



## Inflammation: Bridging Age, Menopause and APOEε4 Genotype to Alzheimer's Disease

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Neuro-inflammatory processes that contribute to development of Alzheimer's are evident early in the latent prodromal phase and worsen during the course of the disease. Despite substantial mechanistic and clinical evidence of inflammation, therapeutic approaches targeting inflammation have failed to alter the course of the disease. Disparate results from epidemiological and clinical trials targeting inflammation, highlight the complexity of the inflammatory process. Herein we review the dynamics of the inflammatory process across aging, midlife endocrine transitions, and the APOE<sub>E</sub>4 genotype and their contribution to progression of Alzheimer's disease (AD). We discuss the chronic inflammatory processes that are activated during midlife chronological and endocrine aging, which ultimately limit the clearance capacity of microglia and lead to immune senescence. Aging, menopause, and APOE<sub>E4</sub> combine the three hits of a compromised bioenergetic system of menopause with the chronic low grade innate inflammation of aging with the APOEε4 dyslipidemia and adaptive immune response. The inflammatory immune response is the unifying factor that bridges across each of the risk factors for AD. Immune system regulators that are specific to stage of disease and inflammatory phenotype would provide a therapeutic strategy to disconnect the bridge that drives disease. Outcomes of this analysis provide plausible mechanisms underlying failed clinical trials of anti-inflammatory agents in Alzheimer's patients. Further, they highlight the need for stratifying AD clinical trial cohorts based on inflammatory phenotype. Combination therapies that include targeted use of anti-inflammatory agent's specific to the immune phenotype are considered.

Keywords: inflammation, Alzheimer's disease, menopause, APOEe4, aging

## INTRODUCTION

Alzheimer's disease (AD) is characterized by an extended prodromal phase of typically 10–20 years duration prior to clinical manifestation of cognitive decline (Amieva et al., 2008). The prodromal phase of AD consists of both- pre-stage symptoms and mild cognitive impairment (MCI) (Wilson et al., 2011). Clinical studies have shown that the prodromal phase is characterized by metabolic dysfunction, amyloid- $\beta$  (A $\beta$ ) deposition in the brain, mild to moderate cognitive dysfunction, and chronic low-grade inflammation (Habeck et al., 2012; Olsson et al., 2013; Brinkmalm et al., 2014;

#### **OPEN ACCESS**

#### Edited by:

Anne Eckert, Universitäre Psychiatrische Kliniken Basel, Switzerland

#### Reviewed by:

Walter A. Rocca, Mayo Clinic, United States Amandine Grimm, Universität Basel, Switzerland

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Received: 08 August 2018 Accepted: 18 September 2018 Published: 09 October 2018

#### Citation:

Mishra A and Brinton RD (2018) Inflammation: Bridging Age, Menopause and APOEc4 Genotype to Alzheimer's Disease. Front. Aging Neurosci. 10:312. doi: 10.3389/fnagi.2018.00312 Wirz et al., 2014; Rajan et al., 2015; Mosconi et al., 2017a,b). These hallmark pathologies have aided in the development of biomarkers predictive of disease pathogenesis. In some populations, A $\beta$ 42 is one of the first biomarkers to appear in the cerebrospinal fluid (CSF) (Jack et al., 2013; Brinkmalm et al., 2014; Calsolaro and Edison, 2016). Despite the recent and widespread recognition of neuroinflammation in the pathogenesis of AD, there has been sparse expansion of inflammation-based biomarkers and preventive strategies. Understanding the dynamic interplay between inflammation and the risk factors such as age, APOE genotype, and endocrine transition states, can aid in this process. This review addresses the interaction between inflammation and risk factors key to the pathogenesis of AD. Strategies to specifically target these processes are also considered.

#### EARLY ACTIVATION OF INFLAMMATION AND RISK FOR ALZHEIMER'S DISEASE

Substantial evidence documents reactive microgliosis around plaque deposition and is now a hallmark of AD pathology (McGeer et al., 1988; Mattiace et al., 1990; Xiang et al., 2006). Reactive microgliosis and neuroinflammation in AD patients is considered a consequence of A $\beta$  plaque deposition (McGeer et al., 1987). Microgliosis in AD is evidenced both microscopically and biochemically with increased levels of the proinflammatory cytokines including tumor necrosis factor- $\alpha$  (TNF $\alpha$ ), IL-6, and IL-1 $\beta$  (Itagaki et al., 1989; Dickson et al., 1993; Ferretti and Cuello, 2011; Eikelenboom et al., 2012; Latta et al., 2014). While the inflammatory response to A $\beta$  plaque deposition is irrefutable, it is a late stage response in the inflammatory cascade. Indicators of earlier inflammatory responses are apparent in multiple conditions that are risk factors for later development of AD.

Associations between the occurrence of systemic infections and chronic inflammatory conditions with Alzheimer's disease, suggests an active participation of inflammation in early stages of disease development. Patients with higher erythrocyte sedimentation rate (ESR), which is a clinical indicator of nonspecific inflammation, are at greater risk of developing AD (Li et al., 2012). This is further corroborated by epidemiological studies that show that patients who suffer from chronic periodontal infection (Van Den Heuvel et al., 2007) and HIV have a higher risk of developing AD (Stanley et al., 1994; Alisky, 2007; Xu and Ikezu, 2009; Chakradhar, 2018). Recapitulating the clinical effect, Krstic et al. (2012) established an animal model that displayed AD like neuropathology by inducing chronic inflammation prenatally using a viral antigen, thereby showing that chronic inflammation potentiates the development of AD. Traumatic brain injury (TBI) also increases the risk of developing AD. Lesions that develop during TBI lead to an acute inflammatory response that includes microglial activation to facilitate debris removal and neuroprotection (Van Den Heuvel et al., 2007; Breunig et al., 2013; Habib et al., 2014). Incomplete resolution of the acute inflammatory response in TBI, however, is often followed by hypoxia and oxidative stress, which leads to

the chronic activation of microglia and the release of neurotoxic proinflammatory cytokines (Van Den Heuvel et al., 2007; Breunig et al., 2013; Habib et al., 2014).

Chronic inflammatory conditions such as autoimmune disorders alter the risk of development of dementia. A recent study found patients admitted to the hospital for an autoimmune disorder have greater risk for subsequent hospitalization due to dementia (Wotton and Goldacre, 2017). This association was particularly significant for multiple sclerosis and systemic lupus erythematosus for AD. While patients with rheumatoid arthritis (RA) had a reduced risk of developing Alzheimer's disease, they had an increased risk of vascular dementia (Wotton and Goldacre, 2017). Multiple studies indicate that AD incidence is lower in persons with RA (Policicchio et al., 2017). Some attribute this reduction in incidence to the regular use of non-steroidal anti-inflammatory drugs (NSAIDs) (McGeer et al., 1996; Etminan et al., 2003). An alternative mechanism involves upregulation of granulocyte macrophage-colony stimulating factor (GM-CSF) with a probable gain of function in myeloid cells, thus enabling effective debris clearance is also hypothesized to reduce the incidence of AD in RA patients (McGeer et al., 1996; Boyd et al., 2010). In mice, increased levels of GM-CSF (both intrahippocampal and subcutaneous administration) significantly reduced amyloidosis and reversed cognitive impairment (McGeer et al., 1996; Boyd et al., 2010). More recent findings indicate that RA and risk of AD can be stratified based on treatment. Case-controlled study conducted on electronic medical records from 8.5 million commercially insured adults, indicate that RA patients treated with an anti-TNFa therapy, etanercept, had a lower risk of AD whereas those on other anti-inflammatory agents had increased risk of AD (Chou et al., 2016).

Disparate results from epidemiological studies and randomized clinical trials highlight the complexity of response to anti-inflammatory agents (Thal et al., 2005). Epidemiological analyses indicated that long-term NSAIDs users have a lower risk of developing AD (McGeer et al., 1996; Vlad et al., 2008). Based on epidemiological findings, a clinical study - ADAPT (Alzheimer's Disease Anti-Inflammatory Prevention Trial) was conducted in cognitively intact elderly individuals with a family history of AD. In this trial, the selective cyclooxygenase-2 (COX-2) inhibitor Celecoxib and non-selective COX inhibitor Naproxen, were used as preventive therapies. The trial was discontinued 15 months after randomization, due to the increased cardiovascular risk of these therapies. On extended follow-up after 7 years, treatment with celecoxib or naproxen for 1-3 years did not prevent cognitive decline (Group, 2007, 2008; Breitner et al., 2011; Alzheimer's Disease Anti-inflammatory Prevention Trial Research Group, 2013). Collectively, these findings indicate that timing of NSAID treatment for chronic inflammatory conditions are a critical factor impacting the efficacy of NSAID therapy to prevent or delay progression of Alzheimer's disease (McGeer et al., 1996; Vlad et al., 2008).

Discrepancies between the epidemiological and clinical trial findings indicate the need for greater refinement in considering patient populations and anti-inflammatory therapies. Elucidating the inflammatory phenotype which emerges during the

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progression of AD requires consideration of the triggers that initiate chronic inflammation. In the sections below, we address how age, endocrine status, and APOE genotype impact inflammatory processes across AD progression from risk to late stage disease.

#### AGE-RELATED NEUROINFLAMMATION

Aging has a broad systems-level effect on human biology, which is evidenced by alterations in physiological function, metabolism, cognition, and inflammation (Smith et al., 2005). The effects of aging on immune responses are extensive and complex. In some individuals, adaptive immune responses will decline with age, whereas, others will experience aberrant immune responses leading to autoimmune disorders (Goronzy and Weyand, 2012; Vadasz et al., 2013; Fougère et al., 2017). Aging is associated with accumulation of oxidative stress and DNA damage and chronic low-grade inflammation (Cui et al., 2012). Though the effect of aging on cognitive function is variable, age remains the greatest risk factor for Alzheimer's in which inflammation is an early and persistent hallmark of the disease (von Bernhardi et al., 2015).

Central nervous system microglia plays a prominent role in innate immunity. Microglia constantly conducts surveillance of brain parenchyma to detect foreign pathogens and clear debris (Streit et al., 2004; von Bernhardi et al., 2015). Microglia detects and responds to a broad range of triggers including TBI, infections and damage associated molecular patterns (DAMPs). Reactive oxygen species (ROS), extracellular DNA and ATP all act as DAMPs (von Bernhardi et al., 2015; Gulke et al., 2018).

Innate immune responses by microglia are phenotypically typified by enlargement of the cell body, and molecularly by the upregulation of CD68, major histocompatibility complex-II (MHC-II) along with costimulatory molecules and secretion of pro and anti-inflammatory cytokines (Kim and Joh, 2006). The onset of innate immune responses leads to activation of the adaptive immune response. The innate activation of the adaptive immune response results in infiltration of peripheral immune cells, particularly T cell invasion of the brain (Kim and Joh, 2006). Together the innate and adaptive immune responses create the chronic low-grade inflammation typical of aging (Kim and Joh, 2006; see **Figures 1, 2**).

Microglial phenotype is dynamic. It changes with age and is typified by both homeostatic and disease phenotypes (Butovsky et al., 2014; Keren-Shaul et al., 2017; Krasemann et al., 2017). The telomerase deficient accelerated aging mouse model exhibits reduced microglial numbers and deficient morphological and cellular processes (Khan et al., 2015). Further, microglial response to activation is stage of development dependent. Production of cytokines and trophic factors by microglia increases linearly with age (Lai et al., 2013). Microglia isolated from younger mice (2–4 months) exhibit a lower expression of proinflammatory cytokines: TNF $\alpha$ , IL-6, and IL-1 $\beta$  than older mice (Helenius et al., 1996; Crain et al., 2013; Latta et al., 2015). On activation by ATP, microglia isolated from neonatal rats and 13–15 month



old adult rats have a more robust inflammatory response exhibited by upregulation of nitric oxide,  $TNF\alpha$ , and brain derived neurotrophic factor (BDNF) in comparison to microglia derived from younger animals (2–11 months) (Lai et al., 2013). Aging distinctly affects migratory function of microglia; younger microglia on encountering activating signals exhibit an increase in motility and rapid extension of ramifications, whereas older microglia are less dynamic. Transcriptomic studies corroborate a reduction in migratory ability of microglia with age as age affects actin cytoskeleton reogranization which is vital in both phagocytosis and migration (Damani et al., 2011; Orre et al., 2014).

