



A Novel L-ascorbate Peroxidase 6 Gene, ScAPX6, Plays an Important Role in the Regulation of Response to Biotic and Abiotic Stresses in Sugarcane

OPEN ACCESS

Edited by:

Sagadevan G. Mundree,
Queensland University of Technology,
Australia

Reviewed by:

Jian Li Yang,
Zhejiang University, China
Taras P. Pasternak,
Albert Ludwigs University of Freiburg,
Germany

*Correspondence:

Youxiong Que
queyouxiong@126.com
Yachun Su
syc2009mail@163.com

†These authors have contributed
equally to this work.

Specialty section:

This article was submitted to
Plant Biotechnology,
a section of the journal
Frontiers in Plant Science

Received: 25 July 2017

Accepted: 27 December 2017

Published: 17 January 2018

Citation:

Liu F, Huang N, Wang L, Ling H,
Sun T, Ahmad W, Muhammad K,
Guo J, Xu L, Gao S, Que Y and Su Y
(2018) A Novel L-ascorbate
Peroxidase 6 Gene, ScAPX6, Plays an
Important Role in the Regulation of
Response to Biotic and Abiotic
Stresses in Sugarcane.
Front. Plant Sci. 8:2262.
doi: 10.3389/fpls.2017.02262

Feng Liu^{1†}, Ning Huang^{1†}, Ling Wang¹, Hui Ling¹, Tingting Sun¹, Waqar Ahmad¹,
Khushi Muhammad², Jinxin Guo¹, Liping Xu¹, Shiwu Gao¹, Youxiong Que^{1,3*} and
Yachun Su^{1*}

¹ Key Laboratory of Sugarcane Biology and Genetic Breeding, Ministry of Agriculture, Fujian Agriculture and Forestry
University, Fuzhou, China, ² Department of Genetics, Hazara University, Mansehra, Pakistan, ³ Guangxi Collaborative
Innovation Center of Sugarcane Industry, Guangxi University, Nanning, China

The L-ascorbate peroxidase 6 gene (APX6) is one of the most important genes for scavenging H₂O₂ and plays a vital role in plant resistance to environmental stresses. In this study, a novel ScAPX6 gene (GenBank Accession No. KT907352) was obtained from a sugarcane variety (ROC22). Bioinformatics analysis showed that ScAPX6 has a cDNA length of 1,086 bp and encoded 333 amino acid residues. Subcellular localization confirmed that ScAPX6 was located in the chloroplast. Enhanced growth of *Escherichia coli* BL21 cells that expressed ScAPX6 showed high tolerance under copper (Cu) stress. Real-time quantitative PCR analysis revealed that ScAPX6 was constitutively expressed wherein with the highest expression levels in sugarcane pith and leaf and the lowest in the root. ScAPX6 was down-regulated by salicylic acid (SA), hydrogen peroxide (H₂O₂), polyethylene glycol (PEG) and sodium chloride (NaCl) stimuli. Interestingly, it was significantly up-regulated under the stresses of abscisic acid (ABA) and methyl jasmonate (MeJA) wherein with the highest inducible expression levels at 6 h at 6.0- and 70.0-times higher, respectively than that of control. Overexpression of ScAPX6 in *Nicotiana benthamiana* leaves enhanced the resistance to the infection of tobacco pathogens *Pseudomonas solanacearum* and *Fusarium solani* var. *coeruleum*. These results implied that ScAPX6 might positively respond to ABA, MeJA, and Cu, but might negatively respond to the stresses of SA, H₂O₂, PEG, and NaCl. Keeping in view the current investigation, ScAPX6 could be associated with the hypersensitive response (HR) or immunity of sugarcane, which will provide a baseline for the function identification of sugarcane ScAPX6.

Keywords: sugarcane, L-ascorbate peroxidase 6 gene, subcellular localization, biotic and abiotic stresses, transient overexpression

INTRODUCTION

In addition to *Oryza sativa*, *Triticum aestivum*, and *Zea mays*, sugarcane is the fourth largest staple food for the people of China. Sugarcane planting and production are of great significance in sugar supply (Li, 2000). However, the growth and development of sugarcane is severely affected by various abiotic and biotic stresses, such as drought, cold, salinity, heavy metals, high temperature, viruses, fungi, and so on (Li, 2000; Xu et al., 2008). As reported, environmental stimuli can induce active oxygen system which may cause injury to plant cells (Mittler et al., 2004, 2011). Peroxidases (EC number 1.11.1.x), including glutathione peroxidase (GPX), catalase (CAT), and ascorbate peroxidase (APX), are widespread in organisms and can remove the reactive oxygen (Shigeoka et al., 2002; Apel and Hirt, 2014). APX, belonging to type I heme peroxidase and copper oxidase family, is widely employed in plants and can rapidly scavenge hydrogen peroxide (H_2O_2) in the ascorbic acid (ASA) and glutathione (GSH) cycle (Shigeoka et al., 2002). The role of APX is highly specific to ascorbic acid, that is, to help electron donor to oxidation (Chen and Asada, 1989; Mittler, 2002; Foyer and Noctor, 2005).

According to the orientation characteristic, there are three mainly APX subfamilies in plants, such as cytoplasm APX (cAPX), thylakoid APX (tAPX), and APx-R (Apx-Related) (Mano et al., 1997; Shigeoka et al., 2002; Chew et al., 2003; Dunand et al., 2011). APXs gene have been reported in several plants such as *Nelumbo nucifera* (Chen et al., 2011), *Hordeum vulgare* (Shi et al., 2001), *Solanum tuberosum* (Kawakami et al., 2002), *Z. mays* (Breusegem et al., 1995), and *Vitis pseudoreticulata* (Lin et al., 2006). The expression of APX can be regulated by environmental stimuli, such as salt (Badawi et al., 2004), temperature (Kawakami et al., 2002), high light (Maruta et al., 2010), and heavy metal stresses (Pallavi and Dubey, 2007). Research has also shown that APX induced by adversity stress can regulate the content of H_2O_2 in the cell and redox signaling, and then affect plant tolerance to the osmotic stress (Andréia et al., 2012). This characteristic of APX enzyme activity may be treated as one of the physiological and biochemical indexes measuring crop resistance to biotic and abiotic stresses, for instance, water deficit (Nayyar and Gupta, 2006) and high temperature (Almeselmani et al., 2006). Korniyev et al. (2001) transferred the chloroplast APX gene into *Gossypium* spp., and it was found that APX activity in the transgenic cotton leaf was higher than that of the wild type. Overexpression of tAPX genes increased the resistance of *Nicotiana tabacum* and *Arabidopsis thaliana* to the oxidative stress induced by methyl violet essence (Yabuta et al., 2002; Murgia et al., 2004).

Until now, there are four nucleotide sequences of APX genes, which have been identified in sugarcane. Wang Z. Q. et al. (2015) indicated that the APX enzyme activity in sugarcane smut resistant variety Yacheng05-179 was significantly higher than the susceptible variety Liucheng03-182 after inoculated with *Sporisorium scitamineum*. As a result the expression level of one sugarcane ScAPX gene (GenBank Acc. No. KJ7565501) increased under the stresses of salicylic acid (SA), methyl jasmonate (MeJA), abscisic acid (ABA), H_2O_2 , sodium chloride

(NaCl), and polyethylene glycol (PEG). Wang S. et al. (2015) demonstrated that TAPX gene (GenBank Acc. No. JQ958327) played a part in sugarcane resistance to osmotic stress. Huang et al. (2013) found that sugarcane S-APX2 gene showed highly homologous with rice APX (GenBank Acc. No. XP_002463451.1) and maize APX (GenBank Acc. No. DAA41857.1). Another sugarcane APX gene (GenBank Acc. No. KX235995) was found in *Saccharum arundinaceum*, but its function was unclear.

From all the above, cloning the APXs gene of different isozymes is necessary to better understand the APX gene family and know more about their expression levels under different stress conditions. In the present study, a putative APX6 unigene, named as ScAPX6, was cloned and identified based on our previous transcriptome data of sugarcane in response to sorghum mosaic virus (SrMV) infection (Bioproject number: PRJNA379719). The sequence characters of ScAPX6 was analyzed by bioinformatics analysis, and the expression patterns of ScAPX6 gene after exposure to various stresses, such as ABA, MeJA, SA, H_2O_2 , PEG, NaCl, and copper (Cu), were detected by real-time quantitative polymerase chain reaction (qRT-PCR). Furthermore, its expression in *Escherichia coli*, subcellular localization and transient overexpression in *Nicotiana benthamiana* were also investigated. This study will be helpful to understand the gene function of ScAPX6 in sugarcane.

MATERIALS AND METHODS

Plant Material and Treatments

For the analysis of the tissue-specific expression of ScAPX6, six healthy plants of 10 months old of ROC22 were used, and then +1 leaf, root, bud, skin and pith were collected. The samples were fixed in liquid nitrogen and stored at $-80^{\circ}C$ until the extraction of total RNA.

For the abiotic treatment, uniform tissue cultured plantlets of ROC22 at the five or six leaf stage were transferred to water for one week and then treated by the following six stress conditions with root dipping of $5\text{ mmol}\cdot\text{L}^{-1}$ SA, $100\text{ }\mu\text{mol}\cdot\text{L}^{-1}$ MeJA, $100\text{ }\mu\text{mol}\cdot\text{L}^{-1}$ ABA, $10\text{ }\mu\text{mol}\cdot\text{L}^{-1}$ H_2O_2 , 25.0% PEG 8000, $250\text{ mmol}\cdot\text{L}^{-1}$ NaCl, and $100\text{ mmol}\cdot\text{L}^{-1}$ copper chloride ($CuCl_2$), at $28^{\circ}C$ with 16 h light and 8 h darkness (Su et al., 2014a). The whole plantlets under SA, MeJA, H_2O_2 , PEG, and NaCl stresses were harvested at 0, 6, 12, and 24 h, respectively. Another set of plantlets under Cu stress was harvested at 0, 12, 24, and 48 h, respectively. Three plantlets per time point were gathered and immediately fixed in liquid nitrogen, and stored at $-80^{\circ}C$ until the extraction of total RNA.

Total RNA Extraction and the First-strand cDNA Synthesis

Total RNA of the treated samples was extracted by Trizol[®] Reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's instructions. The first-strand cDNA synthesis was performed using Prime-Script[™] RT Reagent Kit (TaKaRa, Dalian, China) following manufacturer's instructions and tested by 1% agarose gel electrophoresis.

