



Acute and Chronic Effects of Endurance Running on Inflammatory Markers: A Systematic Review

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In order to understand the effect of endurance running on inflammation, it is necessary to quantify the extent to which acute and chronic running affects inflammatory mediators. The aim of this study was to summarize the literature on the effects of endurance running on inflammation mediators. Electronic searches were conducted on PubMED and Science Direct with no limits of date and language of publication. Randomized controlled trials (RCTs) and non-randomized controlled trials (NRCTs) investigating the acute and chronic effects of running on inflammation markers in runners were reviewed by two researchers for eligibility. The modified Downs and Black checklist for the assessments of the methodological quality of studies was subsequently used. Fifty-one studies were finally included. There were no studies with elite athletes. Only two studies were chronic interventions. Results revealed that acute and chronic endurance running may affect anti- and pro-inflammatory markers but methodological differences between studies do not allow comparisons or generalization of the results. The information provided in this systematic review would help practitioners for better designing further studies while providing reference values for a better understanding of inflammatory responses after different running events. Further longitudinal studies are needed to identify the influence of training load parameters on inflammatory markers in runners of different levels and training background.

Keywords: running, inflammation, marathon, half-marathon, athletes, immunology

INTRODUCTION

Running is an important natural ability of our species that has contributed to our survival and body adaptations (Bramble and Lieberman, 2004). In the Paleolithic Era, survival was dependent on hunting and gathering, and therefore it has been suggested that the ancient physical activity pattern included mostly prolonged, low-intensity physical activities, including endurance running, interspersed with high-intensity bursts of activity (O'Keefe et al., 2010; Boullosa et al., 2013). Nowadays, endurance running is probably the most popular sport worlwide and it is practiced for recreational, health and competitive purposes (Chiampas and Goyal, 2015).

There is a close link between endurance running and the activity of the immune system. The importance of this relationship has led to important investigations over the last decades. Previously, Peters and Bateman (1983) identified an increased prevalence of upper respiratory tract infection (URTI) in 150 runners following a 56.0 km ultramarathon. Subsequently, specialized literature

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has suggested that even highly trained individuals, when subjected to frequent strenuous exercise, could develop a proinflammatory condition that favors the onset of a number of health problems, including damage to myocardial cells and connective tissues, overload of the atria and right ventricle, coronary artery disease (CAD), and coronary artery calcification among others (Peters and Bateman, 1983; Febbraio and Pedersen, 2005; Petersen and Pedersen, 2005; Zaldivar et al., 2006; Mohlenkamp et al., 2007; Hubble et al., 2009; Nieman, 2009; Meeusen et al., 2013; O'Keefe and Lavie, 2013; Taylor et al., 2014). However, little is known about whether the development of these chronic pathologies is the result of an excess of training volume, intensity, or both, associated with an insufficient recovery, which often promotes an increased susceptibility to infections and subsequent reduction in performance (Smith, 2004; Zaldivar et al., 2006; Hubble et al., 2009). Furthermore, systematic and non-systematic inflammation after running might be related with functional overreaching (Steinacker et al., 2004). In contrast, it has been suggested that periodised training with adequate recovery may be associated with positive adaptations including an adequate balance between pro-inflammatory and anti-inflammatory responses (Febbraio and Pedersen, 2005; Petersen and Pedersen, 2005; Zaldivar et al., 2006).

A growing body of evidence highlights the importance of studying inflammation promoted by endurance running as a factor which is linked to the physiopathology of a number of cardiovascular diseases (Mohlenkamp et al., 2007). It has also been suggested a link between myocardial damage and small thrombotic or even atherosclerotic emboli following a marathon, or after a quick session of exercise, accompanied by a transient monocytosis (about 2 h) (Walsh et al., 2011). The tissue factor is known as the key initiator of coagulation, and is highly dependent on vascular injury and mediators of inflammation such as tumor necrosis factor alpha (TNF- α), which has been reported to increase during and predominantly after marathon running (O'Brien, 2012; Gill et al., 2015b).

Contrary to other endurance sports, eccentric muscle contractions play a key role in running exercises, leading up to the occurrence of different levels of damage in muscle, connective and bone tissues (Suzuki et al., 2003; Jarvinen et al., 2013). The repair of these tissues involves the presence of inflammatory cells into the damaged site, which stimulates the release of pro-inflammatory cytokines such as TNF- α and interleukin-1 beta (IL-1 β), thus triggering inflammation (Nieman et al., 1989, 1990). However, little is known about the impact of this chronic cycle tissue damage and repair in runners.

On the other hand, it is also important to emphasize that signaling promoted by repeated muscle contractions as in running, stimulates the production of anti-inflammatory mediators by myocytes, especially interleukin-6 (IL-6), which acts as an inhibitor of pro-inflammatory cytokines such as TNF- α by stimulating the production of its soluble receptor antagonists (Pedersen, 2013). In addition, IL-6 also stimulates the production of interleukin-10 (IL-10) and interleukin-1 receptor antagonist (IL-1ra), generating an anti-inflammatory environment which may counterbalance the pro-inflammatory responses associated to repetitive eccentric actions (Pedersen and Febbraio, 2012).

Despite the growing body of evidence regarding the effects of endurance running on inflammation, the link between transient acute responses and chronic adaptations needs to be addressed (Gleeson, 2007). This information would be important to shed light on the possible role of the inflammatory *milieu* in the pathophysiology of a number of diseases, especially the cardiovascular ones. Thus, the aim of this systematic review was to investigate the effects of different doses (i.e., training and competitive loads) of endurance running on the acute and chronic inflammatory responses, and the immune effects of this practice on runners of different levels and training backgrounds.

METHODS

Search Strategy

A systematic review was conducted and the recommendations from the Preferred Reporting Itens for Systematic Review and Meta-Analyses (PRISMA) were considered (Liberati et al., 2009). The search strategies were reported to ensure the integrity of the results and allow the updating using the same methods to bring emerging evidence into the review. The Boolean and proximity operators were used and the search strategy was correctly adapted for each database used (**Table 1**) (Sampson et al., 2008, 2009). Studies were identified by searching the following electronic databases: PubMED/MEDLINE (via National Library of Medicine) (2000–2017) and Science Direct (Elsevier) (2000–2017). The last search was conducted in February 2017.

Once the abstracts were reviewed, the complete versions of the papers that met the criteria were obtained. In addition, the reference lists of the papers that fulfilled the inclusion criteria were analyzed for identification of additional studies. The exclusion of studies with irrelevant content and duplicates was carried out after the title, abstract and full-text were read.

Definition of Terms

An "athlete" was defined according to the Medical Subject Headings (MeSH) and was considered to be an individual who has developed skills, physical fitness and strength, or who has participated in sports running (MeSH, 2015). We have considered the definition proposed by Stirling and Kerr (2006) that defines a "recreational athlete" as being an individual who plays on a sports team at an amateur level, works out 1– 4 times a week, does not train and compete nationally or internationally, and does not train for the same activity for more than 8 h per week. Novice runners were those individuals who had not been running on a regular basis in the previous 12 months (10 km total in all training sessions in the previous 12 months), and recreational runners were considered as individuals running a mean of 24.94 km/week (Videbaek et al., 2015).

The following thesaurus terms registered in the database from MeSH were also used: "running," "aerobic exercise," "inflammation," and "cytokines." These terms were associated with the free terms "recreational runners," "novice runners," "marathon runners," and "ultramarathon."

TABLE 1 | Search strategies.

Database	Search strategy	Hits	No. (%) of trials finally selected
PubMED/MEDLINE—via national library of medicine	1. Inflammation AND aerobic exercise AND runners;	128	50
	2. Cytokines AND runn* AND (marathon runners or novice runners);	64	
	· · · · · · · · ·	Total: 192	
Science Direct (Elsevier)	1. "marathon runners" OR "novice runners" AND "cytokines";	94	4
		Total: 94	
Other sources (reference lists of the papers that		Total: 19	6
fulfilled the inclusion criteria were analyzed for			
the identification of additional studies)			

*Truncation or wildcard.

Inclusion and Exclusion Criteria

The inclusion criteria were as follows: randomized controlled trials (RCTs) and non-randomized controlled trials (NRCTs); studies investigating the acute and chronic effects of running on markers of inflammation in marathon runners, recreational runners and novice runners; the terms runners, marathon runners, recreational runners and novice runners should be cited in the paper; only healthy participants; only full-text article citations with no restriction on languages; with individuals aged over 19 as the World Health Organisation (WHO) defines adolescence as the period in human growth and development that occurs between childhood and adulthood, from ages 10-19 (WHO, 2015)¹. Meeting abstracts, unpublished data, observational studies, review articles, studies using walking and jogging as independent variables, and studies on the effects of any kind of supplements on running, diet restrictions, use of devices (e.g., equipment, compression garments), comparisons between running and other sports, and effects of environmental conditions (ex. dry and hot) were excluded.

Outcome Measures

The outcome measures assessed for acute and chronic effects of marathon and recreational running were interleukin (IL): IL-6, IL-10, IL-8, IL-1ra, IL-1 β , IL-2 and IL-12, TNF- α , C reactive-protein (CRP), interferon-gamma (IFN- γ), soluble receptors, and transformation growth factor-beta (TGF- β). These mediators were chosen after an initial analysis and review of the literature. They were identified as the main outcomes in studies published with marathon runners, recreational runners and novice runners (Nieman et al., 2005; Santos et al., 2007; Scott et al., 2011; Abbasi et al., 2013; Jee et al., 2013; Shin and Lee, 2013).

Quality Assessment

The quality and assessment of all eligible articles was evaluated using a modified version of the Downs and Black checklist (Downs and Black, 1998). Disagreements between authors were discussed and subsequently solved. This modified version consists of 27 objective questions (Downs and Black, 1998).

RESULTS

Research Strategy

Results of the research strategy are presented in **Figure 1**. Initially, 60 studies were selected, with 51 studies being finally included according to the inclusion/exclusion criteria. Nine studies were excluded as follows: one study was excluded due to the use of heat stress, three because the subjects were adolescents, one following the reading of the full paper, two because of comparison with other sporting activities, and one because of medication use; in addition, one paper was not available in full-text version (Saravia et al., 2010). A total of 49 studies verified the acute effect of running on inflammation and two studies focused on the chronic effects.

Methodological Quality Assessment

Quality assessment of the studies according to the modified Downs and Black scale is summarized in **Table 2**. One important finding was that characteristics of the patients' included were not cleary described in 32 studies. Important adverse events and description of patients' characteristics lost to follow-up was not reported in 34 and 32 studies, respectively. None of the studies were randomized controlled trials and power was provided in 4 studies (**Table 2**).

Characteristics of the Studies and Summary of Outcome Measures

An overview of the studies' characteristics is provided in **Table 3** with sample size, age, sex, and exercise protocols. A summary of outcome measures in selected studies is presented on **Table 4**.

The 51 studies included resulted in a total of 1,421 subjects, of whom 163 were female and 1,234 males; 24 subjects were not identified by sex in one study (Neidhart et al., 2000). All trials provided age ranges for the subjects and the mean age was 39.16 years.

The common protocols adopted included marathon in 17 studies, ultra-marathon in 22, half-marathon in three studies, different distance protocols (42.195, 21.1, 12, 10 km and treadmill) in seven studies, and chronic training only in two studies (see **Table 3**).

¹World Health Organization; Available online at: http://www.who.int/ maternal_child_adolescent/topics/adolescence/dev/en/ (Acessed November 1, 2015).



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TABLE 2 Methodological quality assessment scores of the included studies.			
TABLE	Study		

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Suzuki et al., 2000

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Auersperger et al., 2012

Reihmane et al., 2013

Millet et al., 2011

Karstoft et al., 2013 Wilhelm et al., 2014 Bernecker et al., 2013

Chimenti et al., 2009

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Mrakic-Sposta et al., 2015		-	0	-	-	-	0	-	0	*0	*0	-	*0	*0	0	0		*0	-		0	0	0	0	-	-
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Nieman et al., 2016		-	0	-	-	-	0	0		*0	*0	-	*0	*0	0		-	*0	-		0	0	0	0	0	0
Arakawa et al., 2016	-		0	-	-	-	0	0	0	*0	*0	-	*0	*0	0			*0	-		0	0	0	0	0	0
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Krzeminski et al., 2016		-	0	-	-	-	0	0	0	*0	*0	-	*0	*0	0		-	*0	-	0	0	0	0	0	0	0
Nielsen et al., 2016	÷	. 		0	-		0	0	-	*0	*0	-	*0	*0	0	-	-	*0	-		0	0	0	0	0	0
1 = yes and 0 = no for questions 1, 2, 3, 4, 6, 7, 8, 9, and 10. For question 5: 2 = yes, $1 = partially, and 0 = no$. For other questions 1 = yes, $0 = no$, and $0^* = unable to determine.$	is 1, 2, . tially, ai = no, ar	3, 4, (7d 0 = 7d 0* :	3, 7, 8 = no. = unal	, 9, ar ble to	id 10. deterr	nine.																				

