



# Implications of Zoonoses From Hunting and Use of Wildlife in North American Arctic and Boreal Biomes: Pandemic Potential, Monitoring, and Mitigation

Lucy O. Keatts<sup>1\*</sup>, Martin Robards<sup>2</sup>, Sarah H. Olson<sup>1</sup>, Karsten Hueffer<sup>3</sup>, Stephen J. Insley<sup>4,5</sup>, Damien O. Joly<sup>6</sup>, Susan Kutz<sup>7</sup>, David S. Lee<sup>8</sup>, Cheryl-Lesley B. Chetkiewicz<sup>4</sup>, Stéphane Lair<sup>9</sup>, Nicholas D. Preston<sup>10</sup>, Mathieu Pruvot<sup>1,7</sup>, Justina C. Ray<sup>4</sup>, Donald Reid<sup>4</sup>, Jonathan M. Sleeman<sup>11</sup>, Raphaela Stimmelmayr<sup>12,13</sup>, Craig Stephen<sup>14,15</sup> and Chris Walzer<sup>1,16</sup>

#### **OPEN ACCESS**

#### Edited by:

Pierre Echaubard, SOAS University of London, United Kingdom

#### Reviewed by:

Simon Rodrigo Rüegg, University of Zurich, Switzerland Alison J. Peel, Griffith University, Australia

#### \*Correspondence:

Lucy O. Keatts lkeatts@wcs.org

#### Specialty section:

This article was submitted to Planetary Health, a section of the journal Frontiers in Public Health

Received: 09 November 2020 Accepted: 30 March 2021 Published: 05 May 2021

#### Citation:

Keatts LO, Robards M, Olson SH, Hueffer K, Insley SJ, Joly DO, Kutz S, Lee DS, Chetkiewicz C-L, Lair S, Preston ND, Pruvot M, Ray JC, Reid D, Sleeman JM, Stimmelmayr R, Stephen C and Walzer C (2021) Implications of Zoonoses From Hunting and Use of Wildlife in North American Arctic and Boreal Biomes: Pandemic Potential, Monitoring, and Mitigation. Front. Public Health 9:627654. doi: 10.3389/fpubh.2021.627654 <sup>1</sup> Wildlife Conservation Society Health Program, Bronx, NY, United States, <sup>2</sup> Wildlife Conservation Society, Arctic Beringia Program, Fairbanks, AK, United States, <sup>3</sup> Department of Veterinary Medicine & Arctic and Northern Studies Program, University of Alaska Fairbanks, Fairbanks, AK, United States, <sup>4</sup> Wildlife Conservation Society Canada, Toronto, ON, Canada, <sup>5</sup> Department of Biology, University of Victoria, Victoria, BC, Canada, <sup>6</sup> Nyati Health Consulting, Nanaimo, BC, Canada, <sup>7</sup> Department of Ecosystem and Public Health, Faculty of Veterinary Medicine, University of Calgary, Calgary, AB, Canada, <sup>8</sup> Department of Wildlife and Environment, Nunavut Tunngavik Inc., Ottawa, ON, Canada, <sup>9</sup> Canadian Wildlife Health Cooperative, Université de Montréal, Montreal, QC, Canada, <sup>10</sup> Salmon Coast Field Station, Echo Bay, BC, Canada, <sup>11</sup> United States Geological Survey National Wildlife Health Center, Madison, WI, United States, <sup>12</sup> North Slope Department of Wildlife Management, Utqiagvik, AK, United States, <sup>13</sup> Institute of Arctic Biology, University of Alaska Fairbanks, Fairbanks, AK, United States, <sup>14</sup> University of British Columbia, Vancouver, BC, Canada, <sup>15</sup> Ross University School of Veterinary Medicine, Basseterre, Saint Kitts and Nevis, <sup>16</sup> Conservation Medicine Unit, Department of Interdisciplinary Life Sciences, Research Institute of Wildlife Ecology, University of Veterinary Medicine, Vienna, Austria

The COVID-19 pandemic has re-focused attention on mechanisms that lead to zoonotic disease spillover and spread. Commercial wildlife trade, and associated markets, are recognized mechanisms for zoonotic disease emergence, resulting in a growing global conversation around reducing human disease risks from spillover associated with hunting, trade, and consumption of wild animals. These discussions are especially relevant to people who rely on harvesting wildlife to meet nutritional, and cultural needs, including those in Arctic and boreal regions. Global policies around wildlife use and trade can impact food sovereignty and security, especially of Indigenous Peoples. We reviewed known zoonotic pathogens and current risks of transmission from wildlife (including fish) to humans in North American Arctic and boreal biomes, and evaluated the epidemic and pandemic potential of these zoonoses. We discuss future concerns, and consider monitoring and mitigation measures in these changing socio-ecological systems. While multiple zoonotic pathogens circulate in these systems, risks to humans are mostly limited to individual illness or local community outbreaks. These regions are relatively remote, subject to very cold temperatures, have relatively low wildlife, domestic animal, and pathogen diversity, and in many cases low density, including of humans. Hence, favorable conditions for emergence of novel diseases or major amplification of a spillover event are currently not present. The greatest risk to northern communities

1

from pathogens of pandemic potential is via introduction with humans visiting from other areas. However, Arctic and boreal ecosystems are undergoing rapid changes through climate warming, habitat encroachment, and development; all of which can change host and pathogen relationships, thereby affecting the probability of the emergence of new (and re-emergence of old) zoonoses. Indigenous leadership and engagement in disease monitoring, prevention and response, is vital from the outset, and would increase the success of such efforts, as well as ensure the protection of Indigenous rights as outlined in the United Nations Declaration on the Rights of Indigenous Peoples. Partnering with northern communities and including Indigenous Knowledge Systems would improve the timeliness, and likelihood, of detecting emerging zoonotic risks, and contextualize risk assessments to the unique human-wildlife relationships present in northern biomes.

Keywords: wildlife, hunting, zoonotic, pandemic, Arctic, boreal, Indigenous, One Health

# INTRODUCTION

Emerging infectious diseases (EIDs) are a significant burden on public health and economies, and are increasingly recognized as a global threat (1). EIDs are currently defined by the World Health Organization (WHO) as those that "have newly appeared in a population or have existed but are rapidly increasing in incidence or geographic range (2)." This definition does not clarify between different categories of emergence or reemergence, nor does it clearly differentiate novel diseases with pandemic potential, such as COVID-19, from those that are variants of old pathogens, new detections of old pathogens with novel technologies, or re-emergence of old pathogens in new regions (2). Thus, the definition does not reflect the very different drivers and significance between diseases and pathogens in terms of global vs. local burden, threat and origin (2). The majority of EIDs (over 60%) are considered zoonotic, with zoonoses defined as "any infection that is naturally transmissible from vertebrate animals to humans" (3-7). There is, however, a need to better differentiate between diseases that originate in animals but are subsequently independently perpetuated in human populations, and those that require an animal host for pathogen persistence, to target research, control, policy and mitigation efforts (2). Infection of people with zoonotic pathogens occurs through contact with infected animals via a variety of mechanisms including: direct contact with bodily fluids (e.g., saliva, blood, urine, feces); indirect contact with surfaces contaminated with an animal's infectious secretions; vector-borne through biting arthropods; foodborne through consumption of contaminated raw or undercooked food; and waterborne, via contaminated drinking water (8). The definition of "risk" usually considers two dimensions: how likely the uncertainty is to occur (probability), and what the effect would be if it happened (impact) (9). Both components are important when establishing the local vs. global risks of zoonoses and emerging diseases of animal-origin, and in considering policy or other interventions.

Severe Acute Respiratory Syndrome (SARS)-CoV-2, considered a novel zoonotic coronavirus, and the causative agent for COVID-19, emerged in December of 2019 in Wuhan, China and rapidly spread globally, to devastating effect. Comparative

genomic analysis indicates that SARS-CoV-2 evolved naturally, with bats the likely ancestral reservoir host (10). Given that the animal reservoir for SARS-CoV-2 is yet to be identified, others propose the virus be classified an "emerging infectious disease (EID) of probable animal origin" rather than a zoonosis (2). Many, but not all, early cases of COVID-19 were associated with a market in Wuhan that traded in wildlife (11), and plausible scenarios have been put forth to explain the origin of SARS-CoV-2: evolution in bats or an intermediate animal host before zoonotic spillover to humans in the market or market trade chain (12, 13); or natural evolution in humans following direct zoonotic transfer from bats (12), though purifying selection of SARS-CoV-2 in humans since the start of the pandemic has been weak compared with the significant positive diversifying selection that has occurred in bats since SARS-CoV-2 evolved from its closest known relative RmYN02 (13). SARS-CoV-2 shares some genetic similarity with SARS-CoV, and prevailing evidence suggests that SARS-CoV spilled over into humans via an intermediate host-likely masked palm civets (Paguma larvata)-at a wildlife market in southern China in 2002 (14-17). The most devastating pandemics in human history, the Black Death (in the 1300s), Spanish influenza (1918), and HIV/AIDS, all resulted from an initial zoonotic spillover from wildlife (18). Of currently defined emerging zoonoses, over 70% originate in wildlife (6, 19-21) however, very few diseases of wild animal origin persist with ongoing zoonotic transmission, with most human infections from such diseases being acquired through human-human transmission. The frequency of zoonotic disease spillover into humans is increasing (1, 19, 22, 23). This rise has been linked to changing human ecology, due to the growing global human population, and its demand for food, land, and natural resources (5, 19, 20, 24). Escalating anthropogenic activities are increasing contact rates between humans, domestic animals, and wildlife (19, 25-28).

Wild animal consumption-based food systems have been implicated in the emergence of diseases with zoonotic origins (including HIV, SARS, Ebola virus disease, Avian Influenza A), and mounting evidence indicates substantial human health risks from the trade in live wildlife (29–40). Members of the WHO team investigating the origins of the SARS-CoV-2

pandemic recently reported that wildlife farms were the most likely source (41). The trade of wild-harvested meat for food fills a continuum from subsistence-based rural consumption, to extensive commercial trade networks to meet growing urban and international demand for wild animal meat as a luxury product (42). There are calls from scientific, health, conservation organizations, and government officials to end commercial trade in wildlife for human consumption. Closing wild animal markets and the trade in wildlife, is viewed as an expedient measure to reduce the risk of future viral outbreaks, like the COVID-19 pandemic, that threaten human health, well-being, economies, and security at local, regional, and global scales (43, 44). Public health organizations, including the WHO, support the rigorous enforcement of bans on the sale and trade of wildlife for food (45). However, some populations are dependent on wild-harvested food to meet basic nutritional requirements. In the face of disruptions to food supply chains during the COVID-19 pandemic, harvesting and sharing of local foods has helped maintain food and nutrition security for isolated boreal and Arctic communities (46), and harvesting of wildlife, with secondary use of wildlife by-products (i.e., pelts, claws, skulls), forms an important part of traditional economies and socioeconomic-cultural well-being of Indigenous Peoples. The World Organization for Animal Health (OIE) considers wildlife all of: (a) wild animals (b) free-ranging feral [domesticated] animals, and (c) non-domestic animals in captivity or farming, though these different categories and interfaces likely pose quite different risks for spillover to humans (2). Infection cycles in densely populated wildlife farms, along trade chains and live markets, are very different to those in natural, free-ranging populations. These crowded and stressful environments are much more likely to facilitate cross-species transmission of pathogens with pandemic potential (2, 32, 47). Based on past experiences, wellmeaning initiatives aimed at halting the hunting, trapping, and use of wildlife risk violating the rights of Indigenous Peoples as outlined in the United Nations (UN) Declaration on the Rights of Indigenous Peoples (48), and also threaten the subsistence of non-Indigenous hunter communities.

Indigenous Peoples and local communities across the boreal and Arctic regions (or "northern communities") have extensive and essential relationships with their environments, including through the harvest and sharing of nutritionally and spiritually important native plants, and wildlife, also known as traditional and country foods (49-53). Limited access to traditional and country foods is a strong predictor of health disparities in Indigenous Peoples across the North, and is correlated to diseases such as diabetes, cardiovascular disease and mental illness (54). Throughout boreal and Arctic ecosystems, subsistence economies support rich and diverse cultures that include the use, sharing, and consumption of wildlife. Subsistence is considered as: the personal consumption of wildlife for food, fuel, shelter, clothing, tools, or transportation; the barter, trade, or sharing of wildlife products in their harvested form with relatives, with others in the local community or with persons in locations other than the local community with whom local residents share familial, social, cultural, or economic ties; and the making and selling of handicrafts from wildlife products, when the wildlife are harvested for the purposes defined above (55). Subsistence, together with guided hunting and fishing, by Indigenous Peoples in the Arctic and boreal is best considered through the lens of food security and sovereignty, cultural security, and livelihoods (56, 57). Food security is considered to exist when "all people, at all times, have physical, social, and economic access to sufficient, safe, and nutritious food to meet their dietary needs, and food preferences, for a healthy and active life" (58). Food sovereignty refers to the "ability and right of people to define their own policies and strategies for sustainable production, distribution, and consumption of food that guarantees the right to food for the entire population" (59). The rights of Indigenous Peoples to determine and maintain these relationships with the environment are embedded in the UNDRIP (48), in the Canadian Constitution (60), and in the Truth and Reconciliation Commission of Canada's Calls to Action (61), but have received less formalized protections within the United States. The relationships between Indigenous Peoples in North America and the environment, particularly the harvest and use of wildlife, are the basis for historic numbered Treaties, and modern land claim agreements in Canada and the United States, Native to Native agreements, and Indigenous advisory and co-management institutions (see Table 1 for specific examples). Such agreements however, have not been as effective in addressing the needs of Indigenous Peoples and local communities as they are forced to adapt to the impacts of biodiversity loss and climate change within historically colonial approaches to environmental management (62). The use of wildlife by non-Indigenous local communities are largely managed through legislation and regulations developed, and enforced, by governments and regional authorities.

Indigenous Peoples in the Arctic and boreal are often at the forefront of protecting wild food systems, livelihoods, and cultural values (63, 64), and concerns have been raised about potential impacts to Indigenous rights and food sovereignty due to policy initiatives focused on hunting, consumption, sharing, and local trade of wildlife (55, 65-70). Negative impacts have occurred when socio-economic, cultural, and nutritional dimensions of Indigenous subsistence practices were not considered. For example, previous culturally insensitive and poorly developed communication outreach efforts regarding health and traditional and country foods (on contaminants) resulted in negative health consequences for affected populations from avoidance of traditional foods altogether, given limited healthy alternatives (71, 72). Zoonotic health risks and concerns are also relevant to local communities of hunters, trappers, and fishers across the region. This paper was written to respond to some of the issues related to northern food sovereignty, and to apprehension expressed by researchers and veterinarians working with these northern communities around potential routes for introduction of emerging zoonoses, such as COVID-19, into remote Indigenous and local communities where subsistence based on traditional and mixed economies remains a vital necessity. The manuscript aims to (i) review zoonotic pathogens of wildlife origin in Arctic and boreal systems, in the context of wildlife use by northern communities; (ii) provide a reference for northern communities, and wildlife 

 TABLE 1 | Examples of formal agreements and co-management institutions protecting the rights of Indigenous Peoples in Arctic and boreal biomes across North

 America regarding land, hunting, and use of wildlife.

| Type of agreement   | Specific examples of agreements  |
|---|--|
| Agreements with federal government of<br>Canada                                 | Inuvialuit final agreement; Nunavut land claims agreement; James Bay and Northern Québec agreement and the Northeastern Québec agreement; Nunavik inuit land claims agreement; Eeyou marine region land claims agreement; Labrador Inuit land claims agreement; Umbrella final agreement (Yukon first nations), Historic treaties and agreements <sup>a</sup> across the boreal, including Treaty No. 5 (Manitoba), Treaty no. 8 (Alberta, Northwest Territories, British Columbia), James Bay treaty–treaty no. 9 and adhesions made in 1929 and 1930 (Ontario), Treaty no. 10 (Saskatchewan)   |
| Agreement with national government of the United States                         | Alaska native claims settlement act  |
| Indigenous Nation to Nation agreements  | Inuvialuit-Inupiat polar bear management agreement (IIA); Alaska and inuvialuit beluga whale committee (AIBWC)   |
| Indigenous advisory and wildlife<br>co-management institutions (not exhaustive) | Alaska: alaska eskimo whaling commission (AEWC); Alaska beluga whale committee (ABWC); Iceseal committee (ICS);<br>Alaska nannut co-management council (ANCC); Eskimo walrus commission (EWC); Indigenous people's council for<br>marine mammals (IPCoMM); Association of traditional marine mammal hunters of chukotka (ATMMHC); Wildlife<br>management advisory council; kitikmeot regional wildlife board<br>Canada: Nunavut wildlife management board, Inuvialuit game council, Fisheries joint management committee, Hunting<br>fishing trapping coordinating committee, Nunavik marine region wildlife board, Eeyou marine region wildlife board,<br>Torngat wildlife & plants co-management board, Sahtu renewable resources board, Gwich'in renewable resources<br>board, Wek'eezhii renewable resources board, wildlife management advisory council (NT, NS), Yukon fish and wildlife |

<sup>a</sup>https://www.rcaanc-cirnac.gc.ca/eng/1100100028574/1529354437231.

disease and public health researchers, to consider and compare, the potential community and broader health implications of zoonoses transmissible via traditional use of wildlife; (iii) examine zoonotic pathogen spillover, amplification, epidemic or pandemic spread, and relevant dynamics in northern biomes; (iv) describe some future zoonotic concerns; and (v) discuss considerations for current and future monitoring, surveillance and risk reduction approaches. The results section, with **Table 2**, addresses aims 1 and 2, and the remaining aims are addressed in the discussion.

## MATERIALS AND METHODS

We carried out a qualitative literature review to identify publications focused on zoonotic diseases in Arctic and boreal biomes with a potential wildlife origin. The review focused on zoonoses associated with the hunting, trapping, butchering, sharing, use, and trade of wildlife (including fish), and only considered agricultural or domestic species if related to a sylvatic cycle. Given this focus, vector-borne zoonoses were not included in the review, but are considered in the discussion. In addition, we asked for the contributions of experts on the topics. Only published materials were included. Our search included articles, reviews, proceedings papers, reports and book chapters in the English language. The Web of Science database and Google Scholar search engine were utilized to conduct the search in late April, 2020. Publications from any year were considered.

To gain an overview of the literature pertaining to existing zoonotic disease reports for communities in Arctic and boreal regions associated with consumption and/or hunting of wildlife, search terms were conducted using the following keywords and phrases:

"zoonoses" OR "zoonotic"

- AND "wildlife" OR "fish" OR "hunter" OR "hunting" OR "hunted" OR "subsistence" OR "country food" OR "traditional food" OR "Indigenous"
- AND "Arctic" OR "boreal" OR "Alaska" OR "northern Canada"

Based on the titles and abstracts of the identified studies, we excluded those publications considered irrelevant to part one of our review, including those for which the full manuscript was not available, publications pertaining to vector-borne zoonoses in Northern regions; those describing zoonoses in other regions, or for which hunting, consumption, use or sharing of wildlife have not been reported to facilitate transmission; and those referring to experimental rather than natural infections. Relevance was attributed if the publications discussed case studies of zoonotic diseases transmitted to humans via the hunting, consumption, preparation or sharing of wildlife in northern regions of North America; or if they described the ecology and epidemiology of zoonoses with one or more wild hosts and potential transmission to humans through the hunting, consumption, preparation or sharing of wildlife in North American Arctic and boreal biomes; details on these relevant aspects were recorded in a table.