Sugarcane ScAPX6 Gene Isolation and Gateway Entry Vector Construction

The sequence of a putative *APX6* unigene (*ScAPX6*) from our previous transcriptome data of sugarcane in response to SrMV infection was used to design the cloning primer APX6-1F/1R (Table 1). The first-strand cDNA of ROC22 was used as amplification template. The reverse transcription–polymerase chain reaction (RT-PCR) procedure was 94°C for 4 min; 94°C for 30 s, 55°C for 30 s, 72°C for 2 min, 35 cycles; and 72°C for 10 min. RT-PCR products were gel-purified and cloned into pMD19-T vector (TaKaRa, Dalian, China), and then transformed into *E. coli* DH5 α competent cells and sequenced (Sangon, Shanghai, China).

The open reading frame (ORF) of *ScAPX6* with Gateway entry adapters attB1 and attB2 was amplified from the plasmid of pMD19-T-*ScAPX6* by the primers (APX6-3F/3R) (Table 1). The touchdown PCR procedure was 94°C for 4 min; 94°C for 30 s, 70°C for 30 s and then each loop drop 0.5°C, 72°C for 1 min and 30 s, 35 cycles; and 72°C for 10 min. The PCR amplification products were gel-purified and transformed into the Gateway[®] donor vector of pDONR221 (Invitrogen, USA) following the manufacturer's instructions of Gateway[®] BP Clonase[™] II Enzyme Mix (Invitrogen, USA). The mixture of BP reaction was transformed into DH5 α competent cells and sequenced (Sangon, Shanghai, China). The positive plasmid pDONR221-*ScAPX6* was achieved and then used for the constructions of prokaryotic expression vector and eukaryotic expression vector.

Bioinformatics Analysis

The ORF was translated and analyzed by ORF Finder (<https://www.ncbi.nlm.nih.gov/orffinder/>). Conserved domain of *ScAPX6* was predicted by the SMART program (<http://smart.embl-heidelberg.de/>) and NCBI Conserved Domains Database (CDD) (<http://www.ncbi.nlm.nih.gov/Structure/cdd/wrpsb.cgi>). The ExPASy tool (<http://us.expasy.org/tools>) was used to predict the basic physical and chemical properties of *ScAPX6*. The cleavage sites of the signal peptides were predicted by SignalP 4.1 Server (<http://www.cbs.dtu.dk/services/SignalP/>). Prediction of transmembrane helices in *ScAPX6* protein was performed by TMHMM Server v. 2.0 (<http://www.cbs.dtu.dk/services/TMHMM-2.0/>). Psort software was used to predict the subcellular localization of *ScAPX6*. GOR IV software (https://npsa-prabi.ibcp.fr/cgi-bin/npsa_automat.pl?page=/NPSA/npsa_gor4.html) was used to analyze the secondary structure of *ScAPX6*. The protein 3D model was predicted by SWISSMODEL software (<http://swissmodel.expasy.org/>). The homologous sequences of *ScAPX6* were obtained using Blastp in NCBI. DNAMAN software was used for the multiple sequence alignment. The phylogenetic tree of *ScAPX6* was constructed with amino acid sequences from other species by the neighbor-joining (NJ) method (1,000 bootstrap replicates) using the MEGA 6.06 (Saitou and Nei, 1987).

Subcellular Colocalization Assay

The ORF of *ScAPX6* was amplified by the primer APX6-4F/4R, and then was inserted into the *Bsa*I and *Eco*31I restriction sites of

the pBWA(V)HS-ccdb-GLogfp vector. Then the recombinant vector pBWA(V)HS-*ScAPX6*-GLogfp and the chloroplast marker vector were co-transformed in rice protoplasts with PEG solution (40% W/V PEG 4000, 0.2 mol·L⁻¹ mannitol and 0.1 mol·L⁻¹ calcium chloride). The mixture was cultured in dark for 30 min, and then the protoplasts was gathered and cultured in dark for 16–24 h. The method of transformation of rice protoplasts was followed by Datta and Datta (1999). The subcellular localization of the fusion protein was observed by a confocal laser scanning microscope Leica TCS SP5 (Germany).

Expression of ScAPX6 in E. coli BL21 (DE3) Strain

The plasmid of pDONR221-*ScAPX6* was digested with *Ava*I and then gel-purified for LR reaction with prokaryotic expressive vector of pEZYHb according to the manufacturer's instructions of LR Clonase[™] II Enzyme Mix (Invitrogen, USA). The recombinant plasmid of pEZYHb-*ScAPX6* was transformed into the competent cells *E. coli* BL21 (DE3) and then induced by 1.0 mmol·L⁻¹ isopropyl β -D-thiogalactoside (IPTG) at 28°C for 0, 2, 4, and 8 h (Guo et al., 2008). LB medium with *E. coli* BL21 (blank) and BL21+pEZYHb (control) were induced by 1.0 mmol·L⁻¹ IPTG for 0 and 8 h, respectively. The collected bacterial protein was analyzed by 12% sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE).

Spot assay was conducted to study the responses of *E. coli* BL21 cells expressing the *ScAPX6* gene under abiotic stress, such as NaCl, Cu and PEG. When OD₆₀₀ of *E. coli* BL21 cells in LB medium (containing 80 μ g·mL⁻¹ ampicillin) reached to 0.6, IPTG with a concentration of 1.0 mmol·L⁻¹ was added, and then the cells were grown at 37°C for 12 h. The cultures were diluted to OD₆₀₀ = 0.6, and then diluted to two levels of 10⁻³ and 10⁻⁴ (Guo et al., 2012). Ten microliters from each level was spotted on LB plates containing NaCl (250, 500, and 750 mmol·L⁻¹), CuCl₂ (250, 500, and 750 μ mol·L⁻¹) and PEG (15, 30, and 45%), respectively (Su et al., 2013). All plates were cultured in 37°C overnight and photographed.

Gene Expression Patterns of ScAPX6

SYBR Green Master (ROX) (Roche, China) and a 7500 qRT-PCR system (Applied Biosystems, South San Francisco, CA, USA) were applied to analyze gene expression levels of *ScAPX6* in different tissues and in response to various stresses. The sequence-specific primer of *ScAPX6* (APX6-2F/2R) (Table 1) was designed by Premier 5.0 software. The primer combination of clathrin adaptor complex (CAC) and cullin (*CUL*) (Table 1) was regarded as the internal control (Guo et al., 2014). The 20 μ L reaction system containing 10 μ L SYBR Green Master Mix, 0.8 μ L each of 10 μ mol·L⁻¹ upstream and downstream primers, 2 μ L cDNA templates (20 \times diluted cDNA) and 6.4 μ L double distilled water. Each qRT-PCR was conducted in triplicate. The qRT-PCR procedure was 50°C for 2 min; 95°C for 10 min; 35 cycles of 95°C for 15 s, and 60°C for 1 min. The 2^{- $\Delta\Delta$ Ct} method (Livak and

TABLE 1 | Primers used in this study.

Primer	Sequence information (5'–3')	Strategy
APX6-1F	CTTGAGAAGGCAAGCCAGGA	Gene cloning
APX6-1R	CGAGACACTGGTACAGGGGA	Gene cloning
APX6-2F	GATTTGATTGCCGTGGCTGG	qRT-PCR analysis
APX6-2R	TCTTCAGGAAGTTTGCCAGTTG	qRT-PCR analysis
CUL-F	TGCTGAATGTGTTGAGCAGC	qRT-PCR analysis
CUL-R	TTGTCGCGCTCCAAGTAGTC	qRT-PCR analysis
CAC-F	ACAACGTCAGGCAAAGCAAA	qRT-PCR analysis
CAC-R	AGATCAACTCCACCTCTGCG	qRT-PCR analysis
APX6-3F	<u>GGGGACAAGTTTGTACAAAAAAGCAGGCTTCATGGAGCTCACCAACATCCC</u>	Gateway entry vector construction and RT-PCR analysis
APX6-3R	<u>GGGGACCACCTTTGTACAAAGAAAGCTGGGTCAGCTGTTCTCCACGAGGCTC</u>	Gateway entry vector construction and RT-PCR analysis
APX6-4F	CAGTGGTCTCACAACATGGAGCTCACCAACATCCC	Subcellular localization vector construction
APX6-4R	CAGTGGTCTCATAAAGCTGTTCTCCACGAGGCTC	Subcellular localization vector construction
NtHSR201-F	CAGCAGTCTTTGGCGTTGTC	qRT-PCR analysis
NtHSR201-R	GCTCAGTTTAGCCGAGTTGTG	qRT-PCR analysis
NtHSR203-F	TGGCTCAACGATTACGCA	qRT-PCR analysis
NtHSR203-R	GCACGAAACCTGGATGG	qRT-PCR analysis
NtHSR515-F	TTGGGCAGAATAGATGGGTA	qRT-PCR analysis
NtHSR515-R	TTTGGTGAAAGTCTTTGGCTC	qRT-PCR analysis
NtPR-1a/c-F	AACCTTTGACCTGGGACGAC	qRT-PCR analysis
NtPR-1a/c-R	GCACATCCAACACGAACCGA	qRT-PCR analysis
NtPR2-F	TGATGCCCTTTTGGATTCTATG	qRT-PCR analysis
NtPR2-R	AGTTCCTGCCCGCTTT	qRT-PCR analysis
NtPR3-F	CAGGAGGGTATTGCTTTGTTAGG	qRT-PCR analysis
NtPR3-R	CGTGGGAAGATGGCTTGTGTGC	qRT-PCR analysis
NtEFE26-F	CGGACGCTGGTGGCATAAT	qRT-PCR analysis
NtEFE26-R	CAACAAGAGCTGGTGGCTGGATA	qRT-PCR analysis
NtAcCoDeaminase-F	TCTGAGTTACTGATTTGGATTGG	qRT-PCR analysis
NtAcCoDeaminase-R	TGGACATGGTGGATAGTTGCT	qRT-PCR analysis
NtEF1- α -F	TGCTGCTGTAACAAGATGGATGC	qRT-PCR analysis and RT-PCR analysis
NtEF1- α -R	GAGATGGGGACAAAGGGGATT	qRT-PCR analysis and RT-PCR analysis

attB1 and *attB2* adapters were underlined in the forward primer APX6–3F and in the reverse primer APX6–3R, respectively.

