody 12G5 can reduce the number of osteosarcoma pulmonary metastases having a diameter of <0.1 mm but not those of larger dimensions (Brennecke et al., 2014). This finding can be explained by the fact that chemoattraction of cancer cells can be mediated by several pairs of chemokine-receptor interaction (for example, SDF-1 can bind either CXCR4 or CXCR7 or both, and CXCR6 can be activated by CXCL16). In addition, cells can activate the so-called compensation mechanisms *in vivo* to maintain their migration capacity. Indeed, it has been observed that neural progenitor cells (NPCs) are able to migrate in response to SDF-1 *via* the activation of the CXCR7 receptor in response to the blockade of the CXCR4 receptor (Chen et al., 2015). Therefore, in order to stop the migration of tumor cells completely, all receptors implicated in *in vivo* chemoattraction should be identified and blocked, making the task nearly impossible. Moreover, this strategy could only work if the tumor cells have yet to begin migrating. In the particular case of GBM, cancer cells usually have already invaded the surrounding tissues at the time of diagnosis (Giese et al., 2003). Thus, it may be more useful to direct the migration of cells toward a desired location instead of blocking the migration process altogether.

Chemokines are already being used to attract cells into a scaffold for regenerative medicine purposes. The tumor trap concept can benefit from the existing knowledge in this field of application. Water-retaining polymer networks such as hydrogels and swellable matrices, which have been widely

used in tissue engineering and regeneration, are pivotal platforms that are transferrable to the tumor trap application. For this purpose, biocompatible polymers capable of *in situ* formation of three-dimensional gels (Schesny et al., 2014; Shen et al., 2014; Addington et al., 2015) or matrices (Li et al., 2013; Yoon et al., 2013; Azarin et al., 2015; De Vlieghere et al., 2015; Ding et al., 2015; Autier et al., 2018) may be used to exert chemotaxis (based on a gradient of soluble attractant or repellent) or haptotaxis (based on a gradient of substrate-bound extracellular matrix proteins) (Figure 4). Of particular interest is the potential exploitation of the CXCR4-SDF-1 axis due to its prominent roles in regulating the migration of many types of cancer cells (Andreas et al., 2014; Schesny et al., 2014; Addington et al., 2015; Goffart et al., 2015). Examples of biomaterials used to deliver SDF-1 for regenerative medicine are presented in Table 1. Recently, the development of SDF-1-releasing scaffolds to attract tumor cells has received increasing attention. Giarra et al. (2018) designed a temperature-responsive gel loaded with SDF-1 based on methylcellulose (MC) or poloxamers with or without hyaluronic acid (HA) for the purpose of attracting CXCR4-expressing GBM cells (Figure 1 Scheme II-3-f). Haji Mansor et al. (2018), on the other hand, encapsulated the chemokine in nanoparticles composed of PLGA and a (PEG)-PLGA copolymer to achieve sustained release (Figure 1 Scheme II-3-g). However, in both papers, no *in vivo* assessment on the ability of SDF-1 to attract migrating cancer cells was performed.

Challenges Associated With the Clinical Translation of the Tumor Trapping Strategy

While promising preclinical results have been obtained from the use of tumor traps as a diagnostic and/or therapeutic tool, there are multiple issues that must be addressed before this approach can enter the clinic. Main concerns include identifying suitable means for *in vivo* monitoring of the recruitment of cancer cells into the scaffolds to allow one to decide on the optimal time point for killing the trapped cancer cells. Prolonged duration of cancer cell recruitment may lead to overcrowding of the tumor trap and subsequent cell escape, reducing the purpose of the synthetic niche to merely a “relay” for the cancer cells en route to their natural metastatic sites.

