



Associations of *MAP2K3* Gene Variants With Superior Memory in SuperAgers

Matthew J. Huentelman¹, Ignazio S. Piras¹, Ashley L. Siniard¹, Matthew D. De Both¹, Ryan F. Richholt¹, Chris D. Balak¹, Pouya Jamshidi², Eileen H. Bigio^{2,3}, Sandra Weintraub^{2,4}, Emmaleigh T. Loyer², M.-Marsel Mesulam^{2,5}, Changiz Geula² and Emily J. Rogalski^{2,4}* for The Alzheimer's Disease Neuroimaging Initiative[†]

OPEN ACCESS

Edited by:

Ashok Kumar, University of Florida, United States

Reviewed by:

Jorge L. Del-Aguila, Washington University in St. Louis, United States Manav Kapoor, Icahn School of Medicine at Mount Sinai, United States

> *Correspondence: Emily J. Rogalski erogalski@gmail.com

[†]Data used in preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu). As such, the investigators within the ADNI contributed to the design and implementation of ADNI and/or provided data but did not participate in analysis or writing of this report. A complete listing of ADNI investigators can be found at: http://adni.loni.usc.edu/ wp-content/uploads/ how_to_apply/ADNI_ Acknowledgement_List.pdf

> Received: 30 January 2018 Accepted: 07 May 2018 Published: 29 May 2018

Citation:

Huentelman MJ, Piras IS, Siniard AL, De Both MD, Richholt RF, Balak CD, Jamshidi P, Bigio EH, Weintraub S, Loyer ET, Mesulam M-M, Geula C and Rogalski EJ for The Alzheimer's Disease Neuroimaging Initiative (2018) Associations of MAP2K3 Gene Variants With Superior Memory in SuperAgers. Front. Aging Neurosci. 10:155. doi: 10.3389/fnagi.2018.00155 ¹Neurogenomics Division, Translational Genomics Research Institute, Phoenix, AZ, United States, ²Cognitive Neurology & Alzheimer's Disease Center, Northwestern University Feinberg School of Medicine (NU FSM), Chicago, IL, United States, ³Department of Pathology, Northwestern University Feinberg School of Medicine (NU FSM), Chicago, IL, United States, ⁴Department of Psychiatry and Behavioral Sciences, Northwestern University Feinberg School of Medicine (NU FSM), Chicago, IL, United States, ⁵Department of Neurology, Northwestern University Feinberg School of Medicine (NU FSM), Chicago, IL, United States

Introduction: SuperAgers are adults age 80+ with episodic memory performance that is *at least* as good as that of average middle-aged adults. Understanding the biological determinants of SuperAging may have relevance to preventing age-related cognitive decline and dementia. This study aimed to identify associations between genetic variations and the SuperAging phenotype using Whole Exome Sequencing (WES).

Methods: Sequence Kernel Association Combined (SKAT-C) test was conducted at the gene level including both rare and common variants in 56 SuperAgers and 22 cognitively-average controls from the Alzheimer's disease Neuroimaging Initiative (ADNI).

Results: The SuperAging phenotype was associated with variants in the Mitogen-Activated Protein Kinase Kinase 3 (*MAP2K3*) gene. Three single nucleotide polymorphisms (SNPs) contributed to the significance (rs2363221 [intron 1], rs2230435 [exon 5], rs736103 [intron 7]).

Conclusions: MAP2K3 resides in a biological pathway linked to memory. It is in a signaling cascade associated with beta-amyloid mediated apoptosis and has enriched expression in microglia. This preliminary work suggests MAP2K3 may represent a novel therapeutic target for age-related memory decline and perhaps Alzheimer's disease (AD).

