

Macrophages in homeostatic immune function

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INTRODUCTION

Leukocytes, consisting of diverse cell types, are the cellular constituents of the body's immune system. As they patrol throughout the blood and lymphatic systems, these cells become recruited to infected or damaged tissue and then act to restore the integrity of the site. Leucocytes share a common origin from haematopoietic stem cells and develop into a variety of subsets according to distinct differentiation stimuli. The mononuclear phagocyte system (MPS) constitutes a subgroup of leukocytes comprising monocytes, macrophages, and dendritic cells; the latter representing specialized antigen-presenting cells linking innate and adaptive immune responses (reviewed in Geissmann et al., 2010).

The cells of the MPS are not only involved in inflammatory and anti-infective processes but also play an important homeostatic role in maintaining the steady state of the tissue (Medzhitov, 2008; Lutz and Kurts, 2009; Pollard, 2009); for example, monocytes derived from the spleen act to rapidly promote tissue repair after myocardial infarction (Swirski et al., 2009). Particularly, macrophages have essential and diverse roles in regulating tissue homeostasis. Over a century ago Metchnikoff proposed that macrophages are not only involved in combating invading intruders, but play other roles within the body to maintain homeostasis. He used the terms "physiological"

Macrophages are not only involved in inflammatory and anti-infective processes, but also play an important role in maintaining tissue homeostasis. In this review, we summarize recent evidence investigating the role of macrophages in controlling angiogenesis, metabolism as well as salt and water balance. Particularly, we summarize the importance of macrophage tonicity enhancer binding protein (TonEBP, also termed nuclear factor of activated T-cells 5 [NFAT5]) expression in the regulation of salt and water homeostasis. Further understanding of homeostatic macrophage function may lead to new therapeutic approaches to treat ischemia, hypertension and metabolic disorders.

Keywords: macrophage polarization, arteriogenesis, angiogenesis, lipid metabolism, peroxisome proliferator activator receptor (PPAR), salt, Na⁺ storage, tonicity enhance binding protein (TonEBP)/nuclear factor of activated T-cells 5(NFAT5)

and "pathological" (e.g., induced by pathogens) tissue insults to describe the initial "disharmony" facing different cells within a multicellular organism. Whilst these two types of insults have different and competing demands on the tissue, in both situations macrophages have a central homeostatic role to re-establish the steady state of the tissue (Tauber, 2003). Recent classification proposes that macrophages have a continuum of phenotypic subsets, each with overlapping functions ranging from classically activated (M1) to alternatively activated (M2) macrophages, where the latter includes both wound healing and regulatory macrophages (Gordon, 2003; Mosser and Edwards, 2008; Murray and Wynn, 2011; Locati et al., 2013; Mantovani et al., 2013). In addition to these well recognized activities, macrophages are important for various other functions including tissue development, such as neuronal patterning, bone morphogenesis, and generation of adipose tissue (Pollard, 2009), promoting angiogenesis and arteriogenesis (Murdoch et al., 2008; Pollard, 2009), and the maintenance of internal body fluids or the "milieu interieur." Regulation of the *mileu interiur* is especially important for tissue homeostasis as, according to Bernard, this guarantees a "free and independent" life (Bernard, 1957). In this review, we will summarize recent developments of the role of macrophages in regulating blood supply and metabolism. Finally, we will highlight the role of macrophages as central regulators of internal body fluids *via* the expression of NFAT5. Further understanding into this emerging concept of homeostatic macrophage function may offer new insight into the regulation of energy and electrolyte metabolism, and thus offer new therapeutics to so-called "Western diseases."

MACROPHAGES AS ANGIOGENIC AND ARTERIOGENIC ACCESSORY CELLS

Tissue hypoxia drives the development of new blood vessels from existing blood vessels (angiogenesis), and the remodeling of existing collateral vessels (arteriogenesis), in order to ensure sufficient tissue perfusion and hence, tissue oxygenation (Potente et al., 2011). It has been shown that macrophages play an important role in both processes, as angiogenic, and arteriogenic accessory cells (Pollard, 2004; Murdoch et al., 2008; Coffelt et al., 2009; David Dong et al., 2009; Nucera et al., 2011; Chambers et al., 2013; Owen and Mohamadzadeh, 2013). Myeloid cells are first attracted to the site of injury, for example by the chemokine CCL2 (Low-Marchelli et al., 2013). Accordingly, interference with local monocyte attraction to ischemic tissue resulted in flap necrosis due to impaired flap revascularization (Khan et al., 2013). Once at the ischemic site, macrophages are exposed to vesseland tissue-derived cytokines [such as Angiopoietin (ANG) 1, ANG2, vascular endothelial growth factor A (VEGF)], which reprogram them to become highly angiogenic and arteriogenic accessory cells (Avraham-Davidi et al., 2013; Hamm et al., 2013). The expression and composition of these vessel- and tissuederived cytokines are tightly regulated at the ischemic site, and may critically affect the angiogenic and arteriogenic function of macrophages (Folkman, 2006; Saharinen et al., 2010; Saharinen and Alitalo, 2011). For example, ANG1-mediated macrophage reprogramming resulted in the repression of the oxygen-sensitive prolyl hydroxylase domain (PHD) protein 2 (PHD2). PHD2 repression promoted a M2-like, proarteriogenic phenotype of macrophages by activating canonical nuclear factor 'kappa-lightchain-enhancer' of activated B-cells (NF-kB) signaling (Takeda et al., 2011). Furthermore, ANG1-mediated Phd2-repression enhanced the expression of ANG-receptor TIE2, which amplifies ANG-dependent TIE2 signaling in a positive feedback loop and hence promoted vessel maturation (Hamm et al., 2013). In addition, local tissue hypoxia and endothelial cell derived signals might maintain this regulatory circuit by promoting TIE2 expression (Lewis et al., 2007; He et al., 2012).

