



Transceptors at the boundary of nutrient transporters and receptors: a new role for *Arabidopsis* SULTR1;2 in sulfur sensing

Zhi-Liang Zheng^{1,2*}, Bo Zhang¹ and Thomas Leustek^{3*}

¹ Plant Nutrient Signaling and Fruit Quality Improvement Laboratory, Citrus Research Institute, Southwest University, Chongqing, China

² Department of Biological Sciences, Lehman College, City University of New York, Bronx, NY, USA

³ Department of Plant Biology and Pathology, Rutgers University, New Brunswick, NJ, USA

Edited by:

Stanislav Kopriva, University of Cologne, Germany

Reviewed by:

Markus Wirtz, Centre for Organismal Studies, Germany
Cornelia Herschbach, Albert-Ludwigs-Universität Freiburg, Germany
Peter Buchner, Rothamsted Research, UK

*Correspondence:

Zhi-Liang Zheng, Department of Biological Sciences, Lehman College, City University of New York, 250 Bedford Park Boulevard West, Bronx, NY 10468, USA
e-mail: zhiliang.zheng@lehman.cuny.edu;

Thomas Leustek, Department of Plant Biology and Pathology, Rutgers University, 59 Dudley Road, New Brunswick, NJ 08901, USA
e-mail: leustek@aesop.rutgers.edu

Plants have evolved a sophisticated mechanism to sense the extracellular sulfur (S) status so that sulfate transport and S assimilation/metabolism can be coordinated. Genetic, biochemical, and molecular studies in *Arabidopsis* over the past 10 years have started to shed some light on the regulatory mechanism of the S response. Key advances in transcriptional regulation (SLIM1, MYB, and miR395), involvement of hormones (auxin, cytokinin, and abscisic acid) and identification of putative sensors (OASTL and SULTR1;2) are highlighted here. Although our current view of S nutrient sensing and signaling remains fragmented, it is anticipated that through further studies a sensing and signaling network will be revealed in the near future.

Keywords: sulfate, SULTR1;2, transporter, sensor, transceptor

TIGHTLY REGULATED SULFATE UPTAKE IS REQUIRED FOR SULFUR ASSIMILATION AND UTILIZATION

Plants have evolved a biosynthetic pathway to assimilate sulfate (SO_4^{2-}), a primary source of the essential nutrient sulfur (S), into Cys and Met, which are then used for synthesis of proteins and various S-containing compounds including glucosinolates and glutathione (GSH; Takahashi et al., 2011). SO_4^{2-} is taken up from the rhizosphere by roots and is subsequently translocated into shoots. Therefore, SO_4^{2-} transport and assimilation must be tightly coordinated to meet the dynamic demand for S. SO_4^{2-} uptake and translocation is mediated by transporters (SULTR) with specific gene products performing distinct and also overlapping functions (Gigolashvili and Kopriva, 2014). In *Arabidopsis*, two members of group 1 (SULTR1;1 and SULTR1;2) are high affinity SO_4^{2-} transporters and mediate SO_4^{2-} uptake into roots (Gigolashvili and Kopriva, 2014). Several members of Groups 2 and 3 are likely involved in SO_4^{2-} translocation from roots to shoots, while Group 4 (SULTR4;1 and SULTR4;2) functions in vacuolar export of SO_4^{2-} (Takahashi et al., 2011; Gigolashvili and Kopriva, 2014). In response to S deficiency, many of the transporter genes are transcriptionally up-regulated. The two most

studied transporters, SULTR1;1 and SULTR1;2, have been shown to act redundantly in controlling SO_4^{2-} uptake from roots, with SULTR1;2 having a major role (Takahashi et al., 2011; Gigolashvili and Kopriva, 2014). This tightly regulated transport system is critical for plant response and adaptation to the dynamically changing S nutrient environment.

KNOWLEDGE OF S SENSING AND SIGNALING REMAINS FRAGMENTED

To understand the regulatory mechanism of S sensing, transport and signaling, significant efforts have been made and exciting progress is summarized below.

