



Selenium Accumulation Characteristics and Biofortification Potentiality in Turnip (*Brassica rapa* var. *rapa*) Supplied with Selenite or Selenate

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Li X, Wu Y, Li B, Yang Y and Yang Y (2018) Selenium Accumulation Characteristics and Biofortification Potentiality in Turnip (Brassica rapa var. rapa) Supplied with Selenite or Selenate. Front. Plant Sci. 8:2207. doi: 10.3389/fpls.2017.02207 Selenium (Se) is an essential trace element for humans. About 70% of the regions in China, including most of the Tibetan Plateau, are faced with Se deficiency problems. Turnip is mainly distributed around the Tibetan Plateau and is one of the few local crops. In the present study, we compared the absorption and translocation differences of Se (IV) selenite and Se (VI) selenate in turnip. The results showed that Se treatment, either by soil addition (0.2–2 mg Se kg⁻¹ dry soil) or by foliar spraying (50–200 mg L⁻¹ Se), could significantly increase the Se concentrations in turnips, and 0.5 mg Se (IV) or Se (VI) kg⁻¹ dry matter in soils could improve the biomasses of turnips. Moreover, turnip absorbed significantly more Se (VI) than Se (IV) at the same concentration and also transferred much more Se (VI) from roots to leaves. Based on the Se concentrations, as well as the bioconcentration factors and translocation coefficients, we considered that turnip might be a potential Se indicator plant. Subsequently, we estimated the daily Se intake for adults based on the Se concentrations in turnip roots. The results indicated that Se (IV) should be more suitable as an artificial Se fertilizer for turnips, although the levels found in most samples in this study could cause selenosis to humans. In addition, we also estimated the optimum and maximum Se concentrations for treating turnips based on the linear relations between Se concentrations in turnip roots and Se treatment concentrations. The results provided preliminary and useful information about Se biofortification in turnips.

Keywords: selenium deficiency, turnip, selenite, selenate, tibetan plateau

INTRODUCTION

Selenium (Se) is an essential trace element for human beings and animals (White and Brown, 2010). Se deficiency is a serious threat to human health, and is associated with cardiovascular disease, a weakened immune system, hypothyroidism, male infertility, cognitive decline and increased risks of various cancers (Fairweather-Tait et al., 2011; Rayman, 2012; White, 2016). The World Health Organization (WHO) recommended a dietary allowance of about from 55 to 200 μ g Se d⁻¹ for different groups of people (Wu et al., 2015). Unfortunately, a great number of people around the

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world may lack sufficient Se for their well-being, mainly because of the uneven distribution of Se resources on the earth (Combs, 2001; Fairweather-Tait et al., 2011; Joy et al., 2014; Wu et al., 2015; White, 2016). Taking China as an example, approximately 70% of its regions are faced with Se deficiency in varying degrees (Zhu et al., 2009; Wu et al., 2015), though several areas are also found to possess seleniferous soils (Wu et al., 2015). Because much of Se in human bodies is derived either directly or indirectly from edible plants, Se in diet greatly depends on grain and vegetable production in soils with substantial Se content or Se phytoavailability (Broadley et al., 2006; White and Broadley, 2009; Chilimba et al., 2011; Joy et al., 2015). However, excessive dietary Se intakes can also produce toxic effects in humans and animals (Fairweather-Tait et al., 2011; Rayman, 2012; Sperotto et al., 2014). The symptoms of selenosis in humans are similar to those caused by heavy metals, and include dermatitis, cracking of nails, hair loss, garlicky breath, acute respiratory distress, myocardial infarction, and renal failure (White, 2016). The Institute of Medicine (USA) has suggested a tolerable upper intake of 400 μ g Se d⁻¹ for adults (White, 2016).

Although some studies have focused on the potential of plants with high-Se accumulation capacities in phytoremediation of Se-contaminated soils (Banuelos and Dhillon, 2011; Wu et al., 2015), much interest has been developed toward the value of crops, vegetables, and edible mushrooms (Zhao et al., 2004; Maseko et al., 2014; Dogan et al., 2016), for producing Seenriched food because of the severe problem of Se deficiency. Indeed, the application of inorganic Se fertilizers, especially in non-seleniferous areas, has been an effective way to increase Se content of diets and to improve the Se status and health of both animals and humans (White and Broadley, 2009; Alfthan et al., 2015). Se concentrations in plants are directly associated with both Se phytoavailability in the soil and the ability of plants to accumulate Se (Dhillon and Dhillon, 2009). Angiosperm species can be divided into three ecological types based on their Se accumulation ability in tissues, which include nonaccumulator, Se-indicator and Se-accumulator species (White et al., 2007; White, 2016). Most angiosperm species belong to non-accumulator species, which are unable to tolerate tissue Se concentrations >10-100 mg Se kg⁻¹ dry weight (DW) and can hardly survive in seleniferous soils (Dhillon and Dhillon, 2009; White, 2016). In comparison, the Se-indicator species can tolerate tissue Se concentrations approaching 1 g Se kg⁻¹ DW and survive in both non-seleniferous and seleniferous soils (Moreno Rodriguez et al., 2005; White, 2016). For Seaccumulator species, whose distribution is usually constrained within seleniferous soils, their tissue Se concentrations can exceed 1 g Se kg⁻¹ DW (White, 2016). In particular, an extreme sub-set of Se-accumulator species are defined as Sehyperaccumulators, with leaves containing at least 1 g Se kg⁻¹ DW in natural environments (Terry et al., 2000), although some scientists suggested that this threshold should be lowered to 100 μ g Se g⁻¹ DW (van der Ent et al., 2012). To date, known Se-accumulator species include several members of Asteraceae, Brassicaceae, and Fabaceae, reported from America, Australia and China, respectively (Freeman et al., 2006, 2010; Yuan et al., 2013; White, 2016). These species accommodate high Se concentrations in leaf trichomes and epidermal cells (Freeman et al., 2006, 2010). Several members of the Lecythidaceae family are also well-known for accumulating high Se concentrations in their fruits and seeds (Chang et al., 1995; Hammel et al., 1996; Dernovics et al., 2007).

Brassicaceae species have always attracted much attention for their Se accumulation characteristics (Suarez et al., 2003; Seppanen et al., 2010; Hladun et al., 2011). The major reason is that Brassicaceae plants include a large proportion of the commonly cultivated vegetables around the world, such as pakchoi, cabbage, broccoli, mustard, radish and turnip, etc. Undoubtedly, Brassicaceae crops provide a considerable way to supplement Se intake for humans. However, Brassicaceae species, containing a large number of members (Couvreur et al., 2010), may have variable Se accumulation abilities that range from nonaccumulator to Se-hyperaccumulator species (Suarez et al., 2003; Seppanen et al., 2010; Yuan et al., 2013). Therefore, it is necessary to independently discover the Se accumulation ability of each individual species. Turnip (Brassica rapa var. rapa), a cruciferous biennial plant, has been widely cultivated as a vegetable or fodder crop in Europe, America and Asia over a long history. It is rich in vitamin C, riboflavin, dietary fiber, and a variety of mineral elements, but is low in calories (Parveen et al., 2015). It is also considered to have antioxidants and can lower the risk of high blood pressure and diabetes, as well as various cancers (Parveen et al., 2015). In China, the distribution center of turnip is the Tibetan Plateau and its surrounding areas, which are faced with severe Se deficiency problems. Thus, it is of great interest to explore the absorption and accumulation characteristics of this species in China for Se nutrition supply to local people. Few relevant studies have been performed to date, according to a recent review from China (Wu et al., 2015).