TABLE 3 | Characteristics of the studies.

Citation	Age (years)	Exercise group (<i>N</i>) sex	Total subjects (N) male/female	Control group (N) sex	Level of runners	Method (distance)		vention ects)
							Acute	Chronic
Grabs et al., 2015	45.0 ± 8.0	20	20 (්)	-	ATL	Marathon		
Fallon et al., 2001	47.0 ± 7.0	8	8 (7♂-1♀)	-	ATL	Ultra-Marathon: 6 days		
Kim et al., 2015	50.8 ± 8.2	40	40 (්)	-	LNR	Marathon		
Gill et al., 2015b	41.0 ± 8.0 49.0 ± 4.0	19 (13ơ-6ọ)	31 (18♂-13♀)	12 (5♂-7♀)	LNR	Ultra-marathon: 230 km Five stage (37, 48, 38, 69, 39 km)	•	
Mattusch et al., 2000	EG: 25–40 CG: 31–52	14	25 (♂)	11	REC	Training		
Neidhart et al., 2000	EG: 25–34 CG: 24–35	8	24 sex (NR)	16	REC	Marathon	•	
Vaisberg et al., 2013	41.4 ± 9.4	Asymptomatic: 15 Symptomatic: 7	22 (♂)		ATL	Marathon	•	
Niess et al., 2000	EG: 32.3 ± 3.3 CG: 25.0 ± 2.2	½ Marathon Group: 10	18 (♂)	8	NOV	Half-marathon and treadmill		
Kasprowicz et al., 2013	44.5 ± 13.5	6	6 (♂)	-	ATL	Ultra-marathon: 100 km	•	
Saugy et al., 2013	45.4 ± 10.3 CG: 29.3 ± 8.1	25	33 (ơ²)	8	ATL	Ultra-marathon: 330 km (Mountain)	•	
Jee et al., 2013	EG: 49.75 ± 5.65 CG: 46.75 ± 5.44	8	16 (♂¹)	8	ATL	Ultra-marathon: 308 km	•	
Karstoft et al., 2013	44 ± 2	8	7 (♂) 1 (♀)	-	ATL	Marathon		
Wilhelm et al., 2014	34.9 ± 4.2	11	11 (♂)	-	ATL	Marathon (Mountain)		
Reihmane et al., 2013	Half-Marathon: 26 ± 5 Marathon: 27 ± 5	22 (♂) 18 (♂)	40 (ơ [*])	-	REC	Half-marathon Marathon	•	
Millet et al., 2011	40.2	22	22 (ơ [*])	-	ATL	Ultra-marathon 166 km	•	
Auersperger et al., 2012	Interval Group: 32.9 ± 5.7 Continuous Group: 31.6 ± 4.8	10 8	18 (ç)		REC	Chronic Training		•
Bernecker et al., 2013	43 (33–53)	12	12 (ീ		REC	Marathon		
Chimenti et al., 2009	40.3 ± 3.8	9 (♂)	9 (ơ [*])		REC	Half-marathon, fall (21 km), winter (12 km) and summer (10 km)	•	
Papassotiriou et al., 2008	42.8 ± 1.4	15	15 (♂)		ATL	Ultra-marathon 246 km	•	
Kim et al., 2007	45.7 ± 5.1	54	54 (♂)		ATL	Ultra-marathon 200 km	•	
Peters et al., 2004	Fast group: 35.4 ± 1.84 Slow group: 41.4 ± 2.77	9 10	30 (♂¹)	-	ATL	Ultra-marathon 90 km	•	
Suzuki et al., 2003	31.7 ± 5.0	10	10 (♂)	-	ATL	Marathon		
Bachi et al., 2015	Sedentary group: 35.5 ± 7 Marathon runners: 35.7 ± 9	20	40 (♂)	20	REC	Marathon	•	
Kłapcinska et al., 2013	45.4 ± 9.2	7	7 (♂)		ATL	Ultra-marathon 48 h		
Rehm et al., 2013	40.95 ± 9.38	19	19 (14♂-5♀)		REC	Marathon		
Fehrenbach et al., 2000	32.3 ± 9.3	12	24 (ి)	12	REC	Half-marathon		
Schobersberger et al., 2000	36.3 (22–50)	13	13 (♂)	-	ATL	Ultra-marathon 67 km	-	

Citation	Age (years)	Exercise group (<i>N</i>) sex	Total subjects (N) male/female	Control group (N) sex	Level of runners	Method (distance)		vention ects)
							Acute	Chronic
Suzuki et al., 2000	21–39	16	16 (♂)	_	ATL	Marathon		
Vaisberg et al., 2012	Sedentary Group: 37.5 ± 4 Athletes Group: 38 ± 7	14	42 (♂ [*])	28	REC	Marathon	•	
Tomaszewski et al., 2003	Lean, BMI <25 kg/m2 Marathon runners: 43.1 \pm 8.4 Control: 42.5 \pm 10.4 Non-lean, BMI > 25 kg/m2 Marathon runners: 45.6 \pm 12.3 Control: 43.1 \pm 7.5	55 12	110 (ơ')	30 13	ATL	Ultra-marathon	•	
Bonsignore et al., 2002	41.3 ± 13.4	Half-marathon: 8 Marathon: 8	25 (ở)	9	ATL	Half-marathon and Marathon	•	
Nickel et al., 2012	LE: 40 \pm 7; LNE 40 \pm 6; ONE 40 \pm 6	LE: 16; LNE: 16	47 (ơ [*])	15	ATL and REC	Marathon		
Waśkiewicz et al., 2012	43.0 ± 10.8	14	14 (♂)	-	ATL	Ultra-marathon 24 h		
Chimenti et al., 2010	NR	15	15 (♂)	-	ATL	Half-marathon		
Ng et al., 2008	25 (21–32)	30	30 (්)	-	NRL	Half-marathon		
Siegel et al., 2007	49 ± 10	33	33 (ి)	-	NRL	Marathon		
Shin and Lee, 2013	52.8 ± 5.0	18	18 (ơ')	-	ATL	Ultra-marathon 308 km	•	
Jee and Jin, 2012	49.5 (47–54)	24	24 (ి)	-	ATL	Ultra-marathon 308 km		
Santos et al., 2013	Athletes 35.2 ± 3.6 Non-athletes 31.6 ± 2.3	Athletes: 15	27 (්)	Non- athletes: 12	ATL	Marathon	•	
Hewing et al., 2015	50.3 (22–72)	167	167 (78♂) and (89♀)	-	ATL	Marathon	•	
Nieman et al., 2003	46.9 (33–65)	31	31 (22♂) and (9♀)	-	ATL	Ultra-marathon 160 km	•	
Uchakin et al., 2003	WR: 37.8 ± 3.9 CT: 40.3 ± 7.7	WR: 8	15	CT: 7	ATL	Marathon	•	
Mrakic-Sposta et al., 2015	45.04 ± 8.75	46	46 (♂ [°])	-	ATL	Mountain Ultra-Marathon 330 km	•	
Stuempfle et al., 2016	With nausea: 44.3 ± 10.5 Without nausea: 41.8 ± 9.1	20	(15♂-05♀)	-	ATL	Ultra-Marathon 161-km	•	
Nieman et al., 2016	22–45	20	(10♂-10♀)	_	ATL	1.5 h on treadmills at ~70% VO ₂ max followed by 30 min of downhill running	•	
Arakawa et al., 2016	52.1 ± 12.1	25	25 (♂)	-	ATL	Ultra-Marathon		
Mohamed et al., 2016	SS: 23.9 \pm 4.20 LDR: 22.70 \pm 3.70 MDR: 21 \pm 1.80	SS (n = 8) LDR (n = 9) MDR (n = 8)	24 (♂)	-	ATL	Incremental Event (VAMEVAL test) Supra-Maximal Exhausting Race (Limited-Time Test)	•	
Cairns and Hew-Butler, 2015	43.7 ± 9.8	Normonatremic: 5 Hyponatremic: 10	15 (12♂ 3♀)	-	ATL	100 km (103.7 km) and 100 miles (173.7 km)	•	
Gill et al., 2015a	40 ± 7	17	(14♂-03♀)	17 (04♂-08♀)	ATL	Ultra-Marathon 24-H	•	

Citation	Age (years)	Exercise group (N) sex	Total subjects (N) male/female	Control group (N) sex	Level of runners	Method (distance)		ention ects)
							Acute	Chronic
Krzeminski et al., 2016	30 ± 1.0	09	09 (්)	-	ATL	Ultra-Marathon 100 km	•	
Nielsen et al., 2016	40 (29–56)	Half-Marathon: (09ଟ'-09ହ) Marathon: (14ଟ')	32	-	REC	Half- Marathon/Marathon		

♂, male; ♀, female; EG, exercise group; CG, control group; PBMCs, peripheral blood mononuclear cells; LE, lean elite group; LNE, non-elite group; ONE, obese non-elite group; BMI, body mass index; WR, White Rock marathon; CT, CowTown marathon; SS, sedentary subjects; LDR, long-distance runners; MDR, Middle-distance runners; ALT, Athlete; REC, Recreational; NOV, Novice; Not reportable level (No reported details by authors), NRL; LNR, Level not reported.

DISCUSSION

The aim of this systematic review was to analyze studies that verified the effects of different endurance running exercises on acute and chronic inflammatory responses in runners of different training background. The present systematic review allows an initial understanding of this issue. It seems that acute and chronic endurance running may affect anti- and proinflammatory markers. However, important differences between studies in terms of methods as well as in runners' charactersitics do not allow appropriate comparison or generalization of results.

Inflammatory Markers

The timing in data collection could be considered an important limiting factor for adequate comparisons, given the heterogeneity observed across the reported acute (i.e., 5–20 min) (Fallon et al., 2001; Kim et al., 2007; Vaisberg et al., 2013; Grabs et al., 2015) and delayed responses (24 h to 8 days) (Uchakin et al., 2003; Siegel et al., 2007; Hewing et al., 2015). This is not a simple issue given the different kinetics and biological availability of the molecules considered as IL-6 and CRP (Kasprowicz et al., 2013; Reihmane et al., 2013). Of note, some inflammatory markers (e.g., monocyte chemoattractant protein-1, granulocyte colonystimulating factor) (Suzuki et al., 2003) have not been included in the current review. Further studies should elaborate appropriate study designs that consider both the appropriateness of the inflammatory markers selected as well as their kinetics.