Comparison of the transcriptomic profiles of young and aged microglia revealed that microglial receptors (Trem2c, P2yr12, P2yr13, and Adora) involved in recognizing DAMPs such as, oxidized low-density lipoprotein, mitochondrial DNA, extracellular ATP decreased with age (Orre et al., 2014). In contrast, the expression of receptors that recognize pathogens and microbes (Tlr2, CD74, Ltf, Clec7a, Cxcl16, and Ifitm6) increases with age (Orre et al., 2014). Age-related changes in



(mt-DNA), ATP. State of chronic low-grade inflammation leads to recruitment of adaptive responses, in turn affecting glial metabolism.

microglial transcriptome are not ubiquitous as the expression of phagocytic receptors (Cd14, Cd68, Cd11b, and ICAM) remained unaltered in aging (Hickman et al., 2013; Smith and Dragunow, 2014). However, activation of microglial phagocytosis is diminished in aging. Studies characterizing microglial phagocytic capacity, report a reduction in phagocytosis with age which is especially evident following activation (Li M.D. et al., 2015; Ritzel et al., 2015). These findings indicate that despite stable expression of phagocytic receptors, the functional capacity of microglia decreases with age (Li M.D. et al., 2015; Ritzel et al., 2015). For example, the ability of microglia to phagocytose AB is affected by age, with microglia isolated from postnatal animals effectively phagocytosing Aß fibrils, whereas adult microglia lose their capacity to do so (Floden and Combs, 2011). Other contributing factors to microglial senescence are age-related myelin degeneration and lysosomal storage in microglia, which in turn burden microglial clearance function (Holtman et al., 2015; Safaiyan et al., 2016).

Systemic inflammation and aging cause microglial priming. Primed microglia have a lower threshold for activation, are hypersensitive, develop an exaggerated immune response on activation, and have a distinct molecular signature from the M1–M2 phenotype (Perry and Teeling, 2013; Holtman et al., 2015; Ojo et al., 2015). The molecular signatures of primed microglia include the overexpression of antigen presentation, redox pathways, oxidative phosphorylation, and lysosomes.

In summary, during aging primed microglia generate a pro-inflammatory cascade due to their lower threshold of activation, enhanced reactivity, and limited functional capacity on encountering secondary triggers. The chronic activation of microglia coupled with age-related microglial priming hastens the process of senescence to cause loss of function over time (Franceschi et al., 2000; Streit and Xue, 2014). Senescent microglia appear to have lesser ramifications and stouter cell bodies, often referred to as dystrophic microglia.

#### Immunometabolic Sensor of Aging: Targeting Aging as a Disease

The inflammasome complex is a sensor of DAMPs. DAMPs act as an inflammatory challenge to the host defense mechanism and lead to the activation of the NLRP3 (Nod-like receptor pyrin domain 3) inflammasome complex (Youm et al., 2013; Zhang et al., 2013). Within the family of innate inflammasome sensors, the NLRP3 inflammasome has the unique ability to detect sterile inflammatory triggers. It can detect a wide range of metabolic and aging-related DAMPS, such as ROS production, glucose tolerance, and insulin resistance (Vandanmagsar et al., 2011; Salminen et al., 2012; Walsh et al., 2014), lipotoxic fatty acids, ceramides, free cholesterol, uric acid, and ATP, and it releases IL-1 $\beta$  and IL-18 (Youm et al., 2012, 2013; Zhang et al., 2013; see **Figure 2**).

The NLRP3 inflammasome complex activation is a two-step process. Molecular pathogens like lipopolysaccharides (LPS) have been shown to prime cells, leading to the activation of pattern recognition receptors (PRRs), the release of IL-1 $\beta$ , and increased expression of NLRP3. When followed by a secondary trigger such as ATP, this process causes the inflammasome to assemble and causes further activation. NLRP3 is also activated by the accumulation of damaged mitochondria due to the inhibition of autophagy, resulting in excessive production of ROS. oxidized mitochondrial DNA is also implicated in the activation of NLRP3 (Dixit, 2013). The activation of NLRP3 by ROS is mediated by thioredoxin interacting protein (TXNIIP) (Youm et al., 2012, 2013; Zhang et al., 2013). The activation patterns of NLRP3 are similar in both macrophages and microglia. NLRP3 activation leads to the priming of microglia and reducing the threshold for activation (Halle et al., 2008; Heneka et al., 2013). Increase in caspase-1 activity in postmortem MCI and AD brains indicates the possible participation of the NLRP3 inflammasome in AD pathogenesis (Heneka et al., 2013). Targeting NLRP3 and NF- $\kappa$ B (nuclear factor kappa-light-chain-enhancer of activated B cells) has been associated with a reduction in pathology of AD (Tang et al., 2015). In animal models carrying AD pathology, NLRP3 knockout and caspase-1 knockout caused spatial memory to remain intact. Moreover, the microglial phenotype in the NLRP3 knockout model shifted to the M2 anti-inflammatory phenotype with greater neuroprotection improved clearance of the plaque burden (Halle et al., 2008; Heneka et al., 2013).

With several studies linking NLRP3 inflammasome activation to chronic low-grade inflammation observed in aging, therapeutics targeting this sensor has also emerged. In a recent study, the ketone body –  $\beta$ -hydroxybutyrate (BHB) was found to suppress NLRP3 activation caused by urate crystals, lipotoxic fatty acids, and ATP. The levels of BHB increase with starvation, caloric restriction, and high-intensity exercise. Aging also marks a shift in the fuel usage and dependence on different fuel mechanisms. The inhibition of NLRP3 by BHB resulted in a decrease of IL-1 $\beta$  and IL-18 production by monocytes. BHB also reduced caspase-1 activation and IL-1 $\beta$  secretion in mouse models of NLRP3-mediated chronic inflammatory diseases like Muckle-Wells syndrome (Youm et al., 2015).

Another molecule, MCC950, blocked canonical and non-canonical activation of NLRP3 and attenuated experimental autoimmune encephalomyelitis (EAE). Both, MCC950 and BHB, were used in a mouse model of Muckle-Wells syndrome, which is characterized by chronic inflammation mediated by NLRP3. Thus, targeting NLRP3 in aging and aging-related disorders could be an important therapeutic strategy (Coll et al., 2015).

#### MENOPAUSE, INFLAMMATION, AND AD

The endocrine transition of the perimenopause to the post-menopause, while associated with loss of reproductive function (Brinton et al., 2015), is also associated with rise in chronic low-grade inflammation (Yin et al., 2015). Chronic systemic inflammation accelerates ovarian failure (Ağaçayak et al., 2016). Conversely, depleting proinflammatory cytokines IL-1 $\alpha$  and IL-1 $\beta$  extends ovarian function and lifespan (Uri-Belapolsky et al., 2014). Concurrent with the chronic low-grade inflammation, the perimenopausal transition is typified by decline in brain glucose metabolism and mitochondrial respiration (Yao et al., 2010; Ding et al., 2013; Brinton et al., 2015; Yin et al., 2015; Mosconi et al., 2017a,b), myelin catabolism (Klosinski et al., 2015) and loss of white matter volume (Mosconi et al., 2017a,b), beta-amyloid deposition in brain (Mosconi et al., 2017a,b) and changes in neurological function (Brinton et al., 2015). Later age at natural and surgical menopause is associated with better verbal memory (Kuh et al., 2018). Surgically induced menopause prior to natural menopause is associated with rapid cognitive decline and earlier onset of AD (Rocca et al., 2008; Bove et al., 2014). Further, studies have shown that post-menopausal women with higher estradiol levels have a reduced risk of developing AD (Manly et al., 2000).

Post-menopausal women are at higher risk for developing autoimmune disorders and obesity (Doran et al., 2002; Bove, 2013). The incidence of RA is higher in peri- and post-menopausal women (Doran et al., 2002). The pathology of multiple sclerosis worsens after menopause (Tutuncu et al., 2013). Post-menopausal women are more prone to robust immune responses. The lack of ovarian steroidal hormones potentiates the inflammatory process predisposing menopausal women to immune disorders (Benedusi et al., 2012; Kireev et al., 2014; Sharma et al., 2018). Menopause and the associated lack of steroidal hormones further potentiate inflammation, which is reflected in levels of circulating cytokine levels and inflammatory responses. IL-6 and sIL-6 levels are higher in postmenopausal women (Giuliani et al., 2001). IL-4 and IL-2 levels also increase with menopause, which can be reversed by hormone therapy (Yasui et al., 2007). Serum IFN-y levels increase during early menopause but decrease in later menopause (Goetzl et al., 2010). Peripheral blood mononuclear cells (PBMCs) isolated from postmenopausal women produced higher IL-6, IL-1β, and TNFα upon induction by LPS than PBMCs isolated from premenopausal women (Brooks-Asplund et al., 2002).

In addition to an altered cytokine profile, changes in T cell biology occur in women during this endocrine transition. Pre-menopausal women have higher CD4 counts than men and thus a more robust response to vaccination. Menopause causes a reduction in CD4 T cell numbers. This eventually causes an inversion of the CD4/CD8 T cell ratio, which is indicative of aging and can be correlated with increased oxidative stress (Larbi et al., 2008; Gameiro et al., 2010; Muller et al., 2015). The number of B2 cells (involved with antibody production) also decreases with menopause, especially during late menopause in comparison to perimenopause (Kamada et al., 2001). In mice, ovariectomy causes a reduction in the LPS-induced proliferation of leukocytes and subsequent chemotaxis, which is indicative of premature immune senescence (Baeza et al., 2011). Ovariectomized animals generally have a reduced and delayed adaptive response to vaccination, leading to decreased IgG titers in comparison to animals with intact ovaries (Haberthur et al., 2010). These changes are indicative of immune senescence occurring during menopausal transition. In the context of AD, the systemic effect of menopause on inflammation combined with effects on neurological function indicates cruciality of the menopausal transition in AD pathogensis.

## Molecular Neuro-Inflammatory Mechanisms in Menopause

Menopause is composed of three transitions; the perimenopause that precedes menopause, the cessation of reproductive capacity, menopause, and the years following menopause, post-menopause. Concomitant with this endocrine aging is chronological aging as the endocrine transition states span multiple years. Each of these endocrine stages is characterized by complex hormonal fluctuations (Brinton et al., 2015).

Decline in estradiol level during perimenopause and menopause coincides with a bioenergetic deficit in brain (Ding et al., 2013). Estradiol is a master regulator of metabolic function in the female (Rettberg et al., 2014). Clinical evidence of decline in glucose metabolism in brain and the coincident bioenergetic deficit is evidenced by reduced uptake of 18-fludeoxyglucose detected by positron emission tomography (PET) in perimenopausal and menopausal women (Mosconi et al., 2017a,b). The bioenergetic deficit precedes a shift to utilization of ketone bodies as a compensatory response to decline in brain glucose as bioenergetic fuel to generate ATP in brain. This shift to utilizing an auxiliary fuel during female endocrine aging activates catabolism of white matter as an endogenous lipid source of ketone bodies in brain (Klosinski et al., 2015) and a concomitant increase in microglial and astrocytic reactivity (Xie et al., 2013; Suenaga et al., 2015).

