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Primary cilia: The central role in the electromagnetic field induced bone healing

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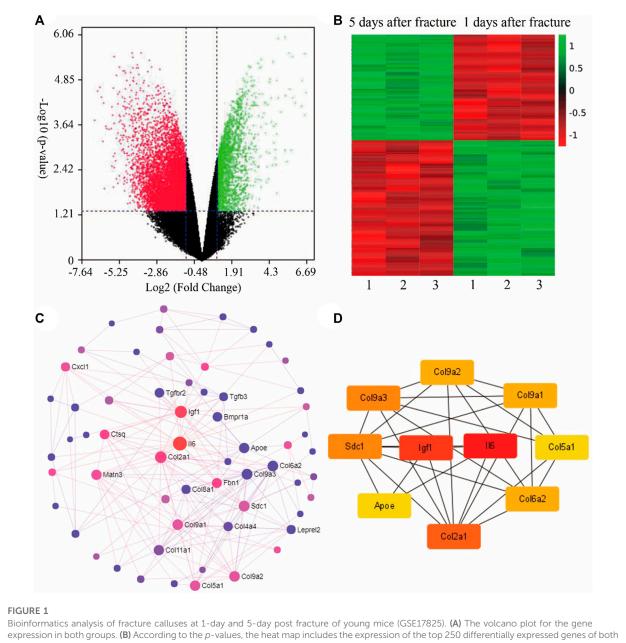
Primary cilia have emerged as the cellular "antenna" that can receive and transduce extracellular chemical/physical signals, thus playing an important role in regulating cellular activities. Although the electromagnetic field (EMF) is an effective treatment for bone fractures since 1978, however, the detailed mechanisms leading to such positive effects are still unclear. Primary cilia may play a central role in receiving EMF signals, translating physical signals into biochemical information, and initiating various signaling signaling pathways to transduce signals into the nucleus. In this review, we elucidated the process of bone healing, the structure, and function of primary cilia, as well as the application and mechanism of EMF in treating fracture healing. To comprehensively understand the process of bone healing, we used bioinformatics to analyze the molecular change and associated the results with other studies. Moreover, this review summarized summarized some limitations in EMFs-related research and provides an outlook for ongoing studies. In conclusion, this review illustrated the primary cilia and related molecular mechanisms in the EMF-induced bone healing process, and it may shed light on future research.

KEYWORDS

electromagnetic fields, primary cilia, osteogenesis, signaling pathway, bone

1 Introduction

The skeletal system is the framework that not only provides the architectural stability of the connective tissues but also protects the organs. Therefore, bone fractures caused by injury, osteoporosis, cancer, or other systemic diseases are one of the most serious diseases (De Mattos et al., 2012). Considering there are 7.9 million fractures in the United States annually, with the increase in disabilities and the prolonged recovery time, bone fractures exacerbate heavy socioeconomic and healthcare burdens (Victoria et al., 2009). Although lots of research and great progress have been made in this field, 5%–10% of fractures worldwide still developed into delayed-union or nonunion (Claes et al., 2012). Thus, further research for improving the clinical outcomes is urgently necessary.



Bioinformatics analysis of fracture calluses at 1-day and 5-day post fracture of young mice (CSE1/825). (A) The volcano plot for the gene expression in both groups. (B) According to the *p*-values, the heat map includes the expression of the top 250 differentially expressed genes of both groups. (C) The protein-protein interaction (PPI) of the top 100 genes generated by String and Network Analyst. (D) The top 10 hub genes analyzed by Cytoscape, *i.e.*, Igf1, II6, Col9a2, Col9a3, Sdc1, Apoe, Col2a1, Col6a2, and Col5a1. Col9a1 (figures were plotted by https://www.bioinformatics. com.cn (last accessed on 30 July 2022), an online platform for data analysis and visualization).

For thoroughly exploring the treatment of fracture healing, the physiological and pathological activities of bone should be clarified first. In general, the skeletal system is a dynamic and highly coordinated system, which involves the balance of both osteoblastic bone formation and osteoclastic bone resorption (Kaku and Komatsu, 2017). After the fracture, the healing process follows a regular timeline: 1) Intramembranous ossification: within the first 5 days, mesenchymal stem cells (MSCs) and osteoprogenitors migrate to the fracture site, then they proliferate and differentiate at the subperiosteal callus region. 2) Endochondral ossification: during days 5–9, transforming growth factor- β (TGF- β), osteocalcin (OCN), and various cytokines are released to promote chondrocytes proliferation. 3) During days 9–14, the chondrocytes become mature, the proliferation decreases, and even undergo apoptosis gradually. But the osteogenesis sustains in this process, the soft callus is mineralized with matrix deposition, the woven bone is generated, the vascular invasion is initiated gradually. 4) Between

days 14–21, the new bone is regenerated actively. 5) After day 21, cellular proliferation ceases, the bone remodeling process initiates (Daish et al., 2018).

To observe the change in gene expression during fracture healing, gene expression datasets (GSE17825) of fracture was analyzed from the Gene Expression Omnibus (GEO) database (Khan et al., 2008). Gene expression analysis was performed on the groups of 1 day and 5 days post fracture of young mice. The volcano plot indicated that there is a total of 13,661 genes with significant difference between 1 and 5 days post fracture (Figure 1A). The expression of top 250 genes is shown in the heat map, which confirmed that gene expression is clustered by timepoint, and the differences can be observed (Figure 1B). To discern the relationship between the top 250 genes, the STRING database was used to analyse the protein-protein interactions (PPI) (Figure 1C). Then the top 10 hub genes were determined by the Cytoscape software (Figure 1D), which demonstrated that the processes of inflammation, extracellular matrix, and cartilage development are involved.

However, during the healing process, there are various biochemical and biophysical stimuli, such as fluid shear, microgravity, as well as the electromagnetic field, which can affect cellular activities and the clinical outcomes. Therefore, it is promising and worthwhile to explore, optimize, and modulate these physiochemical signals to positively promote bone fractures. To reach this goal, several prerequisites should be fulfilled by applying physiochemical signals to affect cellular activities: 1) there should exist organelles which can perceive these extracellular signals, 2) the specific translator that can translate these signals into the biochemical messages, 3) an effective signalingsignaling pathway which can transmit this information to the nucleus (Temiyasathit and Jacobs, 2010). Therefore, we focused on the cellular "antenna"-primary cilium. In this review, we intend to delineate the key role of and the molecular mechanism of the primary cilium in the process of bone healing.

2 The structure and function of the primary cilium

The primary cilium is a microtubule-based non-motile organelle that protrudes from the membrane in nearly all eukaryotic cells. Compared with motile cilium, primary cilium lacks the two central microtubules (9 + 0 structure) and other axonemal components, such as radial spokes and dynein arms (Schwartz et al., 1997). Primary cilium can detect, transmit, and interpret physiochemical signals from the local environment into cells to regulate proliferation, migration, and differentiation (Prevo et al., 2017). This function owe to the ciliary membrane that contains lots of receptors that are associated with various signalingsignaling pathways (Seeger-Nukpezah and Golemis, 2012).

Primary cilium cannot synthesize proteins for themselves, so the existence of a transport system between primary cilia and the cell body is necessary for delivering necessary proteins for genesis and maintenance of the cilium. In 1993, Rosenbaum et al. verified this transport system by finding the bidirectional movement of particles in the space between the membrane of flagellar and the doublet microtubules (Kozminski et al., 1993). This orderly movement is the intraflagellar transport (IFT) system, which is a complex and highly regulated cellular activity initiated immediately after the early stage of ciliogenesis and operated by various IFT proteins and motors (Taschner et al., 2012). The IFT proteins are required for the structure and function of the primary cilium. IFT proteins can be classified into two types, and functions as retrograde or anterograde transporting proteins between cell body and primary cilium (Table 1). Nevertheless, the components of the IFT-B complex, both IFT25 and IFT27, can also deliver the proteins of hedgehog (Hh) signaling through the retrograde IFT (Keady et al., 2012; Eguether et al., 2014). IFT 140 was highly related to the osteogenic ability given that the ageing and ovariectomy (OVX) induced osteoporosis models expressed significantly lower IFT 140 (Zhang C. Y. et al., 2019). Further study showed that the targeted deletion of IFT 80 (Yuan et al., 2016) or IFT 88 (Tummala et al., 2010) resulted in impaired osteogenic ability. Recently, primary cilium was proven to be an important organelle to promote bone healing by reacting and transducing extracellular physiochemical signals, such as electromagnetic fields (EMFs).

3 The development of electromagnetic fields in bone healing

In 1978, Bassett *et al.* successfully applied pulsed electromagnetic fields (PEMFs) in treating bone fractures (Bassett et al., 1978). After that, the United States Food and Drug Administration (FDA) approved PEMF as a kind of safe and effective physical treatment for osteoporosis, delayed union, and nonunion fractures. Nowadays, it has been widely recognized that PEMFs can bring tremendous clinical benefits because of its noninvasive property, safety, low-cost, home implementation, and none-adverse effects (Yuan et al., 2018).

To better understand the potential mechanism, it is necessary to elucidate the character of EMFs. EMF exists everywhere in our daily life and can be categorized into different types depending on the origin (from permanent magnets/electric currents), production (natural/man-made), status (static/dynamic), and ionization ability (ionizing/non-ionizing radiation). In the field of regenerative medicine, EMFs with specific frequency receive substantial attention. Based on the frequency, EMFs can be classified into: 1) extremely low-frequency EMFs (ELFintermediate frequency EMFs, < 300 Hz), 2)EMFs (300 Hz-10 MHz), and 3) radiofrequency EMFs TABLE 1 The classification and function of the intraflagellar transport (IFT) system.