In natural environments, selenite and selenate are the main water-soluble forms of Se in oxic and anaerobic soils, respectively (White, 2016). Plants usually show differential accumulation ability for these two Se forms and they may have different toxicity thresholds in plants (Fu et al., 2011; Mao et al., 2011; Longchamp et al., 2015; Schiavon et al., 2016). In the present study, we compared the absorption and translocation differences of selenite and selenate in turnip and estimated the Se intake safety in turnip foodstuffs based on the experimental results. We would like to understand the ability of turnips to accumulate Se and to assess their potential for producing Se-supplemented food in natural seleniferous soils or via artificial Se fertilizers.

MATERIALS AND METHODS

Plant Cultivation and Treatment

Turnip seeds were collected from Ninglang County of China, from a population constituting a local landrace (NO. KTRG-B54) (Li et al., 2016). The seeds were germinated and grown outdoors in mid-April. To compare the differences of absorption and translocation of Se forms of Se (IV) selenite and Se (VI) selenate in turnip, concentrations of 0, 0.2, 0.5, and 2 mg Se kg⁻¹

Abbreviations: BCF, bioconcentration factor; DW, dry weight; Se, selenium; TC, translocation coefficient.

DM were established through fertilization of artificial Se-free mucky soil (**Table 1**) with sodium selenite (Na₂SeO₃) and sodium selenate (Na₂SeO₄), respectively. Three uniform $35 \times 28 \times 21$ -cm boxes were prepared for each Se treatment and 5 kg mucky soil was fitted into the boxes. After 25-day growth, three turnip seedlings with consistent size were neatly transplanted in each box. The boxes were placed under a transparent plastic shed with appropriate watering. At 45 day post-transplantation, the plants of each treatment were harvested for subsequent measurement.

In order to compare the effects of Se supply by foliar application, seedlings of the same size were transplanted into uniform (15-cm diam., 15-cm high) flowerpots filled with equivalent aforementioned soils (one seedling in each pot). The pots were divided into four groups (nine pots for each group) and the plants were supplied with 0, 50, 100, or 200 mg L^{-1} of Se (IV) by foliar application 15 and 30 day after transplanting, respectively. The pots were placed under a transparent plastic shed with appropriate watering. After transplanting for 45 day, the plants in each treatment were harvested for subsequent measurement.

Sample Preparation and Biomass Measurement

The roots and leaves of the treated plants were harvested, and the roots were washed with distilled water. The fresh weights of the root and leaf samples were measured, and the samples dried in an 80°C oven for 48 h prior to dry biomass measurement. A conversion factor for converting fresh weight of the experimental plants to dry weight was calculated. Three biological replications were performed for each sample.

Se Concentration Determination

The total Se concentration in the leaves and roots for all treatment samples (excluding control samples) was determined by hydride generation-atomic fluorescence spectrometry (HG-AFS) based on the national food safety standard of China (GB 5009.93-2010). Briefly, approximately 0.5-1.0 g dried samples were added to digestion bottles. A 10-mL acid mixture (9:1 nitric acid: perchloric acid) was subsequently injected into the bottle and the samples were left to digest overnight. Afterwards, the mixtures were heated until the solutions ran clear (2 mL solution left). When the temperature cooled, the solutions were combined with 5.0 mL HCl (4.10) and subjected to continued heating until the solutions ran clear with emission of white smoke from the samples. When Se (VI) was reduced to Se (IV), the solutions were cooled and then transferred to 50-mL volumetric flasks and fixed to a volume of 50 mL after rinsing three times using ultrapure water. The blank control was treated using the same method. A 10-mL digestion solution was mixed with 2.0 mL HCl (4.30) and 1.0 mL ferricyanide solution (4.70) in a 15-mL centrifuge tube for determining concentration using an atomic fluorescence spectrometer. The Se concentrations were calculated according to a standard curve. To draw the standard curve, 0, 0.1, 0.2, 0.3, 0.4, and 0.5 mL of standard solutions (1 μ g mL⁻¹) were injected into 15-mL centrifuge tubes and volume fixed to 10 mL in each tube. The solutions were individually mixed with 2.0 mL HCl (4.30) and 1.0 mL ferricyanide solution (4.70) for detection. TABLE 1 | Parameters of the soil used in the experiment.

Parameter	Unit	Soil D
н	/	7.44
Organic matter	g kg ⁻¹ DW	332.9
Humus	g kg ⁻¹ DW	191.2
Total N	g kg ⁻¹ DW	0.79
Total P	g kg ⁻¹ DW	0.37
Available K	mg kg ⁻¹ DW	275.2
Exchangeable Ca	cmol kg ⁻¹ DW	12.15
Exchangeable Mg	cmol kg ⁻¹ DW	1.18
Total Se	${ m mg}~{ m kg}^{-1}~{ m DW}$	ND ^a

 a ND indicates that the data were not detected. The detection limit in the experiment was 0.02 mg kg^{-1}.

A standard curve was not drawn until the linear correlation coefficient was greater than 0.99. The detection limit of total Se was 0.02 mg kg^{-1} . Three biological replications were performed for each sample.

Parameter Calculation

- Bioconcentration factor (BCF) = Se concentration in plant (mg kg⁻¹)/Se concentration in soil (mg kg⁻¹) (Liao et al., 2013).
- (2) Translocation coefficient (TC) = Se concentration in leaf (mg kg⁻¹)/Se concentration in root (mg kg⁻¹) (Huang et al., 2012).
- (3) Daily intake of Se (μg) = C_{Se} × C_{factor} × D_{foodintake} × 1000. C_{Se}, C_{factor}, and D_{foodintak} represent the Se concentrations in plants (mg kg⁻¹ DW), conversion factor, and daily intake of vegetables, respectively. The conversion factor 0.107 is used to convert fresh weight to dry weight for turnip fleshy roots in the present study. The average daily vegetable intake for adults was considered to be 0.345 kg person⁻¹ day⁻¹ (Parveen et al., 2015).
- (4) Optimum or maximum C_{Se} in turnip (mg kg⁻¹ DW) = Optimum or maximum daily Se intake/ $(C_{\text{factor}} \times D_{\text{foodintake}} \times 1000)$. The optimum and maximum daily Se intake is 55– 200 µg and 400 µg, respectively. C_{Se} , C_{factor} , and $D_{\text{foodintak}}$ represent Se concentrations, conversion factor and daily intake of vegetables, respectively. The conversion factor 0.107 is used to convert fresh weight to dry weight for fleshy turnip roots in the present study. The average daily vegetable intake for adults is 0.345 kg person⁻¹ day⁻¹ (Parveen et al., 2015).

Statistical Analysis

Statistical analyses were performed using SPSS version 18.0. Parametric (one-way ANOVA or independent-samples *t*-test) or nonparametric (K independent-samples or 2 independent-samples) statistical tests, respectively, were applied.