Schmittgen, 2001) was employed to analyze the qRT-PCR data.

Transient Overexpression of ScAPX6 in *N. benthamiana*

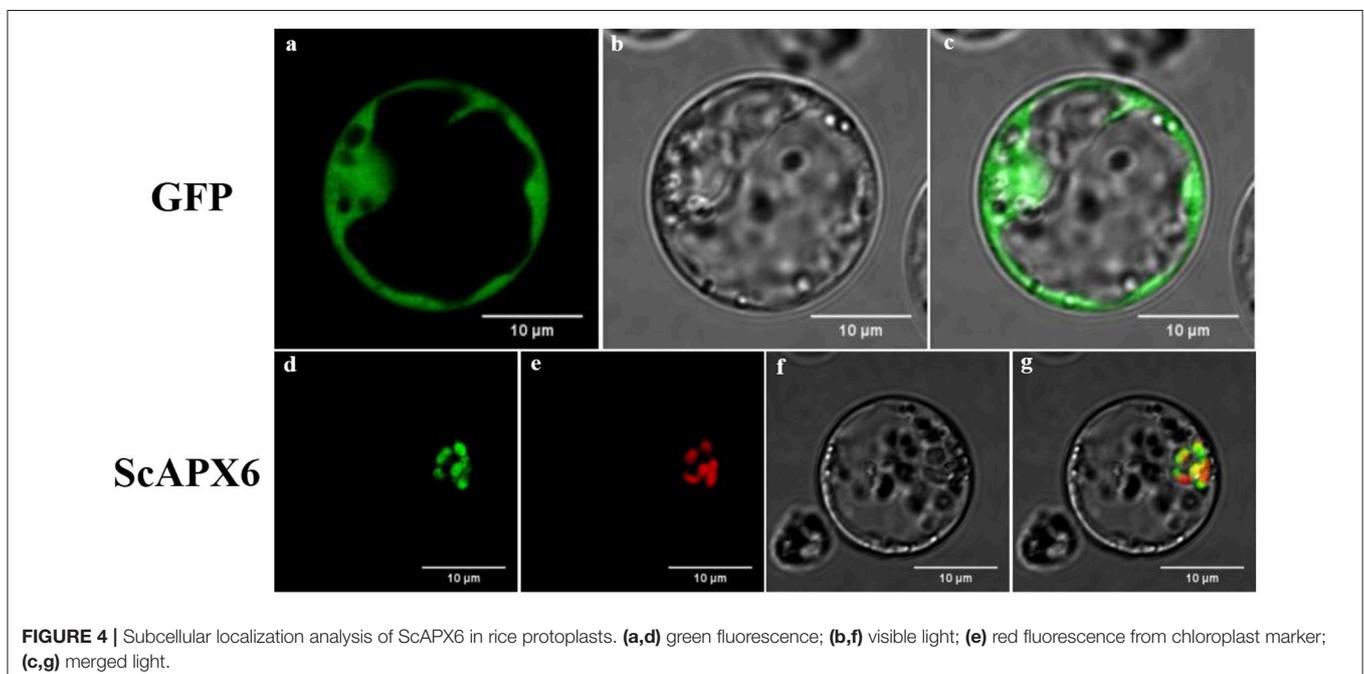
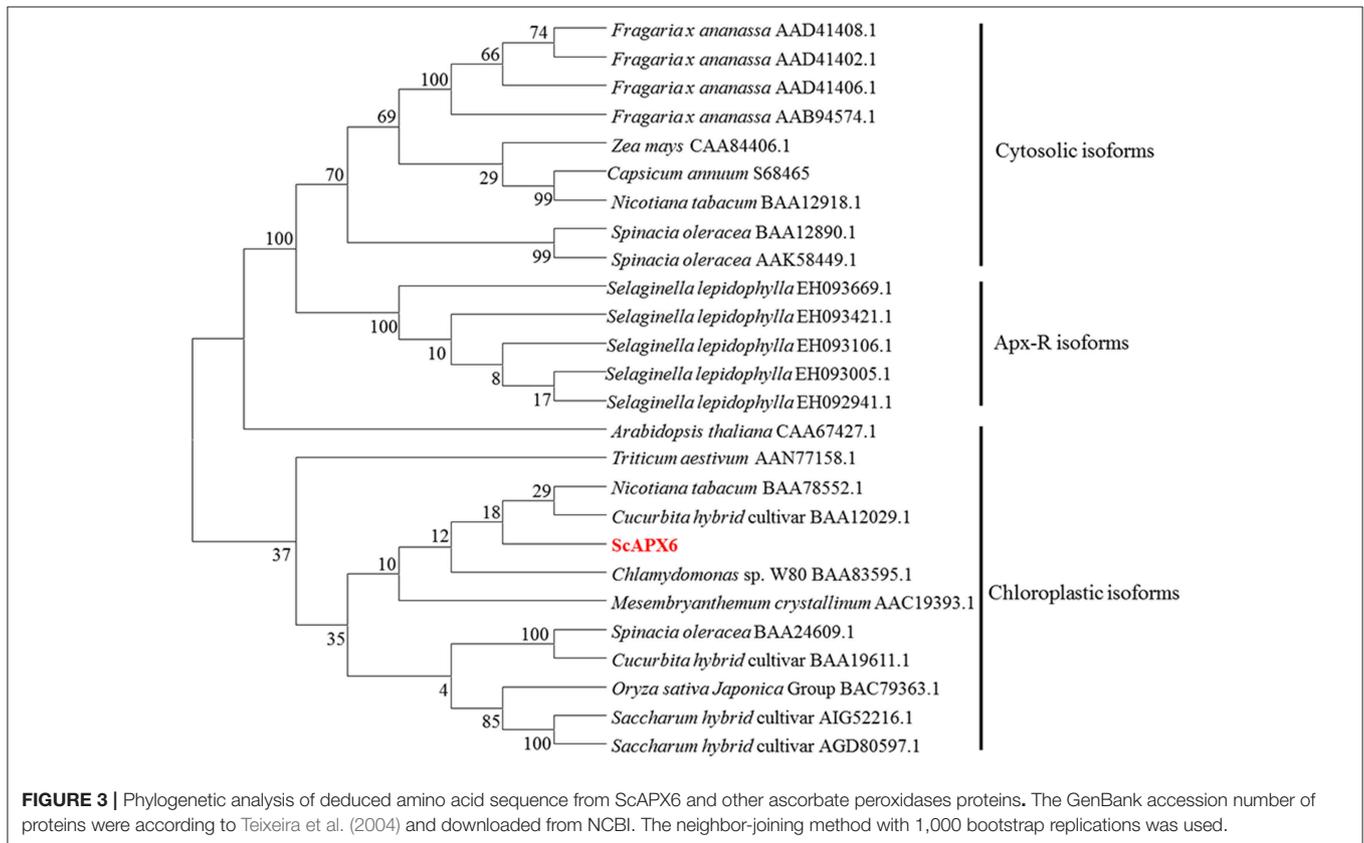
To study the role of ScAPX6 in response to pathogen infection and its hypersensitive reaction in plant, an overexpressed vector pEarleyGate 203-ScAPX6 was constructed by Gateway cloning technique according to the manufacturer's instructions of LR ClonaseTM II Enzyme Mix (Invitrogen, USA). *N. benthamiana* leaves was inoculated with the vector of pEarleyGate 203-ScAPX6 by an *Agrobacterium*-mediated transient expression method conducted by Su et al. (2014b). Two important tobacco pathogens, *Pseudomonas solanacearum* and *Fusarium solani* var. *coeruleum*, were cultured in potato dextrose water (PDW) liquid medium at 28°C. When the two pathogens cells were cultured to an OD₆₀₀ of 0.8, they were separately infected into the treated leaves that were agroinfiltrated with pEarleyGate 203-ScAPX6 for 24 h. Then the 3,3'-diaminobenzidine (DAB) staining, trypan blue staining and transcript analysis of the

eight tobacco immunity-associated marker genes (Table 1), were conducted by the treated *N. benthamiana* leaves according to Su et al. (2016). RT-PCR was used to detect whether ScAPX6 has been overexpressed in *N. benthamiana*, with the RNA of treated leaves and ScAPX6 specific primer (APX6-3F/3R, Table 1), the *NtEF1- α* was treated as control. RT-PCR procedure was 94°C for 4 min; 94°C for 30 s, 72°C for 30 s, 72°C for 2 min, 35 cycles; and 72°C for 10 min. All treatment materials were cultured at 24°C (16 h light/8 h darkness) and then photographed at 1 and 7 day (d) separately. Each test was repeated three times.

DAB and trypan blue staining were used for histochemical analysis of *Agrobacterium*-infiltrated leaves. The leaves was soaked in DAB-HCl solution (1.0 mg·mL⁻¹, pH 5.8), and then cultured in the dark for 12 h. The leaves were destained with 95% ethanol at 100°C for 10 min (Su et al., 2014a), and then was imaged for H₂O₂ detection with a stereoscopic microscope (Nikon, Tokyo, Japan) and a light microscope (Leica, Wetzlar, Germany). Three biological replicates were prepared. The leaves were also deal with

residues. ScAPX6 had a molecular mass of 36.21 kDa and an isoelectric point (pI) of 6.91. CDD search of NCBI showed that ScAPX6 belonged to a member of the plant-peroxidase-like

superfamily (**Figure 1**). The instability index of ScAPX6 protein was 48.10, suggesting that ScAPX6 might be an unstable acid hydrophilic protein (Walker, 2005). Secondary structure



prediction of ScAPX6 predicted that the percentages of alpha-helix, random coil, and extended strand were 44.74, 42.94, and 12.31%, respectively.

Furthermore, SWISSMODEL program showed that the main spatial structures of ScAPX6 were alpha-helix and random coil (Figure 2). Comparing ScAPX6 with *O. sativa* Japonica Group APX6 (EAZ43377.1), *S. italic* APX6 (XP_004973913.1), and *S. bicolor* APX6 (XP_002445876.1), we found that the spatial structure of these four APX6 was basically in line with each other, suggesting that ScAPX6 owned high conservation of spatial structure with different plant species. Psort software predicted that ScAPX6 might be located in the chloroplast thylakoid membrane, plasma membrane, chloroplast stroma, and chloroplast thylakoid space with the probabilities of 71.9, 65.0, 56.1, and 56.1%, respectively.