Runners' and Training Load Characteristics

Another important confounding factor is the different experience of runners. Thus, 26 studies (4 studies with male and female participants, and 15 studies with only male participants) reported a range of 4–17.5 years of experience with competitions (e.g., marathon, and ultra-marathon experience) (Schobersberger et al., 2000; Fallon et al., 2001; Nieman et al., 2003, 2016; Suzuki et al., 2003; Tomaszewski et al., 2003; Kim et al., 2007, 2015; Millet et al., 2011; Jee and Jin, 2012; Vaisberg et al., 2012, 2013; Waśkiewicz et al., 2012; Bernecker et al., 2013; Jee et al., 2013; Karstoft et al., 2013; Kasprowicz et al., 2013; Kłapcinska et al., 2013; Saugy et al., 2013; Shin and Lee, 2013; Wilhelm et al., 2014; Grabs et al., 2015; Hewing et al., 2015; Mrakic-Sposta et al., 2015; Krzeminski et al., 2016; Mohamed et al., 2016). However, 25 studies did not report this information (Fehrenbach et al., 2000; Mattusch et al., 2000; Neidhart et al., 2000; Niess et al., 2000; Suzuki et al., 2000; Bonsignore et al., 2002; Uchakin et al., 2003; Peters et al., 2004; Siegel et al., 2007; Ng et al., 2008; Papassotiriou et al., 2008; Chimenti et al., 2009, 2010; Auersperger et al., 2012; Nickel et al., 2012; Rehm et al., 2013; Santos et al., 2013; Bachi et al., 2015; Gill et al., 2015b; Arakawa et al., 2016; Nielsen et al., 2016; Stuempfle et al., 2016; Vernillo et al., 2017). This is not a trivial issue given that training experience of runners could have a potentially additive effect to the influence of runners' age on inflammation that warrants further research. Besides, only 34 articles especified the training preparation (km/week that ranged between 21.4 and 161 and time that ranged between 3.9 and 10 h/week) of runners before races (Fehrenbach et al., 2000; Mattusch et al., 2000; Niess et al., 2000; Suzuki et al., 2000, 2003; Fallon et al., 2001; Bonsignore et al., 2002; Nieman et al., 2003, 2016; Tomaszewski et al., 2003; Uchakin et al., 2003; Peters et al., 2004; Chimenti et al., 2009; Millet et al., 2011; Auersperger et al., 2012; Jee and Jin, 2012; Nickel et al., 2012; Vaisberg et al., 2012, 2013; Waśkiewicz et al., 2012; Bernecker et al., 2013; Karstoft et al., 2013; Kłapcinska et al., 2013; Rehm et al., 2013; Reihmane et al., 2013; Santos et al., 2013; Shin and Lee, 2013; Wilhelm et al., 2014; Bachi et al., 2015; Grabs et al., 2015; Hewing et al., 2015; Mrakic-Sposta et al., 2015; Arakawa et al., 2016; Krzeminski et al., 2016). Furthermore, only two studies cited the control of intensity during training (Mattusch et al., 2000; Auersperger et al., 2012). This aspect would be important for a better understanding of the relationship between running and inflammation from a doseresponse perspective. For instance, higher values for IL-6 after a limited-time test were observed in sedentary individuals when compared to long- and middle-distance runners, but with no differences between groups for TNF- α (Mohamed et al., 2016). Given the growing popularity of races that last various days, further studies are warranted to elucidate if chronic exposure to high volumes of endurance running are detrimental for health. From an evolutionary perspective, this is an interesting topic given that daily running volumes of modern huntergatherers are far below (e.g., ~10-15 km) (O'Keefe et al., 2010;

TABLE 4 | Summary of outcome measures.

Citation	Outcomes	Exercise		Control		
		Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	р value ^t
Grabs et al., 2015	IL-6 (mg/L)	2.0 (0.0)	33.1 (24.1–37.00) ^a			
	hs-CRP (mg/L)	0.83 (0.57-1.18)	9.13 (6.48–13.63) ^a			
Fallon et al., 2001	CRP (mg/L)	0.19 (0.14)	1.84 (0.88) ^a			
Kim et al., 2015	hs-CRP (mg/L)	0.06 (0.07)	0.10 (0.09) ^a			
Gill et al., 2015b	CRP (mg/L)	0.00 (0.07)	0.10 (0.00)			
Gill 01 Gil, 20100	Stage 1	1.1 (1.7)	1.6 (2.4)	1.4 (0.7)		NS
	Stage 2	7.4 (5.3) [†]	8.8 (5.4)	-		NO
	Stage 3	10.0 (5.7) [†]	9.6 (5.9)	1.3 (0.8) ^b		< 0.05
	Stage 4	9.2 (5.9) [†]	10.0 (6.7) ^a	_		
	Stage 5	8.8 (5.6)†	11.0 (6.4) ^a	1.3 (0.8) ^b		< 0.05
	IL-6 (pg/L)					
	Stage 1	8.2 (4.5)	27.9 (23.4) ^a	7.5 (2.5)		NS
	Stage 2	20.8 (18.5) [†]	20.7 (14.8)			0.05
	Stage 3	20.7 (16.8) [†] 19.2 (14.1) [†]	25.3 (24.3) ^a 21.7 (12.6) ^a	5.5 (7.1) ^b		<0.05
	Stage 4 Stage 5	18.2 (14.1) ⁺ 18.2 (11.6) [†]	23.4 (13.1) ^a	6.5 (5.7) ^b		<0.05
		10.2 (11.0)	20.1(10.1)	0.0 (0.1)		<0.00
	IL-1β (pg/L) Stage 1	0.6 (0.3)	1.0 (0.3) ^a	0.7 (0.2)		NS
	Stage 2	1.1 (0.4) [†]	1.1 (0.4)	-		110
	Stage 3	1.2 (0.4) [†]	1.2 (0.4)	1.2 (0.2)		NS
	Stage 4	1.1 (0.3)†	1.4 (0.4) ^a	-		
	Stage 5	1.2 (0.4)†	1.4 (0.4) ^a	1.3 (0.5)		NS
	TNF-α (pg/L)					
	Stage 1	3.1 (2.9)	6.3 (5.0) ^a	1.3 (0.4)		NS
	Stage 2	6.1 (4.5) [†]	6.6 (3.7)	- -		
	Stage 3	6.9 (4.4) [†] 6.5 (4.2) [†]	6.1 (3.8) ^a 8.1 (4.3) ^a	1.8 (0.7) ^b		<0.05
	Stage 4	7.1 (3.8) [†]	8.3 (5.0)	_ 2.3 (0.7) ^b		<0.05
	Stage 5	111 (0.0)	0.0 (0.0)	2.0 (0.1)		<0.00
	IFN-γ (IU/ml) Stage 1	9.3 (5.5)	12.9 (6.0) ^a	16.8 (5.5)		NS
	Stage 2	15.2 (6.8)	16.9 (5.7)	-		
	Stage 3	16.7 (6.7) [†]	15.2 (5.2)	14.3 (2.0)		NS
	Stage 4	16.2 (7.2)	19.9 (8.3) ^a	-		
	Stage 5	18.8 (10.0) [†]	22.7 (9.9) ^a	16.8 (5.1)		NS
	IL-10 (pg/ml)					
	Stage 1	0.7 (0.6)	7.9 (10.1) ^a	0.6 (0.1)		NS
	Stage 2	7.0 (10.8) [†]	7.9 (9.1)	- 1 4 (0 0)b		0.05
	Stage 3	9.0 (10.2) [†] 9.0 (12.2) [†]	8.0 (9.4) 9.3 (10.1)	1.4 (0.3) ^b		<0.05
	Stage 4 Stage 5	8.2 (11.2) [†]	10.9 (15.0) ^a	1.4 (0.7) ^b		<0.05
		- ()		(-)		
	IL-1ra (pg/ml) Stage 1	22.9 (8.0)	70.3 (28.1) ^a	23.4 (7.3)		NS
	Stage 2	39.8 (12.4) [†]	61.0 (39.8) ^a	_		
	Stage 3	45.5 (20.6) [†]	53.9 (19.0) ^a	36.4 (9.2) ^b		< 0.05
	Stage 4	37.9 (14.7)†	56.3 (30.4) ^a	-		
	Stage 5	47.1 (22.4)†	63.2 (28.1) ^a	33.1 (9.3) ^b		<0.05
Mattusch et al., 2000	CRP (mg/L)	1.19 (1.63)	0.82 (0.94)	0.77 (2.18)	1.55 (9.17)	NS
Neidhart et al., 2000	CRP (mg/L)					
	Before run (T0)	NR		NR	NR	NS
					NR	NS
	()				NR NR	NS NS
	. ,				NR	< 0.05
		NR ^{b↑,c↑}			NR	< 0.05
	After 31 km (T1) After 42 km (T2) 2 h after run (T3) 24 h after run (T4) 48 h after run (T5)	NR NR NR ^{b∱,c↑} NR ^{b∱,c↑}		NR NR NR NR	ח ח ח	NR NR NR

Citation	Outcomes	Exercise		Control		
		Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	<i>p</i> value ^b
	IL-1β (ng/ml)					
	Before run (T0)	NR		NR	NR	NS
	After 31 km (T1)	NR		NR	NR	NS
	After 42 km (T2)	NR		NR	NR	NS
	2 h after run (T3)	NR		NR	NR	NS
	24 h after run (T4)	NR		NR	NR	NS
	48 h after run (T5)	NR		NR	NR	NS
	IL-1ra (ng/ml)					
	Before run (T0)	95		NR	NR	NS
	After 31 km (T1)	260 ^{b↑,c↑}		NR	NR	<0.05
	After 42 km (T2)	485 ^{b↑,c↑}		NR	NR	<0.05
	2 h after run (T3)	1195 ^{b↑,c↑}		NR	NR	<0.05
	24 h after run (T4)	NR		NR	NR	NS
	48 h after run (T5))	NR		NR	NR	NS
	IL-6 (ng/ml)	bl				
	Before run (T0)	1.8 ^{b↓}		NR	NR	<0.05
	After 31 km (T1)	8.7 ^{c↑}		NR	NR	NS
	After 42 km (T2)	9.8 ^{c↑}		NR	NR	NS
	2 h after run (T3)	5.3 ^{c↑}		NR	NR	NS
	24 h after run (T4)	2.2		NR	NR	NS
	48 h after run (T5)	2.1		NR	NR	NS
	TNF-α (pg/ml)					
	Before run (T0)	9.7		NR	NR	NS
	After 31 km (T1)	16.6 ^{c↑}		NR	NR	NS
	After 42 km (T2)	14.3 ^{c↑}		NR	NR	NS
	2 h after run (T3)	15.1 ^{¢↑}		NR	NR	NS
	24 h after run (T4)	13.1 ^{¢↑}		NR	NR	NS
	48 h after run (T5)	10.2 ^{c↑}		NR	NR	NS
	sIL-6R (pg/ml)					
	Before run (T0)	NR ^{b↑}		NR	NR	<0.05
	After 31 km (T1)	NR		NR	NR	NS
	After 42 km (T2)	NR		NR	NR	NS
	2 h after run (T3)	NR		NR	NR	NS
	24 h after run (T4)	NR		NR	NR	NS
	48 h after run (T5)	NR		NR	NR	NS
	sTNFRII (pg/ml)					
	Before run (T0)	3180		NR	NR	NS
	After 31 km (T1)	NR		NR	NR	NS
	After 42 km (T2)	NR		NR	NR	NS
	2 h after run (T3)	NR		NR	NR	NS
	. ,	NR		NR	NR	NS
	24 h after run (T4) 48 h after run (T5)	NR		NR	NR	NS
Veisbaue et al. 0010						NO
Vaisberg et al., 2013	II-6 nasal cell extract	Symptomatic		Asymptomatic		NS
	(pg/mg)					NS
	Baseline	0.07 (0.11)		0.22 (0.47)		
	Immediately	0.33 (0.17) ^{d↑}		0.49 (0.89) ^{d↑}		NS
	72 h	2.49 (2.35) ^d ↑		0.94 (1.1) ^d ↑		
	II-6 serum (pg/ml)					
	Baseline	8.2 (20.8)		14.9 (32.3)		NS
	Immediately	40.9 (30.9) ^d ↑		39.2 (26.5) ^{d↑}		NS
	72 h	10.6 (20.4)		16.9 (34.3)		NS
	IL-10 nasal cell extract					
	(pg/mg)					
	Baseline	0.22 (0.30)		0.28 (0.69)		NS
	Immediately	0.15 (0.15)		1.0 (2.68)		NS
	72 h	5.33 (8.00) ^{d,e↑}		10.26 (13.31) ^{b,d,e↑}		NS