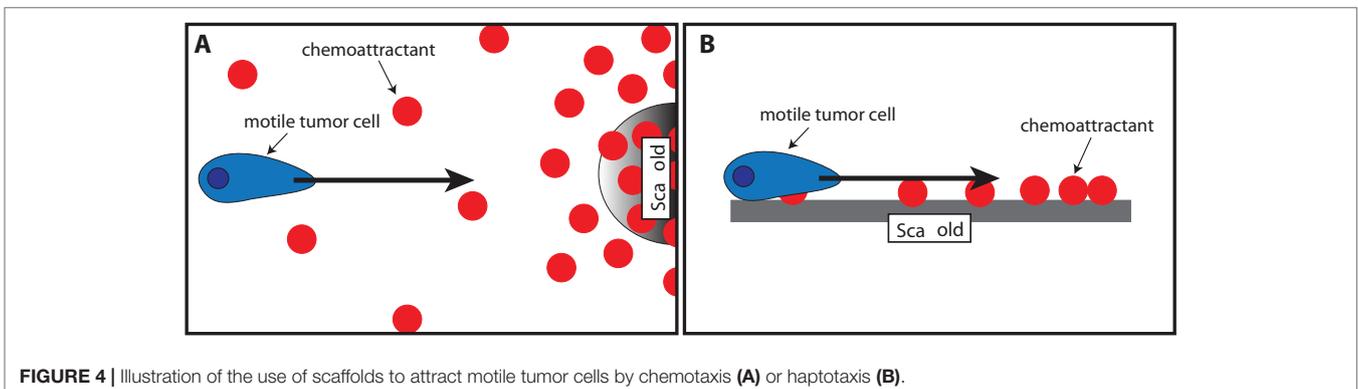


TABLE 1 | Strategies to load SDF-1 into different biomaterials.

Bonding strategy	Type of biomaterial	Composition	Target site for regeneration	Ref	
Adsorption	Hydrogel	Hyaluronic acid	Cardiac tissue	(Purcell et al., 2012)	
		PPCN	Wound healing	(Zhu et al., 2016)	
	3D scaffold	Collagen		Cartilage	(Chen et al., 2015)
				Tendon	(Sun et al., 2018)
		Collagen/silk fibroin	Bone	(Hu et al., 2018)	
		Hydroxyapatite	Bone	(Zhang et al., 2018)	
		Decellularized skeletal muscle	Muscle	(Rajabi et al., 2018)	
		Collagen/PLA	Bone	(Ritz et al., 2017)	
		PLGA	Cartilage	(Wang et al., 2017)	
		PCL/gelatin	Bone	(Ji et al., 2013)	
Immobilization through specific heparin-mediated interaction	Membrane	Heparin/PEG	Cardiac tissue	(Prokoph et al., 2012)	
	Hydrogel	Heparin/PEG	Blood vessel	(Krieger et al., 2016)	
			Cardiac tissue	(Baumann et al., 2013)	
	3D scaffold	Heparin/PLCL	Blood vessel	(Shafiq et al., 2017)	
		Heparin/PGS	Blood vessel	(Lee et al., 2013)	
		Heparin/PLLA	Blood vessel	(Yu et al., 2012)	
	Systems with nano/microparticles	Microspheres	Alginate	Bone	(Xu et al., 2018)
		Hydrogel/	Hydrogel: CS/GP	Bone	(Mi et al., 2017)
		Nanoparticles	nano: CS/CMCS		
		Microcapsules	Dex-GMA/gelatin/PNIPAAm	Wound healing	(Chen et al., 2013)
Particles		PLGA	Cardiac tissue	(Zamani et al., 2015)	

PPCN, poly (polyethylene glycol citrate-co-N-isopropylacrylamide); PLA, polylactide; PLGA, poly(lactic-co-glycolic acid); PCL, polycaprolactone; PEG, poly(ethylene-glycol); PLCL, poly(L-lactide-co-ε-caprolactone) (PLCL); PGS, poly(glycerol sebacate); PLLA, poly(L-lactic acid); CS, chitosan; GP, beta-glycerol phosphate disodium salt; CMCS, carboxymethyl-chitosan; Dex-GMA, glycidyl methacrylated dextran; PNIPAAm, poly(N-isopropylacrylamide).