Analyses of microarray data obtained from the brains of postmenopausal women made available by NCBI revealed an inflammatory gene expression profile in the post central and superior frontal gyrus (Sárvári et al., 2012). In comparison to pre-menopausal women, post-menopausal women showed an upregulation in microglial markers CD14, CD18, and CD45, as well as TLR4 and MHC-II markers CD74 and C3 (Sárvári et al., 2012). These findings in the human female brain were consistent with the pattern of gene expression in the frontal cortex of ovariectomized middle-aged rats (13 months old) (Sárvári et al., 2012). Ovariectomy caused an upregulation of microglial reactivity markers CD11b, C18, CD45, and CD86, complement pathway C3, and phagocytic markers Msr2 and CD32. Together, these data are indicative of a shift in the microglial phenotype to an activated state (see **Figure 3**).

Hippocampal inflammatory gene expression in middle-aged rats drastically changed upon ovariectomy. A similar upregulation of microglial markers (CD45, IBA1, CD68, CD11b, CD18, Fcgr1a, and Fcgr2b) to that witnessed in the cortex was observed in the hippocampus. The gene expression results imply a possible activation of microglia. This effect was mitigated by treatment with estradiol and selective estrogen receptor- $\alpha$  (ER $\alpha$ ) and estrogen receptor- $\beta$  (ER $\beta$ ) agonists (Sarvari et al., 2015). Human post-menopausal gene expression in the hippocampus corroborated the inflammatory gene expression pattern in ovariectomized rats, with an upregulation of microglial reactivity markers CD11b, CD18, IBA1, CD14, and complement C3 (Sarvari et al., 2015).

In parallel, aging is associated with a marked upregulation in genes encoding the major histocompatibility complex class I and class II (MHC-I and MHC-II) (VanGuilder et al., 2011), alterations in Toll-like receptors (TLRs) (Shaw et al., 2011), and the complement pathway (Reichwald et al., 2009). This effect is more pronounced in women and represents the sexual dimorphism of the immune system (Blalock et al., 2003; Berchtold et al., 2008). The dynamics involved between age and menopause-related increase in myelin degeneration and microglial priming can be a tipping point in the neuro-inflammatory system. The increased myelin antigen load and upregulation of antigen presentation by microglia can set





forth a cascade that leads to dysregulated glial metabolism and hypertrophy, eventually causing altered extracellular matrix (Blalock et al., 2003). Each event is pivotal in the development of AD.

#### Hormone Therapy: A Potential Anti-inflammatory Preventive Intervention for Alzheimer's Disease?

Hormone therapy (HT) promotes neuronal survival and has been shown to improve cognitive function and episodic memory in perimenopausal and postmenopausal women (Morrison et al., 2006; Brinton, 2008). Epidemiological studies have shown that HT delays the onset of AD as well as reduces the risk of developing AD (Tang et al., 1996; Persad et al., 2009; Dye et al., 2012). Women transitioning through their menopause benefit most from HT as compared to women who have already transitioned (Girard et al., 2017). Results from several clinical trials have emphasized on the timing of treatment with HT and the drawbacks of missing the window of treatment (Hogervorst et al., 2000; Zandi et al., 2002; Shumaker et al., 2003). This effect of estradiol in HT has been explained by two theories: the healthy cell bias of estrogen action (Brinton, 2008) and critical window hypothesis (Maki, 2013). Healthy cell bias highlights that neuronal viability and health at baseline are important for estradiol to exert its therapeutic efficacy, whereas the critical window hypothesis focuses on the perimenopausal transition, when cells are still healthy, being a key phase for using HT. The use of estradiol in HT provides a therapeutic opportunity to target inflammatory pathways that simultaneously modulate metabolic functions, thereby providing a supportive milieu for neuronal survival and growth (Vegeto et al., 2008; Zhao et al., 2014). HT restores the hormonal levels in post-menopausal women to those of premenopausal women. Post-menopausal women using HT have higher lymphocyte numbers and higher circulating monocytes in comparison to post-menopausal women who are not on HT (Kamada et al., 2000). Likewise, levels of B2 cells involved in antibody production are significantly higher in HT users in comparison to non-users (Kamada et al., 2001).

Estrogen receptor alpha (ERa) and estrogen receptor beta (ER<sub>β</sub>) are abundantly expressed in astrocytes, microglia, and neurons, and both  $ER\alpha$  and  $ER\beta$  are involved in regulating the immunomodulatory responses exerted by astrocytes and microglia (Liu et al., 2003; Khan and Ansar Ahmed, 2015). In ovariectomized middle-aged rats, estradiol induces downregulation of the complement pathway and macrophageassociated genes in the frontal cortex. This effect was mediated through ERα and ERβ (Sárvári et al., 2011). Estradiol treatment in microglial cells induces a dose-dependent attenuation in superoxide release, phagocytic activity, and a concomitant increase in iNOS activity, without altering NF- $\kappa$ B expression (Bruce-Keller et al., 2000; Drew and Chavis, 2000). Some studies have shown that sex steroids reduce neuroinflammation via inhibiting the inflammasome complex, a possible downstream effect mediated by ER $\alpha$  and ER $\beta$  (Slowik and Beyer, 2015).

Astrocytes also participate in mediating the neuroprotective anti-inflammatory effect of estradiol via  $\text{ER}\alpha$ 

(Spence et al., 2011). In contrast to microglial cells, estradiol inhibits NF- $\kappa$ B expression in astrocytes (Giraud et al., 2010; Acaz-Fonseca et al., 2014). Estradiol inhibits secretion of proinflammatory cytokines IL-6, TNF- $\alpha$ , IL-1 $\beta$ , expression of matrix metalloproteinases 9 (MMP-9), and interferon gamma-inducible protein 10 (IP-10) in astrocytes. Estradiol also reduces proinflammatory cytokines secreted by astrocytes when exposed to A $\beta$  (Giraud et al., 2010; Acaz-Fonseca et al., 2014).

Much like estradiol, selective estrogen receptor modulators (SERMs) exert a neuroprotective effect by reducing neuroinflammation. Tamoxifen and raloxifene both reduce microgliosis, astrogliosis, and the production of proinflammatory cytokines IL-6 and IP-10 induced by LPS administration (Arevalo et al., 2012). They have also been demonstrated to protect neurons against neurotoxicity caused by neuroinflammation through an ER mediated pathway (Ishihara et al., 2015). SERMs reduce the proinflammatory response produced by astrocytes and are helpful in potentiating their neurotrophic function (Tapia-Gonzalez et al., 2008; Cerciat et al., 2010; Arevalo et al., 2012; Ishihara et al., 2015).

## APOEε4, INFLAMMATION, AND ALZHEIMER'S DISEASE

APOE¢4 is the primary genetic risk factor for the late onset form of AD (Scacchi et al., 1995; Liu et al., 2014; Manning et al., 2014). The human form of the APOE gene possesses three polymorphic alleles: E2, E3, and E4. The E3 allele occurs more frequently (77%) than E2 (8%) and E4 (14%) (Eisenberg et al., 2010). The E4 allele occurs in 40% of AD patients (Farrer et al., 1997). However, 91% of homozygous E4 carriers and 47% heterozygous carriers go on to develop AD (Corder et al., 1993).

Female APOE $\varepsilon$ 4 carriers are more susceptible to developing AD than males (Altmann et al., 2014; Ungar et al., 2014; Neu et al., 2017). Prospective cohort studies have also suggested that female APOE $\varepsilon$ 4 carriers are at greater risk of converting from MCI to AD than males (Altmann et al., 2014). Female APOE $\varepsilon$ 4 carriers also have a higher rate of cognitive decline than APOE $\varepsilon$ 3 carriers (Holland et al., 2013). Leukocyte telomere length is also greatly reduced in APOE $\varepsilon$ 4 carrier's relative to age-matched controls, reflecting premature aging of female APOE $\varepsilon$ 4 carriers (Jacobs et al., 2013).

Apolipoprotein E (ApoE) is a key regulator of lipid homeostasis and in the brain, it functions to shuttle lipid molecules from astrocytes and microglia to neurons, via lipoprotein complexes (Liu et al., 2013). In the periphery, ApoE is expressed in macrophages and liver. In the central nervous system, ApoE is mainly produced by astrocytes and microglia (Liu et al., 2013).

# Sources of Neuroinflammation in APOEε4

ApoE is known to exert an immunosuppressive effect by inhibiting lymphocyte proliferation, Ig synthesis, and neutrophil activation. ApoE also exerts this immunosuppressive property on microglial activation (Guo et al., 2004; Baitsch et al., 2011; Christensen et al., 2011). Relative to APOEɛ2 and APOEɛ3 carriers, APOEɛ4 carriers generate less ApoE protein in periphery and brain (Larson et al., 2000). Given the reduced amounts of ApoE protein in APOEɛ4 carriers, this population could be predisposed to a heightened inflammatory response when encountering a sterile inducer or infection (Ukkola et al., 2009; Gale et al., 2014; Tai et al., 2015).

Due to the differences in cysteine and arginine residues translated at positions 112 and 158, each of the three different isoforms of APOE exhibit different conformations. The conformational change in the protein affects its stability, folding characteristics, and the propensity to bind lipoprotein particles (Jofre-Monseny et al., 2008). APOEɛ4 has a globule-like structure and preferably binds to very low-density lipoprotein (vLDL) and low-density lipoprotein (LDL) particles. However, APOEE2 and APOEE3 tend to bind high-density lipoprotein (HDL) particles (Jofre-Monseny et al., 2008). This difference in protein structure affects the lipid shuttling ability of ApoE, which leads to hypercholesterolemia in APOEE4 carriers and increases the predisposition for generation of plaques (Jofre-Monseny et al., 2008). ApoE also exerts an inhibitory effect on the oxidation of LDL in an isoform-specific manner (E2>E3>E4) (Miyata and Smith, 1996). Among smokers, APOEE4 carriers have significantly higher amounts of oxidized LDL (ox-LDL) (Jofre-Monseny et al., 2008).

Cholesterol, ox-LDL, and A $\beta$  are sterile inducers of inflammation called DAMPs (Miller et al., 2011; Clark and Vissel, 2015). DAMPs are recognized by PRRs expressed on macrophages, dendritic cells, monocytes, microglia, and neutrophils, which trigger the activation of an inflammatory process (Silverstein and Febbraio, 2009). One such PRR is CD36. In the context of recognizing DAMPs and plaque formation, it was recently demonstrated that CD36 (expressed on monocytes, macrophages, and microglia) recognizes soluble ligands such as oxidized LDL and soluble A $\beta$  and converts them to crystals and fibrils, respectively. This leads to the assembly and activation of the NLRP3 inflammasome and the consequent release of the proinflammatory cytokine IL-1 $\beta$  (Sheedy et al., 2013; Oury, 2014).

Due to the increased probability of APOE&4 carriers to develop plaques, cholesterol crystals, and amyloid depositions, cellular immune function and reactivity are affected. In APOE&4 carriers there is a reduced clearance and efflux of cholesterol from macrophages (Cash et al., 2012). Moreover, increased nitric oxide production in APOE&4 causes increased platelet aggregation and secretion of adhesion molecules, further enabling the plaque formation in the periphery. In the brain, microglial and astrocytic clearance of debris is also diminished (Guo et al., 2006).

ApoE also affects  $A\beta$  uptake and oligomerization and thus can be a key factor in  $A\beta$  turnover. AD patients possessing the APOE&4 allele were found to have higher levels oligomeric  $A\beta$ in their brains as compared to APOE&3 carriers, implicating an association between ApoE with  $A\beta$  (Hashimoto et al., 2012). *In vitro* experiments correspond to clinical findings and have shown that APOE&4 has the greatest effect on promoting  $A\beta$ oligomerization in comparison to other isoforms (Hashimoto et al., 2012). Blocking  $A\beta$  and ApoE interaction by ApoE  $A\beta$  antagonist in hippocampal neuronal and astrocytic co-culture systems led to decreased accumulation and oligomerization of A $\beta$ . Treatment with ApoE A $\beta$  antagonists also inhibited the loss of synaptic proteins induced by A $\beta$  accumulation (Kuszczyk et al., 2013; Liu S. et al., 2014).