According to the classification method by Teixeira et al. (2004), the phylogenetic tree was separated into three groups, including cytosolic isoforms, Apx-R isoforms and chloroplastic isoforms (Figure 3). ScAPX6 was clustered into group chloroplastic isoforms. Two APX proteins reported in *Saccharum* hybrid cultivar, which were ScAPX (AIG52216.1) and TAPX (AGD80596.1), were also clustered into the same clade as ScAPX6.

Subcellular Localization

The recombinant vector pBWA(V)HS-ScAPX6-GLogfp was constructed to understand the subcellular location of ScAPX6. The results showed that ScAPX6 and the chloroplast marker were located in the same place, so it was confirmed that ScAPX6 was located in the chloroplast, which is in accordance with the results of prediction (Figure 4).

Expression of ScAPX6 in *E. coli* BL21 (DE3) Strain

The ScAPX6 gene was combined with the expression vector pEZYHb and then was transformed into *E. coli* BL21 cell. The SDS-PAGE analysis (Figure 5) showed that ScAPX6 was expressed as a recombinant protein in the BL21 cells. In Figure 5, after induced by 1.0 mmol·L⁻¹ IPTG at 28°C for 2, 4, and 8 h, an obvious accumulation protein (including the 6× His-tag) at approximate 55 kDa was observed.

Overexpression of ScAPX6 in *E. coli* Enhances Cell Growth under Cu Stress

It has been reported that the APX activity was up-regulated by abiotic stress, such as heavy metal, high salinity, drought, high temperature, and wounding (Shi et al., 2001). In this study, spot assay was performed to study the function of ScAPX6 in response to abiotic stress *in vivo*. The control (BL21+pEZY-Hb) and the gene-expressed cells (BL21+pEZY-Hb-ScAPX6) grew in LB plates containing NaCl, CuCl₂, and PEG were performed (Figure 6). It had been recorded that the recombinant ScAPX6 cells showed a more rapid growth than the control in LB plates with PEG and CuCl₂ supplement, but not with NaCl, suggesting that the overexpression of ScAPX6 in *E. coli* could enhance its tolerance to PEG and Cu stress.

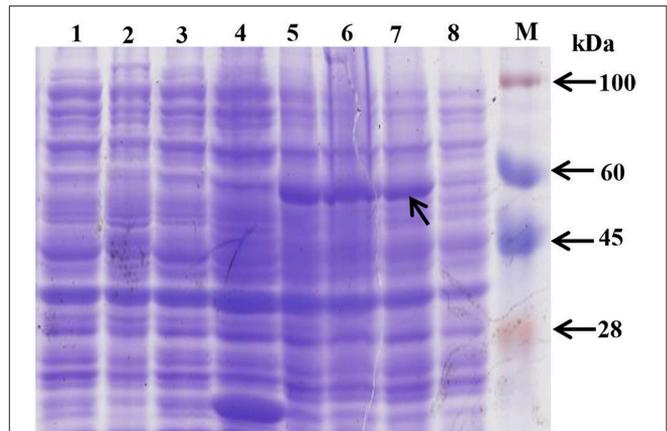


FIGURE 5 | Prokaryotic expression of pEZY-Hb-ScAPX6 fusion protein in *Escherichia coli* BL21 (DE3). M, protein marker; 1, BL21 cell induction for 8 h; 2, BL21 cell without induction; 3, pEZY-Hb without induction; 4, pEZY-Hb induction for 8 h; 5–8, pEZY-Hb-ScAPX6 induction for 8, 4, 2, and 0 h, respectively.

Tissue-Specific Expression of ScAPX6

qRT-PCR analysis showed that ScAPX6 was constitutively expressed in all five kinds of sugarcane tissues, including root, bud, skin, leaf, and pith, but with different expression levels (Figure 7). ScAPX6 showed the highest expression levels in the pith and leaf, and then on the skin, while the transcript in root was at the lowest level.

Gene Expression Patterns of ScAPX6 in Response to Abiotic Stress

qRT-PCR analysis revealed that the ScAPX6 gene exhibited different expression characteristics in response to ABA, MeJA, SA, H₂O₂, PEG, NaCl, and Cu stimuli (Figures 8A,B). As shown in Figure 8A, the transcripts of ScAPX6 were remarkably up-regulated under the stresses of ABA and MeJA, and with the highest inducible expression levels at 6 h, which were 6.0- and 70.0-times higher than that of control, respectively. However, ScAPX6 was down-regulated during the SA treatment and rapidly decreased at 6 h. Under the stress of PEG, the expression of ScAPX6 showed no change at 6 h, and then decreased at 12 h. ScAPX6 was down-regulated after the treatment of H₂O₂ and NaCl, but was up-regulated by the CuCl₂. These results demonstrated that ScAPX6 might positively respond to ABA, MeJA, and Cu stresses but negatively respond to SA, H₂O₂, PEG, and NaCl stresses.

Transient Overexpression of ScAPX6 Induces a Defense Response in *N. benthamiana*

After transient overexpression of ScAPX6 in *N. benthamiana* leaves for 1 day (d), the transcripts of ScAPX6 were detected by qRT-PCR (Figure 9A). As shown in Figures 9A,B darker DAB staining color and more intense trypan blue staining cells were observed in ScAPX6 leaves than that in the control (35S::00) after infiltration for 2 and 6 d, respectively. The

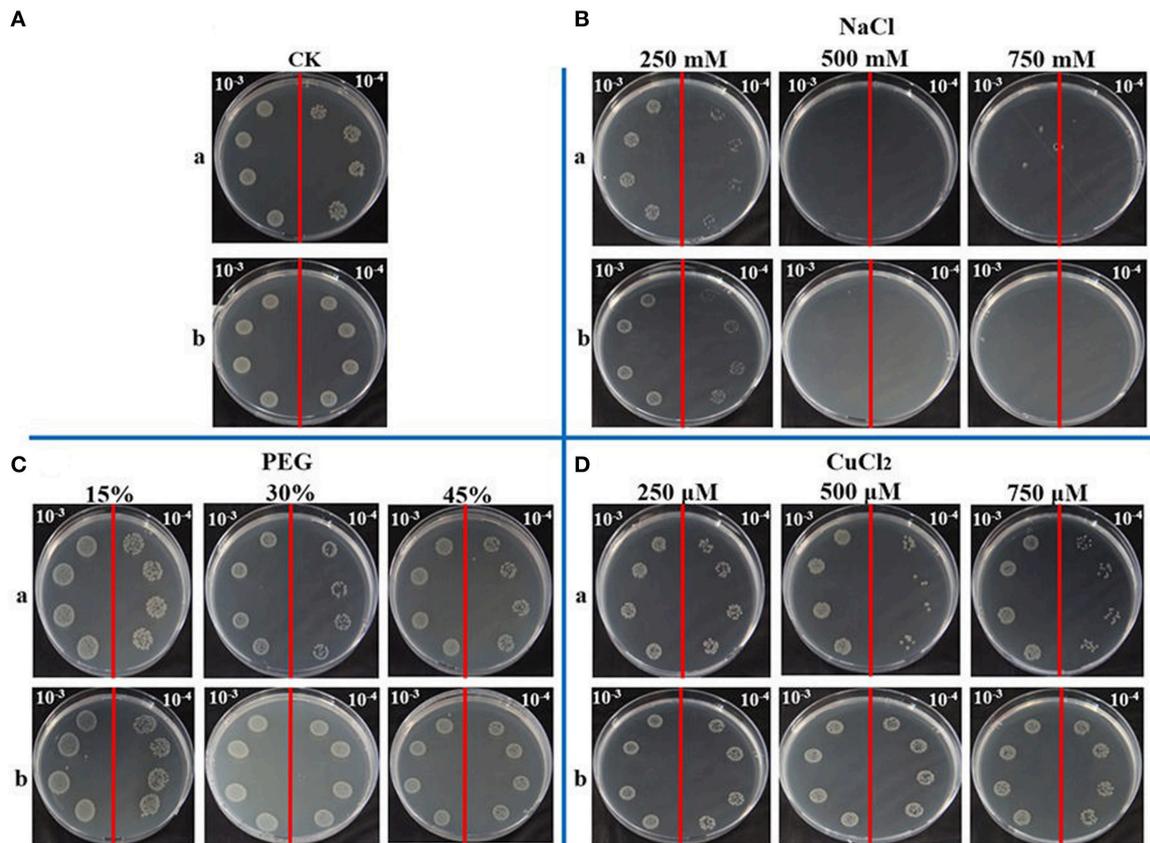


FIGURE 6 | Spot assays of BL21+pEZY-Hb-ScAPX6 (**b**) and BL21+pEZY-Hb (control) (**a**) on LB plates with NaCl, PEG and CuCl₂. Isopropyl β-D-thiogalactoside (IPTG) was added to the cultures of BL21+pEZY-Hb-ScAPX6 and BL21+pEZY-Hb to induce the expression of recombinant protein. The cultures were adjusted to OD₆₀₀ = 0.6. Ten microliters from 10⁻³ (left side of the red line on the plate) to 10⁻⁴ (right side of the red line on the plate) dilutions were spotted onto LB plates without any supplement (CK) (**A**) or with NaCl (250, 500, and 750 mmol·L⁻¹) (**B**), PEG (15, 30, and 45%) (**C**) and CuCl₂ (250, 500, and 750 μmol·L⁻¹) (**D**), respectively. NaCl, sodium chloride; PEG, polyethylene glycol; CuCl₂, copper chloride.