In many different models of angiogenesis, M2-like alternatively activated macrophages supported the proliferation and migration of endothelial cells and vessel sprouting (Jetten et al., 2014; Fantin et al., 2010; Marchetti et al., 2011). Further to serving as a source of the potent angiogenic mediators such as VEGF and fibroblast growth factor (FGF)-2 (Schulze-Osthoff et al., 1990; Xiong et al., 1998; Dirkx et al., 2006), IL-4-driven alternatively activated macrophages promoted the release of VEGF from the tissue matrix, thereby enhancing the sprouting of vessels (Zajac et al., 2013). This was shown to be as a result of a blockade of the tissue inhibitor of metalloproteinase 1 (TIMP1) gene expression, that promotes the secretion of highly angiogenic matrix metalloproteinase (MMP) 9, which in turn results in the release of matrix-sequestered angiogenic growth factors such as VEGF and FGF-2 from the tissue (Zajac et al., 2013). Release of macrophage-derived MMP9 might be further augmented by hypoxia, as MMP9 expression is known to be governed by hypoxia inducible factor (HIF) 2a(alpha) -signaling (Yang et al., 2010).

In addition to the above described proangiogenic role of IL-4-dependent M2-like macrophages (Zajac et al., 2013; Jetten et al., 2014), non-IL-4 stimulated macrophages also have important roles. In this context, ANG2-dependent TIE2-signaling in macrophages was also found to promote angiogenesis in models of inflammation and cancer (Coffelt et al., 2010, 2011; Mazzieri et al., 2011; Krausz et al., 2012). Additionally, hypoxia was found to enhance ANG2 expression in murine and human macrophages, which may subsequently boost their proangiogenic function (Fang et al., 2009).

These data demonstrate that macrophage cell function is critically involved in angio- and arteriogenesis (**Figure 1**). Given the role of T cell-derived cytokines on macrophage polarization and activation it is obvious that alteration in T cell activation will also affect angiogenesis and arteriogenesis, and will thus bring another level of complexity to the effect of immune cells on vascular biology (Starnes et al., 2001; Naldini et al., 2003; Stabile et al., 2003, 2006; Facciabene et al., 2012).

MACROPHAGES AS GLUCOSE AND LIPID SENSORS

Whilst cytokines and chemokines are the main drivers of the activation and function of macrophages, recent studies have revealed that macrophages also respond to environmental cues in the form of small metabolites such as glucose, lipids, and sodium chloride (to be discussed further in the following section). These small metabolites also influence programming of macrophages into either classical or alternative subsets, and can thus modulate macrophage function.

Macrophages infiltrate and reside in nearly every tissue, including adipose. Accompanied with the observation that macrophages accumulate within adipose tissue with obesity (Weisberg et al., 2003; Xu et al., 2003), there has been great



tissue- and T cell-derived signals.

interest on the effect of lipids on macrophage function and activation. Macrophages take up lipids via scavenger receptors, such as CD36 and scavenger receptor A (SR-A). This process is not subject to a negative feedback mechanism and as such, where there is excess lipid present macrophages can become loaded with lipid and form pro-atherogenic foam cells (Nagy et al., 2012). In cases of overnutrition, where the adipose tissue is overwhelmed with nutrients resulting in various amounts of cellular stress (reviewed by Odegaard and Chawla, 2013), macrophages accumulate within adipose tissue and subsequently switch from an alternative activated (M2) phenotype to a classically activated (M1), suggesting that excess fat can enhance the activation of inflammatory signaling pathways (Lumeng et al., 2007a). This has also been demonstrated by in vitro experiments, where incubation of macrophage with free fatty acids led to the activation of Toll-like receptor 4 signaling, NF-kB activation and subsequently fatty acid-induced insulin resistance (Shi et al., 2006; Pal et al., 2012). The JNK signaling pathway has additionally been shown to be involved in the activation of inflammatory M1 macrophages and the development of obesity and insulin-resistance (Han et al., 2013). This was also shown by the deletion of JNK1 in hematopoietic-derived cells, which subsequently resulted in protection against diet-induced inflammation and insulin resistance without affecting obesity (Solinas et al., 2007).

Under normal conditions, the transcriptional effect of the uptake of lipids by macrophages is the activation of peroxisome proliferator activator receptors (PPAR) (Ricote et al., 1998). In humans, there are three PPAR subtypes (α , δ , and γ), which are expressed in a variety of cell types and tissues. These three PPAR's act as transcription factors and coordinate the transcription of molecules important for every facet of fatty acid metabolism (reviewed by Desvergne et al., 2006). In terms of macrophage activation, both PPAR-8 and -y have been shown to be especially important in modulating alternative macrophage activation (reviewed extensively by Nagy et al., 2012). In a recent study, PPAR-8 was implicated in Salmonella replication, where upon infection M2 macrophages had elevated PPAR-8 expression. As the deletion of PPAR- δ in macrophages prevented the replication and survival of the bacteria, the authors propose that Salmonella have evolved to utilize the metabolic state of M2 macrophages in order to survive (Eisele et al., 2013). PPAR- γ seems to be of particular importance to alternative macrophage activation, as its expression is induced by IL-4 (Huang et al., 1999). It has subsequently been shown that signal transducer and activator of transcription 6 (STAT6), the downstream transcription factor of IL-4 signaling, physically interacts with PPAR-y at transcriptionally important regions of M2 signature genes to augment their expression (Szanto et al., 2010). Macrophage-specific PPARy knockout mice have also been shown to have an attenuated M2 phenotype: the expression of M2 signature genes such as Arg1 were blunted, which was coupled with a reduced fatty acid oxidative metabolism (Odegaard et al., 2007). Surprisingly PPAR-y knockout mice had an increased weight gain on a high fat diet, which was attributed to a decrease in the number of alternatively activated macrophages, and thus a decrease in the homeostatic capacity provided by these macrophages under conditions of overnutrition to ensure that efficient lipid metabolism is

occurring (Odegaard et al., 2007). It is tempting to speculate that PPAR- γ -dependent signaling pathways may empower the macrophages to efficiently clear cholesterol from tissue (Martel et al., 2013).