Keywords: aging, Alzheimer's disease (AD), Alzheimer's dementia, Alzheimer's, successful aging, genetics, whole exome sequencing, cognition, episodic memory

INTRODUCTION

Average episodic memory capacity is significantly higher in populations of 50–60 year olds than in populations of 80+ year olds. We defined SuperAgers as individuals 80 or older with episodic memory at least average for cognitively average individuals in their 50 s and 60 s. Thus, SuperAgers have superior memory capacity for age and are potentially resistant to age-related decline

1

TABLE 1 | Demographic and neuropsychological performance.

| Demographics | SuperAgers ^a n = 56 | Cognitively average controls ^{a,b} n = 22 |
|---|-----------------------------------|---|
| Age (years) | 83.0 ± 3.3 | 82.8 ± 2.6 |
| | [80–95] | [79–89] |
| Education (years) | 15.8 ± 2.3 | $17.7 \pm 1.8^{*}$ |
| | [12–20] | [14–20] |
| Sex Men : Women | 17:39 | 19:3* |
| Neuropsychological test performance | | |
| Rey Auditory Verbal Learning Test (RAVLT) | 11.6 ± 1.7 | $4.4 \pm 1.4^{*}$ |
| Delay Raw Score (out of 15) | [9–15] | [3–7] |
| Category fluency (animals) | 22.8 ± 4.9 | 19.9 ± 3.6 |
| | [12–33] | [15–27] |
| Boston Naming Test (BNT; 30-item) | 28.4 ± 1.8^{b} | 28.4 ± 1.7 |
| | [21–30] | [25–30] |
| Trail Making Test B | 88.6 ± 34.5 | 84.0 ± 32.9 |
| | [38–231] | [41–170] |
| Mini Mental Status Exam (MMSE) | 29.2 ± 1.1 | 29.0 ± 1.0 |
| | [25-30] | [27-30] |

Data are provided as Means \pm Standard Deviations and [Ranges]. ^aData were from non-Hispanic individuals. ^bThe control genotypes used in the preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu). ^bBNT scores for the SuperAging group were based on n = 55. *Indicates significant differences between the groups. RAVLT: 15-item list learning test of episodic memory with possible scores ranging from 0 to 15. Category fluency: Semantic fluency task in which individuals say as many items from a given category (i.e., animals) as they can in 1 min. BNT: measure of object naming with possible scores ranging from 0 to 30. Trail Making Test B: timed test that assesses executive attention. Test discontinued at 300 s if not completed. MMSE: brief measurement of cognitive functioning with possible scores ranging from 0 to 30.

(Harrison et al., 2012; Rogalski et al., 2013; Gefen et al., 2015; Cook et al., 2017). Previous work has shown that SuperAgers are less vulnerable to age-related cortical atrophy (Harrison et al., 2012; Rogalski et al., 2013; Cook et al., 2017), and have thicker anterior cingulate cortex when compared to their "cognitively average" peers (Harrison et al., 2012; Rogalski et al., 2013). Postmortem studies showed more von Economo neurons and less Alzheimer pathology in the anterior cingulate of SuperAgers compared to their peers (Rogalski et al., 2013; Gefen et al., 2015). Here we investigated the association between genetic variation and the SuperAging phenotype using Whole Exome Sequencing (WES).

MATERIALS AND METHODS

Analyses included 56 SuperAgers from the Northwestern SuperAging Program. Genotypes were obtained from 22 cognitively-average controls from the Alzheimer's disease neuroimaging initiative (ADNI) Whole Genome Sequencing (WGS) database¹. The ADNI was launched in 2003 as a publicprivate partnership, led by Principal Investigator Michael W. Weiner, MD. The primary goal of ADNI has been to test whether serial magnetic resonance imaging (MRI), positron emission tomography (PET), other biological markers, and clinical and neuropsychological assessment can be combined to measure the progression of mild cognitive impairment (MCI) and early Alzheimer's disease (AD).

SuperAgers met previously established criteria (Rogalski et al., 2013). Briefly, they were adults \geq 80 years scoring at or above average normative values for adults 50–65 years on an episodic memory test and at least average-for-age in other

¹http://adni.loni.usc.edu/

cognitive domains. This study was carried out in accordance with the recommendations of the Institutional Review Board. The protocol was approved by the Institutional Review Board at Northwestern University and at all of the ADNI sites. All subjects gave written informed consent in accordance with the Declaration of Helsinki.