Coupled with the A $\beta$  oligomerization, ApoE also affects A $\beta$  uptake. Astrocytes secrete ApoE as a lipoparticle into the interstitial fluid, where it binds with A $\beta$ . Neurons endocytose and internalize these lipoparticles, thus promoting A $\beta$  uptake. APOE $\epsilon$ 4 isoform has maximal binding affinity to A $\beta$  and thus causes a greater uptake of A $\beta$  by neurons in comparison to other isoforms (Mulder et al., 2014). APOE $\epsilon$ 4 prevents the uptake of oligomeric A $\beta$  by astrocytes and the uptake of both oligomeric and fibrillar A $\beta$  by microglia thereby inhibiting its clearance (Mulder et al., 2014).

ApoE also modulates A $\beta$  clearance by microglia by regulating Aß clearing enzymes such as neprilysin intracellularly and insulin degrading enzyme extracellularly. Effective degradation of AB depends on Liver X Receptor (LXR) activation, the isoform of APOE expressed, and the lipidation status of ApoE particles (Hashimoto et al., 2012; Kuszczyk et al., 2013; Liu S. et al., 2014). Activation of LXR/RXR (Retinoid X receptor) potentiates Aβ clearance as it compensates for the loss of APOEE4 function and induces the expression of ATP-binding cassette transporter subfamily A member 1 (ABCA1) and ApoE, thus inducing clearance by microglia and astrocytes (Lefterov et al., 2007; Terwel et al., 2011; Lee et al., 2012; Mandrekar-Colucci et al., 2012; Tai et al., 2014). Therefore, the presence of APOEE4 promotes the production of  $A\beta$  and uptake by neurons while preventing clearance and enabling the production of DAMPs and chronic low-grade inflammation.

Given the dysregulated lipid metabolism and impairment of cellular function with age, the APOE64 allele is associated with increased systemic inflammation. It is expected that this would be reflected in cytokine levels, which are inflammatory markers such as C-Reactive Protein (CRP) measured in plasma/serum. Surprisingly, however, this is not the case, at least with CRP. Studies have consistently shown that CRP levels are lower in APOE64 carriers (Ukkola et al., 2009; Lima et al., 2014; Metti et al., 2014; Yun et al., 2015). Marz et al. (2004) proposed that the metabolism of CRP might be associated with the mevalonate/cholesterol synthetic pathway, which might be downregulated in APOE64 carriers. On the other hand, studies have shown that levels of IL-1 $\beta$  and vascular inflammatory marker: vascular cell adhesion molecule-1 (VCAM-1) are higher in E4 carriers (Olgiati et al., 2010; see Figure 4).

The APOE&4 allele also accelerates aging, which is reflected in the shorter telomeres in APOE&4 women in comparison to APOE&3 women (Jacobs et al., 2013). The accelerated aging phenotype is also evident in the reduction of T cell numbers. Age-related reduction in T cells for women is more dramatic during menopause, which is even more pronounced if the woman is an APOE&4 carrier (Begum et al., 2014). In comparison to APOE&3 derived microglia, estradiol has a reduced anti-inflammatory effect on microglia derived from APOE&4 (Brown et al., 2008). The APOE&4 allele is also a risk factor for metabolic syndrome and has been associated with RA, thus



increasing the risk of comorbidities that can, in turn, affect systemic inflammation (Sima et al., 2007; Gungor et al., 2012; Toms et al., 2012; Johnson et al., 2013). APOEc4 also affects immune function by causing blood barrier dysfunction (Nelson et al., 2016).

#### Innate Immune Responses in APOE<sub>6</sub>4

Inflammatory responses triggered by innate immune agonists are highest in APOE $\varepsilon$ 4 carriers (Vitek et al., 2009; Gale et al., 2014). This finding holds true in cells isolated from both humans and rodents and in both the periphery and the brain (Gale et al., 2014; Li X. et al., 2015). Therefore, inflammatory triggers such as TBI, infection, and DAMPs produced from metabolic syndrome significantly increase the inflammatory response in APOE $\varepsilon$ 4 carriers. This may potentially lead to the incomplete resolution of inflammation to initiate chronic inflammatory processes that result in neurotoxicity. This trend is evident in HIV-associated dementia, for which E4 carriers have increased risk (Chang et al., 2011).

A heightened inflammatory response occurred in microglia derived from humanized APOE  $\epsilon4$  knock-in mice upon on

treatment with the TLR3 and TLR4 activator LPS compared to APOE $\epsilon$ 3 mice (Vitek et al., 2009; Heneka et al., 2015). The inflammatory response was characterized by altered cell morphology, increased nitric oxide production, COX-2 expression, prostaglandin E2 (PGE2) expression, and cytokine production (IL-6, TNF- $\alpha$ , and IL-12p40). In contrast, TREM2 expression was decreased (Vitek et al., 2009; Heneka et al., 2015). A comparable inflammatory response was observed in peripheral macrophages isolated from APOE $\epsilon$ 4 mice. The E4 allele increases the reactivity of glial and peripheral immune cells, thus aggravating the neurotoxic proinflammatory response (Vitek et al., 2009; Heneka et al., 2015).

### DISCUSSION

The etiology of the prodromal phase of AD presents as a complex interplay between several risk factors, which is relevant to therapeutic interventions and preventive strategies (see **Figures 5**, **6**). This implies that therapeutic strategies should employ stratification of patient populations regarding parameters



**FIGURE 5** Inflammation integrates Alzheimer's disease risk factors of female sex, chronological age, endocrine aging, and APOE¢4 genotype. The three hit model of Alzheimer's risk: aging, menopause, and APOE¢4 genotype collectively induce a compromised bioenergetic system in brain that is impacted by the chronic low grade innate inflammation of aging coupled with APOE¢4 dysregulated cholesterol homeostasis lead to activation of the adaptive immune response. The inflammatory immune response is the factor that bridges across each of the risk factors for AD. Immune system regulators that are specific to stage of disease and inflammatory phenotype would provide a therapeutic strategy to disconnect the bridge that drives disease.



**FIGURE 6** | Immune drivers involved in aging, menopause, and APOE£4 genotype related inflammation. Key immune drivers contributing to inflammation due to aging, menopause and APOE£4 genotype related inflammation are detailed to give a global picture of immune dynamics (upward and downward arrows indicate increased and decreased expression, respectively).

of age, sex, and APOE genotype. It also calls for the use of combination therapies that modulate inflammation, lipid-based metabolism in APOE¢4 carriers, and loss of estrogenic control in menopausal women. For example, preventive strategies to reduce age-related inflammation could include a combination of NSAIDs and statins in middle-aged adults (45–55 years). In women, this therapy could be modified to include HT during their perimenopausal transition. Patient's medical histories and electronic health records are a source of indicators for chronic inflammation. These strategies should be tailored to the patient's metabolic profile, genetic history, and endocrine-related transition states (see **Figures 5, 6**).

This understanding of the disease progression also calls for change in design of clinical trials that target the amyloidogenic pathway and treat later stages of the disease pathogenesis. Trial design should incorporate the identification of persons with an increased risk of developing AD, and utilize a risk-factor-based responder analysis. Inflammation-mediated therapeutic and preventive strategies will largely depend upon this stratification of patient populations.

The inflammatory response is influenced by age, chromosomal sex, endocrine transition – menopause and APOE genotype. Inflammation is characteristic of each of these modifying factors and can be a driving force for development of AD. Thus, inflammation has been a therapeutic target in multiple clinical trials for AD. However, each of these trials have failed to meet primary endpoints. Reviewed herein is a consideration of the multiple factors that contribute to and modify the inflammatory phenotype. Going forward, in both discovery and clinical science, it will be important to delineate the etiology of the inflammatory response, the stage of the inflammatory cascade, and the activated network of inflammatory signaling. Inflammation is a moving target and thus requires a precision approach to identifying etiology, stage and appropriate therapeutic target.

#### REFERENCES

- Acaz-Fonseca, E., Sanchez-Gonzalez, R., Azcoitia, I., Arevalo, M. A., and Garcia-Segura, L. M. (2014). Role of astrocytes in the neuroprotective actions of 17β-estradiol and selective estrogen receptor modulators. *Mol. Cell. Endocrinol.* 389, 48–57. doi: 10.1016/j.mce.2014.01.009
- Ağaçayak, E., Yaman Görük, N., Küsen, H., Yaman Tunç, S., Başaranoğlu, S., Içe, M. S., et al. (2016). Role of inflammation and oxidative stress in the etiology of primary ovarian insufficiency. *J. Turk. Soc. Obstet. Gynecol.* 13, 109–115. doi: 10.4274/tjod.00334
- Alisky, J. M. (2007). The coming problem of HIV-associated Alzheimer's disease. Med. Hypotheses 69, 1140–1143. doi: 10.1016/j.mehy.2007.02.030
- Altmann, A., Tian, L., Henderson, V. W., Greicius, M. D., and Alzheimer's Disease Neuroimaging Initiative, I. (2014). Sex modifies the APOE-related risk of developing Alzheimer disease. Ann. Neurol. 75, 563–573. doi: 10.1002/ana. 24135
- Alzheimer's Disease Anti-inflammatory Prevention Trial Research Group (2013). Results of a follow-up study to the randomized Alzheimer's Disease Antiinflammatory Prevention Trial (ADAPT). *Alzheimers Dement.* 9, 714–723. doi: 10.1016/j.jalz.2012.11.012
- Amieva, H., Goff, M. L., Millet, X., Orgogozo, J. M., Pérès, K., Barberger-Gateau, P., et al. (2008). Prodromal Alzheimer's disease: successive emergence of the clinical symptoms. *Ann. Neurol.* 64, 492–498. doi: 10.1002/ana.21509

#### **CONCLUDING REMARKS**

In summary, neuro-inflammatory processes are evident early in the latent prodromal phase and worsen during the course of the Alzheimer's. Disparate results from epidemiological and clinical trials targeting inflammation, highlight the complexity of the inflammatory process. The inflammatory processes that occur during aging, midlife endocrine transitions, and in the APOEE4 carrier contribute to risk and progression of AD. The chronic inflammatory processes that are activated during midlife chronological and endocrine aging, ultimately limit the clearance capacity of microglia and lead to immune senescence. The loss of estrogenic control of bioenergetic function in the brain coupled with dysregulated lipid metabolism in the APOEE4 genotype adversely impact microglial function and clearance mechanisms. The dynamic and context specific activation pattern of the inflammatory processes provide plausible mechanisms underlying failed clinical trials of anti-inflammatory agents in Alzheimer's patients. Collectively, these considerations highlight the rationale for stratifying AD clinical trial cohorts based on their inflammatory phenotype. Combination therapies that include targeted use of anti-inflammatory agent's specific to the immune phenotype could have a higher probability of successfully modifying risk and progression of Alzheimer's disease.

#### **AUTHOR CONTRIBUTIONS**

AM and RB wrote and reviewed the manuscript.

### FUNDING

This work was supported by National Institute on Aging (NIA) grants P01AG026572 and R37AG053589 to RB.