Transporting direction	IFT subtypes	Components of IFT proteins	Motor protein
retrograde transport: from the cilium to the cell body	IFT-A	IFT 43, IFT 121, IFT 122, IFT 139, IFT 140, and IFT 144	dynein-2
anterograde transport: from the cell body to the cilium	IFT-B Core subcomplex	IFT 22, IFT 25, IFT 27, IFT 46, IFT 52, IFT 56, IFT 70, IFT 74, IFT 81, and IFT 88/Polaris	kinesin-2
anterograde transport: from the cell body to the cilium	IFT-B Peripheral subcomplex	IFT 20, IFT 38, IFT 54, IFT 57, IFT 80, and IFT 172	kinesin-2

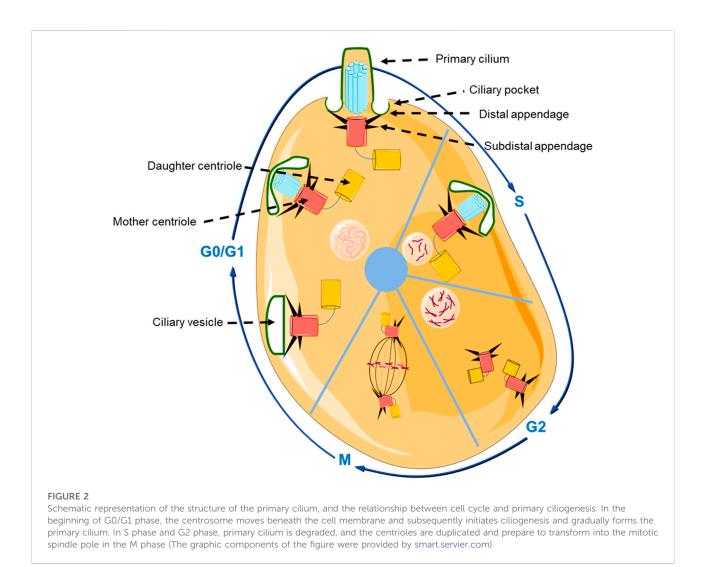
(10 MHz-300 GHz) (Wang and Zhang, 2017). Interestingly, electric fields (EFs) and magnetic fields (MFs) are different because Efs exert electronic forces on charged molecules, the cell membrane, and the cellular organelles directly (Ross et al., 2015). Thus, Efs can mediate the cell migration by directly regulating the dynamics of microfilament and microtubules (Finkelstein et al., 2004). MFs can penetrate through the cell, influent the biochemical reaction, and regulate protein synthesis (Funk et al., 2009). Therefore, MFs may play a more important role in stimulating bone cells that were surrounded by tissues and bone. From numerous types of EMFs, the ELF-PEMFs combine various advantages, which can generate a series of magnetic pulses, and each magnetic pulses can also generate an electrical signal in turn (Goodman and Shirley-Henderson, 1990). Most importantly, ELF-EMFs were proven to be the non-ionizing, low-energy, and significant influences on the cellular and subcellular activities (Bassett, 1993).

The generation of EMFs whichs in accord with Faraday's Lawwhichch declared that EFs or MFs are generated when electrons move at a constant velocity, while EMFs are generated when electrons move at an accelerated velocity, and the change of EFs or MFs can generate each other mutually (Ross et al., 2015). Therefore, it is theoretically feasible to regulate the endogenous EMFs at the wound site with the exogenous EMFs device (Ross et al., 2015). Yasuda et al. reported that physiological EMFs exist in the human body because the piezoelectric phenomenon (The classic, 1977). The further study verified that the physiological frequency of EMFs during the movement of the musculoskeletal system ranges from 5 to 30 Hz (Antonsson and Mann, 1985). Under pathological conditions, Becker et al. (1962) reported that during the healing process, the differences in electric potentials around the wound would gradually decline to zero. This phenomenon could be explained by the endogenous EFs formation between the lower electric potential in the wounded site, and the higher electric potential in the healthy tissue; thus, the transepithelial potential difference (TEP) is generated in the local wound (Song et al., 2002). Therefore, EMFs provide lots of benefits for wound healing and are a potential treatment for fracture healing.

4 The function of primary cilia in transducing electromagnetic field signals to promote bone healing

Primary cilia of osteoblast cells were first discovered by Tonna and Lampen, (1972). After that, the fundamental function of primary cilia in bone metabolism attracted extensive attention. Osteocytes, a subset of osteoblasts located in the "lacunae", play an important role in transferring the extracellular mechanical signals into the bone to regulate the balance of osteogenesis and osteoclastogenesis. Primary cilium of osteocyte works as: 1) most osteocytes stay in G0/G1 stage, so they exhibit matured primary cilia (Kaku and Komatsu, 2017); 2) The primary cilium of osteocyte touches the lacunar tightly because the length of primary cilia (2-9 µm) is much longer than the lacunae space $(0.1-3 \,\mu\text{m})$ (Jacobs et al., 2010); 3) After receiving extracellular signals by primary cilia, osteocytes can communicate with surrounding cells through the gap junction (Jacobs et al., 2010). Therefore the length of the primary cilium is most important for its function for two reasons: 1) if the length of primary cilia is too short to touch the wall of the "lacunae", the osteocytes cannot perceive mechanical forces, and 2) longer primary cilia have a larger surface area, more receptors, ion channels, thus are more sensitive to extracellular signals (Yuan and Yang, 2016).

Primary cilia are indispensable in transducing extracellular signals. For example, under the treatment of fluid flow, osteocyte cell lines MLO-Y4 showed excellent osteogenic ability (Malone et al., 2007). However, after damaging primary cilia by using the chloral hydrate or IFT88 siRNA, the expression of OPG/RANKL decreased (Malone et al., 2007), and the osteogenesis was impaired (Tummala et al., 2010). The polycystin-1 (PC1), a component of primary cilium that plays a key role in mechanotransduction, PC1 knockdown reduced the expression of both the early and late osteogenic genes, such as runt-related transcription factor 2 (Runx2), OCN, and osterix (OSX) (Wang et al., 2014). Thus, the methods to



increase the length of primary cilia is an effective treatment to promote osteogenesis, such as suppressing of adenylate cyclase III by lithium, inhibiting the anterograde direction IFT *via* deletion of Dynlt1 (Palmer et al., 2011), and regulating the actin cytoskeleton by using actin polymerization inhibitor cytochalasin D (Shi et al., 2017), as well as EMFs treatment.

To shed light on primary cilia related osteogenesis, it is necessary to mention the interesting relationship between ciliogenesis and the cell cycle (Figure 2). Primary cilia can be formed *via* either extracellular way or intracellular way (Ghossoub et al., 2011). Firstly, in the extracellular pathway, the centrioles will move beneath the cell membrane, and transform to the basal body (BB). Then, the BB will bind to the cell membrane, after that the axonemal microtubules are formed. In contrast, during the intracellular pathways, the BB will dock to the Golgi derived ciliary vesicles (CV) firstly, then the axoneme elongates, and the CV fused with the plasm membrane (Huber et al., 1993).

The BB plays a central role during the genesis of primary cilia. The BB is a cylindrical organelle that consists of nine symmetrically triplet microtubules and membranes, and the formation of BB is regulated by the cell cycle (Figure 2). During the G0/G1 phase, the centrosome moves beneath the cell membrane, the mother centriole that was used for axoneme nucleation transforms into the BB and initiates ciliogenesis (Kobayashi and Dynlacht, 2011). Then, primary cilia start to be degraded in the G2 phase and S phase. This phenomenon occurs because the centrioles are duplicating and preparing to form the mitotic spindle pole in the M phase (Ford et al., 2018). In addition, there is a ciliary transition zone (TZ) that exists in the space between the BB and the primary cilium. The special structure of TZ consists of Y-links, transition fibers, and a ciliary necklace, that not only functions as a physical boundary that can separate primary cilia from the cytoplasm (Hildebrandt et al., 2011), but also exhibits an indispensable role in controlling the movement of proteins by only selecting proteins of proper size (~10 nm) (Kee et al., 2012). Those proteins larger than 10 nm accumulate at the ciliary pocket (CiPo) instead of entering into cilia (Deane et al., 2001). Therefore, it is plausible that CiPo involves in transducing or amplifying of extracellular signals toward primary cilia (Benmerah, 2013).

Moreover, the cell cycle can also be regulated by the length of primary cilia. It was reported that an IFT88 knockdown promoted hASCs proliferation (Bodle et al., 2013). The loss of Nde1, which is a centrosome protein, leaded to the longer primary cilia, and retarded cell cycle re-entry (Kim and Tsiokas, 2011). In addition, the phosphorylation of a dynein subunit Tctex-1 resulted in disassembly of the primary cilia and maintenance in S phase (Li et al., 2011).

Due to the relationship between the cell cycle and primary cilia, the serum starvation is used to promote the assembly of primary cilia by arresting cells in the G0 phase. In contrast, serum stimulation is used to disassemble primary cilia by promoting cell cycle re-entry. It was reported the influence of serum stimulation on the disassembly of primary cilia takes effect immediately after 1–2 and 18–24 h of treatment (Pugacheva et al., 2007). Moreover, PEMF (15 Hz, 0.5 mT) can also regulate the cell cycle to inhibit apoptosis of osteocyte-like cells in the presence of primary cilia in an intensity-dependent manner (Wang et al., 2019). Altogether, considering that ciliogenesis initiates at the interphase, the cell proliferation and differentiation/ciliogenesis should be mutually exclusive. Therefore, the presence of primary cilia could be a maker to distinguish the cell cycle and differentiation.

5 The mechanism between ELF-PEMFs, primary cilia, and bone fracture

Recently, the potential mechanism of ELF-PEMFs induced bone healing has received more attention. It was reported that ELF-PEMFs (26 Hz) could regulate the level of various proteins in hMSCs, such as Akt, Erk 1/2, Hsp27, and p53, in a frequency and time-dependent exposure manner (Poh et al., 2018). Therefore, it is plausible that ELF-PEMFs can enhance osteogenesis by modulating specific signaling pathways, such as TGF- β /BMP signaling, Hedgehog (Hh) signaling, mitogenactivated protein kinase (MAPK) signaling, platelet-derived growth factor (PDGF) signaling, Wnt signaling, and calcium ion channel. In this review, the major purpose is to clarify the interaction between primary cilia, ELF-PEMFs, and TGF- β /BMP signaling.

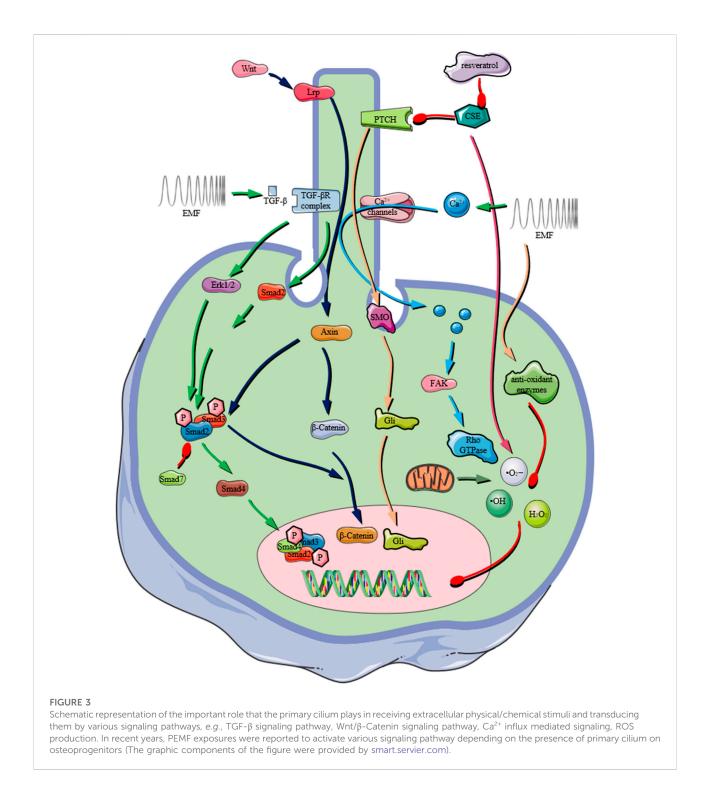
5.1 TGF- β and BMP signaling pathways

The TGF- β superfamily contains TGF- β s, bone morphogenic proteins (BMPs), activins (ACTs), and

inhibins (INHs), from which both TGF-ßs and BMPs are most outstanding in regulating the bone healing. TGF- β is a potent chemotactic stimulator that plays numerous key functions in the development and maintenance of bone system, such as cell migration, proliferation, osteogenic differentiation, and matrix mineralizationmineralization (Tsiridis et al., 2007). Typically, TGF-ßs are classified into TGF-\$1, TGF-\$2, and TGF-\$3. They share highly homologous amino acid sequences with each other (Yue and Mulder, 2001). Without the stimulation, TGF-β connects with the matrix to maintain an inactive status, under the stimulation of some bioactive molecules, such as matrix metalloproteinases (MMPs), BMPs, or reactive oxygen species (ROS), the inactive TGF- β will be released from the matrix, and transform to an active TGF-B to play various biological functions (Horiguchi et al., 2012).