INSIGHTS INTO TRANSCRIPTIONAL CONTROL IN S DEFICIENCY RESPONSE

Several transcriptome profiling studies reported that more than 1500 genes in *Arabidopsis* are up- or down-regulated by S deficiency (Hirai et al., 2003, 2004; Maruyama-Nakashita et al., 2003, 2006). These studies confirmed up-regulation of SULTR1;2 and other transporter genes, and led to the identification of two novel S-responsive genes, *BGLU28* and *SD11*, which have

received considerable attentions. *BGLU28* is the most strongly up-regulated gene in several of the studies and is hypothesized to act by releasing S from glucosinolate, which is potentially a major S storage compound in the vacuole (Maruyama-Nakashita et al., 2003, 2006; Dan et al., 2007). *SDII* is annotated as a protein similar to male sterility family protein MS5 and recent evidence suggests that its expression level can be used as a biosensor of S nutrient status (Howarth et al., 2009). Interestingly, a *cis* element has been identified called SURE that is necessary for S-deficiency control including transcriptional regulation of *BLGU28* (Maruyama-Nakashita et al., 2005). Furthermore, transcriptional regulators have been identified. The *SLIM1* mutants lack the ability to up-regulate S-response gene expression including that of *SULTR1;2* (Maruyama-Nakashita et al., 2006). Although many of S-responsive genes (including *BGLU28* and *SDII*) are under *SLIM1* control, others (e.g., *APR2* and *APR3*) were not affected, strongly suggesting that although *SLIM1* may be a major S-response transcription factor, additional transcriptional regulators are also involved. Consistent with this, several MYB transcription factors, in particular *MYB28* and *MYB29* which are transcriptionally repressed by S-deficiency, have been shown critical for transcriptional regulation of genes for the biosynthesis of glucosinolate which potentially serves as a critical S storage compound (Yatusevich et al., 2010). Recently, a microRNA gene (miR395) was shown to be important for regulating several target genes involved in S-deficiency response including *SULTR2;1/AST68* and *APS4* (Kawashima et al., 2009, 2011). Interestingly, miR395 was shown to be controlled by *SLIM1* (Kawashima et al., 2011). Taken together, these studies provided an important foundation for understanding the transcriptional events in the nucleus.

EMERGING UNDERSTANDING OF THE ROLE OF PROTEIN PHOSPHORYLATION, DEGRADATION, AND HORMONES IN S DEFICIENCY RESPONSE

SULTR1;2 was shown to be regulated posttranscriptionally (Yoshimoto et al., 2007). The effects of inhibitors of protein kinase and proteasome have indicated that protein phosphorylation (Maruyama-Nakashita et al., 2004a) and degradation (Pootakham et al., 2010) are involved in regulating S transport and S-starvation response in *Arabidopsis* and *Chlamydomonas*, respectively. On the other hand, the role of hormones has been increasingly recognized as a key factor in S response. Based on surveys for the impact of several hormones on the S deficiency-activated expression of beta-conglycinin (Ohkama et al., 2002), *SULTR1;2* (Maruyama-Nakashita et al., 2004b), and *BGLU28* (Dan et al., 2007), it seems that auxin, cytokinin, and abscisic acid (ABA) are involved in negatively regulating S deficiency response. Cytokinin seems to have a broader effect in S response as all of the above three S response genes could be suppressed by exogenous application of this hormone. Furthermore, genetic evidence using a cytokinin receptor mutant *cre1* demonstrated the negative regulatory role of cytokine on S uptake (Maruyama-Nakashita et al., 2004b). The negative regulatory role of ABA was first implicated by the observed suppression by S deficiency of an ABA response marker *RD29B:GUS* and down-regulation of *BGLU28* by externally applied ABA (Dan et al., 2007). A role for ABA biosynthesis in S response was recently reported (Cao et al., 2014). Compared