- Arevalo, M. A., Diz-Chaves, Y., Santos-Galindo, M., Bellini, M. J., and Garcia-Segura, L. M. (2012). Selective oestrogen receptor modulators decrease the inflammatory response of glial cells. *J. Neuroendocrinol.* 24, 183–190. doi: 10. 1111/j.1365-2826.2011.02156.x
- Baeza, I., De Castro, N. M., Arranz, L., Fdez-Tresguerres, J., and De La Fuente, M. (2011). Ovariectomy causes immunosenescence and oxi-inflamm-ageing in peritoneal leukocytes of aged female mice similar to that in aged males. *Biogerontology* 12, 227–238. doi: 10.1007/s10522-010-9317-0
- Baitsch, D., Bock, H. H., Engel, T., Telgmann, R., Muller-Tidow, C., Varga, G., et al. (2011). Apolipoprotein E induces antiinflammatory phenotype in macrophages. *Arterioscler. Thromb. Vasc. Biol.* 31, 1160–1168. doi: 10.1161/ATVBAHA.111. 222745
- Begum, A. N., Cunha, C., Sidhu, H., Alkam, T., Scolnick, J., Rosario, E. R., et al. (2014). Women with the Alzheimer's risk marker ApoE4 lose Abeta-specific CD4<sup>+</sup> T cells 10-20 years before men. *Transl. Psychiatry* 4:e414. doi: 10.1038/ tp.2014.51
- Benedusi, V., Meda, C., Della Torre, S., Monteleone, G., Vegeto, E., and Maggi, A. (2012). A lack of ovarian function increases neuroinflammation in aged mice. *Endocrinology* 153, 2777–2788. doi: 10.1210/en.2011-1925
- Berchtold, N. C., Cribbs, D. H., Coleman, P. D., Rogers, J., Head, E., Kim, R., et al. (2008). Gene expression changes in the course of normal brain aging are sexually dimorphic. *Proc. Natl. Acad. Sci. U.S.A.* 105, 15605–15610. doi: 10.1073/pnas.0806883105

- Blalock, E. M., Chen, K.-C., Sharrow, K., Herman, J. P., Porter, N. M., Foster, T. C., et al. (2003). Gene microarrays in hippocampal aging: statistical profiling identifies novel processes correlated with cognitive impairment. *J. Neurosci.* 23, 3807–3819. doi: 10.1523/JNEUROSCI.23-09-03807.2003
- Bove, R. (2013). Autoimmune diseases and reproductive aging. *Clin. Immunol.* 149, 251–264. doi: 10.1016/j.clim.2013.02.010
- Bove, R., Secor, E., Chibnik, L. B., Barnes, L. L., Schneider, J. A., Bennett, D. A., et al. (2014). Age at surgical menopause influences cognitive decline and Alzheimer pathology in older women. *Neurology* 82, 222–229. doi: 10.1212/ WNL.000000000000033
- Boyd, T. D., Bennett, S. P., Mori, T., Governatori, N., Runfeldt, M., Norden, M., et al. (2010). GM-CSF upregulated in rheumatoid arthritis reverses cognitive impairment and amyloidosis in Alzheimer mice. J. Alzheimers Dis. 21, 507–518. doi: 10.3233/JAD-2010-091471
- Breitner, J. C., Baker, L. D., Montine, T. J., Meinert, C. L., Lyketsos, C. G., Ashe, K. H., et al. (2011). Extended results of the Alzheimer's disease antiinflammatory prevention trial. *Alzheimers Dement*. 7, 402–411. doi: 10.1016/ j.jalz.2010.12.014
- Breunig, J. J., Guillot-Sestier, M. V., and Town, T. (2013). Brain injury, neuroinflammation and Alzheimer's disease. *Front. Aging Neurosci.* 5:26. doi: 10.3389/fnagi.2013.00026
- Brinkmalm, A., Brinkmalm, G., Honer, W. G., Frölich, L., Hausner, L., Minthon, L., et al. (2014). SNAP-25 is a promising novel cerebrospinal fluid biomarker for synapse degeneration in Alzheimer's disease. *Mol. Neurodegener.* 9:53. doi: 10.1186/1750-1326-9-53
- Brinton, R. D. (2008). The healthy cell bias of estrogen action: mitochondrial bioenergetics and neurological implications. *Trends Neurosci.* 31, 529–537. doi: 10.1016/j.tins.2008.07.003
- Brinton, R. D., Yao, J., Yin, F., Mack, W. J., and Cadenas, E. (2015). Perimenopause as a neurological transition state. *Nat. Rev. Endocrinol.* 11, 393–405. doi: 10. 1038/nrendo.2015.82
- Brooks-Asplund, E. M., Tupper, C. E., Daun, J. M., Kenney, W. L., and Cannon, J. G. (2002). Hormonal modulation of interleukin-6, tumor necrosis factor and associated receptor secretion in postmenopausal women. *Cytokine* 19, 193–200. doi: 10.1006/cyto.2002.1963
- Brown, C. M., Choi, E., Xu, Q., Vitek, M. P., and Colton, C. A. (2008). The APOE4 genotype alters the response of microglia and macrophages to 17beta-estradiol. *Neurobiol. Aging* 29, 1783–1794. doi: 10.1016/j.neurobiolaging.2007.04.018
- Bruce-Keller, A. J., Keeling, J. L., Keller, J. N., Huang, F. F., Camondola, S., and Mattson, M. P. (2000). Antiinflammatory effects of estrogen on microglial activation. *Endocrinology* 141, 3646–3656. doi: 10.1210/endo.141.10.7693
- Butovsky, O., Jedrychowski, M. P., Moore, C. S., Cialic, R., Lanser, A. J., Gabriely, G., et al. (2014). Identification of a unique TGF-beta-dependent molecular and functional signature in microglia. *Nat. Neurosci.* 17, 131–143. doi: 10.1038/nn.3599
- Calsolaro, V., and Edison, P. (2016). Neuroinflammation in Alzheimer's disease: current evidence and future directions. *Alzheimers Dement.* 12, 719–732. doi: 10.1016/j.jalz.2016.02.010
- Cash, J. G., Kuhel, D. G., Basford, J. E., Jaeschke, A., Chatterjee, T. K., Weintraub, N. L., et al. (2012). Apolipoprotein E4 impairs macrophage efferocytosis and potentiates apoptosis by accelerating endoplasmic reticulum stress. J. Biol. Chem. 287, 27876–27884. doi: 10.1074/jbc.M112.377549
- Cerciat, M., Unkila, M., Garcia-Segura, L. M., and Arevalo, M. A. (2010). Selective estrogen receptor modulators decrease the production of interleukin-6 and interferon-gamma-inducible protein-10 by astrocytes exposed to inflammatory challenge in vitro. *Glia* 58, 93–102. doi: 10.1002/glia.20904
- Chakradhar, S. (2018). A tale of two diseases: aging HIV patients inspire a closer look at Alzheimer's disease. Nat. Med. 24, 376–377. doi: 10.1038/ nm0418-376
- Chang, L., Andres, M., Sadino, J., Jiang, C. S., Nakama, H., Miller, E., et al. (2011). Impact of apolipoprotein E epsilon4 and HIV on cognition and brain atrophy: antagonistic pleiotropy and premature brain aging. *Neuroimage* 58, 1017–1027. doi: 10.1016/j.neuroimage.2011.07.010
- Chou, R. C., Kane, M., Ghimire, S., Gautam, S., and Gui, J. (2016). Treatment for rheumatoid arthritis and risk of Alzheimer's disease: a nested case-control analysis. *CNS Drugs* 30, 1111–1120. doi: 10.1007/s40263-016-0374-z
- Christensen, D. J., Ohkubo, N., Oddo, J., Van Kanegan, M. J., Neil, J., Li, F., et al. (2011). Apolipoprotein E and peptide mimetics modulate inflammation

by binding the SET protein and activating protein phosphatase 2A. J. Immunol. 186, 2535–2542. doi: 10.4049/jimmunol.1002847

- Clark, I. A., and Vissel, B. (2015). Amyloid beta: one of three danger-associated molecules that are secondary inducers of the proinflammatory cytokines that mediate Alzheimer's disease. *Br. J. Pharmacol.* 172, 3714–3727. doi: 10.1111/ bph.13181
- Coll, R. C., Robertson, A. A., Chae, J. J., Higgins, S. C., Munoz-Planillo, R., Inserra, M. C., et al. (2015). A small-molecule inhibitor of the NLRP3 inflammasome for the treatment of inflammatory diseases. *Nat. Med.* 21, 248–255. doi: 10.1038/ nm.3806
- Corder, E. H., Saunders, A. M., Strittmatter, W. J., Schmechel, D. E., Gaskell, P. C., Small, G. W., et al. (1993). Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* 261, 921–923. doi: 10.1126/science.8346443
- Crain, J. M., Nikodemova, M., and Watters, J. J. (2013). Microglia express distinct M1 and M2 phenotypic markers in the postnatal and adult central nervous system in male and female mice. J. Neurosci. Res. 91, 1143–1151. doi: 10.1002/ jnr.23242
- Cui, H., Kong, Y., and Zhang, H. (2012). Oxidative stress, mitochondrial dysfunction, and aging. J. Signal. Transduct. 2012:646354. doi: 10.1155/2012/ 646354
- Damani, M. R., Zhao, L., Fontainhas, A. M., Amaral, J., Fariss, R. N., and Wong,
  W. T. (2011). Age-related alterations in the dynamic behavior of microglia. *Aging Cell* 10, 263–276. doi: 10.1111/j.1474-9726.2010.00660.x
- Dickson, D. W., Lee, S. C., Mattiace, L. A., Yen, S. -H. C., and Brosnan, C. (1993). Microglia and cytokines in neurological disease, with special reference to AIDS and Alzheimer's disease. *Glia* 7, 75–83. doi: 10.1002/glia.440070113
- Ding, F., Yao, J., Rettberg, J. R., Chen, S., and Brinton, R. D. (2013). Early decline in glucose transport and metabolism precedes shift to ketogenic system in female aging and Alzheimer's mouse brain: implication for bioenergetic intervention. *PLoS One* 8:e79977. doi: 10.1371/journal.pone.0079977
- Dixit, V. D. (2013). Nlrp3 inflammasome activation in type 2 diabetes: is it clinically relevant? *Diabetes* 62, 22–24. doi: 10.2337/db12-1115
- Doran, M. F., Pond, G. R., Crowson, C. S., O'fallon, W. M., and Gabriel, S. E. (2002). Trends in incidence and mortality in rheumatoid arthritis in Rochester, Minnesota, over a forty-year period. *Arthritis Rheum.* 46, 625–631. doi: 10.1002/ art.509
- Drew, P. D., and Chavis, J. A. (2000). Female sex steroids: effects upon microglial cell activation. J. Neuroimmunol. 111, 77–85. doi: 10.1016/S0165-5728(00) 00386-6
- Dye, R. V., Miller, K. J., Singer, E. J., and Levine, A. J. (2012). Hormone replacement therapy and risk for neurodegenerative diseases. *Int. J. Alzheimers Dis.* 2012:258454. doi: 10.1155/2012/258454
- Eikelenboom, P., Van Exel, E., Veerhuis, R., Rozemuller, A. J., Van Gool, W. A., and Hoozemans, J. J. (2012). Innate immunity and the etiology of late-onset Alzheimer's disease. *Neurodegener. Dis.* 10, 271–273. doi: 10.1159/000334287
- Eisenberg, D. T., Kuzawa, C. W., and Hayes, M. G. (2010). Worldwide allele frequencies of the human apolipoprotein E gene: climate, local adaptations, and evolutionary history. *Am. J. Phys. Anthropol.* 143, 100–111. doi: 10.1002/ajpa. 21298
- Etminan, M., Gill, S., and Samii, A. (2003). Effect of non-steroidal antiinflammatory drugs on risk of Alzheimer's disease: systematic review and meta-analysis of observational studies. *BMJ* 327:128. doi: 10.1136/bmj.327. 7407.128
- Farrer, L. A., Cupples, L. A., Haines, J. L., Hyman, B., Kukull, W. A., Mayeux, R., et al. (1997). Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease. A meta-analysis. APOE and Alzheimer Disease Meta Analysis Consortium. *JAMA* 278, 1349–1356. doi: 10.1001/jama.1997.03550160069041
- Ferretti, M. T., and Cuello, A. C. (2011). Does a pro-inflammatory process precede Alzheimer's disease and mild cognitive impairment? *Curr. Alzheimer Res.* 8, 164–174. doi: 10.2174/156720511795255982
- Floden, A. M., and Combs, C. K. (2011). Microglia demonstrate age-dependent interaction with amyloid-beta fibrils. J. Alzheimers Dis. 25, 279–293. doi: 10. 3233/JAD-2011-101014
- Fougère, B., Boulanger, E., Nourhashémi, F., Guyonnet, S., and Cesari, M. (2017). Chronic inflammation: accelerator of biological aging. J. Gerontol. Ser. A 72, 1218–1225.