In addition to obesity, the onset of diabetes (type 1 or 2) is closely linked to macrophage activation and function. Type 2 diabetes in particular is closely linked to macrophage function, where the infiltration of macrophages into adipose tissue and their modulation of the inflammation level of this tissue greatly contributes to the development of insulin resistance (Arkan et al., 2005; Kanda et al., 2006; Weisberg et al., 2006; Lumeng et al., 2007b). In addition, macrophages have been shown to directly infiltrate the pancreatic islet and subsequently modulate the function of this organ to promote the onset of diabetes (Ehses et al., 2007). Conversely, glucose itself seems to have an influence on macrophage function (de Souza et al., 2008; Kanter et al., 2012). Furthermore, the state of glucose metabolism within the cell is also important for macrophage polarization. Whilst M1 macrophages rely on glycolytic metabolism via HIF1a (Cramer et al., 2003), M2 macrophages utilize fatty acid oxidation via PPAR (as discussed above; Vats et al., 2006). More recently, two studies have identified pathways important for mediating the metabolic activity of macrophages, and thus their activation state. Haschemi et al. demonstrated that the carbohydrate kinaselike (CARKL) protein controls M1-M2 activation by directing the metabolic reprogramming of the macrophage from oxidative phosphorylation to glycolysis (Haschemi et al., 2012). It has also been shown that the mammalian target of rapamycin (mTOR), a master signaling pathway involved in growth and metabolism, is involved in regulating the switch from M2 to M1 activation (Byles et al., 2013). These studies all point toward glucose itself being a "physiological insult" which influences macrophage activation and eventually, their homeostatic function.

MACROPHAGES ACT AS LOCAL SENSORS AND REGULATORS OF ELECTROLYTE COMPOSITION IN THE SKIN INTERSTITIUM

Recently, Machnik et al attributed a new role to macrophages in regulating internal body fluids (Machnik et al., 2009, 2010). The conventional key regulator for salt and water regulation is the kidney. This traditional concept is based on the idea that body fluids readily equilibrate and that Na⁺ is the major extracellular cation and holds water in the extracellular space due to its osmotic activity. Consequently, it is believed that renal excretion of excess Na⁺ is sufficient to govern intravascular and interstitial electrolyte content. Thus, researchers interested in the regulation of intravascular and interstitial fluids have previously focused on mechanisms as to how the kidneys handle Na⁺. In contrast to this view, there is now ample evidence that Na⁺ is stored without commensurate water retention in the skin (reviewed in Titze and Machnik, 2010; Titze, 2014). This suggests that the electrolytes in the skin do not readily equilibrate with plasma, and hence escape renal homeostatic control. These findings support the idea that the intravascular and interstitial spaces are two distinct extracellular electrolyte compartments which are regulated separately. In a series of experiments by our laboratory, we have identified that the electrolyte composition of the skin is regulated

by macrophages and by its local interstitial lymph capillary system (Machnik et al., 2009, 2010; Wiig et al., 2013).

Macrophages infiltrate to the sites of Na⁺ and Cl⁻ overload in the skin which display a hypertonic microenvironment, indicating that the salt-gradient may be the driving force of macrophage cell attraction (Muller et al., 2013). These recruited macrophages sense the interstitial electrolyte composition and subsequently upregulate the transcription factor, tonicity enhancer binding protein [TonEBP, also termed nuclear factor of activated T-cells 5 (NFAT5)], which is an essential transcription factor required for the expression of osmoprotective genes in response to hypertonicity-induced osmotic stress (Halterman et al., 2012).

The molecular events which lead to NFAT5 activation has been the focus of many studies. The p38/mitogen activated protein kinase (MAPK) signaling pathway is one which is activated by osmotic stress in both mammalian (Han et al., 1994) and yeast cells [Brewster et al. (1993); via its MAPK homolog high osmolarity glycerol response protein 1 (HOG1)]. This has been confirmed in many other studies (Shapiro and Dinarello, 1995; Nadkarni et al., 1999; Ko et al., 2002; Morancho et al., 2008; Kuper et al., 2009; Roth et al., 2010; Kleinewietfeld et al., 2013). However, what remains to be shown are the macrophage-specific molecular events which occur upon osmotic stress, upstream of the activation of p38/ MAPK. One possibility is MAPK1 phosphatase-1 (MKP-1), which augments p38/ MAPK signaling upon high concentrations of NaCl (Zhou et al., 2008). Whether this molecule is also important for macrophages has not been shown. In addition to p38/ MAPK, several other signaling cascades have been induced in response to osmotic stress. These include the phosphatidylinositol 3-kinase signaling cascades (Irarrazabal et al., 2004), protein kinase A dependent processes (Ferraris et al., 2002), as well as a Rac1/osmosensing scaffold (Zhou et al., 2011). More recently, a sucrose nonfermenting-1-related serine/ threonine kinase (SIK1) was identified as a sensor of extracellular Na⁺ gradients, subsequently transducing this information into signaling cascades which modulates cellular function (Sjostrom et al., 2007). Importantly, this axis has been demonstrated in macrophages, where inhibition of SIK1 activity was shown to affect M2 regulatory macrophage activation (Clark et al., 2012). Also, serum glucocorticoid kinase-1 (SGK1) is another saltinducible kinase (reviewed extensively by Lang et al., 2006) which has recently been identified as an important modulator of IL-17 producing CD4⁺ T helper (Th17) cell activation (Wu et al., 2013). How the salt-inducible kinases such as SGK1 and SIK1 interact with NFAT5 in macrophages is unknown. Furthermore, how these signaling cascades are induced and interplay with each other and which effect they have on NFAT5 activation especially in macrophages is yet to be elucidated.

Our laboratory has demonstrated one biological outcome for the activation of NFAT5 in macrophages. The induction of NFAT5 in macrophages of the skin was shown to directly govern the expression of vascular endothelial growth factor C (VEGF-C), resulting in the hyperplasia of lymph capillaries *via* and interaction with the VEGF receptor 3 (VEGFR3) (Wiig et al., 2013). Similar to a breakdown of the renal salt and water excretion, failure of this local extrarenal macrophage-dependent control



mechanism to regulate interstitial electrolyte and water homeostasis, resulted in arterial hypertension and massive disturbances in skin electrolyte composition (Machnik et al., 2009, 2010; Wiig et al., 2013). Thus, these data indicate that the traditional model for electrolyte and water homeostasis, which considers the intravascular and interstitial space as one functional extracellular compartment, is an oversimplification. Instead, the interstitial space is a distinct compartment that relies on tissue-specific regulatory mechanisms for controlling its electrolyte content; this role being fulfilled by macrophages (**Figure 2**).