The data were fitted to linear [y = ax+b] and polynomial $[y = ax^2+bx+c]$ models to analyze the correlations. Based on R^2 and P values, the model (either linear or polynomial) best fit to the data was selected. All analyses were conducted with SigmaPlot 10.0.

RESULTS

Effects of Different Treatment Modes on Turnip Growth

Leaf biomass was significantly greater when 0.5 mg kg⁻¹ DW Se (IV) was added to soils (P < 0.05) whereas Se (VI) showed no significant effects on the leaf biomass accumulation in turnip at 0.2–2 mg kg⁻¹ DW concentrations (**Figure 1A** top). The root biomass was markedly improved by both Se (IV) and Se (VI) at a concentration of 0.5 mg kg⁻¹ DW in soil (P < 0.05) (**Figure 1A** bottom). However, the growth of turnips (both the leaves and roots) was not influenced by 50–200 mg L⁻¹ Se (IV) used for foliar application in the present study (**Figure 1B**).

Accumulation Characteristics of Se (IV) and Se (VI) in Turnip

Se concentration in turnip leaves was 0.06, 0.27, and 2.73 mg kg^{-1} DW when treated by 0.2, 0.5, and $2 mg kg^{-1}$ DW Se (IV), respectively (Figure 2A top), while the values were 7.13, 37.53, and 150.33 mg kg⁻¹ DW, respectively, under the same Se (IV) concentrations (Figure 2A top). Se concentration in turnip leaves was significantly increased with the increasing soil Se concentrations of both Se (IV) and Se (VI) (P < 0.05) (Figure 2A top). Additionally, we found that turnip leaves treated by Se (VI) accumulated obviously more Se compared with those treated by Se (IV) under the same concentration (P < 0.001) (Figure 2A top). Se concentrations in turnip roots showed similar change patterns with those in leaves (Figure 2A bottom), which ranged from 2.96 to 12.44 mg kg⁻¹ DW and from 6.30 to 158.33 mg kg⁻¹ DW in roots treated by Se (IV) and Se (VI), respectively (Figure 2A bottom). When treated by a foliar application of 50- 200 mg L^{-1} Se (IV), turnip leaves and roots had Se concentrations of $18.71-35.70 \text{ mg kg}^{-1}$ DW and $6.50-19.77 \text{ mg kg}^{-1}$ DW, respectively (Figure 2B). Se concentrations were similar between 50 and 100 mg L^{-1} Se (IV) treatment but were significantly higher when the treatment concentration reached 200 mg L^{-1} (P < 0.05) (Figure 2B). Interestingly, the Se concentrations in both leaves and roots of turnips had highly significant positive correlations with the Se treatment concentrations in different modes (P < 0.01), which showed representative linear relations (Table 2).

In order to reflect the ability of turnips to accumulate different Se forms, we calculated the BCFs in leaves and roots for Se (IV) and Se (VI) treatment by soil addition. The BCFs in leaves of turnip treated by Se (IV) (from 0.2 to 2 mg kg^{-1} DW) ranged from 0.31 to 1.36 whereas those of turnip treated with Se (VI) increased from 35.67 to 75.17 (Figure 3A). Under the same Se concentration, BCF of Se (VI) in turnip leaves was significantly higher than that of Se (IV) (P < 0.001) (Figure 3A). Interestingly, BCFs of Se (IV) in turnip roots were relatively stable but showed a decreasing trend (from 14.82 to 6.22) with increasing Se concentrations (from 0.2 to 2 mg kg^{-1} DW) (Figure 3B). However, the BCFs in roots of turnip treated with Se (VI) were similar between 0.2 and 0.5 mg kg⁻¹ DW concentrations (31.50 and 32.27) but were significantly higher when the treatment concentration reached 2 mg kg^{-1} DW (79.17) (P < 0.05) (Figure 3B). As a result, the BCFs in turnip roots treated by Se (VI) were significantly higher than those of Se (IV) treatment samples under 0.2 and 2 mg kg⁻¹ DW treatment concentrations (**Figure 3B**). TCs in turnips treated by Se (IV) were similar under 0.2 and 0.5 mg kg⁻¹ DW concentrations (0.02 and 0.05) but were significantly higher when the treatment concentration reached 2 mg kg⁻¹ DW (0.25) (**Figure 3C**); however, TCs in turnips treated by Se (VI) reached the maximum at 0.5 mg kg⁻¹ DW concentration (2.33) while the values of 0.2 and 2 mg kg⁻¹ DW concentrations were similar (1.13 and 0.95) (**Figure 3C**).

Se Biofortification Potentiality Analysis

Estimated daily Se intake levels are showed in **Table 3**. For adults, the daily intake of Se from turnip roots treated with 0.2–2 mg kg⁻¹ DW Se (IV) added in soil was estimated to be 108.93–457.27 μ g, while those from samples treated with Se (VI) was 231.58–5,820.05 μ g (**Table 3**). The daily intake of Se from turnip roots treated with a foliar application of 50–200 mg L⁻¹ Se (IV) ranged from 238.86 to 726.71 μ g (**Table 3**).

We also estimated optimum and maximum Se concentrations in turnip roots based on the optimum (55-200 µg) and maximum (400 μ g) daily intake of Se for adults, and further estimated the corresponding exogenous Se treatment concentrations by different modes based on above analyzed linear equations (Table 2) between Se treatment concentrations and Se concentrations in plants. The optimum Se concentration in turnip roots was $1.50-5.44 \text{ mg kg}^{-1}$ DW while the optimum Se concentration in turnip roots was 10.88 mg kg⁻¹ DW (**Table 4**). Maximum Se treatment concentrations were $1.60 \text{ mg kg}^{-1} \text{ DW}$, 0.26 mg kg⁻¹ DW, and 108.42 mg L^{-1} for soil Se (IV), soil Se (VI), and foliar Se (IV) treatments, respectively (Table 4). Optimum Se treatment concentrations were $0.15-0.19 \text{ mg kg}^{-1}$ DW and 10.44–51.62 mg L^{-1} for soil Se (VI) and foliar Se (IV) treatments, respectively (Table 4). Unfortunately, the values of optimum Se treatment concentrations for soil Se (IV) treatment were unavailable using the linear models (Table 4).