Ninety five publications were identified for inclusion in the review. The lead author read each full article and extracted relevant information for inclusion in the manuscript. The other authors reviewed this selection and recommended additional papers, where available, that met inclusion criteria described above. All authors then considered together how the review findings combine with risk factors for zoonotic disease emergence, amplification and pandemic spread, and key cultural, socio-economic, political and ecological factors in Arctic and boreal regions, to inform on the risks of zoonotic disease emergence and amplification from Northern wildlife use and trade, and how such risks are being and could be better addressed. TABLE 2 Zoonotic pathogens of Arctic and boreal systems that can be transmitted to humans through hunting, consumption or other use of wildlife, noting pathogen potential for local outbreak clusters, human-to-human transmission, and epidemic/pandemic spread.

| Type of<br>pathogen | Disease/Pathogen   | Wild Host(s)* in arctic<br>and boreal biomes   | Route of infection to humans  | Disease in humans   | Local outbreak clusters possible?                                 | Human to human transmission? | Epidemic or<br>pandemic<br>potential? |
|---------------------|--|--|---|---|---|------------------------------|---------------------------------------|
| Parasites           | Anisakidosis/<br>Roundworms of genus<br><i>Anisakis</i> (73, 74)   | Definitive host: Bearded<br>seals, ringed seals and<br>beluga whale<br>Intermediate: Fish and squid  | Consumption of raw fish   | Gastritis with ulcerative lesions of stomach wall   | Possible through<br>consumption of shared<br>contaminated product | No                           | No                                    |
|                     | Fluke infection<br><i>Cryptocotyle lingua</i><br>(fish trematode) (75, 76)   | Intermediate host: Fish<br>Reservoir hosts: Fish-eating<br>birds and mammals<br>including foxes, gulls, terns,<br>and herons   | Ingestion of raw or improperly<br>cooked fish from fresh and<br>brackish water  | Liver and intestinal damage   | Possible through<br>consumption of shared<br>contaminated product | No                           | No                                    |
|                     | Cystic Echinococcosis/<br>Echinococcus<br>canadensis (22, 77–83)   | Definitive host: Wolf, coyote<br>(dog)<br>Intermediate host: Mainly<br>caribou/reindeer and moose<br>(also muskox, elk, bison,<br>and white-tailed and<br>mule deer)   | Canids: Ingestion of viscera of<br>infected intermediate host.<br>Humans: Via accidental ingestion<br>of eggs shed in canid feces (e.g.,<br>from fur during fox skinning), or<br>from a water or food source<br>(e.g., plants, berries)<br>contaminated with eggs | Relatively benign cyst formation in liver<br>and lung   | Possible through contaminated water source                        | No                           | No                                    |
|                     | Alveolar<br>Echinococcosis/<br><i>Echinococcus</i><br><i>multilocularis</i><br>(79, 84, 85)                              | Definitive host: Fox, felid,<br>wolf, and coyote (also dog)<br>Intermediate host: rodents<br>e.g., vole, deer mice,<br>lemming, muskrat (ground<br>squirrels and shrews on St.<br>Lawrence Island)                     | <i>Canids:</i> Ingestion of viscera of infected intermediate host. <i>Humans:</i> Accidental ingestion of eggs shed in canid or felid feces (e.g., from fur during fox skinning); or from contaminated water or food source (e.g., plants, berries)               | Alveolar hydatid disease with parasitic<br>tumor growth in liver, lungs, brain, and<br>other organs and much higher mortality<br>than for <i>Echinococcus canadensis</i><br>infection | Possible through contaminated water source                        | No                           | No                                    |
|                     | Tapeworms/<br>Diphyllobothrium latum<br>& Diphyllobothrium<br>dendriticum &<br>Diphyllobothrium<br>nihonkaiense (73, 86) | Diphyllobothrium latum:<br>Fish-eating mammals (e.g.,<br>bear, wolf, otters, and mink)<br>Diphyllobothrium<br>dendriticum: Fish,<br>fish-eating mammals, and<br>birds<br>Diphyllobothrium<br>nihonkaiense: Wild salmon | Consumption of undercooked fish meat or livers  | Asymptomatic or causes mild chronic intermittent diarrhea   | Possible through<br>consumption of shared<br>contaminated product | No                           | No                                    |
|                     | Toxocariasis/ <i>Toxocara</i><br><i>canis</i> (primarily) &<br><i>Toxocara cati</i> **<br>(79, 82, 83, 87–90)            | <i>Toxocara canis:</i> Wolves,<br>coyotes, and foxes (small<br>mammals), (dog)<br><i>Toxocara cati</i> : Felids (e.g.,<br>lynx) and rodents  | Mainly through accidental<br>ingestion of eggs from<br>contaminated; possible via<br>consumption of uncooked meat<br>of small mammal paratenic hosts  | Ocular and visceral larval migrans  | Possible through contaminated water source                        | No                           | No                                    |

(Continued)

#### TABLE 2 | Continued

| Type of<br>pathogen | Disease/Pathogen  | Wild Host(s)* in arctic<br>and boreal biomes   | Route of infection to humans  | Disease in humans   | Local outbreak clusters possible?   | Human to human transmission?   | Epidemic o<br>pandemic<br>potential? |
|---------------------|---|--|---|---|---|--|--------------------------------------|
|                     | Toxoplasmosis/<br><i>Toxoplasma gondii</i><br>(79, 84, 91–101)                                      | Definitive host: Felids (e.g.,<br>lynx)<br>Intermediate hosts: many<br>northern animals including<br>caribou, walrus, birds<br>and seal  | Consumption of raw or<br>undercooked meat; or via water<br>or soil contaminated with felid<br>feces containing infective<br>oocysts   | Often asymptomatic, possible association<br>with mental health issues e.g., depression;<br>severe disease in immune- compromised<br>individuals (e.g., encephalitis/<br>chorioretinitis); fetal morbidity and mortality<br>during pregnancy   | Possible through<br>consumption of shared<br>contaminated product   | No   | No                                   |
|                     | Trichinellosis/ <i>Trichinella native</i> (30, 73, 79, 82, 84, 102–106)                             | Walrus, seal, bear (polar,<br>black and grizzly), fox, wolf,<br>and wolverine  | Ingestion of raw or undercooked meat  | From asymptomatic to nausea, diarrhea,<br>vomiting, abdominal pain, muscle pain,<br>fever swelling of eyes, weakness/fatigue,<br>headache, and (rarely) fatality if heart<br>affected   | Possible through<br>consumption of shared<br>contaminated product   | No   | No                                   |
|                     | Giardiasis ( <i>Giardia</i> spp.)<br>& Cryptosporidiosis<br>( <i>Cryptosporidium</i> spp.)<br>(107) | Mammals including beaver,<br>muskrats, muskoxen, and<br>others   | Via water contaminated with<br>feces containing infective<br>oocysts  | Diarrheal disease   | Possible through contaminated water source  | No   | No                                   |
| Bacteria            | Anthrax/ <i>Bacillus</i><br><i>anthracis</i> (108–111)  | Wild ungulates (e.g.,<br>white-tailed and mule deer,<br>bison, moose, and reindeer)  | Ingestion or inhalation, or<br>contamination of wounds by,<br>bacterial spores  | Cutaneous: skin sores<br>Inhalational: chest pain, shortness of<br>breath, cough, nausea, vomiting, stomach<br>pains, headache, sweats, fatigue, body<br>aches<br>Gastrointestinal: Fever, swelling of neck<br>glands, sore throat, nausea, vomiting and<br>diarrhea, headache, fainting, swelling of<br>abdomen.<br>All types have potential, if untreated, to<br>spread throughout body, causing severe<br>illness and even death | Possible through shared<br>contaminated water or food<br>source. Rarely direct<br>human-to- human<br>transmission | Very rare reports<br>from cutaneous<br>form. Not<br>considered<br>contagious | No                                   |
|                     | Brucellosis/ <i>Brucella</i><br>spp. (81, 84, 112, 113)   | Wild mammals including:<br>caribou/reindeer, elk,<br>muskoxen, bison,<br>white-tailed and mule deer,<br>goats, sheep, moose, wolf,<br>fox, rodents, hares, mink,<br>and marine mammals | Handling of carcasses, fetuses,<br>and newborn calves from<br>infective animals; or<br>consumption of raw (including<br>frozen or dried) meat and<br>marrow<br>Dogs: Consumption of<br>uncooked infected tissue | Systemic bacterial disease (acute or<br>insidious): intermittent fever with<br>headache, weakness, sweating, chills,<br>joint pain and weight loss; also cerebral<br>forms; can be fatal  | Possible, through<br>consumption of shared<br>contaminated product  | Extremely rare<br>(e.g., through<br>breastmilk)                              | No                                   |
|                     | Botulism/ <i>Clostridium</i><br><i>botulinum</i> *** (114–119)                                      | Fish (especially salmon) and<br>many mammals<br>Main source in north is<br>marine mammals, especially<br>seals and whales  | Consumption of raw or parboiled<br>seal meat, fish, seal oil, or other<br>wild meat that has undergone<br>faulty fermentation or aging  | Multiple clinical symptoms including:<br>blurred vision, nausea, vomiting, paralysis<br>of the motor nerves, and respiratory<br>paralysis in fatal cases  | Possible, through<br>consumption of shared<br>contaminated product  | No   | No                                   |

(Continued)

Zoonoses and Northern Wildlife Use

#### TABLE 2 | Continued

| Type of<br>pathogen | Disease/Pathogen   | Wild Host(s)* in arctic<br>and boreal biomes   | Route of infection to humans   | Disease in humans  | Local outbreak clusters possible?  | Human to human<br>transmission?   | Epidemic or<br>pandemic<br>potential? |
|---------------------|--|--|--|--|--|---|---------------------------------------|
|                     | Erysipelas/ <i>Erisipelothrix</i><br><i>rhusiopathiae</i><br>(56, 120–128)   | Terrestrial and aquatic<br>mammals including<br>muskoxen, white-tailed and<br>mule deer, caribou, birds,<br>fish, and arthropods                                 | Exposure to infected animals or<br>fish or animal products via skin<br>wounds or via ingestion;<br>environmental sources of<br>infection also reported   | Localized skin infections; or severe cases<br>with diffuse cutaneous or systemic<br>disease, septicemia, endocarditis;<br>infrequently pneumonia, abscesses,<br>meningitis, arthritis  | Unlikely   | No  | No                                    |
|                     | Leptospirosis/<br><i>Leptospira interrogans</i><br>(129–131)   | Beavers, coyotes,<br>white-tailed and mule deer,<br>foxes, opossums, otters,<br>raccoons, skunks, and<br>Northern fur seals                                      | Direct contact with contaminated urine or animals  | From no symptoms, to kidney damage,<br>meningitis, liver failure, respiratory distress,<br>and death   | Possible through contaminated water source   | No  | No                                    |
|                     | Lyme disease/ <i>Borrelia</i><br><i>burgdorferi</i> (132, 133)   | Seabirds, song birds, and wild ungulates   | Bites from ticks that have fed on an infected animal   | Fever, rash, facial paralysis, arthritis   | Possible if infected host and tick densities high  | No  | No                                    |
|                     | Pasteurellosis/<br>Pasteurella multocida;<br>Bisgaardia<br>hudsonensis (others)<br>(134–137)                                 | Pinnipeds, including seals<br>and walruses, many<br>terrestrial mammals, birds,<br>and reptiles  | Animal bites or contact with<br>nasal secretions of infected<br>animal   | Skin and soft tissue infections: rapidly<br>spreading edema, erythema and<br>tenderness at site of the bite or scratch;<br>abscessation; enlarged local lymph nodes  | No   | No  | No                                    |
|                     | Q fever/ <i>Coxiella burnetii</i><br>(138–140)   | Northern fur seals and sea<br>birds  | Inhalation of dust contaminated<br>by infected animal feces, urine,<br>milk, or birth products;<br>contaminated water source; or<br>ingestion of infected animal<br>products e.g., milk or cheese                                      | Mild: Fever, fatigue, headache, muscle<br>aches, vomiting, diarrhea, chest or<br>stomach pain, weight loss, cough<br>Severe: pneumonia or hepatitis. Infection<br>during pregnancy can cause miscarriage,<br>stillbirth, pre-term delivery, low infant<br>birth weight | Occasionally, through<br>shared contaminated water<br>or food source, or<br>human-to-human<br>transmission                         | Rare: although<br>highly<br>transmissible from<br>animal-human it is<br>not highly<br>transmissible from<br>human-human | No                                    |
|                     | Seal finger/<br><i>Mycoplasma</i> spp.<br>(114, 134, 141)  | Seals and whales   | Marine mammal bites; or broken<br>skin contact with infectious<br>material from marine mammal  | Swollen, painful, and suppurative lesion on finger; rarely systemic, with fever and lymphangitis   | No   | No  | No                                    |
|                     | Tuberculosis and<br>Mycobacteriosis/<br>Mycobacterium bovis,<br>M. tuberculosis, M.<br>pinnipedii & others<br>(114, 142–144) | Marine mammals: Wild<br>seals (more commonly) and<br>cetaceans (rarely)<br>Terrestrial mammals: bison,<br>elk, moose, white-tailed<br>deer, mule deer and wolves | Multiple routes: inhalation,<br>ingestion of raw/undercooked<br>meat or unpasteurized milk<br>products, and direct contact with<br>breaks in the skin e.g., when<br>dressing infected ungulates  | Pulmonary (cough, shortness of breath)<br>and cutaneous (localized skin infections)<br>disease   | Possible, through<br>consumption of shared<br>contaminated product and<br>direct human- to-human<br>transmission                   | Yes   | Yes                                   |
|                     | Tularemia/ <i>Francisella<br/>tularensis</i><br>(84, 145–148)  | Muskrats, beavers, hares,<br>voles, squirrels, wolves,<br>bears, and other northern<br>wildlife  | Consumption of insufficiently<br>cooked meat or contaminated<br>water and dust; or through bites<br>from infected vectors such as<br>mosquitoes and ticks; and<br>through direct contact i.e.,<br>skinning; touching hare<br>carcasses | Skin lesions or ulcerations,<br>lymphadenomegaly, vomiting, diarrhea,<br>abdominal pain, conjunctivitis, pneumonia,<br>septicemia, and hepatosplenomegaly  | Rare, but possible through<br>consumption of shared<br>contaminated product or<br>contaminated water source<br>or vector abundance | No  | No                                    |

(Continued)

Zoonoses and Northern Wildlife Use

| Type of<br>pathogen | Disease/Pathogen   | Wild Host(s)* in arctic<br>and boreal biomes  | Route of infection to humans   | Disease in humans  | Local outbreak clusters possible?  | Human to human transmission?   | Epidemic or<br>pandemic<br>potential?  |
|---------------------|--|---|--|--|--|--|--|
|                     | Yersiniosis****/Yersinia<br>pseudotuberculosis &<br>Y. enterocolitica<br>(94, 142, 149–151)  | Reservoirs in rodents<br>(beaver, muskrat, ground<br>squirrels), lagomorphs<br>(snowshoe hare), and<br>outbreaks in muskoxen  | Ingestion of raw or undercooked<br>meat, or water contaminated<br>with infected fecal matter   | Fever, abdominal pain, and diarrhea  | Possible through<br>consumption of shared<br>contaminated product or<br>contaminated water source                    | Very rarely; not<br>highly<br>transmissible                                |  |
| Viruses             | Avian influenza/<br>Influenza A viruses<br>(114, 152–155)  | Wild birds, especially waterfowl  | During preparation of infected<br>birds for eating (plucking,<br>cleaning, butchering) or<br>consumption of raw meat from<br>infected bird | Mild to severe illness, sometimes death.<br>Fever, chills, cough, sore throat,<br>congestion, body aches, headache,<br>fatigue; vomiting, and diarrhea in children   | Yes  | Yes  | Yes  |
|                     | Caliciviruses/(marine<br>caliciviruses: serotypes<br>of vesicular exanthema<br>of swine virus)<br>(114, 156)                                     | Arctic marine mammals,<br>including fur seals, elephant<br>seals, walrus, and whales<br>(including bowhead and<br>gray)   | Broken skin contact with<br>infectious animal or their<br>secretions   | Fluid-filled blisters on the extremities   | Rare, but possible through handling same animal  | Possible but rare,<br>through broken<br>skin contact with<br>blister fluid | Not for<br>marine<br>serotypes   |
|                     | Sealpox/ <i>Parapox virus</i><br>(114, 157, 158)   | Harbor and gray seals   | Direct contact via pox lesions on<br>infected mammals  | Painful, nodular lesions   | Rare, but possible through handling same animal  | No   | No   |
|                     | Orf/ <i>Parapox virus</i><br>(157–159)   | Muskoxen, mountain goats,<br>Dall's sheep, caribou and<br>white-tailed, and mule deer   | Direct contact via pox lesions on<br>infected mammals  | Painful, nodular lesions   | Rare but possible through<br>handling same animal or if<br>multiple animals infected in<br>herd                      | No   | No   |
|                     | Rabies<br>(38, 81, 160–170)  | Principle reservoir hosts:<br>Arctic foxes, red foxes,<br>wolves, and bats<br>Less commonly: Caribou,<br>beaver, black and polar<br>bears, racoons, lynx,<br>and wolverine) | Humans and domestic dogs: via<br>bites from infected wildlife  | Almost always fatal if untreated. Affects<br>central nervous system: general weakness<br>or discomfort, fever, headache; prickling<br>sensation at site of the bite, anxiety,<br>confusion, agitation, delirium,<br>hallucinations, hydrophobia (fear of water),<br>and insomnia | Possible through rabid<br>animal in community  | Extremely rare:<br>through bite or<br>organ transplant                     | No   |
|                     | Hepatitis E***** N.B.<br>transmission from<br>wildlife to humans not<br>yet confirmed in Arctic<br>or boreal regions, but<br>suspected (171–173) | Free-ranging deer, possibly caribou   | Humans via undercooked meat<br>or food contaminated with feces<br>from infected animal   | Acute viral hepatitis, mortality a concern in pregnant women   | Possible through<br>consumption of shared<br>contaminated undercooked<br>meat or shared<br>contaminated water source | Rare: mainly via<br>maternal-infant<br>transmission                        | Outbreaks<br>possible via<br>fecal<br>contamination<br>of drinking<br>water source |
| Fungal              | None as yet  | Future concern in warming<br>Arctic and boreal systems.<br>Dearth of data for fish (174)  | Future concern in warming Arctic and boreal systems  | N/A  | N/A  | N/A  | No   |

(Continued)

| Disease/Pathogen  | Wild Host(s)* in arctic<br>and boreal biomes  | Route of infection to humans Disease in humans  | Disease in humans   | Local outbreak clusters<br>possible?  | Human to human Epidemic or<br>transmission? pandemic<br>potential?   | Epidemic or<br>pandemic<br>potential?  |
|---|---|---|---|---|--|--|
| Prion Diseases Chronic Wasting<br>Disease (CWD)******<br>(175, 176) | Wild cervids: Moose,<br>white-tailed and mule deer,<br>elk and reindeer in<br>Fennoscandia (potentially<br>caribou) | No documented transmission to<br>humans as yet, but experimental<br>evidence of CWD transmission<br>to non-human primates. Other<br>TSEs have spread from animals<br>to humans via consumption of<br>infected offal, so public health<br>officials still advise caution | N/A   | Potentially possible through consumption of shared contaminated product   | N/A  | 2  |
| S8 <  | /D)*****  | *   | Wild cervids: Moose,<br>white-tailed and mule deer,<br>elk and reindeer in<br>Fennoscandia (potentially<br>carlbou) | Wild cervids: Moose,<br>white-tailed and mule deer,<br>elk and reindeer in<br>Fennoscandia (potentially<br>carlbou) | Wild cervids: Moose, No documented transmission to N/A<br>white-tailed and mule deer, humans as yet, but experimental<br>ek and reindeer in evidence of CWD transmission<br>Fennoscandia (potentially to non-human primates. Other<br>caribou) TSEs have spread from animals | Wild cervids: Moose, No documented transmission to N/A<br>white-tailed and mule deer, humans as yet, but experimental<br>ek and reindeer in evidence of CWD transmission<br>Fennoscandia (potentially to non-human primates. Other<br>caribou) TSEs have spread from animals |

transmission

for

shown potential

research has

a very prolonged incubation period in people; experimental I

more longitudinal research is warranted (176).

to non-human primates and ability of CWD to convert human prion protein to a misfolded state; CWD is an emerging disease and

| Zoonoses and Northern Wildlife Use |
|------------------------------------|
|------------------------------------|

#### RESULTS

Our review identified 25 zoonotic bacterial (n = 12), parasitic (n= 9), and viral (n = 4) diseases described across Arctic and boreal regions of Canada and Alaska that can be transmitted to humans through the hunting, consumption, preparation, or other use of wildlife (22, 56, 73, 75-84, 86-95, 97, 99, 102-110, 112-117, 120-125, 129-134, 138-142, 145, 149-154, 157-170, 177-193). Table 2 summarizes the diseases, their causative agents, wild hosts, modes of transmission, and potential for local, epidemic or pandemic spread, with additional details given in the Appendix 1. Potential wild species hosts of these 25 zoonoses include many subsistence species important to northern Indigenous Peoples and local communities. These come from various taxa, including carnivores, ungulates, rodents, birds, marine mammals, and fish. In contrast to the global overrepresentation of viruses as emerging and pandemic human pathogens (21, 194-196), viruses comprise the lowest proportion of northern zoonoses. Of the 25 zoonotic diseases reported, 12 were identified as having potential to cause local outbreak clusters through the sharing of contaminated wild animal meat, or other products for consumption (anisakidosis, trematodosis, tapeworms, toxoplasmosis, trichinellosis, anthrax, botulism, brucellosis, mycobacteriosis including tuberculosis, Q fever, tularemia, yersiniosis); 8 have potential for human to human transmission, [anthrax, brucellosis (via breastmilk), influenzas, fever, tuberculosis/mycobacterial disease, versiniosis, 0 caliciviruses, and rabies]; and only 2 of these [influenzas and tuberculosis/mycobacterial disease] are considered as having true epidemic or pandemic potential. Many of these pathogens can threaten human health directly as zoonoses, but also indirectly via food and economic insecurity, by causing mortality events and declines in wildlife, or reduced quality of wildlife products on which northern Indigenous Peoples and local communities depend (120, 178, 179, 193). Thus, these zoonotic diseases are of local significance, even where EID risk of more global significance is low.

A study of 36 Inuit communities for 4 zoonotic parasites, found highest levels of exposure to Toxoplasma gondii (27.2%) and trichinella (18.6%), with overall seropositivity related to age, education, and consumption of marine mammals and seafood (180). Serological surveys of hunters and trappers in Cree communities in northern Québec (Eastmain, Wemindji, and Mistissini) found 44% to be positive for at least one zoonotic pathogen, with risk correlated to hunting, fishing, and trapping activities, as well as consumption of smoked game, and domestic dog ownership (91, 181). Free-ranging domestic dogs serve not only as sentinels for disease, but also as a conduit for diseases from wildlife to people e.g., for rabies and zoonotic parasites which dogs acquire from wild meat (78, 87, 182-184). The role of dogs as potential sources or amplifying hosts for zoonotic diseases highlights the importance of integrating improved domestic animal management and healthcare alongside that for humans in northern communities (197).

While wildlife use in Arctic and boreal biomes is not currently considered a major concern in terms of being the source of an epidemic or pandemic, zoonotic diseases of wildlife remain important for northern communities in the Arctic and boreal biomes: changes in quality and quantity of wildlife, access to traditional and country foods, and potential for disease risk, can create public health, food safety, and food security concerns. Indigenous and local communities in northern and remote areas of these biomes also tend to have limited access to health care services, along with suboptimal housing, infrastructure such as potable water, and sewage treatment facilities. These underlying factors contribute to the cumulative impact of additional stressors like zoonoses on the community (198).