Recently, it has become possible to measure the concentration of Na⁺ in the tissue of humans *via* a non-invasive technique using ²³Na-MRI (Kopp et al., 2012a). These studies revealed that there is an increased local sodium storage in humans that suffer from hyperaldosteronism, hyernatriemia, and hypertension (Kopp et al., 2012a,b, 2013). Harnessing the potential of macrophages to regulate salt and water balance may therefore be of interest to physicians that aim to treat arterial hypertension as well as salt-balance disorders.

CONCLUSION

The new homeostatic functions reviewed here, extends the classical role of macrophages as cells which remove foreign microorganisms from the body, to those which tightly regulate the microenvironments of the body to ensure correct blood supply and concentrations of metabolites, electrolytes, and maintain proper tissue function. In light of the amount of overnutrition and excessive dietary intake of sugar, salt and saturated fats in western civilizations, further understanding of the molecular details by which macrophages sense these metabolites is warranted. The identification of other metabolites and electrolytes which influence macrophage polarization and function may reveal new mechanisms by which diseases such as hypertension, type II diabetes and autoimmune diseases may occur.

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REFERENCES

- Arkan, M. C., Hevener, A. L., Greten, F. R., Maeda, S., Li, Z. W., Long, J. M., et al. (2005). IKK-beta links inflammation to obesity-induced insulin resistance. *Nat. Med.* 11, 191–198. doi: 10.1038/nm1185
- Avraham-Davidi, I., Yona, S., Grunewald, M., Landsman, L., Cochain, C., Silvestre, J. S., et al. (2013). On-site education of VEGF-recruited monocytes improves their performance as angiogenic and arteriogenic accessory cells. *J. Exp. Med.* 210, 2611–2625. doi: 10.1084/jem.20120690
- Bernard, C. (1957). An Introduction to the Study of Experimental Medicine. New York, NY: Dover Publications.
- Brewster, J. L., de Valoir, T., Dwyer, N. D., Winter, E., and Gustin, M. C. (1993). An osmosensing signal transduction pathway in yeast. *Science* 259, 1760–1763.
- Byles, V., Covarrubias, A. J., Ben-Sahra, I., Lamming, D. W., Sabatini, D. M., Manning, B. D., et al. (2013). The TSC-mTOR pathway regulates macrophage polarization. *Nat. Commun.* 4:2834. doi: 10.1038/ncomms3834
- Chambers, S. E., O'Neill, C. L., O'Doherty, T. M., Medina, R. J., and Stitt, A. W. (2013). The role of immune-related myeloid cells in angiogenesis. *Immunobiology* 218, 1370–1375. doi: 10.1016/j.imbio.2013.06.010
- Clark, K., MacKenzie, K. F., Petkevicius, K., Kristariyanto, Y., Zhang, J., Choi, H. G., et al. (2012). Phosphorylation of CRTC3 by the salt-inducible kinases controls the interconversion of classically activated and regulatory macrophages. *Proc. Natl. Acad. Sci. U.S.A.* 109, 16986–16991. doi: 10.1073/pnas.1215450109
- Coffelt, S. B., Chen, Y, Muthana, M., Welford, A. F., Tal, A. O., Scholz, A., et al. (2011). Angiopoietin 2 stimulates TIE2-expressing monocytes to suppress T cell activation and to promote regulatory T cell expansion. *J. Immunol.* 186, 4183–4190. doi: 10.4049/jimmunol.1002802
- Coffelt, S. B., Hughes, R., and Lewis, C. E. (2009). Tumor-associated macrophages: effectors of angiogenesis and tumor progression. *Biochim. Biophys. Acta* 1796, 11–18. doi: 10.1016/j.bbcan.2009.02.004
- Coffelt, S. B., Tal, A. O., Scholz, A., De Palma, M., Patel, S., Urbich, C., et al. (2010). Angiopoietin-2 regulates gene expression in TIE2-expressing monocytes and augments their inherent proangiogenic functions. *Cancer Res.* 70, 5270–5280. doi: 10.1158/0008-5472.CAN-10-0012
- Cramer, T., Yamanishi, Y., Clausen, B. E., Forster, I., Pawlinski, R., Mackman, N., et al. (2003). HIF-1alpha is essential for myeloid cell-mediated inflammation. *Cell* 112, 645–657. doi: 10.1016/S0092-8674(03)00154-5
- David Dong, Z. M., Aplin, A. C., and Nicosia, R. F. (2009). Regulation of angiogenesis by macrophages, dendritic cells, and circulating myelomonocytic cells. *Curr. Pharm. Des.* 15, 365–379. doi: 10.2174/138161209787315783
- de Souza, L. F., Jardim, F. R., Sauter, I. P., de Souza, M, and Bernard, E. A. (2008). High glucose increases RAW 264.7 macrophages activation by lipoteichoic acid from *Staphylococcus aureus*. *Clin. Chim. Acta* 398, 130–133. doi: 10.1016/j.cca.2008.09.007
- Desvergne, B., Michalik, L., and Wahli, W. (2006). Transcriptional regulation of metabolism. *Physiol. Rev.* 86, 465–514. doi: 10.1152/physrev.00025.2005
- Dirkx, A. E., Oude Egbrink, M. G., Wagstaff, J., and Griffioen, A. W. (2006). Monocyte/macrophage infiltration in tumors: modulators of angiogenesis. *J. Leukoc. Biol.* 80, 1183–1196. doi: 10.1189/jlb.0905495
- Ehses, J. A., Perren, A., Eppler, E., Ribaux, P., Pospisilik, J. A., Maor-Cahn, R., et al. (2007). Increased number of islet-associated macrophages in type 2 diabetes. *Diabetes* 56, 2356–2370. doi: 10.2337/db06-1650
- Eisele, N. A., Ruby, T., Jacobson, A., Manzanillo, P. S., Cox, J. S., Lam, L., et al. (2013). Salmonella require the fatty acid regulator PPARdelta for the establishment of a metabolic environment essential for long-term persistence. *Cell Host Microbe* 14, 171–182. doi: 10.1016/j.chom.2013.07.010
- Facciabene, A., Motz, G. T., and Coukos, G. (2012). T-regulatory cells: key players in tumor immune escape and angiogenesis. *Cancer Res.* 72, 2162–2171. doi: 10.1158/0008-5472.CAN-11-3687
- Fang, H. Y., Hughes, R., Murdoch, C., Coffelt, S. B., Biswas, S. K., Harris, A. L., et al. (2009). Hypoxia-inducible factors 1 and 2 are important transcriptional effectors in primary macrophages experiencing hypoxia. *Blood* 114, 844–859. doi: 10.1182/blood-2008-12-195941
- Fantin, A., Vieira, J. M., Gestri, G., Denti, L., Schwarz, Q., Prykhozhij, S., et al. (2010). Tissue macrophages act as cellular chaperones for vascular anastomosis