Controls were adults \geq 80 years, scoring within the averagefor-age normative range on cognitive tests, including episodic memory.

SuperAging DNA samples were extracted from whole blood using the QIAamp DNA Blood Midi Kits (Qiagen, USA) and analyzed with WES, using the Illumina technology. Libraries were prepared with either the TruSeq 62Mb Exome Enrichment Kit or the Nextera 62Mb Rapid Capture Expanded Exome Kit (Illumina, Inc., San Diego, CA, USA), following manufacturers' protocols. Exome libraries were sequenced by 100-bp paired-end sequencing on a HiSeq 2500 System (Illumina, Inc., San Diego, CA, USA).

Control WGS data were used to run association analysis and to compare to SuperAger genetic profiles. Control DNA samples were obtained from National Cell Repository for Alzheimer's Disease (NCRAD) for validation of candidate variants by Sanger sequencing.

Demographic variables were compared using the Wilcoxon-rank sum test or Fisher's Exact Test. FASTQs were processed with the open-source bcbio-nextgen pipeline², using bwa-mem 0.7.10 for reads alignment, Picard 1.96 to mark duplicates, and GATK Haplotype Caller 3.2 for variants call. The resulting VCF file was filtered to include: Single Nucleotide Variants (SNVs), single nucleotide polymorphisms (SNPs) call rate >85%, Depth >7, and Quality Score >30.

²https://github.com/chapmanb/bcbio-nextgen



FIGURE 1 | Manhattan plots showing the results of the Sequence Kernel Association Combined (SKAT-C) analysis between SuperAgers and Alzheimer's disease neuroimaging initiative (ADNI) controls, without covariates (**A**) and with covariates (**B**) included in the model. The red line indicates the significance threshold after Bonferroni correction (p = 3.09E-06), whereas the blue line indicates the nominal significance threshold (p = 0.05). (**C**) Allele frequencies for the single nucleotide polymorphisms (SNPs) in the Mitogen-Activated Protein Kinase Kinase 3 (MAP2K3) gene in SuperAging, Control and ExAC cohorts (European non-Finnish population sample).

VCF files containing WGS data for controls (n = 22) were downloaded from ADNI³. We applied the same filters used in the SuperAgers, and excluded SNPs in Hardy-Weinberg disequilibrium (p < 1.0E-05). Then, the same SNPs obtained in the SuperAgers were selected after filtering. After combining datasets, a final SNP call rate filter was applied (>95%), and variants were annotated using the "*BumpHunter*" R-package (Jaffe et al., 2012) referencing to the GRCh37/Hg19 assembly.

Population stratification was assessed with the λ inflation factor using the R-package *snpStats* (Clayton, 2015). Association analysis was performed at the gene-level using the Sequence

³http://adni.loni.usc.edu/

Kernel Association Combined test (SKAT-C; Ionita-Laza et al., 2013) including SNPs located in the promoter and the gene body. Briefly, the SKAT-C test computes a multi-marker statistic assigning the same weight to common and rare variants. Rare variants were defined using the Minimum Allele Frequency (MAF) cutoff computed as MAF = 1/SQRT(2N), where N is total sample size. Analyses using SKAT R-package, accounted for significant covariates. Significant signals were further checked by visual inspection of BAM file reads. *P*-values were Bonferroni corrected. Linkage Disequilibrium between the candidate SNPs was computed using PLINK 1.9 (Purcell et al., 2007).

The frequency of candidate variants was compared with the European (non-Finnish) population data in the $ExAC^4$ and the gnomAD databases⁵.

Sanger sequencing of candidate variants was completed on five SuperAgers (selected to include the different genotypes) and 22 Controls. The PCR reactions were performed in 25 µL with AmpliTaq Gold system according to protocol (Applied Biosystems GmbH, Weiterstadt, Germany). Each sample underwent PCR separately for SNPs rs2363221, rs2230435, rs736103 using the primers (5'-3') Fwd: CTGCTATGAGGCTGGAGTATG+Rev:ACCTCATGCCTTGG GATTT, Fwd:CTGTCATGAGTGTGGGGTGTT+Rev:ACCTTCC GTGTCAACTTTAGG and Fwd:TGTCTGCACCAGGATTGT TAG+Rev:CACCTTTCTCAGGACAGAAGTC, respectively. PCR reactions were purified with Agencourt AMPure XP PCR purification (Beckman-Coulter) at a sample/bead ratio of 1:1.8 and sequenced. Samples with ambiguous electropherograms were sequenced twice.

