



Understory Community Assembly Following Wildfire in Boreal Forests: Shift From Stochasticity to Competitive Exclusion and Environmental Filtering

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Understory vegetation accounts for the majority of plant species diversity and serves as a driver of overstory succession and nutrient cycling in boreal forest ecosystems. However, investigations of the underlying assembly processes of understory vegetation associated with stand development following a wildfire disturbance are rare, particularly in Eurasian boreal forests. In this study, we measured the phylogenetic and functional diversity and trait dispersions of understory communities and tested how these patterns changed with stand age in the Great Xing'an Mountains of Northeastern China. Contrary to our expectation, we found that understory functional traits were phylogenetically convergent. We found that random patterns of phylogenetic, functional, and trait dispersions were dominant for most of our surveyed plots, indicating that stochastic processes may play a crucial role in the determination of understory community assembly. Yet, there was an evidence that understory community assembly was also determined by competitive exclusion and environmental filtering to a certain degree, which was demonstrated by the observed clustered phylogenetic and functional patterns in some plots. Our results showed that phylogenetic diversity significantly decreased, while functional diversity increased with stand age. The observed shift trends in phylogenetic and functional patterns between random to clustering along with stand age, which suggested that understory community assembly shifted from stochasticity to competitive exclusion and environmental filtering. Our study presented a difference to community assembly and species coexistence theories insisted solely on deterministic processes. These findings indicated that Eurasian boreal understory communities may be primarily regulated by stochastic processes, providing complementary evidence that stochastic processes are crucial in the determination of community assembly both in tropical and boreal forests.

Keywords: chronosequence, community assembly, environmental filtering, functional diversity, trait conservatism, trait dispersion, phylogenetic diversity, overdispersion

INTRODUCTION

Wildfires are common and widespread in ecosystems, serving as a global "herbivore" in the determination of plant distribution, and therefore, community composition (Bond and Keeley, 2005). Wildfire is a potent evolutionary force for fire-tolerant and firedependent plant species (Bond and Keeley, 2005). Functional traits that enable rapid colonization and efficient post-fire regeneration are crucial for the successful establishment and persistence of fire-adaptive plant communities (Pausas et al., 2004; Pausas and Keeley, 2014). Although global burned areas have declined over the last two decades due to anthropogenic activities (Andela et al., 2017), fire frequency and severity are anticipated to increase in many ecosystems as a consequence of projected climate change in the coming decades (Stephens et al., 2013; Jolly et al., 2015). Thus, an understanding of how forest communities are assembled following wildfire, is useful to land managers in the preparation of post-fire strategies for vegetation regeneration and future fuel management.

Two core processes have been proposed to explain vegetation assembly following stand-replacing disturbances (e.g., wildfire, wind, logging), namely stochastic processes and deterministic processes. The stochastic class emphasizes the importance of dispersal limitations and stochastic demographics in determining community assembly (Bell, 2000; Hubbell, 2001). In contrast, the deterministic class assumes that plant communities are generally driven by two opposing forces: environmental filtering, which gives rise to co-occurring species that are intimately related under similar environmental conditions, and competitive exclusion that decreases the similarity of co-occurring species (Weiher et al., 1998; Chesson, 2000; Cornwell et al., 2006). Previous studies have suggested that stochastic processes are dominant in the determination of community assembly in tropical forests (Condit et al., 2002; Chase, 2010), while niche based deterministic processes are predominant in mid-latitude temperate regions (Clark and McLachlan, 2003; Gilbert and Lechowicz, 2004).

Over the last two decades, phylogenetic- and functional trait-based approaches were increasingly adopted for studies of tropical and temperate forest community assembly; however, there have been few for boreal forests (Kraft and Ackerly, 2010; Swenson et al., 2012a; Wang et al., 2015). Phylogenetic and functional patterns could reflect different ecological processes acting on community assembly (e.g., environmental filtering and competitive exclusion) with diversified evolution of functional traits (Figure 1). Studies have shown that determining the evolutionary patterns of functional traits is the prerequisite to interpreting the mechanisms of community phylogenetic and functional structure (Uriarte et al., 2010; Bennett et al., 2013; Fritschie et al., 2014). When functional traits are phylogenetically conserved (i.e., closely related species are ecologically similar with traits being a legacy from their ancestors), environmental filtering generally results in phylogenetic and functional clustering (Figure 1A); while competitive exclusion results in phylogenetic and functional overdispersion (Figure 1B) (Webb et al., 2002; Cavender-Bares et al., 2004; Kembel, 2009). In addition, environmental filtering will drive functional traits of co-occurring species to be more similar than expected by chance, namely trait convergence (Weiher et al., 1998; Cornwell et al., 2006; Grime, 2006) (**Figure 1A**). Conversely, competitive exclusion will drive functional traits of co-occurring species to be less similar than expected by chance, namely trait divergence (**Figure 1B**) (Chesson et al., 2004; Wilson, 2007; Wilson and Stubbs, 2012). When functional traits are phylogenetically convergent (i.e., species presented in different lineages have similar functional traits), environmental filtering would generate phylogenetic overdispersion, functional clustering and trait convergence (**Figure 1C**), while competitive exclusion will generate phylogenetic clustering, functional overdispersion and trait divergence (**Figure 1D**) (Cavender-Bares et al., 2004; Kembel, 2009; Uriarte et al., 2010; Bennett et al., 2013; Fritschie et al., 2014).

Previous studies have reported that stand age imparts potent influences on plant community assembly, due to the time required for colonization, and changes in resource availability as the stand develops (Hart and Chen, 2006; Verdú and Pausas, 2007; Kumar et al., 2017). According to the observed temporal changes in phylogenetic or functional patterns, successional studies have demonstrated that community assembly processes are altered as stands develop following stand-replacing disturbances (Norden et al., 2012; Purschke et al., 2013; Li et al., 2015; Muscarella et al., 2016). Stands at early successional stages are typically dominated by shade intolerant, nutrient demanding, and fast growing species, which results in phylogenetic and functional clustering, due to the availability of abundant resources subsequent to a disturbance (Verdú et al., 2009; Letcher et al., 2012). In contrast, late-successional communities are more often dominated by shade tolerant, slow growing and distantly related species with dissimilar functional traits, which are characterized by phylogenetic and functional over dispersion caused by competitive exclusion as resources become limited (Verdú et al., 2009; Letcher et al., 2012). Such transformation has been generally attributed to shifts in community assembly processes, from environmental filtering to competitive exclusion (Purschke et al., 2013).