downstream of VEGF-mediated endothelial tip cell induction. *Blood* 116, 829–840. doi: 10.1182/blood-2009-12-257832

- Ferraris, J. D., Persaud, P., Williams, C. K., Chen, Y., and Burg, M. B. (2002). cAMP-independent role of PKA in tonicity-induced transactivation of tonicityresponsive enhancer/ osmotic response element-binding protein. *Proc. Natl. Acad. Sci. U.S.A.* 99, 16800–16805. doi: 10.1073/pnas.222659799
- Folkman, J. (2006). Angiogenesis. Annu. Rev. Med. 57:1–18. doi: 10.1146/annurev.med.57.121304.131306
- Geissmann, F., Manz, M. G., Jung, S., Sieweke, M. H., Merad, M., and Ley, K. (2010). Development of monocytes, macrophages, and dendritic cells. *Science* 327, 656–661. doi: 10.1126/science.1178331
- Gordon, S. (2003). Alternative activation of macrophages. Nat. Rev. Immunol. 3, 23–35. doi: 10.1038/nri978
- Halterman, J. A., Kwon, H. M., and Wamhoff, B. R. (2012). Tonicity-independent regulation of the osmosensitive transcription factor TonEBP (NFAT5). Am. J. Physiol. Cell Physiol. 302, C1–C8. doi: 10.1152/ajpcell.00327.2011
- Hamm, A., Veschini, L., Takeda, Y., Costa, S., Delamarre, E., Squadrito, M. L., et al. (2013). PHD2 regulates arteriogenic macrophages through TIE2 signalling. *EMBO Mol. Med.* 5, 843–857. doi: 10.1002/emmm.201302695
- Han, J., Lee, J. D., Bibbs, L., and Ulevitch, R. J. (1994). A MAP kinase targeted by endotoxin and hyperosmolarity in mammalian cells. *Science* 265, 808–811.
- Han, M. S., Jung, D. Y., Morel, C., Lakhani, S. A., Kim, J. K., Flavell, R. A., et al. (2013). JNK expression by macrophages promotes obesity-induced insulin resistance and inflammation. *Science* 339, 218–222. doi: 10.1126/science.1227568
- Haschemi, A., Kosma, P., Gille, L., Evans, C. R., Burant, C. F., Starkl, P., et al. (2012). The sedoheptulose kinase CARKL directs macrophage polarization through control of glucose metabolism. *Cell Metab.* 15, 813–826. doi: 10.1016/j.cmet.2012.04.023
- He, H, Xu, J. Y., Warren, C. M., Duan, D., Li, X. M., Wu, L., et al. (2012). Endothelial cells provide an instructive niche for the differentiation and functional polarization of M2-like macrophages. *Blood* 120, 3152–3162. doi: 10.1182/blood-2012-04-422758
- Huang, J. T., Welch, J. S., Ricote, M., Binder, C. J., Willson, T. M., Kelly, C., et al. (1999). Interleukin-4-dependent production of PPAR-gamma ligands in macrophages by 12/15-lipoxygenase. *Nature* 400, 378–382. doi: 10.1038/ 22572
- Irarrazabal, C. E., Liu, J. C., Burg, M. B., and Ferraris, J. D. (2004). ATM, a DNA damage-inducible kinase, contributes to activation by high NaCl of the transcription factor TonEBP/OREBP. *Proc. Natl. Acad. Sci. U.S.A.* 101, 8809–8814. doi: 10.1073/pnas.0403062101
- Jetten, N., Verbruggen, S., Gijbels, M. J., Post, M. J., De Winther, M. P., and Donners, M. (2014). Anti-inflammatory M2, but not pro-inflammatory M1 macrophages promote angiogenesis *in vivo*. Angiogenesis 17, 109–118. doi: 10.1007/s10456-013-9381-6
- Kanda, H., Tateya, S., Tamori, Y., Kotani, K., Hiasa, K., Kitazawa, R., et al. (2006). MCP-1 contributes to macrophage infiltration into adipose tissue, insulin resistance, and hepatic steatosis in obesity. J. Clin. Invest. 116, 1494–1505. doi: 10.1172/JCI26498
- Kanter, J. E., Kramer, F., Barnhart, S., Averill, M, Vivekanandan-Giri, A., Vickery, T., et al. (2012). Diabetes promotes an inflammatory macrophage phenotype and atherosclerosis through acyl-CoA synthetase 1. *Proc. Natl. Acad. Sci. U.S.A.* 109, E715–E724. doi: 10.1073/pnas.1111600109
- Khan, B., Rangasamy, S., McGuire, P. G., and Howdieshell, T. R. (2013). The role of monocyte subsets in myocutaneous revascularization. J. Surg. Res. 183, 963–975. doi: 10.1016/j.jss.2013.02.019
- Kleinewietfeld, M., Manzel, A., Titze, J., Kvakan, H., Yosef, N., Linker, R. A., et al. (2013). Sodium chloride drives autoimmune disease by the induction of pathogenic TH17 cells. *Nature* 496, 518–522. doi: 10.1038/nature11868
- Ko, B. C., Lam, A. K., Kapus, A., Fan, L., Chung, S. K., and Chung, S. (2002). Fyn and p38 signaling are both required for maximal hypertonic activation of the osmotic response element-binding protein/tonicity-responsive enhancerbinding protein (OREBP/TonEBP). J. Biol. Chem. 277, 46085–46092. doi: 10.1074/jbc.M208138200
- Kopp, C., Linz, P., Dahlmann, A., Hammon, M., Jantsch, J., Muller, D. N., et al. (2013). 23Na magnetic resonance imaging-determined tissue sodium in healthy subjects and hypertensive patients. *Hypertension* 61, 635–640. doi: 10.1161/HYPERTENSIONAHA.111.00566