Citation	Outcomes	Exercise		Control		
		Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	<i>p</i> value ^b
	IL-10 serum (pg/ml) Baseline Immediately 72 h	0.76 (0.1) ^b 57.2 (32.7) ^{d↑} 1.1 (0.9)		17.4 (34.0) 30.7 (25.3) ^{d↑} 4.5 (11.4)		<0.05 NS NS
Niess et al., 2000	IL-8 (pg/ml) Baseline 0 h 3 h 24 h 48 h TNF-α**	NR NR∱ ^{d,b} NR NR NR		NR NR NR NR		_ <0.05 _ _ _
Kasprowicz et al., 2013	CRP (ng/ml) Baseline 25 km 50 km 75 km Post 14 h after IL-6 (pg/ml) Baseline 25 km 50 km 75 km Post	NR NR NR NR↑ ^{d,f} NR↑ ^{d,f} NR↑ ^d ,f NR↑ ^{d,f} NR↑ ^{d,f}				
	14 h after	NR↓ ^{g,h,i}				
Saugy et al., 2013	CRP (mg/dl)	0.31 (0.32)	13.11 (7.51) ^a	1.05 (1.04)	0.65 (0.61) ^b	<0.05
Jee et al., 2013	CRP (mg/dl) Baseline 100 km 200 km 308 km	0.31 (0.21) 3.97 (4.58) 25.37 (18.24) ^{d,j} NR		0.39 (0.61) 4.27 (5.75) 25.09 (14.54) ^{d,j} 22.48 (12.90) ^{d,j}		NS NS NS NS
Karstoft et al., 2013	CRP (mg/dl)	1.0 (0.0)	6.0 (1.1) ^a	_	-	_
Wilhelm et al., 2014	TNF-α (pg/ml) Baseline Post race Follow-up 1 Follow-up 2 IL-6 (pg/ml) Baseline Post race Follow-up 1 Follow-up 2 hsCRP (mg/dl) Baseline Post race Follow-up 1 Follow-up 1 Follow-up 2	NR NR↑ ^d NR NR NR↑ ^d NR NR NR NR NR NR↑ ^d NR				
Reihmane et al., 2013	IL-6			Marathon runners		
	(pg/ml)-Half-marathon Pre-race 15 min post-race 28 h post-race TNF-α	NR NR∱* ^b NR		NR NR NR		
	(pg/ml)–Half-marathon Pre-race 15 min post-race 28 h post-race	NR NR↑ ^{*b} NR		NR NR NR		

Citation	Outcomes	Exercise		Control		
		Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	<i>p</i> value ^b
Saravia et al., 2010	THE PAPER WAS NOT AVAILABLE					
Millet et al., 2011	CRP (mg/dl)					
	Pre	2.0 (0.0)				
	Post	46.8 (24.8)*				
	2 days after	30.0 (19.7)*				
	5 days after	7.2 (3.7)*				
	9 days after	2.5 (1.2)				
	16 days after	2.3 (0.6)				
Auersperger et al., 2012	IL-6 (pg/ml)** Hs CRP (mg/dl) – Interval			Continuos		
		0.00 (1.40)		1 40 (1 0 4)		
	Baseline	0.88 (1.40)		1.48 (1.04)		
	3 weeks of training	1.13 (1.45)		1.27 (0.99)		
	Recovery 1	0.50 (0.66)		1.62 (2.92)		
	3 weeks of training	0.98 (0.88)		1.60 (1.52)		
	Recovery 2	0.49 (0.59)		1.92 (2.99)		
	Post	4.85 (12.54)		1.01 (1.28)		
Bernecker et al., 2013	IL-6 (ng/L)	2.06 (1.98–2.20)	31.93 (20.68–41.47) ^a			
	TNF-α (ng/L)	9.01 (7.16–10.26)	10.26 (9.33–12.31) ^a			
Chimenti et al., 2009	IL-8 (ng/ml)–Fall					
	Baseline	NR				
	Race	NR				
	TNF-α (pg/ml)–Fall					
	Baseline	NR				
	Race	NR				
	IL-8 (ng/ml)–Winter					
	Baseline	NR				
	Race	NR				
	TNF-α (pg/ml)–Winter					
	Baseline	NR				
	Race	NR				
	IL-8 (ng/ml)–Summer					
	Baseline	NR				
	Race	NR				
	TNF-α (pg/ml)–Summer					
	Baseline	NR				
	Race	NR				
Papassotiriou et al.,	CRP (mg/L)					
2008	Baseline	0.8 (0.1)				
2000	End of race	93.0 (12.6) ^d				
	48 h post	70.6 (11.5) ^{d,k}				
	IL-6 (ng/L)					
	Baseline	0.8 (0.1)				
	End of race	8376.0 (1819.8) ^d				
	48 h post	0.7 (0.1) ^{d,k}				
	TNF-α (ng/L)					
	Baseline	3.9 (0.9)				
	End of race	4.0 (0.8) ^d				
	48 h post	3.7 (0.7) ^{d,k}				
Kim et al., 2007	Hs CRP (IU/L)	. *				
1 MIT OL UL, 2007	Pre	2.0 (4.0)				
	100 km	6.0 (6.0)*				
	200 km	46.0 (28.0)* ^{,j}				
	IL-6 (pg/ml)	- \/				
	Pre	0.86 (0.17)				
	100 km	104.3 (45.5)				
	200 km	108.6 (28.4) ^j				
	200 NIII					

Citation	Outcomes	Exercise		Control		
		Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	р value ^b
	TNF-α					
	Pre	2.35 (1.56)				
	100 km	2.60 (1.43)				
	200 km	2.77 (1.82)				
Peters et al., 2004	CRP (mg/l)–Well trained	NR	NR↑ ^a	Less trained NR	NR↑ ^a	
Suzuki et al., 2003	TNF-α (pg/ml)					
	Plasma Urine**	0.31 (0.44)	0.29 (0.38)			
	IL1-β (pg/ml)					
	Plasma	0.43 (0.27)	0.52 (0.23)			
	Urine	1.7 (3.7)	7.1 (5.1) ^a			
	IL-6 (pg/ml)					
	Plasma Urine	1.27 (1.19) 2.86 (6.91)	101.40 (50.34) ^a 23.60 (19.94)			
	IL-8 (pg/ml)					
	Plasma Urine**	1.16 (0.70)	0.06 (6.95) ^a			
	IL-10 (pg/ml)					
	Plasma	8.0 (2.1)	32.8 (14.5) ^a			
	Urine	19.3 (6.3)	22.8 (3.8) ^a			
Bachi et al., 2015	IL-8 (pg/ml) PBMCs					
	Baseline	NR↑b		NR		
	Post	NR		NR		
	72 h post	NR↓d		NR		
	IL-8 (pg/ml) serum					
	Baseline	NR		NR		
	Post	NR↑d		NR		
	72 h post	NR↓I		NR		
	IL-10 (pg/ml) PBMCs					
	Baseline	NR↑b		NR		
	Post	NR NR↓d		NR NR		
	72 h post	inn↓u				
	IL-10 (pg/ml) serum					
	Baseline	NR NR∱d		NR NR		
	Post 70 h post	NR↓I		NR		
	72 h post	i vi tų t		INI I		
íłapcinska et al., 2013	CRP (mg/l)	0.8 (0.8)				
	Baseline 12 h running	3.4 (17.7) ^d				
	24 h running	30.0 (8.9) ^d				
	48 h running	63.5 (31.5) ^d				
	24 h post-race	45.5 (37.8) ^d				
	48 h post-race	28.0 (38.2) ^d				
	IL-6 (pg/ml)					
	Baseline	0.64 (0.34)				
	12 h running	35.86 (17.35) ^d				
	24 h running	33.25 (16.54) ^d				
	48 h running	23.20 (18.85) ^d				
	24 h post-race	7.39 (13.32) ^d				
	48 h post-race	2.19 (3.67)				
Rehm et al., 2013	sIFNγ (x1.000) pg/ml					
	Baseline	42.24 (26.75)				
	Pre-race	28.86 (23.16) ^d				
	Recovery	38.86 (28.76)				
	sIL4 pg/ml					
	Baseline	3.59 (5.08)				
	Pre-race	8.65 (10.71) ^{d,m}				
	Recovery	4.02 (6.06)				

Citation	Outcomes	Exercise		Control		
		Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	р value ^b
	slL10 pg/ml					
	Baseline	389.73 (254.22)				
	Pre-race	248.9 (191.8) ^d				
	Recovery	323.79 (240.5)				
Fehrenbach et al., 2000	IL-8 pg/ml					
	Baseline	5.0 (6.5)				
	0 h	30.7 (5.3) ^d				
	3 h	8.9 (11.3)				
	24 h	3.9 (6.3)				
	TNF-α pg/ml					
	Baseline	0.3 (0.2)				
	0 h	1.2 (0.9) 0.6 (0.6)				
	3 h	0.8 (0.8)				
	24 h	0.3 (0.5)				
Schobersberger et al.,	IL-6 pg/ml	0.0 (0–1.75)				
2000	Baseline	60.0 (40.5–180.0) ^d				
	0 h 2 h	65.0 (22.3–81.0) ^d				
	Z n Day1	0.0 (0.0–5.0)				
	Day3	0.0 (0.0–0.8)				
	Day5	0.0 (0.0–4.0)				
	IL1-ra pg/ml					
	Baseline	23.0 (18.0–33.5)				
	0 h	720.0 (370.0–42.01) ^d				
	2 h	733.0 (443.0–41.63) ^d				
	Day1	100.0 (43.0–221.0) ^d				
	Day3	84.0 (25.0–246.0)				
	Day5	28.0 (17.0–215.0)				
	TNF-α pg/ml					
	Baseline	13.0 (10.3–15.0)				
	0 h	17.5 (15.0–30.0) ^d				
	2 h	18.0 (14.3–24.8) ^d				
	Day1	16.5 (12.8–28.0) ^d				
	Day3	16.2 (11.0–17.5) ^d				
	Day5	16.0 (12.0–20.0) ^d				
	sTNF-RI ng/ml					
	Baseline	2.4 (2.1–2.9)				
	0 h	5.7 (4.5–11.7) ^d				
	2 h	6.2 (4.2–9.0) ^d				
	Day1	3.9 (3.2–5.3) ^d				
	Day3	3.5 (2.8–5.0) ^d 2.8 (2.3–3.7) ^d				
	Day5	2.0 (2.3-3.7)				
	sTNF-RII ng/ml	11 1 (7 0 10 0)				
	Baseline	11.1 (7.2–13.9) 11.8 (10.4–22.4) ^d				
	0 h 2 h	12.9 (10.6–21.3) ^d				
	Z n Day1	15.7 (10.4–19.7) ^d				
	Day3	15.1 (9.8–19.9) ^d				
	Day5	11.6 (8.1–17.7) ^d				
Suzuki et al., 2000	IL-1β pg/ml	1.8 (3.6)	1.4 (2.1)			
0020NI 6L al., 2000	IL-rp pg/ml	59.0 (37.0)	12629.0 (12360.0) ^a			
	IL-2 pg/ml	73.0 (44.0)	50.0 (49.0) ^a			
	IL-4 pg/ml	3.5 (1.45)	4.9 (4.1)			
	IL-6 pg/ml	<1.1 (1.3)	120.0 (79.0) ^a			
	IL-8 pg/ml	22.0 (19)	5.5 (25.0) ^a			
	IL-10 pg/ml	13.9 (12.3)	47.9 (23.1) ^a			
	IL-12 pg/ml**	_	_			
	TNF-α pg/ml**	-	-			
	IFN-α pg/ml**	-	-			
	IFN-γ pg/ml	1.5 (0.8)	1.4 (1.0)			

Citation	Outcomes	Exercise		Control		
		Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	<i>p</i> value ^b
Vaisberg et al., 2012	IL-6 pg/ml					
	Baseline	NR		NR		NS
	Immediatly	NR ^{d,n↑}		-		NS
	72 h	NR		-		NS
	TNF-α					
	Baseline	NR ^{b↑}		NR		< 0.05
	Immediatly	NR ^{d,n↑}		-		NS
	72 h	NR		-		NS
Tomaszewski et al., 2003 (64)	CRP mg/L	0.3 (0.2–0.7)	1.8 (1.0–3.4) ^a			
Bonsignore et al., 2002	TNF-α pg/ml–Half-marathon	TNF- α pg/ml–Marathon				
	Baseline	NR		NR		
	End of race ^{d↑}	NR		NR		
	Post	NR		NR		
	IL-6					
	pg/ml–Half-marathon	IL-6 pg/ml–Marathon				
	Baseline	NR		NR		NS
	End of race ^{d↑}	NR ^{d,b↑}		NR		NS
	Post	NR		NR		NS
Tomaszewski et al.,	CRP mg/dl–Lean Elite	CRP mg/dl–Lean		Obese non-elite		
2003	Baseline	Non-elite		NR		
	Marathon ^{d↑}	NR		NR ^d ↑		
	24 h ^{d↑}	NR ^{d↑} NR ^{d↑}		NR ^d ↑b(in relation to Lean Elit	e)↑	
	IL-6 pg/ml–Lean Elite Baseline	IL-6 pg/ml–Lean non-elite		Obese non-elite NR ^{b(in} relation to Lean Elite)↑		
	Immediately post	NR ^{b(in relation Lean Elite)↑}				
	marathon ^{d↑}			NR ^{d↑}		
	24 h ^{d↑,e↓}	NR ^{d↑} NR ^{d↑,e↓}		NR ^{d↑,e↓}		
	TNF-α pg/ml–Lean Elite	TNF-α pg/ml–Lean Non-elite		Obese non-elite NR		
	Baseline	NR		NR		
	Immediately post	NR		NR ^{d↑,e↑}		
	marathon 24 h ^{d↑,e↑}	NR ^{d↑,e↑}				
	IL-10 pg/ml–Lean Elite	IL-10 pg/ml–Lean		Obese non-elite		
	Baseline	Non-elite		NR		
	b(in relation to obese non-elite)			NR ^{d↑}		
		b(in relation to obese non-elite)↓		NR ^{e↓}		
	Immediately post					
	marathon ^d ↑ 24 h ^{e↓}	NR ^{d↑} NB ^{e↓}				
Waśkiewicz et al., 2012	IL-6 mg/dL					
11001001102 01 01., 2012	Baseline	0.87 (0.68)				
	Immediately post Marathon	20.29 (7.77) ^d				
	Post 12 h	27.36 (7.67) ^d				
	Post 24 h	28.49 (11.99) ^d				
	hsCRP mg/dL	17(07)				
	Baseline	1.7 (2.7)				
	Immediately post	1.7 (2.5)				
	Marathon	8.7 (4.6) ^d				
	Post 12 h	39.2 (16.7) ^d				
	Post 24 h	09.2 (10.7)-				