To date, much research has primarily focused on overstory tree community assembly (Kooyman et al., 2011; Whitfeld et al., 2012), yet little empirical work exists for understory species (Azeria et al., 2011). In boreal region, forest understory communities constitutes the majority of plant diversity, serving as a critical driver for nutrient cycling and overstory succession (Nilsson and Wardle, 2005; Hart and Chen, 2006). Therefore, understanding the underlying mechanisms of understory community assembly may provide complementary information toward the elucidation of community assembly mechanisms following disturbances in boreal forest ecosystems. Here, we attempted to infer understory assembly processes from the temporal trends of phylogenetic and functional patterns coupling with trait dispersion in boreal forest. We surveyed understory species abundance and measured key functional traits over a chronosequence of 200 years following wildfire in a larch forest of Northeastern China. Specially, we tested phylogenetic trait conservatism to determine how understory functional traits evolved through quantifying phylogenetic signal and expected that understory functional traits were Conserved traits (Phylogenetic trait conservatism): traits are a legacy from their common ancestors and closely related species in phylogeny had similar traits

Convergent traits: traits have similar form or function but species are present in different lineages **Phylogenetic clustering**: co-occurring species are more closely related in phylogeny than expected by chance

Functional clustering: co-occurring species are more functionally related with similar traits than expected by chance

Phylogenetic overdispersion: co-occurring species are more distantly related in phylogeny than expected by chance

Functional overdispersion: co-occurring species are less functionally related with dissimilar traits than expected by chance

Trait convergence: functional traits of co-occurring species are more similar than expected by chance

Trait divergence: functional traits of co-occurring species are less similar than expected by chance



FIGURE 1 | Environmental filtering and competitive exclusion give rise to opposite expectations about the phylogenetic, functional and trait patterns. Which ecological force can be invoked to explain a given phylogenetic, functional and trait patterns depends on whether trait evolution is conserved or convergent. When functional traits are phylogenetically conserved, environmental filtering drives clustering patterns (A) and competitive exclusion drives overdispersion patterns (B). When functional traits are phylogenetically convergent, competitive exclusion drives phylogenetic clustering, functional overdispersion and trait divergence patterns (C), and environmental filtering drives phylogenetic overdispersion, functional clustering and trait convergence patterns (D). In each figure, there are three communities represented by rectangles. Within each community, colored circles represent species. Black lines represent the source of species from phylogenetic tree for each community. Colored lines with different length represent traits with different values. Figure adapted from Webb et al. (2002), Cavender-Bares et al. (2004), Kembel (2009), and Bernard-Verdier et al. (2012).

phylogenetically conserved. We tested the effects of stand age (years since fire) on understory phylogenetic and functional diversity and trait dispersion. We expected that phylogenetic and functional diversity and trait dispersion would change with stand age due to its strong control in the shifting of understory species composition (Hart and Chen, 2008; Kumar et al., 2017). By comparing the phylogenetic and functional patterns and trait dispersion of understory vegetation, we assessed the changes in relative importance of environmental filtering and competitive exclusion with stand development. We hypothesized that increasing stand age would promote phylogenetic and functional overdispersion and trait divergence as a result of increasing competitive exclusion (Spasojevic and Suding, 2012; Purschke et al., 2013).

MATERIALS AND METHODS

Study Area

The present study was conducted at the southern margin of the Eurasian boreal forest, which is located in the Great Xing'an Mountains of Northeastern China. This study area is a mountainous region with elevations that range from 239 m in the Northeast, to 1,488 m in the Southwest. The climate in this region is typical terrestrial monsoon with a mean annual temperature of -4.4° C, ranging from -2.7° C to -5.3° C, and average annual precipitation \sim 500 mm. Fire comprises the primary natural disturbance, with a mean fire return interval of approximately 120-150 years (Chang et al., 2007). The typical vegetation type of this region belongs to the cool temperate coniferous forest (Zhou, 1991). The most dominant species is larch (Larix gmelini (Rupr.) Kuzen.), which is widely distributed in this region; typically forming pure stands. Birch (Betula platyphylla Suk.) is the widely distributed broadleaf species that intersperses in the larch forests at xeric sites. Pine (Pinus sylvestris L. var. mongolica Litv.), spruce (Picea koraiensis Nakai), aspen (Populus davidiana Dode, Populus suaveolens Fisch.) and willow (Chosenia arbutifolia (Pall.) A. Skv.) are also interspersed in larch forests, with a small area of distribution (<2%). The most diverse component of the boreal forests is the understory vegetation, where common understory plants include Betula fruticosa Pall., Rhododendron dauricum L., Vaccinium uliginosum L., Carex schmidtii, and Chamerion angustifolium.

Sampling Design

To examine the effects of stand age on understory vegetation, stands originating subsequent to wildfire were selected using a chronosequence approach. The chronosequence approach is recommended for the investigation of successional processes, over decadal to millennial time scales (Walker et al., 2010). Based on available fire-originating stands within the study area, we selected seven age classes of fire-originating stands, representing early stand initiation (4-year), late stand initiation (14-year), early stem exclusion (27-year), late stem exclusion (55-year), early canopy transition (76-year), late canopy transition (98year), and gap dynamic (203-year) stages of stand development (Chen and Popadiouk, 2002). All stands were sampled on welldrained brown coniferous forest soil, which is the dominant soil type in this region (Gong, 2001). The selected stands were visually homogeneous in terms of stem density and composition within each stand age.

For the sampling of stands \leq 50 years old, time since last fire was determined according to the fire occurrence records, which have been reported since 1965 (Liu et al., 2012). For the sampling of stands >50 years old, we confirmed fire to be the primary disturbance factor based on the black carbon in soil and the burned stump for forest stands over 50 years old. The stands we selected had cohorts of overstory seedlings and trees with similar age. Stand ages were determined through dendrochronology analysis (Chen et al., 2013). We selected larch to determine the time since fire for all sampled stands. For each stand, three to five trees were selected, and a core was extracted at breast height (1.3 m above root collar) from each tree. The cores collected in the field were stored and transported in plastic straws. In the laboratory, the cores were mounted on grooved, wooden core strips and sanded to make the growth rings visible. Subsequently, we counted the tree rings using a hand-held magnifier until the same number was obtained following three successive counts. In order to precisely represent stand age, we corrected the tree ages by adding 8 years to the ring counts made at breast height, accounting for the number of years required by trees to grow to breast height.

Field Survey

A field survey was conducted during the peak vegetation cover from July to August. For each sampled stand, we randomly established a 400 m² square plot for all measurements. In order to ensure accessibility and to avoid edge effects, each plot was selected within a walking distance of 50–2,000 m from a road. For stands of >4 years, the diameters at breast height (1.3 m above root collar) of all trees within the plot were measured and recorded. Tree and sapling density and basal area were summed to the plot level and calculated per hectare (**Table S1**). For stands in the 4-year age class, seedling basal areas were not reported here due to negligible values.

