



# A Combination of Long-Day Suppressor Genes Contributes to the

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Northward Expansion of Rice Yongxia Cui<sup>1†</sup>, Junrui Wang<sup>1†</sup>, Li Feng<sup>1†</sup>, Sha Liu<sup>1</sup>, Jiagi Li<sup>1</sup>, Weihua Qiao<sup>1</sup>, Yue Song<sup>1</sup>,

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Growing cultivated rice with a moderate heading date is the key to expanding its cultivation area and maintaining stable yields. The genes that regulate heading date are largely cloned; however, it remains unclear how genetic mutations and their combinations affect the heading date and adaptability of cultivated rice. Here, we report the analysis of genetic variation in eight long-day flowering suppressor genes (Hd1, DTH8, Ghd7, OsCOL4, DTH7, Hd6, Se5, and PhyB) and the phylogenetic relationship of eight genes. Genetic variations in DTH8, Ghd7, Hd1, DTH7, PhyB, and OsCOL4 are correlated with differences in heading date and the correlation between the genetic diversity of Hd6 and Se5 and rice heading data are weak. One group of haplotypes of DTH8, Ghd7, Hd1, DTH7, PhyB, and OsCOL4 are associated with earlier heading dates and appear to have accumulated during the northward expansion of rice cultivation. A minimum of four group A alleles of DTH8, Ghd7, Hd1, DTH7, PhyB, and OsCOL4 are required for the growth of cultivated rice at latitudes above 30°N. This study presents a preliminary investigation of the genetic patterns and adaptation mechanisms of long-day flowering suppressor genes and provides a useful reference for the molecular breeding of rice cultivars for various environments and farming systems.

Keywords: Oryza sativa, long-day suppressor genes, domestication, northward expansion, heading date

#### INTRODUCTION

To make full use of sunlight and temperature resources and to ensure food security, it is necessary to selectively breed rice varieties whose heading dates are suited to the climatic characteristics of different regions (Itoh et al., 2018). The ancestral species of cultivated rice (Oryza rufipogon Griff.) is a short-day sensitive plant that grows mainly in low-latitude tropical and subtropical regions. After natural selection and artificial domestication, the geographical distribution of cultivated rice gradually expanded from 45°N to 23°S (Monfreda et al., 2008; Huang et al., 2012), growing under both long-day and short-day conditions. During the expansion process, it was important to cultivate varieties with appropriate heading dates. Therefore, exploring the genetic mechanisms that

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underlie heading date diversity in cultivated rice is important for understanding its domestication history and the cultivation of eurytopic rice species.

Previous studies have shown that photoperiod is an important factor that affects the heading date of cultivated rice (Shang et al., 2012). To date, many genes in the rice photoperiodic regulatory network have been cloned (Gao et al., 2014). These include Heading date 3a (Hd3a) and RICE FLOWERING-LOCUS T 1 (RFT1), two florigen genes that act as biological signals to complete the phase transition from vegetative to reproductive growth (Kojima et al., 2002; Komiya et al., 2008). Heading date 1 (Hd1) and Early heading date 1 (Ehd1) are two hub genes in the photoperiodic regulatory pathway that have different responses to day length. They control the formation of florigen genes by receiving multiple regulatory signals from upstream genes and then influencing heading date (Yano et al., 2000; Wei et al., 2016). In rice, Hd1 suppresses flowering under long-day conditions by interacted with Ghd7 to repress the expression of Ehd1 (Nemoto et al., 2016), but promotes flowering under short-day conditions (Yano et al., 2000). Ehd1 promotes flowering under both long- and short-day conditions (Wei et al., 2016). Under short-day conditions, Ehd1 is regulated by multiple upstream genes that include OsGIGANTEA (OsGI), Early heading date 2 (Ehd2), and MADS BOX GENE 51 (OsMAD51) (Matsubara et al., 2008; Ryu et al., 2009). Under long-day conditions, Ehd1 is directly or indirectly regulated by MADS BOX GENE 50 (OsMAD50), Early heading date 2 (Ehd2), Early heading date 3 (Ehd3), and Early heading date 4 (Ehd4). Furthermore, HEADING DATE 6 (Hd6), Heading date (QTL)-5 (t) (DTH8), Heading date 2 (DTH7), CONSTANS-like gene 4 (OsCOL4), HEADING DATE 7 (Ghd7), PHYTOCHROME B (*PhyB*), and PHOTOSENSITIVITY 5 (*Se5*) also directly or indirectly suppress Ehd1 expression (Dehesh et al., 1991; Takahashi et al., 2001; Xue et al., 2008; Lee et al., 2010; Wei et al., 2010; Hyuk et al., 2013; Du et al., 2017). In summary, the photoperiodic regulatory network in cultivated rice can be divided into short-day promotion, long-day promotion, and long-day suppression pathways.