to ABA and cytokinin, the role of auxin in S response has received more attentions. Auxin was first implicated as a regulator of S deficiency response by the observed up-regulation of auxin-inducible genes (such as *IAA28*) and *NITs* (likely involved in auxin synthesis) under S deficiency (Nikiforova et al., 2003), although S deficiency did not significantly alter auxin level (Kutz et al., 2002). However, evidence obtained from the use of *DR5:GUS*, an auxin response marker, suggests that S deficiency inhibits auxin accumulation or response (Dan et al., 2007). Such an inhibitory effect of auxin biosynthesis was confirmed recently (Zhao et al., 2014). Furthermore, by applying auxin externally, the S deficiency-activated *BGLU28* expression is down-regulated. The role of auxin response regulators such as *IAA28* and *ARF2* in controlling expression of S metabolism genes has been implicated using a transgenic approach (Falkenberg et al., 2008), and a definite role of auxin was demonstrated by two genetic studies. An auxin signaling component called *AXR1*, which is a component of the 26S proteasome, was shown to be involved in the S deficiency response (Dan et al., 2007), in agreement with the subsequently reported role of protein degradation in *Chlamydomonas* S response (Pootakham et al., 2010). Another S response mutant is allelic to *BIG* (a calossin-like protein involved in polar auxin transport), indicating a role for auxin transport as well as auxin biosynthesis or response in S signaling (Kasajima et al., 2007).

Most interestingly, putative S sensors or sensing components have been reported. Cys homeostasis is tightly controlled by the Cys synthase complex which consists of Ser acetyltransferase (SAT, the enzyme producing the substrate for Cys biosynthesis) and O-acetylserine (thiol) lyase (OASTL, the enzyme producing L-Cys; Yi et al., 2010). *Arabidopsis* OASTL has three isoforms, OASTL-A1, OASTL-B, and OASTL-C, which are located in the cytosol, plastids, and mitochondria, respectively. OASTL-A1, the most abundant isoform, has been demonstrated *in vitro* to specifically interact with the STAS domain of *SULTR1;2* (Shibagaki and Grossman, 2010). Interestingly, this interaction may be physiologically relevant as demonstrated in a heterologous yeast system. The interaction could enhance OASTL-A1 Cys synthesis activity at the same time it inhibits *SULTR1;2* transport activity. This reciprocal activity regulation has led to the proposal that OASTL-A1 is involved in sensing of S status (Shibagaki and Grossman, 2010). OASTL-C has also been reported to act in Cys sensing (Wirtz et al., 2012). The questions remain whether these two differentially localized OASTL members sense Cys or SO_4^{2-} located in different compartments and how they act to sense S status.

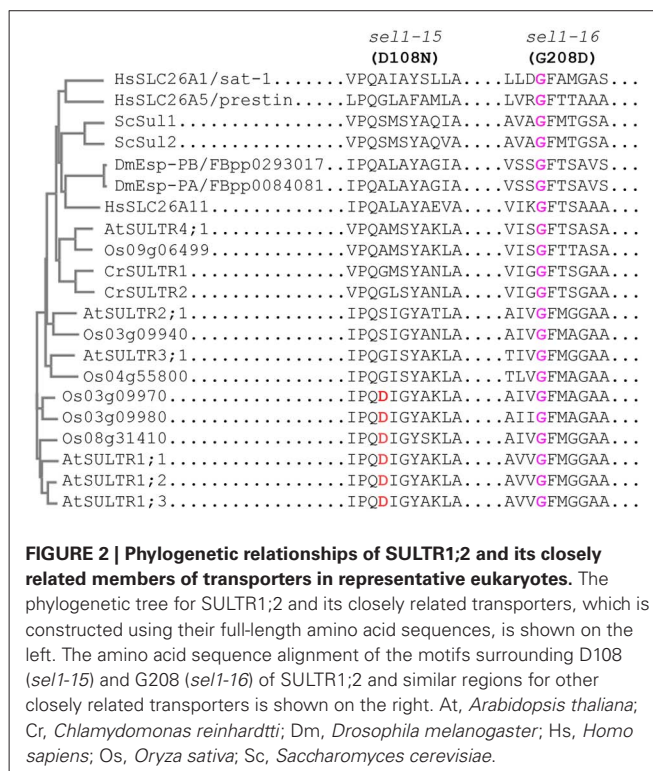
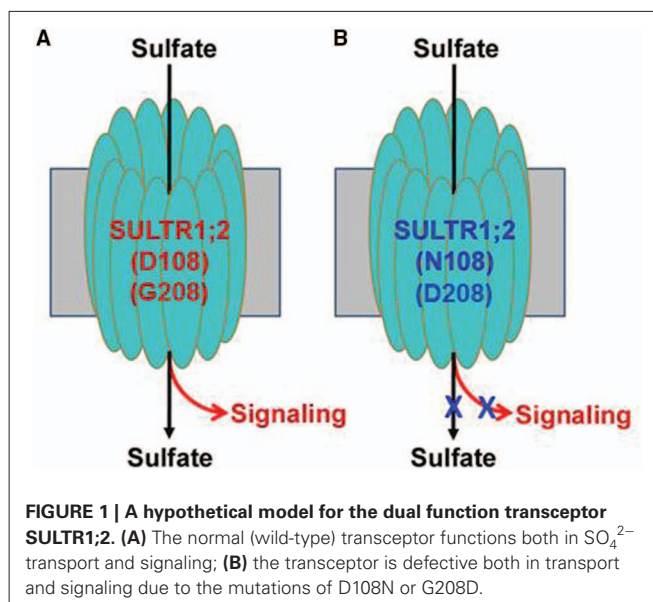
Most recent genetic and physiological evidence obtained from our groups have shown that besides its high affinity transport function, *SULTR1;2* has a novel regulatory function (Zhang et al., 2014). Using *BGLU28* promoter:GUS as a mutant screening tool, two novel alleles of *SULTR1;2* were isolated that exhibit high GUS activity even under sufficient S conditions: *sel1-15* (D108N) and *sel1-16* (G208D). These two mutations lie in the predicted transmembrane (TM) helices TM11 and TM5. In contrast to all prior studies in which up-regulation of S response genes in *sel1* mutants were interpreted as the result of compromised SO_4^{2-} uptake and consequently lower accumulation of internal SO_4^{2-} or its metabolites (Shibagaki et al., 2002; Maruyama-Nakashita