The incorporation of chemoattractant molecules such as SDF-1 into the tumor trap may also introduce additional complexities. In particular, there are concerns regarding the selectivity of SDF-1-mediated chemotaxis. Indeed, in addition to its role in recruiting cancer cells to local and distant sites of colonization, SDF-1 is also implicated in the homing of other cell lines such as immune cells and stem cells (Crump et al., 1997; Wang et al., 2017; Hu et al., 2018). Moreover, the potential off-target effects may also be exacerbated by the fact that this chemokine is known to be involved in various processes that support tumor progression, angiogenesis, metastasis, and survival (Teicher and Fricker, 2010). It is therefore necessary to study in more detail the effect of injecting such proteins near tumor cells *in vivo* and to carefully evaluate the entire risks before moving to the clinic.

Further down the development timeline, the most effective way to kill the recruited cancer cells should be elucidated. It may be tempting to suggest direct removal of the trap to achieve an immediate eradication of the disease. However, this approach will necessitate an additional surgery, a requirement that may be very difficult to fulfill especially in patients who are terminally ill. A less invasive solution would be to use stereotactic radiotherapy (SRT). SRT is a treatment where radiation beam is directed to a well-defined spot, usually the tumor site, from many different angles around the body. The procedure ensures the targeted site receives much higher dose of radiation than the surrounding tissues. At the moment, SRT seems to be a viable option for killing the trapped cancer cells. This said, other selective approaches should also be considered and evaluated.

CONCLUSION

A good understanding of the escape pathways of a prey allows the hunter to capture it more efficiently; the same rule of thumb can be applied to tumor cells. Using this principle, it is possible to design tumor traps for diagnostic and/or therapeutic applications. For the latter purpose, it is necessary that the trapped cells are killed by the application of existing therapies. The different therapeutic strategies (surgery, chemotherapy, targeted therapy, ...) may not be sufficient on their own to cure every cancer type, but they can be used in combination to achieve the best clinical outcomes. Jain et al. used a chemotherapeutic agent in the form of cyclophosphamide alongside their tumor trap to shrink down the size of GBM tumors (Jain et al., 2014). It would also be interesting to combine the trap with radiosensitizers, focus x-ray, or γ -ray microbeams. Since the trap would concentrate the tumor cells, the efficiency of chemo- and radiotherapies can potentially be improved, while the associated side effects are likely to decrease. Immunotherapy, which can be broadly described as the activation of immune cells to make them able to recognize and eliminate tumor cells, could also be used. Indeed, one of the major difficulties in immunotherapy is to make the cancer cells accessible to the activated immune cells. This is particularly true in the brain, as there is a need to overcome the blood-brain barrier (Lyon et al., 2017). If immune cells can be preloaded or attracted into the trap *via* chemoattraction, this will facilitate the killing of the trapped cancer cells. Indeed, immune cells are also sensitive to a gradient of chemokines such as SDF-1 (Krieger et al., 2016)

and can therefore be recruited into the trap together with the cancer cells of interest. Overall, this bio-integrative approach can be seen as counterintuitive insofar as the factors governing the trapping of tumor cells are also involved in other signaling pathways that may lead to effects that are opposite to the initial will (Komarova, 2015). Our current knowledge on the mechanisms driving the migration of cancerous cells might not be sufficient to develop a trap that only impacts tumor cells in a safe manner. The translation to the clinic will therefore require further investigations on the efficacy and safety of such systems. Nevertheless, as Albert Einstein pointed out, “we do not solve problems with the modes of thought that have engendered them,” and this unique approach therefore deserves further investigations.

AUTHOR CONTRIBUTIONS

MN wrote the manuscript. CA-L and EG contributed to the conception and design of the work. MH, FB, CA-L and EG contributed to manuscript revision. All authors read and approved the submitted version.

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Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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