Citation	Outcomes	Exercise		Control		
		Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	<i>p</i> value ^b
Chimenti et al., 2010	IL-8 ng/ml–Octuber IL-8 ng/ml–May IL-8 ng/ml–November	NR NR NR	NR ^a NR ^a NR ^a			
Ng et al., 2008	IL-6 pg/ml IL-10 pg/ml IL-1ra TNF-α pg/ml IL1-β	9.2 (4.1) ^a 6.4 (2.4) ^a 154.1 (0.4) NR NR	15.2 (5.3) 9.6 (3.0) 189.8 (61.9) NR NR			
Siegel et al., 2007	IL-6 pg/mL Baseline 2 h post race 24 h post race	1.6 (0.45) 66.6 (11.9) ^d 4.3 (0.6)				
	CRP ng/dL Baseline 2 h post race 24 h post race	0.10 (0.02) 0.10 (0.03) 2.3 (0.53) ^d				
Shin and Lee, 2013	CRP IU/L Baseline 100 km 200 km 308 km	NR NR ^{d↑} NR ^{d↑} NR ^{d↑}				
	IL-6 pg/ml Baseline 100 km 200 km 308 km	NR NR ^d ↑ NR ^d ↑ NR ^d ↑				
	IL-10 pg/ml Baseline 100 km 200 km 308 km	NR NR ^d ↑ NR ^d ↑ NR ^d ↑				
	IL-8 pg/ml Baseline 100 km 200 km 308 km	NR NR ^d ↑ NR ^d ↑ NR ^d ↑				
Jee and Jin, 2012	hsCRP IU/L Baseline 100 km 200 km 308 km	0.40 (0.10) 5.06 (1.46) ^d 25.56 (3.82) ^{d,i} 21.87 (3.49) ^{d,i}				
	TNF-α pg/ml Baseline 100 km 200 km 308 km	3.68 (0.15) 4.00 (0.20) 3.37 (0.18) ⁱ 4.50 (0.36) ^{d,o}				
Santos et al., 2013	IL-6 pg/ml IL1-ra pg/ml TNF-α pg/ml IL-8 pg/ml** IL-10 pg/ml** CPB LI	106.00 (38.5) 18.0 (12.0) 32.3 (13.3) - - 5.2 (0.5)	435.0 (145.5) ^a 2708.0 (355.0) ^a 32.4 (7.7) 40.4 (20.2) 32.0 (11.2) 5 3 (0 7)			
Hewing et al., 2015	CRP UL CRP mg/dl Baseline Post 14 days lates	5.2 (0.5) 0.10 (0.05–0.21) 0.06 (0.04–0.12) ^d 0.10 (0.06–0.18)	5.3 (0.7)			

Citation	Outcomes	Exercise		Control		
		Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	<i>p</i> value ^b
Nieman et al., 2003	IL-10 pg/ml Baseline 90 km 160 km	4.65 (0.40) 39.7 (8.0) ^d 49.0 (8.2) ^d				
	IL1-ra pg/ml Baseline 90 km 160 km	229.0 (14.0) 2330.0 (421.0) ^d 1616.0 (255.0) ^d				
	IL-6 pg/ml Baseline 90 km 160 km	1.19 (0.15) 58.6 (4.6) ^d 60.9 (9.4) ^d				
	IL-8 pg/ml Baseline 90 km 160 km	6.31 (1.09) 20.4 (2.1) ^d 22.0 (2.4) ^d				
Uchakin et al., 2003	IL-2 (IU/ml) Baseline 0 h 1 h	6.0 (2.5) 2.0 (0.4) ^d 1.6 (0.2) ^d				
	24 h 48 h 5 days 8 days	6.6 (1.0) 5.0 (0.6) 8.4 (1.2) ^d 5.7 (1.0)				
	INF-γ (IU/ml) Baseline 0 h 1 h	210.8 (26.6) 17.7 (3.5) ^d 14.4 (3.3) ^d				
	24 h 48 h 5 days 8 days	196.4 (15.8) 154.8 (20.0) 272.8 (23.9) ^d 141.0 (18.1)				
	IL-10 (pg/ml) Baseline 0 h 1 h	445.3 (69.3) 310.0 (44.3) 463.7 (146.9)				
	24 h 48 h 5 days 8 days	262.3 (27.0) 288.2 (33.2) 441.8 (73.9) 355.0 (47.9)				
	TNF-α (pg/ml) Baseline 0 h 1 h 24 h	16937.0 (1800.8) 9594.0 (1421.5) ^d 1394.0 (1522.8) ^d 12859.0 (1585.0) ^d				
	48 h 5 days 8 days	12899.0 (1720.8) ^d 12276.0 (12276.7) ^d 14043.0 (1231.2)				
	IL1-β (pg/ml) Baseline 0 h 1 h	4377.1 (664.5) 2937.1 (696.8) 3162.9 (617.1) 3520.0 (743.3)				
	24 h 48 h 5 days 8 days	2342.8 (359.6) ^d 2388.6 (481.9) ^d 2817.1 (243.6)				

Citation	Outcomes	Exercise	Control			
		Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	<i>p</i> value ^l
	IL-6 (pg/ml)					
	Baseline	16571.4 (2058.1)				
	0 h	13585.7 (3105.4)				
	1 h	16200.0 (1740.2)				
	24 h	6514.3 (985.0) ^d				
	48 h	8414.3 (1470.9) ^d				
	5 days	10642.8 (2291.1)				
	8 days	14271.4 (1331.8)				
Vrakic-Sposta et al., 2015	IL-6 (pg/ml) Plama	1.29 ± 0.54	66.42 ± 36.92^{a} 1.33 ± 0.56^{a}			
	IL-6 (pg/ml) Urine	0.71 ± 0.17				
Stuempfle et al., 2016		Without Nausea	Without Nausea	With Nausea	With Nausea	NS
	IL-6 (pg/ml)	0.9 ± 0.4	105.7 ± 53.6 ^a	1.0 ± 0.7	78.6 ± 62.5 ^a	NS
	CRP (ng/ml)	323 ± 487	31,448 ± 13,149 ^a	1686 ± 2607	46,361 ± 29,708 ^a	
Nieman et al., 2016	IL-6 (pg/ml)	Male		Female		
	Pre-run	3.17 ± 0.41		2.88 ± 0.91		
	Post-run	$11.8 \pm 2.23^{*}$		7.46 ± 0.89		
	1-h Post-run	$9.03 \pm 1.41^{*}$		7.37 ± 1.56		
	24-h Post run	2.36 ± 0.31		2.99 ± 1.19		
	IL-8 (pg/ml) Pre-run	9.99 ± 1.00		8.88 ± 0.68		
	Post-run	9.99 ± 1.00 22.4 ± 2.9*		16.1 ± 1.1		
	1-h Post-run	22.4 ± 2.9 18.1 ± 1.5*		16.6 ± 2.1		
	24-h Post run	8.12 ± 0.50		9.17 ± 0.68		
	IL-10 (pg/ml)	0.12 ± 0.00		0.17 ± 0.00		
	Pre-run	2.50 ± 0.18		3.31 ± 0.64		
	Post-run	$9.33 \pm 1.77^*$		6.20 ± 0.91		
	1-h Post-run	10.5 ± 1.9* ^b		6.03 ± 0.81		
	24-h Post run	2.51 ± 0.53		3.51 ± 0.81		
	IL-1ra (pg/ml)					
	Pre-run	111 ± 10.8		125 ± 18.4		
	Post-run	216 ± 14.8		215 ± 40.5		
	1-h Post-run	385 ± 83.8		300 ± 80.4		
	24-h Post run	111 ± 9.8		117 ± 9.3		
	CRP (mg/l)					
	Pre-run	0.71 ± 0.15		0.70 ± 0.19		
	Post-run	0.67 ± 0.17		0.71 ± 0.22		
	1-h Post-run	0.62 ± 0.16		0.70 ± 0.19		
	24-h Post run	$2.56 \pm 0.50^{*}$		2.02 ± 0.73		
Arakawa et al., 2016	IL-6 (pg/ml)	0.77 + 0.00				
	Baseline	0.77 ± 0.26 26.52 $\pm 5.05^{d}$				
	Day 1	19.28 ± 1.99^{d}				
	Day 2	3.35 ± 0.96				
	Day 3 Day 5	6.53 ± 4.16				
	Day 5 Day 7	1.40 ± 0.38				
	CRP (mg/dl)	1110 ± 0.00				
	Baseline	0.07 ± 0.03				
	Day 1	0.07 ± 0.03				
	Day 2	0.72 ± 0.14^{d}				
	Day 3	1.45 ± 0.29^{d}				
	Day 5	0.64 ± 0.18				
	Day 7	0.57 ± 0.28				
	TNF-α (pg/ml)					
	Baseline	0.91 ± 0.06				
	Day 1	0.95 ± 0.09				
	Day 2	0.84 ± 0.06				
	Day 3	0.95 ± 0.08				
	Day 5	0.98 ± 0.07				
	Day 7	1.15 ± 0.17				

Citation	Outcomes	Exercise				
		Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	<i>p</i> value ^b
Mohamed et al., 2016	IL-6 (pg/ml)	Sedentary Subjects	Long-Distance Runners	Middle-Distance		
	VAMEVAL test			Runners		
	Before	NR	NR	NR		
	After	NR ^{d↑}	NR ^d ↑	NR ^{d↑}		
	IL-6 (pg/ml)					
	Limited-time test					
	Before	NR	NR	NR		
	After	NR ^{d,b↑}	NR ^{d↑}	NR ^{d↑}		
	TNF-α (pg/ml)					
	VAMEVAL test					
	Before	NR	NR	NR		
	After	NR ^{d↑}	NR ^{d↑}	NR ^{d↑}		
	TNF-α (pg/ml)					
	Limited-time test					
	Before	NR	NR	NR		
	After	NR ^{d↑}	NR ^{d↑}	NR ^{d↑}		
Cairns and Hew-Butler,	IL-6 (pg/ml)	With hyponatremia	Non-hyponatremia			
2015	Before	0.1 ± 0.2	0.5 ± 0.4			
	After	$10.6 \pm 6.1^{*}$	$8.4 \pm 2.8^{*}$			
Gill et al., 2015a	IL-6 (pg/ml)					
olin oc ally 20 roa	Before	0.4 (0.3 to 0.5)				
	After	14.5 (9.3 to 19.7)*				
	IL-1 β (pg/ml)					
	Before	0.1 (0.0 to 0.3)				
	After	0.6 (0.1 to 1.1)*				
	TNF-α (pg/ml)					
	Before	2.8 (2.5 to 3.2)				
	After	3.8. (3.5 to 4.2)*				
	IFN-γ (pg/ml)					
	Before	1.0 (0.6 to 1.4)				
	After	1.2 (0.3 to 2.2)				
	IL-10 (pg/ml)					
	Before	2.1 (1.3 to 2.9)				
	After	12.8 (7.3 to 18.2)*				
	IL-8 (pg/ml)					
	Before	11.4 (9.4 to 13.4)				
	After	38.7 (26.3 to 51.1)*				
Krzeminski et al., 2016	TNF-α (pg/ml)					
,	Pre-race	1.39 ± 0.09				
	Post-race	$1.63 \pm 0.09^{*}$				
	90 min post-race	1.54 ± 0.09				
	IL-6 (pg/ml)					
	Pre-race	0.54 ± 0.07				
	Post-race	$47.35 \pm 8.48^{*}$				
	90 min post-race	$37.67 \pm 7.94^{*1}$				
	IL-10 (pg/ml)					
	Pre-race	0.31 ± 0.06				
	Post-race	$5.04 \pm 1.34^{*}$				
	90 min post-race	$1.24 \pm 0.35^{*}$				
	IL-18 (pg/ml)					
	Pre-race	75.08 ± 9.46				
	Post-race	$96.74 \pm 9.92^{*}$				
	90 min post-race	101.25 ± 9.28				
	IL-1β (pg/ml)					
	Pre-race	0.76 ± 0.24				
	Post-race	1.30 ± 0.27				
	90 min post-race	0.70 ± 0.12				