According to previous studies (Chipman and Johnson, 2002; Hart and Chen, 2008), the understory vegetation was surveyed in four randomly allocated 1×1 m quadrats. The percentage of cover for each shrub and herb species <1.3 m in height within each quadrat was visually estimated (Mueller Dombois and Ellenberg, 1974). Specimens of any unidentified species were collected in the field and transferred to the Herbarium of Northeast China for identification. Species nomenclature followed the Flora Republicae Popularis Sinicae (http://frps. eflora.cn) and Flora of China databases (http://foc.eflora.cn/).

Trait Selection and Measurements

Six morphological functional traits were selected to characterize the vegetative phase of understory vegetation according to Pérez-Harguindeguy et al. (2013). Leaf area (LA, mm²) is a major determinant of the ability of a species to sequester light resource, which can affect the photosynthetic rate. Leaf carbon content (LCC, %) and leaf dry matter content (LDMC, %) are related to physical plant resistance, leaf life span, and relative growth rate. Leaf nitrogen content (LNC, %) is a key foliar trait, which is strongly correlated with nutritional quality, the photosynthetic rate, and productivity. Plant height (PH, cm) is employed to quantify the light that is available for capture by understory plants. Specific leaf area (SLA, cm²/g) represents strategies for plant growth and survival, such as structural investment, leaf life spans, and photosynthetic rates. Functional trait values were determined following the new handbook for worldwide functional traits measurement (Pérez-Harguindeguy et al., 2013). We sampled at least five individuals for each observed species in each plot. The individuals we selected were reproductively mature and visually healthy. Following measurements, functional trait values were averaged at the species level for each sample plot as interspecific variability inclines to exceed intraspecific variability for understory traits (Burton et al., 2017). All understory species and the values of functional traits for each species were listed in the Table S2.

Measurement of Trait Phylogenetic Conservatism

Before quantifying phylogenetic conservatisms for the measured functional traits, a phylogenetic tree for our study species was constructed using the online tool *Phylomatic* (http:// phylodiversity.net/phylomatic/). This program generates a megatree with modern family and genus names based on previously published phylogenies (Phylogenetic tree version: zanne2014) (Zanne et al., 2014). Family and genus name resolutions are based on the Angiosperm Phylogeny Website (Stevens, 2001).

To assess the degree of species phylogenetic trait conservatism, we employed a widely used Blomberg's K statistic to quantify the phylogenetic signals of the six continuous traits (Blomberg et al., 2003). K was the ratio of the mean squared error of the tip data measured from the phylogenetically correct mean (MSE_0), divided by the mean squared error of the data (MSE), which was quantified using the phylogenetic variance-covariance matrix, derived from the candidate tree. Afterward, K is calculated as:

$$K = observed \frac{MSE_0}{MSE} / expected \frac{MSE_0}{MSE}$$
(1)

K is a continuous value, ranging from zero to infinity. When the K value is close to zero (p > 0.05), it implies a random or convergent pattern of trait evolution, and a weak phylogenetic signal. When the K value is close to one (p < 0.05), it implies that there is a strong phylogenetic signal, and that a trait has evolved based on the Brownian motion model. When the K value is higher than 1 (p < 0.05), it indicates a strong phylogenetic signal of traits, and closely related species are more similar than expected under a Brownian motion model of trait evolution. The K values were calculated in R 3.4.1 (R Development Core Team, 2017) using the "picante" package (Kembel et al., 2010). Additionally, we projected phylogenetic tree into trait space using traitgram and extended this to incorporate uncertainty about ancestral trait values along branches and at nodes, which was performed in R with package "phylotools" (Ackerly, 2009; Revell, 2013).

Measurement of Phylogenetic and Functional Diversity and Trait Dispersion

The phylogenetic diversity within individual plots at each stand age was quantified via the mean pairwise distance (MPD), which measures the sum of the branch lengths of all co-occurring species within each community (Webb, 2000; Webb et al., 2002). We performed this analysis by using the phylogenetic dendrogram and weighted the pairwise distances among species by their relative coverage. An identical framework was utilized to quantity functional diversity using the trait dendrogram from Euclidean trait distances, referred to as mean pairwise functional distance (MFD) (Li et al., 2015). Community weighted trait variance (CWV) within individual plots at each stage was employed to depict trait dispersion (Bernard-Verdier et al., 2012), which was computed for each trait in each plot as follows:

$$CWV = \sum_{i=1}^{S} p_i \times (x_i \text{-} CWM)^2$$
(2)

where CWV is the community weighted trait variance for a given functional trait, p_i is the relative abundance (percent coverage) of species *i* (*i* = 1, 2, ..., S), x_i is the trait value of species *i*, and CWM is the community weighted mean trait value of species *i*. The community weighted mean trait value (CWM) for each functional trait (Garnier et al., 2004) was calculated as follows:

$$CWM = \sum_{i=1}^{S} p_i x_i \tag{3}$$

where CWM is the community weighted mean value for a given functional trait, p_i is the relative abundance (percent coverage) of species *i* (*i* = 1, 2, ..., S), and x_i is the trait value of species *i*. In order to account for the potential scale sensitivities of community patterns, all calculations of individual plots were replicated at a larger spatial scale by summing the plot composition within each stand age (n = 7).

Null Model Testing

The null model was employed to determine whether the observed value of a metric varied from its random expectations. In order to achieve this goal, we implemented three null models. Null model 1 and 2 were generated through shuffling the names of taxa across the tips of the phylogenetic dendrogram and trait dendrogram, respectively (Kembel, 2009). Null model 3 was constructed to test the hypothesis that species abundance (percent coverage) within a community is randomly distributed with respect to trait values through shuffling the abundance values with unchanged traits (Mason et al., 2008; Bernard-Verdier et al., 2012). We then calculated the standardized effect size (SES) of MPD, MFD, and CWV as follows:

$$SES = \frac{Metric_{observed} - mean(Metric_{null})}{sd(Metric_{null})}$$
(4)

where *Metric*_{observed} is the observed value of MPD, MFD, and CWV, *Metric*_{null} is the mean values of random assemblages, and

TABLE 1 Six measured species functional traits: leaf area (LA), leaf carbon
concentration (LCC), leaf dry matter content (LDMC), leaf nitrogen concentration
(LNC), plant height (PH), and specific leaf area (SLA).