RESULTS

Table 1 provides group demographics and neuropsychological performance data. The groups were similar in age (p = 0.56), but differed in sex (p = 8.8E-06) and education level (p = 2.8E-03).

For WES analysis, after quality controls filtering, there were 145,891 SNPs shared between SuperAgers and Controls. Intergenic SNPs were removed, obtaining 104,731 SNPs in 16,195 genes. Population stratification was not detected ($\lambda = 1.013$). After removing one false positive by visual inspection of BAM file reads, a significant association for Mitogen-Activated Protein Kinase Kinase 3 (*MAP2K3*) gene was detected in two SKAT-C association analyses conducted with sex and education as covariates and without covariates (Q = 37, $p_{adj} = 0.018$; Q = 73.4, $p_{adj} = 0.0011$, respectively; **Figure 1**). Three SNPs contributed to the significance (rs2363221, rs2230435, rs736103) located in intron 1, exon 5 and intron 7, respectively. SNPs rs2230435 and rs736103 showed high Linkage Disequilibrium ($r^2 = 0.790$), whereas other pairwise r^2 values between SNPs showed $r^2 < 0.30$ for all comparisons.

SuperAger and Control allele frequencies were also compared with frequencies in the European ExAC cohort and gnomAD database. ExAC and gnomAD cohort allele frequencies were comparable to Controls, but not SuperAgers (**Figure 1C**).

Using Sanger sequencing, the three variants were successfully validated in 22 ADNI controls and five SuperAgers. The genotype in one sample for the rs2363221 SNP could not be determined.

DISCUSSION

The SuperAging phenotype was associated with variants in the *MAP2K3* gene. MAP2K3 (or MKK3) is a dual specificity kinase activated by environmental and mitogenic stress residing in a biological pathway linked to memory (Peng et al., 2010) and within a signaling cascade associated with beta-amyloid mediated

⁴ExAC.broadinstitute.org

⁵http://gnomad.broadinstitute.org/

apoptosis (Zhou et al., 2014). *MAP2K3* is located in chr17:21, 187, 968-21, 218, 552 (GRCh37/hg19) including 11 introns and 12 exons. It is upstream of MAPK14/p38-MAPK, an enzyme altered in AD and a recent therapeutic focus with inhibitors demonstrating positive effects on AD relevant pathology in mouse models (Alam and Scheper, 2016).

MAP2K3 expression is enriched in microglia in mouse cortex (Zhang et al., 2014) and is part of the signaling cascade leading to inflammation (Swaroop et al., 2016), suggesting this variant may be contributing to SuperAging via brain immune system regulation. The role of microglia in AD has recently gained interest due to the association of rare variants in triggering receptor expressed on myeloid cells 2 (TREM2) with significantly altered AD risk (Bellenguez et al., 2017; Sims et al., 2017). The *MAP2K3* gene is significantly upregulated in expression in the middle temporal gyrus of AD patients compared to controls, indicating this gene is regulated to some extent in response to AD.

In view of the small study sample sizes, strict statistical approaches were used to identify the association within *MAP2K3*. Furthermore, data from ADNI were used as controls as these individuals have rigorous phenotype information. We also examined allele frequencies of associated variants in ExAC the largest public human genomic database. Of note, ExAC allele frequencies closely matched our control frequencies. It is possible SuperAger allele frequencies could drift closer to Controls with larger sample sizes; therefore, independent replication is needed.