Previous research suggests that there is a correlation between genetic diversity of long-day suppressor genes and heading date, and that this diversity was critical for the expansion of cultivated rice production to higher latitudes (Wu et al., 2012; Zheng et al., 2016). Takahashi et al. (2009) found that genetic diversity of Hd1 was an important reason for flowering date diversity in cultivated rice. They showed that a loss of function of Hd1 occurred 8,000 to 10,000 years ago and played an important role in the adaptation of rice crops to higher latitudes and in the development of various growing strategies (Takahashi et al., 2009). Wu et al. (2012) reported that genetic variation in DTH2 was closely related to the northward expansion of Asian cultivated rice (Wu et al., 2012). Koo et al. (2013) found that genetic mutations in OsPRR37 and Ghd7 could promote early flowering of cultivated rice during heading under long-day conditions, thereby adapting cultivated rice to higher latitudes (Koo et al., 2013). Gao et al. (2014) showed that different haplotype combinations of the long-day suppressor genes

*DTH7*, *DTH8*, and *Ghd7* were closely related to heading date diversity in cultivated rice. Among three-gene haplotype combinations, the combination that included loss of function in *DTH7*, *DTH8*, and *Ghd7* was found mainly in cultivated rice in northern China (Gao et al., 2014).

The aforementioned studies were performed on a limited number of long-day suppressor genes. Less is known about the genetic diversity of long-day suppressor gene pathways and about the accumulation effects of different haplotype combinations of eight long day suppression gene on heading date in cultivated rice. Therefore, in this study, we collected rice cultivars from around the world and analyzed the genetic diversity of eight important long-day suppressor genes: *Hd1*, *DTH8*, *Ghd7*, *OsCOL4*, *DTH7*, *Hd6*, *Se5*, and *PhyB*. We then analyzed the effects of individual haplotypes and haplotype combinations of different long-day suppressor genes on heading date and geographic distribution to better understand the molecular underpinnings of heading date diversity in cultivated rice.

## MATERIALS AND METHODS

## **Plant Materials and Phenotyping**

One hundred and forty-eight samples, including 83 from cultivated rice (Oryza sativa L.) and 65 from wild rice (O.nivara and O.rufipogon) were included in the study. Among these accessions, the 83 cultivated rice accessions covered approximately all subgroups and rice planting areas from Northeastern to Southern China and from Central Asia to South Asia and Southeast Asia (Table S1, Figure S1). The 65 wild rice accessions covered most of tropical and subtropical of Asia from Southern China (28°N) to Indonesia (10°S). Detailed information about the accessions, including names, countries of origin, geographic locations, and subpopulation classifications, is presented in Table S1. All seeds were breaked dormancy by keeping at 50°C, 72 h, then soaked for 48 h in water; they were then planted in nutritious soil and raised in wet-bed condition for 23-26 days for reaching the seedling stage. Seedlings were then transplanted into the same agricultural field in Peking (116° 13'E, 39°54'N) on May 5, 2016, May 5, 2017, and May 5, 2018, respectively during the summer under natural long-day conditions. Seedlings from each accession were planted in a single row of 10 plants in each of the years. Field management was performed according to typical rice cultivation practices. Heading date was scored for each plant as the number of days from the sowing date to the date at which the first panicle measured 1-2 cm; The mean heading date of 30 plants per accession for the 3 years was calculated (Table S1). Of the 83 cultivated rice accessions, 15 did not have heading date data because the seedlings did not survive.

## DNA Isolation, PCR Amplification and Sequencing

Fresh leaves were collected from field-grown rice plants after rice flowering, and genomic DNA was isolated from the fresh tissue

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by a modified CTAB method and used to amplify the coding regions of *DTH8*, *Ghd7*, *Hd1*, *DTH7*, *PhyB*, *OsCOL4*, *Hd6*, and *Se5*. PCR amplification methods followed standard PCR protocols as described in Zheng and Ge (Zheng and Ge, 2010). The genome sequence of Nipponbare (*O. sativa* ssp. *japonica*) was downloaded from NCBI and used as the reference sequence for primer design (**Figure S2** and **Table S2**). PCR products were sequenced using an ABI 3730 automatic sequencer (Applied Biosystem, Foster City, CA).

#### **Sequence Analysis**

DNA Sequence alignment was performed with ClustalX 2.1 (Larkin et al., 2007) and manually adjusted in BioEdit (http://www.mbio.ncsu.edu). Polymorphic sites were analyzed using DnaSP5.0 (Huelsenbeck and Ronquist, 2001), and insertions/deletions (indels) were included in the analysis. Gaps with a length greater than one were considered to be single mutations. Network trees for the eight genes of interest were constructed using the median-joining model of DnaSP5.0 and Network 4.0 (Bandelt et al., 1999), and divided *DTH8*, *Ghd7*, *Hd1*, *DTH7*, *PhyB*, and *OsCOL4* into two groups (A and B) based on genetic distance.

#### **Statistical Analysis**

To determine whether haplotype in groups A and B of the six long-day suppression genes (DTH8, Ghd7, Hd1, DTH7, PhyB and OsCOL4) differed in heading date, we constructed boxplots of rice heading date data using R version 3.5.2 and used paired ttest to assess significant differences. We then investigated differences in frequency of haplotype groups A and B with latitude from 45°N to 23°S by constructing a heatmap in Excel based on rice haplotype coordinate data. Ultimately, to analyze the geographic distribution of group A and group B samples, geographic distribution maps of rice from haplotype groups A and B of each gene were constructed using leaflet function in R based on geographic coordinates. To explore the association between haplotype combinations and climate factors, raw data for Minimum Temperature of Coldest Month were download from the WorldClim Version 2 database (http://worldclim.org/ bioclim) at a 30 s (~1 km) resolution. Using sp and rgdal function in R, the data were converted into a climate density map, and haplotype combination data points for OsCOL4, DTH7, PhyB, Ghd7, and DTH8 were overlaid onto the map based on geographic coordinate data. To determine the relationships between haplotype combination and heading date or latitude, we again constructed boxplots in R. The effect of haplotype combination was assessed using the Anova function in R, and mean separations were performed using the LSD function in R (Grunsky, 2002).