Citation	Outcomes	Exercise		Control		
		Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	Pre [mean (SD)] or median (IQR)	Post [mean (SD)] or median (IQR)	<i>p</i> value ^b
Nielsen et al., 2016	IL-1β (pg/ml)	Marathon		Half-Marathon		
	Pre-race	NR		NR		
	Post-race	NR		NR		
	II-6 (pg/ml)					
	Pre-race	NR		NR		
	Post-race	NR*↑		NR*↑		
	IL-8 (pg/ml)					
	Pre-race	NR		NR		
	Post-race	NR*↑		NR*↑		
	IL-10 (pg/ml)					
	Pre-race	NR		NR		
	Post-race	NR*↑		NR		
	TNF-α (pg/ml)					
	Pre-race	NR		NR		
	Post-race	NR		NR		
	IFN-γ (pg/ml)					
	Pre-race	NR		NR		
	Post-race	NR		NR		

NS, not significant; –, not compared or evaluated; **, below the detectable plasma concentrations; NR, not reported; PBMCs, produced by peripheral blood mononuclear cells; IL-6, interleukin 6; CRP, C-reactive protein; IL-8, interleukin- eight; IL-2, interleukin two; IL-4, interleukin four; IL-10, interleukin tem; IL-12, interleukin 12; IFN- γ , interferon gama; TNF- α , turmor necroses factor alpha; IL-1-ra, receptor antogonist of interleukin one; IL-1- β , interleukin beta; sTNF-R, soluble receptor for turmor necroses factor alpha; IL-6, soluble receptor antagonista for interleukin six; 0 h, immediately post-race; [†] Significant difference (p < 0.05) vs. pre-stage 1; *Significant difference (p < 0.05) vs. pre; ^aSignificant difference (p < 0.05) vs. pre; ^aSignificant difference (p < 0.05) vs. bre; ^aSignificant difference (p < 0.05) vs. bre; ^bSignificant difference (p < 0.05) vs. bre; ^aSignificant difference (p < 0.05) vs. bre; ^bSignificant difference (p < 0.05) vs. bre; ^bSignificant difference (p < 0.05) vs. bresetage 1; *Significant difference (p < 0.05) vs. bresetage 1; between pre and post for the same group; ^bSignificant difference (p < 0.05) between pre and post for the same group; ^bSignificant difference (p < 0.05) vs. 25 km; ^gSignificant difference (p < 0.05) vs. 50 km; ^hSignificant difference (p < 0.05) vs. baseline; ^bSignificant difference (p < 0.05) vs. immediately; ^fSignificant difference (p < 0.05) vs. 25 km; ^gSignificant difference (p < 0.05) vs. 50 km; ^hSignificant difference (p < 0.05) vs. post; ^hSignificant difference (p < 0.05) vs. 100 km; ^kSignificant difference (p < 0.05) vs. end of race; ^lSignificant difference (p < 0.05) vs. post; ^mSignificant difference (p < 0.05) vs. post; ^mSignificant difference (p < 0.05) vs. post; ^lSignificant difference (p < 0.05) vs

Boullosa et al., 2013) the training and competitive volumes of runners competing in ultra-endurance events. In addition, another new aspect that must be raised in further studies is the imbalance between training phases and recovery not reported in the studies included in this systematic review. Thus, since functional overreaching might be related to inflammation (Steinacker et al., 2004), further studies should explore these relationship in conjunction with other biological markers of overreaching.

Another important characteristic for a better characterization of runners is their fitness level. For instance, maximum oxygen consumption (VO₂max), which is the gold standard for aerobic evaluation, has been reported only in 14 articles (3 studies with participants of both sexes (Nieman et al., 2003, 2016; Ng et al., 2008; Chimenti et al., 2009; Jee et al., 2013), and 11 studies with males (Millet et al., 2011; Jee and Jin, 2012; Waśkiewicz et al., 2012; Kłapcinska et al., 2013; Shin and Lee, 2013; Wilhelm et al., 2014; Kim et al., 2015; Krzeminski et al., 2016; Mohamed et al., 2016). The participants of these studies could be considered recreational runners when classified by their actual VO₂max (44-51 and 35-41 mL.kg-1.min-1, for males and females, respectively) (Martin and Coe, 2007). In contrast, no study included elite runners when classified by their actual VO2max (70-85 mL.kg-1.min-1 and 61-73 mL.kg-1.min-1 for males and females respectively) (Martin and Coe, 2007). Moreover, runners' classification in the current review has been challenging when using the selected criteria (see Table 3) (Stirling and Kerr, 2006; MeSH, 2015). Thus, further studies should provide all these informations for a better characterization of runners. As we did not perceive an influence of aerobic fitness on inflammatory markers, further studies should elaborate on this relationship while controlling other runners' characteristics as training experience. Additionally, the influence of other fitness components as muscle strength capacity should be assessed in further studies for verifying the potentially protective effect for muscle damage and therefore on inflammation.

Another important limitation for generalization of the results refers to the heterogeneity of running exercises (e.g., distance, intensity) used for evaluation of acute inflammatory responses. Furthermore, ambient characteristics (e.g., altitude, temperature) and race profile (e.g., uphill and downhill running) which have been suggested to influence muscle contraction and physiological responses (Vernillo et al., 2017), have not been always reported (Millet et al., 2011; Saugy et al., 2013). These aspects should be controlled in further studies for isolating the relative effect of every specific factor on inflammation.

Body Fatness and Inflammation

Given the relationship between adipose tissue and inflammation (Pedersen and Febbraio, 2012), attention should be paid to overweight and obese runners. For instance, a higher level of CRP 24 h following a marathon has been observed in obese nonelite runners when compared to lean elite runners (Nickel et al., 2012). Furthermore, obese non-elite runners when compared to lean elite and lean non-elite runners demonstrated a higher level of IL-6 and a lower level of IL-10 serum levels at baseline (Nickel et al., 2012). However, all groups presented an increase for serum IL-10 and TNF- α , and a decrease for serum IL-6 levels, immediately post-marathon (Nickel et al., 2012). It must be considered that increments in IL-10 induced by exercise may be responsible for the elevation in IL1-ra which exerts an anti-inflammatory action by antagonizing IL-1 and IL-1 β (Dinarello, 2000; Moldoveanu et al., 2001; Petersen and Pedersen, 2005; Pedersen, 2011). Nevertheless, it could be suggested that, in overweighted individuals, a higher pro-inflammatory status at baseline and post-marathon could be observed, with unknow consequences for health in the long term.

Inflammation and Cardiovascular Health

One relevant issue refers to the link between inflammation and cardiovascular health. Interestingly, the exercise-induced increase of IL-6 after the marathon in 20 lean male runners was associated with a lower prevalence of arrhythmias during and after the marathon race (Grabs et al., 2015). When produced by muscle contraction, IL-6 stimulates the synthesis of other anti-inflammatory cytokines such as IL-1ra and IL-10, thus providing an inhibitory effect on pro-inflammatory cytokines such as IL-1 β and TNF- α (Pedersen and Febbraio, 2012; Pedersen, 2013). However, CRP, a strong predictor of cardiovascular events, is an acute phase protein synthesized in the liver by the stimulation of IL-6 (Ridker et al., 2002). Chronic endurance training may decrease CRP values, especially when accompanied by a loss in fat mass, therefore promoting further reduction of risk for cardiovascular events (Fallon et al., 2001; Tomaszewski et al., 2003; Walsh et al., 2011; Grabs et al., 2015; Kim et al., 2015). Of note, CRP may be more susceptible to chronically decrease in individuals presenting higher baseline levels (Barnett et al., 2005). Therefore, caution should be taken when evaluating the anti- and pro-inflammatory effects of running in individuals with different characteristics regarding cardiovascular risk factors (e.g., body composition) (Moldoveanu et al., 2000; Petersen and Pedersen, 2005; Walsh et al., 2011).

Studies' Characteristics

Most studies included in this systematic review were acute interventions (49 studies). However acute changes in inflammatory markers might not be related with anti- and pro-inflammatory outcomes during chronic aerobic training interventions. For instance, there were divergent responses for CRP changes in chronic interventions (Mattusch et al., 2000; Auersperger et al., 2012). Thus, while Mattusch et al. (2000) observed a reduction in CRP levels, Auersperger et al. (2012) did not observe any change. Therefore, further studies must

REFERENCES

Abbasi, A., Fehrenbach, E., Hauth, M., Walter, M., Hudemann, J., Wank, V., et al. (2013). Changes in spontaneous and LPS-induced *ex vivo* cytokine production and mRNA expression in male and female athletes following prolonged exhaustive exercise. *Exerc. Immunol. Rev.* 19, 8–28. consider this important limitation, while providing training load charactersitics as volume, intensity, and frequency of training sessions. An important question to be answered refers to the minimal training load required for runners of different levels when preparing different competitive distances, while analyzing the impact of these factors on inflammatory markers. Additionally, there is a prevalence of male runners on literature therefore more studies with female runners are needed.

CONCLUSION

In summary, our results revealed that acute and chronic endurance running may affect anti- and pro-inflammatory markes but methodological differences between studies do not allow comparisons or generalization of the results. Only two studies were chronic interventions. There are no studies with elite athletes. Thus, RCTs are urgently needed to identify the appropriate dose of endurance running (volume, intensity, and frequency) required to elicit improvements in inflammatory markers in runners of different levels and training background. External (e.g., ambient characteristics, race profile) and internal factors (e.g., fitness level, training experience) to runners should be considered in further studies for a better understanding of the relationship between running and the mediators of inflammation. The information provided in this systematic review would help practitioners for better designing further studies while providing reference values for a better understanding of inflammatory responses after different running events.

AUTHOR CONTRIBUTIONS

Conception and design: EB, CC, ON, JP, and DB. Search: EB, DN, and FS. Eligibility and outcome measures: EB and DN. Quality assessment: DN and JP. Writing of the manuscript: EB, DN, JP, ON, CC, and DB. Revision and approval of the final manuscript version and interpretation of the results: EB, DN, JP, ON, CC, FS, and DB.