Trait	Type of variable	Mean (SE)	к	p
LA	Continuous (mm ²)	1921.06 (326.89)	0.176	0.022
LCC	Continuous (%)	43.20 (0.56)	0.218	0.001
LDMC	Continuous (%)	31.13 (0.97)	0.083	0.445
LNC	Continuous (%)	1.71 (0.08)	0.139	0.032
PH	Continuous (cm)	28.59 (3.10)	0.087	0.348
SLA	Continuous (cm ² /g)	284.32 (16.78)	0.158	0.047

The phylogenetic signal was measured using the K statistic. K values closer to 1 indicate a phylogenetic signal similar to a Brownian motion model of trait evolution. Values >1 indicate a strong phylogenetic signal, and values <1 indicate a weak phylogenetic signal. P-values <0.05 indicate that traits have a greater phylogenetic signal than the traits predicted by random association.

 $sd(Metric_{null})$ is the standard deviation of the random value. The negative values of SES.MPD and SES.MFD with low quantiles (p < 0.05) indicated smaller phylogenetic or functional distances among co-occurring species than expected by chance, namely, significant clustering. In contrast, positive values with high quantiles (p > 0.95) indicated greater phylogenetic or functional distances among species than expected by chance, namely, significant overdispersion (Webb et al., 2002). For SES.CWV, the negative values with low quantiles (p < 0.05) indicated trait convergence, whereas positive values with high quantiles (p > 0.95) indicated trait divergence. Non-significant positive or negative values indicated observed values close to the median of the random dispersion. The metrics of SES.MPD, SES.MFD, and SES.CWV were all calculated in R using the "*picante*" package with an "*abundance.weighted*" argument (Kembel et al., 2010).

Statistical Analysis

The differences in SES.MPD, SES.MFD, and SES.CWV among stand ages were assessed using a non-parametric approach, as our data failed to meet the assumptions of normality and homogeneous variances. We employed rank-based one-way ANOVA to test the effects of stand age on SES.MPD, SES.MFD, and SES.CWV. These analyses were performed in R using the "oneway.rfit" function with "*Rfit*" package (Kloke and McKean, 2012). The significance of the differences among stand ages was tested by Dunn's *Post Hoc* test via the "dunn.test" function in R using the "pgirmess" package (Giraudoux, 2017). We used chi-squared tests to detect whether and how phylogenetic and functional patterns and trait dispersions changed from early successional stage (4- and 14-year age classes combined) to the late successional stage (98- and 203-year age classes combined).

RESULTS

Trait Conservatism and Evolution

All measured traits exhibited small degrees of trait conservatism, with K values ranging from 0.083 for LDMC, to 0.218 for LCC (**Table 1**). This indicated that these traits were largely

phylogenetically convergent, in contrast to the traits that were evolved under a Brownian motion model (all trait K values <1; Table 1). Among these traits, four functional traits (LA, LCC, LNC, and SLA) were more conserved than the those predicted by a random association (four traits with p < 0.05; Table 1), whereas LDMC and PH were closer to zero, which corresponded to a random pattern of evolution. Mapping the six functional traits on the phylogenetic tree, we found functional traits displayed significant differences among phylogenetic related species (Figure S1). LA values of most species exhibited a widely range of 14.21-4,000 mm², which was far away from the estimated ancestral value of LA (1,228 mm²) for all species (Figure S1a). As well, SLA had a widely range of values from 150 to 350 cm²/g, while the ancestral trait value is 234 cm²/g (Figure S1f). Conversely, LCC, LDMC, LNDC, and PH displayed a relatively narrow range of trait values, which were close to their estimated ancestral ancestral values (Figures S1b-e).

Phylogenetic and Functional Diversity and Trait Dispersion of Environmental Filtering and Competitive Exclusion

In the 76- and 203-year age classes, more than half of the plots had negative SES.MPD values, which were significantly different from those expected via the null model, indicating a clustered phylogenetic structure (Table 2). In other age classes, more than half, and even all plots, had SES.MPD values that were not significantly different from those expected by the null model, which indicated a randomly distributed phylogenetic structure (Table 2). The clustered patterns of phylogenetic structure were driven by competitive exclusion, as the traits were largely convergent. In the 4- and 55-year age classes, more than half of the plots had negative SES.MFD values, which were significantly different from those expected by the null model. This indicated a clustered functional structure that was driven by environmental filtering as the traits were largely convergent (Table 2). For other age classes, more than half, and even all plots, had SES.MFD values that were not significantly different from those expected by the null model, indicating a randomly distributed functional structure (Table 2). For the six measured functional traits, more than two-thirds, and even all plots had SES.CWV values that were not significantly different from those expected by the null model across all age classes, indicating random trait dispersions for most plots (Table S3).

Phylogenetic and Functional Patterns and Trait Dispersions Across Stand Age

The SES.MPD decreased significantly from the 4-year age class and attained the lowest level in the 76-year age class (**Figure 2A**). Subsequently, the SES.MPD increased significantly and remained stable in the 98 and 203-year stages (**Figure 2A**). The SES.MFD attained the lowest value in the 4-year age class (**Figure 2B**). The SES.MFD then increased considerably and attained the highest level in the 27-year age class (**Figure 2B**). Further, the SES.MFD initially decreased and then increased between the 27 and 76-year age classes, with a further decrease and increase,

Stand age	Phylogenetic structure			Functional structure		
	Clustering	Random	Overdispersion	Clustering	Random	Overdispersion
4	0	100	0	56	44	0
14	0	100	0	33	67	0
27	25	75	0	0	100	0
55	50	50	0	67	33	0
76	83	17	0	8	92	0
98	42	58	0	33	67	0
203	58	42	0	25	75	0

TABLE 2 | Percentage of clustering, random, and overdispersion patterns for phylogenetic (SES.MPD) and functional (SES.MFD) structures in each age class.



FIGURE 2 | Standardized effect size of phylogenetic diversity (SES.MPD, **A**) and functional diversity (SES.MFD, **B**) along with stand age which were measured as the mean pairwise phylogenetic distance and the mean pairwise functional distance of the 88 plots over 203 years of succession, based separately on phylogeny and the functional dendrograms. Stand ages from 4 to 203 years represent early stand initiation, late stand initiation, early stem exclusion, late stem exclusion, early canopy transition, late canopy transition, and gap dynamics stages, respectively. The boxplot midlines correspond to the median value for each stand age; upper and lower hinges represent the first and third quartiles. Letters indicate significant differences of the mean SES values (p < 0.05) across the stand age. Red dots correspond to the SES values at a larger spatial scale by summing the plot composition for each stand age, and asterisks indicate a significant difference from the null model.

from the 76 to 203-year age classes (**Figure 2B**). At the larger scale, by summing the plot composition for each age class, the SES.MPD was significantly different from the null model for the 55- to 203-year age classes (**Figure 2A**), and the SES.MFD was significantly different from the null model in only the 4-year age class (**Figure 2B**). The chi-squared test indicated that there was a pronounced shift of random phylogenetic pattern to clustering from early to late successional stages (Chi-squared = 18.2, p < 0.01), while there was no pronounced shift of functional pattern from early to late successional stage (Chi-squared = 1.6, p = 0.2).