- Franceschi, C., Bonafè, M., Valensin, S., Olivieri, F., De Luca, M., Ottaviani, E., et al. (2000). Inflamm-aging: an evolutionary perspective on immunosenescence. *Ann. N. Y. Acad. Sci.* 908, 244–254. doi: 10.1111/j.1749-6632.2000.tb06651.x
- Gale, S. C., Gao, L., Mikacenic, C., Coyle, S. M., Rafaels, N., Murray Dudenkov, T., et al. (2014). APOepsilon4 is associated with enhanced in vivo innate immune responses in human subjects. J. Allergy Clin. Immunol. 134, 127–134. doi: 10. 1016/j.jaci.2014.01.032
- Gameiro, C. M., Romao, F., and Castelo-Branco, C. (2010). Menopause and aging: changes in the immune system–a review. *Maturitas* 67, 316–320. doi: 10.1016/j. maturitas.2010.08.003
- Girard, R., Metereau, E., Thomas, J., Pugeat, M., Qu, C., and Dreher, J. C. (2017). Hormone therapy at early post-menopause increases cognitive control-related prefrontal activity. *Sci. Rep.* 7:44917. doi: 10.1038/srep44917
- Giraud, S. N., Caron, C. M., Pham-Dinh, D., Kitabgi, P., and Nicot, A. B. (2010). Estradiol inhibits ongoing autoimmune neuroinflammation and NFkappaBdependent CCL2 expression in reactive astrocytes. *Proc. Natl. Acad. Sci. U.S.A.* 107, 8416–8421. doi: 10.1073/pnas.0910627107
- Giuliani, N., Sansoni, P., Girasole, G., Vescovini, R., Passeri, G., Passeri, M., et al. (2001). Serum interleukin-6, soluble interleukin-6 receptor and soluble gp130 exhibit different patterns of age- and menopause-related changes. *Exp. Gerontol.* 36, 547–557. doi: 10.1016/S0531-5565(00)00220-5
- Goetzl, E. J., Huang, M. -C., Kon, J., Patel, K., Schwartz, J. B., Fast, K., et al. (2010). Gender specificity of altered human immune cytokine profiles in aging. *FASEB J.* 24, 3580–3589. doi: 10.1096/fj.10-160911
- Goronzy, J. J., and Weyand, C. M. (2012). Immune aging and autoimmunity. Cell. Mol. Life Sci. 69, 1615–1623. doi: 10.1007/s00018-012-0970-0
- Group, A. R. (2007). Naproxen and celecoxib do not prevent AD in early results from a randomized controlled trial. *Neurology* 68, 1800–1808. doi: 10.1212/01. wnl.0000260269.93245.d2
- Group, A. R. (2008). Cognitive function over time in the alzheimer's disease anti-inflammatory prevention trial (adapt): results of a randomized, controlled trial of naproxen and celecoxib. Arch. Neurol. 65, 896–905. doi: 10.1001/ archneur.2008.65.7.nct70006
- Gulke, E., Gelderblom, M., and Magnus, T. (2018). Danger signals in stroke and their role on microglia activation after ischemia. *Ther. Adv. Neurol. Disord.* 11:1756286418774254. doi: 10.1177/1756286418774254
- Gungor, Z., Anuurad, E., Enkhmaa, B., Zhang, W., Kim, K., and Berglund, L. (2012). Apo E4 and lipoprotein-associated phospholipase A2 synergistically increase cardiovascular risk. *Atherosclerosis* 223, 230–234. doi: 10.1016/j. atherosclerosis.2012.04.021
- Guo, L., Ladu, M. J., and Van Eldik, L. J. (2004). A dual role for apolipoprotein E in neuroinflammation. J. Mol. Neurosci. 23, 205–212. doi: 10.1385/JMN:23:3:205
- Guo, S., Wang, S., Kim, W. J., Lee, S. R., Frosch, M. P., Bacskai, B. J., et al. (2006). Effects of apoE isoforms on beta-amyloid-induced matrix metalloproteinase-9 in rat astrocytes. *Brain Res.* 1111, 222–226. doi: 10.1016/j.brainres.2006.06.041
- Habeck, C., Risacher, S., Lee, G. J., Glymour, M. M., Mormino, E., Mukherjee, S., et al. (2012). Relationship between baseline brain metabolism measured using [(1)(8)F]FDG PET and memory and executive function in prodromal and early Alzheimer's disease. *Brain Imaging Behav.* 6, 568–583. doi: 10.1007/s11682-012-9208-x
- Haberthur, K., Engelman, F., Barron, A., and Messaoudi, I. (2010). Immune senescence in aged nonhuman primates. *Exp. Gerontol.* 45, 655–661. doi: 10. 1016/j.exger.2010.06.001
- Habib, P., Slowik, A., Zendedel, A., Johann, S., Dang, J., and Beyer, C. (2014). Regulation of hypoxia-induced inflammatory responses and M1-M2 phenotype switch of primary rat microglia by sex steroids. *J. Mol. Neurosci.* 52, 277–285. doi: 10.1007/s12031-013-0137-y
- Halle, A., Hornung, V., Petzold, G. C., Stewart, C. R., Monks, B. G., Reinheckel, T., et al. (2008). The NALP3 inflammasome is involved in the innate immune response to amyloid-beta. *Nat. Immunol.* 9, 857–865. doi: 10.1038/ni.1636
- Hashimoto, T., Serrano-Pozo, A., Hori, Y., Adams, K. W., Takeda, S., Banerji, A. O., et al. (2012). Apolipoprotein E, especially apolipoprotein E4, increases the oligomerization of amyloid beta peptide. *J. Neurosci.* 32, 15181–15192. doi: 10.1523/JNEUROSCI.1542-12.2012
- Helenius, M., Hänninen, M., Lehtinen, S. K., and Salminen, A. (1996). Changes associated with aging and replicative senescence in the regulation of transcription factor nuclear factor-kappa B. *Biochem. J.* 318, 603–608. doi: 10.1042/bj3180603

- Heneka, M. T., Carson, M. J., Khoury, J. E., Landreth, G. E., Brosseron, F., Feinstein, D. L., et al. (2015). Neuroinflammation in Alzheimer's disease. *Lancet Neurol.* 14, 388–405. doi: 10.1016/S1474-4422(15)70016-5
- Heneka, M. T., Kummer, M. P., Stutz, A., Delekate, A., Schwartz, S., Vieira-Saecker, A., et al. (2013). NLRP3 is activated in Alzheimer's disease and contributes to pathology in APP/PS1 mice. *Nature* 493, 674–678. doi: 10.1038/ nature11729
- Hickman, S. E., Kingery, N. D., Ohsumi, T. K., Borowsky, M. L., Wang, L. C., Means, T. K., et al. (2013). The microglial sensome revealed by direct RNA sequencing. *Nat. Neurosci.* 16, 1896–1905. doi: 10.1038/nn.3554
- Hogervorst, E., Williams, J., Budge, M., Riedel, W., and Jolles, J. (2000). The nature of the effect of female gonadal hormone replacement therapy on cognitive function in post-menopausal women: a meta-analysis. *Neuroscience* 101, 485–512. doi: 10.1016/S0306-4522(00)00410-3
- Holland, D., Desikan, R. S., Dale, A. M., Mcevoy, L. K., and Alzheimer's Disease Neuroimaging, I. (2013). Higher rates of decline for women and apolipoprotein E epsilon4 carriers. *AJNR Am. J. Neuroradiol.* 34, 2287–2293. doi: 10.3174/ajnr. A3601
- Holtman, I. R., Raj, D. D., Miller, J. A., Schaafsma, W., Yin, Z., Brouwer, N., et al. (2015). Induction of a common microglia gene expression signature by aging and neurodegenerative conditions: a co-expression meta-analysis. *Acta Neuropathol. Commun.* 3:31. doi: 10.1186/s40478-015-0203-5
- Ishihara, Y., Itoh, K., Ishida, A., and Yamazaki, T. (2015). Selective estrogenreceptor modulators suppress microglial activation and neuronal cell death via an estrogen receptor-dependent pathway. J. Steroid Biochem. Mol. Biol. 145, 85–93. doi: 10.1016/j.jsbmb.2014.10.002
- Itagaki, S., Mcgeer, P. L., Akiyama, H., Zhu, S., and Selkoe, D. (1989). Relationship of microglia and astrocytes to amyloid deposits of Alzheimer disease. *J. Neuroimmunol.* 24, 173–182. doi: 10.1016/0165-5728(89)90115-X
- Jack, C. R., Knopman, D. S., Jagust, W. J., Petersen, R. C., Weiner, M. W., Aisen, P. S., et al. (2013). Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers. *Lancet Neurol.* 12, 207–216. doi: 10.1016/S1474-4422(12)70291-0
- Jacobs, E. G., Kroenke, C., Lin, J., Epel, E. S., Kenna, H. A., Blackburn, E. H., et al. (2013). Accelerated cell aging in female APOE-epsilon4 carriers: implications for hormone therapy use. *PLoS One* 8:e54713. doi: 10.1371/journal.pone. 0054713
- Jofre-Monseny, L., Minihane, A. M., and Rimbach, G. (2008). Impact of apoE genotype on oxidative stress, inflammation and disease risk. *Mol. Nutr. Food Res.* 52, 131–145. doi: 10.1002/mnfr.200700322
- Johnson, L. A., Kim, H. S., Knudson, M. J., Nipp, C. T., Yi, X., and Maeda, N. (2013). Diabetic atherosclerosis in APOE\*4 mice: synergy between lipoprotein metabolism and vascular inflammation. J. Lipid Res. 54, 386–396. doi: 10.1194/ jlr.M031435
- Kamada, M., Irahara, M., Maegawa, M., Yasui, T., Takeji, T., Yamada, M., et al. (2000). Effect of hormone replacement therapy on post-menopausal changes of lymphocytes and T cell subsets. *J. Endocrinol. Invest.* 23, 376–382. doi: 10.1007/ BF03343741
- Kamada, M., Irahara, M., Maegawa, M., Yasui, T., Yamano, S., Yamada, M., et al. (2001). B cell subsets in postmenopausal women and the effect of hormone replacement therapy. *Maturitas* 37, 173–179. doi: 10.1016/S0378-5122(00) 00180-8
- Keren-Shaul, H., Spinrad, A., Weiner, A., Matcovitch-Natan, O., Dvir-Szternfeld, R., Ulland, T. K., et al. (2017). A unique microglia type associated with restricting development of Alzheimer's disease. *Cell* 169, 1276–1290.e17. doi: 10.1016/j.cell.2017.05.018
- Khan, A. M., Babcock, A. A., Saeed, H., Myhre, C. L., Kassem, M., and Finsen, B. (2015). Telomere dysfunction reduces microglial numbers without fully inducing an aging phenotype. *Neurobiol. Aging* 36, 2164–2175. doi: 10. 1016/j.neurobiolaging.2015.03.008
- Khan, D., and Ansar Ahmed, S. (2015). The immune system is a natural target for estrogen action: opposing effects of estrogen in two prototypical autoimmune diseases. *Front. Immunol.* 6:635. doi: 10.3389/fimmu.2015.00635
- Kim, Y. S., and Joh, T. H. (2006). Microglia, major player in the brain inflammation: their roles in the pathogenesis of Parkinson's disease. *Exp. Mol. Med.* 38, 333–347. doi: 10.1038/emm.2006.40
- Kireev, R. A., Vara, E., Vina, J., and Tresguerres, J. A. (2014). Melatonin and oestrogen treatments were able to improve neuroinflammation and apoptotic

processes in dentate gyrus of old ovariectomized female rats. Age 36:9707. doi: 10.1007/s11357-014-9707-3