### DISCUSSION

## Zoonotic Pathogen Spillover, Amplification, Epidemic or Pandemic Spread, and Relevant Dynamics in Northern Biomes

Spillover and amplification of novel diseases is a rare, dynamic, and complex process with multiple factors at play (25, 47, 199-203). Zoonotic disease emergence events can result in deadend "spillover" infections in which the pathogen is unable to establish stable onward transmission in the novel (human) host (27, 204). Despite this, spillover events are thought to be under-reported, as they can occur in remote regions where people have limited access to healthcare and reporting, with a lack of surveillance and diagnostic test services, and limited infrastructure, while also living in relative isolation from other human populations (47). Novel diseases are likely often misdiagnosed, or not diagnosed at all in mild cases that resolve without treatment (47). The risk of pathogen spillover from reservoir hosts to humans, or other animals, depends upon the intensity of infection within reservoir host populations, and human contact with the reservoir population, and is affected by a cascade of events involving multiple factors associated with hosts (reservoir, intermediate/vector, recipient), the environment and the pathogen itself (205-208). This includes release of infectious material from reservoir hosts, pathogen survival in the environment, behaviors that drive exposure of a novel host, and biologically driven susceptibility of that host (199, 209). For a spillover event to result in epidemic or pandemic spread, it may need to overcome multiple barriers (210), and the pathogen must adapt to efficiently spread between individuals of the recipient population, with genetic, physiological and immunological attributes of the recipient host, together with dose and route of exposure, affecting the susceptibility to infection (206, 211). The pathogen can then be considered as transformed from a zoonotic EID to a pathogen specific for the recipient population (2). High densities, and gregariousness/sociability of the recipient host species can then facilitate pathogen exposure and transmission. Similarly, connectivity through travel, and trade networks facilitate regional, international, and global pandemic spread of pathogens in humans and domestic animals (212, 213).

Of human pathogens with potential for widespread transmission and global dissemination i.e., pandemic potential, a disproportionate number are viruses (20, 21, 26, 194–196, 214). Viruses transmitted to humans during practices that facilitate

mixing of diverse animal species have significantly higher host plasticity, and higher pandemic potential (201). Zoonotic viruses capable of infecting a more diverse range of host species have higher pandemic potential in humans than those with a narrower host specificity: they are more likely to be amplified by human-to-human transmission, and spread on a global scale (201, 215). Host traits play a role in transmission, and the proportion of zoonotic viruses per mammal species has been linked with phylogenetic relatedness to humans, host taxonomy, host biomass and density, and opportunities for human contact (194, 211, 216–219). Certain mammalian groups (bats, rodents, primates) have been suggested as more likely to host zoonotic viruses (194, 215); though recent analysis suggest bats and rodents host high numbers of zoonoses simply due to the high degree of species richness within these orders, not as a result of intrinsic or ecological differences (220). Whilst wildlife are described as the source of the majority of emerging zoonotic diseases, the term "wildlife" is defined by OIE as "all free-roaming wild animals, feral animals and captive or farmed wildlife" (2). It is valuable to differentiate spillover risk posed by free-roaming wildlife, vs. captive-managed "farmed" or traded wildlife that may have wild origins, but have either been caught and maintained, or bred in captivity. Wildlife trade chains and markets that trade and process live animals or fresh meat from wildlife frequently represent high-risk interfaces due to high wildlife-human contact rates, and large numbers of admixed species (including mixing of wild and domestic animals), with their potential to shed and share viruses for extended periods prior to on-site slaughter or onward sale (221, 222). The proportion of wild rodents testing positive for coronaviruses has been shown to significantly increase along the live animal supply chain from local traders (21%), to large markets (32%), and to restaurants (56%) (32). Environmental and physiological stressors, such as the poor transportation and holding conditions along wildlife trade chains, impact animal immune function, and can lead to increased shedding and amplification of viruses (210, 223-225).

The risk of spillover of zoonotic diseases from wildlife increases when wildlife-human, and wildlife-domestic animalhuman contact rates increase (19-21, 25, 42). Ecosystem degradation, land-use change for agricultural intensification or other industrial expansion, wildlife trade, and wildlife markets, all contribute to increasing contact rates, and therefore increase risk of disease emergence, particularly when they occur in areas of high biodiversity (19-21, 25, 47, 226-241). A higher diversity of zoonotic agents is found where the diversity of host animals is higher (239, 242). Although higher animal host and pathogen species diversity is associated with an increased risk of disease emergence (243), complex host, environmental and pathogen relationships ultimately determine spillover risk (206, 244). A high level of biodiversity can serve as a source of pathogens, but can also serve as a regulating factor (termed the "dilution" effect), where by loss of habitat specialists and predators due to habitat disturbance or hunting, can allow more generalist reservoir host species, such as some rodent species, to proliferate (240, 245). Zoonotic viral richness correlates strongly with mammalian species diversity and abundance (2), and domestic

species harbor, on average, 19.3 zoonotic viruses compared to an average for free-ranging wildlife of 0.23 (2). Wildlife make up <1% of the world's non-human mammal biomass, thus, whilst wildlife is of concern as a source of uncommon but significant animal-origin EIDs, land-use change for agricultural intensification to supply human food systems, particularly for livestock, is of concern as a significant driver of EID risk (19, 24, 238). Higher biodiversity tends to be observed along ecosystem edges (e.g., along roads built into pristine ecosystems, or fragmented forest edges) where differing ecological systems meet, resulting in increased contact opportunities between a wider range of different host and pathogen species, and increased potential pathways for spillover (237, 246, 247). In biodiverse areas, livestock can become intermediate or amplifier hosts in which pathogens can evolve and spill over into humans (238) and over three-quarters of livestock pathogens are capable of infecting multiple host species (219).

# Wildlife Use, Spillover and Emerging Zoonotic Diseases in North American Arctic and Boreal Regions: Social, Cultural, Political, and Ecological Considerations

Arctic and boreal socio-ecological systems are currently less predisposed for novel zoonotic disease emergence, due in part to a relatively low species diversity; low population density of human and livestock; limited and less intensive land uses, such as for commercial forestry and agriculture; and a lack of live wildlife markets, which are not a part of the cultural values of northern communities (19, 25, 206, 220, 239, 248-252). In addition, cold temperatures, photoperiod extremes, and geographic isolation restrict the diversity of pathogens, and often require specialized pathogen adaptations in order to persist in these environments (250, 253) (Figure 1). Recent mapping of global hotspots of relative risk for zoonotic EID events based on demographic, environmental, and biological correlates found Arctic and boreal biomes to be within the lowest risk index (25). Low human density, and reduced connectivity among human populations, also decrease the likelihood of onward pandemic spread of any newly emerged pathogen (28, 213).

Commercialization of wildlife for food has been, and continues to be promoted in northern communities and regions. The type, and context, of sharing and trade of wildlife for food in northern communities is entirely different in character from the commercial trade in wildlife for human consumption, including wildlife markets, in parts of Asia and Africa. The trade is also not characterized by the same mixing of species carrying potential pandemic zoonotic pathogens as commonly occurs in wildlife trade chains throughout tropical regions. As an example, a Nunavut Wildlife Harvest Study found only 86 records (from over 145,000, and after removing the separately-monitored commercial harvest of muskoxen (Ovibos moschatus) and sales to fish or meat plants) indicating a harvest was sold commercially, vs. for personal use (254). Larger-scale commercial trade of wildlife from Arctic and boreal biomes is highly regulated, and falls under similar food safety inspection as meat from domestic animals, which not only lowers the risk of zoonotic disease transmission, but also protects the food and economic security of Indigenous and local communities who rely on wildlife (255– 257). Whilst commercial trade still forms only a small component of wildlife use, the dual roles of wildlife harvesting in northern subsistence and mixed economies, including sport hunting, remain important (258–260). Maintaining and strengthening current surveillance systems for wildlife health and zoonoses, based on populations with the highest contact rates with wildlife, is an important component of an improved health infrastructure across the North. These systems can help detect, define, and control local human emergence of zoonoses while still geographically confined.

Risk for emergence of zoonotic-origin pandemic pathogens from wildlife use in Arctic and boreal biomes is currently minimal, indicating that new policies restricting these traditional and subsistence activities in the name of pandemic prevention would be unfounded at the current time.

Our review did find that endemic zoonotic concerns persist in northern communities (Table 2, Appendix 1), and increased risk is associated with consumption of raw meat, the practice of meat and fish fermentation (e.g., igunaq, muktuk), and exposure to the bodily fluids of animals through methods of harvesting and butchering (56, 118). For many Indigenous cultures around the Arctic, wildlife as food, and food sharing, are fundamental components of a cultural value system that emphasizes generosity, reciprocity, and cooperation, and usually operates within networks related to kinship and family social groups within the community (261-267). Alaska prohibits the sale of most wildlife hunted by Alaska Natives to non-Alaska Natives, but, across Canada, different jurisdictions recognize different Indigenous rights and responsibilities with respect to disposition of country food. For example, in Nunavut, an Inuk has the right to dispose freely to any person any wildlife lawfully harvested (268). Food exchanges are important because of the nutritional value of the items exchanged, and also because they carry cultural and economic values (53, 269, 270). Local sharing of foods can facilitate community zoonotic disease outbreaks (e.g., trichinella or botulism, as noted in Table 2); however, the limited potential for human-to-human transmission of current pathogens spread via sharing, and the low human population density, makes the likelihood of epidemics and pandemics originating from such activities very low. While this traditional way of sharing remains, there also exist other broader networks of sharing practices established to support urban hubs such as Anchorage, and others, across the United States (265, 266). Although low levels of consumption and sharing of traditional foods by Indigenous Peoples in urban centers in Canada have been previously reported (64, 271), relatively new sharing networks have emerged as a result of social media (272, 273) that facilitate the broader distribution of country food, often at the expense of local and traditional sharing practices and values (274), and potentially facilitating the spread of pathogens across larger geographic regions. Zoonotic disease risk is lowered through tight regulation of legal trade of wildlife and wildlife products from northern regions, however international spread of a zoonosis has been associated with sport hunting and the illegal export of wild meat (275).



FIGURE 1 | Transmission of Zoonoses of Wildlife Origin in Arctic and Boreal Regions: Characteristics That Lower Risk for Novel Pathogen Emergence a Spread (Created with BioRender.com).

Despite the perceived current low risk for zoonotic EID events, Arctic and boreal biomes are undergoing rapid ecological change. As the earth warms, and permafrost thaws, concerns have been raised for the potential release of dormant pathogens and contaminants that could affect wildlife and humans. Climate change, migration, introduction of industrial landuse, alongside increasing connectivity of people through development of tourism, travel and trade networks, will alter pathogen dynamics and could facilitate the future spread of emerging pathogens both to and from Arctic and boreal biomes (28, 213, 276, 277). These trends are already reflected in range expansion of existing pathogens (278, 279), and could promote reemergence of past diseases, novel hostpathogen relationships, and the emergence of new zoonotic diseases including vector-borne diseases (121, 185, 277, 280-282). Novel pathogens have recently emerged in northern regions: A second case of Alaskapox virus was reported in Fairbanks in October, 2020, with suspected, but as yet unknown, zoonotic origin (283), and Erysipelothrix has emerged as an apparently new disease causing agent in muskoxen in Canada, with a genotype distinct from strains found in other regions (121).

The greatest risk for pathogens of pandemic potential being introduced into northern communities is currently from people coming into the region from more densely populated southern areas (e.g., via cruise ship passengers). Travel and tourism were found to be the most significant and frequent drivers of epidemic events in Europe, and during the ongoing COVID-19 pandemic (284, 285). The current COVID-19 outbreak is an excellent example of travel-related introduction into northern regions, and subsequent community spread of a suspected wild-origin EID maintained by human-human transmission (286). As a result, many remote northern communities, who experienced impacts from previous epidemics such as influenzas, smallpox, and tuberculosis (287), have imposed strict travel restrictions and prohibited outsiders from entering their communities during the current pandemic (288). Northern residents traveling to and from regional hubs for medical procedures and other essential activities should be considered in future risk scenario planning, particularly during a pandemic. Severe Acute Respiratory Syndrome (SARS) like coronaviruses have not been identified in any wild animals in Arctic and boreal biomes. Natural and experimental infection indicates that SARS-CoV-2 appears to have the ability to infect a broad range

of distantly related mammals (289). Analysis of Angiotensinconverting enzyme 2 (ACE2) receptors, suggests that some cetacean species are hypothetically susceptible, including beluga (*Delphinapterus leucas*) and narwhal (*Monodon monoceros*) (261) which are hunted by northern communities. However, entry of a virus into a cell is a complex biochemical process, with multiple other factors at play beyond the binding to a receptor, and actual infection of marine mammals is undocumented. Marine mammals are not deemed a risk based on current knowledge. Other coronaviruses have previously been identified in marine mammals, but there has been no evidence of zoonotic transmission from these animals to humans (290–293).

Reverse zoonotic transmission of human pathogens to nonhuman animals (anthropozoonoses), including wildlife, occurs more frequently than previously thought (294), and this directional flow of infection for SARS-CoV-2 from humans to wildlife is currently more likely than the reverse in Arctic and boreal biomes. A recent assessment found a non-negligible risk of transmission of SARS-CoV-2 from humans to bats (295), and, whilst the COVID-19 pandemic is ongoing, suspension of field work that involves direct interaction with bats has been recommended (296). Preventing human-to-wildlife SARS-CoV-2 transmission is important for protecting animals from disease, but also to avoid establishment of reservoirs in wild animals (e.g., bats), complicating disease control efforts, with potential for future spillover to other wildlife (e.g., Mustelidae) and spillback to humans (297, 298). Both anthropozoonotic, and zoonotic transmission of SARS-CoV-2 has been reported on mink (Mustela vison) farms across Europe, and the United States (299, 300). Mustelids are highly susceptible to SARS-CoV-2, an important consideration given that trapping of marten (Martes americana) and wolverine (Gulo gulo) forms a critically important industry in many northern communities (301). Contaminated wastewater and refuse from human settlements, commercial vessels, and cruise ships could theoretically pose a risk of transmission of SARS-CoV-2 from humans to marine mammals in the Arctic (261, 302). However, it is unclear if the virus remains viable under these varying environmental conditions and risk likely remains low (303), and is further attenuated by the current ban on cruise ships in the Artic.

## **Future Zoonotic Disease Concerns**

The circumpolar North is uniquely vulnerable to the health impacts of climate change, including, but not limited to, alterations in the distribution and ecology of infectious diseases, expansion of zoonotic disease vectors, changing migration patterns, impacts on food security, limited resources of northern communities to respond to medical emergencies, and changes in water availability and quality (65, 227).

Climate change can impact distribution, life cycle, and physiological status of hosts, pathogens, and vectors, and can drive novel cross-species viral transmission (276, 304, 305). For example, a marked increase in leptospirosis was observed in Ontario following the warmest and third wettest autumn in a decade (186). Leptospirosis has not yet been recorded in humans in the Arctic, but serosurveys of Alaskan wildlife found antibodies to *Leptospira* serovars in caribou (*Rangifer*)

tarandus), moose (Alces alces), and bears (Ursidae) (186). The cumulative impacts of environmental and climatic changes may be increasing the susceptibility to the bacteria Erysipelothrix rhusiopathiae in certain wild animal host populations, including multiple unusual mortality events of muskoxen in Nunavut, the Northwest Territories, and Alaska, and of moose and caribou in British Columbia (120-122). In addition to the health risk for people who interact with these animals or their environments, there is potential for reduced food security for northern communities, through the direct loss of animals from E. rhusiopathiae (56, 120, 123). Climate change is already and will lead to further changes in wildlife community dynamics, including range shifts and increasing overlap between marine and terrestrial ecosystems as a result of sea ice loss and other climate-related changes. Yersinia pestis, a bacterium that persists in rodent-flea communities and causes plague in humans, is not yet reported in the Arctic, though it has been identified twice from wildlife in Canada [in bushy-tailed woodrats (Neotoma cinerea) and a Prairie dog (genus Cynomys)] (149), and appears to occur enzootically in southern Alberta, Saskatchewan, and British Columbia (150). If warming leads to northern expansion of rodent reservoirs, the disease could affect Arctic and more boreal communities in the future. Northward expansion in mammal ranges are already noted, for example in beavers (Castor canadensis) increasingly colonizing the north. Such expansion impacts zoonotic disease ecology, with beavers, for e.g., capable of carrying tularemia, and amplifying and maintaining Giardia (249). Migratory terrestrial and marine intermediate hosts have been implicated in the introduction of other zoonoses to northern regions. For example, human seroprevalence for the protozoan parasite Toxoplasma gondii is high in some parts of the Canadian Arctic, with infection associated with consumption of under-cooked country foods (306). The parasite, thought originally to be of South American origin, but now ubiquitous around the globe, has felids as the only known definitive host. However, seropositive wildlife species have been detected in the Arctic, where wild felids do not occur, and hypotheses for arrival in the region include spread via migratory waterfowl or marine mammal intermediate hosts (306, 307). As environmental conditions change, bacterial infections routinely reported from direct contact with, or ingestion of, fish in more southern regions may also become a greater concern for northern communities who depend on fish for subsistence and as part of their food sovereignty (126). While there are currently no known zoonotic viruses of fish origin, there are many viruses among fish that share evolutionary history with modern human viral pathogens, and viruses have been shown to readily jump between species in the aquatic environment (308). The possibility of some diseases decreasing in relevance warrants consideration when assessing future potential risks from climate change (187, 253, 309).

Thawing of permafrost has already contributed to changing patterns of traditional food consumption, and forced some northern communities to abandon the use of traditional ice cellars and increasingly utilize preservation methods such as smoking, pickling, and salting, and other ways to store traditional and country foods (281).

If pathogens emerge from thawing permafrost, ice patches, glaciers, or graves, wildlife may be the first to be affected. Wildlife may be both important sentinels and amplifying hosts, and a food safety concern (281). In 2016 in the Russian Arctic, one fatality from anthrax occurred among native reindeer herders, and several were hospitalized. One proposed explanation for the outbreak was that warm temperatures melted permafrost, exposing the corpses of reindeer that had died of anthrax almost a 100 years earlier, and releasing infectious Bacillus anthracis spores into nearby waterways. However, another valid explanation is the large increase in reindeer herd sizes, discontinuation of routine anthrax vaccination of reindeer (110), loss of Indigenous Knowledge among herders, and lack of veterinary experience to recognize signs of the disease in reindeer (111). Certain zoonotic disease agents survive particularly well in cold northern climates, including spore-forming bacteria, Mycobacterium species, protozoan cysts/oocysts, some helminth eggs, prions, non-enveloped viruses, and pox viruses (281). Some fungi can survive in permafrost for extended periods, and nematodes can be viable after long-term cryobiosis in Arctic permafrost (310). Most viruses are rapidly inactivated outside host cells: while RNA from the 1918 influenza strain was detected from buried Arctic peoples almost 100 years after their death (18), the material was non-infectious. Giant viruses appear to be more resilient: a giant virus trapped for around 30,000 years was recently isolated from Siberian permafrost (311), and two plant viruses recovered from 700-yr-old caribou feces still demonstrated infectivity (312). In addition to revealing dormant pathogens, climate warming also leads to release of persistent environmental pollutants, heavy metals, carbon and other natural elements from soils and rocks (313). These have cumulative effects on the immune systems and health of wildlife and humans, increasing susceptibility to diseases, particularly when combined with additional climate-related stressors such as reduced or altered habitat and food availability for wildlife, and, in turn, for humans (186).

In terms of pathogens of epidemic or pandemic potential, climate-driven changes that influence wild Arctic water bird habitat use, distribution, and migration could be a factor in the global distribution of avian viral agents, and possibly the emergence of a new pandemic influenza strain (186). Several recent studies provide evidence that migratory birds serve as effective long-distance vectors of wildlife and zoonotic pathogens to the Arctic (282), with boreal regions providing stop-over sites for many migratory species (314–316).

Most vector-borne disease agents have limited pandemic potential, as their spread tends to be restrained by geographically and climatologically restricted vector habitats (317). However, with warming environmental temperatures, and changing precipitation levels, vector-borne pathogens will become of increasing concern for epidemics in Arctic and boreal biomes, as vector ranges expand. Such viruses include those spread through mosquito bites like West Nile virus and Sindbis virus, which are maintained in wild birds; California serogroup viruses, such as Jamestown Canyon virus (antibodies to which have been found in bison (*Bison bison*), Dall's sheep (*Ovis dalli*), snowshoe hare (*Lepus americanus*), and Arctic fox (*Vulpes lagopus*) in Alaska); and Snowshoe hare virus which is maintained in mammalian hosts including rodents, deer, and hares (186, 276). Tick borne-encephalitis virus can also infect multiple hosts including ungulates, birds, rodents, and carnivores, and is spread from animals to humans through tick bites. In the Russian Arctic, climate associated increases in mosquito and tick populations, and rodent-borne infections transmitted by arthropods are already having an impact on population health (281). Insect repellent and avoidance of tick bites will likely become increasingly important for people living in northern communities.

As existing agricultural regions are threatened by climate change, warming of high latitude regions, and increasing food demands may lead to northward expansion of global agriculture (318). Agriculture is associated with the emergence of more than 50% of zoonotic infectious diseases in humans (319), and future rates of zoonotic disease emergence and/or reemergence are predicted to be closely linked to the evolution of the agriculture-environment nexus (42, 202, 238, 320). As opportunities for agriculture, and other industries, increase with warming of Arctic and boreal biomes, there is potential for disruption and dramatic change of these ecosystems from an influx of people, domestic livestock, and pets, and the disease agents they harbor (226, 243, 321). Beyond prospective increases in infectious disease emergence, climate-driven agricultural expansion will have major impacts on biodiversity, on downstream water resources, and on carbon storage (322). Multiple links exist between human health and anthropogenic environmental degradation. Land-use modifications that lead to the loss of ecosystem integrity, and increased human-domestic animal-wildlife contact rates can increase susceptibility to emerging zoonoses directly, but also indirectly via impacts on the immune system, mental health issues, environmental contaminants, and endemic diseases, paired with reduced access to fundamental services such as timber, freshwater, wild foods, medicines, and decreased air quality (248). A systematic, interdisciplinary, and holistic approach to understand and address environmental health, zoonotic, and other disease concerns around agricultural and other developments are key to mitigating negative impacts (323-325).