#### RESULTS

### **Genetic Variation**

One hundred and forty-eight rice samples were collected from major rice cropping areas, including 83 representative cultivated rice samples and 65 wild rice samples (**Table S1**, **Figure S1**). The coding regions of eight long-day suppressor genes (DTH8, Ghd7, Hd1, DTH7, PhyB, OsCOL4, Hd6, and Se58) were sequenced using the primers in Table S2. Diagrams of their gene structures are shown in Figure S2. Exon numbers ranged from one to ten, with one exon in DTH8 and 10 in Hd6. Sequence lengths ranged from 774 bp to 3516 bp, and 233 SNPs were found among all the sequences (25, 47, 41, 59, 30, 19, 10, and 2 SNPs in DTH8, Ghd7, Hd1, DTH7, PhyB, OsCOL4, Hd6, and Se5, respectively). Among these SNPs, all mutations in Se5 were silent mutations that did not affect the coding for amino acids. More than half of the remaining seven genes had SNPs that led to changes in amino acid sequence, and most of the biologically significant mutations were located in the first exon (Figure S3). Three, 2, 1, 5, and 1 SNPs that generated premature stop codons were identified in DTH8, DTH7, Ghd7, Hd1, and Hd6, respectively (Figure S3). Three, 4, 2, 6, 3, 1, and 1 indels were identified in DTH8, DTH7, Ghd7, Hd1, PhyB, OsCOL4, and Hd6, respectively (Figure S3); there were no indels in Se5 (Figure S3). There was greater genetic diversity in DTH8, DTH7, Ghd7, Hd1, PhyB, and OsCOL4 than in *Hd6* and *se5*.

#### **Haplotype Analysis**

To assess the genetic relationships among taxa we first calculated the numbers of haplotypes and constructed network tree. Mutation sites of each gene in the 148 samples were analyzed, and 18, 41, 41, 42, 28, 22, 3, and 13 haplotypes were found in DTH8, DTH7, Ghd7, Hd1, PhyB, OsCOL4, Se5, and Hd6, respectively (Figure S3). The number of haplotypes with a frequency greater than 5% was 6, 6, 2, 6, 2, 5, 2, and 2 in DTH8, DTH7, Ghd7, Hd1, PhyB, OsCOL4, Se5, and Hd6, respectively. Frequencies of these haplotypes ranged from 5.10% to 86% (Table S3). Network trees for the eight genes were constructed using the median-joining (MJ) method. Based on the relationship among haplotypes and the haplotype frequency in population, we found that DTH8, DTH7, Ghd7, Hd1, PhyB, and OsCOL4 were all centered around highfrequency haplotypes and clearly differentiated into two groups, which we called group A and group B (Figure 1 and Figure S3). In DTH8, DTH7, Ghd7, PhyB, and OsCOL4, we found that the ancestral haplotypes in group B were mainly composed of *indica* and wild rice, whereas the ancestral haplotypes in group A were mainly composed of japonica and wild rice grown in China (Figure 1).

Additionally, We also analyzed the geographical distribution of samples from both haplotype groups. Group A haplotypes were found at the greatest frequency in cultivated rice from highlatitude regions basically, including Heilongjiang, Jilin, Liaoning, Shanxi, Beijing, and Tianjin in China, as well as from Japan and South Korea. Group B haplotypes were found at the greatest frequency mainly in cultivated rice from the southern regions of China, as well as from tropical and subtropical regions of South and Southeast Asia (**Figures 1** and **2**, **Table S1**). By contrast, although the *Hd1* gene was also centered on high-frequency haplotypes and separated into two groups (i.e. group A and group B), the composition and geographical distribution of subcultivars in groups A and B were different from those of *DTH8*, *DTH7*, *Ghd7*, *PhyB*, and *OsCOL4*. Specifically, both