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- Arakawa, K., Hosono, A., Shibata, K., Ghadimi, R., Fuku, M., Goto, C., et al. (2016). Changes in blood biochemical markers before, during, and after a 2-day ultramarathon. *Open Access J Sports Med.* 7, 43–50. doi: 10.2147/OAJSM.S97468
- Auersperger, I., Knap, B., Jerin, A., Blagus, R., Lainscak, M., Skitek, M., et al. (2012). The effects of 8 weeks of endurance running on hepcidin concentrations,

inflammatory parameters, and iron status in female runners. Int. J. Sport Nutr. Exerc. Metab. 1, 55–63. doi: 10.1123/ijsnem.22.1.55

- Bachi, A. L., Rios, F. J., Vaisberg, P. H., Martins, M., de Sá, M. C., Victorino, A. B., et al. (2015). Neuro-immuno-endocrine modulation in marathon runners. *Neuroimmunomodulation* 22, 196–202. doi: 10.1159/000363061
- Barnett, A. G., van der Pols, J. C., and Dobson, A. J. (2005). Regression to the mean: what it is and how to deal with it. *Int. J. Epidemiol.* 34, 215–220. doi: 10.1093/ije/dyh299
- Bernecker, C., Scherr, J., Schinner, S., Braun, S., Scherbaum, W. A., and Halle, M. (2013). Evidence for an exercise induced increase of TNF-alpha and IL-6 in marathon runners. *Scand. J. Med. Sci. Sports* 23, 207–214. doi: 10.1111/j.1600-0838.2011.01372.x
- Bonsignore, M. R., Morici, G., Santoro, A., Pagano, M., Cascio, L., Bonanno, A., et al. (2002). Circulating hematopoietic progenitor cells in runners. J. Appl. Physiol. 93, 1691–1697. doi: 10.1152/japplphysiol.00376.2002
- Boullosa, D. A., Abreu, L., Varela-Sanz, A., and Mujika, I. (2013). Do olympic athletes train as in the Paleolithic era? *Sports Med.* 43, 909–917. doi: 10.1007/s40279-013-0086-1
- Bramble, D. M., and Lieberman, D. E. (2004). Endurance running and the evolution of Homo. *Nature* 432, 345–352. doi: 10.1038/nature03052
- Cairns, R. S., and Hew-Butler, T. (2015). Incidence of exercise-associated hyponatremia and its association with nonosmotic stimuli of arginine vasopressin in the GNW100s ultra-endurance marathon. *Clin. J. Sport Med.* 25, 347–354. doi: 10.1097/JSM.00000000000144
- Chiampas, G. T., and Goyal, A. V. (2015). Innovative operations measures and nutritional support for mass endurance events. *Sports Med.* 45, 61–69. doi: 10.1007/s40279-015-0396-6
- Chimenti, L., Morici, G., Paterno, A., Bonanno, A., Vultaggio, M., Bellia, V., et al. (2009). Environmental conditions, air pollutants, and airway cells in runners: a longitudinal field study. *J. Sports Sci.* 27, 925–935. doi: 10.1080/02640410902946493
- Chimenti, L., Morici, G., Paternò, A., Santagata, R., Bonanno, A., Profita, M., et al. (2010). Bronchial epithelial damage after a half-marathon in nonasthmatic amateur runners. Am. J. Physiol. Lung Cell. Mol. Physiol. 298, L857–L862. doi: 10.1152/ajplung.00053.2010
- Dinarello, C. A. (2000). The role of the interleukin-1-receptor antagonist in blocking inflammation mediated by interleukin-1. N. Engl. J. Med. 343, 732–734. doi: 10.1056/NEJM200009073431011
- Downs, S. H., and Black, N. (1998). The feasibility of creating a checklist for the assessment of the methodological quality both of randomised and nonrandomised studies of health care interventions. *J. Epidemiol. Commun. Health* 52, 377–384. doi: 10.1136/jech.52.6.377
- Fallon, K. E., Fallon, S. K., and Boston, T. (2001). The acute phase response and exercise: court and field sports. *Br. J. Sports Med.* 35, 170–173. doi: 10.1136/bjsm.35.3.170
- Febbraio, M. A., and Pedersen, B. K. (2005). Contraction-induced myokine production and release: is skeletal muscle an endocrine organ? *Exerc. Sport Sci. Rev.* 33, 114–119. doi: 10.1097/00003677-200507000-00003
- Fehrenbach, E., Niess, A. M., Schlotz, E., Passek, F., Dickhuth, H. H., and Northoff, H. (2000). Transcriptional and translational regulation of heat shock proteins in leukocytes of endurance runners. J. Appl. Physiol. 89, 704–710.
- Gill, S. K., Hankey, J., Wright, A., Marczak, S., Hemming, K., Allerton, D. M., et al. (2015a). The impact of a 24-h ultra-marathon on circulatory endotoxin and cytokine profile. *Int. J. Sports Med.* 36, 688–695. doi: 10.1055/s-0034-1398535
- Gill, S. K., Teixeira, A., Rama, L., Prestes, J., Rosado, F., Hankey, J., et al. (2015b). Circulatory endotoxin concentration and cytokine profile in response to exertional-heat stress during a multi-stage ultra-marathon competition. *Exerc. Immunol. Rev.* 21, 114–128.
- Gleeson, M. (2007). Immune function in sport and exercise. J. Appl. Physiol. 103, 693–699. doi: 10.1152/japplphysiol.00008.2007
- Grabs, V., Peres, T., Zelger, O., Haller, B., Pressler, A., Braun, S., et al. (2015). Decreased prevalence of cardiac arrhythmias during and after vigorous and prolonged exercise in healthy male marathon runners. *Am. Heart J.* 170, 149–155. doi: 10.1016/j.ahj.2015.04.001
- Hewing, B., Schattke, S., Spethmann, S., Sanad, W., Schroeckh, S., Schimke, I., et al. (2015). Cardiac and renal function in a large cohort of amateur marathon runners. *Cardiovasc. Ultrasound.* 13:13. doi: 10.1186/s12947-015-0007-6

- Hubble, K. M., Fatovich, D. M., Grasko, J. M., and Vasikaran, S. D. (2009). Cardiac troponin increases among marathon runners in the Perth Marathon: the Troponin in Marathons (TRIM) study. *Med. J. Aust.*, 190, 91–93.
- Jarvinen, T. A., Jarvinen, M., and Kalimo, H. (2013). Regeneration of injured skeletal muscle after the injury. *Muscles Ligaments Tendons J.* 3, 337–345.
- Jee, H., and Jin, Y. (2012). Effects of prolonged endurance exercise on vascular endothelial and inflammation markers. *J. Sports Sci. Med.* 11, 719–726.
- Jee, H., Park, J., Oh, J.-G., Lee, Y.-H., Shin, K.-A., and Kim, Y.-J. (2013). Effect of a prolonged endurance marathon on vascular endothelial and inflammation markers in runners with exercise-induced hypertension. Am. J. Phys. Med. Rehabil. 92, 513–522. doi: 10.1097/PHM.0b013e31829232db
- Karstoft, K., Solomon, T. P., Laye, M. J., and Pedersen, B. K. (2013). Daily marathon running for a week-the biochemical and body compositional effects of participation. *J. Strength Cond. Res.* 27, 2927–2933. doi: 10.1519/JSC.0b013e318289e39d
- Kasprowicz, K., Ziemann, E., Ratkowski, W., Laskowski, R., Kaczor, J. J., Dadci, R., et al. (2013). Running a 100-km ultra-marathon induces an inflammatory response but does not raise the level of the plasma iron-regulatory protein hepcidin. J. Sports Med. Phys. Fitness. 53, 533–537.
- Kim, H. J., Lee, Y. H., and Kim, C. K. (2007). Biomarkers of muscle and cartilage damage and inflammation during a 200 km run. *Eur. J. Appl. Physiol.* 99, 443–447. doi: 10.1007/s00421-006-0362-y
- Kim, Y. J., Ahn, J. K., Shin, K. A., Kim, C. H., Lee, Y. H., and Park, K. M. (2015). Correlation of cardiac markers and biomarkers with blood pressure of middleaged marathon runners. J. Clin. Hypertens. 17, 868–873. doi: 10.1111/jch.12591
- Kłapcinska, B., Waskiewicz, Z., Chrapusta, S. J., Sadowska-Krepa, E., Czuba, M., and Langfort, J. (2013). Metabolic responses to a 48-h ultra-marathon run in middle-aged male amateur runners. *Eur. J. Appl. Physiol.* 113, 2781–2793. doi: 10.1007/s00421-013-2714-8
- Krzeminski, K., Buraczewska, M., Miśkiewicz, Z., Dabrowski, J., Steczkowska, M., Kozacz, A., et al. (2016). Effect of ultra-endurance exercise on left ventricular performance and plasma cytokines in healthy trained men. *Biol. Sport.* 33, 63–69. doi: 10.5604/20831862.1189767
- Liberati, A., Altman, D. G., Tetzlaff, J., Mulrow, C., Gøtzsche, P. C., Ioannidis, J. P., et al. (2009). The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate healthcare interventions: explanation and elaboration. *Br. Med. J.* 339:b2700. doi: 10.1136/bmj.b2700
- Martin, D. E., and Coe, P. N. (2007). Entrenamiento para Corredores de Fondo y Medio Fondo. Paidotribo.
- Mattusch, F., Dufaux, B., Heine, O., Mertens, I., and Rost, R. (2000). Reduction of the plasma concentration of C-reactive protein following nine months of endurance training. *Int. J. Sports Med.* 21, 21–24. doi: 10.1055/s-2000-8852
- Meeusen, R., Duclos, M., Foster, C., Fry, A., Gleeson, M., Nieman, D., et al. (2013). Prevention, diagnosis, and treatment of the overtraining syndrome: joint consensus statement of the European College of Sport Science and the American College of Sports Medicine. *Med. Sci. Sports Exerc.* 45, 186–205. doi: 10.1249/MSS.0b013e318279a10a
- MeSH (2015). MeSH (Medical Subject Headings) is the NLM Controlled Vocabulary Thesaurus Used for Indexing Articles for PubMed. Available online at: http:// www.ncbi.nlm.nih.gov/mesh/ (Acessed November 1, 2015).
- Millet, G. Y., Tomazin, K., Verges, S., Vincent, C., Bonnefoy, R., Boisson, R. C., et al. (2011). Neuromuscular consequences of an extreme mountain ultramarathon. *PLoS ONE* 6:e17059. doi: 10.1371/journal.pone.0017059
- Mohamed, S., Lamya, N., and Hamda, M. (2016). Effect of maximal versus supra-maximal exhausting race on lipid peroxidation, antioxidant activity and muscle-damage biomarkers in long-distance and middle-distance runners. *Asian J. Sports Med.* 7:e27902. doi: 10.5812/asjsm.27902
- Mohlenkamp, S., Bose, D., Mahabadi, A. A., Heusch, G., and Erbel, R. (2007). On the paradox of exercise: coronary atherosclerosis in an apparently healthy marathon runner. *Nat. Clin. Pract. Cardiovasc. Med.* 4, 396–401. doi: 10.1038/ncpcardio0926
- Moldoveanu, A. I., Shephard, R. J., and Shek, P. N. (2000). Exercise elevates plasma levels but not gene expression of IL-1beta, IL-6, and TNF-alpha in blood mononuclear cells. *J. Appl. Physiol.* 4, 1499–1504.
- Moldoveanu, A. I., Shephard, R. J., and Shek, P. N. (2001). The cytokine response to physical activity and training. Sports Med. 31, 115–144. doi: 10.2165/00007256-200131020-00004