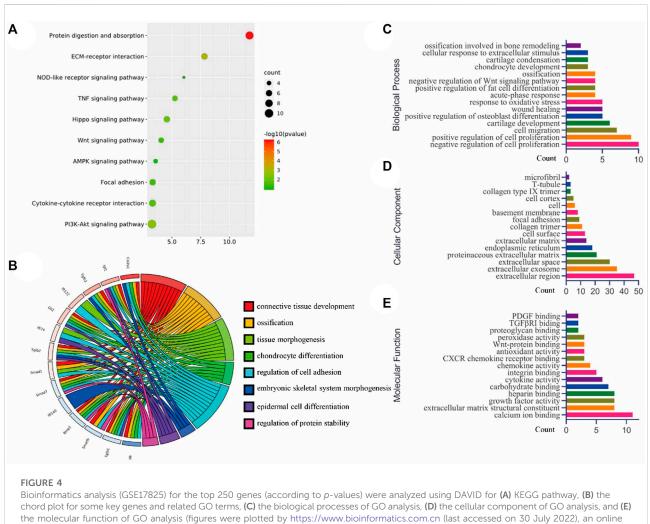
The TGF-β receptor (TGF-β RI/ALK5 and TGF-β RII) is located on the surface of the ciliary membrane, which is a heterotetrameric receptor of serine/threonine kinases (Clement et al., 2013). TGF-B RII, in the activated state after binding with ligands, will phosphorylate TGF-B RI. In canonical signaling, TGF-B RI will then phosphorylate Smad2/3. The p-Smad2/3 will bind to Smad4 to enter into the cell nucleus to regulate cellular activities. Besides, TGF-ßs also involve in many non-canonical signaling pathways, such as NF-kB signaling (Luo, 2017) and ERK/MAPK signaling (Clement et al., 2013), at the ciliary base. In 2013, Clement et al. (2013) proved that both TGF-β RI and TGF-β RII, after being stimulated by TGF-\u00b31, will move from the ciliary membrane to the ciliary base and CP. Similarly, blocking the formation of the clathrin-coated pit (CCP) and clathrin-coated vesicle (CCV) by dynasore can also impair the phosphorylation and subsequent nuclear translocation of Smad2/3, and inhibit osteogenesis (Clement et al., 2013). Thus, the TGF- β signaling is highly associated with primary cilia.

Besides, as a part of the TGF- β superfamily, BMPs play an important role in bone healing. In the canonical signaling pathway, BMP ligands first bind to the BMPRII (such as ACVR2A and ACVR2B) and phosphorylate BMPRI (such as ALK-1, -2, -3, and -6). Then, intracellular BMP effector proteins Smad1, Smad5, and Smad8 are phosphorylated. Like TGF-ß signaling, further transcription of canonical BMP signaling depends on the binding of Smad1/5/8 with Smad4, which then translocates into the cell nucleus to regulate gene transcription (Rahman et al., 2015). In 1998, it was reported that PEMF at 15 Hz promoted the expression of BMP2 and BMP4 in osteoblasts (Bodamyali et al., 1998). Further study demonstrated that PEMF promote osteogenesis by activating BMP-Smad1/5/8 signaling (Yan et al., 2015). Moreover, it was reported that PEMF (50 Hz, 0.6 mT) could promote the expression of BMPRIB and



BMPRII in the membrane of primary cilia, and activated canonical BMP signaling to enhance osteogenesis, but IFT88 siRNA treatment abolished the PEMF induced osteogenic ability (Xie et al., 2016). Furthermore, the combination of PEMF (75 Hz, 1.5 mT) and BMP2 also

showed a strong synergistic ability in promoting osteogenic differentiation of hMSCs through the canonical BMP signaling pathway (Martini et al., 2020). Take together, TGF- β s/BMPs play a central role in PEMF induced bone healing.



platform for data analysis and visualization).

5.2 The interaction between TGF- β and BMP, and the implications in the treatment of bone fracture

The biphasic effect between TGF- β and BMP plays a fundamental role in the elaborated bone dynamics during bone healing. Immediately after the bone fracture, TGF- β is released and activated at the fracture site, which leads to a robust increase in the concentration of TGF- β . The higher concentration of TGF- β brings benefits to the recruitment and proliferation of MSCs and osteoprogenitors. The major aim of the early stage is to get a sufficient cell number basis at the local fracture site, thus the BMP signaling is inhibited due to the high concentration of TGF- β . The interaction of TGF- β and BMP is dose-dependent, and TGF- β 1 at a high concentration (20 ng/ml) showed negative influences on the osteogenic ability of BMP9 (Li et al., 2012). Further study clarified that the high concentration of TGF- β 1 could inhibit BMP2 by changing its biding site directly,

and upregulate the expression of tomoregulin1 (Temff1) to suppress the BMP signaling indirectly (Xu et al., 2020).

As the concentration of TGF-\u03b3 decreases, TGF-\u03b31 exhibits positive effects on BMP signaling. At this stage, the major aim is to promote matrix mineralization and new bone formation; therefore, lots of energy and resources are regulated to promote the osteogenic ability. Thus, cell proliferation ceases, matrix mineralization initiates, and the primary cilia start to perceive the extracellular signals to regulate the healing process. A recent study demonstrated that TGF-B1 at a lower concentration (1 ng/ml) could activate Smad3, and enhanced the expression of BMP2 by promoting the combination of Smad3 and the BMP promoter (Xu et al., 2020). TGF-B1 at a low concentration (5 ng/ml) has a synergistic effect with BMP9 in inducing bone formation (Li et al., 2012). Moreover, TGF-β1 was able to prolong the duration of BMP2 activated Smad1/5/ 8 signaling, thus enhancing osteogenesis (Asparuhova et al., 2018).

Many fracture patients suffer from the complex relationship between TGF- β and BMP. Since the excessively high concentration of TGF- β exist in patients with delayed union or nonunion, BMP2 has no effect on 36% of patients with fractures (Ehnert et al., 2012). Considering the complex relationship between TGF- β and BMP in bone healing, it may be more effective and feasible to optimise EMF with the spatiotemporal specificity in promoting bone healing. Within a few days after a bone fracture, we use the EMF with a TGF- β signaling specific parameters to promote the migration and proliferation of MSCs and osteoprogenitors at the fracture site. After that, we change the EMF parameters according to a BMP specific one that activates BMP signaling to promote osteogenesis.

5.3 Crosstalk with other signaling pathways

For both TGF- β /BMP signaling pathways, they have extensive crosstalk with other signaling networks. For example, the activation of BMPs involves the phosphorylation of almost 400 proteins (Kim et al., 2009). Therefore, it is necessary to understand the influences of EMF on the crosstalk between TGF- β /BMP signaling and other signalings (Figure 3).

Moreover, we explored the deeper mechanism for post bone fracture by analysing the gene array (Series GSE17825) from the GEO database. To have an insightful understanding of the interaction between these differentially expressed genes, we analyzed the KEGG pathways. The results indicated that different signaling pathways, such as Hippo signaling, Wnt signaling, AMPK signaling, among others, were involved in the bone healing between 1-day and 5-day post fracture (Figure 4A). The relationships between key genes and gene ontology (GO) terms are shown in the chord plot (Figure 4B). The GO terms of the biological process (Figure 4C), cellular component (Figure 4D), and molecular function (Figure 4E) were summarized.

5.3.1 Wnt signaling

Wnt signaling plays a pivotal role in the development and maintenance of stem cells. The Wnt ligand can bind to the receptor Frizzled, then Lrp5 or Lrp6 interacts with Axin. After that, the function of GSK-3 β is blocked after the formation of the Dishevelled- Axin-GSK-3 β complex. As a result, the stabilised β -catenin is able to regulate gene transcription (Nusse, 2012). Nevertheless, Wnt signaling has an extensive crosstalk with TGF- β signaling, and both signaling pathways have synergistic effects in regulating cellular differentiation (Nusse, 2012). For example, Axin can promote the phosphorylation of Smad3, and Smad3 facilitates the nuclear entrance of β -catenin (Furuhashi et al., 2001). Jing et al. (2014) demonstrated PEMF (15 Hz, 2.4 mT) was excellent therapy for both disuse osteoporosis of hindlimb unloaded rats and ovariectomy-induced osteoporosis, by regulating Wnt/Lrp5/ β-Catenin signaling in vivo (Jing et al., 2013). Recently, their group proved PEMF is the effective treatment for improving bone mass and the microarchitecture of the glucocorticoidsinduced osteoporosis, through protecting the canonical Wnt/β-Catenin signaling impaired by the glucocorticoids (Cai et al., 2020a). Similarly, sinusoidal EMF (50 Hz, 1.8 mT) was also proven to contribute to enhancing bone mass through upregulating Wnt10b/β-Catenin signaling in a primary ciliadependent manner (Zhou et al., 2019). Another study showed that PEMF (15 Hz, 2.0 mT) could increase serum levels of OCN, P1NP, and upregulate the gene expression of alkaline phosphatase (ALP) and Runx2 in vivo, through activating the canonical Wnt/β-catenin signaling (Cai et al., 2020b). Moreover, ELF-PEMF (50 Hz, 20 mT, 30 min/day) can promote the osteogenic ability of adipose-derived MSCs by increasing the expression of Wnt1, Wnt3a, β-Catenin, and Lrp5 (Fathi and Farahzadi, 2017). Interestingly, the influences of PEMF on bone healing is age-dependent. It exhibited strong positive effects in 5-month rats, while showing nearly no impact on 20-month rats (Cai et al., 2020b).