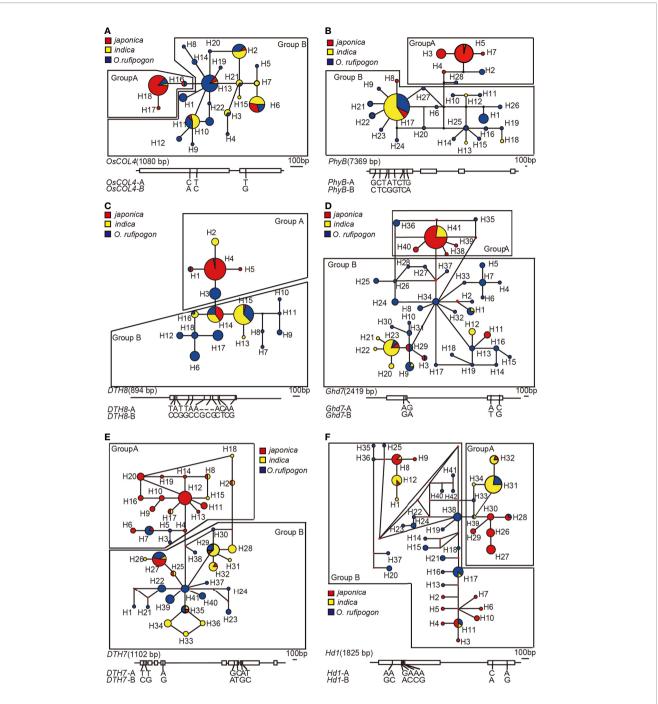
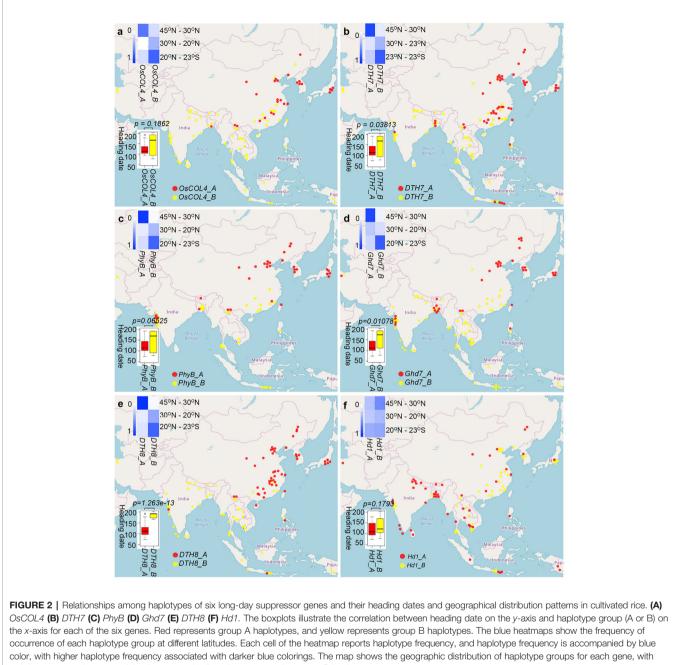


FIGURE 1 | Network trees of six long-day suppressor genes in cultivated rice. (A) OsCOL4: H\_2, H\_6, H\_11, H\_13, and H\_18 are high-frequency haplotypes of OsCOL4. (B) PhyB: H\_5 and H\_17 are high-frequency haplotypes of PhyB. (C) DTH8: H\_3, H\_4, H\_6, H\_14, H\_15, and H\_17 are high-frequency haplotypes of DTH8. (D) Ghd7: H\_23 and H\_41 are high-frequency haplotypes of Ghd7. (E) DTH7: H\_7, H\_12, H\_27, H\_29, and H\_32 are high-frequency haplotypes of DTH7. (F) Hd1: H\_8, Hap\_11, H\_12, H\_17, H\_27 and H\_31 are high-frequency haplotypes of Hd1. The size of the circle in each gene's network tree represents the haplotype frequency. Yellow and red represent the two subcultivars of cultivated rice, *indica* and *japonica*, and blue represents O.rufipogon. Each gene's network tree corresponds to the SNP on the diagram of long-day suppressor genes below, which illustrates the haplotype-specific SNPs between group A and group B of the long-day suppressor genes.



red dots representing group A haplotypes and yellow dots representing group B haplotypes.

*japonica* and *indica* were found in group A and group B, and cultivated rice species in both groups were distributed from 45°N to 23°S. *Hd6* and *Se5* had only one high-frequency haplotype apiece, and these haplotypes accounted for 81% and 86% of the total haplotypes of the two genes, respectively (**Figure S4**).

#### Relationship Between Heading Date of Cultivated Rice and Haplotype of Long-Day Suppressor Genes

To explore the heading date differences between group A and group B. Next, we compared long-day suppressor gene haplotypes

with heading dates in cultivated rice. Due to the low genetic diversity of *Se5* and *Hd6*, we only analyzed heading date differences between group A and group B haplotypes for the remaining six genes (**Figure 2**; **Table S4**). For cultivated rice in haplotype group A of *DTH8*, *DTH7*, *Ghd7*, *PhyB*, *OsCOL4*, and *Hd1*, the average heading dates were 116.9, 120.8, 121.1, 121.6, 124.8, and 119.5 days, respectively. By contrast, for cultivated rice in haplotype group B of the same genes, the average heading dates were 185.0, 148.7, 156.5, 147.5, 143.2, and 137.6 days, respectively. Therefore, under long-day conditions, the heading dates of the group A samples were earlier than those of the group B samples.

Differences in heading date between group A and group B were statistically significant for DTH8 (t = -10.802, P = 1.263e-13, ttest), DTH7 (t = -2.1884, P = 0.03813, t-test), and Ghd7 (t = -2.7768, P = 0.01078, *t*-test) (Figure 2). The difference between the two groups for *PhyB* was close to significance (t = -1.9479, P =0.06525, t-test), and differences for the remaining genes were not significant: OsCOL4 (t = -1.3603, P = 0.1862, t-test) and Hd1 (t =-1.4024, P = 0.1793, *t*-test). Taken together, our association studies suggest that group A and group B of DTH8, DTH7, Ghd7, PhyB, OsCOL4, and Hd1 are different in terms of heading date regulation. We have added 10 random sequences (Zhu et al., 2007; Wu et al., 2012). Based on these neutral sequence data, congruence between data sets was evaluated using the partition homogeneity test (Farris et al., 1995), as implemented in PAUP with 1,000 replicates, random taxon addition, and one tree saved per replicate. There was significant incongruence between any gene (p=0.0017 for DTH8, p=0.024 for Ghd7, p=0.0033 for Hd1, p=0.0042 for DTH7, p=0.019 for PhyB, p=0.0063 for OsCOL4). The resulting *p*-value was used to determine whether the data sets had significant incongruence.