# Current and Future Considerations for Monitoring, Surveillance, and Risk Reduction Approaches

Given the changing socio-ecological, -economic, and political systems of Arctic and boreal biomes, future research would be beneficial to understand how the increasing anthropogenic impacts across northern regions are altering ecological processes, and thereby potentially converting microbial hazards in naturally occurring pathogen diversity into risks to human health (277, 326, 327).

Human health monitoring for zoonotic diseases in Arctic and boreal biomes is typically coordinated through relevant local health authorities and health surveys (e.g., Inuit Health Survey). Outbreaks can lead to new sampling programs, such as the sampling of walrus (Odobenus rosmarus) meat before sharing and distribution in the community (189). In general, there has been minimal research on the risks associated with traditional and country food preparation practices, despite awareness of zoonotic pathogens found in harvested wildlife and a clear need for risk reduction (118). Good hygiene during butchering and skinning, and thorough cooking of food can prevent transmission of many of the endemic zoonotic pathogens reported (118). Most health status measures and outcomes are consistently poorer for Indigenous Peoples in comparison to the rest of the North American population (63, 328) and investing in health infrastructure across Arctic and boreal biomes is critical to meet the broader needs of these populations. These investments would enhance emerging infectious disease surveillance, contribute toward a better understanding of the patterns of exposure and immunity, and reduce risks from both endemic and emerging zoonoses (79, 248, 329). Ensuring ownership of health initiatives by local communities in general, and specifically in the necessary prioritization process, is essential, with ample evidence demonstrating that failure to engage and build trust with local political and thought leaders, Elders, traditional health workers, and community groups in disease detection and control, will delay both diagnosis and response for emerging diseases (330). Identifying priority pathogens is a valuable starting point given present-day perceived financial constraints (281), though broader monitoring systems capturing syndromic trends may be more sensitive to detect emerging pathogens. Furthermore, every community has location-specific, individual risk profiles that will help determine the discrete approaches needed. To provide timely, accurate, and pertinent information on zoonotic disease risks across northern regions, the current capacity for northern communities to establish or continue disease monitoring and diagnosis needs to be expanded (186, 193, 331).

National, regional, state and territorial level veterinarians and programs monitor the health of wildlife across North American Arctic and boreal biomes. These programs range from highly standardized to less formal efforts, and vary in their objectives, from basic data collection, to in-depth species specific research. Recommendations regarding the reduction of disease risk from wildlife are detailed in a variety of local guidance publications (see Table 3). Furthermore, many groups publish regular or as-needed health updates through websites, comanagement meetings, and scientific gatherings. The Canadian Wildlife Health Cooperative (332) forms a cross-Canada network of collaborators dedicated to wildlife health and a national surveillance program for wildlife diseases, but the approach to date has been essentially reactive to emerging issues. A new "Pan-Canadian Approach to Wildlife Health" was initiated in 2018 (333), and represents a deliberate transition to proactive health promotion; however, increased funding commitments, and jurisdictional collaboration and coordination, are necessary to enable its execution. In Alaska, the U.S. Geological Survey's National Wildlife Health Center (NWHC) (334) supports wildlife disease detection, control, and prevention, and conducts wildlife disease outbreak investigations as part of a national, general surveillance program. The Alaska Department of Fish and Game has a longstanding wildlife disease monitoring program and wildlife health monitoring also occurs at local levels: the North Slope Borough Department of Wildlife Management has multiple harvest monitoring programs for subsistence species [e.g., bowhead whale (*Balaena mysticetus*), beluga, ice seals (Phocidae), walrus, polar bears (*Ursus maritimus*)], and has a well-developed wildlife health research program for these mammals. This program, and an increasing number of other programs in northern regions, are community-based, and work with hunters and communities to answer questions about health and diseases in wildlife in a changing Arctic environment. Some examples of current monitoring institutions and efforts regarding health of wildlife and traditional foods are shown in **Table 4**.

Community-based monitoring, Indigenous Knowledge, participatory epidemiology, and citizen science have become increasingly relevant as ways of centering research on community needs and priorities, and obtaining invaluable information on health and the environment (190, 193, 197, 201, 331, 335-338). Rich knowledge, including on preventing certain food-borne illness, results from the long-standing relationships of Indigenous Peoples and local communities with their environment, including their harvesting of nutritionally and spiritually important native plants, fish, and wildlife (56, 57, 118, 339, 340). Indigenous knowledge; the prevention, monitoring, and surveillance of zoonotic agents; and education are considered the most important methods to reduce human health risks associated with the consumption of traditional and country foods (118). Many wildlife harvesters in Arctic and boreal biomes are keenly aware of concerns regarding wildlife health and food safety. In general, parasites, lesions (discoloration, tracts, fungal growths, etc.), or abnormal behavior of hunted animals are often observed by Indigenous and local communities. Indigenous Peoples were some of the first to draw attention to traditional and country food safety concerns related to environmental contaminants, because of changes they detected in quality of the animals and fish they hunted (118). Community-based monitoring programs contribute valuable information. For example, the Nunavik Trichinellosis Prevention Program provides rapid carcass-side testing for trichinella in country foods for Nunavik communities (341), giving community members control to obtain the information they need to make informed decisions about food preparation, consumption, and carcass disposal to prevent further transmission. Recent working groups assessing Unusual Mortality Events (UMEs) for marine mammals and other species around Alaska include Indigenous perspectives and observations (191, 342) and, across Canada, several Indigenous-led Guardians Programs monitor ecological health, maintain cultural sites, and protect sensitive areas and species, while also playing a vital role in creating conservation plans, and supporting Indigenous Protected and Conserved Areas (343). Table 3 lists some examples of the existing monitoring efforts for wildlife health, and country and traditional foods. Ongoing academic-government-community collaborative partnerships have resulted in several successful community-based wildlife health surveillance programs that aim to address community concerns about wildlife health and food safety while simultaneously addressing key research questions about wildlife ecology and dynamics (193, 197, 338).

#### TABLE 3 | Examples of Existing Guidance on Safety for Hunters.

| Hunter guidance document and source  | Reference and website for accessing document  |
|--|---|
| Safety manual for harvesters of fish and wildlife in nunavut                               | Canadian Wildlife Health Cooperative and Government of Nunavut. (2011). Safety Manual for Harvesters of Fish and Wildlife in Nunavut: An Illustrated Guide to Common Diseases and Parasites.<br>https://www.gov.nu.ca/sites/default/files/files/Safety%20Manual%20for%20Harvesters%20of%20Fish%20&%20Wildlife%20in<br>%20Nunavut.pdf (accessed March 10, 2021)                      |
| Disease precautions for hunters  | American Veterinary Medical Association (2021).<br>https://www.avma.org/resources/public-health/disease-precautions-hunters#protecting (webpage only) (accessed March 10,<br>2021)  |
| Diseases you can get from wildlife   | Government of British Columbia (2017). Diseases You Can Get From Wildlife: A Field-guide for Hunters, Trappers, Anglers and<br>Biologists.<br>https://www2.gov.bc.ca/assets/gov/environment/plants-animals-and-ecosystems/wildlife-wildlife-habitat/wildlife-health/<br>wildlife-health-documents/diseases_you_can_get_from_wildlife_field_guide_2017.pdf (accessed March 10, 2021) |
| A field guide to common wildlife<br>diseases and parasites in the<br>northwest territories | Government of Northwest Territories (2017). A Field Guide to Common Wildlife Diseases and Parasites in the Northwest<br>Territories'; 6th Edition, March 2017.<br>https://www.enr.gov.nt.ca/sites/enr/files/field_guide_wildlife_diseases.pdf (accessed March 10, 2021)   |
| Common wildlife parasites and diseases   | Alaska Department of Fish and Game (2021). Common Wildlife Parasites and Diseases.<br>Available online at: https://www.adfg.alaska.gov/static/home/library/pdfs/wildlife/brochures_newsletters/common_wildlife_<br>parasites_diseases.pdf<br>and<br>https://www.adfg.alaska.gov/index.cfm?adfg=disease.main (webpage only) (accessed March 10, 2021)                                |

TABLE 4 | Examples of current health monitoring institutions and efforts for health of wildlife and traditional foods (not intended to be comprehensive).

| Institution   | Website   |
|---|---|
| Alaska Native Tribal Health Consortium                      | https://anthc.org/what-we-do/traditional-foods-and-nutrition/   |
| North Slope Borough Department of Wildlife Management       | http://www.north-slope.org/departments/wildlife-management/studies-and-research-projects/health-<br>assessment-of-subsistence-resources |
| Unusual Mortality Events (UMEs)                             | https://www.fisheries.noaa.gov/insight/understanding-marine-mammal-unusual-mortality-events   |
| Community-Based Monitoring                                  | https://www.inuitcircumpolar.com/wp-content/uploads/2019/01/cbm_report_final.pdf  |
| LEO Network   | https://www.leonetwork.org/en/#lat=60.71611&lng=-135.05375&zoom=7   |
| Canadian Wildlife Health Cooperative                        | http://www.cwhc-rcsf.ca/  |
| JS Geological Survey (USGS) National Wildlife Health Center | https://www.usgs.gov/centers/nwhc   |
| US National Oceanic and Atmospheric Administration (NOAA)   | https://www.fisheries.noaa.gov/national/marine-life-distress/marine-mammal-health-and-stranding-response-program                        |
| USGS Alaska Science Center                                  | https://www.usgs.gov/centers/asc/science-topics/wildlife-disease  |

Inuit, Métis, First Nations, and Alaskan Native societies across the Arctic and boreal biomes (and globally) maintain vast and holistic Indigenous Knowledge Systems, across generations, about the natural environment and how it is changing. Such awareness and presence of eyes-in-the-field is irreplaceable for the early detection of changes in wildlife populations, and the environment. Combining Indigenous Knowledge with scientific understanding improves wildlife surveillance, fosters reconciliation, and advances Indigenous Peoples' selfdetermination in research, while creating mutual health and conservation benefits (331, 344-348). However, a history of colonialism, relocations, residential schools, and loss of Indigenous languages has led to generational changes in diet, life-style and relationships with the environment for many Indigenous communities. Loss of transgenerational "hands-on" and oral transmission of knowledge, especially regarding harvesting, butchering, food safety and zoonoses, may increase zoonotic risk, increase meat wastage, and limit the ability to engage with "two-eyed-seeing" within participatory epidemiology (331). In addition to fostering reconciliation with Indigenous Peoples, programs that assist Elders to pass on Indigenous Knowledge to Indigenous youth, and brings communities together to prepare and share country and traditional foods, contributes to future monitoring of wildlife, human and environmental health, and supports a conservation economy focused on the land and wildlife. The participation and leadership of men and women involved in the harvest and preparation of wild foods, and youth in wildlife and community health monitoring and research, builds trust between researchers and local communities, particularly when focused on community-raised concerns and priorities (197, 331). Continued investment in existing communitybased wildlife and human health monitoring efforts, paired with consistent and well-conveyed methodologies, would be a valuable approach to tracking wild harvest use and trade, zoonotic and other emerging pathogens, host species, and environmental impacts from the changing climate across northern communities.

Early recognition and intervention during an emerging infectious zoonotic disease event is essential to limit spread (349). Wild animals, and domestic dogs, can serve as sentinels for zoonotic diseases and other health concerns such as contaminants and it appears essential that existing veterinary and wildlife surveillance systems for zoonotic pathogens are closely integrated with public health surveillance, to better control such pathogens before they affect human health. Surveillance would benefit from the exploration and cooperative development with local and Indigenous leaders of practical ways to integrate and share surveillance-generated information for humans, domestic animals and wildlife. This would ensure that the surveillance systems are risk-based, spatially and temporally targeting species and geographic locations with the highest risk for spillover, and guaranteeing that the information gathered can be used by the local communities to deploy interventions in a timely manner. Expansion of regional, transdisciplinary One Health networks that involve community members and diverse stakeholders and collate real-time multifactorial data and explicit observations, (e.g., the Echo Network for health and conservation, the Circumpolar Climate Change and Infectious Diseases Workgroup, or the LEO network<sup>1</sup>) (350) and integration with public health networks such as the International Circumpolar Surveillance (ICS) project (192), can facilitate earlywarning systems, rapid response, and mitigation for health threats to animals, humans, and the environment, while also advancing our understanding of endemic, and emerging zoonotic health risks for these communities.

It is important that the desire to increase awareness of the potential risk of zoonotic infections be balanced with the unintended consequences on other determinants of health. Past experience with poorly designed communication strategies and materials with respect to country and traditional foods, such as contaminants in wild-caught foods, provides stark lessons on the need to avoid provoking unnecessary fear when addressing food safety risk in Indigenous and local communities (71, 72). For example, scientific communication to Nunavut Inuit communities in the late 1980s that women's breast milk had high levels of polychlorinated biphenyls (PCBs), failed to consider broader understanding of contamination issues in the region, and failed to include the Inuit directly on messaging (72). The resulting alarm and confusion across Inuit and other northern communities led many women to stop breastfeeding, and stopped consumption of country and traditional foods, leading to other more insidious health issues, poor nutrition, and food insecurity (72). Similar issues were associated with the research and communication of contaminants in wildlife in Alaska. Participation in subsistence activities, including the practice of hunting by Indigenous Peoples, is considered a protective mental health factor for Indigenous circumpolar youth (351), and there are different and pertinent health concerns associated with the consumption of store-bought, processed, imported market foods increasingly eaten by younger generations. Such foods are often low in nutritional value, high in sugar, and sometimes harbor contaminants including microplastics and pesticides (56, 352-354). In Alaska, the introduction of modern materials such as plastic bags, plastic pails and glass jars in the preparation of rendered and aged foods (muktuk and igunaq) actually increased outbreaks of botulism (118). Indigenous and non-Indigenous health professionals promoted the return to traditional methods thereby reducing fatality rates by leveraging knowledge from community members, Elders, and survivors of botulism. Communications and management approaches around zoonotic disease risk from traditional and country foods are better co-developed with Indigenous and other hunting/trapping communities, with mindfulness that hunters and their families are keenly aware of changes in the behavior of wildlife or condition of wildlife products; these foods are usually the most cost-effective and healthy sources of nutrition in northern and remote communities; and country and traditional foods have important cultural, spiritual, and social values to northern communities (355, 356).

Combining diverse approaches and ways of knowing, including Indigenous Knowledge, provides a more complete understanding of the socio-ecological system than applying Western science alone. Working together can help bridge gaps in scientific monitoring, and bring the best available knowledge to more effectively monitor, and respond to, the impacts of disease and climate change on the health of Arctic and boreal inhabitants (338).

# CONCLUSION

An essential connection exists between subsistence activities, and well-being and resilience within Indigenous Peoples and local communities across the Arctic and boreal biomes in North America. The current threat of pandemic zoonotic disease emergence from hunting, consumption, and use of wildlife in the North American Arctic and boreal biomes is low, and policies restricting these traditional and subsistence activities in the name of pandemic prevention would be greatly misplaced. Health threats from endemic zoonotic diseases in northern biomes remain, with plausible hypotheses that environmental alterations from development, and climate change could alter epidemic and pandemic risks. Whilst commercialization of trade in wild food and products, and agricultural expansion have been identified as factors important in developing resilience in the North (351, 356, 357), they will bring a different set of challenges around zoonotic disease emergence, and disruption of ecosystems (238, 323). Co-development of any future policies and interventions with Indigenous Peoples and local communities may more effectively address zoonotic disease emergence in these rapidly

<sup>&</sup>lt;sup>1</sup>The LEO Network collates and shares information on unusual events, climate and other drivers of environmental change. The online platform is multi-lingual, including Indigenous languages, and was designed to be accessible from remote locations, in order to gather input from local community members, using holistic observational data, and welcoming diverse knowledge systems. The network facilitates connectivity between communities, environmental and public health experts, and allows researchers and agencies to use data as a surveillance system for local updates on emerging issues and vulnerabilities, and to provide event specific consultations.

changing northern regions. Combining Indigenous Knowledge Systems with science provides a more holistic understanding of zoonotic risks in Arctic and boreal ecosystems, informs opportunities for mitigation and monitoring, and improves disease risk communications, while avoiding potentially negative repercussions (331, 356, 358, 359). Monitoring, research, and response efforts would all benefit from employing more inclusive One Health approaches that draw on all knowledge systems and types of expertise, and proactively incorporate the complexity and interrelatedness of the environmental, biological, economic, political, cultural, and social dimensions of zoonotic disease emergence in northern biomes.

## **AUTHOR CONTRIBUTIONS**

LK, CW, MR and JR contributed to conception and design of the study. LK organized the database, performed the review and analysis and wrote the first draft of the manuscript. MR and C-LC

## REFERENCES

- 1. Tabish S. Recent trends in emerging infectious diseases. Int J Health. (2009) 3:V-VIII.
- Haider N, Rothman-Ostrow P, Osman AY, Arruda LB, Macfarlane-Berry L, Elton L, et al. COVID-19—Zoonosis or emerging infectious disease? Front Public Health. (2020) 8:596944. doi: 10.3389/fpubh.2020.596944
- Li HY, Zhu GJ, Zhang YZ, Zhang LB, Hagan EA, Martinez S, et al. A qualitative study of zoonotic risk factors among rural communities in southern China. Int Health. (2020) 12:77–85. doi: 10.1093/inthealth/ihaa001
- López-Robles G, Montalvo-Corral M, Caire-Juvera G, Ayora-Talavera G, Hernández J. Seroprevalence and risk factors for swine influenza zoonotic transmission in swine workers from Northwestern Mexico. *Transbound Emerg Dis.* (2012) 59:183–8. doi: 10.1111/j.1865-1682.2011.01250.x
- Morse SS. Factors in the emergence of infectious diseases. In: Price-Smith AT, editor. *Plagues Politics*. London: Palgrave Macmillan (2001). p. 8–26. doi: 10.1057/9780230524248\_2
- Taylor LH, Latham SM, Woolhouse MEJ. Risk factors for human disease emergence. *Philos Trans R Soc London Ser B Biol Sci.* (2001) 356:983–9. doi: 10.1098/rstb.2001.0888
- Woolhouse MEJ, Haydon DT, Antia R. Emerging pathogens: the epidemiology and evolution of species jumps. *Trends Ecol Evol.* (2005) 20:238–44. doi: 10.1016/j.tree.2005.02.009
- Loh EH, Zambrana-Torrelio C, Olival KJ, Bogich TL, Johnson CK, Mazet JAK, et al. Targeting transmission pathways for emerging zoonotic disease surveillance and control. *Vector Borne Zoonotic Dis.* (2015) 15:432–7. doi: 10.1089/vbz.2013.1563
- Hillson D, Hulett D. Assessing Risk Probability: Impact Alternative Approaches. Prague: PMI® Global Congress 2004—EMEA (2004). Available online at: https://www.pmi.org/learning/library/assessing-risk-probabilityimpact-alternative-approaches-8444 (accessed March 10, 2021).
- Zhang L, Shen F, Chen F, Lin Z. Origin and evolution of the 2019 novel coronavirus. *Clin Infect Dis.* (2020) 71:882–3. doi: 10.1093/cid/ciaa112
- 11. Zhang X, Chen X, Zhang Z, Roy A, Shen Y. Strategies to trace back the origin of COVID-19. J Infect. (2020) 80:e39–40. doi: 10.1016/j.jinf.2020.03.032
- Andersen KG, Rambaut A, Lipkin WI, Holmes EC, Garry RF. The proximal origin of SARS-CoV-2. *Nat Med.* (2020) 26:450–2. doi: 10.1038/s41591-020-0820-9
- MacLean OA, Lytras S, Weaver S, Singer JB, Boni MF, Lemey P, et al. Natural selection in the evolution of SARS-CoV-2 in bats created a generalist virus and highly capable human pathogen. *PLOS Biol.* (2021) 19:e3001115. doi: 10.1371/journal.pbio.3001115

also wrote sections of the manuscript. All authors contributed to manuscript revision and additions, further development of the research concept and read, and approved the submitted version.

## ACKNOWLEDGMENTS

The authors would like to acknowledge and thank Ms. Jean Allen, and Ms. Sharon Edmunds of Nunavut Tunngavik Inc., and Robert Suydam of North Slope Department of Wildlife Management, for their valuable input to the manuscript. Any use of trade, firm, or product names is for descriptive purposes only and does not imply endorsement by the U.S. Government.

## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpubh. 2021.627654/full#supplementary-material

- Lau SKP, Woo PCY, Li KSM, Huang Y, Tsoi HW, Wong BHL, et al. Severe acute respiratory syndrome coronavirus-like virus in Chinese horseshoe bats. *Proc Natl Acad Sci USA*. (2005) 102:14040–5. doi: 10.1073/pnas.0506735102
- Wang M, Yan M, Xu H, Liang W, Kan B, Zheng B, et al. SARS-CoV infection in a restaurant from palm civet. *Emerg Infect Dis.* (2005) 11:1860–5. doi: 10.3201/eid1112.041293
- Wang LF, Eaton BT. Bats, civets and the emergence of SIRS. Curr Topics Microbiol Immunol. (2007) 315:325–344. doi: 10.1007/978-3-540-70962-6\_13
- Song H-D, Tu C-C, Zhang G-W, Wang S-Y, Zheng K, Lei L-C, et al. Cross-host evolution of severe acute respiratory syndrome coronavirus in palm civet and human. *Proc Natl Acad Sic USA*. (2005) 102:2430–5. doi: 10.1073/pnas.0409608102
- Taubenberger JK, Hultin JV, Morens DM. Discovery and characterization of the 1918 pandemic influenza virus in historical context. *Antivir Ther.* (2007) 12:581–91.
- Jones KE, Patel NG, Levy MA, Storeygard A, Balk D, Gittleman JL, et al. Global trends in emerging infectious diseases. *Nature*. (2008) 451:990–3. doi: 10.1038/nature06536
- Karesh WB, Dobson A, Lloyd-Smith JO, Lubroth J, Dixon MA, Bennett M, et al. Ecology of zoonoses: natural and unnatural histories. *Lancet.* (2012) 380:1936–45. doi: 10.1016/S0140-6736(12)61678-X
- Morse SS, Mazet JAK, Woolhouse M, Parrish CR, Carroll D, Karesh WB, et al. Prediction and prevention of the next pandemic zoonosis. *Lancet.* (2012) 380:1956–65. doi: 10.1016/S0140-6736(12)61684-5
- Schurer JM, Bouchard E, Bryant A, Revell S, Chavis G, Lichtenwalner A, et al. Echinococcus in wild canids in Québec (Canada) and Maine (USA). *PLoS Negl Trop Dis.* (2018) 12:e0006712. doi: 10.1371/journal.pntd.0006712
- Temmam S, Desnues C. Zoonotic viruses: how to monitor their emergence? Virologie. (2016) 20:231–44. doi: 10.1684/vir.2016.0660
- McMichael AJ. Environmental and social influences on emerging infectious diseases: past, present and future. *Philos Trans R Soc London Ser B Biol Sci.* (2004) 359:1049–58. doi: 10.1098/rstb.2004.1480
- Allen T, Murray KA, Zambrana-Torrelio C, Morse SS, Rondinini C, Di Marco M, et al. Global hotspots and correlates of emerging zoonotic diseases. *Nat Commun.* (2017) 8:1124. doi: 10.1038/s41467-017-00923-8
- Carroll D, Daszak P, Wolfe ND, Gao GF, Morel CM, Morzaria S, et al. The global virome project. *Science.* (2018) 359:872–4. doi: 10.1126/science.aap7463
- Geoghegan JL, Senior AM, Giallonardo F Di, Holmes EC. Virological factors that increase the transmissibility of emerging human viruses. *Proc Natl Acad Sci USA*. (2016) 113:4170–5. doi: 10.1073/pnas.1521582113

- Pike J, Bogich T, Elwood S, Finnoff DC, Daszak P. Economic optimization of a global strategy to address the pandemic threat. *Proc Natl Acad Sci USA*. (2014) 111:18519–23. doi: 10.1073/pnas.1412661112
- Cantlay JC, Ingram DJ, Meredith AL. A review of zoonotic infection risks associated with the wild meat trade in Malaysia. *Ecohealth*. (2017) 14:361–88. doi: 10.1007/s10393-017-1229-x
- Chomel BB, Belotto A, Meslin FX. Wildlife, exotic pets, and emerging zoonoses. *Emerg Infect Dis.* (2007) 13:6–11. doi: 10.3201/eid1301.060480
- 31. Travis DA, Watson RP, Tauer A. The spread of pathogens through trade in wildlife. *Rev Sci Tech OIE*. (2011) 30:219–39. doi: 10.20506/rst.30.1.2035
- 32. Huong NQ, Nga NTT, Long N Van, Luu BD, Latinne A, Pruvot M, et al. Coronavirus testing indicates transmission risk increases along wildlife supply chains for human consumption in Viet Nam, 2013-2014. PLoS ONE. (2020) 15:e0237129. doi: 10.1371/journal.pone.0237129
- Durand B, Lecollinet S, Beck C, Martínez-López B, Balenghien T, Chevalier V. Identification of hotspots in the european union for the introduction of four zoonotic arboviroses by live animal trade. *PLoS ONE.* (2013) 8:e70000. doi: 10.1371/journal.pone.0070000
- Fèvre EM, Bronsvoort BMDC, Hamilton KA, Cleaveland S. Animal movements and the spread of infectious diseases. *Trends Microbiol.* (2006) 14:125–31. doi: 10.1016/j.tim.2006.01.004
- 35. Greatorex ZF, Olson SH, Singhalath S, Silithammavong S, Khammavong K, Fine AE, et al. Wildlife trade and human health in lao PDR: an assessment of the zoonotic disease risk in markets. *PLoS ONE.* (2016) 11:e0150666. doi: 10.1371/journal.pone.0150666
- Hoffmann B, Tappe D, Höper D, Herden C, Boldt A, Mawrin C, et al. A variegated squirrel bornavirus associated with fatal human encephalitis. N Engl J Med. (2015) 373:154–62. doi: 10.1056/NEJMoa1415627
- Karesh WB, Cook RA, Bennett EL, Newcomb J. Wildlife trade and global disease emergence. *Emerg Infect Dis.* (2005) 11:1000–2. doi: 10.3201/eid1107.050194
- Kurpiers LA, Schulte-Herbrüggen B, Ejotre I, Reeder DM. Bushmeat and emerging infectious diseases: lessons from Africa. *Probl Wildl.* (2016) 507– 51. doi: 10.1007/978-3-319-22246-2\_24
- Marano N, Arguin PM, Pappaioanou M. Impact of globalization and animal trade on infectious disease ecology. *Emerg Infect Dis.* (2007) 13:1807–9. doi: 10.3201/eid1312.071276
- Pavlin BI, Schloegel LM, Daszak P. Risk of importing zoonotic diseases through wildlife trade, United States. *Emerg Infect Dis.* (2009) 15:1721–6. doi: 10.3201/eid1511.090467
- ProMED. ProMED-Mail Post: Subject: PRO/AH/EDR> COVID-19 Update (102): China, Origin, Farmed Wild Animals susp. (2021). Available online at: https://promedmail.org/promed-post/?id=8251684 (accessed March 17, 2021).
- 42. Murray KA, Allen T, Loh E, Machalaba C, Daszak P. Emerging viral zoonoses from wildlife associated with animal-based food systems: risks and opportunities. In: Jay-Russell M, Doyle M, editors. *Food Safety Risks from Wildlife*. Cham: Springer International Publishing (2015). p. 31–57. doi: 10.1007/978-3-319-24442-6\_2
- Dobson AP, Pimm SL, Hannah L, Kaufman L, Ahumada JA, Ando AW, et al. Ecology and economics for pandemic prevention. *Science*. (2020) 369:379–81. doi: 10.1126/science.abc3189
- Walzer C. COVID-19 and the curse of piecemeal perspectives. Front Vet Sci. (2020) 7:582983. doi: 10.3389/fvets.2020.582983
- 45. WHO. WHO Director-General's Opening Remarks at the Media Briefing on COVID-19-17 April 2020. (2020). Available online at: https://www.who. int/dg/speeches/detail/who-director-general-s-opening-remarks-at-themedia-briefing-on-covid-19-\$-\$17-april-2020 (accessed May 26, 2020).
- Zavaleta-Cortijo C, Ford JD, Arotoma-Rojas I, Lwasa S, Lancha-Rucoba G, García PJ, et al. Climate change and COVID-19: reinforcing indigenous food systems. *Lancet Planet Health*. (2020) 4:e381–2. doi: 10.1016/S2542-5196(20)30173-X
- 47. Johnson CK, Hitchens PL, Pandit PS, Rushmore J, Evans TS, Young CCW, et al. Global shifts in mammalian population trends reveal key predictors of virus spillover risk. *Proc R Soc B Biol Sci.* (2020) 287:20192736. doi: 10.1098/rspb.2019.2736
- 48. United Nations. United Nations Declaration on the Rights of Indigenous Peoples. New York, NY: United Nations (2007).

- Kuhnlein HV, Receveur O, Chan HM. Traditional food systems research with Canadian indigenous peoples. Int J Circumpolar Health. (2001) 60:112–22.
- Walch A, Bersamin A, Loring P, Johnson R, Tholl M. A scoping review of traditional food security in Alaska. Int J Circumpolar Health. (2018) 77:1419678. doi: 10.1080/22423982.2017.1419678
- 51. Bjerregaard P, Kue Young T, Dewailly E, Ebbesson SOE. Review article: indigenous health in the Arctic: an overview of the circumpolar inuit population. *Scand J Public Health.* (2004) 32:390–5. doi: 10.1080/14034940410028398
- Loring PA, Gerlach SC. Food, culture, and human health in Alaska: an integrative health approach to food security. *Environ Sci Policy*. (2009) 12:466–78. doi: 10.1016/j.envsci.2008.10.006
- Collings P, Marten MG, Pearce T, Young AG. Country food sharing networks, household structure, and implications for understanding food insecurity in Arctic Canada. *Ecol Food Nutr.* (2016) 55:30–49. doi: 10.1080/03670244.2015.1072812
- 54. McGrath-Hanna NK, Greene DM, Tavernier RJ, Bult-Ito A. Diet and mental health in the Arctic: is diet an important risk factor for mental health in circumpolar peoples?–A review. *Int J Circumpolar Health*. (2003) 62:228–41. doi: 10.3402/ijch.v62i3.17560
- Reeves RR. The origins and character of "aboriginal subsistence" whaling: a global review. *Mamm Rev.* (2002) 32:71–106. doi: 10.1046/j.1365-2907.2002.00100.x
- CCA. Aboriginal Food Security in northern Canada: An Assessment of the State of Knowledge. Ottawa, ON: Council of Canadian Academies (2014).
- 57. ICC. Alaskan Inuit Food Security Conceptual Framework: How To Assess The Arctic From An Inuit Perspective. Anchorage, AK: Inuit Circumpolar Council-Alaska (2015).
- Food and Agriculture Organisation. An Introduction to the Basic Concepts of Food Security. Rome: Food and Agriculture Organisation (2008).
- 59. World Forum on Food Sovereignty. *Final Declaration of the World Forum on Food Sovereignty*. Havana: World Forum on Food Sovereignty (2001).
- 60. Government of Canada. *Government of Canada Constitution Acts*, 1867 to 1982: Rights of the Aboriginal Peoples of Canada. Ottawa, ON: Government of Canada (2016).
- 61. Truth and Reconciliation Commission. *Truth and Reconciliation Commission of Canada: Calls to Action.* Winnipeg, MB: Truth and Reconciliation Commission (2015).
- 62. Lougheed T. The changing landscape of arctic traditional food. *Environ Health Perspect*. (2010) 118:a386–93. doi: 10.1289/ehp.118-a386
- Kuhnlein H, Erasmus B, Creed-Kanashiro H, Englberger L, Okeke C, Turner N, et al. Indigenous peoples' food systems for health: finding interventions that work. *Public Health Nutr.* (2006) 9:1013–19. doi: 10.1017/PHN2006987
- Elliott B, Jayatilaka D, Brown C, Varley L, Corbett KK. "We are not being heard": aboriginal perspectives on traditional foods access and food security. J Environ Public Health. (2012) 2012:1–9. doi: 10.1155/2012/ 130945
- 65. Farquhar SD. Inuit seal hunting in canada: emerging narratives in an old controversy. *Arctic.* (2020) 73:13–9. doi: 10.14430/arctic69833
- 66. Chan L, Receveur O, Batal M, Sadik T, Schwartz H, Ing A, et al. First Nations Food, Nutrition and Environment Study (FNFNES): Results from Saskatchewan (2015). Ottawa, ON: University of Ottawa (2018).
- 67. Goettke E, Reynolds J. "It's all interconnected... like a spider web": a qualitative study of the meanings of food and healthy eating in an Indigenous community. *Int J Circumpolar Health.* (2019) 78:1648969. doi: 10.1080/22423982.2019.1648969
- 68. Hanemaayer R, Anderson K, Haines J, Lickers KRL, Lickers Xavier A, Gordon K, et al. Exploring the perceptions of and experiences with traditional foods among first nations female youth: a participatory photovoice study. *Int J Environ Res Public Health.* (2020) 17:2214. doi: 10.3390/ijerph17072214
- Kuhnlein HV, Receveur O. Dietary change and traditional food systems of indigenous peoples. *Annu Rev Nutr.* (1996) 16:417–42. doi: 10.1146/annurev.nu.16.070196.002221
- Moses SK, Whiting A V., Bratton GR, Taylor RJ, O'Hara TM. Inorganic nutrients and contaminants in subsistence species of Alaska: linking wildlife and human health. *Int J Circumpolar Health.* (2009) 68:53–74. doi: 10.3402/ijch.v68i1.18294

- Furgal CM, Powell S, Myers H. Digesting the message about contaminants and country foods in the Canadian north: a review and recommendations for future research and action. *Arctic.* (2005) 58:103–14. doi: 10.14430/ARCTIC404
- Krümmel EM, Gilman A, Krü EM. An update on risk communication in the Arctic. Int J Circumpolar Health. (2016) 75:33822. doi: 10.3402/ijch.v75.33822
- Pufall EL, Jones-Bitton A, McEwen SA, Brown TM, Edge VL, Rokicki J, et al. Prevalence of zoonotic anisakid nematodes in inuit-harvested fish and mammals from the Eastern Canadian Arctic. *Foodborne Pathog Dis.* (2012) 9:1002–9. doi: 10.1089/fpd.2012.1186
- 74. Cawthorn M. Meat Consumption From Stranded Whales and Marine Mammals in New Zealand: Public Health and Other Issues (1997). Wellington: Department of Conservation.
- Gordon JE, Frye WW, Babbott FL. Intestinal parasites of man in arctic greenland\*. Am J Trop Med Hyg. (1961) 10:185–90. doi: 10.4269/ajtmh.1961.10.185
- Chai JY, Jung BK. Fishborne zoonotic heterophyid infections: An update. *Food Waterbor Parasitol.* (2017) 8–9:33–63. doi: 10.1016/j.fawpar.2017.09.001
- Schurer JM, Gesy KM, Elkin BT, Jenkins EJ. Echinococcus multilocularis and *Echinococcus canadensis* in wolves from western Canada. *Parasitology*. (2014) 141:159–63. doi: 10.1017/S0031182013001716
- Rausch R. Cystic Echinococcosis in the Arctic and Sub-Arctic (2003). doi: 10.1017/s0031182003003664
- Jenkins EJ, Castrodale LJ, de Rosemond SJC, Dixon BR, Elmore SA, Gesy KM, et al. Tradition and transition: parasitic zoonoses of people and animals in Alaska, northern Canada, and Greenland. *Adv Parasitol.* (2013) 82:33–204. doi: 10.1016/B978-0-12-407706-5.00002-2
- Kutz SJ, Ducrocq J, Verocai GG, Hoar BM, Colwell DD, Beckmen KB, et al. Parasites in Ungulates of Arctic North America and Greenland. *Adv Parasitol.* (2012) 79:99–252. doi: 10.1016/B978-0-12-398457-9.00002-0
- Lantis M. Zoonotic diseases in the Canadian and Alaskan North. *Études Inuit Stud.* (1981) 5:83–107.
- Ross P, Olpinski S, Curtis M. Relationships between dietary practice and parasite zoonoses in Northern Québec Inuit communities. *Études Inuit Stud.* (1989) 13:33–47.
- Schurer JM, Pawlik M, Huber A, Elkin B, Cluff HD, Pongracz JD, et al. Intestinal parasites of gray wolves (*Canis lupus*) in northern and western Canada. *Can J Zool.* (2016) 94:643–50. doi: 10.1139/cjz-2016-0017
- Hueffer K, Parkinson AJ, Gerlach R, Berner J. Zoonotic infections in Alaska: disease prevalence, potential impact of climate change and recommended actions for earlier disease detection, research, prevention and control. *Int J Circumpolar Health.* (2013) 72. doi: 10.3402/ijch.v72i0.19562
- Zhang W, Wen H, Li J, Lin R, McManus DP. Immunology and immunodiagnosis of cystic echinococcosis: an update. *Clin Dev Immunol.* (2012) 2012:1–10. doi: 10.1155/2012/101895
- Kuchta R, Oros M, Ferguson J, Scholz T. Diphyllobothrium nihonkaiense tapeworm larvae in Salmon from North America. *Emerg Infect Dis.* (2017) 23:351–3. doi: 10.3201/eid2302.161026
- Salb AL, Barkema HW, Elkin BT, Thompson RCA, Whiteside DP, Black SR, et al. Dogs as sources and sentinels of parasites in humans and wildlife, Northern Canada. *Emerg Infect Dis.* (2008) 14:60–3. doi: 10.3201/eid1401.071113
- Bradbury RS, Panicker IS. Toxocara seroprevalence in Canada climate, environment and culture. *Adv Parasitol.* (2020) 109:291–316. doi: 10.1016/bs.apar.2020.03.003
- Wapenaar W, Barkema HW, O'Handley R. Fecal shedding of toxocara canis and other parasites in foxes and coyotes on prince Edward Island, Canada. J Wildl Dis. (2013) 49:394–7. doi: 10.7589/2012-04-113
- Rausch RL, Fay FH. Toxascaris leonina in rodents, and relationship to eosinophilia in a human population. *Comp Parasitol.* (2011) 78:236–44. doi: 10.1654/4504.1
- Lévesque B, Messier V, Bonnier-Viger Y, Couillard M, Côté S, Ward BJ, et al. Seroprevalence of zoonoses in a cree community (Canada). *Diagn Microbiol Infect Dis.* (2007) 59:283–6. doi: 10.1016/j.diagmicrobio.2007. 06.002

- Messier V, Lévesque B, Proulx J-F, Rochette L, Libman MD, Ward BJ, et al. Seroprevalence of toxoplasma gondii among nunavik inuit (Canada). Zoo Public Health. (2009) 56:188–97. doi: 10.1111/j.1863-2378.2008.01177.x
- Dubey J., Lewis B, Beam K, Abbitt B. Transplacental toxoplasmosis in a reindeer (*Rangifer tarandus*) fetus. Vet Parasitol. (2002) 110:131–5. doi: 10.1016/S0304-4017(02)00320-5
- 94. Sørensen KK, Mørk T, Sigurð\*ardóttir ÓG, Åsbakk K, Åkerstedt J, Bergsjø B, Fuglei E. Acute toxoplasmosis in three wild arctic foxes (Alopex lagopus) from Svalbard; one with co-infections of *Salmonella enteritidis* PT1 and *Yersinia pseudotuberculosis* serotype 2b. *Res Vet Sci.* (2005) 78:161–7. doi: 10.1016/j.rvsc.2004.07.010
- Sandström CAM, Buma AGJ, Hoye BJ, Prop J, van der Jeugd H, Voslamber B, et al. Latitudinal variability in the seroprevalence of antibodies against *Toxoplasma gondii* in non-migrant and Arctic migratory geese. *Vet Parasitol.* (2013) 194:9–15. doi: 10.1016/j.vetpar.2012.12.027
- Torrey EF, Yolken RH. Toxoplasma gondii and schizophrenia. Emerg Infect Dis. (2003) 9:1375–80. doi: 10.3201/eid0911.030143
- Elmore SA, Huyvaert KP, Bailey LL, Milhous J, Alisauskas RT, Gajadhar AA, et al. *Toxoplasma gondii* exposure in arctic-nesting geese: a multi-state occupancy framework and comparison of serological assays. *Int J Parasitol Parasites Wildl.* (2014) 3:147–53. doi: 10.1016/j.ijppaw.2014.05.005
- Gill CO. Microbiological conditions of meats from large game animals and birds. *Meat Sci.* (2007) 77:149–60. doi: 10.1016/j.meatsci.2007.03.007
- Jenkins EJ, Simon A, Bachand N, Stephen C. Wildlife parasites in a one health world. *Trends Parasitol.* (2015) 31:174–80. doi: 10.1016/j.pt.2015.01.002
- Massie GN, Ware MW, Villegas EN, Black MW. Uptake and transmission of Toxoplasma gondii oocysts by migratory, filter-feeding fish. *Vet Parasitol.* (2010) 169:296–303. doi: 10.1016/j.vetpar.2010.01.002
- 101. Chegeni TN, Sharif M, Sarvi S, Moosazadeh M, Montazeri M, Aghayan SA, et al. Is there any association between toxoplasma gondii infection and depression? A systematic review and meta-analysis. *PLoS ONE.* (2019) 14:e0218524. doi: 10.1371/journal.pone.0218524
- McIntyre L, Pollock SL, Fyfe M, Gajadhar A, Isaac-Renton J, Fung J, et al. Trichinellosis from consumption of wild game meat. *Can Med Assoc J.* (2007) 176:449–51. doi: 10.1503/cmaj.061530
- 103. Seymour J, Horstmann-Dehn L, Rosa C, Lopez JA. Occurrence and genotypic analysis of *Trichinella species* in Alaska marine-associated mammals of the bering and chukchi seas. *Vet Parasitol.* (2014) 200:153–64. doi: 10.1016/j.vetpar.2013.11.015
- 104. Forbes LB. The occurrence and ecology of trichinella in marine mammals. *Vet Parasitol.* (2000) 93:321–34. doi: 10.1016/S0304-4017(00)00349-6
- 105. Sharma R, Thompson P, Elkin B, Mulders R, Branigan M, Pongracz J, et al. Trichinella pseudospiralis in a wolverine (Gulo gulo) from the Canadian North. *Int J Parasitol Parasites Wildl.* (2019) 9:274–80. doi: 10.1016/j.ijppaw.2019.06.005
- 106. Curtis MA, Rau ME, Tanner CE, Prichard RK, Faubert GM, Olpinski S, et al. Parasitic zoonoses in relation to fish and wildlife harvesting by inuit communities in Northern Quebec, Canada. Arctic Me Res. (1988) 47 (Suppl. 1):693–6.
- 107. Olson ME, Roach PD, Stabler M, Chan W. Giardiasis in ringed seals from the Western Arctic. J Wildl Dis. (1997) 33:646–8. doi: 10.7589/0090-3558-33.3.646
- Leighton FA. Wildlife pathogens and diseases in Canada. In: Canadian Biodiversity: Ecosystem Status and Trends 2010, Technical Thematic Report No. 7. Ottawa, ON: Canadian Councils of Resource Ministers (2011). p. 53.
- Shury TK, Frandsen D, O'Brodovich L. Anthrax in free-ranging bison in the prince albert national park area of Saskatchewan in 2008. *Can Vet J.* (2009) 50:152–4.
- Hueffer K, Drown D, Romanovsky V, Hennessy T. Factors contributing to anthrax outbreaks in the circumpolar North. *Ecohealth.* (2020) 17:174–80. doi: 10.1007/s10393-020-01474-z
- 111. Timofeev V, Bahtejeva I, Mironova R, Titareva G, Lev I, Christiany D, et al. Insights from Bacillus anthracis strains isolated from permafrost in the tundra zone of Russia. *PLoS ONE.* (2019) 14:e0209140. doi: 10.1371/journal.pone.0209140
- 112. Tessaro SV. The existing and potential importance of brucellosis and tuberculosis in canadian wildlife: a review. *Can Vet J.* (1986) 27:119–24.