5.3.2 Notch signaling

Notch signaling is activated following the binding of a ligand (such as Jagged and Delta) with the Notch receptor, and release of the Notch intracellular domain (NICD). Then NICD translocates into the nucleus to regulate Notch target genes. However, while they are important signaling in controlling cell proliferation and differentiation, Notch signaling and TGF-B/BMP signaling have extensive crosstalk and exhibit biphasic effects on bone healing. For example, the osteogenic ability of BMP can be enhanced by the ligands of Notch signaling (Tezuka et al., 2002), or inhibited by Notch1 siRNAs (Nobta et al., 2005). However, Notch signaling can also impair BMP signaling, because the overexpression of NICD produces Hey1, which impairs osteogenesis by suppressing Runx2 (Zamurovic et al., 2004). A study showed that PEMF (75 Hz, 1.5 mT) could activate canonical Notch signaling to promote the osteogenic ability of hMSCs via enhancing the expression of Notch ligand Dll4, Notch 4 receptor, as well as the nuclear target genes Hey1, Hes1, and Hes5 (Bagheri et al., 2018).

5.3.3 Hedgehog signaling

Hh signaling is an evolutionarily conserved signaling pathway that controls tissue development and regeneration. The Hh signaling pathway is regulated after the binding of Hh ligands to the Patched receptor (PTCH). Afterwards, the smoothened (SMO) is released and move to primary cilia and activate the glioma-associated oncogene homolog (Gli). Then, Gli is translocated into the nucleus to regulate gene expression.⁸⁴ Therefore, the primary cilium exerts distinct effects in transducing Hh-Gli signaling during bone healing. Moreover, TGF- β can promote the expression of Gli2 in a SMO/Hh independent manner,⁸⁵ while BMP can inhibit the expression of SMO and Gli1 (Rios et al., 2004).

IFT is also involved in Hh signaling. An IFT80 deletion blocks the canonical Hh signaling pathway, and promotes the non-canonical Hh-Gai-RhoA-ROCK signaling pathway to damage osteogenesis (Yuan et al., 2016). The knockout of IFT 140 resulted in the impaired matrix deposition *via* inhibiting the expression of PTCH1, SMO, and Gli1-3 (Li G. et al., 2018). In addition, kruppel-like factor 4 (KLF4), a transcription factor can repress osteogenesis by blocking the Hh signaling pathway (Takeuchi et al., 2018). In our previous study, we demonstrated that the cigarette smoke extract (CSE) damaged MSCs osteogenesis by impairing primary cilia and inhibiting Hh-Gli2 signaling. Meanwhile, resveratrol protected the CSEinduced dysfunction of the primary cilium and Hh-Gli2 signaling, and therefore, rescued the osteogenic ability (Sreekumar et al., 2018).

5.3.4 Calcium ion channels

Ca2+ is an important second messenger in various physiological and pathological reactions e.g., cell proliferation, metabolism, differentiation, and apoptosis (Mellstrom et al., 2008; Zhou et al., 2022). Therefore, PEMF may affect many cellular activities by reorienting charged biomolecules such as Ca²⁺ (Yan et al., 2022). It was reported that PEMF has the ability to elicit Ca2+ oscillation and increase the concentration of intracellular Ca2+ to regulate osteogenesis (Petecchia et al., 2015). In addition, a recent study reported that EMF (15 Hz, 1 mT) exhibited a lasting impact on promoting BMSCs proliferation, osteogenic ability both in vitro and in vivo, but the mechanism was unclear (Tu et al., 2018). It is plausible that Ca²⁺ channels play an important role in the legacy effects of EMF because $\mathrm{Ca}^{\scriptscriptstyle 2+}$ channels can maintain the inward current to a longer process physiologically. To investigate the signaling of Ca2+ induced-cellular activities, recent research focused on the non-selective Ca²⁺ channel piezo1 (Wang B. et al., 2022; Liu et al., 2022). Interestingly, in our unpublished work, we proved that piezo1-induced Ca2+ influx in osteoprogenitors is highly related with the improved fracture healing, the pharmaceutically blocking piezo1 impaired the fracture healing improved by PEMF exposure (Yangmengfan Chen et al., 2022). Moreover, cell migration is also an important factor in promoting wound healing, especially for orthopaedic surgery. A recent study showed that the EMF (50 Hz, 1 mT) could promote the migration of hMSCs by increasing intracellular Ca2+ and enhancing FAK/Rho GTPase pathways (Zhang et al., 2018). However, a conflicting study reported that the mechanosensory ability of primary cilia was proven to be a Ca²⁺ channel-independent because bone cells without primary cilia also show Ca²⁺ flux under fluid flow. Therefore, further study should be performed to determine the role of primary cilia in Ca²⁺ related signaling transduction.

5.4 Reactive oxygen species

ROS, a crucial second messenger, play a pivotal role in regulating cellular metabolism. ROS consists of hydrogen peroxide (H₂O₂), hydroxyl radical (•OH), superoxide anion (•O₂⁻), and hypochlorous acid (HOCl) (Wang and Zhang, 2017). These free radicals can widely affect downstream signaling, from which H2O2 can sensitively reacts to extracellular stimuli and transfuse cellular membranes to elicit a transduction cascades (Sheppard et al., 2022). Nevertheless, in high concentrations, ROS can induce strong cytotoxicity to damage cellular membranes, proteins, and DNA (Zeng et al., 2022). As a result, excessive ROS will impair the normal cellular function, structure (Yin et al., 2016), and harm the process of fracture healing (Wang K. et al., 2022). Recent studies showed that the close relationship between EMF and ROS (Li J. et al., 2018). EMF (50 Hz, 2 mT, 6 h/day) was reported to stimulate the generation of ROS (Chen et al., 2014). The same result was also reported under the treatment of 7.5 Hz EMF (Tang et al., 2016). In addition, it is interesting that the high expression of ROS can return to the base levels if the EMF exposure time is prolonged (Tang et al., 2016). However, there are some opposing results showed that 15 and 20 Hz EMF can reduce ROS generation caused by ischemia-reperfusion injury (Ma et al., 2013). The conflicting results might result from the different EMF parameters, cell types, and exposure strategy. In our previous study, the single exposure to ELF-PEMF (16 Hz) generated excessive oxidative stress on human osteoblasts, while the repetitive exposure to ELF-PEMF not only reduced the ROS level by activating various anti-oxidant enzymes but also promoted the cells viability and osteogenic ability (Ehnert et al., 2017). Therefore, applying EMF with proper parameters and exposure strategy could be a safe and effective therapy for modulating ROS level to regulate the process of fracture healing.

5.5 Other signaling pathways

There exist various serine/threonine kinases in different signaling pathways that can regulate R-Smads in the TGF- β / BMP signaling. JAK-STAT signaling is important in modulating cell proliferation, differentiation, and apoptosis. After ligands bind, the JAK kinases are phosphorylated and dimerized to phosphorylate and activate STAT. However, STAT3 was reported to inhibit the integration of Smad3/4 and nucleus translocation *via* competitive combination with Smad3 (Wang et al., 2016). NF- κ B signaling is highly related to arthritis, and chronic inflammation. TGF- β 1 can promote I κ B, and inhibit NF- κ B signaling that activated by tumour necrosis factor- α (TNF- α) (Monteleone et al., 2004). In turn, NF- κ B activated by proinflammatory cytokines that can upregulate Smad7 to suppress TGF- β signaling (Bitzer et al., 2000). Hippo signaling is conserved signaling, which regulates the development and

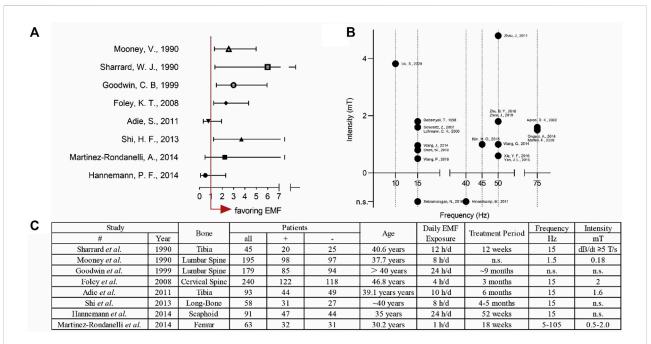


FIGURE 5

Overview of both clinical and experimental applications of EMFs. (A) Forest plot for analysing the clinical treatment of EMFs with odds ratio (OR) \pm 95% confidence interval (B) Scatter plot for summarising the parameters used in the basic experiments. (C) The detailed data for the clinical application of EMFs.

homeostasis of various organs, and tissue. Both YAP and TAZ in Hippo signaling are reported to be modulated by the mechanotransduction (Dupont et al., 2011). Therefore, the Hippo signaling pathway might involve in the primary ciliarelated signal transduction. In addition, YAP can regulate TGF-β signaling by binding to Smad1 (Alarcon et al., 2009), Smad7 (Ferrigno et al., 2002), and modulating the translocation of Smad2/3/4 complexes (Varelas et al., 2008). The MAPK pathway also has extensive crosstalk with TGF-B signaling. For example, TGF- β can promote the function of Erk and p38 (Takekawa et al., 2002); however, Erk enables the inhibition of BMP signaling by blocking the nuclear translocation of Smad1 and promoting the degradation of Smad1 through eliciting sequential phosphorylation (Sapkota et al., 2007; Eivers et al., 2009). The sinusoidal EMF (15 Hz, 1 mT) can promote AP activity, the expression of BMP2, Runx2, and OCN significantly by phosphorylating p38 and Erk1/2 (Yong et al., 2016). The PI3K-AKT pathway has complex crosstalk activities with TGF-B signaling. For example, after strong activation of PI3 signaling, Erk is suppressed, while GSK-3 β and β -catenin are activated. Then, Smad2/3 stimulates the expression of Nanog, which is involved in the self-renewal. Meanwhile, modest activation of PI3K signaling promotes Smad2/3 to regulate mesendoderm differentiation (Singh et al., 2012). Recently, it was reported that EMF (15 Hz, 1 mT) could promote osteogenesis by promoting the ion

channel P2X7, then activating Akt/GSK-3 β/β -catenin signaling *in vitro* and *in vivo* (Zhang Y. C. et al., 2019). IFT80 depletion also impairs the formation of mineralization significantly by blocking the expression and crosstalk between Hh-Gli signaling and FGF2-FGFR1-PI3K-Akt signaling (Yuan et al., 2019).

6 The clinic significance

The PEMFs were widely reported for their excellent ability to accelerate fracture healing and shorten the recovery time. We summarized the clinical cases that using ELF-PEMFs to treat bone fractures. The results indicated that ELF-PEMFs were an effective treatment for various types of bone fractures (Figures 5A,C). Recent studies showed out of 44 patients suffering from delayed bone union, or nonunion, 77.3% (34 patients) achieved clinical bone union after being treated by the PEMFs (the median treatment duration was 29.5 weeks) (Assiotis et al., 2012). The average bone healing time can be shorted from 14.7 to 8.9 weeks after the PEMFs treatment (Streit et al., 2016). In a recent prospective clinical study that involved 43 women with postmenopausal osteoporosis, PEMFs (16-22 Hz, 3-3.6 mT, 50 min/day) enable the increased expression of bone-specific AP, and decrease expression of bone resorption marker CTX by activating Wnt/ β -catenin signaling (Catalano et al., 2018). In our previous study, ELF-PEMF (16 Hz, 6–282 μ T, 7 min/day for 30 days) was used to treat 37 patients after high tibial osteotomy, and the results showed ELF-PEMF treatment could promote osseous consolidation, especially for those patients older than 50 years of age (Ziegler et al., 2019). However, there also exist many difficulties in evaluating the clinical effects of PEMFs, such as the inconsistency of fracture types, surgery impacts, PEMF treatment, and even the criteria for fracture union.