#### Geographical Evolution Models of Long-Day Suppressor Genes

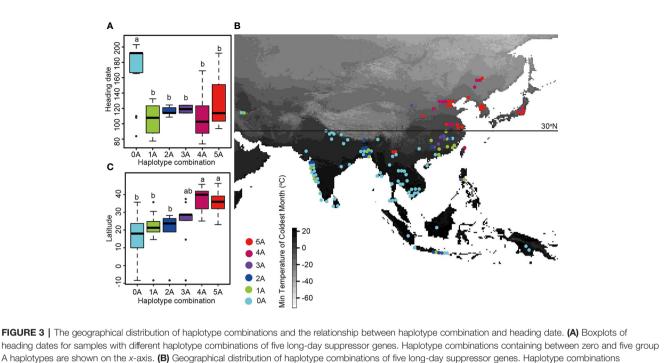
Generally, a single gene does not fully explain the genetic basis of different cultivated rice to adapt to different ecological regions. So we next analyzed the geographical distribution of haplotype combinations of the long-day suppressor genes (Figure 3, Table S1). Because the composition and geographical distribution of Hd1 subcultivars in group A and group B were different from those of the other genes, we only analyzed the geographical distribution of haplotype combinations of the five long-day suppressor genes for group A and group B haplotypes in DTH8, DTH7, Ghd7, PhyB, and OsCOL4 (Figure 2, Table S1). Twentyone samples contained all five group A haplotypes and accounted for 14.2% of all individuals. These samples were composed of japonica and were distributed in Japan, South Korea, the Chinese region north to the latitude of 30 degree (Zhejiang, Shanghai, Anhui, Jiangsu, Tianjin, and Shanxi), the high latitudes of Yunnan Province, etc. The average heading date of these samples was 127.12 days. Eleven samples had four group A haplotypes and accounted for 7.4% of all individuals. These samples were composed of japonica and were distributed in Taiwan, Fujian, Anhui, Jiangsu, Shanxi, Beijing, Jilin, Heilongjiang, etc. The average heading date of these samples was 108.9 days. Ten samples had three group A haplotypes and accounted for 6.8% of all individuals. These samples were composed primarily of japonica and wild rice, and they were mainly distributed near the Yangtze River in China (Hunan and Jiangxi), as well as in Japan, Indonesia, and high latitudes of the Philippines. The average heading date of these samples was 119.07 days. Twelve samples had two group A haplotypes and accounted for 8.1% of all individuals. They comprised three japonica, six indica and three wild rice. These individuals were distributed in Hunan and Jiangxi, China, as well as in Indonesia, India, Bangladesh, Bhutan, etc. The average heading date of these samples was 116.39 days. Eighteen samples had one group A haplotype and

accounted for 12.2% of all individuals. They comprised three *japonica*, eight *indica*, and three wild rice, and they were distributed in the Philippines, India, Indonesia, Guangdong, Hainan, Bangladesh, and Iran, as well as in Jiangxi and Anhui, China. The average heading date of these samples was 96.3 days. Seventy-six samples had all five group B haplotypes and accounted for 51.4% of all individuals. They were composed of 22 *indica* and 53 wild rice, and they were mainly distributed in India, South and Southeast Asia, and a few regions in southern China. The average heading date of these samples was 171.94 days. These results indicate that if any of the six genes has a group A haplotype, the sample can grow north of the Tropic of Cancer. As cropping regions move northward, a greater number of haplotypes fall into group A (Gao et al., 2014).

## DISCUSSION

Flowering time is one of the best-studied ecologically significant traits for adaptation of plants to specific natural environments. Hd3a and RTF1 are two major florigen genes in the photoperiod regulation pathways. The upstream regulation pathway of Hd3a and RTF1 mainly include long-day suppression pathway, and long-day and short-day promote pathway. Among of them, the long-day suppression genes inhibits the expression of Hd3a and RTF1 under long-day condition, then inhibits the flowering of rice cultivars. So functional change of the long-day suppression genes is the genetic basis of rice flowering in advance. Some studies have also found that the long-day suppression gene as a "bridge" to affect the heading date of cultivated rice (Nemoto et al., 2016). Gao et al. (2014) found that rice flowering is controlled to combinations of DTH7, Ghd7, or DTH8 under various environments, especially under long-day conditions. In details, when the three genes are all non-function or weakfunction, the heading date of the cultivated rice is obviously earlier than the only one or two genes which are non-function. Zheng et al. (2016) also confirmed that the non-function or weak-function of the long-day suppression genes makes the optimal heading date of the cultivated rice by exploring longday suppression genes of Hd1, DTH8, Ghd7, and DTH7 (Zheng et al., 2016). In this study, we not only analyzed the genetic diversity, but also the genetic relationships of these haplotypes, and the relationship between haplotype combination and geographical distribution. These results give us some clues of interaction effects on flowering time difference and skewed distribution in the rice cultivars.