The SES.CWV revealed individualistic trends for the six measured functional traits along with stand age (**Figure 3**). The SES.CWV.LA (leaf area) decreased significantly from the 4-year age class, and attained the lowest level in the 14-year age class,

which was followed by an increase to attain the highest level in the 27-year age class (**Figure 3A**). The SES.CWV.LA then decreased and attained the lowest level in the 76- and 98-year age classes, followed by an increase to attain the highest level in the 203-year age class. The SES.CWV.LCC (leaf carbon content) increased from the 4-year age class and attained the highest level in the 14-year age class, which then decreased and remained stable from the 55-year age class to the 98-year age class, followed by an increase to attain the highest level in the 203-year age class (**Figure 3B**). The SES.CWV.LDMC (leaf dry matter content) attained the highest value in the 4-year age class, which then decreased to attain the lowest level in the 76-year age class, followed by an increase to attain the highest level in the 76-year age class, followed by an increase to attain the highest level in the 76-year age class, followed by an increase to attain the highest level in the 76-year age class, followed by an increase to attain the highest level in the 76-year age class, followed by an increase to attain the highest level in the 76-year age class, followed by an increase to attain the highest level in the 203-year age class (**Figure 3C**). The SES.CWV.LNC (leaf nitrogen content) had the





lowest value in the 4-year age class, which then increased and attained lowest level in the 27-year age class, followed by a regular increase and decrease from the 27-year age class to the 203year age class (Figure 3D). The SES.CWV.SLA increased from the 4-year age class and attained the highest level in the 27-year age class, which then decreased and reached lowest level in the 98-year age class, followed by an increase to attain the highest level in the 203-year age class (Figure 3F). At the larger scale by summing the plot composition for each stand age, the values of SES.CWV.LA were significantly different from the null model, except in the 4- and 27-year age classes (Figure 3A). Similarly, the values of SES.CWV.LCC were significantly different from the null model in only the 203-year age class (Figure 3B), and the values of SES.CWV.LDMC were significantly different from the null model in the 4- and 14-year age classes (Figure 3C). The SES values of LNC, PH, and SLA were not significantly different from the null model across all age classes (Figures 3D-F).

DISCUSSION

In this study, we firstly tested the degree of phylogenetic trait conservatism for boreal forest understory through quantifying phylogenetic signal (Losos, 2008; Revell et al., 2008; Ackerly, 2009). According to previous studies, phylogenetically conserved traits may result in low rates of trait evolution; conversely, less phylogenetically conserved traits may result in a random or convergent pattern of trait evolution (Webb et al., 2002; Ackerly, 2004). The degree of phylogenetic trait conservatism has been widely tested in tropical and temperate forests, and studies in support of phylogenetic trait conservatism appear to be dominant (Ackerly, 2003). However, there were also counterexamples which suggested that key functional traits, especially leaf morphological traits, were evolutionarily labile for tree species (Kraft and Ackerly, 2010; Swenson et al., 2012b). In our study, the phylogenetic signals of four measured functional traits were >0 (p < 0.05) but <1, which indicated that the key functional traits related to understory light capture strategies were less phylogenetically conserved (Ackerly, 2009). Furthermore, the traitgrams of the six functional traits implied that these functional traits were convergently evolved (Ackerly, 2009).

Contrary to our expectations, the phylogenetic and functional diversity of most plots were not significantly different from those expected by the null models, which indicated that most understory communities were randomly assembled (Webb et al., 2002; Kembel, 2009). These random patterns of co-occurring understory species might be a consequence of stochastic processes, such as external spatially random colonization and germination from the soil seed bank (Hubbell, 2001; Ulrich et al., 2016). Aside from random patterns, there was a clustered phylogenetic pattern, which might be caused by competitive exclusion as trait evolution was largely convergent (Cavender-Bares et al., 2004; Pausas and Verd,ú, 2010). The phylogenetic structure was inclined to shift from random to clustering, which suggested that the importance of competitive exclusion increased from early to late successional stages. In addition, we also found

that the clustered functional pattern, which might be caused by environmental filtering as trait evolution was largely convergent (Cavender-Bares et al., 2004). Although we observed clustered functional patterns in the 4- and 55-year age classes, we failed to observe shifts between clustered and random patterns, from early to late successional stages. This suggested that environmental filtering had only a minimal impact on understory assemblages in some specific age classes. We found that the community weighted trait variance for most communities were equal to those expected by the null model, which indicated that random trait dispersion was dominant in random community assemblages (Bernard-Verdier et al., 2012).

Our study suggested that wildfire substantially altered understory phylogenetic and functional diversity, and community trait dispersions. In our study, phylogenetic diversity decreased while functional diversity increased along with stand age, which showed different temporal trends compared with previous studies. For instance, Li et al. (2015) showed a unimodal relationship of phylogenetic diversity, and a linear relationship of functional diversity, along with stand age, respectively. In contrast, Purschke et al. (2013) found that both the phylogenetic and functional diversity of grassland communities increased along with long-term succession. In addition, we found that phylogenetic and functional diversity exhibited different temporal trends compared with species richness (Figure S2), which initially increased and then decreased along with stand age (Hart and Chen, 2006). This suggested that phylogenetic and functional diversity had weak correlations with species richness in Eurasian boreal forests.

Our results failed to support the assumption that increased local phylogenetic diversity corresponded to increased local functional diversity. This might have been caused by the degree of phylogenetic trait conservatism of the measured functional traits in our study (Gerhold et al., 2015). In addition, we found that the community weighted trait variance for the five traits that we measured, changed significantly but with various temporal trends along with stand age. The differences in phylogenetic and functional diversity, and community weighted trait variance among stand age classes might be attributed to random species colorization and extinction (Lapiedra et al., 2015).