- Klosinski, L. P., Yao, J., Yin, F., Fonteh, A. N., Harrington, M. G., Christensen, T. A., et al. (2015). White matter lipids as a ketogenic fuel supply in aging female brain: implications for Alzheimer's disease. *EBioMedicine* 2, 1888–1904. doi: 10.1016/j.ebiom.2015.11.002
- Krasemann, S., Madore, C., Cialic, R., Baufeld, C., Calcagno, N., El Fatimy, R., et al. (2017). The TREM2-APOE pathway drives the transcriptional phenotype of dysfunctional microglia in neurodegenerative diseases. *Immunity* 47, 566– 581.e9. doi: 10.1016/j.immuni.2017.08.008
- Krstic, D., Madhusudan, A., Doehner, J., Vogel, P., Notter, T., Imhof, C., et al. (2012). Systemic immune challenges trigger and drive Alzheimer-like neuropathology in mice. J. Neuroinflammation 9:151. doi: 10.1186/1742-2094-9-151
- Kuh, D., Cooper, R., Moore, A., Richards, M., and Hardy, R. (2018). Age at menopause and lifetime cognition: findings from a British birth cohort study. *Neurology* 90, e1673–e1681. doi: 10.1212/WNL.00000000005486
- Kuszczyk, M. A., Sanchez, S., Pankiewicz, J., Kim, J., Duszczyk, M., Guridi, M., et al. (2013). Blocking the interaction between apolipoprotein E and Abeta reduces intraneuronal accumulation of Abeta and inhibits synaptic degeneration. *Am. J. Pathol.* 182, 1750–1768. doi: 10.1016/j.ajpath.2013.01.034
- Lai, A. Y., Dibal, C. D., Armitage, G. A., Winship, I. R., and Todd, K. G. (2013). Distinct activation profiles in microglia of different ages: a systematic study in isolated embryonic to aged microglial cultures. *Neuroscience* 254, 185–195. doi: 10.1016/j.neuroscience.2013.09.010
- Larbi, A., Franceschi, C., Mazzatti, D., Solana, R., Wikby, A., and Pawelec, G. (2008). Aging of the immune system as a prognostic factor for human longevity. *Physiology* 23, 64–74. doi: 10.1152/physiol.00040.2007
- Larson, I. A., Ordovas, J. M., Deluca, C., Barnard, J. R., Feussner, G., and Schaefer, E. J. (2000). Association of apolipoprotein (Apo)E genotype with plasma apo E levels. *Atherosclerosis* 148, 327–335. doi: 10.1016/S0021-9150(99)00280-4
- Latta, C. H., Brothers, H. M., and Wilcock, D. M. (2014). Neuroinflammation in Alzheimer's disease; a source of heterogeneity and target for personalized therapy. *Neuroscience* 302, 103–111. doi: 10.1016/j.neuroscience.2014.09.061
- Latta, C. H., Sudduth, T. L., Weekman, E. M., Brothers, H. M., Abner, E. L., Popa, G. J., et al. (2015). Determining the role of IL-4 induced neuroinflammation in microglial activity and amyloid-beta using BV2 microglial cells and APP/PS1 transgenic mice. J. Neuroinflammation 12:41. doi: 10.1186/s12974-015-0243-6
- Lee, C. Y., Tse, W., Smith, J. D., and Landreth, G. E. (2012). Apolipoprotein E promotes beta-amyloid trafficking and degradation by modulating microglial cholesterol levels. J. Biol. Chem. 287, 2032–2044. doi: 10.1074/jbc.M111.295451
- Lefterov, I., Bookout, A., Wang, Z., Staufenbiel, M., Mangelsdorf, D., and Koldamova, R. (2007). Expression profiling in APP23 mouse brain: inhibition of Abeta amyloidosis and inflammation in response to LXR agonist treatment. *Mol. Neurodegener.* 2:20. doi: 10.1186/1750-1326-2-20
- Li, L., Ruau, D., Chen, R., Weber, S., and Butte, A. J. (2012). "Systematic identification of risk factors for alzheimer's disease through shared genetic architecture and electronic medical records," in *Proceedings of the Pacific Symposium on Biocomputing 2013.* (Singapore: World Scientific), 224–235. doi: 10.1142/9789814447973\_0022
- Li, M. D., Burns, T. C., Kumar, S., Morgan, A. A., Sloan, S. A., and Palmer, T. D. (2015). Aging-like changes in the transcriptome of irradiated microglia. *Glia* 63, 754–767. doi: 10.1002/glia.22782
- Li, X., Montine, K. S., Keene, C. D., and Montine, T. J. (2015). Different mechanisms of apolipoprotein E isoform-dependent modulation of prostaglandin E(2) production and triggering receptor expressed on myeloid cells 2 (TREM2) expression after innate immune activation of microglia. *FASEB* J. 29, 1754–1762. doi: 10.1096/fj.14-262683
- Lima, T. A., Adler, A. L., Minett, T., Matthews, F. E., Brayne, C., Marioni, R. E., et al. (2014). C-reactive protein, APOE genotype and longitudinal cognitive change in an older population. Age Ageing 43, 289–292. doi: 10.1093/ageing/aft193
- Liu, C. C., Kanekiyo, T., Xu, H., and Bu, G. (2013). Apolipoprotein E and Alzheimer disease: risk, mechanisms and therapy. *Nat. Rev. Neurol.* 9, 106–118. doi: 10. 1038/nrneurol.2012.263
- Liu, H.-B., Loo, K. K., Palaszynski, K., Ashouri, J., Lubahn, D. B., and Voskuhl, R. R. (2003). Estrogen receptor α mediates estrogen's immune protection in autoimmune disease. *J. Immunol.* 171, 6936–6940. doi: 10.4049/jimmunol.171. 12.6936

- Liu, M., Bian, C., Zhang, J., and Wen, F. (2014). Apolipoprotein E gene polymorphism and Alzheimer's disease in Chinese population: a meta-analysis. *Sci. Rep.* 4:4383. doi: 10.1038/srep04383
- Liu, S., Breitbart, A., Sun, Y., Mehta, P. D., Boutajangout, A., Scholtzova, H., et al. (2014). Blocking the apolipoprotein E/amyloid beta interaction in triple transgenic mice ameliorates Alzheimer's disease related amyloid beta and tau pathology. J. Neurochem. 128, 577–591. doi: 10.1111/jnc.12484
- Maki, P. M. (2013). Critical window hypothesis of hormone therapy and cognition: a scientific update on clinical studies. *Menopause* 20, 695–709. doi: 10.1097/ GME.0b013e3182960cf8
- Mandrekar-Colucci, S., Karlo, J. C., and Landreth, G. E. (2012). Mechanisms underlying the rapid peroxisome proliferator-activated receptor-gammamediated amyloid clearance and reversal of cognitive deficits in a murine model of Alzheimer's disease. J. Neurosci. 32, 10117–10128. doi: 10.1523/JNEUROSCI. 5268-11.2012
- Manly, J. J., Merchant, C. A., Jacobs, D. M., Small, S. A., Bell, K., Ferin, M., et al. (2000). Endogenous estrogen levels and Alzheimer's disease among postmenopausal women. *Neurology* 54, 833–837. doi: 10.1212/WNL.54.4.833
- Manning, E. N., Barnes, J., Cash, D. M., Bartlett, J. W., Leung, K. K., Ourselin, S., et al. (2014). APOE 64 is associated with disproportionate progressive hippocampal atrophy in AD. *PLoS One* 9:e97608. doi: 10.1371/journal.pone. 0097608
- Marz, W., Scharnagl, H., Hoffmann, M., Boehm, B., and Winkelmann, B. (2004). The apolipoprotein E polymorphism is associated with circulating C-reactive protein (the Ludwigshafen risk and cardiovascular health study). *Eur. Heart J.* 25, 2109–2119. doi: 10.1016/j.ehj.2004.08.024
- Mattiace, L. A., Davies, P., and Dickson, D. W. (1990). Detection of HLA-DR on microglia in the human brain is a function of both clinical and technical factors. *Am. J. Pathol.* 136, 1101–1114.
- McGeer, P. L., Itagaki, S., Boyes, B. E., and Mcgeer, E. G. (1988). Reactive microglia are positive for HLA-DR in the substantia nigra of Parkinson's and Alzheimer's disease brains. *Neurology* 38, 1285–1291. doi: 10.1212/WNL.38.8. 1285
- McGeer, P. L., Itagaki, S., Tago, H., and Mcgeer, E. G. (1987). Reactive microglia in patients with senile dementia of the Alzheimer type are positive for the histocompatibility glycoprotein HLA-DR. *Neurosci. Lett.* 79, 195–200. doi: 10. 1016/0304-3940(87)90696-3
- McGeer, P. L., Schulzer, M., and Mcgeer, E. G. (1996). Arthritis and antiinflammatory agents as possible protective factors for Alzheimer's disease: a review of 17 epidemiologic studies. *Neurology* 47, 425–432. doi: 10.1212/WNL. 47.2.425
- Metti, A. L., Yaffe, K., Boudreau, R. M., Simonsick, E. M., Carnahan, R. M., Satterfield, S., et al. (2014). Trajectories of inflammatory markers and cognitive decline over 10 years. *Neurobiol. Aging* 35, 2785–2790. doi: 10.1016/j. neurobiolaging.2014.05.030
- Miller, Y. I., Choi, S. H., Wiesner, P., Fang, L., Harkewicz, R., Hartvigsen, K., et al. (2011). Oxidation-specific epitopes are danger-associated molecular patterns recognized by pattern recognition receptors of innate immunity. *Circ. Res.* 108, 235–248. doi: 10.1161/CIRCRESAHA.110.223875
- Miyata, M., and Smith, J. D. (1996). Apolipoprotein E allele-specific antioxidant activity and effects on cytotoxicity by oxidative insults and [beta]-amyloid peptides. *Nat. Genet.* 14, 55–61. doi: 10.1038/ng0996-55
- Morrison, J. H., Brinton, R. D., Schmidt, P. J., and Gore, A. C. (2006). Estrogen, menopause, and the aging brain: how basic neuroscience can inform hormone therapy in women. J. Neurosci. 26, 10332–10348. doi: 10.1523/JNEUROSCI. 3369-06.2006
- Mosconi, L., Berti, V., Guyara-Quinn, C., Mchugh, P., Petrongolo, G., Osorio, R. S., et al. (2017a). Perimenopause and emergence of an Alzheimer's bioenergetic phenotype in brain and periphery. *PLoS One* 12:e0185926. doi: 10.1371/journal. pone.0185926
- Mosconi, L., Berti, V., Quinn, C., Mchugh, P., Petrongolo, G., Varsavsky, I., et al. (2017b). Sex differences in Alzheimer risk: brain imaging of endocrine vs chronologic aging. *Neurology* 89, 1382–1390. doi: 10.1212/WNL. 000000000004425
- Mulder, S. D., Nielsen, H. M., Blankenstein, M. A., Eikelenboom, P., and Veerhuis, R. (2014). Apolipoproteins E and J interfere with amyloid-beta uptake by primary human astrocytes and microglia in vitro. *Glia* 62, 493–503. doi: 10.1002/glia.22619