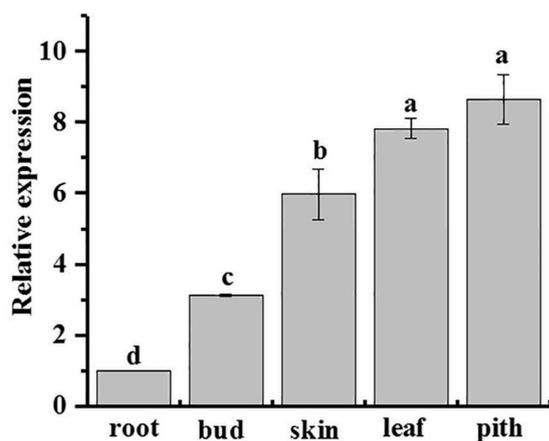
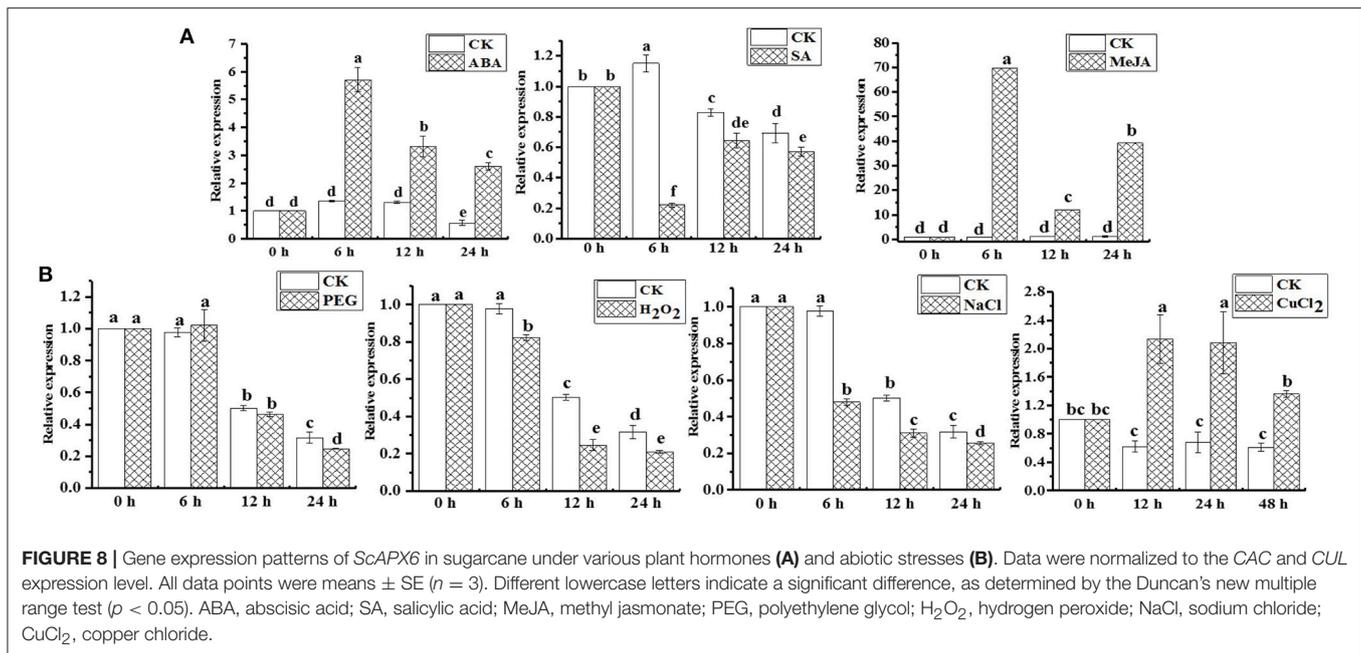


FIGURE 7 | Tissue-specific expression analysis of ScAPX6 in sugarcane. The error bars represented the standard error of each treating group ($n = 3$). Data were normalized to the CAC and CUL expression level. All data points were means \pm SE ($n = 3$). Different lowercase letters indicate a significant difference, as determined by the Duncan's new multiple range test ($p < 0.05$).

eight immunity-associated marker genes in *N. benthamiana* were induced by transient overexpression of ScAPX6 (Figure 9C). The hypersensitive response (HR) marker genes, *NtHSR201* and *NtHSR203*, showed no change in transcript, while *NtHSR515* was up-regulated. The expression level of SA-responsive gene *NtPR2* remained unchanged, while *NtPR-1a/c* and *NtPR3* and two ethylene synthesis dependent genes, *NtEFE26* and *NtAccdeaminase*, were all down-regulated. Compared with the control leaves, 35S::ScAPX6 exhibited darker color reflecting high levels of H₂O₂ accumulation and intense hypersensitivity response.

To further investigate the response of ScAPX6 to pathogen, two tobacco pathogens, *P. solanacearum* and *F. solani* var. *coeruleum*, were separately injected into *N. benthamiana* containing 35S::ScAPX6 or the control. After inoculation with *P. solanacearum*, no disease symptom was found between 35S::ScAPX6 and the control leaves for 1 d, while 35S::ScAPX6 exhibited darker color than the control at 1 d by DAB staining. With elongated treatment time, although the DAB staining showed no difference between 35S::ScAPX6 and the



control leaves, the leaves in the control showed slight yellow phenomenon and necrotic spot, while the 35S::*ScAPX6* only exhibited the faint wilting symptom after inoculation at 7 d (Figure 9D). After challenging with *P. solanacearum* for 1 and 7 d (Figure 9E), the expression levels of *NtHSR201*, *NtHSR203*, and *NtPR2*, were unchanged or down-regulated in the control and 35S::*ScAPX6* leaves. The transcripts of *NtPR-1a/c* and *NtPR3* were significantly down-regulated at 1 d and increased at 7 d in the control leaves, but were down-regulated or remained unchanged in the 35S::*ScAPX6* leaves. The expression levels of *NtHSR515* and *NtEFE26* were unchanged at 1 and 7 d and *NtAccdeaminase* was up-regulated at 7 d in the control leaves, while these three genes in the 35S::*ScAPX6* leaves were all unchanged at 1 d and reached the peak values at 7 d after inoculation.

Likewise, for DAB staining, the 35S::*ScAPX6* leaves showed darker color than the control after inoculation with *F. solani* var. *coeruleum* for 1 d and exhibited no difference at 7 d. No apparent disease symptom differences between 35S::*ScAPX6* and the control were found at 1 d after inoculation (Figure 9F). Some symptoms, such as wilting, decay phenomenon and necrotic spot, were observed in the control leaves at 7 d, but not in the 35S::*ScAPX6* leaves. After challenging with *F. solani* var. *coeruleum* for 1 and 7 d (Figure 9G), the expression levels of *NtHSR201* and *NtPR2* were unchanged or down-regulated in the control and 35S::*ScAPX6* leaves. The transcripts of *NtAccdeaminase*, *NtHSR203*, and *NtHSR515*, were significantly up-regulated at 1 or 7 d in the control leaves, while stayed stable or showed a small rise in the 35S::*ScAPX6* leaves. The transcripts of *NtPR-1a/c*, *NtPR3*, and *NtEFE26*, were down-regulated or remained unchanged in the control leaves, but were all significantly up-regulated in the 35S::*ScAPX6* leaves.

DISCUSSION

Plant APXs are a multi-gene family (Shigeoka et al., 2002). Many plants APXs genes have been cloned and identified, including eight in *O. sativa* (Teixeira et al., 2005, 2006), four in *Vigna unguiculata* (D'Arcylameta et al., 2006) and *Spinacia oleracea*, respectively (Ishikawa et al., 1995, 1996, 1998), six in *Eucalyptus grandis* (Teixeira et al., 2005), seven in *Lycopersicon esculentum* (Najami et al., 2008), and nine in *A. thaliana* (Panchuk et al., 2002; Mittler et al., 2004; Narendra et al., 2006). In the present study, based on a putative APX6 unigene sequence from our previous transcriptome data, a sugarcane *ScAPX6* gene (GenBank Acc. No. KT907352) was cloned (Figure S1), which was different from the other already reported sugarcane APX genes in NCBI (*ScAPX*: GenBank Acc. No. KJ7565501; *TAPX*: GenBank Acc. No. JQ958327; *APX*: GenBank Acc. No. KX235995), and shared only 19.79% identity at the amino acid sequence level. Najami et al. (2008) found that in *Solanum lycopersicum* three cytosolic *SlAPX* genes, *SlAPX1*, *SlAPX2*, and *SlAPX3* showed a high sequence identity (>90%). Teixeira et al. (2004) found that APX contained two isoforms, chloroplastic and nonchloroplastic isoforms. Furthermore, APX-R was found as a new heme-containing protein functionally associated with ascorbate peroxidase (Lazzarotto et al., 2011). In this study, *ScAPX6* was clustered in chloroplastic isoforms (Figure 3), which was consistent with the study conducted by Teixeira et al. (2004). Subcellular localization of *ScAPX6* in rice protoplast showed that *ScAPX6*::GFP was targeted at chloroplast (Figure 4), which was consistent with the result of bioinformatics predicted localization. Similar to other plant species, such as the APXs from *Cucumis melo* (Cheng et al., 2009) and *A. andraeanum* (Liu et al., 2013), *ScAPX6* also contained a plant peroxidase like superfamily and the heme binding site and