DISCUSSION

Effects of Se on Turnip Growth

Although Se is not an essential element for angiosperms, it is considered to be a beneficial element since it can stimulate growth, confer tolerance to abiotic stresses, and provide resistance to pathogens or herbivory (Quinn et al., 2007; Pilon-Smits et al., 2009; Feng et al., 2013). Some studies have reported that Se could improve plant growth and grain production (Lyons et al., 2009). However, Se toxicity also has been often observed (Ximenez-Embun et al., 2004; Fu et al., 2011; Mao et al., 2011; Longchamp et al., 2015). For example, when grown with 12 μ mol L⁻¹ selenite, white lupine and sunflower were reported to experience a biomass reduction of 20% and 40%, respectively (Ximenez-Embun et al., 2004). Indeed, the effects of Se on plant growth usually show the principle of lowconcentration promotion and high-concentration inhibition; this has been observed in many plants including radish, Chinese cabbage, pakchoi, rapeseed, and spinach (Fu et al., 2011; Mao et al., 2011). The results of our study supported this assertion, where both selenite and selenate had optimal promotion effects









at 0.5 mg Se kg⁻¹ dry soil and then began to inhibit plant growth as the Se concentrations increased. The results of Se (IV) treatment by foliar application also indicated that there was an optimum concentration for turnip growth near 100 mg L⁻¹. Dhillon and Dhillon (2009) obtained similar results, in which the authors found that dry matter accumulation of turnip and several other vegetables decreased when 1.25-5.0 mg kg⁻ selenate-Se was applied to the soil (Dhillon and Dhillon, 2009). The effect intensity of Se (IV) vs. Se (VI) on plant growth is inconclusive. Mao et al. (2011) found that Se (VI) had a significant promoting effect on cauliflower at low concentrations and lower toxicity to wheat and alfalfa at high concentrations compared with Se (IV) (Mao et al., 2011). Longchamp et al. (2015) also reported that Se (VI) produced lower inhibition effect on the growth of maize crops compared to Se (IV) at $12\,\mu M$ Se concentration (Longchamp et al., 2015). However, we found similar effects of Se (IV) vs. Se (VI) on turnip growth at 0.2- 2 mg kg^{-1} Se concentrations. This might be because the range of Se concentrations for promoting or inhibiting plant growth was greatly distinct in different species, attributed to the differences of plant tolerance to Se or various environmental conditions.

TABLE 2 | Results of regression analysis between exogenous Se treatment concentrations and Se accumulation concentrations in turnip leaves and roots.

Treatment modeSe formTissue TissueRegression equation R^2 FSoil additionSe (IV)Leaf Root $y = 1.433x - 0.20$ 0.980477.155Soil additionSe (IV)Leaf V = 5.569xx + 1.9980.59414.654Se (VI)Leaf V = 76.691x - 3.0160.9973335.329Root $y = 82.665x - 10.607$ 0.980497.228Foliar applicationSe (IV)Leaf V = 0.164x + 4.6980.85659.413							
addition Root $y = 5.569xx + 1.998$ 0.594 14.654 Se (VI) Leaf $y = 76.691x - 3.016$ 0.997 3335.329 Root $y = 82.665x - 10.607$ 0.980 497.228 Foliar Se (IV) Leaf $y = 0.164x + 4.698$ 0.856 59.413		Se form	Tissue	U U	R ²	F	Р
Root $y = 5.569X + 1.998$ 0.394 14.654 Se (VI) Leaf $y = 76.691x - 3.016$ 0.997 3335.329 Root $y = 82.665x - 10.607$ 0.980 497.228 Foliar Se (IV) Leaf $y = 0.164x + 4.698$ 0.856 59.413		Se (IV)	Leaf	y = 1.433 x - 0.20	0.980	477.155	<0.001
Root $y = 82.665x - 10.607$ 0.980 497.228 Foliar Se (IV) Leaf $y = 0.164x + 4.698$ 0.856 59.413	dition		Root	y = 5.569xx + 1.998	0.594	14.654	0.003
Foliar Se (IV) Leaf $y = 0.164x + 4.698$ 0.856 59.413		Se (VI)	Leaf	y = 76.691 x - 3.016	0.997	3335.329	< 0.001
application			Root	y = 82.665x - 10.607	0.980	497.228	<0.001
application	Foliar application	Se (IV)	Leaf	y = 0.164x + 4.698	0.856	59.413	<0.001
Root $y = 0.096x + 0.496$ 0.883 75.217			Root	y = 0.096x + 0.496	0.883	75.217	< 0.001

Se Absorption and Translocation in Turnip

Se (IV) and Se (VI) are two main Se forms in the natural soils and can be absorbed by plant roots (White and Broadley, 2009). Se forms and soil environment, especially soil pH and pe, greatly affect Se uptake and accumulation in plants (Longchamp et al., 2015). Our results showed that turnips more easily accumulated Se (VI) in the soil humus in comparison with Se (IV). This was consistent with previous reports finding that Se (VI) was relatively mobile in soil solution and thus the addition of Se (VI) to soils facilitated immediate Se accumulation by plants while Se (IV) provided a longer-lasting Se source (Broadley et al., 2006). However, many studies on rice, wheat, soybean, or maize have also shown that Se (IV) can accumulate as much as (Zaved et al., 1998; Li et al., 2008) or even more than (Zhang et al., 2003; Longchamp et al., 2015) Se (VI). For example, Longchamp et al. (2015) found that maize absorbed more Se (IV) than Se (VI) in whole plants, as well as in roots and grains, although the Se concentrations in stems and leaves supplied with Se (VI) were much higher than those supplied

TABLE 3	Daily intake of Se (μ g) for adults from fleshy turnip roots.
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Treatment mode	Se form	Concentration	Daily intake of Se (μ g)
Soil addition	Se (IV)	0.2 mg kg ⁻¹ DW	108.93
		0.5 mg kg ⁻¹ DW	280.22
		2.0 mg kg ⁻¹ DW	457.27
	Se (VI)	0.2 mg kg ⁻¹ DW	231.58
		$0.5\mathrm{mg}\mathrm{kg}^{-1}$ DW	593.03
		2.0 mg kg ⁻¹ DW	5820.05
Foliar application	Se (IV)	50 mg kg ⁻¹ DW	238.86
		100 mg kg ⁻¹ DW	340.12
		$200\mathrm{mg}\mathrm{kg}^{-1}\mathrm{DW}$	726.71

The daily intake of turnip root for adults used in the present study is 0.345 kg fresh weight.





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Optimum Se concentration in plant (mg kg ⁻¹ DW)	Optimum Se treatment concentration	Maximum Se concentration in plant (mg kg ⁻¹ DW)	Maximum Se treatment concentration	Treatment mode
1.50–5.44	Unavailable	10.88	1.60 mg kg ⁻¹ DW	Se (IV) (Soil)
	$0.15-0.19 \mathrm{mg kg^{-1}}$ DW		0.26 mg kg ⁻¹ DW	Se (VI) (Soil)
	10.44–51.62 mg L ⁻¹		$108.42 \mathrm{mg}\mathrm{L}^{-1}$	Se (IV) (Foliar)

TABLE 4 | Estimated optimum and maximum Se treatment concentrations for different treatment modes based on the optimum (55–200 µg) and maximum (400 µg) daily Se intake for adults from fleshy turnip roots (0.345 kg fresh weight).