Our current hypothesis is MAP2K3 activity in the SuperAger brain is slightly decreased due to genetic factors present from birth. None of the identified SuperAger variants are predicted to fully impair MAP2K3 activity. This is notable because MAP2K3 signaling is likely critical for normal cell physiology. This alteration may result in lowered p38-MAPK activity in neuronal cells and may reduce inflammation mediated by microglia (Swaroop et al., 2016), the cell type with the highest known MAP2K3 expression in brain. Further work is necessary to fully explain mechanistic changes and how they are realized in each cell type in SuperAgers. Based on our findings, we postulate MAP2K3 inhibitors may represent a novel therapeutic strategy for enhanced cognition and resistance to AD.

AUTHOR CONTRIBUTIONS

Conceptualization of this study was primarily provided by ER, MH, IP, CG and M-MM. Data were collected by ER, EB, SW, EL and M-MM or provided by (ADNI). Data analysis was completed by MH, IP, AS, MD, RR, CB, PJ, EL, M-MM, ER and CG. All authors participated in the writing or revision of this manuscript: MH, IP, AS, MD, RR, CB, PJ, EB, SW, EL, M-MM, CG and ER.

ACKNOWLEDGMENTS

This work was funded in part by grants from the National Institutes of Health, including R01 AG045571 and P30 AG13854 from the National Institute on Aging as well as by a grant from the Davee Foundation. Samples from the National Cell Repository for Alzheimer's Disease (NCRAD), which receives government support under a cooperative agreement grant (U24 AG21886) awarded by the National Institute of Aging (NIA), were used in this study. We thank contributors who collected samples used in this study, as well as participants, whose help and participation made this work possible. Data collection and sharing for this project was funded by the Alzheimer's Disease Neuroimaging Initiative (ADNI; National Institutes of Health Grant U01 AG024904) and DOD ADNI (Department of Defense award number W81XWH-12-2-0012). ADNI is funded by the National Institute on Aging, the National Institute of Biomedical Imaging and Bioengineering, and through generous contributions from the following: AbbVie, Alzheimer's Association; Alzheimer's Drug Discovery Foundation; Araclon Biotech; BioClinica, Inc.; Biogen; Bristol-Myers Squibb Company; CereSpir, Inc.; Cogstate; Eisai Inc.; Elan Pharmaceuticals, Inc.; Eli Lilly and Company; EuroImmun;

REFERENCES

- Alam, J., and Scheper, W. (2016). Targeting neuronal MAPK14/p38α activity to modulate autophagy in the Alzheimer disease brain. *Autophagy* 12, 2516–2520. doi: 10.1080/15548627.2016.1238555
- Bellenguez, C., Charbonnier, C., Grenier-Boley, B., Quenez, O., Le Guennec, K., Nicolas, G., et al. (2017). Contribution to Alzheimer's disease risk of rare variants in TREM2, SORL1, and ABCA7 in 1779 cases and 1273 controls. *Neurobiol. Aging* 59, 220.e1–220.e9. doi: 10.1016/j.neurobiolaging.2017. 07.001
- Clayton, D. (2015). snpStats: SnpMatrix and XSnpMatrix classes and methods. R package version 1.20.0.
- Cook, A. H., Sridhar, J., Ohm, D., Rademaker, A., Mesulam, M. M., Weintraub, S., et al. (2017). Rates of cortical atrophy in adults 80 years and older with superior vs. average episodic memory. *JAMA* 317, 1373–1375. doi: 10.1001/jama. 2017.0627
- Gefen, T., Peterson, M., Papastefan, S. T., Martersteck, A., Whitney, K., Rademaker, A., et al. (2015). Morphometric and histologic substrates of cingulate integrity in elders with exceptional memory capacity. *J. Neurosci.* 35, 1781–1791. doi: 10.1523/JNEUROSCI.2998-14.2015
- Harrison, T. M., Weintraub, S., Mesulam, M. M., and Rogalski, E. (2012). Superior memory and higher cortical volumes in unusually successful cognitive aging. J. Int. Neuropsychol. Soc. 18, 1081–1085. doi: 10.1017/s13556177120 00847
- Ionita-Laza, I., Lee, S., Makarov, V., Buxbaum, J. D., and Lin, X. (2013). Sequence kernel association tests for the combined effect of rare and common variants. *Am. J. Hum. Genet.* 92, 841–853. doi: 10.1016/j.ajhg.2013. 04.015
- Jaffe, A. E., Murakami, P., Lee, H., Leek, J. T., Fallin, M. D., Feinberg, A. P., et al. (2012). Bump hunting to identify differentially methylated regions in epigenetic epidemiology studies. *Int. J. Epidemiol.* 41, 200–209. doi: 10.1093/ije/ dyr238
- Peng, X. Q., Damarla, M., Skirball, J., Nonas, S., Wang, X. Y., Han, E. J., et al. (2010). Protective role of PI3-kinase/Akt/eNOS signaling in mechanical stress through inhibition of p38 mitogen-activated protein kinase in mouse lung. *Acta Pharmacol. Sin.* 31, 175–183. doi: 10.1038/aps.2009.190