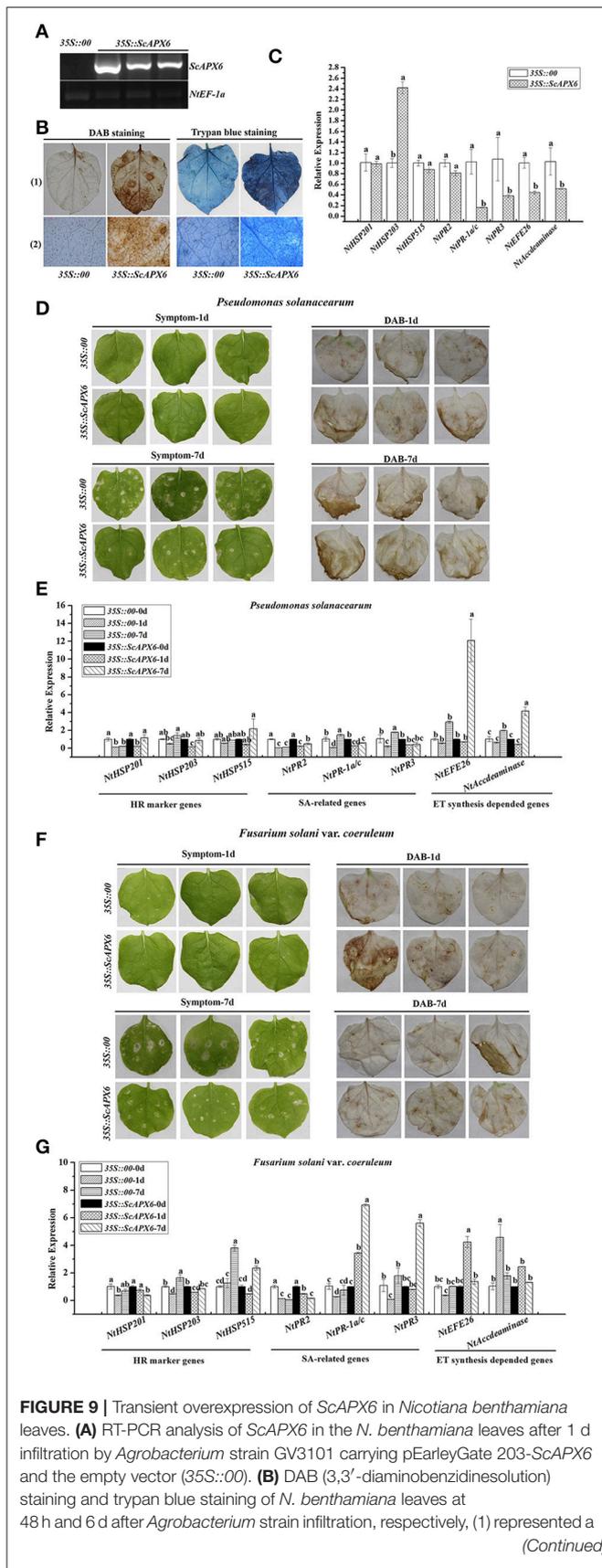


FIGURE 9 | stereoscopic microscope and (2) represented a light microscope. **(C)** The transcripts of eight immunity-associated marker genes in the *N. benthamiana* leaves at 24 h after infiltration. **(D, F)** Disease symptoms and DAB staining results of *N. benthamiana* leaves by *P. solanacearum* and *F. solani* var. *coeruleum* infection after infiltration with 35S::00 (control) or 35S::ScAPX6-containing *Agrobacterium* strain. Disease symptoms of infected leaves were observed at 1 and 7 d post-inoculation. **(E, G)** The transcripts of immunity-associated marker genes in the *N. benthamiana* leaves after inoculation with *P. solanacearum* or *F. solani* var. *coeruleum* for 1 and 7 d. *NtEF1-α* was used for normalization of the transcript levels. All data points were expressed as the mean \pm SE ($n = 3$). Different lowercase letters indicate a significant difference, as determined by the Duncan's new multiple range test ($p < 0.05$). *NtHSR201*, *NtHSR203*, and *NtHSR515*, hypersensitive response marker genes; *NtPR2*, *NtPR-1a/c*, and *NtPR3*, a salicylic acid pathway-related gene; *NtEFE26* and *NtAccdeaminase*, the ethylene synthesis-dependent genes. Control, the *Agrobacterium* strain carrying 35S::00.

shared 94.29 and 82.93% similarities with the APX homologs from *S. italic* APX6 (XP_004973913.1) and *S. bicolor* APX6 (XP_002445876.1), suggesting that ScAPX6 belongs to a member of APX family.

APX plays an important role in response to biotic and abiotic stresses (Andréia et al., 2012). Cheng et al. (2009) observed that in *C. melo*, the gene expression level of *CmAPX* varied in different tissues, and with the highest expression in leaves and roots. Chen et al. (2011) found that *NuAPX* showed higher expression levels in leaf stalks than in root, due to the reasons that the tissues of the leaf stalks and leaf were rich in chloroplast and mitochondria, which was the leading source of reactive oxygen species (ROS) through the electron-transport chain of photosynthesis. In this study, ScAPX6 was constitutively expressed in sugarcane tissues and with the highest expression in pith but the lowest in root (Figure 7).

For abiotic stress, Agarwal et al. (2005) have found that the enzyme activities of APX, SOD, and CAT in wheat seedlings could be increased by 500 $\text{mmol}\cdot\text{L}^{-1}$ ABA treatment. It was shown previously that in *H. vulgare*, the transcript level of *HvAPX1* was remarkably up-regulated by the treatments of ABA and NaCl (Shi et al., 2001). In *Brassica oleracea* var. *italica*, Jiang et al. (2012) have found that the expression of *BoAPX2* increased after H_2O_2 , SA, and NaCl treatments. Previous research on sugarcane showed that the transcripts of ScAPX increased under the treatment of ABA, MeJA, SA, H_2O_2 , PEG, and NaCl (Wang Z. Q. et al., 2015). In this study, the expression of ScAPX6 was also up-regulated by both ABA and MeJA, but down-regulated by SA and H_2O_2 treatments (Figure 8A). As reported, the expression of *TAPX* gene in sugarcane was significantly induced by NaCl and PEG stresses (Wang S. et al., 2015). Previous investigations have identified the transcripts of *OsAPX7* and *OsAPX8*, which were separately down regulated by 300 $\text{mmol}\cdot\text{L}^{-1}$ NaCl in rice roots and leaves (Hong and Kao, 2007; Yamane et al., 2010). Similarly, in this study, down regulation of the transcripts of ScAPX6 were noted with NaCl and PEG treatment (Figure 8B), which was consistent with the results of the spot assay that the recombinant protein of ScAPX6 expressed in *E. coli* BL21 did not show better growth than the control under both two

treatments (Figure 6). Previous reports have provided evidence that over-expressed plant stress tolerance genes in *E. coli* cells could enhance their growth under abiotic stress (Gupta et al., 2010; Guo et al., 2013). For example, Su et al. (2014b) have tested a chitinase gene *ScChi* in *E. coli*, which showed better growth under NaCl, Cu, CdCl₂, and ZnSO₄ treatments. Duan et al. (2006) have indicated that the transgenic *O. sativa* with *HvAPX1* gene was more tolerant to cadmium stress when compared with the wild type. In this study, the transcript of *ScAPX6* was also up-regulated by the treatment of Cu (Figure 8B), which was in line with the results that the recombinant protein of *ScAPX6* expressed in *E. coli* BL21 resulted in a better growth under Cu stress (Figure 6). Therefore, it was predicted that *ScAPX6* could be helpful for the tolerance of sugarcane to Cu. These findings suggested *ScAPX6* might be a positive response to ABA, MeJA, and Cu stresses, while showed the negative response to SA, H₂O₂, PEG, and NaCl stresses. However, what should be pointed out here is that, we only use treatment with some hormones, such as SA, ABA, and MeJA with one concentration, and we cannot directly link effect of our treatment with specific hormone pathway because the specific concentration of the hormones was not determined. These points need to be considered in future.

Previous studies have revealed that the overexpression of *tApx* gene in tobacco enhanced tolerance to chilling, methylviologen, and high-intensity light (Yabuta et al., 2002). Transgenic potato with simultaneous overexpression of *APX*, choline oxidase (*codA*), and *SOD*, increased tolerance of SSAC plants and lower levels of H₂O₂ under methylviologen, drought and salt-mediated oxidative stresses (Ahmad et al., 2010). Investigations have revealed that cell death could induce R gene expression, ion fluxes, stimulation of ROS, and defense-related hormones, which can efficiently restrict pathogen growth and development (Li et al., 2010; Melech-Bonfil and Sessa, 2010; Du et al., 2012). Thordal-Christensen et al. (1997) proved that DAB-uptake method can serve H₂O₂ detection at a subcellular level. Although the DAB reactions reflect increases in local H₂O₂, this method has been adapted to many other plant species for in situ detection of H₂O₂. Thordal-Christensen et al. (1997) speculated that this could be related either to the fact that the optimal pH (5.5–6.0) for DAB precipitation coincides with the expected pH level in the leaf, or to the strong polymerization observed in the plant tissue. Lai et al. (2013) found that *35S::BrERF11* transgenic tobacco plants showed significantly increased HR and H₂O₂ accumulation compared with wild-type plants according to trypan blue and DAB staining. In the present study, a darker DAB staining color was found after overexpression of *ScAPX6* in *N. benthamiana* leaves and after inoculation with *P. solanacearum* and *F. solani* var. *coeruleum* at 1 d compared with that in the leaves of control (Figures 9B,D,F), which was indicative of the accumulation of H₂O₂ and resulted in intense hypersensitivity response, but with elongated treatment time, the DAB staining color became lighter, which was consistent with the fact that APX can rapidly scavenge H₂O₂ in the plant by the ASA-GSH cycle (Shigeoka et al., 2002). However, since the DAB buffer is far

from real physiological conditions, whether itself results in the accumulation of H₂O₂ remains an open question. Together, the antimicrobial action against the tobacco pathogens after overexpression of *ScAPX6* in *N. benthamiana*, suggesting that *ScAPX6* may enhance the resistance to *P. solanacearum* and *F. solani* var. *coeruleum*.

CONCLUSIONS

In this study, a novel ascorbate peroxidase gene, *ScAPX6* (GenBank Acc. No. KT907352), was isolated and characterized. The cDNA of *ScAPX6* gene was 1,086 bp long with a complete 1,002 bp ORF, encoding 333 amino acids. Subcellular localization revealed that *ScAPX6* was targeted in chloroplast. After inducing by IPTG, the accumulation protein of pEZYHb-*ScAPX6* at 55 KDa led to a better growth of *E. coli* BL21 under Cu stress. *ScAPX6* was constitutively expressed in sugarcane tissues. Besides, *ScAPX6* showed positive response to ABA, MeJA, and Cu stresses, but negative response to the stresses of SA, H₂O₂, PEG, and NaCl. The overexpression of *ScAPX6* in *N. benthamiana* leaves showed positive response against the attack of *P. solanacearum* and *F. solani* var. *coeruleum*. These results suggested that *ScAPX6* plays an important role in the HR or immunity of sugarcane.

AUTHOR CONTRIBUTIONS

FL, YS, and YQ: Conceived, designed, and initiated the project; NH, HL, and SG: Prepared materials; FL, NH, LW, TS, WA, and JG: Performed experiments and contributed to data analysis and validation; FL and YS: Drafted the manuscript; LX, KM, YS, and YQ: Revised the manuscript. All authors read and approved the final manuscript.

ACKNOWLEDGMENTS

This work was supported by Natural Science Foundation of Fujian province, China (2015J06006), the National Natural Science Foundation of China (31501363), the Research Funds for Distinguished Young Scientists in Fujian Agriculture and Forestry University (xjq201630), the Research Funds for Distinguished Young Scientists in Fujian Provincial Department of Education, the earmarked fund for China Agricultural Research System (CARS-17) and the Program for New Century Excellent Talents in Fujian Province University (JA14095).