7 Outlook and discussion

With the deepening of EMFs-related research, the influences of EMFs signals on DNA raised a concern about the safety in clinical treatment. For example, EMFs can affect stem cells during early embryonic development, reduce the rate of successful assisted reproductive technologies, and increase the incidence rates of various congenital malformations for newborns, such as Wiedemann syndrome and Angelman syndrome (Maher et al., 2003). In fact, the structure of DNA is broken only under EMF with energetic radiofrequency or ionizing frequency. The EMFs with extremely low frequency are safe because this frequency can only regulate gene expression and protein synthesis (Blank and Goodman, 2009).

Primary cilia are of great importance in the development of various tissues and organs. The mutation of ciliary genes will result in more than 30 ciliopathies (Reiter and Leroux, 2017). As a manifestation of ciliopathy disorders, bone dysplasia present in many diseases such as Jeune syndrome, Ellis-van Creveld syndrome, and orofacial digital syndrome (Yuan and Yang, 2015). The Jeune syndrome mice model established by IFT140 knockout showed dwarf phenotypes with decreased bone length, density, growth retardation, and disappearance of the growth plate (Tao et al., 2019). The mutation of IFT88 resulted in attenuated mandibular development and damaged osteogenesis by inhibiting Hedgehog signaling (Kitamura et al., 2020). Considering the positive effects of ELF-EMFs on primary cilia, it is theoretically plausible that ELF-EMFs could be an effective therapy for ciliopathy disorders in the future.

In regard to the clinical efficacy of EMFs, there exist a nonlinear effect of EMF on cell activities. Chen et al. compared the influences of different intensities of PEMFs (0.6–3.6 mT, 50 Hz) on osteoblasts, the results showed all intensities of PEMF promoted cell proliferation, from which the lowest intensity (0.6 mT) was significantly higher than other intensities (Yan et al., 2015). In addition, 0.6 mT PEMF showed the best pro-osteogenic ability, while 2.4 mT PEMF exhibited almost the same ability as the control (Yan et al., 2015).

Even though many studies demonstrated that ELF-PEMFs could promote the new bone formation effectively (Ehnert et al., 2015), it is still difficult to reach the best curative effect in the clinic, because of the lack of consistency in EMFs-related research. These inconsistencies could be classified as follows:

- The EMF generating devices are different. The generating devices consist of either a single solenoid coil or a pair of Helmholtz coils. Besides, the size, component, and structure of the compartment can also affect the EMF signals.
- 2) Different laboratories choose different EMF parameters, such as frequency, intensity, and exposure time. This phenomenon often leads readers to speculate that the parameters were random, which makes it difficult for researchers to improve the efficacy by modulating the parameters. Therefore, it is necessary to fix variates and figure out the best parameters (Ross et al., 2015). We summarized the EMFs frequency and intensity used in studies that related to bone healing. The result demonstrated that the most popular frequency is 15 and 50 Hz; besides, most of the intensity is lower than 4 mT (Figure 5B). This conclusion could shed light on further study to explore the effective parameters.
- 3) The type and components of the culture medium are different and may lead to the inconsistency. For example, the content of fetal bovine serum ranges from 0.1% to 20%.
- 4) Our previous study demonstrated that ELF-PEMFs could effectively promote the formation of new bone, especially when the osteogenic ability was suppressed (Ehnert et al., 2015). Moreover, excessive exposure not only showed no positive influences on bone healing but also damaged cell activity. PEMF was demonstrated to promote cellular proliferation for these cells in the proliferation stage, while stimulating the matrix formation in the differentiation stage. In addition, the inconsistencies result from the position, dimension of culture plate on the EMF generating devices, as well as cell densities in the plate.
- 5) The process of bone healing is a continuous activity without a cut-off point for total union. Therefore, it is necessary to have uniform criteria to define the bone union (Bhandari et al., 2012).
- 6) One of the advances of using EMF to treat bone nonunion is the cost-effectiveness; thus, it is better to analyze the healthcare cost, and consider the socioeconomic burden, such as increasing the EMF efficacy, and shorting the exposure duration (Button et al., 2009).

In summary, this review elucidated the important functions and insightful signaling pathways of EMFs and primary cilia on the bone healing process, which has essential clinical value and research implications for the development and optimization of EMFs for treating bone fractures in the future.

Author contributions

Conceptualization, YC; methodology, YC; software, YC and KC; formal analyses, YC; investigation, YC; resources YC; data curation, YC and KC; writing—original draft preparation, YC and CL and KC; writing—review and editing, all authors; visualization, YC and KC; supervision, XS and YC; project ad-ministration KC; funding acquisition KC; All authors have read and agreed to the published version of the manuscript.

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References

Alarcon, C., Zaromytidou, A. I., Xi, Q. R., Gao, S., Yu, J. Z., Fujisawa, S., et al. (2009). Nuclear CDKs drive Smad transcriptional activation and turnover in BMP and TGF-beta pathways. *Cell* 139, 757–769. doi:10.1016/j.cell.2009.09.035

Antonsson, E. K., and Mann, R. W. (1985). The frequency content of gait. J. Biomech. 18, 39-47. doi:10.1016/0021-9290(85)90043-0

Asparuhova, M. B., Caballe-Serrano, J., Buser, D., and Chappuis, V. (2018). Boneconditioned medium contributes to initiation and progression of osteogenesis by exhibiting synergistic TGF- β 1/BMP-2 activity. *Int. J. Oral Sci.* 10, 20. doi:10.1038/ s41368-018-0021-2

Assiotis, A., Sachinis, N. P., and Chalidis, B. E. (2012). Pulsed electromagnetic fields for the treatment of tibial delayed unions and nonunions. A prospective clinical study and review of the literature. *J. Orthop. Surg. Res.* 7, 24. doi:10.1186/1749-799X-7-24

Bagheri, L., Pellati, A., Rizzo, P., Aquila, G., Massari, L., De Mattei, M., et al. (2018). Notch pathway is active during osteogenic differentiation of human bone marrow mesenchymal stem cells induced by pulsed electromagnetic fields. *J. Tissue Eng. Regen. Med.* 12, 304–315. doi:10.1002/term.2455

Bassett, C. A. L. (1993). Beneficial-effects of electromagnetic-fields. J. Cell. Biochem. 51, 387-393. doi:10.1002/jcb.2400510402

Bassett, C. A., Mitchell, S. N., Norton, L., and Pilla, A. (1978). Repair of nonunions by pulsing electromagnetic fields. *Acta Orthop. belg.* 44, 706–724.

Becker, R. O., Bachman, C. H., and Friedman, H. (1962). The direct current control system. A link between environment and organism. *N. Y. State J. Med.* 62, 1169–1176.

Benmerah, A. (2013). The ciliary pocket. Curr. Opin. Cell Biol. 25, 78-84. doi:10. 1016/j.ceb.2012.10.011

Bhandari, M., Fong, K., Sprague, S., Williams, D., and Petrisor, B. (2012). Variability in the definition and perceived causes of delayed unions and nonunions: A cross-sectional, multinational survey of orthopaedic surgeons. *J. Bone Jt. Surg. Am.* 94, e1091–e1096. doi:10.2106/JBJS.K.01344

Bitzer, M., von Gersdorff, G., Liang, D., Dominguez-Rosales, A., Beg, A. A., Rojkind, M., et al. (2000). A mechanism of suppression of TGF– β /SMAD signaling by NF- κ B/RelA. *Genes Dev.* 14, 187–197. doi:10.1101/gad.14.2.187

Blank, M., and Goodman, R. (2009). Electromagnetic fields stress living cells. Pathophysiology 16, 71-78. doi:10.1016/j.pathophys.2009.01.006

Bodamyali, T., Bhatt, B., Hughes, F. J., Winrow, V. R., Kanczler, J. M., Simon, B., et al. (1998). Pulsed electromagnetic fields simultaneously induce osteogenesis and upregulate transcription of bone morphogenetic proteins 2 and 4 in rat osteoblasts *in vitro*. *Biochem. Biophys. Res. Commun.* 250, 458–461. doi:10.1006/bbrc.1998.9243

Bodle, J. C., Rubenstein, C. D., Phillips, M. E., Bernacki, S. H., Qi, J., Banes, A. J., et al. (2013). Primary cilia: The chemical antenna regulating human adiposederived stem cell osteogenesis. *Plos One* 8, e62554. doi:10.1371/journal.pone. 0062554

Button, M. L., Sprague, S., Gharsaa, O., Latouche, S., and Bhandari, M. (2009). Economic evaluation of bone stimulation modalities: A systematic review of the literature. *Indian J. Orthop.* 43, 168–174. doi:10.4103/0019-5413.50852

Conflict of interest

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Cai, J., Shao, X., Yan, Z. D., Liu, X. Y., Yang, Y. Q., Luo, E. P., et al. (2020b). Differential skeletal response in adult and aged rats to independent and combinatorial stimulation with pulsed electromagnetic fields and mechanical vibration. *Faseb J.* 34, 3037–3050. doi:10.1096/fj.201902779R

Cai, J., Shao, X., Yang, Q., Yang, Y., Yan, Z., Luo, E., et al. (2020a). Pulsed electromagnetic fields modify the adverse effects of glucocorticoids on bone architecture, bone strength and porous implant osseointegration by rescuing bone-anabolic actions. *Bone* 133, 115266. doi:10.1016/j.bone.2020.115266

Catalano, A., Loddo, S., Bellone, F., Pecora, C., Lasco, A., and Morabito, N. (2018). Pulsed electromagnetic fields modulate bone metabolism via RANKL/OPG and wnt/ β -catenin pathways in women with postmenopausal osteoporosis: A pilot study. *Bone* 116, 42–46. doi:10.1016/j.bone.2018.07.010

Chen, Y., Hong, L., Zeng, Y., Shen, Y., and Zeng, Q. (2014). Power frequency magnetic fields induced reactive oxygen species-related autophagy in mouse embryonic fibroblasts. *Int. J. Biochem. Cell Biol.* 57, 108–114. doi:10.1016/j. biocel.2014.10.013

Claes, L., Recknagel, S., and Ignatius, A. (2012). Fracture healing under healthy and inflammatory conditions. *Nat. Rev. Rheumatol.* 8, 133–143. doi:10.1038/nrrheum.2012.1

Clement, C. A., Ajbro, K. D., Koefoed, K., Vestergaard, M. L., Veland, I. R., de Jesus, M. P. R. H., et al. (2013). TGF-Beta signaling is associated with endocytosis at the pocket region of the primary cilium. *Cell Rep.* 3, 1806–1814. doi:10.1016/j. celrep.2013.05.020