- 113. Shury TK, Nishi JS, Elkin BT, Wobeser GA. Tuberculosis and brucellosis in wood bison (bison bison athabascae) in northern Canada: a renewed need to develop options for future management. J Wildl Dis. (2015) 51:543–54. doi: 10.7589/2014-06-167
- 114. Tryland M. Zoonoses of arctic marine mammals. *Infect Dis Rev.* (2000) 2:55–64. Available online at: https://scholar.google.com/scholar\_lookup? title=Zoonoses+of+arctic+marine+mammals&author=Tryland,+M.& publication\_year=2000&journal=Infect.+Dis.+Rev.&volume=2&pages=55 %E2%80%9364
- Hauschild AHW, Gauvreau L. Food-borne botulism in Canada, 1971-84. Can Med Assoc J. (1985) 133:1141–6.
- Leclair D, Fung J, Isaac-Renton JL, Proulx J-F, May-Hadford J, Ellis A, et al. Foodborne botulism in Canada, 1985–2005. *Emerg Infect Dis.* (2013) 19:961–8. doi: 10.3201/eid1906.120873
- Shaffer N, Wainwright RB, Middaugh JP, Tauxe R V. Botulism among Alaska natives: the role of changing food preparation and consumption practices. *West J Med.* (1990) 153:390–3.
- 118. Food Safety Network. Safe Preparation and Storage of Aboriginal Traditional/Country Foods: A Review. Ottawa, ON: FSN (2008).
- 119. Franco DA. Botulism. In: Beran GW, Steele JH, Benenson AS, Torten M, Dreesen DW, Ristic M, Pier AC, editors. *Handbook of Zoonoses*. Boca Raton, FL: CRC Press (2009). p. 361–7. doi: 10.1201/9781003006107-27
- 120. Kutz S, Bollinger T, Branigan M, Checkley S, Davison T, Dumond M, et al. Erysipelothrix rhusiopathiae associated with recent widespread muskox mortalities in the Canadian Arctic. *Can Vet J.* (2015) 56:560–3.
- 121. Forde TL, Orsel K, Zadoks RN, Biek R, Adams LG, Checkley SL, et al. Bacterial genomics reveal the complex epidemiology of an emerging pathogen in arctic and boreal ungulates. *Front Microbiol.* (2016) 7:1759. doi: 10.3389/fmicb.2016.01759
- 122. Mavrot F, Orsel K, Hutchins W, Adams LG, Beckmen K, Blake JE, et al. Novel insights into serodiagnosis and epidemiology of *Erysipelothrix rhusiopathiae*, a newly recognized pathogen in muskoxen (Ovibos moschatus). *PLoS ONE*. (2020) 15:e0231724. doi: 10.1371/journal.pone.0231724
- 123. Chiu A, Goddard E, Parlee B. Caribou consumption in northern Canadian communities. J Toxicol Environ Heal Part A. (2016) 79:762–97. doi: 10.1080/15287394.2016.1174011
- 124. Groeschel M, Forde T, Turvey S, Joffe AM, Hui C, Naidu P, et al. An unusual case of *Erysipelothrix rhusiopathiae* prosthetic joint infection from the Canadian Arctic: whole genome sequencing unable to identify a zoonotic source. *BMC Infect Dis.* (2019) 19:282. doi: 10.1186/s12879-019-3913-7
- 125. Canadian Wildlife Health Cooperative. *CWHC Manuals and Guides: Safety Manual for Harvesters of Fish and Wildlife in Nunavut.* Saskatoon, SK: Canadian Cooperative Wildlife Health Centre (2012).
- 126. Washington State University IACUC. Zoonoses Associated with Fish | Institutional Animal Care and Use Committee. Washington State University. Available online at: https://iacuc.wsu.edu/zoonoses-associated-with-fish/ (accessed October 27, 2020).
- 127. Schwantje H, Macbeth JB, Kutz S, Elkin B. British Columbia Boreal Caribou Health Program Progress Report: Year 1 (November 1, 2013 – December 31, 2014), Victoria, BC: The British Columbia Oil and Gas Research and Innovation Society (OGRIS) (2014).
- Brooke CJ, Riley TV. Erysipelothrix rhusiopathiae: bacteriology, epidemiology and clinical manifestations of an occupational pathogen. *J Med Microbiol.* (1999) 48:789–99. doi: 10.1099/00222615-48-9-789
- 129. Kassim SS, Dibernardo A, Lindsay LR, Wuerz TC. Locally acquired leptospirosis in expedition racer, manitoba, Canda. *Emerg Infect Dis.* (2018) 24:2386–8. doi: 10.3201/eid2412.181015
- Smith AW, Brown RJ, Skilling DE, Bray HL, Keyes MC. Naturally-Occurring leptospirosis in Northern fur seals (*Callorhinus ursinus*). J Wildl Dis. (1977) 13:144–8. doi: 10.7589/0090-3558-13.2.144
- Messier V, Lévesque B, Proulx JF, Rochette L, Serhir B, Couillard M, et al. Seroprevalence of seven zoonotic infections in Nunavik, Quebec (Canada). *Zoo Public Health.* (2012) 59:107–17. doi: 10.1111/j.1863-2378.2011. 01424.x
- Larsson C, Comstedt P, Olsen B, Bergström S. First record of lyme disease borrelia in the Arctic. Vector Borne Zoo Dis. (2007) 7:453–6. doi: 10.1089/vbz.2006.0644

- Scott JD, Anderson JF, Durden LA. Widespread dispersal of borrelia burgdorferi–infected ticks collected from songbirds across Canada. J Parasitol. (2012) 98:49–59. doi: 10.1645/GE-2874.1
- Sundeep S, Cleeve V. Isolation of *Bisgaardia hudsonensis* from a seal bite. Case report and review of the literature on seal finger. *J Infect*. (2011) 63:86–8. doi: 10.1016/j.jinf.2011.04.006
- 135. Hansen MJ, Bertelsen MF, Christensen H, Bisgaard M, Bojesen AM. occurrence of pasteurellaceae bacteria in the oral cavity of selected marine mammal species. J Zoo Wildl Med. (2012) 43:828–35. doi: 10.1638/2011-0264R1.1
- 136. Blanchong JA, Samuel MD, Goldberg DR, Shadduck DJ, Lehr MA. Persistence of pasteurella multocida in wetlands following avian cholera outbreaks. J Wildl Dis. (2006) 42:33–9. doi: 10.7589/0090-3558-42.1.33
- Crawford RL, Blyde D, Blackall PJ, Forde BM, Beatson SA, Harris LM, et al. Novel insights into pasteurellosis in captive pinnipeds. *Vet Microbiol.* (2019) 231:232–7. doi: 10.1016/j.vetmic.2019.03.017
- Miernyk KM, Bruden D, Parkinson AJ, Hurlburt D, Klejka J, Berner J, et al. Human Seroprevalence to 11 zoonotic pathogens in the U.S. Arctic, Alaska. *Vector Borne Zoo Dis.* (2019) 19:563–75. doi: 10.1089/vbz.2018.2390
- 139. Duncan C, Savage K, Williams M, Dickerson B, Kondas A V., Fitzpatrick KA, et al. Multiple strains of coxiella burnetii are present in the environment of St. Paul Island, Alaska. *Transbound Emerg Dis.* (2013) 60:345–50. doi: 10.1111/j.1865-1682.2012.01353.x
- 140. Duncan C, Dickerson B, Pabilonia K, Miller A, Gelatt T. Prevalence of Coxiella burnetii and Brucella spp. in tissues from subsistence harvested northern fur seals (*Callorhinus ursinus*) of St. Paul Island, Alaska. *Acta Vet Scand.* (2014) 56:67. doi: 10.1186/s13028-014-0067-x
- 141. Westley BP, Horazdovsky RD, Michaels DL, Brown DR. Identification of a novel mycoplasma species in a patient with septic arthritis of the hip and seal finger. *Clin Infect Dis.* (2015) 62:491–3. doi: 10.1093/cid/civ875
- 142. Wobeser G, Campbell GD, Dallaire A, McBurney S. Tularemia, plague, yersiniosis, and tyzzer's disease in wild rodents and lagomorphs in Canada: a review. *Can Vet J.* (2009) 50:1251–6.
- 143. Barnett JEF, Woodley AJ, Hill TJ, Turner L. Conditions in grey seal pups (Halichoerus grypus) presented for rehabilitation. *Vet Rec.* (2000) 147:98– 104. doi: 10.1136/vr.147.4.98
- 144. Sunstrum J, Shoyinka A, Power LE, Maxwell D, Stobierski MG, Signs K, et al. Notes from the field: zoonotic mycobacterium bovis disease in deer hunters — michigan, 2002–2017. MMWR Morb Mortal Wkly Rep. (2019) 68:807–8. doi: 10.15585/mmwr.mm6837a3
- 145. Hansen CM, Vogler AJ, Keim P, Wagner DM, Hueffer K. Tularemia in Alaska, 1938 - 2010. Acta Vet Scand. (2011) 53:61. doi: 10.1186/1751-0147-53-61
- 146. Desvars A, Furberg M, Hjertqvist M, Vidman L, Sjöstedt A, Rydén P, et al. Epidemiology and ecology of tularemia in Sweden, 1984–2012. *Emerg Infect Dis.* (2015) 21:32–9. doi: 10.3201/eid2101.140916
- 147. Zellner B, Huntley JF. Ticks and tularemia: do we know what we don't know? Front Cell Infect Microbiol. (2019) 9:146. doi: 10.3389/fcimb.2019.00146
- 148. Posautz A, Gyuranecz M, Dénes B, Knauer F, Dier H, Walzer C. Tularemia possible increase and new risk factors. *Int J Infect Dis.* (2019) 79:60. doi: 10.1016/j.ijid.2018.11.155
- 149. Antonation KS, Shury TK, Bollinger TK, Olson A, Mabon P, Van Domselaar G, et al. Sylvatic plague in a canadian black-tailed prairie dog (*Cynomys ludovicianus*). J Wildl Dis. (2014) 50:699–702. doi: 10.7589/2013-08-215
- 150. Liccioli S, Stephens T, Wilson SC, McPherson JM, Keating LM, Antonation KS, et al. Enzootic maintenance of sylvatic plague in Canada's threatened black-tailed prairie dog ecosystem. *Ecosphere*. (2020) 11:e03138. doi: 10.1002/ecs2.3138
- Blake JE, McLean BD, Gunn A. Yersiniosis in free-ranging muskoxen on banks island, northwest territories, Canada. J Wildl Dis. (1991) 27:527–33. doi: 10.7589/0090-3558-27.4.527
- 152. Reed C, Bruden D, Byrd KK, Veguilla V, Bruce M, Hurlburt D, et al. Characterizing wild bird contact and seropositivity to highly pathogenic avian influenza A (H5N1) virus in Alaskan residents. *Influ Other Respi* Viruses. (2014) 8:516–23. doi: 10.1111/irv.12253
- USDA. USDA APHIS | USDA Confirms Highly Pathogenic H5N2 Avian Influenza in a Wild Mallard Duck in Alaska. (2016). Available online at:

https://www.aphis.usda.gov/aphis/newsroom/stakeholder-info/sa\_by\_date/ stakeholder-announcements-2016/sa-08/hpai-ak (accessed June 27, 2020).

- Nielsen O, Clavijo A, Boughen JA. Serologic evidence of influenza a infection in marine mammals of Arctic Canada. J Wildl Dis. (2001) 37:820–5. doi: 10.7589/0090-3558-37.4.820
- 155. Webster R, Geraci J, Petursson G, Skirnisson K. Conjunctivitis in human beings caused by influenza a virus of seals. N Engl J Med. (1981) 304:911. doi: 10.1056/NEJM198104093041515
- 156. Smith AW, Berry ES, Skilling DE, Barlough JE, Poet SE, Berke T, et al. *In vitro* isolation and characterization of a calicivirus causing a vesicular disease of the hands and feet. *Clin Infect Dis.* (1998) 26:434–9. doi: 10.1086/516311
- 157. Tomaselli M, Dalton C, Duignan PJ, Kutz S, van der Meer F, Kafle P, et al. Contagious ecthyma, rangiferine brucellosis, and lungworm infection in a muskox (*Ovibos moschatus*) from the Canadian Arctic, 2014. J Wildl Dis. (2016) 52:719–24. doi: 10.7589/2015-12-327
- Vikøren T, Lillehaug A, Åkerstedt J, Bretten T, Haugum M, Tryland M. A severe outbreak of contagious ecthyma (orf) in a free-ranging musk ox (*Ovibos moschatus*) population in Norway. *Vet Microbiol.* (2008) 127:10–20. doi: 10.1016/j.vetmic.2007.07.029
- 159. Tryland M, Beckmen KB, Burek-Huntington KA, Breines EM, Klein J. Orf virus infection in Alaskan mountain goats, dall's sheep, muskoxen, caribou and sitka black-tailed deer. Acta Vet Scand. (2018) 60:12. doi: 10.1186/s13028-018-0366-8
- 160. Charlton KM, Tabel H. *Epizootiology of Rabies in Canada*. Shepherd and Itoh (1976).
- 161. Hueffer K, Murphy M. Rabies in Alaska, from the past to an uncertain future. Int J Circumpolar Health. (2018) 77:1475185. doi: 10.1080/22423982.2018.1475185
- 162. Rausch RL. Some observations on rabies in alaska, with special reference to wild canidae. *J Wildl Manage*. (1958) 22:246–280.
- 163. Ducrocq J, Proulx J, Lévesque B, De Serres G, Wood H, Lemire M. Assessment of naturally acquired neutralizing antibodies against rabies lyssavirus in a subset of nunavik's inuit population considered most at risk of being exposed to rabid animals. *Zoo Public Health.* (2019) 66:533–9. doi: 10.1111/zph.12561
- 164. Mørk T, Prestrud P. Arctic rabies a review. Acta Vet Scand. (2004) 45:1–9. doi: 10.1186/1751-0147-45-1
- 165. Aenishaenslin C, Simon A, Forde T, Ravel A, Proulx J-F, Fehlner-Gardiner C, et al. Characterizing rabies epidemiology in remote inuit communities in Québec, Canada: a "one health" approach. *Ecohealth.* (2014) 11:343–55. doi: 10.1007/s10393-014-0923-1
- Rosatte RC. Rabies in Canada history, epidemiology and control. *Can Vet J.* (1988) 29:362–5.
- 167. Alaska Department of Fish and Game. Rabies in Alaska: Rabid Wolverine Found on North Slope. (2012). Available online at: http://www.adfg.alaska. gov/index.cfm?adfg=wildlifenews.view\_article&articles\_id=582 (accessed October 27, 2020).
- Odegaard O, Krogsrud J. Rabies in Svalbard: infection diagnosed in arctic fox, reindeer and seal. Vet Rec. (1981) 109:141-2. doi: 10.1136/vr.109. 7.141
- 169. Rausch R. Tropical problems in the Arctic: infectious and parasitic diseases, a common denominator. *Ind Trop Health.* (1974) 8:63–70.
- Tabel H, Corner AH, Webster WA, Casey CA. History and epizootiology of rabies in Canada. *Can Vet J.* (1974) 15:271–81.
- 171. Minuk G, Sun A, Sun D, Uhanova J, Nicolle L, Larke B, et al. Serological evidence of hepatitis e virus infection in an indigenous north american population. *Can J Gastroenterol.* (2007) 21:439–42. doi: 10.1155/2007/289059
- 172. Weger S, Elkin B, Lindsay R, Bollinger T, Crichton V, Andonov A. Hepatitis E virus seroprevalence in free-ranging deer in Canada. *Transbound Emerg Dis.* (2017) 64:1008–11. doi: 10.1111/tbed.12462
- Teshale EH, Hu DJ. Hepatitis E: epidemiology and prevention. World J Hepatol. (2011) 3:285–91. doi: 10.4254/wjh.v3.i12.285
- 174. Gozlan RE, Marshall WL, Lilje O, Jessop CN, Gleason FH, Andreou D. Current ecological understanding of fungal-like pathogens of fish: what lies beneath? *Front Microbiol.* (2014) 5:62. doi: 10.3389/fmicb.2014.00062
- 175. Güere ME, Våge J, Tharaldsen H, Benestad SL, Vikøren T, Madslien K, et al. Chronic wasting disease associated with prion protein gene (PRNP)

variation in Norwegian wild reindeer (*Rangifer tarandus*). Prion. (2020) 14:1-10. doi: 10.1080/19336896.2019.1702446

- 176. Waddell L, Greig J, Mascarenhas M, Otten A, Corrin T, Hierlihy K. Current evidence on the transmissibility of chronic wasting disease prions to humans-a systematic review. *Transbound Emerg Dis.* (2018) 65:37–49. doi: 10.1111/tbed.12612
- 177. Revich B, Tokarevich N, Parkinson AJ. Climate change and zoonotic infections in the Russian Arctic. *Int J Circumpolar Health.* (2012) 71:18792. doi: 10.3402/ijch.v71i0.18792
- Davidson R, Simard M, Kutz SJ, Kapel CMO, Hamnes IS, Robertson LJ. Arctic parasitology: why should we care? *Trends Parasitol.* (2011) 27:239–45. doi: 10.1016/j.pt.2011.02.001
- Ferguson MAD. rangiferine brucellosis on Baffin Island. J Wildl Dis. (1997) 33:536–43. doi: 10.7589/0090-3558-33.3.536
- 180. Goyette S, Cao Z, Libman M, Ndao M, Ward BJ. Seroprevalence of parasitic zoonoses and their relationship with social factors among the Canadian inuit in Arctic regions. *Diagn Microbiol Infect Dis.* (2014) 78:404–10. doi: 10.1016/j.diagmicrobio.2013.08.026
- 181. Campagna S, Lévesque B, Anassour-Laouan-Sidi E, Côté S, Serhir B, Ward BJ, et al. Seroprevalence of 10 zoonotic infections in 2 Canadian cree communities. *Diagn Microbiol Infect Dis.* (2011) 70:191–9. doi: 10.1016/j.diagmicrobio.2011.01.009
- Schurer JM, Quewezance H, Ndao M, Elmore SA, Jenkins EJ. People, pets, and parasites: one health surveillance in Southeastern Saskatchewan. Am J Trop Med Hyg. (2014) 90:1184–90. doi: 10.4269/ajtmh.13-0749
- 183. Schurer JM, Ndao M, Skinner S, Irvine J, Elmore SA, Epp T, et al. Parasitic zoonoses: one health surveillance in Northern Saskatchewan. *PLoS Negl Trop Dis.* (2013) 7:e2141. doi: 10.1371/journal.pntd.0002141
- 184. Jenkins EJ, Schurer JM, Gesy KM. Old problems on a new playing field: Helminth zoonoses transmitted among dogs, wildlife, and people in a changing northern climate. *Vet Parasitol.* (2011) 182:54–69. doi: 10.1016/j.vetpar.2011.07.015
- 185. Parkinson AJ, Berner J. Climate change and impacts on human health in the Arctic: an international workshop on emerging threats and the response of Arctic communities to climate change. *Int J Circumpolar Health.* (2009) 68:84–91. doi: 10.3402/ijch.v68i1.18295
- 186. Bradley MJ, Kutz SJ, Jenkins E, O'Hara TM. The potential impact of climate change on infectious diseases of Arctic fauna. *Int J Circumpolar Health*. (2005) 64:468–77. doi: 10.3402/ijch.v64i5.18028
- Gallana M, Ryser-Degiorgis MP, Wahli T, Segner H. Climate change and infectious diseases of wildlife: altered interactions between pathogens, vectors and hosts. *Curr Zool.* (2013) 59:427–37. doi: 10.1093/czoolo/59.3.427
- Elmore SA, Jenkins EJ, Huyvaert KP, Polley L, Root JJ, Moore CG. Toxoplasma gondii in circumpolar people and wildlife. *Vector Borne Zoo Dis.* (2012) 12:1–9. doi: 10.1089/vbz.2011.0705
- Proulx J, MacLean JD, Gyorkos TW, Leclair D, Richter A, Serhir B, et al. Novel prevention program for trichinellosis in inuit communities. *Clin Infect Dis.* (2002) 34:1508–14. doi: 10.1086/340342
- 190. Tomaselli M, Elkin B, Kutz S, Harms NJ, Nymo HI, Davison T, et al. A transdisciplinary approach to brucella in muskoxen of the western Canadian Arctic 1989–2016. *Ecohealth.* (2019) 16:488–501. doi: 10.1007/s10393-019-01433-3
- 191. Bodenstein B, Beckmen K, Sheffield G, Kuletz K, Van Hemert C, Berlowski B, et al. Avian cholera causes marine bird mortality in the bering sea of Alaska. *J Wildl Dis.* (2015) 51:934–7. doi: 10.7589/2014-12-273
- Bruce M, Zulz T, Koch A. Surveillance of infectious diseases in the Arctic. Public Health. (2016) 137:5–12. doi: 10.1016/j.puhe.2016.06.014
- 193. Tomaselli M, Kutz S, Gerlach C, Checkley S. Local knowledge to enhance wildlife population health surveillance: conserving muskoxen and caribou in the Canadian Arctic. *Biol Conserv.* (2018) 217:337–48. doi: 10.1016/j.biocon.2017.11.010
- 194. Olival KJ, Hosseini PR, Zambrana-Torrelio C, Ross N, Bogich TL, Daszak P. Host and viral traits predict zoonotic spillover from mammals. *Nature*. (2017) 546:646–50. doi: 10.1038/nature22975
- 195. Woolhouse M, Gaunt E. Ecological origins of novel human pathogens. Crit Rev Microbiol. (2007) 33:231–42. doi: 10.1080/10408410701 647560