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpls.2017.02262/full#supplementary-material>

Figure S1 | Amplification of *ScAPX6* gene in sugarcane. M: DNA marker 2,000 bp; 1: RT-PCR product.

REFERENCES

- Agarwal, S., Sairam, R. K., Srivastava, G. C., Tyagi, A., and Meena, R. C. (2005). Role of ABA, salicylic acid, calcium and hydrogen peroxide on antioxidant enzymes induction in wheat seedlings. *Plant Sci.* 169, 559–570. doi: 10.1016/j.plantsci.2005.05.004
- Ahmad, R., Kim, Y. H., Kim, M. D., Kwon, S. Y., Cho, K., Lee, H. S., et al. (2010). Simultaneous expression of choline oxidase, superoxide dismutase and ascorbate peroxidase in potato plant chloroplasts provides synergistically enhanced protection against various abiotic stresses. *Physiol. Plant.* 138:520. doi: 10.1111/j.1399-3054.2010.01348.x
- Almeselmani, M., Deshmukh, P. S., Sairam, R. K., Kushwaha, S. R., and Singh, T. P. (2006). Protective role of antioxidant enzymes under high temperature stress. *Plant Sci.* 171, 382–388. doi: 10.1016/j.plantsci.2006.04.009
- Andréia, C., Gisele, P., Silvia Barcellos, R., Carolina Werner, R., Fernanda, L., and Márcia, M. P. (2012). Plant responses to stresses: role of ascorbate peroxidase in the antioxidant protection. *Genet. Mol. Biol.* 35, 1011–1019.
- Apel, K., and Hirt, H. (2014). Reactive oxygen species: metabolism, oxidative stress, and signal transduction. *Annu. Rev. Plant Biol.* 55, 728–749. doi: 10.1146/annurev-arplant.55.031903.141701
- Badawi, G. H., Kawano, N., Yamauchi, Y., Shimada, E., Sasaki, R., Kubo, A., et al. (2004). Over-expression of ascorbate peroxidase in tobacco chloroplasts enhances the tolerance to salt stress and water deficit. *Physiol. Plant.* 121, 231–238. doi: 10.1111/j.0031-9317.2004.00308.x
- Breusegem, F. V., Villarroel, R., Montagu, M. V., and Inzé, D. (1995). Ascorbate peroxidase cDNA from maize. *Plant Physiol.* 107, 649–650. doi: 10.1104/pp.107.2.649
- Chen, D., Zheng, X. F., Li, G. L., Pan, C., Zhou, M. Q., and Hu, Z. L. (2011). Cloning and expression of one chloroplastic ascorbate peroxidase gene from *Nelumbo nucifera*. *Biochem. Genet.* 49, 656–664. doi: 10.1007/s10528-011-9440-x
- Chen, G. X., and Asada, K. (1989). Ascorbate peroxidase in tea leaves: occurrence of two isozymes and the differences in their enzymatic and molecular properties. *Plant Cell Physiol.* 30, 987–998.
- Cheng, H., He, Q. W., Huo, Y. M., Hou, L. X., and Lv, J. F. (2009). Molecular cloning, characterization and expression analysis of CmAPX. *Mol. Biol. Rep.* 36, 1531–1537. doi: 10.1007/s11033-008-9345-x
- Chew, O., Whelan, J., and Millar, A. H. (2003). Molecular definition of the ascorbate-glutathione cycle in Arabidopsis mitochondria reveals dual targeting of antioxidant defenses in plants. *J. Biol. Chem.* 278, 46869–46877. doi: 10.1074/jbc.M307525200
- Dang, F. F., Wang, Y. N., Yu, L., Eulgem, T., Lai, Y., Liu, Z. Q., et al. (2013). CaWRKY40, a WRKY protein of pepper, plays an important role in the regulation of tolerance to heat stress and resistance to *Ralstonia solanacearum* infection. *Plant Cell Environ.* 36, 757–774. doi: 10.1111/pce.12011
- D'Arcylameta, A., Ferrariliou, R., Contouransel, D., Phamthi, A. T., and Zuilyfodil, Y. (2006). Isolation and characterization of four ascorbate peroxidase cDNAs responsive to water deficit in cowpea leaves. *Ann. Bot.* 97, 133–140. doi: 10.1093/aob/mcj010
- Datta, K., and Datta, S. K. (1999). Transformation of rice via PEG-mediated DNA uptake into protoplasts. *Methods Mol. Biol.* 111, 335–347. doi: 10.1385/1-59259-583-9:335
- Du, S. C., Hwang, I. S., and Hwang, B. K. (2012). Requirement of the cytosolic interaction between pathogenesis-related protein10 and leucine-rich repeat protein1 for cell death and defense signaling in pepper. *Plant Cell* 24, 1675–1690. doi: 10.1105/tpc.112.095869
- Duan, S. R., Ling, Y., and Xi, S. A. (2006). Overexpressing peroxisomal ascorbate peroxidase gene in rice enhanced tolerance to cadmium stress. *Acta Pedol. Sinica* 43, 111–116.
- Dunand, C., Mathé, C., Lazzarotto, F., Margis, R., and Margispinho, M. (2011). Ascorbate peroxidase-related (APx-R) is not a duplicable gene. *Plant Signal. Behav.* 6, 1908–1913. doi: 10.4161/psb.6.12.18098
- Foyer, C. H., and Noctor, G. (2005). Redox homeostasis and antioxidant signaling: a metabolic interface between stress perception and physiological responses. *Plant cell* 17, 1866–1875. doi: 10.1105/tpc.105.033589
- Guo, F., Chiang, M. Y., Wang, Y. T., and Zhang, Y. Z. (2008). An *in vitro* recombination method to convert restriction- and ligation-independent expression vectors. *Biotechnol. J.* 3, 370–377. doi: 10.1002/biot.200700170
- Guo, J. L., Ling, H., Wu, Q. B., Xu, L. P., and Que, Y. X. (2014). The choice of reference genes for assessing gene expression in sugarcane under salinity and drought stresses. *Sci. Rep.* 4, 7042–7051. doi: 10.1038/srep07042
- Guo, J. L., Xu, L. P., Fang, J. P., Su, Y. C., Fu, H. Y., Que, Y. X., et al. (2012). A novel dirigent protein gene with highly stem-specific expression from sugarcane, response to drought, salt and oxidative stresses. *Plant Cell Rep.* 31, 1801–1812. doi: 10.1007/s00299-012-1293-1
- Guo, J. L., Xu, L. P., Su, Y. C., Wang, H. B., Gao, S. W., Xu, J. S., et al. (2013). ScMT2-1-3, a metallothionein gene of sugarcane, plays an important role in the regulation of heavy metal tolerance/accumulation. *Biomed. Res. Int.* 2013, 86–89. doi: 10.1155/2013/904769
- Gupta, K., Agarwal, P. K., Reddy, M. K., and Jha, B. (2010). SbDREB2A, an A-2 type DREB transcription factor from extreme halophyte *Salicornia brachiata* confers abiotic stress tolerance in *Escherichia coli*. *Plant Cell Rep.* 29, 1131–1137. doi: 10.1007/s00299-010-0896-7
- Hong, C. Y., and Kao, C. H. (2007). Expression of ascorbate peroxidase 8 in roots of rice (*Oryza sativa* L.) seedlings in response to NaCl. *J. Exp. Bot.* 58, 3273–3283. doi: 10.1093/jxb/erm174
- Huang, F., Zhang, Y. H., and Li, X. M. (2013). *In silico* cloning and characterization of ascorbate peroxidase gene from *Saccharum* using bioinformatics tools. *J. Shenyang Norm. Univ.* 31, 295–300
- Ishikawa, T., Sakai, K., Takeda, T., and Shigeoka, S. (1995). Cloning and expression of cDNA encoding a new type of ascorbate peroxidase from spinach. *FEBS Lett.* 367, 28–32. doi: 10.1016/0014-5793(95)00539-L
- Ishikawa, T., Sakai, K., Yoshimura, K., Takeda, T., and Shigeoka, S. (1996). cDNAs encoding spinach stromal and thylakoid-bound ascorbate peroxidase, differing in the presence or absence of their 3'-coding regions. *FEBS Lett.* 384, 289–293. doi: 10.1016/0014-5793(96)00332-8
- Ishikawa, T., Yoshimura, K., Sakai, K., Tamoi, M., Takeda, T., and Shigeoka, S. (1998). Molecular characterization and physiological role of a glyoxysome-bound ascorbate peroxidase from spinach. *Plant Cell Physiol.* 39, 23–34. doi: 10.1093/oxfordjournals.pcp.a029285
- Jiang, M., Zhang, Z. X., and Yuan, L. J. (2012). Cloning and expression analysis of an ascorbate peroxidase gene *BoAPX2* from *Brassica oleracea* var. *italica*. *Acta Phytopathol. Sin.* 42, 374–380.
- Kawakami, S., Matsumoto, Y., Matsunaga, A. S., and Mizuno, M. (2002). Molecular cloning of ascorbate peroxidase in potato tubers and its response during storage at low temperature. *Plant Sci.* 163, 829–836. doi: 10.1016/S0168-9452(02)00232-7
- Kornyejev, D., Logan, B. A., Payton, P., Allen, R. D., and Holaday, A. S. (2001). Enhanced photochemical light utilization and decreased chilling-induced photoinhibition of photosystem II in cotton overexpressing genes encoding chloroplast-targeted antioxidant enzymes. *Physiol. Plant.* 113, 323–331. doi: 10.1034/j.1399-3054.2001.1130304.x
- Lai, Y., Dang, F. F., Lin, J., Yu, L., Shi, Y. L., Xiao, Y. H., et al. (2013). Overexpression of a Chinese cabbage *BrERF11* transcription factor enhances disease resistance to *Ralstonia solanacearum* in tobacco. *Plant Physiol. Biochem.* 62, 70–78. doi: 10.1016/j.plaphy.2012.10.010
- Lazzarotto, F., Teixeira, F. K., Rosa, S. B., Dunand, C., Fernandes, C. L., Fontenele, A. D. V., et al. (2011). Ascorbate peroxidase-related (APx-R) is a new heme-containing protein functionally associated with ascorbate peroxidase but evolutionarily divergent. *New Phytol.* 191, 234–250. doi: 10.1111/j.1469-8137.2011.03659.x
- Li, Q. W. (2000). *Modern Technology for Sugarcane Improvement*. Guangzhou: South China Science and Technology Press.
- Li, Y. Z., Tessaro, M. J., Li, X., and Zhang, Y. L. (2010). Regulation of the expression of plant resistance gene *SN1* by a protein with a conserved BAT2 domain. *Plant Physiol.* 153, 1425–1434. doi: 10.1104/pp.110.156240
- Lin, L., Wang, X. P., and Wang, Y. J. (2006). cDNA clone, fusion expression and purification of the novel gene related to ascorbate peroxidase from Chinese wild *Vitis pseudoreticulata* in *E. coli*. *Mol. Biol. Rep.* 33, 197–206. doi: 10.1007/s11033-006-0008-5
- Liu, H. C., Tian, D. Q., Liu, J. X., Ma, G. Y., Zou, Q. C., and Zhu, Z. J. (2013). Cloning and functional analysis of a novel ascorbate peroxidase (APX) gene from *Anthurium andraeanum*. *J. Zhejiang Univ.* 14, 1110–1120. doi: 10.1631/jzus.B1300105