Daish, C., Blanchard, R., Fox, K., Pivonka, P., and Pirogova, E. (2018). The application of pulsed electromagnetic fields (PEMFs) for bone fracture repair: Past and perspective findings. *Ann. Biomed. Eng.* 46, 525–542. doi:10.1007/s10439-018-1982-1

De Mattos, C. B. R., Binitie, O., and Dormans, J. P. (2012). Pathological fractures in children. *Bone Jt. Res.* 1, 272–280. doi:10.1302/2046-3758.110.2000120

Deane, J. A., Cole, D. G., Seeley, E. S., Diener, D. R., and Rosenbaum, J. L. (2001). Localization of intraflagellar transport protein IFT52 identifies basal body transitional fibers as the docking site for IFT particles. *Curr. Biol.* 11, 1586–1590. doi:10.1016/s0960-9822(01)00484-5

Dupont, S., Morsut, L., Aragona, M., Enzo, E., Giulitti, S., Cordenonsi, M., et al. (2011). Role of YAP/TAZ in mechanotransduction. *Nature* 474, 179–183. doi:10. 1038/nature10137

Eguether, T., San Agustin, J. T., Keady, B. T., Jonassen, J. A., Liang, Y., Francis, R., et al. (2014). IFT27 links the BBSome to IFT for maintenance of the ciliary signaling compartment. *Dev. Cell* 31, 279–290. doi:10.1016/j.devcel. 2014.09.011

Ehnert, S., Falldorf, K., Fentz, A. K., Ziegler, P., Schroter, S., Freude, T., et al. (2015). Primary human osteoblasts with reduced alkaline phosphatase and matrix mineralization baseline capacity are responsive to extremely low frequency pulsed electromagnetic field exposure - clinical implication possible. *Bone Rep.* 3, 48–56. doi:10.1016/j.bonr.2015.08.002

Ehnert, S., Fentz, A. K., Schreiner, A., Birk, J., Wilbrand, B., Ziegler, P., et al. (2017). Extremely low frequency pulsed electromagnetic fields cause antioxidative

defense mechanisms in human osteoblasts via induction of \bullet O2- and H2O2. Sci. Rep. 7, 14544. doi:10.1038/s41598-017-14983-9

Ehnert, S., Zhao, J., Pscherer, S., Freude, T., Dooley, S., Kolk, A., et al. (2012). Transforming growth factor beta1 inhibits bone morphogenic protein (BMP)-2 and BMP-7 signaling via upregulation of ski-related novel protein N (SnoN): Possible mechanism for the failure of BMP therapy? *BMC Med.* 10, 101. doi:10.1186/1741-7015-10-101

Eivers, E., Fuentealba, L. C., Sander, V., Clemens, J. C., Hartnett, L., and Robertis, E. M. (2009). Mad is required for wingless signaling in wing development and segment patterning in Drosophila. *Plos One* 4, e6543. doi:10.1371/journal.pone. 0006543

Fathi, E., and Farahzadi, R. (2017). Enhancement of osteogenic differentiation of rat adipose tissue-derived mesenchymal stem cells by zinc sulphate under electromagnetic field via the PKA, ERK1/2 and Wnt/ β -catenin signaling pathways. *PLoS One* 12, e0173877. doi:10.1371/journal.pone.0173877

Ferrigno, O., Lallemand, F., Verrecchia, F., L'Hoste, S., Camonis, J., Atfi, A., et al. (2002). Yes-associated protein (YAP65) interacts with Smad7 and potentiates its inhibitory activity against TGF-beta/Smad signaling. *Oncogene* 21, 4879–4884. doi:10.1038/sj.onc.1205623

Finkelstein, E., Chang, W., Chao, P. H., Gruber, D., Minden, A., Hung, C. T., et al. (2004). Roles of microtubules, cell polarity and adhesion in electric-field-mediated motility of 3T3 fibroblasts. *J. Cell Sci.* 117, 1533–1545. doi:10.1242/jcs.00986

Ford, M. J., Yeyati, P. L., Mali, G. R., Keighren, M. A., Waddell, S. H., Mjoseng, H. K., et al. (2018). A cell/cilia cycle biosensor for single-cell kinetics reveals persistence of cilia after G1/S transition is a general property in cells and mice. *Dev. Cell* 47, 509–523. doi:10.1016/j.devcel.2018.10.027

Funk, R. H. W., Monsees, T., and Ozkucur, N. (2009). Electromagnetic effects from cell biology to medicine. *Prog. Histochem. Cytochem.* 43, 177–264. doi:10. 1016/j.proghi.2008.07.001

Furuhashi, M., Yagi, K., Yamamoto, H., Furukawa, Y., Shimada, S., Nakamura, Y., et al. (2001). Axin facilitates Smad3 activation in the transforming growth factor beta signaling pathway. *Mol. Cell. Biol.* 21, 5132–5141. doi:10.1128/MCB.21.15. 5132-5141.2001

Ghossoub, R., Molla-Herman, A., Bastin, P., and Benmerah, A. (2011). The ciliary pocket: A once-forgotten membrane domain at the base of cilia. *Biol. Cell* 103, 131–144. doi:10.1042/BC20100128

Goodman, R., and Shirley-Henderson, A. (1990). Exposure of cells to extremely low-frequency electromagnetic fields: Relationship to malignancy? *Cancer* 2, 355–359.

Hildebrandt, F., Benzing, T., and Katsanis, N. (2011). Mechanisms of disease: Ciliopathies. N. Engl. J. Med. 364, 1533-1543. doi:10.1056/NEJMra1010172

Horiguchi, M., Ota, M., and Rifkin, D. B. (2012). Matrix control of transforming growth factor-beta function. J. Biochem. 152, 321-329. doi:10.1093/jb/mvs089

Huber, L. A., Pimplikar, S., Parton, R. G., Virta, H., Zerial, M., and Simons, K. (1993). Rab8, a small gtpase involved in vesicular traffic between the tgn and the basolateral plasma-membrane. *J. Cell Biol.* 123, 35–45. doi:10.1083/jcb.123.1.35

Jacobs, C. R., Temiyasathit, S., and Castillo, A. B. (2010). Osteocyte mechanobiology and pericellular mechanics. *Annu. Rev. Biomed. Eng.* 12, 369–400. doi:10.1146/annurev-bioeng-070909-105302

Jing, D., Cai, J., Wu, Y., Shen, G., Li, F., Xu, Q., et al. (2014). Pulsed electromagnetic fields partially preserve bone mass, microarchitecture, and strength by promoting bone formation in hindlimb-suspended rats. *J. Bone Min. Res.* 29, 2250–2261. doi:10.1002/jbmr.2260

Jing, D., Li, F. J., Jiang, M. G., Cai, J., Wu, Y., Xie, K. N., et al. (2013). Pulsed electromagnetic fields improve bone microstructure and strength in ovariectomized rats through a Wnt/Lrp5/ β -catenin signaling-associated mechanism. *Plos One* 8, e79377. doi:10.1371/journal.pone.0079377

Kaku, M., and Komatsu, Y. (2017). Functional diversity of ciliary proteins in bone development and disease. *Curr. Osteoporos. Rep.* 15, 96–102. doi:10.1007/s11914-017-0351-6

Keady, B. T., Samtani, R., Tobita, K., Tsuchya, M., San Agustin, J. T., Follit, J. A., et al. (2012). IFT25 links the signal-dependent movement of Hedgehog components to intraflagellar transport. *Dev. Cell* 22, 940–951. doi:10.1016/j.devcel.2012.04.009

Kee, H. L., Dishinger, J. F., Blasius, T. L., Liu, C. J., Margolis, B., and Verhey, K. J. (2012). A size-exclusion permeability barrier and nucleoporins characterize a ciliary pore complex that regulates transport into cilia. *Nat. Cell Biol.* 14, 431–437. doi:10. 1038/ncb2450

Khan, S. N., Solaris, J., Ramsey, K. E., Yang, X., Bostrom, M. P., Stephan, D., et al. (2008). Identification of novel gene expression in healing fracture callus tissue by DNA microarray. *HSS J.* 4, 149–160. doi:10.1007/s11420-008-9087-2

Kim, B. G., Lee, J. H., Ahn, J. M., Park, S. K., Cho, J. H., Hwang, D., et al. (2009). 'Two-stage double-technique hybrid (TSDTH)' identification strategy for the analysis of BMP2-induced transdifferentiation of premyoblast C2C12 cells to osteoblast. J. Proteome Res. 8, 4441-4454. doi:10.1021/pr900231a

Kim, S., and Tsiokas, L. (2011). Cilia and cell cycle re-entry More than a coincidence. Cell Cycle 10, 2683–2690. doi:10.4161/cc.10.16.17009

Kitamura, A., Kawasaki, M., Kawasaki, K., Meguro, F., Yamada, A., Nagai, T., et al. (2020). Ift88 is involved in mandibular development. *J. Anat.* 236, 317–324. doi:10.1111/joa.13096

Kobayashi, T., and Dynlacht, B. D. (2011). Regulating the transition from centriole to basal body. J. Cell Biol. 193, 435-444. doi:10.1083/jcb.201101005

Kozminski, K. G., Johnson, K. A., Forscher, P., and Rosenbaum, J. L. (1993). A motility in the eukaryotic flagellum unrelated to flagellar beating. *Proc. Natl. Acad. Sci. U. S. A.* 90, 5519–5523. doi:10.1073/pnas.90.12.5519

Li, A., Saito, M., Chuang, J. Z., Tseng, Y. Y., Dedesma, C., Tomizawa, K., et al. (2011). Ciliary transition zone activation of phosphorylated Tctex-1 controls ciliary resorption, S-phase entry and fate of neural progenitors. *Nat. Cell Biol.* 13, 402–411. doi:10.1038/ncb2218

Li, G., Liu, M., Zhang, S., Wan, H., Zhang, Q., Yue, R., et al. (2018). Essential role of IFT140 in promoting dentinogenesis. *J. Dent. Res.* 97, 423–431. doi:10.1177/0022034517741283

Li, J., Zhang, Q., Ren, C., Wu, X., Zhang, Y., Bai, X., et al. (2018). Low-intensity pulsed ultrasound prevents the oxidative stress induced endothelial-mesenchymal transition in human aortic endothelial cells. *Cell. Physiol. biochem.* 45, 1350–1365. doi:10.1159/000487561

Li, R. D., Deng, Z. L., Hu, N., Liang, X., Liu, B., Luo, J. Y., et al. (2012). Biphasic effects of TGF beta 1 on BMP9-induced osteogenic differentiation of mesenchymal stem cells. *BMB Rep.* 45, 509–514. doi:10.5483/bmbrep.2012.45.9.053

Liu, Y., Tian, H., Hu, Y., Cao, Y., Song, H., Lan, S., et al. (2022). Mechanosensitive Piezo1 is crucial for periosteal stem cell-mediated fracture healing. *Int. J. Biol. Sci.* 18, 3961–3980. doi:10.7150/ijbs.71390