The treatment methods and harvest time were assumed to be those used in the present study.

with Se (IV) (Longchamp et al., 2015). Our results also agreed with findings that Se contents in plants were dependent on the Se concentrations in soils (Fu et al., 2011; Schiavon et al., 2016). In the present study, the Se concentration in roots and leaves of turnip plants was significantly positively correlated with Se (IV) or Se (VI) treatment concentrations applied either by soil addition or foliar application. Plant accumulation ability is essential for both biofortification and phytoremediation (Wu et al., 2015). To date, only a few studies have definitely provided data about Se accumulation in turnips worldwide. An American study reported that Se concentrations in turnips were 60 mg kg^{-1} DW in the presence of $2 \text{ mg } \text{L}^{-1}$ Se in irrigation water and Se concentrations in turnip shoots irrigated by sprinkler were about twice the amount in flood-irrigated plants(Suarez et al., 2003). Unfortunately, these results are different compared to our findings that Chinese turnips accumulated relatively low Se concentrations (about $20-30 \text{ mg kg}^{-1} \text{ DW}$) irrigated by sprinkler using as high as 200 mg L^{-1} Se (IV). This might be attributed to complex reasons, including diverse accumulation abilities in turnip genotypes and the differences in solution compositions. In an Indian study, scientists reported that turnip accumulated $60-70 \text{ mg kg}^{-1}$ Se both in edible and inedible portions when the soil was fertilized with 2.5 mg kg^{-1} selenate-Se (Dhillon and Dhillon, 2009). The results were also inconsistent with our findings that Chinese turnips accumulated less than 15 mg kg⁻¹ DW Se (IV) but more than 150 mg kg^{-1} DW Se (VI) in both roots and leaves at the similar Se concentrations added to the soil. However, Liu et al. (2012) found that turnips from the Tibet region contained 6.33 mg kg⁻¹ Se (Liu et al., 2012); this is an interestingly high value for local areas lacking soil Se. Although significant differences have been observed in different reports worldwide, these results have indicated that turnip has an ability to accumulate Se under artificial or natural conditions. We found that turnip could tolerate more than $150 \text{ mg kg}^{-1} \text{ DW}$ Se in both leaves and roots, which did not significantly affect the plant growth. The results indicate that turnip might belong to potential Se-indicator species as defined in a previous report (White, 2016).

BCF is usually used to reflect the ability of plants to absorb trace elements from the soil environment (Liao et al., 2013). Our results showed that the maximum BCFs of Se (IV) and Se (VI) in turnip roots reached 15 and 79, respectively, reflecting an ability of turnips to absorb Se, but a stronger ability for Se (VI). TC is used to reflect transport of elements or ions in plants (He, 2013), which is closely related to the intake risks of contaminants in leaf vegetables, as well as phytoremediation efficiency for

polluted environments (Gao et al., 2012). Interestingly, the TCs of Se (IV) were far lower than 1 whereas those of Se (VI) were about 1-2. These results indicated a differential ability to transport different forms of Se in turnips. This might be attributed to a differential metabolism and transformation of Se (IV) and Se (VI) in turnips (Wu et al., 2015). In plants, Se (IV) is thought to be rapidly converted to organoselenium compounds in the root whereas Se (VI) is delivered immediately to the xylem (White et al., 2004; Ximenez-Embun et al., 2004; Li et al., 2008) and subsequently assimilated into organoselenium compounds in plastids (Pilon-Smits and Leduc, 2009). With Se (IV) treatment, organoselenium compounds might therefore be produced to a greater extent vs. production in the Se (VI) treatment. Longchamp et al. (2015) found no trace of inorganic Se could be detected in whole plants treated by Se (IV) (Longchamp et al., 2015). However, when treated with Se (VI), the percentage of Se (VI) in roots, stems and leaves was respectively 20, 54, and 39% (Longchamp et al., 2015). In several studies, traces of Se (IV) were only detected at less than 7%, indicating that Se (IV) in plants overwhelmingly consists of organoselenium compounds (Ximenez-Embun et al., 2004). However, some studies have also shown that about 30-97% of Se in the leaves, stems or roots consists of Se (IV) (Ximenez-Embun et al., 2004; Li et al., 2008; Mazej et al., 2008; Longchamp et al., 2015). Thus, further analysis of the metabolism and transformation processes of different Se forms in turnips is needed. More interestingly, we found that the BCFs of Se (IV) in leaves, as well as those of Se (VI) in both leaves and roots, showed maximum values at the highest Se treatment concentration $(2.0 \text{ mg kg}^{-1} \text{ DW})$ used in the present study. A similar result was also observed for the TC value of Se (IV), whereas that of Se (VI) reached the maximum at 0.5 mg kg^{-1} DW Se treatment concentration. These results indicate that higher Se content in soils further improved bioavailability of Se, but this requires further verification and exploration.

Se Biofortification Potentiality in Turnip

Producing and consuming the biofortified agricultural products has been proposed as a promising functional agricultural strategy to increase dietary nutrient intake, e.g., Se, for humans (Wu et al., 2015). Biofortification of Se is closely connected to enhancing the efficiency of Se uptake and accumulation in plants (Vamerali et al., 2014). The Tibetan Plateau is famous for having a hard-living environment for higher organisms, including animals and humans. Unfortunately, most parts of the plateau are considered to be areas where Se deficiency disease

frequently occurs (Zhu et al., 2009). Turnip is a cruciferous vegetable mainly distributed around the Tibetan Plateau in China; through biofortification it may be an excellent candidate to supplement daily Se requirement for local people, since cruciferous plants usually have strong abilities to accumulate micronutrients (e.g., Se, zinc, and iron; White and Broadley, 2009). The results of the present study and several previous reports indicate that turnip has a relatively strong ability to accumulate Se (Dhillon and Dhillon, 2009; Liu et al., 2012) and show that turnip is a possible candidate for developing Sefunctional products. In addition, turnip is rich in nutrients like vitamins, mineral elements, and amino acids (Liu et al., 2012; Ma et al., 2016) and contains a variety of medicinal ingredients (Li et al., 2011; Hu et al., 2016; Ma et al., 2016). Moreover, Se has been found to have beneficial effects in promoting plant nutrition and metabolites (Zhao et al., 2003, 2004). For example, Se can improve the synthesis of glucosinolates, important secondary metabolites found mainly in cruciferous plants, by replacing the sulfur element and promoting the activities of some enzymes (Robbins et al., 2005; Barickman et al., 2013; Avila et al., 2014). Se fertilizer concentrations that are beneficial for turnip are therefore greatly recommended. Moreover, Se also has been demonstrated to improve the tolerance of plants to common abiotic stresses including drought, cold and heavy metals (Chu et al., 2010; Al-Waeli et al., 2012; Lin et al., 2012; Ahmad et al., 2016). This is also worth considering for turnip growth and yield in the Tibetan Plateau.

According to the recommended optimum dietary allowance of Se (55–200 μ g Se d⁻¹) and the maximum critical safety value (400 μ g Se d⁻¹) for adults (Wu et al., 2015), we estimated the daily Se intake for adults based on the Se concentrations in the turnip roots in the present study, which were the main edible portions for local people. Unfortunately, the results produced by most samples exceeded thresholds for supplying daily Se intake. Comparatively, the samples treated by Se (VI) would be much more likely to cause selenosis vs. those treated by Se (IV), either with soil addition or foliar spraying. Thus, Se (IV) should be a priority selection as artificial Se fertilizer for turnip under the similar soil conditions. As Se concentrations in turnip roots were significantly positively correlated with the exogenous Se treatment concentrations, we also estimated the optimum and maximum Se concentrations for treating turnips used in adult diets based on the linear regression equations. The results indicated that the safe Se concentrations for Se (IV) and Se (VI) treated by soil addition were below 1.60 and 0.26 mg kg^{-1} DW, respectively, whereas that for Se (IV) treated by foliar application should be lower than 108.42 mg L^{-1} . The results provide some useful information for Se biofortification in turnip. However, as mentioned above, organic Se is the most healthy Se form for humans; transformation efficiency of the inorganic Se (IV) and Se (VI) in turnips is unknown. Furthermore, soil properties have been demonstrated to affect Se accumulation and transformation

(Vamerali et al., 2014; Wu et al., 2015). In addition, stage of plant growth at treatment, as well as treatment frequency and the duration of post-treatment growth periods, is also closely related to the accumulation of Se in plants. Thus, much more effort is needed to determine the characteristics of turnips to Se and accurately evaluate their practicability for producing Se-enriched foodstuff. However, our results have provided preliminary information and narrowed the scope of relevant research.