- Livak, K. J., and Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta C_T}$ method. *Methods* 25, 402–408. doi: 10.1006/meth.2001.1262
- Mano, S., Yamaguchi, K., Hayashi, M., and Nishimura, M. (1997). Stromal and thylakoid-bound ascorbate peroxidases are produced by alternative splicing in pumpkin. *FEBS Lett.* 413, 21–26. doi: 10.1016/S0014-5793(97)00862-4
- Maruta, T., Tanouchi, A., Tamoi, M., Yabuta, Y., Yoshimura, K., Ishikawa, T., et al. (2010). *Arabidopsis* chloroplastic ascorbate peroxidase isoenzymes play a dual role in photoprotection and gene regulation under photooxidative stress. *Plant Cell Physiol.* 51, 190–200. doi: 10.1093/pcp/pcp177
- Melech-Bonfil, S., and Sessa, G. (2010). Tomato MAPKKKε is a positive regulator of cell-death signaling networks associated with plant immunity. *Plant J.* 64, 379–391. doi: 10.1111/j.1365-313X.2010.04333.x
- Mittler, R. (2002). Oxidative stress, antioxidants and stress tolerance. *Trends Plant Sci.* 7, 405–410. doi: 10.1016/S1360-1385(02)02312-9
- Mittler, R., Sandy, V., Martin, G., and Frank, V. B. (2004). Reactive oxygen gene network of plants. *Trends Plant Sci.* 9, 490–498. doi: 10.1016/j.tplants.2004.08.009
- Mittler, R., Sandy, V., Nobuhiro, S., Gad, M., Tognetti, V. B., Klaas, V., et al. (2011). ROS signaling: the new wave? *Trends Plant Sci.* 16, 300–309. doi: 10.1016/j.tplants.2011.03.007
- Murgia, I., Tarantino, D., Vannini, C., Bracale, M., Carravieri, S., and Soave, C. (2004). *Arabidopsis thaliana* plants overexpressing thylakoidal ascorbate peroxidase show increased resistance to paraquat-induced photooxidative stress and to nitric oxide-induced cell death. *Plant J.* 38, 940–953. doi: 10.1111/j.1365-313X.2004.02092.x
- Najami, N., Janda, T., Barriah, W., Kayam, G., Tal, M., Guy, M., et al. (2008). Ascorbate gene family in tomato: its identification and characterization. *Mol. Genet. Genomics* 279, 171–182. doi: 10.1007/s00438-007-0305-2
- Narendra, S., Venkataramani, S., Shen, G., Wang, J., Pasapula, V., Lin, Y., et al. (2006). The *Arabidopsis* ascorbate peroxidase 3 is a peroxisomal membrane-bound antioxidant enzyme and is dispensable for *Arabidopsis* growth and development. *J. Exp. Bot.* 57, 3033–3042. doi: 10.1093/jxb/erl060
- Nayyar, H., and Gupta, D. (2006). Differential sensitivity of C3 and C4 plants to water deficit stress: association with oxidative stress and antioxidants. *Environ. Exp. Bot.* 58, 106–113. doi: 10.1016/j.envexpbot.2005.06.021
- Pallavi, S., and Dubey, R. S. (2007). Involvement of oxidative stress and role of antioxidative defense system in growing rice seedlings exposed to toxic concentrations of aluminum. *Plant Cell Rep.* 26, 2027–2038. doi: 10.1007/s00299-007-0416-6
- Panchuk, I. I., Volkov, R. A., and Friedrich, S. F. (2002). Heat stress and heat shock transcription factor-dependent expression and activity of ascorbate peroxidase in *Arabidopsis*. *Braz. J. Microbiol.* 129, 838–853. doi: 10.1104/pp.001362
- Saitou, N. N. M., and Nei, M. C. (1987). The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.* 4, 406–425.
- Shi, W. M., Muramoto, Y., Ueda, A., and Takabe, T. (2001). Cloning of peroxisomal ascorbate peroxidase gene from barley and enhanced thermotolerance by overexpressing in *Arabidopsis thaliana*. *Gene* 273, 23–27. doi: 10.1016/S0378-1119(01)00566-2
- Shigeoka, S., Ishikawa, T., Tamoi, M., Miyagawa, Y., Takeda, T., and Yabuta, Y. (2002). Regulation and function of ascorbate peroxidase isoenzymes. *J. Exp. Bot.* 53, 1305–1319. doi: 10.1093/jxb/53.372.1305
- Su, Y. C., Guo, J. L., Ling, H., Chen, S. S., Wang, S. S., Xu, L. P., et al. (2014a). Isolation of a novel peroxisomal catalase gene from sugarcane, which is responsive to biotic and abiotic stresses. *PLoS ONE* 9:e84426. doi: 10.1371/journal.pone.0084426
- Su, Y. C., Wang, Z. Q., Liu, F., Li, Z., Peng, Q., Guo, J. L., et al. (2016). Isolation and characterization of *ScGluD2*, a new sugarcane beta-1,3-glucanase D family gene induced by *Sporisorium scitamineum*, ABA, H₂O₂, NaCl, and CdCl₂ stresses. *Front. Plant Sci.* 7:1348. doi: 10.3389/fpls.2016.01348
- Su, Y. C., Xu, L. P., Fu, Z. W., Yang, Y. T., Guo, J. L., Wang, S. S., et al. (2014b). *ScChi*, encoding an acidic class III chitinase of sugarcane, confers positive responses to biotic and abiotic stresses in sugarcane. *Int. J. Mol. Sci.* 15, 2738–2760. doi: 10.3390/ijms15022738
- Su, Y. C., Xu, L. P., Xue, B. T., Wu, Q. B., Guo, J. L., Wu, L. G., et al. (2013). Molecular cloning and characterization of two pathogenesis-related β-1,3-glucanase genes *ScGluA1* and *ScGluD1* from sugarcane infected by *Sporisorium scitamineum*. *Plant Cell Rep.* 32, 1503–1519. doi: 10.1007/s00299-013-1463-9
- Teixeira, F. K., Menezes-Benavente, L., Galvão, V. C., and Margis-Pinheiro, M. (2005). Multigene families encode the major enzymes of antioxidant metabolism in *Eucalyptus grandis* L. *Genet. Mol. Biol.* 28, 529–538. doi: 10.1590/S1415-47572005000400007
- Teixeira, F. K., Menezes-Benavente, L., Galvão, V. C., Margis, R., and Margis-Pinheiro, M. (2006). Rice ascorbate peroxidase gene family encodes functionally diverse isoforms localized in different subcellular compartments. *Planta* 224, 300–314. doi: 10.1007/s00425-005-0214-8
- Teixeira, F. K., Menezes-Benavente, L., Margis, R., and Margis-Pinheiro, M. (2004). Analysis of the molecular evolutionary history of the ascorbate peroxidase gene family: inferences from the rice genome. *J. Mol. Evol.* 59, 761–770. doi: 10.1007/s00239-004-2666-z
- Thordal-Christensen, H., Zhang, Z. G., Wei, Y. D., and Collinge, D. B. (1997). Subcellular localization of H₂O₂ in plants. H₂O₂ accumulation in papillae and hypersensitive response during the barley-powdery mildew interaction. *Plant J.* 11, 1187–1194. doi: 10.1046/j.1365-313X.1997.11061187.x
- Walker, J. M. (2005). *The Proteomics Protocols Handbook*. Totowa, NJ: Humana Press.
- Wang, S., Zhang, K. K., Huang, X., Fan, Y. J., Yang, L. T., and Li, Y. R. (2015). Cloning and functional analysis of thylakoidal ascorbate peroxidase (*TAPX*) gene in sugarcane. *Sugar Tech* 17, 131–135. doi: 10.1007/s12355-014-0354-x
- Wang, Z. Q., Chen, Y., Yang, Y. T., Su, Y. C., Chen, S. S., Wu, Q. B., et al. (2015). Cloning and expression analysis of ascorbate peroxidase gene (*ScAPX*) in sugarcane (*Saccharum officinarum*). *J. Agric. Biotechnol.* 23, 170–180.
- Xu, D. L., Park, J. W., Mirkov, T. E., and Zhou, G. H. (2008). Viruses causing mosaic disease in sugarcane and their genetic diversity in southern China. *Arch. Virol.* 153, 1031–1039. doi: 10.1007/s00705-008-0072-3
- Yabuta, Y., Motoki, T. K., Takeda, T., Ishikawa, T., and Shigeoka, S. (2002). Thylakoid membrane-bound ascorbate peroxidase is a limiting factor of antioxidative systems under photo-oxidative stress. *Plant J.* 32, 915–925. doi: 10.1046/j.1365-313X.2002.01476.x
- Yamane, K., Mitsuya, S., Taniguchi, M., and Miyake, H. (2010). Transcription profiles of genes encoding catalase and ascorbate peroxidase in rice leaf tissues under salinity. *Plant Prod. Sci.* 13, 164–168. doi: 10.1626/pp.13.164

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Liu, Huang, Wang, Ling, Sun, Ahmad, Muhammad, Guo, Xu, Gao, Que and Su. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.