Rice cultivation covers a wide area, including China, South Asia, Southeast Asia, Japan, and South Korea. Among these regions, rice cultivation is most widely distributed in China, which has six rice cropping areas and sixteen sub-regions, including double-crop paddy areas in South China and Central China, single- and double-crop paddy areas in the Southwest Plateau, single-crop paddy areas in North China, early-maturing single-crop paddy areas in Northeast China, and single-crop paddy areas in the dry regions of Northwest China (**Figure S5**). Among of them, There are significant differences in heading date among these regions,



A haplotype are shown on the x-axis. (B) Geographical distribution of haplotype combinations of five long-day suppressor genes. Haplotype combinations containing between zero and five group A haplotypes are indicated by dots of different colors. The map is shaded to indicate the minimum temperature of the coldest month, with lower minimum temperatures in white and higher minimum temperatures in black. (C) Boxplots of latitude data for samples with different haplotype combinations of five long-day suppressor genes.

with a maximum difference of over 50 days (Wei et al., 2012). Rice is originally a tropical short-day crop; it requires various mutations and haplotype combinations to adapt to the natural environment when expanding into higher latitude cropping areas.

Genetic variation analysis of the long-day suppressor genes *Hd1*, DTH8, DTH7, OsCOL4, Ghd7, and PhyB indicated that Hd1 had the largest amount of the genetic variation. However, samples with the group A Hd1 haplotype were distributed across nearly the entire rice cultivation region from 23°S to 45°N, including South and Southeast Asia, all six rice cropping areas in China, and rice cropping areas in Japan and South Korea (Figure 2F). By contrast, the frequency of group A haplotypes of the remaining five long-day suppressor genes gradually increased when moving northward. Compared with Hd1, the geographic distribution map of DTH8 haplotype showed that group A haplotypes mainly appeared in six rice cropping areas in China, Japan and South Korea (Figure 2E). Compared with the aforementioned two genes, the haplotype geographic distribution map showed an even narrower latitude range for group A haplotypes of DTH7 and OsCOL4. These were mainly distributed in the double-crop paddy areas in Central China, the single- and double-crop paddy areas in the Southwest Plateau, the single-crop paddy areas in North China, the early-maturing single-crop paddy rice areas in Northeast China, the single-crop paddy areas in the dry regions of Northwest China, and other cropping areas in Japan and Korea (Figures 2A, B). Ghd7 and PhyB group A haplotypes were distributed in the single-crop paddy areas in North China, the earlymaturing single-crop paddy rice areas in Northeast China, the single-crop paddy areas in the dry regions of Northwest China, and other cropping areas in Japan and Korea (Figures 2C, D). In

summary, we believe that there is a gradual process of selection acting on long-day suppressor genes as rice cropping areas expand northward. In addition, the numbers of group A haplotypes differ among rice cropping areas, and the number of group A haplotypes of long-day suppressor genes can be increased to further shorten the heading date of cultivated rice in different areas.

Group A haplotypes of DTH8 were first found in double-crop paddy areas in South China, implying that the group A haplotypes of this gene were selected by nature/human and then accumulated in this area in order to grow cultivated rice that met the heading date requirements of the region (Figure 2E). However, with continued northward expansion of cultivated rice, a single DTH8 gene did not permit cultivated rice to adapt to the heading dates of the double-crop paddy areas in Central China and the single- and double-crop paddy areas in the Southwest Plateau. The haplotypes from OsCOL4 and DTH7 group A were selected and then accumulated in these two rice cropping areas (Figures 2A, B), gradually permitting development of cultivated rice adapted to the heading date requirements of these regions. It is worth mentioning that in the aforementioned three rice cropping areas, both group A and group B haplotypes were found in DTH8, OsCOL4, and DTH7 genes. However, when the cultivated rice cropping area moved northward above 30°N, group A haplotypes completely replaced group B haplotypes of these genes. In the rice-cropping areas north of 30°N, including the sub-regions of double-crop paddy areas in Central China, the single- and double-crop paddy areas in the lower-middle reaches of the Yangtze River, the doublematuring single-crop paddy rice areas in the Sichuan-Shaanxi

Basin, the single-crop paddy areas in dry regions of Northwest China, the single-crop paddy areas in North China, the earlymaturing single-crop paddy rice areas in Northeast China, and other cropping areas in Japan and Korea, group A haplotypes of *Ghd7* and *PhyB* began to appear and replaced group B haplotypes completely (**Figures 2C, D**). More than four genes among *Hd1*, *DTH8*, *DTH7*, *OsCOL4*, *Ghd7*, and *PhyB* must be of group A in order to meet the heading date requirements of cultivated rice in rice cropping areas north of 30°N.

In conclusion, our study demonstrates that during the northward expansion of cultivated rice, group A haplotypes of the long-day suppressor genes *DTH8*, *DTH7*, *OsCOL4*, *Ghd7*, and *PhyB* were selected to permit adaptation to various latitudes and climate conditions, resulting in the development of cultivated rice varieties with various heading dates in different cropping areas due to the additive effects of multiple genes. In the rice cropping areas north of 30°N, group A haplotypes in long-day suppressor genes replaced group B haplotypes, forming a clear dividing line at 30°N between haplotypes in groups A and B.