- 196. Woolhouse M, Scott F, Hudson Z, Howey R, Chase-Topping M. Human viruses: discovery and emergence. *Philos Trans R Soc B Biol Sci.* (2012) 367:2864–71. doi: 10.1098/rstb.2011.0354
- 197. Brook RK, Kutz SJ, Millins C, Veitch AM, Elkin BT, Leighton T. Evaluation and delivery of domestic animal health services in remote communities in the Northwest territories: a case study of status and needs. *Can Vet J.* (2010) 51:1115–22.
- Palmater P. Priority Pandemic Response Needed for First Nations. Toronto, ON: Policy Options (2020).
- 199. Alexander KA, Carlson CJ, Lewis BL, Getz WM, Marathe MV, Eubank SG, et al. The ecology of pathogen spillover and disease emergence at the human-wildlife-environment interface. Adv Environ Microbiol. 5:267–98. doi: 10.1007/978-3-319-92373-4\_8
- Brierley L, Vonhof MJ, Olival KJ, Daszak P, Jones KE. Quantifying global drivers of zoonotic bat viruses: a process-based perspective. *Am Nat.* (2016) 187:E53-64. doi: 10.1086/684391
- 201. Kreuder Johnson C, Hitchens PL, Smiley Evans T, Goldstein T, Thomas K, Clements A, et al. Spillover and pandemic properties of zoonotic viruses with high host plasticity. *Sci Rep.* (2015) 5:14830. doi: 10.1038/srep14830
- 202. White RJ, Razgour O. Emerging zoonotic diseases originating in mammals: a systematic review of effects of anthropogenic land-use change. *Mamm Rev.* (2020) 50:336–52. doi: 10.1111/mam.12201
- 203. Borland S, Gracieux P, Jones M, Mallet F, Yugueros-Marcos J. Influenza A virus infection in cats and dogs: a literature review in the light of the "one health" concept. *Front Public Health.* (2020) 8:83. doi: 10.3389/fpubh.2020.00083
- 204. Heeney JL. Zoonotic viral diseases and the frontier of early diagnosis, control and prevention. J Intern Med. (2006) 260:399–408. doi: 10.1111/j.1365-2796.2006.01711.x
- 205. Wardeh M, Sharkey KJ, Baylis M. Integration of shared-pathogen networks and machine learning reveals the key aspects of zoonoses and predicts mammalian reservoirs. *Proc R Soc B Biol Sci.* (2020) 287:20192882. doi: 10.1098/rspb.2019.2882
- 206. Plowright RK, Parrish CR, McCallum H, Hudson PJ, Ko AI, Graham AL, et al. Pathways to zoonotic spillover. *Nat Rev Microbiol.* (2017) 15:502–10. doi: 10.1038/nrmicro.2017.45
- 207. Carver S, Mills JN, Parmenter CA, Parmenter RR, Richardson KS, Harris RL, et al. Toward a mechanistic understanding of environmentally forced zoonotic disease emergence: sin nombre hantavirus. *Bioscience*. (2015) 65:651–66. doi: 10.1093/biosci/biv047
- Fedeli C, Moreno H, Kunz S. Novel insights into cell entry of emerging human pathogenic arenaviruses. J Mol Biol. (2018) 430:1839–52. doi: 10.1016/j.jmb.2018.04.026
- 209. Sokolow SH, Nova N, Pepin KM, Peel AJ, Pulliam JRC, Manlove K, et al. Ecological interventions to prevent and manage zoonotic pathogen spillover. *Philos Trans R Soc B Biol Sci.* (2019) 374:20180342. doi: 10.1098/rstb.2018.0342
- 210. Plowright RK, Peel AJ, Streicker DG, Gilbert AT, McCallum H, Wood J, et al. Transmission or within-host dynamics driving pulses of zoonotic viruses in reservoir–host populations. *PLoS Negl Trop Dis.* (2016) 10:e0004796. doi: 10.1371/journal.pntd.0004796
- 211. Guth S, Visher E, Boots M, Brook CE. Host phylogenetic distance drives trends in virus virulence and transmissibility across the animal-human interface. *Philos Trans R Soc B Biol Sci.* (2019) 374:20190296. doi: 10.1098/rstb.2019.0296
- 212. Tatem AJ, Hay SI, Rogers DJ. Global traffic and disease vector dispersal. *Proc Natl Acad Sic USA*. (2006) 103:6242–7. doi: 10.1073/pnas.0508391103
- 213. Bajardi P, Poletto C, Ramasco JJ, Tizzoni M, Colizza V, Vespignani A. Human mobility networks, travel restrictions, and the global spread of 2009 H1N1 pandemic. *PLoS ONE.* (2011) 6:e16591. doi: 10.1371/journal.pone. 0016591
- 214. Lycett SJ, Duchatel F, Digard P. A brief history of bird flu. *Philos Trans R Soc B Biol Sci.* (2019) 374:20180257. doi: 10.1098/rstb.2018.0257
- Cleaveland S, Haydon DT, Taylor L. Overviews of pathogen emergence: which pathogens emerge, when and why? *Curr Top Microbiol Immunol.* (2007) 315:85–111. doi: 10.1007/978-3-540-70962-6\_5

- 216. Wang B, Harms D, Yang XL, Bock CT. Orthohepevirus C: an expanding species of emerging hepatitis e virus variants. *Pathogens*. (2020) 9:154. doi: 10.3390/pathogens9030154
- Washburne AD, Crowley DE, Becker DJ, Olival KJ, Taylor M, Munster VJ, et al. Taxonomic patterns in the zoonotic potential of mammalian viruses. *PeerJ*. (2018) 6:e5979. doi: 10.7717/peerj.5979
- 218. Luis AD, Hayman DTS, O'Shea TJ, Cryan PM, Gilbert AT, Pulliam JRC, et al. A comparison of bats and rodents as reservoirs of zoonotic viruses: are bats special? *Proc R Soc B Biol Sci.* (2013) 280:20122753. doi: 10.1098/rspb.2012.2753
- 219. Cleaveland S, Laurenson MK, Taylor LH. Diseases of humans and their domestic mammals: pathogen characteristics, host range and the risk of emergence. *Philos Trans R Soc London Ser B Biol Sci.* (2001) 356:991–9. doi: 10.1098/rstb.2001.0889
- Mollentze N, Streicker DG. Viral zoonotic risk is homogenous among taxonomic orders of mammalian and avian reservoir hosts. *Proc Natl Acad Sic USA*. (2020) 117:9423–30. doi: 10.1073/pnas.1919176117
- Achenbach JE, Bowen RA. Transmission of avian influenza A viruses among species in an artificial barnyard. *PLoS ONE.* (2011) 6:e17643. doi: 10.1371/journal.pone.0017643
- 222. Bosco-Lauth AM, Bowen RA, Root JJ. Limited transmission of emergent H7N9 influenza A virus in a simulated live animal market: do chickens pose the principal transmission threat? *Virology*. (2016) 495:161-66. doi: 10.1016/j.virol.2016.04.032
- 223. Gervasi SS, Burgan SC, Hofmeister E, Unnasch TR, Martin LB. Stress hormones predict a host superspreader phenotype in the West Nile virus system. *Proc R Soc B Biol Sci.* (2017) 284:20171090. doi: 10.1098/rspb.2017.1090
- 224. Hing S, Narayan EJ, Thompson RCA, Godfrey SS. The relationship between physiological stress and wildlife disease: consequences for health and conservation. *Wildl Res.* (2016) 43:51. doi: 10.1071/WR15183
- 225. Seeber PA, Quintard B, Sicks F, Dehnhard M, Greenwood AD, Franz M. Environmental stressors may cause equine herpesvirus reactivation in captive grévy's zebras (*Equus grevyi*). *PeerJ.* (2018) 6:e5422. doi: 10.7717/peerj.5422
- 226. Patz JA, Daszak P, Tabor GM, Aguirre AA, Pearl M, Epstein J, et al. Unhealthy landscapes: policy recommendations on land use change and infectious disease emergence. *Environ Health Perspect.* (2004) 112:1092–8. doi: 10.1289/ehp.6877
- 227. Pike BL, Saylors KE, Fair JN, LeBreton M, Tamoufe U, Djoko CF, et al. The origin and prevention of pandemics. *Clin Infect Dis.* (2010) 50:1636–40. doi: 10.1086/652860
- Wolfe ND, Daszak P, Kilpatrick AM, Burke DS. Bushmeat hunting, deforestation, and prediction of zoonotic disease. *Emerg Infect Dis.* (2005) 11:1822–7. doi: 10.3201/eid1112.040789
- 229. Valitutto MT, Aung O, Tun KYN, Vodzak ME, Zimmerman D, Yu JH, et al. Detection of novel coronaviruses in bats in Myanmar. *PLoS ONE*. (2020) 15:e0230802. doi: 10.1371/journal.pone.0230802
- 230. Purse B V., Darshan N, Kasabi GS, Gerard F, Samrat A, George C, et al. Predicting disease risk areas through co-production of spatial models: the example of kyasanur forest disease in india's forest landscapes. *PLoS Negl Trop Dis.* (2020) 14:e0008179. doi: 10.1371/journal.pntd.0008179
- 231. Tijjani M, Majid RA, Abdullahi SA, Unyah NZ. Detection of rodent-borne parasitic pathogens of wild rats in Serdang, Selangor, Malaysia: a potential threat to human health. *Int J Parasitol Parasites Wildl.* (2020) 11:174–82. doi: 10.1016/j.ijppaw.2020.01.008
- 232. Grimm K, Rivera NA, Fredebaugh-Siller S, Weng H-Y, Warner RE, Maddox CW, et al. Evidence Of leptospira serovars in wildlife and leptospiral DNA in water sources in a natural area in east-central Illinois, USA. J Wildl Dis. (2020) 56:316. doi: 10.7589/2019-03-069
- 233. Adesiyun AA, Knobel DL, Thompson PN, Wentzel J, Kolo FB, Kolo AO, et al. Sero-Epidemiological study of selected zoonotic and abortifacient pathogens in cattle at a wildlife-livestock interface in South Africa. *Vector-Borne Zoonotic Dis.* (2020) 20:258–67. doi: 10.1089/vbz.201 9.2519
- 234. Lönker NS, Fechner K, Abd El Wahed A. Horses as a crucial part of one health. *Vet Sci.* (2020) 7:28. doi: 10.3390/vetsci7010028

- Narat V, Alcayna-Stevens L, Rupp S, Giles-Vernick T. Rethinking humannonhuman primate contact and pathogenic disease spillover. *Ecohealth*. (2017) 14:840–50. doi: 10.1007/s10393-017-1283-4
- Reaser JK, Clark EE, Meyers NM. All creatures great and minute: a public policy primer for companion animal zoonoses. *Zoo Public Health.* (2008) 55:385–401. doi: 10.1111/j.1863-2378.2008.01123.x
- 237. Gibb R, Redding DW, Chin KQ, Donnelly CA, Blackburn TM, Newbold T, et al. Zoonotic host diversity increases in human-dominated ecosystems. *Nature*. (2020) 584:398–402. doi: 10.1038/s41586-020-2562-8
- Jones BA, Grace D, Kock R, Alonso S, Rushton J, Said MY, et al. Zoonosis emergence linked to agricultural intensification and environmental change. *Proc Natl Acad Sic USA*. (2013) 110:8399–404. doi: 10.1073/pnas.1208059110
- 239. Keesing F, Belden LK, Daszak P, Dobson A, Harvell CD, Holt RD, et al. Impacts of biodiversity on the emergence and transmission of infectious diseases. *Nature*. (2010) 468:647–52. doi: 10.1038/nature09575
- 240. Morand S, Owers K, Bordes F. Biodiversity and emerging zoonoses. In: Yamada A, Kahn LH, Kaplan B, Monath TP, Woodall J, Conti L, editors. *Confronting Emerging Zoonoses*. Tokyo: Springer (2014). p. 27–41. doi: 10.1007/978-4-431-55120-1\_3
- Murray KA, Daszak P. Human ecology in pathogenic landscapes: two hypotheses on how land use change drives viral emergence. *Curr Opin Virol.* (2013) 3:79–83. doi: 10.1016/j.coviro.2013.01.006
- Anthony SJ, Johnson CK, Greig DJ, Kramer S, Che X, Wells H, et al. Global patterns in coronavirus diversity. *Virus Evol.* (2017) 3:vex012. doi: 10.1093/ve/vex012
- 243. Horby PW, Hoa NT, Pfeiffer DU, Wertheim HFL. Drivers of emerging zoonotic infectious diseases. Confr Emerg Zoonoses. (2014) 13–26. doi: 10.1007/978-4-431-55120-1\_2
- Roberts MG, Heesterbeek JAP. Quantifying the dilution effect for models in ecological epidemiology. J R Soc Interface. (2018) 15:20170791. doi: 10.1098/rsif.2017.0791
- 245. Khalil H, Ecke F, Evander M, Magnusson M, Hörnfeldt B. Declining ecosystem health and the dilution effect. *Sci Rep.* (2016) 6:31314. doi: 10.1038/srep31314
- 246. Borremans B, Faust C, Manlove KR, Sokolow SH, Lloyd-Smith JO. Cross-species pathogen spillover across ecosystem boundaries: mechanisms and theory. *Philos Trans R Soc B Biol Sci.* (2019) 374:20180344. doi: 10.1098/rstb.2018.0344
- 247. Wilkinson DA, Marshall JC, French NP, Hayman DTS. Habitat fragmentation, biodiversity loss and the risk of novel infectious disease emergence. J R Soc Interface. (2018) 15:20180403. doi: 10.1098/rsif.2018.0403
- 248. Evans T, Olson S, Watson J, Gruetzmacher K, Pruvot M, Jupiter S, et al. Links between ecological integrity, emerging infectious diseases originating from wildlife, and other aspects of human health -an overview of the literature. *Wildl Conservat Soc.* (2020) 4:e303. doi: 10.13140/RG.2.2.34736.51205
- 249. Kutz SJ, Jenkins EJ, Veitch AM, Ducrocq J, Polley L, Elkin B, et al. The arctic as a model for anticipating, preventing, and mitigating climate change impacts on host-parasite interactions. *Vet Parasitol.* (2009) 163:217–28. doi: 10.1016/j.vetpar.2009.06.008
- 250. Kutz SJ, Hoberg EP, Molnár PK, Dobson A, Verocai GG. A walk on the tundra: host-parasite interactions in an extreme environment. *Int J Parasitol Parasites Wildl.* (2014) 3:198–208. doi: 10.1016/j.ijppaw.2014.01.002
- 251. Meltofte H, Barry T, Berteaux D, Bültmann H, Christiansen JS, Cook JA, et al. *Arctic Biodiversity Assessment: Status and Trends in Arctic Biodiversity; Synthesis.* Akureyri: Conservation of Arctic Flora and Fauna (2013).
- 252. Ruviniyia K, Abdullah DA, Sumita S, Lim YAL, Ooi PT, Sharma RSK. Molecular detection of porcine *Enterocytozoon bieneusi* infection in Peninsular Malaysia and epidemiological risk factors associated with potentially zoonotic genotypes. *Parasitol Res.* (2020) 119:1663–74. doi: 10.1007/s00436-020-06648-w
- Hueffer K, O'Hara TM, Follmann EH. Adaptation of mammalian hostpathogen interactions in a changing arctic environment. *Acta Vet Scand.* (2011) 53:17. doi: 10.1186/1751-0147-53-17
- 254. Nunavut Wildlife Management Board. *The Nunavut Wildlife Harvest Study Feb 2004.* Iqaluit, NU: Nunavut Wildlife Management Board (2004).
- 255. Aarluk Consulting. Review of Intersettlement Trade Opportunities for Arctic Food Products in Nunavut Final Report. (2005).

- Gombay N. The commoditization of country foods in nunavik: a comparative assessment of its development, applications, and significance. *Arctic.* (2005) 58:115–28. Available online at: https://www.jstor.org/stable/ 40512686
- 257. Turner NJ, Plotkin M, Kuhnlein H. Indigenous Peoples' food systems and well-being: interventions and policies for healthy communities. In: Kuhnlein HV, Erasmus B, Spigelski D, Burlingame B, editors. *Indigenous Peoples' Food Systems and Well-Being*. Rome: Food and Agriculture Organization (2013).
- Dowsley M. The value of a polar bear: evaluating the role of a multiple-use resource in the nunavut mixed economy. *Arctic Anthropol.* (2010) 47:39–56. doi: 10.1353/arc.0.0035
- 259. Natcher D. Subsistence and the social economy of canada's aboriginal north | northern review. North Rev. (2009) 30:83–98. Available online at: https:// thenorthernreview.ca/index.php/nr/article/view/6 (accessed April 12, 2021).
- Reeves RR. The commerce in maktaq at Arctic Bay, northern Baffin Island, NWT. Arctic Anthropol. (1993) 30:79–93. doi: 10.2307/403 16330
- 261. Damas J, Hughes GM, Keough KC, Painter CA, Persky NS, Corbo M, et al. Broad host range of SARS-CoV-2 predicted by comparative and structural analysis of ACE2 in vertebrates. *bioRxiv* [Preprint]. (2020). doi: 10.1101/2020.04.16.045302
- Damas D. Central eskimo systems of food sharing. *Ethnology*. (1972) 11:220. doi: 10.2307/3773217
- 263. Wenzel GW. Ningiqtuq: resource sharing and generalized reciprocity in Clyde River, Nunavut. Arctic Anthropol. (1995) 32:43–60. doi: 10.2307/40316386
- Collings P, Wenzel G, Condon RG. Modern food sharing networks and community integration in the central Canadian Arctic. Arctic. (1998) 51:301-14.
- 265. Kofinas G, BurnSilver SB, Magdanz J, Stotts R, Okada M. Subsistence Sharing Networks and Cooperation: Kaktovik, Wainwright, and Venetie, Alaska. BOEM Report 2015-023; AFES Report MP 2015-02. School of Natural Resources and Extension, University of Alaska Fairbanks.
- 266. Braund SR, Associates, Anchorage, Alaska. *Description of Alaskan Eskimo Bowhead Whale Subsistence Sharing Practices*. Final report submitted to the Alaska Eskimo Whaling Commission (2018).
- 267. Dombrowski K, Khan B, Channell E, Moses J, McLean K, Misshula E. Kinship, family, and exchange in a labrador inuit community. Arctic Anthropol. (2013) 50:89–104. doi: 10.3368/aa.50.1.89
- 268. Justice Law. *Nunavut Land Claims Agreement Act: Article 5.* Iqaluit, NU: Government of Canada (1993).
- 269. Bodenhorn B. It's good to know who your relatives are but we were taught to share with everybody: shares and sharing among inupiaq households. *Senri Ethnol Stud.* (2000) 53:27–60. Available online at: https://www. semanticscholar.org/paper/Meating-the-social%3A-sharing-atiku-euiashin-Castro/3e1a8517b622aa4b1f0ebb332f707424cc06791d
- Berkes F, Jolly D. Adapting to climate change: social-ecological resilience in a Canadian western arctic community. *Ecol Soc.* (2002) 5:18. doi: 10.5751/es-00342-050218
- 271. Bhawra J, Cooke MJ, Hanning R, Wilk P, Gonneville SLH. Community perspectives on food insecurity and obesity: focus groups with caregivers of métis and off-reserve first nations children. *Int J Equity Health.* (2015) 14:96. doi: 10.1186/s12939-015-0232-5
- 272. Dunn L, Gross P. Food-Sharing practices online in the facebook group cambridge bay news. *Études Inuit Stud.* (2019) 40:225–43. doi: 10.7202/1055440ar
- 273. BurnSilver S, Magdanz J, Stotts R, Berman M, Kofinas G. Are mixed economies persistent or transitional? Evidence using social networks from Arctic Alaska. Am Anthropol. (2016) 118:121–9. doi: 10.1111/aman.12447
- 274. CBC News. Kitikmeot Inuit Warned to Stop Selling Caribou Meat Online. CBC News (2014)
- 275. Ancelle T, De Bruyne A, Poisson DM, Dupouy-Camet J. Outbreak of trichinellosis due to consumption of bear meat from Canada, France, September 2005. Wkly Releas. (2005) 10:pii2809. doi: 10.2807/esw.10.41.02809-en
- 276. Parkinson AJ, Evengard B, Semenza JC, Ogden N, Børresen ML, Berner J, et al. Climate change and infectious diseases in the Arctic: establishment of

a circumpolar working group. Int J Circumpolar Health. (2014) 73:25163. doi: 10.3402/ijch.v73.25163