- Kopp, C., Linz, P., Hammon, M., Schofl, C., Grauer, M., Eckardt, K. U., et al. (2012b). Seeing the sodium in a patient with hypernatremia. *Kidney Int.* 82, 1343–1344. doi: 10.1038/ki.2012.314
- Kopp, C., Linz, P., Wachsmuth, L., Dahlmann, A., Horbach, T., Schofl, C., et al. (2012a). (23)Na magnetic resonance imaging of tissue sodium. *Hypertension* 59, 167–172. doi: 10.1161/HYPERTENSIONAHA.111.183517
- Krausz, S., Garcia, S., Ambarus, C. A., de Launay, D., Foster, M., Naiman, B., et al. (2012). Angiopoietin-2 promotes inflammatory activation of human macrophages and is essential for murine experimental arthritis. *Ann. Rheum. Dis.* 71, 1402–1410. doi: 10.1136/annrheumdis-2011-200718
- Kuper, C., Steinert, D., Fraek, M. L., Beck, F. X., and Neuhofer, W. (2009). EGF receptor signaling is involved in expression of osmoprotective TonEBP target gene aldose reductase under hypertonic conditions. *Am. J. Physiol. Renal Physiol.* 296, F1100–F1108. doi: 10.1152/ajprenal.90402.2008
- Lang, F., Bohmer, C., Palmada, M., Seebohm, G., Strutz-Seebohm, N., and Vallon, V. (2006). (Patho)physiological significance of the serum- and glucocorticoidinducible kinase isoforms. *Physiol. Rev.* 86, 1151–1178. doi: 10.1152/physrev.00050.2005
- Lewis, C. E., De Palma, M., and Naldini, L. (2007). Tie2-expressing monocytes and tumor angiogenesis: regulation by hypoxia and angiopoietin-2. *Cancer Res.* 67, 8429–8432. doi: 10.1158/0008-5472.CAN-07-1684
- Locati, M., Mantovani, A., and Sica, A. (2013). Macrophage activation and polarization as an adaptive component of innate immunity. *Adv. Immunol.* 120, 163–184. doi: 10.1016/B978-0-12-417028-5.00006-5
- Low-Marchelli, J. M., Ardi, V. C., Vizcarra, E. A., van Rooijen, N., Quigley, J. P., and Yang, J. (2013). Twist1 induces CCL2 and recruits macrophages to promote angiogenesis. *Cancer Res.* 73, 662–671. doi: 10.1158/0008-5472.CAN-12-0653
- Lumeng, C. N., Bodzin, J. L., and Saltiel, A. R. (2007a). Obesity induces a phenotypic switch in adipose tissue macrophage polarization. J. Clin. Invest. 117, 175–184. doi: 10.1172/JCI29881
- Lumeng, C. N., Deyoung, S. M., and Saltiel, A. R. (2007b). Macrophages block insulin action in adipocytes by altering expression of signaling and glucose transport proteins. *Am. J. Physiol. Endocrinol. Metab.* 292:E166–E174. doi: 10.1152/ajpendo.00284.2006
- Lutz, M. B., and Kurts, C. (2009). Induction of peripheral CD4+ T-cell tolerance and CD8+ T-cell cross-tolerance by dendritic cells. *Eur. J. Immunol.* 39, 2325–2330. doi: 10.1002/ejj.200939548
- Machnik, A., Dahlmann, A., Kopp, C., Goss, J., Wagner, H., van Rooijen, N., et al. (2010). Mononuclear phagocyte system depletion blocks interstitial tonicityresponsive enhancer binding protein/vascular endothelial growth factor C expression and induces salt-sensitive hypertension in rats. *Hypertension* 55, 755–761. doi: 10.1161/HYPERTENSIONAHA.109.143339
- Machnik, A., Neuhofer, W., Jantsch, J., Dahlmann, A., Tammela, T., Machura, K., et al. (2009). Macrophages regulate salt-dependent volume and blood pressure by a vascular endothelial growth factor-C-dependent buffering mechanism. *Nat. Med.* 15, 545–552. doi: 10.1038/nm.1960
- Mantovani, A., Biswas, S. K., Galdiero, M. R., Sica, A., and Locati, M. (2013). Macrophage plasticity and polarization in tissue repair and remodelling. *J. Pathol.* 229, 176–185. doi: 10.1002/path.4133
- Marchetti, V., Yanes, O., Aguilar, E., Wang, M., Friedlander, D., Moreno, S., et al. (2011). Differential macrophage polarization promotes tissue remodeling and repair in a model of ischemic retinopathy. *Sci. Rep.* 1:76. doi: 10.1038/srep 00076
- Martel, C., Li, W., Fulp, B., Platt, A. M., Gautier, E. L., Westerterp, M., et al. (2013). Lymphatic vasculature mediates macrophage reverse cholesterol transport in mice. J. Clin. Invest. 123, 1571–1579. doi: 10.1172/JCI63685
- Mazzieri, R., Pucci, F., Moi, D., Zonari, E., Ranghetti, A., Berti, A., et al. (2011). Targeting the ANG2/TIE2 axis inhibits tumor growth and metastasis by impairing angiogenesis and disabling rebounds of proangiogenic myeloid cells. *Cancer Cell* 19, 512–526. doi: 10.1016/j.ccr.2011.02.005
- Medzhitov, R. (2008). Origin and physiological roles of inflammation. *Nature* 454, 428–435. doi: 10.1038/nature07201
- Morancho, B., Minguillon, J., Molkentin, J. D., Lopez-Rodriguez, C., and Aramburu, J. (2008). Analysis of the transcriptional activity of endogenous NFAT5 in primary cells using transgenic NFAT-luciferase reporter mice. *BMC Mol. Biol.* 9:13. doi: 10.1186/1471-2199-9-13
- Mosser, D. M., and Edwards J. P. (2008). Exploring the full spectrum of macrophage activation. Nat. Rev. Immunol. 8, 958–969. doi: 10.1038/nri2448