#### DATA AVAILABILITY STATEMENT

The datasets generated for this study can be found in the NCBI, Genbank number: DTH8 - MT500364 - MT500499; Ghd7 -MT453138 - MT453268; PhyB - MT460250 - MT460374; DTH7 - MT500239 - MT500363; OsCol4 - MT435799 - MT435928; Hd1 - MT500123 - MT500238; Hd6 - MT453303 - MT453434; and Se5 - MT435659 - MT435798.

#### REFERENCES

- Bandelt, H. J., Forster, P., and Rohl, A. (1999). Median-Joining Networks for Inferring Intraspecific Phylogenies. *Mol. Biol. Evol.* 16 (1), 37–48. doi: 10.1093/ oxfordjournals.molbev.a026036
- Dehesh, K., Tepperman, J., Christensen, A., and Quail, P. (1991). phyB is evolutionarily conserved and constitutively expressed in rice seedling shoots. *Mol. Gen. Genet.* 225 (2), 305–313. doi: 10.1007/BF00269863
- Du, A. P., Tian, W., Wei, M. H., Wei, Y., He, H., Zhou, D., et al. (2017). The DTH8-Hd1 Module Mediates Day-Length-Dependent Regulation of Rice Flowering. *Mol. Plant* 10 (7), 948–961. doi: 10.1016/j.molp.2017.05.006
- Farris, J. S., Mari, K., Kluge, A. G., and Bult, C. (1995). Constructing a Significance Test for Incongruence. Syst. Biol. 4 (4), 570–572. doi: 10.1093/sysbio/44.4.570
- Gao, H., Jin, M., Zheng, X. M., Chen, J., Yuan, D., Xin, Y., et al. (2014). Days to heading 7, a major quantitative locus determining photoperiod sensitivity and regional adaptation in rice. *Proc. Natl. Acad. Sci. U. S. A* 111 (46), 16337– 16342. doi: 10.1073/pnas.1418204111
- Grunsky, E. C. (2002). R: a data analysis and statistical programming environment an emerging tool for the geosciences. *Comput. Geosci.* 28, 1219–1222. doi: 10.1016/S0098-3004(02)00034-1
- Huang, C. L., Hung, C. Y., Chiang, Y. C., Hwang, C. C., Hsu, T. W., Huang, C. C., et al. (2012). Footprints of natural and artificial selection for photoperiod pathway genes in Oryza. *Plant J.* 70 (5), 769–782. doi: 10.1111/j.1365-313X.2012.04915.x
- Huelsenbeck, J. P., and Ronquist, F. (2001). MRBAYES: Bayesian inference of phylogenetic trees. *Bioinformatics* 17 (8), 754–755. doi: 10.1093/ bioinformatics/17.8.754
- Hyuk, K. B., Cheul, Y. S., Woo, P. J., Tak, K. C., Doo, L. B., Gynheung, A., et al. (2013). Natural Variation in OsPRR37 Regulates Heading Date and

#### **AUTHOR CONTRIBUTIONS**

QY and XZ were responsible for study design. LF and SL performed the experiments. JL and YS helped in preparing figures and tables. ZZ, WQ, YLC, and LZ were involved in resources collection. YXC, JW, and LF analyzed the data and wrote the manuscript with editing help from all co-authors. QY and XZ supervised.

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#### SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpls.2020.00864/ full#supplementary-material

Contributes to Rice Cultivation at a Wide Range of Latitudes. *Mol. Plant* 6 (6), 1877–1888. doi: 10.1093/mp/sst088

- Itoh, H., Wada, K. C., Sakai, H., Shibasaki, K., Fukuoka, S., Wu, J., et al. (2018). Genomic adaptation of flowering-time genes during the expansion of rice cultivation area. *Plant J.* 94 (5), 895–909. doi: 10.1111/tpj.13906
- Kojima, S., Takahashi, Y., Kobayashi, Y., Monna, L., Sasaki, T., Araki, T., et al. (2002). Hd3a, a rice ortholog of the Arabidopsis FT gene, promotes transition to flowering downstream of Hd1 under short-day conditions. *Plant Cell Physiol.* 43 (10), 1096–1105. doi: 10.1093/pcp/pcf156
- Komiya, R., Ikegami, A., Tamaki, S., Yokoi, S., and Shimamoto, K. (2008). Hd3a and RFT1 are essential for flowering in rice. *Development* 135 (4), 767–774. doi: 10.1242/dev.008631
- Koo, B. H., Yoo, S. C., Park, J. W., Kwon, C. T., Lee, B. D., An, G., et al. (2013). Natural variation in OsPRR37 regulates heading date and contributes to rice cultivation at a wide range of latitudes. *Mol. Plant* 6 (6), 1877–1888. doi: 10.1093/mp/sst088
- Larkin, M. A., Blackshields, G., Brown, N. P., Chenna, R., McGettigan, P. A., McWilliam, H., et al. (2007). Clustal W and Clustal X version 2.0. *Bioinformatics* 23 (21), 2947–2948. doi: 10.1093/bioinformatics/btm404
- Lee, Y. S., Jeong, D. H., Lee, D. Y., Yi, J., Ryu, C. H., Kim, S. L., et al. (2010). OsCOL4 is a constitutive flowering repressor upstream of Ehd1 and downstream of OsphyB. *Plant J.* 63 (1), 18–30. doi: 10.1111/j.1365-313X.2010.04226.x
- Matsubara, K., Yamanouchi, U., Wang, Z. X., Minobe, Y., Izawa, T., and Yano, M. (2008). Ehd2, a rice ortholog of the maize INDETERMINATE1 gene, promotes flowering by up-regulating Ehd1. *Plant Physiol.* 148 (3), 1425–1435. doi: 10.1104/pp.108.125542
- Monfreda, C., Ramankutty, N., and Foley, J. A. (2008). Farming the planet: 2. Geographic distribution of crop areas, yields, physiological types, and net

primary production in the year 2000. Global Biogeochem. Cycles 22, GB1022. doi: 10.1029/2007GB002947