CONCLUSION

Using both phylogenetic- and functional-centric approaches, we presented that the assembly and maintenance of understory communities were the sum of different ecological processes. Our study provided evidence that stochastic processes dominated in the control of boreal forest understory community assembly as shown by the observed random phylogenetic, functional and trait patterns for most plots. We also found that environmental filtering and competitive exclusion affected understory community assembly to a certain degree, which was indicated by the observed clustered phylogenetic and functional patterns for some plots. Our results demonstrated that understory community assembly following wildfire in boreal forests shifted from stochasticity to competitive exclusion and environmental filtering. Our study presented a difference to community assembly and species coexistence theories insisted solely on deterministic processes. We anticipated that our findings might provide complementary information toward the further elucidation of community assembly mechanisms following disturbances in boreal forest ecosystems.

AUTHOR CONTRIBUTIONS

BL, HC, and JY conceived this research. BL collected and analyzed data. BL, HC, and JY wrote the manuscript.

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REFERENCES

- Ackerly, D. (2003). Community assembly, niche conservatism, and adaptive evolution in changing environments. *Int. J. Plant. Sci.* 164, S165–S184. doi: 10.1086/368401
- Ackerly, D. (2004). Functional strategies of chaparral shrubs in relation to seasonal water deficit and disturbance. *Ecol. Monogr.* 74, 25–44. doi: 10.1890/03-4022
- Ackerly, D. (2009). Conservatism and diversification of plant functional traits: evolutionary rates versus phylogenetic signal. *Proc. Natl. Acad. Sci. U.S.A.* 106(Suppl. 2), 19699–19706. doi: 10.1073/pnas.0901635106
- Andela, N., Morton, D., Giglio, L., Chen, Y., van der Werf, G., Kasibhatla, P., et al. (2017). A human-driven decline in global burned area. *Science* 356, 1356–1362. doi: 10.1126/science.aal4108.
- Azeria, E. T., Bouchard, M., Pothier, D., Fortin, D., and Hébert, C. (2011). Using biodiversity deconstruction to disentangle assembly and diversity dynamics of understorey plants along post-fire succession in boreal forest. *Global Ecol. Biogeogr.* 20, 119–133. doi: 10.1111/j.1466-8238.2010.00580.x
- Bell, G. (2000). The distribution of abundance in neutral communities. *Am. Nat.* 155, 606–617. doi: 10.1086/303345
- Bennett, J. A., Lamb, E. G., Hall, J. C., Cardinal-McTeague, W. M., and Cahill, J. F. (2013). Increased competition does not lead to increased phylogenetic overdispersion in a native grassland. *Ecol. Lett.* 16, 1168–1176. doi: 10.1111/ele.12153
- Bernard-Verdier, M., Navas, M. L., Vellend, M., Violle, C., Fayolle, A., and Garnier, E. (2012). Community assembly along a soil depth gradient: contrasting patterns of plant trait convergence and divergence in a Mediterranean rangeland. J. Ecol. 100, 1422–1433. doi: 10.1111/1365-2745.12003
- Blomberg, S. P., Garland T. Jr., Ives, A. R., and Crespi, B. (2003). Testing for phylogenetic signal in comparative data: behavioral traits are more labile. *Evolution* 57, 717–745. doi: 10.1554/0014-3820(2003)057[0717:TFPSIC]2.0. CO;2
- Bond, W. J., and Keeley, J. E. (2005). Fire as a global 'herbivore': the ecology and evolution of flammable ecosystems. *Trends Ecol. Evol.* 20, 387–394. doi: 10.1016/j.tree.2005.04.025
- Burton, J. I., Perakis, S. S., McKenzie, S. C., Lawrence, C. E., and Puettmann, K. J. (2017). Intraspecific variability and reaction norms of forest understory plant species traits. *Funct. Ecol.* 31, 1881–1893. doi: 10.1111/1365-2435. 12898
- Cavender-Bares, J., Ackerly, D., Baum, D., and Bazzaz, F. (2004). Phylogenetic overdispersion in Floridian oak communities. Am. Nat. 163, 823–843. doi: 10.1086/386375
- Chang, Y., He, H. S., Bishop, I., Hu, Y., Bu, R., Xu, C., et al. (2007). Longterm forest landscape responses to fire exclusion in the Great Xing'an Mountains, China. *Int. J. Wildland Fire* 16, 34–44. doi: 10.1071/WF 05093

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpls.2018. 01854/full#supplementary-material

- Chase, J. M. (2010). Stochastic community assembly causes higher biodiversity in more productive environments. *Science* 328, 1388–1391. doi: 10.1126/science.1187820
- Chen, H. Y., and Popadiouk, R. V. (2002). Dynamics of North American boreal mixedwoods. *Environ. Rev.* 10, 137–166. doi: 10.1139/a02-007
- Chen, Z., Zhang, X., He, X., Davi, N. K., Cui, M., and Peng, J. (2013). Extension of summer (June-August) temperature records for northern Inner Mongolia (1715–2008), China using tree rings. *Quatern. Int.* 283, 21–29. doi: 10.1016/j.quaint.2012.07.005
- Chesson, P. (2000). Mechanisms of maintenance of species diversity. Annu. Rev. Ecol. Syst. 31, 343–366. doi: 10.1146/annurev.ecolsys.31.1.343
- Chesson, P., Gebauer, R. L., Schwinning, S., Huntly, N., Wiegand, K., Ernest, M. S., et al. (2004). Resource pulses, species interactions, and diversity maintenance in arid and semi-arid environments. *Oecologia* 141, 236–253. doi: 10.1007/s00442-004-1551-1
- Chipman, S. J., and Johnson, E. (2002). Understory vascular plant species diversity in the mixedwood boreal forest of western Canada. *Ecol. Appl.* 12, 588–601. doi: 10.1890/1051-0761(2002)012[0588:UVPSDI]2.0.CO;2
- Clark, J. S., and McLachlan, J. S. (2003). Stability of forest biodiversity. *Nature* 423, 635–638. doi: 10.1038/nature01632
- Condit, R., Pitman, N., Leigh, E. G., Chave, J., Terborgh, J., Foster, R. B., et al. (2002). Beta-diversity in tropical forest trees. *Science* 295, 666–669. doi: 10.1126/science.1066854
- Cornwell, W. K., Schwilk, D. W., and Ackerly, D. D. (2006). A trait-based test for habitat filtering: convex hull volume. *Ecology* 87, 1465–1471. doi: 10.1890/0012-9658(2006)87[1465:ATTFHF]2.0.CO;2
- Fritschie, K. J., Cardinale, B. J., Alexandrou, M. A., and Oakley, T. H. (2014). Evolutionary history and the strength of species interactions: testing the phylogenetic limiting similarity hypothesis. *Ecology* 95, 1407–1417. doi: 10.1890/13-0986.1
- Garnier, E., Cortez, J., Billès, G., Navas, M.-L., Roumet, C., Debussche, M., et al. (2004). Plant functional markers capture ecosystem properties during secondary succession. *Ecology* 85, 2630–2637. doi: 10.1890/03-0799
- Gerhold, P., Cahill, J. F., Winter, M., Bartish, I. V., and Prinzing, A. (2015). Phylogenetic patterns are not proxies of community assembly mechanisms (they are far better). *Funct. Ecol.* 29, 600–614. doi: 10.1111/1365-2435.12425
- Gilbert, B., and Lechowicz, M. J. (2004). Neutrality, niches, and dispersal in a temperate forest understory. *Proc. Natl. Acad. Sci. U.S.A.* 101, 7651–7656. doi: 10.1073/pnas.0400814101
- Giraudoux, P. (2017). *Pgirmess: Data Analysis in Ecology. R Package Version 1.6.7.* Available online at: https://CRAN.R-project.org/package=pgirmess
- Gong, Z. (2001). Chinese Soil Taxonomy. Beijing: Science Press.
- Grime, J. P. (2006). Trait convergence and trait divergence in herbaceous plant communities: mechanisms and consequences. J. Veg. Sci. 17, 255–260. doi: 10. 1658/1100-9233(2006)17[255:TCATDI]2.0.CO;2