- Muller, S., Quast, T., Schroder, A., Hucke, S., Klotz, L., Jantsch, J., et al. (2013). Saltdependent chemotaxis of macrophages. *PLoS ONE* 8:e73439. doi: 10.1371/journal.pone.0073439
- Murdoch, C., Muthana, M., Coffelt, S. B., and Lewis, C. E. (2008). The role of myeloid cells in the promotion of tumour angiogenesis. *Nat. Rev. Cancer* 8, 618–631. doi: 10.1038/nrc2444
- Murray, P. J., and Wynn, T. A. (2011). Protective and pathogenic functions of macrophage subsets. *Nat. Rev. Immunol.* 11, 723–737. doi: 10.1038/nri3073
- Nadkarni, V., Gabbay, K. H., Bohren, K. M., and Sheikh-Hamad, D. (1999). Osmotic response element enhancer activity. Regulation through p38 kinase and mitogen-activated extracellular signal-regulated kinase kinase. J. Biol. Chem. 274, 20185–20190.
- Nagy, L., Szanto, A., Szatmari, I., and Szeles, L. (2012). Nuclear hormone receptors enable macrophages and dendritic cells to sense their lipid environment and shape their immune response. *Physiol. Rev.* 92, 739–789. doi: 10.1152/physrev.00004.2011
- Naldini, A., Pucci, A., Bernini, C., and Carraro, F. (2003). Regulation of angiogenesis by Th1- and Th2-type cytokines. *Curr. Pharm. Des.* 9, 511–519. doi: 10.2174/1381612033391423
- Nucera, S., Biziato, D., and De Palma, M. (2011). The interplay between macrophages and angiogenesis in development, tissue injury and regeneration. *Int. J. Dev. Biol.* 55, 495–503. doi: 10.1387/ijdb.103227sn
- Odegaard, J. I., Ricardo-Gonzalez, R, Goforth, M. H., Morel, C. R., Subramanian, V., Mukundan, L., et al. (2007). Macrophage-specific PPARgamma controls alternative activation and improves insulin resistance. *Nature* 447, 1116–1120. doi: 10.1038/nature05894
- Odegaard JI., and Chawla, A. (2013). The immune system as a sensor of the metabolic state. *Immunity* 38, 644–654. doi: 10.1016/j.immuni.2013.04.001
- Owen, J. L., and Mohamadzadeh, M. (2013). Macrophages and chemokines as mediators of angiogenesis. *Front. Physiol.* 4:159. doi: 10.3389/fphys.2013. 00159
- Pal, D., Dasgupta, S., Kundu, R., Maitra, S., Das, G., Mukhopadhyay, S., et al. (2012). Fetuin-A acts as an endogenous ligand of TLR4 to promote lipidinduced insulin resistance. *Nat. Med.* 18, 1279–1285. doi: 10.1038/nm.2851
- Pollard, J. W. (2004). Tumour-educated macrophages promote tumour progression and metastasis. Nat. Rev. Cancer 4, 71–78. doi: 10.1038/nrc1256
- Pollard, J. W. (2009). Trophic macrophages in development and disease. Nat. Rev. Immunol. 9, 259–270. doi: 10.1038/nri2528
- Potente, M., Gerhardt, H., and Carmeliet, P. (2011). Basic and therapeutic aspects of angiogenesis. *Cell* 146, 873–887. doi: 10.1016/j.cell.2011.08.039
- Ricote, M., Huang, J., Fajas, L., Li, A., Welch, J., Najib, J., et al. (1998). Expression of the peroxisome proliferator-activated receptor gamma (PPARgamma) in human atherosclerosis and regulation in macrophages by colony stimulating factors and oxidized low density lipoprotein. *Proc. Natl. Acad. Sci. U.S.A.* 95, 7614–7619.
- Roth, I., Leroy, V., Kwon, H. M., Martin, P. Y., Feraille, E., and Hasler, U. (2010). Osmoprotective transcription factor NFAT5/TonEBP modulates nuclear factorkappaB activity. *Mol. Biol. Cell* 21, 3459–3474. doi: 10.1091/mbc.E10-02-0133
- Saharinen, P., and Alitalo, K. (2011). The yin, the yang, and the angiopoietin-1. J. Clin. Invest. 121, 2157–2159. doi: 10.1172/JCI58196
- Saharinen, P., Bry, M., and Alitalo, K. (2010). How do angiopoietins Tie in with vascular endothelial growth factors? *Curr. Opin. Hematol.* 17, 198–205. doi: 10.1097/MOH.0b013e3283386673
- Schulze-Osthoff, K., Risau, W., Vollmer, E., and Sorg, C. (1990). In situ detection of basic fibroblast growth factor by highly specific antibodies. *Am. J. Pathol.* 137, 85–92.
- Shapiro, L., and Dinarello, C. A. (1995). Osmotic regulation of cytokine synthesis in vitro. Proc. Natl. Acad. Sci. U.S.A. 92, 12230–12234.
- Shi, H., Kokoeva, M. V., Inouye, K., Tzameli, I., Yin, H., and Flier, J. S. (2006). TLR4 links innate immunity and fatty acid-induced insulin resistance. J. Clin. Invest. 116, 3015–3025. doi: 10.1172/JCI28898
- Sjostrom, M., Stenstrom, K., Eneling, K., Zwiller, J., Katz, A. I., Takemori, H., et al. (2007). SIK1 is part of a cell sodium-sensing network that regulates active sodium transport through a calcium-dependent process. *Proc. Natl. Acad. Sci.* U.S.A. 104, 16922–16927. doi: 10.1073/pnas.0706838104
- Solinas, G., Vilcu, C., Neels, J. G., Bandyopadhyay, G. K., Luo, J. L., Naugler, W., et al. (2007). JNK1 in hematopoietically derived cells contributes to dietinduced inflammation and insulin resistance without affecting obesity. *Cell Metab.* 6, 386–397. doi: 10.1016/j.cmet.2007.09.011