F. Hoffmann-La Roche Limited and its affiliated company Genentech, Inc.; Fujirebio Europe; GE Healthcare; IXICO Ltd.; Janssen Alzheimer Immunotherapy Research and Development, LLC.; Johnson and Johnson Pharmaceutical Research and Development LLC.; Lumosity; Lundbeck; Merck and Co., Inc.; Meso Scale Diagnostics, LLC.; NeuroRx Research; Neurotrack Technologies; Novartis Pharmaceuticals Corporation; Pfizer Inc.; Piramal Imaging; Servier; Takeda Pharmaceutical Company; and Transition Therapeutics. The Canadian Institutes of Health Research is providing funds to support ADNI clinical sites in Canada. Private sector contributions are facilitated by the Foundation for the National Institutes of Health (www.fnih.org). The guarantee organization is the Northern California Institute for Research and Education, and the study is coordinated by the Alzheimer's Therapeutic Research Institute at the University of Southern California. ADNI data are disseminated by the Laboratory for Neuro Imaging at the University of Southern California.

- Purcell, S., Neale, B., Todd-Brown, K., Thomas, L., Ferreira, M. A., Bender, D., et al. (2007). PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am. J. Hum. Genet.* 81, 559–575. doi: 10.1086/519795
- Rogalski, E. J., Gefen, T., Shi, J., Samimi, M., Bigio, E., Weintraub, S., et al. (2013). Youthful memory capacity in old brains: anatomic and genetic clues from the Northwestern SuperAging Project. J. Cogn. Neurosci. 25, 29–36. doi:10.1162/jocn_a_00300
- Sims, R., van der Lee, S. J., Naj, A. C., Bellenguez, C., Badarinarayan, N., Jakobsdottir, J., et al. (2017). Rare coding variants in PLCG2, ABI3, and TREM2 implicate microglial-mediated innate immunity in Alzheimer's disease. *Nat. Genet.* 49, 1373–1384. doi: 10.1038/ng.3916
- Swaroop, S., Sengupta, N., Suryawanshi, A. R., Adlakha, Y. K., and Basu, A. (2016). HSP60 plays a regulatory role in IL-1β-induced microglial inflammation via TLR4–p38 MAPK axis. J. Neuroinflammation 13:27. doi: 10.1186/s12974-016-0486-x
- Zhang, Y., Chen, K., Sloan, S. A., Bennett, M. L., Scholze, A. R., O'Keeffe, S., et al. (2014). An RNA-sequencing transcriptome and splicing database of glia, neurons, and vascular cells of the cerebral cortex. *J. Neurosci.* 34, 11929–11947. doi: 10.1523/JNEUROSCI.1860-14.2014
- Zhou, F., Xu, Y., and Hou, X. Y. (2014). MLK3-MKK3/6–P38MAPK cascades following N-methyl-D-aspartate receptor activation contributes to amyloidβ peptide-induced apoptosis in SH-SY5Y cells. *J. Neurosci. Res.* 92, 808–817. doi: 10.1002/jnr.23354

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Huentelman, Piras, Siniard, De Both, Richholt, Balak, Jamshidi, Bigio, Weintraub, Loyer, Mesulam, Geula and Rogalski for The Alzheimer's Disease Neuroimaging Initiative. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.