Luo, K. (2017). Signaling cross talk between TGF-β/smad and other signaling pathways. *Cold Spring Harb. Perspect. Biol.* 9, a022137. doi:10.1101/cshperspect.a022137

Ma, S., Zhang, Z., Yi, F., Wang, Y., Zhang, X., Li, X., et al. (2013). Protective effects of low-frequency magnetic fields on cardiomyocytes from ischemia reperfusion injury via ROS and NO/ONOO. *Oxid. Med. Cell. Longev.* 2013, 529173. doi:10.1155/2013/529173

Maher, E. R., Afnan, M., and Barratt, C. L. (2003). Epigenetic risks related to assisted reproductive technologies: Epigenetics, imprinting, ART and icebergs? *Hum. Reprod.* 18, 2508–2511. doi:10.1093/humrep/deg486

Malone, A. M. D., Anderson, C. T., Tummala, P., Kwon, R. Y., Johnston, T. R., Stearns, T., et al. (2007). Primary cilia mediate mechanosensing in bone cells by a calcium-independent mechanism. *Proc. Natl. Acad. Sci. U. S. A.* 104, 13325–13330. doi:10.1073/pnas.0700636104

Martini, F., Pellati, A., Mazzoni, E., Salati, S., Caruso, G., Contartese, D., et al. (2020). Bone morphogenetic protein-2 signaling in the osteogenic differentiation of human bone marrow mesenchymal stem cells induced by pulsed electromagnetic fields. *Int. J. Mol. Sci.* 21, E2104. doi:10.3390/ijms21062104

Mellstrom, B., Savignac, M., Gomez-Villafuertes, R., and Naranjo, J. R. (2008). Ca2+-operated transcriptional networks: Molecular mechanisms and *in vivo* models. *Physiol. Rev.* 88, 421-449. doi:10.1152/physrev.00041.2005

Monteleone, G., Mann, J., Monteleone, I., Vavassori, P., Bremner, R., Fantini, M., et al. (2004). A failure of transforming growth factor-beta1 negative regulation maintains sustained NF-kappaB activation in gut inflammation. *J. Biol. Chem.* 279, 3925–3932. doi:10.1074/jbc.M303654200

Nobta, M., Tsukazaki, T., Shibata, Y., Xin, C., Moriishi, T., Sakano, S., et al. (2005). Critical regulation of bone morphogenetic protein-induced osteoblastic differentiation by Delta1/Jagged1-activated Notch1 signaling. J. Biol. Chem. 280, 15842–15848. doi:10.1074/jbc.M412891200

Nusse, R. (2012). Wnt signaling. Cold Spring Harb. Perspect. Biol. 4, a011163. doi:10.1101/cshperspect.a011163

Palmer, K. J., MacCarthy-Morrogh, L., Smyllie, N., and Stephens, D. J. (2011). A role for Tctex-1 (DYNLT1) in controlling primary cilium length. *Eur. J. Cell Biol.* 90, 865–871. doi:10.1016/j.ejcb.2011.05.003

Petecchia, L., Sbrana, F., Utzeri, R., Vercellino, M., Usai, C., Visai, L., et al. (2015). Electro-magnetic field promotes osteogenic differentiation of BM-hMSCs through a selective action on Ca(2+)-related mechanisms. *Sci. Rep.* 5, 13856. doi:10.1038/ srep13856

Poh, P. S. P., Seeliger, C., Unger, M., Falldorf, K., Balmayor, E. R., and van Griensven, M. (2018). Osteogenic effect and cell signaling activation of extremely low-frequency pulsed electromagnetic fields in adipose-derived mesenchymal stromal cells. *Stem Cells Int.* 2018, 5402853. doi:10.1155/2018/5402853

Prevo, B., Scholey, J. M., and Peterman, E. J. G. (2017). Intraflagellar transport: Mechanisms of motor action, cooperation, and cargo delivery. *Febs J.* 284, 2905–2931. doi:10.1111/febs.14068

Pugacheva, E. N., Jablonski, S. A., Hartman, T. R., Henske, E. P., and Golemis, E. A. (2007). HEF1-dependent Aurora A activation induces disassembly of the primary cilium. *Cell* 129, 1351–1363. doi:10.1016/j.cell.2007.04.035

Rahman, M. S., Akhtar, N., Jamil, H. M., Banik, R. S., and Asaduzzaman, S. M. (2015). TGF- β /BMP signaling and other molecular events: Regulation of osteoblastogenesis and bone formation. *Bone Res.* 3, 15005. doi:10.1038/boneres. 2015.5

Reiter, J. F., and Leroux, M. R. (2017). Genes and molecular pathways underpinning ciliopathies. *Nat. Rev. Mol. Cell Biol.* 18, 533–547. doi:10.1038/nrm.2017.60

Rios, I., Alvarez-Rodriguez, R., Marti, E., and Pons, S. (2004). Bmp2 antagonizes sonic hedgehog-mediated proliferation of cerebellar granule neurones through Smad5 signalling. *Development* 131, 3159–3168. doi:10.1242/dev.01188

Ross, C. L., Siriwardane, M., Almeida-Porada, G., Porada, C. D., Brink, P., Christ, G. J., et al. (2015). The effect of low-frequency electromagnetic field on human bone marrow stem/progenitor cell differentiation. *Stem Cell Res.* 15, 96–108. doi:10.1016/j.scr.2015.04.009

Sapkota, G., Alarcon, C., Spagnoli, F. M., Brivanlou, A. H., and Massague, J. (2007). Balancing BMP signaling through integrated inputs into the Smad1 linker. *Mol. Cell* 25, 441–454. doi:10.1016/j.molcel.2007.01.006

Schwartz, E. A., Leonard, M. L., Bizios, R., and Bowser, S. S. (1997). Analysis and modeling of the primary cilium bending response to fluid shear. *Am. J. Physiol.* 272, F132–F138. doi:10.1152/ajprenal.1997.272.1.F132

Seeger-Nukpezah, T., and Golemis, E. A. (2012). The extracellular matrix and ciliary signaling. *Curr. Opin. Cell Biol.* 24, 652–661. doi:10.1016/j.ceb.2012.06.002

Sheppard, A. J., Barfield, A. M., Barton, S., and Dong, Y. (2022). Understanding reactive oxygen species in bone regeneration: A glance at potential therapeutics and bioengineering applications. *Front. Bioeng. Biotechnol.* 10, 836764. doi:10.3389/fbioe.2022.836764

Shi, W. G., Xie, Y. F., He, J. P., Zhou, J., Gao, Y. H., Wei, W. J., et al. (2017). Microgravity induces inhibition of osteoblastic differentiation and mineralization through abrogating primary cilia. *Sci. Rep.* 7, 1866. doi:10.1038/s41598-017-02049-9

Singh, A. M., Reynolds, D., Cliff, T., Ohtsuka, S., Mattheyses, A. L., Sun, Y. H., et al. (2012). Signaling Network crosstalk in human pluripotent cells: A smad2/3-regulated switch that controls the balance between self-renewal and differentiation. *Cell Stem Cell* 10, 312–326. doi:10.1016/j.stem.2012.01.014

Song, B., Zhao, M., Forrester, J. V., and McCaig, C. D. (2002). Electrical cues regulate the orientation and frequency of cell division and the rate of wound healing *in vivo. Proc. Natl. Acad. Sci. U. S. A.* 99, 13577–13582. doi:10.1073/pnas.202235299

Sreekumar, V., Aspera-Werz, R., Ehnert, S., Strobel, J., Tendulkar, G., Heid, D., et al. (2018). Resveratrol protects primary cilia integrity of human mesenchymal stem cells from cigarette smoke to improve osteogenic differentiation *in vitro*. *Arch. Toxicol.* 92, 1525–1538. doi:10.1007/s00204-017-2149-9

Streit, A., Watson, B. C., Granata, J. D., Philbin, T. M., Lin, H. N., O'Connor, J. P., et al. (2016). Effect on clinical outcome and growth factor synthesis with adjunctive use of pulsed electromagnetic fields for fifth metatarsal nonunion fracture: A double-blind randomized study. *Foot Ankle Int.* 37, 919–923. doi:10.1177/1071100716652621

Takekawa, M., Tatebayashi, K., Itoh, F., Adachi, M., Imai, K., and Saito, H. (2002). Smad-dependent GADD45beta expression mediates delayed activation of p38 MAP kinase by TGF-beta. *Embo J.* 21, 6473–6482. doi:10.1093/emboj/cdf643

Takeuchi, Y., Kito, A., Itoh, S., Naruse, H., Fujikawa, J., Sadek, K. M., et al. (2018). Kruppel-Like Factor 4 represses osteoblast differentiation via ciliary Hedgehog signaling. *Exp. Cell Res.* 371, 417–425. doi:10.1016/j.yexcr.2018.09.002

Tang, R., Xu, Y., Ma, F., Ren, J., Shen, S., Du, Y., et al. (2016). Extremely low frequency magnetic fields regulate differentiation of regulatory T cells: Potential role for ROS-mediated inhibition on AKT. *Bioelectromagnetics* 37, 89–98. doi:10.1002/bem.21954

Tao, D., Xue, H., Zhang, C., Li, G., and Sun, Y. (2019). The role of IFT140 in osteogenesis of adult mice long bone. *J. Histochem. Cytochem.* 67, 601–611. doi:10. 1369/0022155419847188

Taschner, M., Bhogaraju, S., and Lorentzen, E. (2012). Architecture and function of IFT complex proteins in ciliogenesis. *Differentiation*. 83, S12–S22. doi:10.1016/j. diff.2011.11.001

Temiyasathit, S., and Jacobs, C. R. (2010). Osteocyte primary cilium and its role in bone mechanotransduction. *Ann. N. Y. Acad. Sci.* 1192, 422–428. doi:10.1111/j. 1749-6632.2009.05243.x

Tezuka, K., Yasuda, M., Watanabe, N., Morimura, N., Kuroda, K., Miyatani, S., et al. (2002). Stimulation of osteoblastic cell differentiation by Notch. *J. Bone Min. Res.* 17, 231–239. doi:10.1359/jbmr.2002.17.2.231

The classic (1977). The classic: Fundamental aspects of fracture treatment by Iwao Yasuda, reprinted from J. Kyoto Med. Soc., 4:395-406, 1953. *Clin. Orthop. Relat. Res.* 124, 5–7.