- Hart, S. A., and Chen, H. Y. (2006). Understory vegetation dynamics of North American boreal forests. *Crit. Rev. Plant Sci.* 25, 381–397. doi: 10.1080/07352680600819286
- Hart, S. A., and Chen, H. Y. (2008). Fire, logging, and overstory affect understory abundance, diversity, and composition in boreal forest. *Ecol. Monogr.* 78, 123–140. doi: 10.1890/06-2140.1
- Hubbell, S. P. (2001). *The Unified Neutral Theory of Biodiversity and Biogeography*. Princeton: Princeton University Press.
- Jolly, W. M., Cochrane, M. A., Freeborn, P. H., Holden, Z. A., Brown, T. J., Williamson, G. J., et al. (2015). Climate-induced variations in global wildfire danger from 1979 to 2013. *Nat. Commun.* 6:7537. doi: 10.1038/ncomms8537
- Kembel, S. W. (2009). Disentangling niche and neutral influences on community assembly: assessing the performance of community phylogenetic structure tests. *Ecol. Lett.* 12, 949–960. doi: 10.1111/j.1461-0248.2009.01354.x
- Kembel, S. W., Cowan, P. D., Helmus, M. R., Cornwell, W. K., Morlon, H., Ackerly, D. D., et al. (2010). Picante: R tools for integrating phylogenies and ecology. *Bioinformatics* 26, 1463–1464. doi: 10.1093/bioinformatics/btq166
- Kloke, J. D., and McKean, J. W. (2012). Rfit: Rank-based estimation for linear models. R J. 4, 57-64.
- Kooyman, R., Rossetto, M., Cornwell, W., and Westoby, M. (2011). Phylogenetic tests of community assembly across regional to continental scales in tropical and subtropical rain forests. *Global Ecol. Biogeogr.* 20, 707–716. doi: 10.1111/j.1466-8238.2010.00641.x
- Kraft, N. J., and Ackerly, D. D. (2010). Functional trait and phylogenetic tests of community assembly across spatial scales in an Amazonian forest. *Ecol. Monogr.* 80, 401–422. doi: 10.1890/09-1672.1
- Kumar, P., Chen, H. Y., Thomas, S. C., and Shahi, C. (2017). Linking resource availability and heterogeneity to understorey species diversity through succession in boreal forest of Canada. J. Ecol. 106, 1–11. doi: 10.1111/1365-2745.12861
- Lapiedra, O., Sol, D., Traveset, A., and Vilà, M. (2015). Random processes and phylogenetic loss caused by plant invasions. *Global Ecol. Biogeogr.* 24, 774–785. doi: 10.1111/geb.12310.
- Letcher, S. G., Chazdon, R. L., Andrade, A. C., Bongers, F., van Breugel, M., Finegan, B., et al. (2012). Phylogenetic community structure during succession: evidence from three Neotropical forest sites. *Perspect. Plant Ecol. Evol. Syst.* 14, 79–87. doi: 10.1016/j.ppees.2011.09.005
- Li, S-p., Cadotte, M. W., Meiners, S. J., Hua, Z-s., and Shu W-s. (2015). Species colonisation, not competitive exclusion, drives community overdispersion over long-term succession. *Ecol. Lett.* 18, 964–973. doi: 10.1111/ele.12476
- Liu, Z., Yang, J., Chang, Y., Weisberg, P. J., and He, H. S. (2012). Spatial patterns and drivers of fire occurrence and its future trend under climate change in a boreal forest of Northeast China. *Global Change Biol.* 18, 2041–2056. doi: 10.1111/j.1365-2486.2012.02649.x
- Losos, J. B. (2008). Phylogenetic niche conservatism, phylogenetic signal and the relationship between phylogenetic relatedness and ecological similarity among species. *Ecol. Lett.* 11, 995–1003. doi: 10.1111/j.1461-0248.2008.01229.x
- Mason, N. W., Lanoiselée, C., Mouillot, D., Wilson, J. B., and Argillier, C. (2008). Does niche overlap control relative abundance in French lacustrine fish communities? A new method incorporating functional traits. *J. Anim. Ecol.* 77, 661–669. doi: 10.1111/j.1365-2656.2008.01379.x
- Mueller Dombois, D., and Ellenberg, H. (1974). Aims and Methods of Vegetation Ecology. New York, NY: Wiley.
- Muscarella, R., Uriarte, M., Aide, T. M., Erickson, D. L., Forero-Montaña, J., Kress, W. J., et al. (2016). Functional convergence and phylogenetic divergence during secondary succession of subtropical wet forests in Puerto Rico. J. Veg. Sci. 27, 283–294. doi: 10.1111/jvs.12354
- Nilsson, M.-C., and Wardle, D. A. (2005). Understory vegetation as a forest ecosystem driver: evidence from the northern Swedish boreal forest. *Front. Ecol. Environ.* 3, 421–428. doi: 10.1890/1540-9295(2005)003[0421:UVAAFE]2. 0.CO;2
- Norden, N., Letcher, S. G., Boukili, V., Swenson, N. G., and Chazdon, R. (2012). Demographic drivers of successional changes in phylogenetic structure across life-history stages in plant communities. *Ecology* 93, S70–S82. doi: 10.1890/10-2179.1
- Pausas, J. G., Bradstock, R. A., Keith, D. A., and Keeley, J. E. (2004). Plant functional traits in relation to fire in crown-fire ecosystems. *Ecology* 85, 1085–1100. doi: 10.1890/02-4094