- Nemoto, Y., Nonoue, Y., Yano, M., and Izawa, T. (2016). Hd1,a CONSTANS ortholog in rice, functions as an Ehd1 repressor through interaction with monocot-specific CCT-domain protein Ghd7. *Plant J.* 86 (3), 221–233. doi: 10.1111/tpi.13168
- Ryu, C. H., Lee, S., Cho, L. H., Kim, S. L., Lee, Y. S., Choi, S. C., et al. (2009). OsMADS50 and OsMADS56 function antagonistically in regulating long day (LD)-dependent flowering in rice. *Plant Cell Environ.* 32 (10), 1412–1427. doi: 10.1111/j.1365-3040.2009.02008.x
- Shang, H. X., YE, S. H., Deng, X. M., Zhou, Y., Xiu, F. L., Ji, X. J., et al. (2012). Identification and a fine Mapping of heading date related mutant gene in Rice. *Rice Sci.* 19 (4), 269–276. doi: 10.1016/S1672-6308(12)60051-1
- Takahashi, Y., Shomura, A., Sasaki, T., and Yano, M. (2001). Hd6, a rice quantitative trait locus involved in photoperiod sensitivity, encodes the  $\alpha$  subunit of protein kinase CK2. *Proc. Natl. Acad. Sci. U.S.A.* 98 (14), 7922–7927. doi: 10.1073/pnas.111136798
- Takahashi, Y., Teshima, K. M., Yokoi, S., Innan, H., and Shimamoto, K. (2009). Variations in Hd1 proteins, Hd3a promoters, and Ehd1 expression levels contribute to diversity of flowering time in cultivated rice. *Proc. Natl. Acad. Sci.* U.S.A. 106, 4555–4560. doi: 10.1073/pnas.0812092106
- Wei, X. J., Xu, J. F., Guo, H. N., Jiang, L., Chen, S. H., Yu, C. Y., et al. (2010). DTH8 suppresses flowering in rice, influencing plant height and yield potential simultaneously. *Plant Physiol.* 153 (4), 1747–1758. doi: 10.1104/pp.110.156943
- Wei, X. J., Xu, J. F., Jiang, L., Wang, H. J., Zhou, Z. L., Zhai, H. Q., et al. (2012). Genetic Analysis for the Diversity of Heading Date of Cultivated Rice in China. *Acta Agronomica Sin.* 38 (1), 10–22. doi: 10.3724/SPJ.1006.2012.00010
- Wei, F. J., Tsai, Y. C., Wu, H. P., Huang, L. T., Chen, Y. C., Chen, Y. F., et al. (2016). Both Hd1 and Ehd1 are important for artificial selection of flowering time in cultivated rice. *Plant Sci.* 242, 187–194. doi: 10.1016/j.plantsci.2015.09.005
- Wu, W. X., Zheng, X. M., Lu, G. W., Zhong, Z. Z., Gao, C. H., Li, P., et al. (2012). Association of functional nucleotide polymorphisms at DTH2 with the

northward expansion of rice cultivation in Asia. *Proc. Natl. Acad. Sci. U.S.A.* 110 (8), 2775–2780. doi: 10.1073/pnas.1213962110

- Xue, W. Y., Xing, Y. Z., Weng, X. Y., Yu, Z., Tang, W. J., Wang, L., et al. (2008). Natural variation in Ghd7 is an important regulator of heading date and yield potential in rice. *Nat. Genet.* 40 (6), 761–767. doi: 10.1038/ng.143
- Yano, M., Katayose, Y., Ashikari, M., Yamanouchi, U., Monna, L., Fuse, T., et al. (2000). Hd1, a major photoperiod sensitivity quantitative trait locus in rice, is closely related to the Arabidopsis flowering time gene CONSTANS. *Plant Cell* 12 (12), 2473–2483. doi: 10.1105/tpc.12.12.2473
- Zheng, X. M., and Ge, S. (2010). Ecological divergence in the presence of gene flow in two closely related Oryza species (Oryza rufipogon and O. nivara). *Mol. Ecol.* 19 (12), 2439–2454. doi: 10.1111/j.1365-294X.2010.04674.x
- Zheng, X. M., Li, F., Wang, J. R., Qiao, W. H., Zhang, L. F., Cheng, Y. L., et al. (2016). Nonfunctional alleles of long-day suppressor genes independently regulate flowering time. *J. Integr. Plant Biol.* 58 (6), 540–548. doi: 10.1111/ jipb.12383
- Zhu, Q., Zheng, X., Luo, J., Gaut, B. S., and Ge, S. (2007). Multilocus Analysis of Nucleotide Variation of Oryza sativa and Its Wild Relatives: Severe Bottleneck during Domestication of Rice. *Mol. Biol. Evol.* 24 (3), 875–888. doi: 10.1093/ molbev/msm005

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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