- Dudley JP, Hoberg EP, Jenkins EJ, Parkinson AJ. Climate change in the north american arctic: a one health perspective. *Ecohealth.* (2015) 12:713–25. doi: 10.1007/s10393-015-1036-1
- 278. Kafle P, Peller P, Massolo A, Hoberg E, Leclerc L-M, Tomaselli M, et al. Range expansion of muskox lungworms track rapid arctic warming: implications for geographic colonization under climate forcing. *Sci Rep.* (2020) 10:17323. doi: 10.1038/s41598-020-74358-5
- 279. Wallenhammar A, Lindqvist R, Asghar N, Gunaltay S, Fredlund H, Davidsson Å, et al. Revealing new tick-borne encephalitis virus foci by screening antibodies in sheep milk. *Parasit Vectors.* (2020) 13:185. doi: 10.1186/s13071-020-04030-4
- Brandt JP, Flannigan MD, Maynard DG, Thompson ID, Volney WJA. An introduction to Canada's boreal zone: ecosystem processes, health, sustainability, and environmental issues. *Environ Rev.* (2013) 21:207–226. doi: 10.1139/er-2013-0040
- 281. National Academies of Science Engineering Medicine. Understanding Responding to Global Health Security Risks from Microbial Threats in the Arctic. Everett L, editor. Washington, DC.: National Academies Press (2020). doi: 10.17226/25887
- Van Hemert C, Pearce JM, Handel CM. Wildlife health in a rapidly changing North: focus on avian disease. *Front Ecol Environ.* (2014) 12:548–56. doi: 10.1890/130291
- Gigante CM, Gao J, Tang S, McCollum AM, Wilkins K, Reynolds MG, et al. Genome of Alaskapox virus, a novel orthopoxvirus isolated from Alaska. *Viruses.* (2019) 11:708. doi: 10.3390/v11080708
- 284. Lee EC, Wada NI, Grabowski MK, Gurley ES, Lessler J. The engines of SARS-CoV-2 spread. *Science*. (2020) 370:406–7. doi: 10.1126/science.abd8755
- Semenza JC, Lindgren E, Balkanyi L, Espinosa L, Almqvist MS, Penttinen P, et al. Determinants and drivers of infectious disease threat events in Europe. *Emerg Infect Dis.* (2016) 22:581–9. doi: 10.3201/eid2204.151073
- 286. Alaska State Government. First case of COVID-19 confirmed by Alaska State Public Health Laboratory is an international resident – Mike Dunleavy. (2020). Available online at: https://gov.alaska.gov/newsroom/2020/03/ 12/first-case-of-covid-19-confirmed-by-alaska-state-public-healthlaboratory-is-an-international-resident/ (accessed October 1, 2020).
- Doxey M, Chrzaszcz L, Dominguez A, James RD. A forgotten danger. J Public Heal Manag Pract. (2019) 25:S7–10. doi: 10.1097/PHH.000000000001062
- 288. ProPublica. Remote Alaska Villages Isolate Themselves Further in Effort to Shield Against Coronavirus. Anchorage, AK: ProPublica (2020)
- 289. Gryseels S, De Bruyn L, Gyselings R, Calvignac-Spencer S, Leendertz FH, Leirs H. Risk of human-to-wildlife transmission of SARS-CoV-2. *Mam Rev.* (2021) 51:272–92. doi: 10.1111/mam.12225
- 290. Bossart GD, Schwartz JC. Acute necrotizing enteritis associated with suspected coronavirus infection in three harbor seals (*Phoca vitulina*). J Zoo Wildl Med. (1990) 21:84–7.
- 291. Mihindukulasuriya KA, Wu G, St. Leger J, Nordhausen RW, Wang D. Identification of a novel coronavirus from a beluga whale by using a panviral microarray. *J Virol.* (2008) 82:5084–8. doi: 10.1128/JVI.02722-07
- 292. Nollens HH, Wellehan JFX, Archer L, Lowenstine LJ, Gulland FMD. Detection of a respiratory coronavirus from tissues archived during a pneumonia epizootic in free-ranging pacific harbor seals *Phoca vitulina richardsii*. *Dis Aquat Organ*. (2010) 90:113–20. doi: 10.3354/dao02190
- 293. Woo PCY, Lau SKP, Lam CSF, Tsang AKL, Hui S-W, Fan RYY, et al. Discovery of a novel bottlenose dolphin coronavirus reveals a distinct species of marine mammal coronavirus in gammacoronavirus. *J Virol.* (2014) 88:1318–31. doi: 10.1128/JVI.02351-13
- 294. Messenger AM, Barnes AN, Gray GC. Reverse zoonotic disease transmission (zooanthroponosis): A systematic review of seldom-documented human biological threats to animals. *PLoS ONE.* (2014) 9:e89055. doi: 10.1371/journal.pone.0089055
- 295. Runge MC, Campbell Grant EH, Coleman JTH, Reichard JD, Gibbs SEJ, Cryan PM, et al. Assessing the Risks Posed by SARS-CoV-2 in and Via North American Bats—Decision Framing and Rapid Risk Assessment. Reston, VA: U.S. Geological Survey (2020). doi: 10.3133/ofr20201060
- 296. Canadian Wildlife Health Cooperative. Wildlife Health and COVID-19 in Canada: Bats. (2020). Available online At: http://cwhc-rcsf.ca/

docs/miscellaneous/CWHC Bat health and Covid-19 (Version 1.0 April 23 2020).pdf

- 297. Olival KJ, Cryan PM, Amman BR, Baric RS, Blehert DS, Brook CE, et al. Possibility for reverse zoonotic transmission of SARS-CoV-2 to freeranging wildlife: a case study of bats. *PLOS Pathog.* (2020) 16:e1008758. doi: 10.1371/journal.ppat.1008758
- CDC. Guidance to Reduce the Risk of SARS-CoV-2 Spreading between People and Wildlife. (2021). Available online at: https://www.cdc.gov/coronavirus/ 2019-ncov/your-health/wildlife.html (accessed March 17, 2021).
- 299. Oude Munnink BB, Sikkema RS, Nieuwenhuijse DF, Molenaar RJ, Munger E, Molenkamp R, et al. Jumping back and forth: anthropozoonotic and zoonotic transmission of SARS-CoV-2 on mink farms. *bioRxiv* [Preprint]. (2020). doi: 10.1101/2020.09.01.277152
- USDA. USDA. Confirms SARS-CoV-2 in Mink in Utah. (2020) Available online at: https://www.aphis.usda.gov/aphis/newsroom/stakeholder-info/ sa\_by\_date/sa-2020/sa-08/sare-cov-2-mink
- 301. Lanen JVM, Stevens C, Brown CL, Maracle B, Koster DS. Subsistence Land Mammal Harvests and Uses, Yukon Flats, Alaska: 2008–2010 Harvest Report and Ethnographic Update. Alaska Department of Fish and Game, Division of Subsistence Technical Paper No. 377. Anchorage (2012).
- 302. Wurtzer S, Marechal V, Mouchel J-M, Maday Y, Teyssou R, Richard E, et al. Evaluation of lockdown impact on SARS-CoV-2 dynamics through viral genome quantification in Paris wastewaters. *medRxiv* [*Preprint*]. (2020). doi: 10.1101/2020.04.12.20062679
- 303. Lodder W, de Roda Husman AM. SARS-CoV-2 in wastewater: potential health risk, but also data source. *Lancet Gastroenterol Hepatol.* (2020) 5:533– 4. doi: 10.1016/S2468-1253(20)30087-X
- 304. Carlson CJ, Albery GF, Merow C, Trisos CH, Zipfel CM, Eskew EA, et al. Climate change will drive novel cross-species viral transmission. *bioRxiv* [*Preprint*]. (2020). doi: 10.1101/2020.01.24.918755
- Polley L, Thompson RCA. Parasite zoonoses and climate change: molecular tools for tracking shifting boundaries. *Trends Parasitol.* (2009) 25:285–91. doi: 10.1016/j.pt.2009.03.007
- Reiling SJ, Dixon BR. Toxoplasma gondii: how an amazonian parasite became an inuit health issue. *Can Commun Dis Rep.* (2019) 45:183–90. doi: 10.14745/ccdr.v45i78a03
- 307. Prestrud KW, Åsbakk K, Fuglei E, Mørk T, Stien A, Ropstad E, Tryland M, et al. Serosurvey for Toxoplasma gondii in arctic foxes and possible sources of infection in the high Arctic of Svalbard. *Vet Parasitol.* (2007) 150:6–12. doi: 10.1016/j.vetpar.2007.09.006
- Zhang Y-Z, Wu W-C, Shi M, Holmes EC. The diversity, evolution and origins of vertebrate RNA viruses. *Curr Opin Virol.* (2018) 31:9–16. doi: 10.1016/j.coviro.2018.07.017
- Estrada-Peña A, Ostfeld RS, Peterson AT, Poulin R, de la Fuente J. Effects of environmental change on zoonotic disease risk: an ecological primer. *Trends Parasitol.* (2014) 30:205–14. doi: 10.1016/j.pt.2014.02.003
- 310. Shatilovich A V., Tchesunov A V., Neretina T V., Grabarnik IP, Gubin S V., Vishnivetskaya TA, et al. Viable nematodes from late pleistocene permafrost of the kolyma river lowland. *Dokl Biol Sci.* (2018) 480:100–2. doi: 10.1134/S0012496618030079
- 311. Legendre M, Bartoli J, Shmakova L, Jeudy S, Labadie K, Adrait A, et al. Thirty-thousand-year-old distant relative of giant icosahedral DNA viruses with a pandoravirus morphology. *Proc Natl Acad Sci USA*. (2014) 111:4274– 9. doi: 10.1073/pnas.1320670111
- 312. Ng TFF, Chen L-F, Zhou Y, Shapiro B, Stiller M, Heintzman PD, et al. Preservation of viral genomes in 700-y-old caribou feces from a subarctic ice patch. *Proc Natl Acad Sci USA*. (2014) 111:16842–7. doi: 10.1073/pnas.1410429111
- 313. Rydberg J, Klaminder J, Rosén P, Bindler R. Climate driven release of carbon and mercury from permafrost mires increases mercury loading to sub-arctic lakes. *Sci Total Environ.* (2010) 408:4778–83. doi: 10.1016/j.scitotenv.2010.06.056
- 314. Erskine AJ. Birds In Boreal Canada: Communities, Densities And Adaptations. Canadian Wildlife Services Report. Ottawa, ON: Ministry of Supply and Services (1977).
- 315. Virkkala R, Rajasärkkä A. Climate change affects populations of northern birds in boreal protected areas. *Biol Lett.* (2011) 7:395-8. doi: 10.1098/rsbl.2010.1052

- Post D, Taylor J, Kitchell J, Olson M, Schindler D, Herwig B. The role of migratory waterfowl as nutrient vectors in a managed wetland. *Conserv Biol.* (1998) 12:910–20.
- 317. Adalja AA, Watson M, Toner ES, Cicero A, Inglesby T V. *The Characteristics of Pandemic Pathogens*. Baltimore, MD: Johns Hopkins Bloomberg School of Public Health, Center for Health Security (2018).
- King M, Altdorff D, Li P, Galagedara L, Holden J, Unc A. Northward shift of the agricultural climate zone under 21st-century global climate change. *Sci Rep.* (2018) 8:7904. doi: 10.1038/s41598-018-26321-8
- Rohr JR, Barrett CB, Civitello DJ, Craft ME, Delius B, DeLeo GA, et al. Emerging human infectious diseases and the links to global food production. *Nat Sustain.* (2019) 2:445–56. doi: 10.1038/s41893-019-0293-3
- 320. Morand S. Emerging diseases, livestock expansion and biodiversity loss are positively related at global scale. *Biol Conserv.* (2020) 248:108707. doi: 10.1016/j.biocon.2020.108707
- Charron DF. Potential impacts of global warming and climate change on the epidemiology of zoonotic diseases in Canada. *Can J Public Health.* (2002) 93:334–35. doi: 10.1007/BF03404563
- 322. Hannah L, Roehrdanz PR, Fraser EDG, Donatti CI, Saenz L, Wright TM, et al. The environmental consequences of climate-driven agricultural frontiers. *PLoS ONE*. (2020) 15:e0228305. doi: 10.1371/journal.pone.0228305
- 323. Coker R, Rushton J, Mounier-Jack S, Karimuribo E, Lutumba P, Kambarage D, et al. Towards a conceptual framework to support one-health research for policy on emerging zoonoses. *Lancet Infect Dis.* (2011) 11:326–31. doi: 10.1016/S1473-3099(10)70312-1
- 324. Liverani M, Waage J, Barnett T, Pfeiffer DU, Rushton J, Rudge JW, et al. Understanding and managing zoonotic risk in the new livestock industries. *Environ Health Perspect.* (2013) 121:873–7. doi: 10.1289/ehp. 1206001
- 325. Sonne C, Letcher RJ, Jenssen BM, Desforges J-P, Eulaers I, Andersen-Ranberg E, et al. A veterinary perspective on one health in the arctic. *Acta Vet Scand.* (2017) 59:84. doi: 10.1186/s13028-017-0353-5
- 326. Hosseini PR, Mills JN, Prieur-Richard A-H, Ezenwa VO, Bailly X, Rizzoli A, et al. Does the impact of biodiversity differ between emerging and endemic pathogens? The need to separate the concepts of hazard and risk. *Philos Trans R Soc B Biol Sci.* (2017) 372:20160129. doi: 10.1098/rstb.201 6.0129
- 327. Jones BA, Betson M, Pfeiffer DU. Eco-social processes influencing infectious disease emergence and spread. *Parasitology*. (2017) 144:26–36. doi: 10.1017/S0031182016001414
- 328. Canadian Population Health Initiative. *Improving the Health of Canadians*. Ottawa, ON: Canadian Population Health Initiative (2004).
- 329. Munster VJ, Bausch DG, de Wit E, Fischer R, Kobinger G, Muñoz-Fontela C, et al. Outbreaks in a rapidly changing central Africa — lessons from ebola. N Engl J Med. (2018) 379:1198–201. doi: 10.1056/NEJMp1807691
- Bird BH, Mazet JAK. Detection of emerging zoonotic pathogens: an integrated one health approach. Annu Rev Anim Biosci. (2018) 6:121–39. doi: 10.1146/annurev-animal-030117-014628
- Kutz S, Tomaselli M. "Two-eyed seeing" supports wildlife health. Science. (2019) 364:1135–7. doi: 10.1126/science.aau6170
- 332. Canadian Wildlife Health Cooperative. CWHC-RCSF Canadian Wildlife Health Cooperative/Réseau Canadian Pour la Santé de la Faune. Available online at: http://www.cwhc-rcsf.ca/ (accessed October 26, 2020).
- Stephen C. The pan-canadian approach to wildlife health. Can Vet J. (2019) 60:145–6.
- National Wildlife Health Center. USGS National Wildlife Health Center. Available online at: https://www.usgs.gov/centers/nwhc (accessed May 21, 2020).
- 335. Carlsson A, Veitch A, Popko R, Behrens S, Studies SKO. Monitoring wildlife health for conservation food security in the Canadian Arctic-a case study from the Sahtu settlment area in the Northwest Territories. In: Cork S, Hall D, Liljebjelke K, editors. One Health Case Studies: Addressing Complex Problems in a Changing World. Sheffield: 5m Publishing (2016).
- McKay AJ, Johnson CJ. identifying effective and sustainable measures for community-based environmental monitoring. *Environ Manage*. (2017) 60:484–95. doi: 10.1007/s00267-017-0887-3
- 337. Mosites E, Lujan E, Brook M, Brubaker M, Roehl D, Tcheripanoff M, et al. Environmental observation, social media, and one health action: a

description of the local environmental observer (LEO) network. *One Health.* (2018) 6:29–33. doi: 10.1016/j.onehlt.2018.10.002

- 338. Peacock SJ, Mavrot F, Tomaselli M, Hanke A, Fenton H, Nathoo R, et al. Linking co-monitoring to co-management: bringing together local, traditional, and scientific knowledge in a wildlife status assessment framework. Arct Sci. (2020) 6:247–66. doi: 10.1139/as-2019-0019
- 339. Huntington HP. Using traditional ecological knowledge in science: methods and applications. *Ecol Appl.* (2000) 10:1270-4. doi: 10.2307/26 41282
- 340. Tagalik S. Inuit Qaujimajatuqangit: The Role of Indigenous Knowledge Supporting Wellness in Inuit Communities in Nunavut. British Columbia, CA: National Collaborating Center for Aboriginal Health, Prince George (2010).
- Larrat S, Simard M, Lair S, Bélanger D, Proulx JF. From science to action and from action to science: the nunavik trichinellosis prevention program. *Int J Circumpolar Health.* (2012) 71:18595. doi: 10.3402/ijch.v71i0.18595
- 342. Stimmelmayr R, Sheffield G, Garlich-Miller J, Metcalf V, Goodwin J, Raverty S. The alaska Northern Pinniped unusual mortality event: 2011–2012. In Proceedings of the 44th Conference of the International Association for Aquatic Animal Medicine. Sausalito, CA (2012).
- 343. Indigenous Leadership Initiative. Indigenous Guardians Program Indigenous Leadership Initiative. Available online at: https://www. ilinationhood.ca/our-work/guardians/ (accessed October 26, 2020).
- 344. Parlee BL, Goddard E, First Nation ŁKD, Smith M. Tracking change: traditional knowledge and monitoring of wildlife health in Northern Canada. *Hum Dimens Wildl.* (2014) 19:47–61. doi: 10.1080/10871209.2013. 825823
- 345. Robards MD, Huntington HP, Druckenmiller M, Lefevre J, Moses SK, Stevenson Z, et al. Understanding and adapting to observed changes in the Alaskan Arctic: actionable knowledge co-production with alaska native communities. *Deep Sea Res Part II Top Stud Oceanogr.* (2018) 152:203–13. doi: 10.1016/j.dsr2.2018.02.008
- 346. Ostertag SK, Loseto LL, Snow K, Lam J, Hynes K, Gillman DV. "That's how we know they're healthy": the inclusion of traditional ecological knowledge in beluga health monitoring in the Inuvialuit Settlement Region. *Arct Sci.* (2018) 4:292–320. doi: 10.1139/as-2017-0050
- 347. Moore SE, Hauser DDW. Marine mammal ecology and health: finding common ground between conventional science and indigenous knowledge to track arctic ecosystem variability. *Environ Res Lett.* (2019) 14:075001. doi: 10.1088/1748-9326/ab20d8
- 348. Thompson S, Thapa K, Whiteway N. Sacred harvest, sacred place: mapping harvesting sites in wasagamack first nation. J Agric Food Syst Community Dev. (2019) 9:1–29. doi: 10.5304/jafscd.2019.0 9B.017
- 349. National Research Council (US) Committee on Achieving Sustainable Global Capacity for Surveillance Response to Emerging Zoonotic Diseases. 3. Drivers of zoonotic diseases. In: Keusch GT, Pappaioanou M, Gonzalez MC, Scott KA, editors. Sustaining Global Surveillance Response to Emerging Zoonotic Diseases. Washing, DC: National Academies Press (2009).
- LEO Network. About LEO Network LEO Network. Available online at: https://www.leonetwork.org/en/docs/about/about (accessed October 26, 2020).
- 351. MacDonald JP, Ford JD, Willox AC, Ross NA. A review of protective factors and causal mechanisms that enhance the mental health of indigenous circumpolar youth. *Int J Circumpolar Health.* (2013) 72:21775. doi: 10.3402/ijch.v72i0.21775
- 352. O'Hara TM, Hoekstra PF, Hanns C, Backus SM, Muir DCG. Concentrations of selected persistent organochlorine contaminants in store-bought foods from Northern Alaska. Int J Circumpolar Health. (2005) 64:303–13. doi: 10.3402/ijch.v64i4.18008
- 353. DiNicolantonio JJ, O'Keefe JH. The introduction of refined carbohydrates in the Alaskan Inland inuit diet may have led to an increase in dental caries, hypertension and atherosclerosis. *Open Hear.* (2018) 5:e000776. doi: 10.1136/openhrt-2018-000776
- 354. Willows N, Johnson-Down L, Kenny T-A, Chan HM, Batal M. Modelling optimal diets for quality and cost: examples from inuit and first nations communities in Canada. *Appl Physiol Nutr Metab.* (2019) 44:696–703. doi: 10.1139/apnm-2018-0624

- Kuhnlein H V, Chan HM. Environment and contaminants in traditional food systems of Northern indigenous peoples. *Annu Rev Nutr.* (2000) 20:595–626. doi: 10.1146/annurev.nutr.20.1.595
- 356. Chatwood S, Francois P, Baker GR, Eriksen AMA, Hansen KL, Eriksen H, et al. E. Indigenous values and health systems stewardship in circumpolar countries. *Int J Env Res Public Heal.* (2017) 14:1462. doi: 10.3390/ijerph14121462
- 357. Duhaime G, Bernard N. (eds). Arctic Food Security. Edmonton, AB: Canadian Circumpolar Institute Press (2008). p. 344. Available online at: https://www.researchgate.net/publication/276232632\_DUHAIME\_Gerard\_ and\_Nick\_BERNARD\_eds\_2008\_Arctic\_Food\_Security\_Edmonton\_ Canadian\_Circumpolar\_Institute\_Press\_344\_pages (accessed April 13, 2021).
- 358. Ban NC, Frid A, Reid M, Edgar B, Shaw D, Siwallace P. Incorporate Indigenous perspectives for impactful research and effective management. Nat Ecol Evol. (2018) 2:1680–3. doi: 10.1038/s41559-018-0706-0
- 359. Shea E. The concepts of health of young Inuit adolescents. *Arctic Med Res.* (1991) 309–12.

**Conflict of Interest:** DJ was employed by the company Nyati Health Consulting, British Columbia, Canada. DL was employed by Nunavut Tunngavik Inc., Ottawa, Canada.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer AP declared a past co-authorship with one of the authors SO to the handling Editor.

Copyright © 2021 Keatts, Robards, Olson, Hueffer, Insley, Joly, Kutz, Lee, Chetkiewicz, Lair, Preston, Pruvot, Ray, Reid, Sleeman, Stimmelmayr, Stephen and Walzer. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.