- Pausas, J. G., and Keeley, J. E. (2014). Evolutionary ecology of resprouting and seeding in fire-prone ecosystems. *New Phytol.* 204, 55–65. doi: 10.1111/nph.12921
- Pausas, J. G., and Verd, M. (2010). The jungle of methods for evaluating phenotypic and phylogenetic structure of communities. *Bioscience* 60, 614–625. doi: 10.1525/bio.2010.60.8.7
- Pérez-Harguindeguy, N., Díaz, S., Garnier, E., Lavorel, S., Poorter, H., Jaureguiberry, P., et al. (2013). New handbook for standardised measurement of plant functional traits worldwide. *Aust. J. Bot.* 61, 167–234. doi: 10.1071/BT12225
- Purschke, O., Schmid, B. C., Sykes, M. T., Poschlod, P., Michalski, S. G., Durka, W., et al. (2013). Contrasting changes in taxonomic, phylogenetic and functional diversity during a long-term succession: insights into assembly processes. J. Ecol. 101, 857–866. doi: 10.1111/1365-2745.12098
- R Development Core Team (2017). R: A Language and Environment for Statistical Computing. Version 3.4.1. Vienna: R Foundation for Statistical Computing. Available online at: http://www.r-project.org/
- Revell, L. J. (2013). Two new graphical methods for mapping trait evolution on phylogenies. *Methods Ecol. Evol.* 4, 754–759. doi: 10.1111/2041-210X.12066
- Revell, L. J., Harmon, L. J., and Collar, D. C. (2008). Phylogenetic signal, evolutionary process, and rate. *Syst. Biol.* 57, 591–601. doi: 10.1080/10635150802302427
- Spasojevic, M. J., and Suding, K. N. (2012). Inferring community assembly mechanisms from functional diversity patterns: the importance of multiple assembly processes. J. Ecol. 100, 652–661. doi: 10.1111/j.1365-2745.2011.01945.x
- Stephens, S., Agee, J. K., Ful,é, P., North, M., Romme, W., Swetnam, T., et al. (2013). Managing forests and fire in changing climates. *Science* 342, 41–42. doi: 10.1126/science.1240294
- Stevens, P. (2001). Angiosperm Phylogeny Website. Version 14, July 2017. Available online at: http://www.mobot.org/MOBOT/research/APweb
- Swenson, N. G., Erickson, D. L., Mi, X., Bourg, N. A., Forero-Montaña, J., Ge, X., et al. (2012a). Phylogenetic and functional alpha and beta diversity in temperate and tropical tree communities. *Ecology* 93, S112–S125. doi: 10.1890/11-0402.1
- Swenson, N. G., Stegen, J. C., Davies, S. J., Erickson, D. L., Forero-Montaña, J., Hurlbert, A. H., et al. (2012b). Temporal turnover in the composition of tropical tree communities: functional determinism and phylogenetic stochasticity. *Ecology* 93, 490–499. doi: 10.1890/11-1180.1
- Ulrich, W., Zaplata, M. K., Winter, S., Schaaf, W., Fischer, A., Soliveres, S., et al. (2016). Species interactions and random dispersal rather than habitat filtering drive community assembly during early plant succession. *Oikos* 125, 698–707. doi: 10.1111/oik.02658
- Uriarte, M., Swenson, N. G., Chazdon, R. L., Comita, L. S., John Kress, W., Erickson, D., et al. (2010). Trait similarity, shared ancestry and the structure of neighbourhood interactions in a subtropical wet forest: implications for community assembly. *Ecol. Lett.* 13, 1503–1514. doi: 10.1111/j.1461-0248.2010.01541.x
- Verdú, M., and Pausas, J. (2007). Fire drives phylogenetic clustering in Mediterranean Basin woody plant communities. J. Ecol. 95, 1316–1323. doi: 10.1111/j.1365-2745.2007.01300.x
- Verdú, M., Rey, P. J., Alcantara, J. M., Siles, G., and Valiente-Banuet, A. (2009). Phylogenetic signatures of facilitation and competition in successional communities. J. Ecol. 97, 1171–1180. doi: 10.1111/j.1365-2745.2009.01565.x
- Walker, L. R., Wardle, D. A., Bardgett, R. D., and Clarkson, B. D. (2010). The use of chronosequences in studies of ecological succession and soil development. *J. Ecol.* 98, 725–736. doi: 10.1111/j.1365-2745.2010.01664.x
- Wang, X., Wiegand, T., Swenson, N. G., Wolf, A. T., Howe, R. W., Hao, Z., et al. (2015). Mechanisms underlying local functional and phylogenetic beta diversity in two temperate forests. *Ecology* 96, 1062–1073. doi: 10.1890/14-0392.1
- Webb, C. O. (2000). Exploring the phylogenetic structure of ecological communities: an example for rain forest trees. Am. Nat. 156, 145–155. doi: 10.1086/303378
- Webb, C. O., Ackerly, D. D., McPeek, M. A., and Donoghue, M. J. (2002). Phylogenies and community ecology. Annu. Rev. Ecol. Syst. 33, 475–505. doi: 10.1146/annurev.ecolsys.33.010802.150448
- Weiher, E., Clarke, G. P., and Keddy, P. A. (1998). Community assembly rules, morphological dispersion, and the coexistence of plant species. *Oikos* 81, 309–322.

- Whitfeld, T., Kress, W., Erickson, D., and Weiblen, G. (2012). Change in community phylogenetic structure during tropical forest succession: evidence from New Guinea. *Ecography* 35, 821–830. doi: 10.1111/j.1600-0587.2011.07181.x
- Wilson, J. B. (2007). Trait-divergence assembly rules have been demonstrated: limiting similarity lives! A reply to Grime. J. Veg. Sci. 18, 451–452. doi: 10.1658/ 1100-9233(2007)18[451:TARHBD]2.0.CO;2
- Wilson, J. B., and Stubbs, W. J. (2012). Evidence for assembly rules: limiting similarity within a saltmarsh. J. Ecol. 100, 210–221. doi: 10.1111/j.1365-2745.2011.01891.x
- Zanne, A. E., Tank, D. C., Cornwell, W. K., Eastman, J. M., Smith, S. A., FitzJohn, R. G., et al. (2014). Three keys to the radiation of angiosperms into freezing environments. *Nature* 506, 89–92. doi: 10.1038/nature12872

Zhou, Y. (1991). Vegetation of Da Hinggan Ling in China. Beijing: Science Press.

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