et al., 2003; El Kassis et al., 2007), we have provided two lines of convincing physiological evidence that support the hypothesis that up-regulation of *BGLU28* and three other genes (*SULTR4;2*, *SDI1*, and *LSU1*) could be independent of the compromised SO_4^{2-} uptake and internal S status of the mutants (Zhang et al., 2014). First, under high concentration of SO_4^{2-} (10 mM) which did not lead to a difference in internal SO_4^{2-} concentration and GSH level, *sel1-15/16* and a null allele (*sel1-18*) still had higher gene expression level than their wild-type (WT) backgrounds. Second, treatments with 1 mM Cys or 1 mM GSH in the SO_4^{2-} deficiency medium (which did not lead to any difference in Cys uptake and/or internal GSH contents between the *sel1* alleles and WT) also led to higher gene expression level in *sel1-15/16/18*. These results strongly suggest that the *sel1* seedlings (in particular the expression in roots) grown under sufficient S behave as if they have been treated by certain degrees of S deficiency. In other words, the mutations in *SULTR1;2* reduce sensitivity to the S-induced suppression of S response genes. The evidence points toward a novel function for *SULTR1;2* in regulating S nutrient response besides its transport function. The possibility that *SULTR1;2* acts as an S sensor is discussed in the next section.

CAN SULTR1;2 ACT AS A PUTATIVE PM-LOCALIZED SULFATE TRANSPORTING RECEPTOR?

Dual function transporters, like *SULTR1;2* described above, are not unusual. Studies in yeast and animal nutrient transport and sensing have revealed the existence of classic receptors (which are not involved in transport, e.g., G-protein-coupled receptor Gpr1), transceptors (which are either transporting receptors, e.g., Gap1, or non-transporting receptors, e.g., Snf3) and the majority of common transporters (which do not have a sensing function; Thevelein and Voordeckers, 2009). Therefore, transceptors can be considered at the boundary between receptors and transporters. In general, to demonstrate a receptor function for a transporter molecule, genetic or pharmacological evidence is required that shows decoupling of nutrient transport and signaling, i.e., the signaling output is independent of transport.