CONCLUSIONS

In the present study, we compared the absorption and translocation differences of selenite and selenate in turnip. The results showed that the Se contents in both leaves and roots of turnip were significantly positively correlated with Se treatment concentrations either by soil addition or by foliar spraying. The biomass of turnips was improved at a concentration of 0.5 mg Se (IV) or Se (VI) kg⁻¹ dry soil. Moreover, turnip absorbed markedly more Se (VI) than Se (IV) in the same soils and much more Se (VI) could transfer from roots to leaves in comparison to Se (IV). Based on the relatively high Se concentrations, we considered that turnip might be a potential Se indicator plant, but supporting data is still needed from natural conditions. Results of BCFs and TCs also indicated that higher Se content in soils seemed to further improve bioavailability of Se. According to the optimum and maximum dietary allowance of Se for adults, we estimated the daily Se intake for adults based on our results. Although the Se concentrations in most root samples in this study were too high to be ingested by humans, we consider that Se (IV) should be a priority selection as an artificial Se fertilizer for turnips. In addition, the optimum and maximum Se concentrations for treating turnips were also estimated based on the linear relations between Se concentrations in turnip roots and Se treatment concentrations. The results provided some useful information for Se intake from turnips, but more effort is required to assess the practicability of using fortified turnips for producing Se-enriched foodstuff; open questions include a lack of transform efficiency of inorganic Se (IV) and Se (VI), the effects of soil properties on Se accumulation and transformation, and the effects of treatment stage of plant growth, treatment lasting time and times. Regardless, our results provide preliminary information and narrow the scope of relevant research.

AUTHOR CONTRIBUTIONS

YPY and XL conceived and designed the experiments. XL, YW, and BL performed the experiments. XL analyzed the data. XL wrote the manuscript. YHY supported the data determination.

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REFERENCES

- Ahmad, R., Waraich, E. A., Nawaz, F., Ashraf, M. Y., and Khalid, M. (2016). Selenium (Se) improves drought tolerance in crop plants-a myth or fact? J. Sci. Food Agric. 96, 372–380. doi: 10.1002/jsfa.7231
- Alfthan, G., Eurola, M., Ekholm, P., Venalainen, E. R., Root, T., Korkalainen, K., et al. (2015). Effects of nationwide addition of selenium to fertilizers on foods, and animal and human health in Finland: from deficiency to optimal selenium status of the population. *J. Trace Elem. Med. Biol.* 31, 142–147. doi: 10.1016/j.jtemb.2014.04.009
- Al-Waeli, A., Pappas, A. C., Zoidis, E., Georgiou, C. A., Fegeros, K., and Zervas, G. (2012). The role of selenium in cadmium toxicity: interactions with essential and toxic elements. *Br. Poult. Sci.* 53, 817–827. doi: 10.1080/00071668.2012.751523
- Avila, F. W., Yang, Y., Faquin, V., Ramos, S. J., Guilherme, L. R., Thannhauser, T. W., et al. (2014). Impact of selenium supply on Se-methylselenocysteine and glucosinolate accumulation in selenium-biofortified *Brassica sprouts. Food Chem.* 165, 578–586. doi: 10.1016/j.foodchem.2014.05.134
- Banuelos, G. S., and Dhillon, K. S. (2011). Developing a sustainable phytomanagement strategy for excessive selenium in western United States and India. *Int. J. Phytoremediation* 13(Suppl. 1), 208–228. doi: 10.1080/15226514.2011.568544
- Barickman, T. C., Kopsell, D. A., and Sams, C. E. (2013). Selenium influences glucosinolate and isothiocyanates and increases sulfur uptake in *Arabidopsis thaliana* and rapid-cycling *Brassica oleracea*. J. Agric. Food Chem. 61, 202–209. doi: 10.1021/jf3037227
- Broadley, M. R., White, P. J., Bryson, R. J., Meacham, M. C., Bowen, H. C., Johnson, S. E., et al. (2006). Biofortification of UK food crops with selenium. *Proc. Nutr. Soc.* 65, 169–181. doi: 10.1079/PNS2006490
- Chang, J. C., Gutenmann, W. H., Reid, C. M., and Lisk, D. J. (1995). Selenium content of Brazil nuts from two geographic locations in Brazil. *Chemosphere* 30, 801–802. doi: 10.1016/0045-6535(94)00409-N
- Chilimba, A. D., Young, S. D., Black, C. R., Rogerson, K. B., Ander, E. L., Watts, M. J., et al. (2011). Maize grain and soil surveys reveal suboptimal dietary selenium intake is widespread in Malawi. *Sci. Rep.* 1:72. doi: 10.1038/ srep00072
- Chu, J., Yao, X., and Zhang, Z. (2010). Responses of wheat seedlings to exogenous selenium supply under cold stress. *Biol. Trace Elem. Res.* 136, 355–363. doi: 10.1007/s12011-009-8542-3
- Combs, G. F. Jr. (2001). Selenium in global food systems. Br. J. Nutr. 85, 517–547. doi: 10.1079/BJN2000280
- Couvreur, T. L., Franzke, A., Al-Shehbaz, I. A., Bakker, F. T., Koch, M. A., and Mummenhoff, K. (2010). Molecular phylogenetics, temporal diversification, and principles of evolution in the mustard family (Brassicaceae). *Mol. Biol. Evol.* 27, 55–71. doi: 10.1093/molbev/msp202
- Dernovics, M., Garcia-Barrera, T., Bierla, K., Preud'homme, H., and Lobinski, R. (2007). Standardless identification of selenocystathionine and its gammaglutamyl derivatives in monkeypot nuts by 3D liquid chromatography with ICP-MS detection followed by nanoHPLC-Q-TOF-MS/MS. *Analyst* 132, 439–449. doi: 10.1039/B618637H
- Dhillon, K. S., and Dhillon, S. K. (2009). Accumulation and distribution of selenium in some vegetable crops grown in selenate-Se treated clay loam soil. *Front. Agric. China* 3, 366–373. doi: 10.1007/s11703-009-0070-6
- Dogan, H., Coteli, E., and Karatas, F. (2016). Determination of glutathione, selenium, and malondialdehyde in different edible mushroom species. *Biol. Trace Elem. Res.* 174, 459–463. doi: 10.1007/s12011-016-0715-2
- Fairweather-Tait, S. J., Bao, Y., Broadley, M. R., Collings, R., Ford, D., Hesketh, J. E., et al. (2011). Selenium in human health and disease. *Antioxid. Redox Signal.* 14, 1337–1383. doi: 10.1089/ars.2010.3275
- Feng, R. W., Wei, C. Y., and Tu, S. X. (2013). The roles of selenium in protecting plants against abiotic stresses. *Environ. Exp. Bot.* 87, 58–68. doi: 10.1016/j.envexpbot.2012.09.002
- Freeman, J. L., Tamaoki, M., Stushnoff, C., Quinn, C. F., Cappa, J. J., Devonshire, J., et al. (2010). Molecular mechanisms of selenium tolerance and hyperaccumulation in *Stanleya pinnata*. *Plant Physiol*. 153, 1630–1652. doi: 10.1104/pp.110.156570
- Freeman, J. L., Zhang, L. H., Marcus, M. A., Fakra, S., Mcgrath, S. P., and Pilon-Smits, E. A. (2006). Spatial imaging, speciation, and quantification of selenium

in the hyperaccumulator plants Astragalus bisulcatus and Stanleya pinnata. Plant Physiol. 142, 124–134. doi: 10.1104/pp.106.081158