- Stabile, E., Burnett, M. S., Watkins, C., Kinnaird, T., Bachis, A., la Sala, A., et al. (2003). Impaired arteriogenic response to acute hindlimb ischemia in CD4knockout mice. *Circulation* 108, 205–210. doi: 10.1161/01.CIR.0000079225. 50817.71
- Stabile, E., Kinnaird, T., la Sala, A., Hanson, S. K., Watkins, C., Campia, U., et al. (2006). CD8+ T lymphocytes regulate the arteriogenic response to ischemia by infiltrating the site of collateral vessel development and recruiting CD4+ mononuclear cells through the expression of interleukin-16. *Circulation* 113, 118–124. doi: 10.1161/CIRCULATIONAHA.105.576702
- Starnes, T., Robertson, M., Sledge, G., Kelich, S., Nakshatri, H., Broxmeyer, H. E., et al. (2001). IL-17F, a novel cytokine selectively expressed in activated T-cells and monocytes, regulates angiogenesis and endothelial cell cytokine production. *Blood* 98, 818a–819a.
- Swirski, F. K., Nahrendorf, M., Etzrodt, M., Wildgruber, M., and Cortez-Retamozo, V., Panizzi, P, et al. (2009). Identification of splenic reservoir monocytes and their deployment to inflammatory sites. *Science* 325, 612–616. doi: 10.1126/science.1175202
- Szanto, A., Balint, B. L., Nagy, Z. S., Barta, E., Dezso, B., Pap, A., et al. (2010). STAT6 transcription factor is a facilitator of the nuclear receptor PPARgammaregulated gene expression in macrophages and dendritic cells. *Immunity* 33, 699–712. doi: 10.1016/j.immuni.2010.11.009
- Takeda, Y., Costa, S., Delamarre, E., Roncal, C., Leite de Oliveira, R., Squadrito, M. L., et al. (2011). Macrophage skewing by Phd2 haplodeficiency prevents ischaemia by inducing arteriogenesis. *Nature* 479, 122–126. doi: 10.1038/ nature10507
- Tauber, A. I. (2003). Metchnikoff and the phagocytosis theory. Nat. Rev. Mol. Cell Biol. 4, 897–901. doi: 10.1038/nrm1244
- Titze, J. (2014). Sodium balance is not just a renal affair. Curr. Opin. Nephrol. Hypertens. 23, 101–105. doi: 10.1097/01.mnh.0000441151.55320.c3
- Titze, J., and Machnik, A. (2010). Sodium sensing in the interstitium and relationship to hypertension. *Curr. Opin. Nephrol. Hypertens.* 19, 385–392. doi: 10.1097/MNH.0b013e32833aeb3b
- Vats, D., Mukundan, L., Odegaard, J. I., Zhang, L., Smith, K. L., Morel, C. R., et al. (2006). Oxidative metabolism and PGC-1beta attenuate macrophage-mediated inflammation. *Cell Metab.* 4, 13–24. doi: 10.1016/j.cmet.2006.05.011
- Weisberg, S. P., Hunter, D., Huber, R., Lemieux, J., Slaymaker, S., Vaddi, K., et al. (2006). CCR2 modulates inflammatory and metabolic effects of high-fat feeding. J. Clin. Invest. 116, 115–124. doi: 10.1172/JCI24335
- Weisberg, S. P., McCann, D., Desai, M., Rosenbaum, M., Leibel, R. L., and Ferrante, A. W. Jr. (2003). Obesity is associated with macrophage accumulation in adipose tissue. J. Clin. Invest. 112, 1796–808. doi: 10.1172/JCI19246
- Wiig, H., Schroder, A., Neuhofer, W., Jantsch, J., Kopp, C., Karlsen, T. V., et al. (2013). Immune cells control skin lymphatic electrolyte homeostasis and blood pressure. J. Clin. Invest. 123, 2803–2815. doi: 10.1172/JCI60113

- Wu, C., Yosef, N., Thalhamer, T., Zhu, C., Xiao, S., Kishi, Y., et al. (2013). Induction of pathogenic TH17 cells by inducible salt-sensing kinase SGK1. *Nature* 496, 513–517. doi: 10.1038/nature11984
- Xiong, M., Elson, G., Legarda, D., and Leibovich, S. J. (1998). Production of vascular endothelial growth factor by murine macrophages: regulation by hypoxia, lactate, and the inducible nitric oxide synthase pathway. *Am. J. Pathol.* 153, 587–598. doi: 10.1016/S0002-9440(10)65601-5
- Xu, H., Barnes, G. T., Yang, Q., Tan, G., Yang, D., Chou, C. J., et al. (2003). Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *J. Clin. Invest.* 112, 1821–1830. doi: 10.1172/ JCI19451
- Yang, S., Kim, J., Ryu, J. H., Oh, H., Chun, C. H., Kim, B. J., et al. (2010). Hypoxiainducible factor-2 alpha is a catabolic regulator of osteoarthritic cartilage destruction. *Nat. Med.* 16, 687–693. doi: 10.1038/Nm.2153
- Zajac, E., Schweighofer, B., Kupriyanova, T. A., Juncker-Jensen, A., Minder, P., Quigley, J. P., et al. (2013). Angiogenic capacity of M1- and M2-polarized macrophages is determined by the levels of TIMP-1 complexed with their secreted proMMP-9. *Blood* 122, 4054–4067. doi: 10.1182/blood-2013-05-501494
- Zhou, X., Ferraris, J. D., Dmitrieva, N. I., Liu, Y., and Burg, M. B. (2008). MKP-1 inhibits high NaCl-induced activation of p38 but does not inhibit the activation of TonEBP/OREBP: opposite roles of p38alpha and p38delta. *Proc. Natl. Acad. Sci. U.S.A.* 105, 5620–5625. doi: 10.1073/pnas. 0801453105
- Zhou, X., Izumi, Y., Burg, M. B., and Ferraris, J. D. (2011). Rac1/osmosensing scaffold for MEKK3 contributes via phospholipase C-gamma1 to activation of the osmoprotective transcription factor NFAT5. *Proc. Natl. Acad. Sci. U.S.A.* 108, 12155–12160. doi: 10.1073/pnas.1108107108

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