In the case of *SULTR1;2*, the mutations in TM1 (*sel1-15*) or TM5 (*sel1-16*) could abolish both SO_4^{2-} transport and signaling (as measured by expression of S response genes), but the defect in signaling could be independent of SO_4^{2-} transport and accumulation (Zhang et al., 2014). Because of this, we propose that *SULTR1;2* can function as a putative SO_4^{2-} transceptor (Figure 1). Although *SULTR1;2* cannot be the only S-sensor since the *sel1-15/16* mutants show reduced sensitivity to S but does not entirely abolish the S-limitation response, this finding provides a first intriguing insight into S-sensing in plants given its PM location where extracellular SO_4^{2-} is first in contact with the PM-localized sensors. Note that a dual-affinity nitrate transporter called *NRT1.1* has been demonstrated to act as a nitrate sensor (Ho et al., 2009; Bouguyon et al., 2012), and thus using nutrient transporters to sense the external nutrient status may be evolutionally conserved and advantageous to plants. Indeed, a phosphate transceptor (Pho84) has been reported in yeast (Popova et al., 2010). More encouraging is that in yeast SO_4^{2-} transporters *Sul1/2* have also been described as being transceptors (Conrad et al., 2014). To gain further insights into the



evolutionarily conserved mechanism of using sulfate transporters as sensors, we performed a sequence alignment using transporters from *Arabidopsis*, rice, *Chlamydomonas*, yeast, *Drosophila* and humans that are most closely related to *Arabidopsis* *SULTR1;2*. The result (Figure 2) shows that while D108 is only specific to *SULTR1* group in *Arabidopsis* and rice, G208 is highly conserved in all transporters. It will be interesting to determine whether G208 is critical for SO_4^{2-} transport and signaling in many eukaryotes.

It remains unclear how plants use SULTR1;2 to sense external SO_4^{2-} status and adopt a high or low affinity transport system in response to dynamic S environment. However, studies from the yeast amino acid transceptor Gap1 or *Arabidopsis* nitrate transceptor NRT1.1 may provide some hints for the SULTR1;2-mediated sensing mechanism. In NRT1.1-mediated nitrate sensing and signaling, auxin transport and NRT1.1 phosphorylation have been shown to be critical (Ho et al., 2009; Bouguyon et al., 2012). In yeast, Gap1 uses the same sites for amino acid binding/transport and signaling (Van Zeebroeck et al., 2009; Conrad et al., 2014). Once amino acid is bound to Gap1, it triggers a conformational change in Gap1 that subsequently allows the amino acid to be transported into the cytoplasm and in the same time a signaling cascade is activated. If the amino acid status is perceived to be sufficient, Gap1 undergoes a rapid endocytic process that removes it from the PM and sorts it for degradation.

FUTURE PROSPECT

Exciting findings in the past 10 years have led to the identification of several components from the PM to the cytoplasm and to the nucleus that are involved in S sensing, transport and downstream response. Several outstanding questions remained to be answered. What is the SULTR1;2 topology and does SULTR1;2 have separate sensing and transport domains? Can SULTR1;2 interact with OASTL *in vivo* (if so, which OASTL isoform?) and exert the effect of S sensing in *planta*? How does SULTR1;2 link to various signaling intermediates acting at the PM, the cytoplasm or the nucleus? Are there additional partners that may form a larger SULTR1;2-based S sensing complex? If such complex cannot account for all S responses, what other sensors are involved? Further, what are the roles of these sensing components in local and systemic S signaling (Hubberten et al., 2012)? Although our current view of S sensing and signaling remains fragmented, further studies into these questions will allow us to piece together individual components and ultimately construct the SULTR1;2-mediated S sensing and signaling pathway or network.

ACKNOWLEDGMENTS

Research in our groups was initially funded in part by an NIH grant 3S06GM008225-20S1 (to Zhi-Liang Zheng) and a USDA HATCH grant (to Thomas Leustek) and is currently by a collaborative NSF grant (IOS-1121521 to Thomas Leustek and IOS-1121551 to Zhi-Liang Zheng). We apologize for not being able to cite many original or review articles due to the space limitation. We appreciate the reviewers comments.