- Fu, D. D., Duan, M. L., Liang, D. L., Wang, S. S., and Wu, X. P. (2011). Effects of selenite and selenate on growth and nutrient absorption of pakchoi. *Plant Nutr. Fertilizer Sci.* 17, 358–365.
- Gao, J., Liu, W. Y., and Chen, W. J. (2012). Potentials of 23 plants growing in contaminated soils nearby electroplating plants for accumulating and translocating heavy metals and remedying the soils. *J. Ecol. Rural Environ.* 28, 468–472.
- Hammel, C., Kyriakopoulos, A., Behne, D., Gawlik, D., and Bratter, P. (1996). Protein-bound selenium in the seeds of coco de mono (*Lecythis ollaria*). J. Trace Elem. Med. Biol. 10, 96–102. doi: 10.1016/S0946-672X(96)80017-4
- He, Q. X. (2013). Research progress of screening cadmium hyperaccumulators. Environ. Prot. Circ. Econ. 33, 46–49.
- Hladun, K. R., Parker, D. R., and Trumble, J. T. (2011). Selenium accumulation in the floral tissues of two Brassicaceae species and its impact on floral traits and plant performance. *Environ. Exp. Bot.* 74, 90–97. doi: 10.1016/j.envexpbot.2011.05.003
- Hu, X. D., Zhao, H. L., Wang, B., Wu, P., Yue, Y., and Zhou, F. (2016). Study on anti-hypoxia activity of traditional tibetan medicine *Brassica rapa* L. extract in mice. *Chin. J. Biochem. Pharm.* 36, 37–39.
- Huang, H. Y., Xu, J., Bai, Y., Zhang, W. Q., Zhu, F., Li, T., et al. (2012). Enrichment of heavy metals in *Saccharum arundinaceum* (Retz.) Jeswiet in different soil habitats. *Chin. J. Ecol.* 31, 961–966.
- Joy, E. J., Ander, E. L., Young, S. D., Black, C. R., Watts, M. J., Chilimba, A. D., et al. (2014). Dietary mineral supplies in Africa. *Physiol. Plant.* 151, 208–229. doi: 10.1111/ppl.12144
- Joy, E. J., Broadley, M. R., Young, S. D., Black, C. R., Chilimba, A. D., Ander, E. L., et al. (2015). Soil type influences crop mineral composition in Malawi. *Sci. Total Environ.* 505, 587–595. doi: 10.1016/j.scitotenv.2014.10.038
- Li, H. F., Mcgrath, S. P., and Zhao, F. J. (2008). Selenium uptake, translocation and speciation in wheat supplied with selenate or selenite. *New Phytol.* 178, 92–102. doi: 10.1111/j.1469-8137.2007.02343.x
- Li, S. M., Meng, L. B., Gao, X. Y., and Pan, M. Y. (2011). Analysis of glucosinolate content in two cultivar turnips. *North. Horticult*. 16, 1–3.
- Li, X., Zhang, X. M., Yang, Y., Li, B. Q., Wu, Y. S., Sun, H., et al. (2016). Cadmium accumulation characteristics in turnip landraces from China and assessment of their phytoremediation potential for contaminated soils. *Front. Plant Sci.* 7:1862. doi: 10.3389/fpls.2016.01862
- Liao, Q. L., Liu, C., Cai, Y. M., Zhu, B. W., Wang, C., Hua, M., et al. (2013). A preliminary study of element bioconcentration factors within milled rice and wheatmeal in some typical areas of Jiangsu Province. *Geol. China* 40, 331–340.
- Lin, L., Zhou, W., Dai, H., Cao, F., Zhang, G., and Wu, F. (2012). Selenium reduces cadmium uptake and mitigates cadmium toxicity in rice. J. Hazard. Mater. 23, 343–351. doi: 10.1016/j.jhazmat.2012.08.012
- Liu, Y. F., Gong, L. X., Liu, L. L., Cai, H. F., Wu, X. Q., and Zhang, Y. (2012). Determination on nutritional content of Tibetan turnip (*Brassica rapa* L.) and experimental study on improvement of mice hypoxia tolerance. *Sci. Technol. Food Ind.* 33, 412–416. doi: 10.13386/j.issn1002-0306.2012.09.010
- Longchamp, M., Castrec-Rouelle, M., Biron, P., and Bariac, T. (2015). Variations in the accumulation, localization and rate of metabolization of selenium in mature *Zea mays* plants supplied with selenite or selenate. *Food Chem.* 182, 128–135. doi: 10.1016/j.foodchem.2015.02.137
- Lyons, G., Genc, Y., Soole, K., Stangoulis, J., Liu, F., and Graham, R. (2009). Selenium increases seed production in *Brassica*. *Plant Soil* 318, 73–80. doi: 10.1007/s11104-008-9818-7
- Ma, G. C., Wang, Y. R., and Xuan, Z. Y. (2016). Analysis and comparison of nutritional compositions in Xinjiang turnip (*Brassica rapa L.*). Sci. Technol. Food Ind. 37, 360–364. doi: 10.13386/j.issn1002-0306.2016.04.064
- Mao, H., Wang, Z. H., Graham, L., and Glenn, M. (2011). Effects of selenium valence states and concentration on germination and root growth of six crop species. J. Agro-Environ. Sci. 30, 1958–1965. doi: 10.1016/j.chemosphere.2016.06.045
- Maseko, T., Dunshea, F. R., Howell, K., Cho, H. J., Rivera, L. R., Furness, J. B., et al. (2014). Selenium-enriched *Agaricus bisporus* mushroom protects against increase in gut permeability ex vivo and up-regulates glutathione peroxidase 1 and 2 in hyperthermally-induced oxidative stress in rats. *Nutrients* 6, 2478–2492. doi: 10.3390/nu6062478