Tonna, E. A., and Lampen, N. M. (1972). Electron microscopy of aging skeletal cells. I. Centrioles and solitary cilia. J. Gerontol. 27, 316–324. doi:10.1093/geronj/27.3.316

Tsiridis, E., Upadhyay, N., and Giannoudis, P. (2007). Molecular aspects of fracture healing: Which are the important molecules? *Injury* 38, S11–S25. doi:10. 1016/j.injury.2007.02.006

Tu, C., Xiao, Y. F., Ma, Y. Z., Wu, H., and Song, M. Y. (2018). The legacy effects of electromagnetic fields on bone marrow mesenchymal stem cell self-renewal and multiple differentiation potential. *Stem Cell Res. Ther.* 9, 215. doi:10.1186/s13287-018-0955-5

Tummala, P., Arnsdorf, E. J., and Jacobs, C. R. (2010). The role of primary cilia in mesenchymal stem cell differentiation: A pivotal switch in guiding lineage commitment. *Cell. Mol. Bioeng.* 3, 207–212. doi:10.1007/s12195-010-0127-x

Varelas, X., Sakuma, R., Samavarchi-Tehrani, P., Peerani, R., Rao, B. M., Dembowy, J., et al. (2008). TAZ controls Smad nucleocytoplasmic shuttling and regulates human embryonic stem-cell self-renewal. *Nat. Cell Biol.* 10, 837–848. doi:10.1038/ncb1748

Victoria, G., Petrisor, B., Drew, B., and Dick, D. (2009). Bone stimulation for fracture healing: Whats all the fuss. *Indian J. Orthop.* 43, 117–120. doi:10.4103/0019-5413.50844

Wang, B., Li, G., Zhu, Q., Liu, W., Ke, W., Hua, W., et al. (2022). Bone repairment via mechanosensation of Piezo1 using wearable pulsed triboelectric nanogenerator. *Small* 18, e2201056. doi:10.1002/smll.202201056

Wang, G., Yu, Y., Sun, C., Liu, T., Liang, T., Zhan, L., et al. (2016). STAT3 selectively interacts with Smad3 to antagonize TGF- β signalling. Oncogene 35, 4422. doi:10.1038/onc.2016.145

Wang, H., Sun, W., Ma, J. Q., Pan, Y. C., Wang, L., and Zhang, W. B. (2014). Polycystin-1 mediates mechanical strain-induced osteoblastic mechanoresponses via potentiation of intracellular calcium and Akt/ β -catenin pathway. *Plos One* 9, e91730. doi:10.1371/journal.pone.0091730

Wang, H. Z., and Zhang, X. (2017). Magnetic fields and reactive oxygen species. Int. J. Mol. Sci. 18, E2175. doi:10.3390/ijms18102175

Wang, K., Zhou, C., Li, L., Dai, C., Wang, Z., Zhang, W., et al. (2022). Aucubin promotes bone-fracture healing via the dual effects of anti-oxidative damage and enhancing osteoblastogenesis of hBM-MSCs. *Stem Cell Res. Ther.* 13, 424. doi:10. 1186/s13287-022-03125-2

Wang, P., Tang, C., Wu, J., Yang, Y., Yan, Z., Liu, X., et al. (2019). Pulsed electromagnetic fields regulate osteocyte apoptosis, RANKL/OPG expression, and its control of osteoclastogenesis depending on the presence of primary cilia. *J. Cell. Physiol.* 234, 10588–10601. doi:10.1002/jcp.27734

Xie, Y. F., Shi, W. G., Zhou, J., Gao, Y. H., Li, S. F., Fang, Q. Q., et al. (2016). Pulsed electromagnetic fields stimulate osteogenic differentiation and maturation of osteoblasts by upregulating the expression of BMPRII localized at the base of primary cilium. *Bone* 93, 22–32. doi:10.1016/j.bone.2016.09.008

Xu, J., Liu, J., Gan, Y., Dai, K., Zhao, J., Huang, M., et al. (2020). High-dose TGF- $\beta 1$ impairs mesenchymal stem cell-mediated bone regeneration via Bmp2 inhibition. J. Bone Min. Res. 35, 167–180. doi:10.1002/jbmr.3871

Yan, J. L., Zhou, J., Ma, H. P., Ma, X. N., Gao, Y. H., Shi, W. G., et al. (2015). Pulsed electromagnetic fields promote osteoblast mineralization and maturation needing the existence of primary cilia. *Mol. Cell. Endocrinol.* 404, 132–140. doi:10. 1016/j.mce.2015.01.031

Yan, Z., Wang, D., Cai, J., Shen, L., Jiang, M., Liu, X., et al. (2022). High-specificity protection against radiation-induced bone loss by a pulsed electromagnetic field. *Sci. Adv.* 8, eabq0222. doi:10.1126/sciadv.abq0222

Yangmengfan Chen, M. M. M., Braun, B. J., Falldorf, K., Histing, T., Nussler, A. K., and Ehnert, S. (2022). Intermittent ELF-PEMFs improve osteoporosis through activat-ing piezo1-induced Ca2+ influx. *Pending*.

Yin, C., Luo, X., Duan, Y., Duan, W., Zhang, H., He, Y., et al. (2016). Neuroprotective effects of lotus seedpod procyanidins on extremely low frequency electromagnetic field-induced neurotoxicity in primary cultured hippocampal neurons. *Biomed. Pharmacother.* 82, 628–639. doi:10.1016/j.biopha.2016.05.032

Yong, Y., Ming, Z. D., Feng, L., Chun, Z. W., and Hua, W. (2016). Electromagnetic fields promote osteogenesis of rat mesenchymal stem cells through the PKA and ERK1/2 pathways. *J. Tissue Eng. Regen. Med.* 10, E537–E545. doi:10.1002/term.1864

Yuan, J., Xin, F., and Jiang, W. X. (2018). Underlying signaling pathways and therapeutic applications of pulsed electromagnetic fields in bone repair. *Cell. Physiol. biochem.* 46, 1581–1594. doi:10.1159/000489206

Yuan, X., Cao, J., He, X. N., Serra, R., Qu, J., Cao, X., et al. (2016). Ciliary IFT80 balances canonical versus non-canonical hedgehog signalling for osteoblast differentiation. *Nat. Commun.* 7, 11024. doi:10.1038/ncomms11024

Yuan, X., Liu, M., Cao, X., and Yang, S. Y. (2019). Ciliary IFT80 regulates dental pulp stem cells differentiation by FGF/FGFR1 and Hh/BMP2 signaling. *Int. J. Biol. Sci.* 15, 2087–2099. doi:10.7150/ijbs.27231

Yuan, X., and Yang, S. (2016). Primary cilia and intraflagellar transport proteins in bone and cartilage. J. Dent. Res. 95, 1341–1349. doi:10.1177/0022034516652383

Yuan, X., and Yang, S. Y. (2015). Cilia/Ift protein and motor-related bone diseases and mouse models. *Front. Biosci.* 20, 515–555. doi:10.2741/4323

Yue, J. B., and Mulder, K. M. (2001). Transforming growth factor-beta signal transduction in epithelial cells. *Pharmacol. Ther.* 91, 1–34. doi:10.1016/s0163-7258(01)00143-7

Zamurovic, N., Cappellen, D., Rohner, D., and Susa, M. (2004). Coordinated activation of Notch, Wnt, and transforming growth factor-beta signaling pathways in bone morphogenic protein 2-induced osteogenesis Notch - target gene Heyl inhibits mineralization and Runx2 transcriptional activity. *J. Biol. Chem.* 279, 37704–37715. doi:10.1074/jbc.M403813200

Zeng, Q., Ma, X., Song, Y., Chen, Q., Jiao, Q., and Zhou, L. (2022). Targeting regulated cell death in tumor nanomedicines. *Theranostics* 12, 817–841. doi:10. 7150/thno.67932

Zhang, C. Y., Zhang, S., and Sun, Y. (2019). Expression of IFT140 during bone development. J. Histochem. Cytochem. 67, 723–734. doi:10.1369/0022155419859357

Zhang, Y. C., Li, W. K., Liu, C. X., Yan, J. Y., Yuan, X. F., Wang, W., et al. (2019). Electromagnetic field treatment increases purinergic receptor P2X7 expression and activates its downstream Akt/GSK3 β/β -catenin axis in mesenchymal stem cells under osteogenic induction. *Stem Cell Res. Ther.* 10, 407. doi:10.1186/s13287-019-1497-1

Zhang, Y., Yan, J., Xu, H., Yang, Y., Li, W., Wu, H., et al. (2018). Extremely low frequency electromagnetic fields promote mesenchymal stem cell migration by increasing intracellular Ca(2+) and activating the FAK/Rho GTPases signaling pathways *in vitro. Stem Cell Res. Ther.* 9, 143. doi:10. 1186/s13287-018-0883-4

Zhou, J., Gao, Y. H., Zhu, B. Y., Shao, J. L., Ma, H. P., Xian, C. J., et al. (2019). Sinusoidal electromagnetic fields increase peak bone mass in rats by activating wnt10b/β-catenin in primary cilia of osteoblasts. *J. Bone Min. Res.* 34, 1336–1351. doi:10.1002/jbmr.3704

Zhou, L. Q., Feng, W., Chen, L., Huang, H., Huang, S. J., Chen, Q., et al. (2022). Targeting acidogenic metabolism by engineering self-catalytic siRNA nanocarriers/ nanocatalysts for amplified tumor apoptosis/ferroptosis. *Nano Today* 46, 101623. doi:10.1016/j.nantod.2022.101623

Ziegler, P., Nussler, A. K., Wilbrand, B., Falldorf, K., Springer, F., Fentz, A. K., et al. (2019). Pulsed electromagnetic field therapy improves osseous consolidation after high tibial osteotomy in elderly patients-A randomized, placebo-controlled, double-blind trial. *J. Clin. Med.* 8, E2008. doi:10.3390/jcm8112008

Glossary

OH hydroxyl radical ACTs activins ACVR activin A receptor ALK activin receptor-like kinase ALP alkaline phosphatase **BB** basal body BMPs bone morphogenic proteins CCPs clathrin-coated pits CCVs clathrin-coated vesicles CDE clathrin-dependent endocytosis CiPo ciliary pocket CSE cigarette smoke extract CV ciliary vesicles DIC differential interference contrast ECM extracellular matrix EF electronic field PEMF extremly low frequency-pulsed electromagnetic field EMF electromagnetic field Erk extracellular signal-regulated kinase Gli glioma-associated oncogene homolog H_2O_2 hydrogen peroxide Hh hedgehog HOCl hypochloric acid Hz herz IFT intraflagellar transport

KLF4 kruppel-Like Factor 4 LLC large latent TGF-β1 complex LTBP latent TGF- β binding protein MAPK mitogen-activated protein kinase MF magnetic field MMPs matrix metalloproteinases MSCs mesenchymal stem cells **NF-κB** nuclear factor-κB NICD notch intracellular domain **OCN** osteocalcin **OSX** osterix **OVX** ovariectomy PC1 polycystin-1 PDGF platelet-derived growth factor PEMF pulsed electromagnetic field PTCH patched receptor RANKL receptor Activator of NF-KB Ligand ROS reactive oxygen species Runx2 runt-related transcription factor 2 **SLC** small latent TGF- β complex SMO Smoothened TEP transepithelial potential difference $TGF-\beta$ transforming growth factor- β TGF-β R TGF-β receptor TNF-a tumor necrosis factor-a TZ transition zone

INHs inhibins