REFERENCES

- Bouguyon, E., Gojon, A., and Nacry, P. (2012). Nitrate sensing and signaling in plants. *Semin. Cell Dev. Biol.* 23, 648–654. doi: 10.1016/j.semcdb.2012.01.004
- Cao, M. J., Wang, Z., Zhao, Q., Mao, J. L., Speiser, A., Wirtz, M., et al. (2014). Sulfate availability affects ABA levels and germination response to ABA and salt stress in *Arabidopsis thaliana*. *Plant J.* 77, 604–615. doi: 10.1111/tpj.12407
- Conrad, M., Schothorst, J., Kankipati, H. N., Van Zeebroeck, G., Rubio-Teixeira, M., and Thevelein, J. M. (2014). Nutrient sensing and signaling in the yeast *Saccharomyces cerevisiae*. *FEMS Microbiol. Rev.* 38, 254–299. doi: 10.1111/1574-6976.12065
- Dan, H., Yang, G., and Zheng, Z. L. (2007). A negative regulatory role for auxin in sulphate deficiency response in *Arabidopsis thaliana*. *Plant Mol. Biol.* 63, 221–235. doi: 10.1007/s11103-006-9084-0
- El Kassis, E., Cathala, N., Rouached, H., Fourcroy, P., Berthomieu, P., Terry, N., et al. (2007). Characterization of a selenate-resistant *Arabidopsis* mutant. Root growth as a potential target for selenate toxicity. *Plant Physiol.* 143, 1231–1241. doi: 10.1104/pp.106.091462
- Falkenberg, B., Witt, I., Zanon, M. I., Steinhäuser, D., Mueller-Roeber, B., Hesse, H., et al. (2008). Transcription factors relevant to auxin signalling coordinate broad-spectrum metabolic shifts including sulphur metabolism. *J. Exp. Bot.* 59, 2831–2846. doi: 10.1093/jxb/ern144
- Gigolashvili, T., and Kopriva, S. (2014). Transporters in plant sulfur metabolism. *Front. Plant Sci.* 5:442. doi: 10.3389/fpls.2014.00442
- Hirai, M. Y., Fujiwara, T., Awazuha, M., Kimura, T., Noji, M., and Saito, K. (2003). Global expression profiling of sulfur-starved *Arabidopsis* by DNA microarray reveals the role of O-acetyl-L-serine as a general regulator of gene expression in response to sulfur nutrition. *Plant J.* 33, 651–663. doi: 10.1046/j.1365-313X.2003.01658.x
- Hirai, M. Y., Yano, M., Goodenow, D. B., Kanaya, S., Kimura, T., Awazuha, M., et al. (2004). Integration of transcriptomics and metabolomics for understanding of global responses to nutritional stresses in *Arabidopsis thaliana*. *Proc. Natl. Acad. Sci. U.S.A.* 101, 10205–10210. doi: 10.1073/pnas.0403218101
- Ho, C. H., Lin, S. H., Hu, H. C., and Tsay, Y. F. (2009). CHL1 functions as a nitrate sensor in plants. *Cell* 138, 1184–1194. doi: 10.1016/j.cell.2009.07.004
- Howarth, J. R., Parmar, S., Barraclough, P. B., and Hawkesford, M. J. (2009). A sulphur deficiency-induced gene, *sdi1*, involved in the utilization of stored sulphate pools under sulphur-limiting conditions has potential as a diagnostic indicator of sulphur nutritional status. *Plant Biotechnol. J.* 7, 200–209. doi: 10.1111/j.1467-7652.2008.00391.x