- Mazej, D., Osvald, J., and Stibilj, V. (2008). Selenium species in leaves of chicory, dandelion, lamb's lettuce and parsley. *Food Chem.* 107, 75–83. doi: 10.1016/j.foodchem.2007.07.036
- Moreno Rodriguez, M. J., Cala Rivero, V., and Jimenez Ballesta, R. (2005). Selenium distribution in topsoils and plants of a semi-arid Mediterranean environment. *Environ. Geochem. Health* 27, 513–519. doi: 10.1007/s10653-005-8625-9
- Parveen, T., Hussain, A., and Rao, M. S. (2015). Growth and accumulation of heavy metals in turnip (*Brassica rapa*) irrigated with different concentrations of treated municipal wastewater. *Hydrol. Res.* 46, 60–71. doi: 10.2166/nh. 2014.140
- Pilon-Smits, E. A., Quinn, C. F., Tapken, W., Malagoli, M., and Schiavon, M. (2009). Physiological functions of beneficial elements. *Curr. Opin. Plant Biol.* 12, 267–274. doi: 10.1016/j.pbi.2009.04.009
- Pilon-Smits, E. A., and Leduc, D. L. (2009). Phytoremediation of selenium using transgenic plants. *Curr. Opin. Biotech.* 20, 207–212. doi: 10.1016/j.copbio.2009.02.001
- Quinn, C. F., Galeas, M. L., Freeman, J. L., and Pilon-Smits, E. A. (2007). Selenium: deterrence, toxicity, and adaptation. *Integr. Environ. Assess. Manag.* 3, 460–462. doi: 10.1002/ieam.5630030317
- Rayman, M. P. (2012). Selenium and human health. Lancet 379, 1256–1268. doi: 10.1016/S0140-6736(11)61452-9
- Robbins, R. J., Keck, A. S., Banuelos, G., and Finley, J. W. (2005). Cultivation conditions and selenium fertilization alter the phenolic profile, glucosinolate, and sulforaphane content of broccoli. J. Med. Food 8, 204–214. doi: 10.1089/jmf.2005.8.204
- Schiavon, M., Pilon-Smits, E.,A.,H., Citta, A., Folda, A., Rigobello, M. P., and Dalla Vecchia, F. (2016). Comparative effects of selenate and selenite on selenium accumulation, morphophysiology, and glutathione synthesis in *Ulva australis. Environ. Sci. Pollut. R.* 23, 15023–15032. doi: 10.1007/s11356-016-6649-6
- Seppanen, M. M., Kontturi, J., Heras, I. L., Madrid, Y., Camara, C., and Hartikainen, H. (2010). Agronomic biofortification of Brassica with seleniumenrichment of SeMet and its identification in Brassica seeds and meal. *Plant Soil* 337, 273–283. doi: 10.1007/s11104-010-0523-y
- Sperotto, R. A., Ricachenevsky, F. K., Williams, L. E., Vasconcelos, M. W., and Menguer, P. K. (2014). From soil to seed: micronutrient movement into and within the plant. *Front. Plant Sci.* 5:438. doi: 10.3389/fpls.2014. 00438
- Suarez, D. L., Grieve, C. M., and Poss, J. A. (2003). Irrigation method affects selenium accumulation in forage *Brassica* species. J. Plant Nutr. 26, 191–201. doi: 10.1081/PLN-120016504
- Terry, N., Zayed, A. M., De Souza, M. P., and Tarun, A. S. (2000). Selenium in higher plants. Annu. Rev. Plant Physiol. Plant Mol. Biol. 51, 401–432. doi: 10.1146/annurev.arplant.51.1.401
- Vamerali, T., Bandiera, M., Lucchini, P., Dickinson, N. M., and Mosca, G. (2014). Long-term phytomanagement of metal-contaminated land with field crops: Integrated remediation and biofortification. *Eur. J. Agron.* 53, 56–66. doi: 10.1016/j.eja.2013.11.008
- van der Ent, A., Baker, A. J. M., Reeves, R. D., Pollard, A. J., and Schat, H. (2012). Hyperaccumulators of metal and metalloid trace elements: facts and fiction. *Plant Soil* 362, 319–334. doi: 10.1007/s11104-012-1287-3

- White, P. J. (2016). Selenium accumulation by plants. Ann. Bot. 117, 217–235. doi: 10.1093/aob/mcv180
- White, P. J., and Broadley, M. R. (2009). Biofortification of crops with seven mineral elements often lacking in human diets-iron, zinc, copper, calcium, magnesium, selenium and iodine. *New Phytol.* 182, 49–84. doi: 10.1111/j.1469-8137.2008.02738.x
- White, P. J., and Brown, P. H. (2010). Plant nutrition for sustainable development and global health. *Ann. Bot.* 105, 1073–1080. doi: 10.1093/aob/mcq085
- White, P. J., Bowen, H. C., Marshall, B., and Broadley, M. R. (2007). Extraordinarily high leaf selenium to sulfur ratios define "Se-accumulator" plants. Ann. Bot. 100, 111–118. doi: 10.1093/aob/mcm084
- White, P. J., Bowen, H. C., Parmaguru, P., Fritz, M., and Spracklen, W. P. (2004). Interactions between selenium and sulphur nutrition in *Arabidopsis thaliana*. J. Exp. Bot. 55, 1927–1937. doi: 10.1093/jxb/erh192
- Wu, Z., Banuelos, G. S., Lin, Z. Q., Liu, Y., Yuan, L., Yin, X., et al. (2015). Biofortification and phytoremediation of selenium in China. *Front. Plant Sci.* 6:136. doi: 10.3389/fpls.2015.00136
- Ximenez-Embun, P., Alonso, I., Madrid-Albarran, Y., and Camara, C. (2004). Establishment of selenium uptake and species distribution in lupine, Indian mustard, and sunflower plants. J. Agric. Food Chem. 52, 832–838. doi: 10.1021/jf034835f
- Yuan, L., Zhu, Y., Lin, Z. Q., Banuelos, G., Li, W., and Yin, X. (2013). A novel selenocystine-accumulating plant in selenium-mine drainage area in Enshi, China. *PLoS ONE* 8:e65615. doi: 10.1371/journal.pone.0065615
- Zayed, A., Lytle, C. M., and Terry, N. (1998). Accumulation and volatilization of different chemical species of selenium by plants. *Planta* 206, 284–292. doi: 10.1007/s004250050402
- Zhang, Y. L., Pan, G. X., Chen, J., and Hu, Q. H. (2003). Uptake and transport of selenite and selenate by soybean seedlings of two genotypes. *Plant Soil* 253, 437–443. doi: 10.1023/A:1024874529957
- Zhao, L., Zhao, G. H., Zhao, Z. D., Chen, P., Du, S. T., Tong, J. Y., et al. (2004). Effect of selenium on the nutritional components of *Ganoderma lucidum*. Acta Botanica Boreali-Occidentalia Sinica 24, 1275–1280. doi: 10.1615/IntJMedMushrooms.v7.i3.690
- Zhao, Z. Q., Zheng, H. L., Zhang, C. G., and Ma, J. H. (2003). Advances in the studies on selenium in soil and selenium biological effect. *Chin. J. Ecol.* 22, 22–25. doi: 10.13292/j.1000-4890.2003.0007
- Zhu, Y. G., Pilon-Smits, E. A., Zhao, F. J., Williams, P. N., and Meharg, A. A. (2009). Selenium in higher plants: understanding mechanisms for biofortification and phytoremediation. *Trends Plant Sci.* 14, 436–442. doi: 10.1016/j.tplants.2009.06.006

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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