- Mrakic-Sposta, S., Gussoni, M., Moretti, S., Pratali, L., Giardini, G., Tacchini, P., et al. (2015). Effects of mountain ultra-marathon running on ros production and oxidative damage by micro-invasive analytic techniques. *PLoS ONE* 10:e0141780. doi: 10.1371/journal.pone.0141780
- Neidhart, M., Müller-Ladner, U., Frey, W., Bosserhoff, A. K., Colombani, P. C., Frey-Rindova, P., et al. (2000). Increased serum levels of noncollagenous matrix proteins (cartilage oligomeric matrix protein and melanoma inhibitory activity) in marathon runners. Osteoarthr. Cartil. 8, 222–229. doi: 10.1053/joca.1999.0293
- Ng, Q. Y., Lee, K. W., Byrne, C., Ho, T. F., and Lim, C. L. (2008). Plasma endotoxin and immune responses during a 21-km road race under a warm and humid environment. *Ann. Acad. Med. Singap.* 37, 307–314.
- Nickel, T., Emslander, I., Sisic, Z., David, R., Schmaderer, C., Marx, N., et al. (2012). Modulation of dendritic cells and toll-like receptors by marathon running. *Eur. J. Appl. Physiol.* 112, 1699–1708. doi: 10.1007/s00421-011-2140-8
- Nielsen, H. G., Øktedalen, O., Opstad, P. K., and Lyberg, T. (2016). Plasma cytokine profiles in long-term strenuous exercise. J. Sports Med. 2016;7186137. doi: 10.1155/2016/7186137
- Nieman, D. C. (2009). Immune function responses to ultramarathon race competition. Med. Sport. 13, 189–196. doi: 10.2478/v10036-009-0031-4
- Nieman, D. C., Dumke, C. I., Henson, D. A., McAnulty, S. R., McAnulty, L. S., Lind, R. H., et al. (2003). Immune and oxidative changes during and following the Western States Endurance Run. *Int. J. Sports Med.* 24, 541–547. doi: 10.1055/s-2003-42018
- Nieman, D. C., Dumke, C. L., Henson, D. A., McAnulty, S. R., Gross, S. J., and Lind, R. H. (2005). Muscle damage is linked to cytokine changes following a 160-km race. *Brain Behav. Immun.* 19, 398–403. doi: 10.1016/j.bbi.2005.03.008
- Nieman, D. C., Johanssen, L. M., and Lee, J. W. (1989). Infectious episodes in runners before and after a roadrace. J. Sports Med. Phys. Fitness 29, 289–296.
- Nieman, D. C., Johanssen, L. M., Lee, J. W., and Arabatzis, K. (1990). Infectious episodes in runners before and after the Los Angeles Marathon. J. Sports Med. Phys. Fitness 30, 316–328.
- Nieman, D. C., Meaney, M. P., John, C. S., Knagge, K. J., and Chen, H. (2016).
 9- and 13-Hydroxy-octadecadienoic acids (9+13 HODE) are inversely related to granulocyte colony stimulating factor and IL-6 in runners after 2 h running. *Brain Behav. Immun.* 56, 246–252. doi: 10.1016/j.bbi.2016.03.020
- Niess, A. M., Sommer, M., Schlotz, E., Northoff, H., Dickhuth, H. H., and Fehrenbach, E. (2000). Expression of the inducible nitric oxide synthase (iNOS) in human leukocytes: responses to running exercise. *Med. Sci. Sports Exerc.* 32, 220–1225. doi: 10.1097/00005768-200007000-00006
- O'Brien, M. (2012). The reciprocal relationship between inflammation and coagulation. *Top. Companion Anim. Med.* 27, 46–52. doi: 10.1053/j.tcam.2012. 06.003
- O'Keefe, J. H., and Lavie, C. J. (2013). Run for your life... at a comfortable speed and not too far. *Heart* 99, 516–519. doi: 10.1136/heartjnl-2012-302886
- O'Keefe, J. H., Vogel, R., Lavie, C. J., and Cordain, L. (2010). Achieving huntergatherer fitness in the 21(st) century: back to the future. Am. J. Med. 123, 1082–1086. doi: 10.1016/j.amjmed.2010.04.026
- Papassotiriou, I., Alexiou, V. G., Tsironi, M., Skenderi, K., Spanos, A., and Falagas, M. E. (2008). Severe aseptic inflammation caused by long distance running (246 km) does not increase procalcitonin. *Eur. J. Clin. Invest.* 38, 276–279. doi: 10.1111/j.1365-2362.2008.01935.x
- Pedersen, B. K. (2011). Muscles and their myokines. J. Exp. Biol. 214, 337–346. doi: 10.1242/jeb.048074
- Pedersen, B. K. (2013). Muscle as a secretory organ. Compr. Physiol. 3, 1337–1362. doi: 10.1002/cphy.c120033
- Pedersen, B. K., and Febbraio, M. A. (2012). Muscles, exercise and obesity: skeletal muscle as a secretory organ. *Nat. Rev. Endocrinol.* 8, 457–465. doi: 10.1038/nrendo.2012.49
- Peters, E. M., and Bateman, E. D. (1983). Ultramarathon running and upper respiratory tract infections. An epidemiological survey. S. Afr. Med. J. 64, 582–584.
- Peters, E. M., Robson, P. J., Kleinveldt, N. C., Naicker, V. L., and Jogessar, V. D. (2004). Hematological recovery in male ultramarathon runners: the effect of variations in training load and running time. *J. Sports Med. Phys. Fitness* 44, 315–321.
- Petersen, A. M., and Pedersen, B. K. (2005). The anti-inflammatory effect of exercise. J. Appl. Physiol. 98, 154–1162. doi: 10.1152/japplphysiol.00164.2004

- Rehm, K. E., Elci, O. U., Hahn, K., and Marshall, G. D. Jr. (2013). The impact of self-reported psychological stress levels on changes to peripheral blood immune biomarkers in recreational marathon runners during training and recovery. *Neuroimmunomodulation* 20, 64–176. doi: 10.1159/000346795
- Reihmane, D., Jurka, A., Tretjakovs, P., and Dela, F. (2013). Increase in IL-6, TNF-alpha, and MMP-9, but not sICAM-1, concentrations depends on exercise duration. *Eur. J. Appl. Physiol.* 113, 851–858. doi: 10.1007/s00421-012-2491-9
- Ridker, P. M., Rifai, N., Rose, L., Buring, J. E., and Cook, N. R. (2002). Comparison of C-reactive protein and low-density lipoprotein cholesterol levels in the prediction of first cardiovascular events. *N. Engl. J. Med.* 347, 1557–1565. doi: 10.1056/NEJMoa021993
- Sampson, M., McGowan, J., Cogo, E., Grimshaw, J., Moher, D., and Lefebvre, C. (2009). An evidence-based practice guideline for the peer review of electronic search strategies. J. Clin. Epidemiol. 62, 944–952. doi:10.1016/j.jclinepi.2008.10.012
- Sampson, M., McGowan, J., Tetzlaff, J., Cogo, E., and Moher, D. (2008). No consensus exists on search reporting methods for systematic reviews. J. Clin. Epidemiol. 61, 748–754. doi: 10.1016/j.jclinepi.2007.10.009
- Santos, R. V., Tufik, S., and De Mello, M. T. (2007). Exercise, sleep and cytokines: is there a relation? Sleep Med. Rev. 11, 231–239. doi: 10.1016/j.smrv.2007.03.003
- Santos, V. C., Levada-Pires, A. C., Alves, S. R., Pithon-Curi, T. C., Curi, R., and Cury-Boaventura, M. F. (2013). Changes in lymphocyte and neutrophil function induced by a marathon race. *Cell Biochem. Funct.* 31, 237–243. doi: 10.1002/cbf.2877
- Saravia, S. G., Knebel, F., Schroeckh, S., Ziebig, R., Lun, A., Weimann, A., et al. (2010). Cardiac troponin T release and inflammation demonstrated in marathon runners. *Clin. Lab.* 56, 51–58.
- Saugy, J., Place, N., Millet, G. Y., Degache, F., Schena, F., and Millet, G. P. (2013). Alterations of neuromuscular function after the world's most challenging mountain ultra-marathon. *PLoS ONE* 8:e65596. doi: 10.1371/ journal.pone.0065596
- Schobersberger, W., Hobisch-Hagen, P., Fries, D., Wiedermann, F., Rieder-Scharinger, J., Villiger, B., et al. (2000). Increase in immune activation, vascular endothelial growth factor and erythropoietin after an ultramarathon run at moderate altitude. *Immunobiology* 201, 611–620. doi: 10.1016/S0171-2985(00)80078-9
- Scott, J. P., Sale, C., Greeves, J. P., Casey, A., Dutton, J., and Fraser, W. D. (2011). Effect of exercise intensity on the cytokine response to an acute bout of running. *Med. Sci. Sports Exerc.* 43, 2297–2306. doi: 10.1249/MSS.0b013e3182 2113a9
- Shin, Y. O., and Lee, J. B. (2013). Leukocyte chemotactic cytokine and leukocyte subset responses during ultra-marathon running. *Cytokine* 61, 364–369. doi: 10.1016/j.cyto.2012.11.019
- Siegel, A. J., Verbalis, J. G., Clement, S., Mendelson, J. H., Mello, N. K., Adner, M., et al. (2007). Hyponatremia in marathon runners due to inappropriate arginine vasopressin secretion. *Am. J. Med.* 120, 461.e11–461.e17. doi: 10.1016/j.amjmed.2006.10.027
- Smith, L. L. (2004). Tissue trauma: the underlying cause of overtraining syndrome? J. Strength Cond. Res. 18, 185–193. doi: 10.1519/00124278-200402000-00028
- Steinacker, J. M., Lormes, W., Reissnecker, S., and Liu, Y. (2004). New aspects of the hormone and cytokine response to training. *Eur. J. Appl. Physiol.* 91, 382–391. doi: 10.1007/s00421-003-0960-x
- Stirling, A. E., and Kerr, G. A. (2006). Perfectionism and mood states among recreational and elite athletes. J. Sport Psychol. 8, 13–27.
- Stuempfle, K. J., Valentino, T., Hew-Butler, T., Hecht, F. M., and Hoffman, M. D. (2016). Nausea is associated with endotoxemia during a 161-km ultramarathon. *J. Sports Sci.* 34, 1662–1668. doi: 10.1080/02640414.2015.1130238
- Suzuki, K., Nakaji, S., Yamada, M., Liu, Q., Kurakake, S., Okamura, N., et al. (2003). Impact of a competitive marathon race on systemic cytokine and neutrophil responses. *Med. Sci. Sports Exerc.* 35, 348–355. doi: 10.1249/01.MSS.0000048861.57899.04
- Suzuki, K., Yamada, M., Kurakake, S., Okamura, N., Yamaya, K., Liu, Q., et al. (2000). Circulating cytokines and hormones with immunosuppressive but neutrophil-priming potentials rise after endurance exercise in humans. *Eur. J. Appl. Physiol.* 81, 281–287. doi: 10.1007/s004210050044
- Taylor, B. A., Zaleski, A. L., Capizzi, J. A., Ballard, K. D., Troyanos, C., Baggish, A. L., et al. (2014). Influence of chronic exercise on carotid atherosclerosis

in marathon runners. Br. Med. J. Open 4:e004498. doi: 10.1136/bmjopen-2013-004498

- Tomaszewski, M., Charchar, F. J., Przybycin, M., Crawford, L., Wallace, A. M., Gosek, K., et al. (2003). Strikingly low circulating CRP concentrations in ultramarathon runners independent of markers of adiposity: how low can you go? *Arterioscler. Thromb. Vasc. Biol.* 23, 1640–1644. doi: 10.1161/01.ATV.0000087036.75849.0B
- Uchakin, P. N., Gotovtseva, E. P., and Stray-Gundersen, J. (2003). Immune and neurodocrine alterations in marathon runners. J. Appl. Res. 3, 483–494.
- Vaisberg, M., Bachi, A. L., Latrilha, C., Dioguardi, G. S., Bydlowski, S. P., and Maranhao, R. C. (2012). Lipid transfer to HDL is higher in marathon runners than in sedentary subjects, but is acutely inhibited during the run. *Lipids* 47, 679–686. doi: 10.1007/s11745-012-3685-y
- Vaisberg, M., Suguri, V. M., Gregorio, L. C., Lopes, J. D., and Bachi, A. L. (2013). Cytokine kinetics in nasal mucosa and sera: new insights in understanding upper-airway disease of marathon runners. *Exerc. Immunol. Rev.* 19, 49–59.
- Vernillo, G., Giandolini, M., Edwards, W. B., Morin, J. B., Samozino, P., Horvais, N., et al. (2017). "Biomechanics and physiology of uphill and downhill running." *Sports Med.* 47, 615–629. doi: 10.1007/s40279-016-0605-y
- Videbaek, S., Bueno, A. M., Nielsen, R. O., and Rasmussen, S. (2015). Incidence of running-related injuries per 1000 h of running in different types of runners: a systematic review and meta-analysis. *Sports Med.* 45, 1017–1026. doi: 10.1007/s40279-015-0333-8

- Walsh, N. P., Gleeson, M., Shephard, R. J., Woods, J. A., Bishop, N. C., Fleshner, M., et al. (2011). Position statement. Part one: immune function and exercise. *Exerc. Immunol. Rev.* 17, 6–63.
- Waśkiewicz, Z., Kłapcinska, B., Sadowska-Krepa, E., Czuba, M., Kempa, K., Kimsa, E., et al. (2012). Acute metabolic responses to a 24-h ultra-marathon race in male amateur runners. *Eur. J. Appl. Physiol.* 112, 1679–1688. doi: 10.1007/s00421-011-2135-5
- Wilhelm, M., Zueger, T., De Marchi, S., Rimoldi, S. F., Brugger, N., Steiner, R., et al. (2014). Inflammation and atrial remodeling after a mountain marathon. *Scand. J. Med. Sci. Sports* 24, 519–525. doi: 10.1111/sms.12030
- Zaldivar, F., Wang-Rodriguez, J., Nemet, D., Schwindt, C., Galassetti, P., Mills, P. J., et al. (2006). Constitutive pro- and anti-inflammatory cytokine and growth factor response to exercise in leukocytes. J. Appl. Physiol. 100, 1124–1133. doi: 10.1152/japplphysiol.00562.2005

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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