- Hubberten, H. M., Drozd, A., Tran, B. V., Hesse, H., and Hoefgen, R. (2012). Local and systemic regulation of sulfur homeostasis in roots of *Arabidopsis thaliana*. *Plant J.* 72, 625–635. doi: 10.1111/j.1365-313X.2012.05105.x
- Kasajima, I., Ohkama-Ohtsu, N., Ide, Y., Hayashi, H., Yoneyama, T., Suzuki, Y., et al. (2007). The BIG gene is involved in regulation of sulfur deficiency-response genes in *Arabidopsis thaliana*. *Physiol. Plant.* 129, 351–363. doi: 10.1111/j.1399-3054.2006.00814.x
- Kawashima, C. G., Matthewman, C. A., Huang, S., Lee, B. R., Yoshimoto, N., Koprivova, A., et al. (2011). Interplay of SLIM1 and miR395 in the regulation of sulfate assimilation in *Arabidopsis*. *Plant J.* 66, 863–876. doi: 10.1111/j.1365-313X.2011.04547.x
- Kawashima, C. G., Yoshimoto, N., Maruyama-Nakashita, A., Tsuchiya, Y. N., Saito, K., Takahashi, H., et al. (2009). Sulphur starvation induces the expression of microRNA-395 and one of its target genes but in different cell types. *Plant J.* 57, 313–321. doi: 10.1111/j.1365-313X.2008.03690.x
- Kutz, A., Muller, A., Hennig, P., Kaiser, W. M., Piotrowski, M., and Weiler, E. W. (2002). A role for nitrilase 3 in the regulation of root morphology in sulphur-starving *Arabidopsis thaliana*. *Plant J.* 30, 95–106. doi: 10.1046/j.1365-313X.2002.01271.x
- Maruyama-Nakashita, A., Inoue, E., Watanabe-Takahashi, A., Yamaya, T., and Takahashi, H. (2003). Transcriptome profiling of sulfur-responsive genes in *Arabidopsis* reveals global effects of sulfur nutrition on multiple metabolic pathways. *Plant Physiol.* 132, 597–605. doi: 10.1104/pp.102.019802
- Maruyama-Nakashita, A., Nakamura, Y., Tohge, T., Saito, K., and Takahashi, H. (2006). *Arabidopsis* SLIM1 is a central transcriptional regulator of plant sulfur response and metabolism. *Plant Cell* 18, 3235–3251. doi: 10.1105/tpc.106.046458
- Maruyama-Nakashita, A., Nakamura, Y., Watanabe-Takahashi, A., Inoue, E., Yamaya, T., and Takahashi, H. (2005). Identification of a novel cis-acting element conferring sulfur deficiency response in *Arabidopsis* roots. *Plant J.* 42, 305–314. doi: 10.1111/j.1365-313X.2005.02363.x
- Maruyama-Nakashita, A., Nakamura, Y., Watanabe-Takahashi, A., Yamaya, T., and Takahashi, H. (2004a). Induction of SULTR1;1 sulfate transporter in *Arabidopsis* roots involves protein phosphorylation/dephosphorylation circuit for transcriptional regulation. *Plant Cell Physiol.* 45, 340–345. doi: 10.1093/pcp/pch029
- Maruyama-Nakashita, A., Nakamura, Y., Yamaya, T., and Takahashi, H. (2004b). A novel regulatory pathway of sulfate uptake in *Arabidopsis* roots: implication of CRE1/WOL/AHK4-mediated cytokinin-dependent regulation. *Plant J.* 38, 779–789. doi: 10.1111/j.1365-313X.2004.02079.x
- Nikiforova, V., Freitag, J., Kempa, S., Adamik, M., Hesse, H., and Hoefgen, R. (2003). Transcriptome analysis of sulfur depletion in *Arabidopsis thaliana*:

- interlacing of biosynthetic pathways provides response specificity. *Plant J.* 33, 633–650. doi: 10.1046/j.1365-313X.2003.01657.x
- Ohkama, N., Takei, K., Sakakibara, H., Hayashi, H., Yoneyama, T., and Fujiwara, T. (2002). Regulation of sulfur-responsive gene expression by exogenously applied cytokinins in *Arabidopsis thaliana*. *Plant Cell Physiol.* 43, 1493–1501. doi: 10.1093/pcp/pcf183
- Pootakham, W., Gonzalez-Ballester, D., and Grossman, A. R. (2010). Identification and regulation of plasma membrane sulfate transporters in *Chlamydomonas*. *Plant Physiol.* 153, 1653–1668. doi: 10.1104/pp.110.157875
- Popova, Y., Thayumanavan, P., Lonati, E., Agrochao, M., and Thevelein, J. M. (2010). Transport and signaling through the phosphate-binding site of the yeast Pho84 phosphate transceptor. *Proc. Natl. Acad. Sci. U.S.A.* 107, 2890–2895. doi: 10.1073/pnas.0906546107
- Shibagaki, N., and Grossman, A. R. (2010). Binding of cysteine synthase to the STAS domain of sulfate transporter and its regulatory consequences. *J. Biol. Chem.* 285, 25094–25102. doi: 10.1074/jbc.M110.126888
- Shibagaki, N., Rose, A., McDermott, J. P., Fujiwara, T., Hayashi, H., Yoneyama, T., et al. (2002). Selenate-resistant mutants of *Arabidopsis thaliana* identify Sultr1;2, a sulfate transporter required for efficient transport of sulfate into roots. *Plant J.* 29, 475–486. doi: 10.1046/j.0960-7412.2001.01232.x
- Takahashi, H., Kopriva, S., Giordano, M., Saito, K., and Hell, R. (2011). Sulfur assimilation in photosynthetic organisms: molecular functions and regulations of transporters and assimilatory enzymes. *Annu. Rev. Plant Biol.* 62, 157–184. doi: 10.1146/annurev-arplant-042110-103921
- Thevelein, J. M., and Voordeckers, K. (2009). Functioning and evolutionary significance of nutrient transceptors. *Mol. Biol. Evol.* 26, 2407–2414. doi: 10.1093/molbev/msp168
- Van Zeebroeck, G., Bonini, B. M., Versele, M., and Thevelein, J. M. (2009). Transport and signaling via the amino acid binding site of the yeast Gap1 amino acid transceptor. *Nat. Chem. Biol.* 5, 45–52. doi: 10.1038/nchembio.132
- Wirtz, M., Beard, K. F., Lee, C. P., Boltz, A., Schwarzlander, M., Fuchs, C., et al. (2012). Mitochondrial cysteine synthase complex regulates O-acetylserine biosynthesis in plants. *J. Biol. Chem.* 287, 27941–27947. doi: 10.1074/jbc.M112.372656
- Yatusevich, R., Mugford, S. G., Matthewman, C., Gigolashvili, T., Frerigmann, H., Delaney, S., et al. (2010). Genes of primary sulfate assimilation are part of the glucosinolate biosynthetic network in *Arabidopsis thaliana*. *Plant J.* 62, 1–11. doi: 10.1111/j.1365-313X.2009.04118.x
- Yi, H., Galant, A., Ravilious, G. E., Preuss, M. L., and Jez, J. M. (2010). Sensing sulfur conditions: simple to complex protein regulatory mechanisms in plant thiol metabolism. *Mol. Plant* 3, 269–279. doi: 10.1093/mp/ssp112
- Yoshimoto, N., Inoue, E., Watanabe-Takahashi, A., Saito, K., and Takahashi, H. (2007). Posttranscriptional regulation of high-affinity sulfate transporters in *Arabidopsis* by sulfur nutrition. *Plant Physiol.* 145, 378–388. doi: 10.1104/pp.107.105742
- Zhang, B., Pasini, R., Dan, H., Joshi, N., Zhao, Y., Leustek, T., et al. (2014). Aberrant gene expression in the *Arabidopsis* SULTR1;2 mutants suggests a possible regulatory role for this sulfate transporter in response to sulfur nutrient status. *Plant J.* 77, 185–197. doi: 10.1111/tpp.12376
- Zhao, Q., Wu, Y., Gao, L., Ma, J., Li, C. Y., and Xiang, C. B. (2014). Sulfur nutrient availability regulates root elongation by affecting root indole-3-acetic acid levels and the stem cell niche. *J. Integr. Plant Biol.* doi: 10.1111/jipb.12217 [Epub ahead of print].

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 30 September 2014; accepted: 26 November 2014; published online: 11 December 2014.

Citation: Zheng Z-L, Zhang B and Leustek T (2014) Transceptors at the boundary of nutrient transporters and receptors: a new role for *Arabidopsis* SULTR1;2 in sulfur sensing. *Front. Plant Sci.* 5:710. doi: 10.3389/fpls.2014.00710

This article was submitted to *Plant Physiology*, a section of the journal *Frontiers in Plant Science*.

Copyright © 2014 Zheng, Zhang and Leustek. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.