- Muller, G. C., Gottlieb, M. G. V., Luz Correa, B., Filho, I. G., Moresco, R. N., and Bauer, M. E. (2015). The inverted CD4:CD8 ratio is associated with genderrelated changes in oxidative stress during aging. *Cell. Immunol.* 296, 149–154. doi: 10.1016/j.cellimm.2015.05.006
- Nelson, A. R., Sweeney, M. D., Sagare, A. P., and Zlokovic, B. V. (2016). Neurovascular dysfunction and neurodegeneration in dementia and Alzheimer's disease. *Biochim. Biophys. Acta* 1862, 887–900. doi: 10.1016/j.bbadis.2015.12.016
- Neu, S. C., Pa, J., Kukull, W., Beekly, D., Kuzma, A., Gangadharan, P., et al. (2017). Apolipoprotein E genotype and sex risk factors for alzheimer disease: a meta-analysis. *JAMA Neurol.* 74, 1178–1189. doi: 10.1001/jamaneurol.2017. 2188
- Ojo, J. O., Rezaie, P., Gabbott, P. L., and Stewart, M. G. (2015). Impact of agerelated neuroglial cell responses on hippocampal deterioration. *Front. Aging Neurosci.* 7:57. doi: 10.3389/fnagi.2015.00057
- Olgiati, P., Politis, A., Malitas, P., Albani, D., Dusi, S., Polito, L., et al. (2010). APOE epsilon-4 allele and cytokine production in Alzheimer's disease. *Int. J. Geriatr. Psychiatry* 25, 338–344. doi: 10.1002/gps.2344
- Olsson, B., Hertze, J., Lautner, R., Zetterberg, H., Nagga, K., Hoglund, K., et al. (2013). Microglial markers are elevated in the prodromal phase of Alzheimer's disease and vascular dementia. *J. Alzheimers Dis.* 33, 45–53. doi: 10.3233/JAD-2012-120787
- Orre, M., Kamphuis, W., Osborn, L. M., Melief, J., Kooijman, L., Huitinga, I., et al. (2014). Acute isolation and transcriptome characterization of cortical astrocytes and microglia from young and aged mice. *Neurobiol. Aging* 35, 1–14. doi: 10.1016/j.neurobiolaging.2013.07.008
- Oury, C. (2014). CD36: linking lipids to the NLRP3 inflammasome, atherogenesis and atherothrombosis. *Cell. Mol. Immunol.* 11, 8–10. doi: 10.1038/cmi.2013.48
- Perry, V. H., and Teeling, J. (2013). Microglia and macrophages of the central nervous system: the contribution of microglia priming and systemic inflammation to chronic neurodegeneration. *Semin. Immunopathol.* 35, 601–612. doi: 10.1007/s00281-013-0382-8
- Persad, C. C., Zubieta, J. K., Love, T., Wang, H., Tkaczyk, A., and Smith, Y. R. (2009). Enhanced neuroactivation during verbal memory processing in postmenopausal women receiving short-term hormone therapy. *Fertil. Steril.* 92, 197–204. doi: 10.1016/j.fertnstert.2008.04.040
- Policicchio, S., Ahmad, A. N., Powell, J. F., and Proitsi, P. (2017). Rheumatoid arthritis and risk for Alzheimer's disease: a systematic review and meta-analysis and a Mendelian Randomization study. *Sci. Rep.* 7:12861. doi: 10.1038/s41598-017-13168-8
- Rajan, K. B., Wilson, R. S., Weuve, J., Barnes, L. L., and Evans, D. A. (2015). Cognitive impairment 18 years before clinical diagnosis of Alzheimer disease dementia. *Neurology* 85, 898–904. doi: 10.1212/WNL.000000000001774
- Reichwald, J., Danner, S., Wiederhold, K. H., and Staufenbiel, M. (2009). Expression of complement system components during aging and amyloid deposition in APP transgenic mice. J. Neuroinflammation 6:35. doi: 10.1186/ 1742-2094-6-35
- Rettberg, J. R., Yao, J., and Brinton, R. D. (2014). Estrogen: a master regulator of bioenergetic systems in the brain and body. *Front. Neuroendocrinol.* 35, 8–30. doi: 10.1016/j.yfrne.2013.08.001
- Ritzel, R. M., Patel, A. R., Pan, S., Crapser, J., Hammond, M., Jellison, E., et al. (2015). Age- and location-related changes in microglial function. *Neurobiol. Aging* 36, 2153–2163. doi: 10.1016/j.neurobiolaging.2015.02.016
- Rocca, W. A., Grossardt, B. R., and Maraganore, D. M. (2008). The long-term effects of oophorectomy on cognitive and motor aging are age dependent. *Neurodegener. Dis.* 5, 257–260. doi: 10.1159/000113718
- Safaiyan, S., Kannaiyan, N., Snaidero, N., Brioschi, S., Biber, K., Yona, S., et al. (2016). Age-related myelin degradation burdens the clearance function of microglia during aging. *Nat. Neurosci.* 19, 995–998. doi: 10.1038/nn.4325
- Salminen, A., Kaarniranta, K., and Kauppinen, A. (2012). Inflammaging: disturbed interplay between autophagy and inflammasomes. *Aging* 4, 166–175. doi: 10. 18632/aging.100444
- Sárvári, M., Hrabovszky, E., Kalló, I., Solymosi, N., Likó, I., Berchtold, N., et al. (2012). Menopause leads to elevated expression of macrophage-associated genes in the aging frontal cortex: rat and human studies identify strikingly similar changes. J. Neuroinflammation 9:264. doi: 10.1186/1742-2094-9-264
- Sárvári, M., Hrabovszky, E., Kalló, I., Solymosi, N., Tóth, K., Likó, I., et al. (2011). Estrogens regulate neuroinflammatory genes via estrogen receptors  $\alpha$  and  $\beta$

in the frontal cortex of middle-aged female rats. J. Neuroinflammation 8:82. doi: 10.1186/1742-2094-8-82

- Sarvari, M., Kallo, I., Hrabovszky, E., Solymosi, N., Rodolosse, A., Vastagh, C., et al. (2015). Hippocampal gene expression is highly responsive to estradiol replacement in middle-aged female rats. *Endocrinology* 156, 2632–2645. doi: 10.1210/en.2015-1109
- Scacchi, R., De Bernardini, L., Mantuano, E., Donini, L. M., Vilardo, T., and Corbo, R. M. (1995). Apolipoprotein E (APOE) allele frequencies in late-onset sporadic Alzheimer's disease (AD), mixed dementia and vascular dementia: lack of association of ε4 allele with AD in Italian octogenarian patients. *Neurosci. Lett.* 201, 231–234. doi: 10.1016/0304-3940(95)12190-0
- Sharma, N., Sharma, R. K., Tewari, S., Chauhan, M., and Bhatia, A. (2018). Association of periodontal inflammation, systemic inflammation, and duration of menopausal years in postmenopausal women. *Quintessence Int.* 49, 123–131.
- Shaw, A. C., Panda, A., Joshi, S. R., Qian, F., Allore, H. G., and Montgomery, R. R. (2011). Dysregulation of human Toll-like receptor function in aging. *Ageing Res. Rev.* 10, 346–353. doi: 10.1016/j.arr.2010.10.007
- Sheedy, F. J., Grebe, A., Rayner, K. J., Kalantari, P., Ramkhelawon, B., Carpenter, S. B., et al. (2013). CD36 coordinates NLRP3 inflammasome activation by facilitating intracellular nucleation of soluble ligands into particulate ligands in sterile inflammation. *Nat. Immunol.* 14, 812–820. doi: 10.1038/ni.2639
- Shumaker, S. A., Legault, C., Rapp, S. R., Thal, L., Wallace, R. B., Ockene, J. K., et al. (2003). Estrogen plus progestin and the incidence of dementia and mild cognitive impairment in postmenopausal women: the Women's Health Initiative Memory Study: a randomized controlled trial. *JAMA* 289, 2651–2662. doi: 10.1001/jama.289.20.2651
- Silverstein, R. L., and Febbraio, M. (2009). CD36, a scavenger receptor involved in immunity, metabolism, angiogenesis, and behavior. *Sci. Signal.* 2:re3. doi: 10.1126/scisignal.272re3
- Sima, A., Iordan, A., and Stancu, C. (2007). Apolipoprotein E polymorphisma risk factor for metabolic syndrome. *Clin. Chem. Lab. Med.* 45, 1149–1153. doi: 10.1515/CCLM.2007.258
- Slowik, A., and Beyer, C. (2015). Inflammasomes are neuroprotective targets for sex steroids. J. Steroid Biochem. Mol. Biol. 153, 135–143. doi: 10.1016/j.jsbmb. 2015.02.013
- Smith, A. M., and Dragunow, M. (2014). The human side of microglia. Trends Neurosci. 37, 125–135. doi: 10.1016/j.tins.2013.12.001
- Smith, R. G., Betancourt, L., and Sun, Y. (2005). Molecular endocrinology and physiology of the aging central nervous system. *Endocr. Rev.* 26, 203–250. doi: 10.1210/er.2002-0017
- Spence, R. D., Hamby, M. E., Umeda, E., Itoh, N., Du, S., Wisdom, A. J., et al. (2011). Neuroprotection mediated through estrogen receptor-α in astrocytes. *Proc. Natl. Acad. Sci. U.S.A.* 108, 8867–8872. doi: 10.1073/pnas.1103833108
- Stanley, L. C., Mrak, R. E., Woody, R. C., Perrot, L. J., Zhang, S., Marshak, D. R., et al. (1994). Glial cytokines as neuropathogenic factors in HIV infection: pathogenic similarities to Alzheimer's disease. *J. Neuropathol. Exp. Neurol.* 53, 231–238. doi: 10.1097/00005072-199405000-00003
- Streit, W. J., Mrak, R. E., and Griffin, W. S. (2004). Microglia and neuroinflammation: a pathological perspective. J. Neuroinflammation 1:14. doi: 10.1186/1742-2094-1-14
- Streit, W. J., and Xue, Q. S. (2014). Human CNS immune senescence and neurodegeneration. Curr. Opin. Immunol. 29, 93–96. doi: 10.1016/j.coi.2014. 05.005
- Suenaga, J., Hu, X., Pu, H., Shi, Y., Hassan, S. H., Xu, M., et al. (2015). White matter injury and microglia/macrophage polarization are strongly linked with age-related long-term deficits in neurological function after stroke. *Exp. Neurol.* 272, 109–119. doi: 10.1016/j.expneurol.2015.03.021
- Tai, L. M., Ghura, S., Koster, K. P., Liakaite, V., Maienschein-Cline, M., Kanabar, P., et al. (2015). APOE-modulated Abeta-induced neuroinflammation in Alzheimer's disease: current landscape, novel data, and future perspective. *J. Neurochem.* 133, 465–488. doi: 10.1111/jnc.13072
- Tai, L. M., Koster, K. P., Luo, J., Lee, S. H., Wang, Y. T., Collins, N. C., et al. (2014). Amyloid-beta pathology and APOE genotype modulate retinoid X receptor agonist activity in vivo. *J. Biol. Chem.* 289, 30538–30555. doi: 10.1074/jbc.M114. 600833
- Tang, M. -X., Jacobs, D., Stern, Y., Marder, K., Schofield, P., Gurland, B., et al. (1996). Effect of oestrogen during menopause on risk and age at onset of Alzheimer's disease. *Lancet* 348, 429–432. doi: 10.1016/S0140-6736(96)03356-9

- Tang, Y., Purkayastha, S., and Cai, D. (2015). Hypothalamic microinflammation: a common basis of metabolic syndrome and aging. *Trends Neurosci.* 38, 36–44. doi: 10.1016/j.tins.2014.10.002
- Tapia-Gonzalez, S., Carrero, P., Pernia, O., Garcia-Segura, L. M., and Diz-Chaves, Y. (2008). Selective oestrogen receptor (ER) modulators reduce microglia reactivity in vivo after peripheral inflammation: potential role of microglial ERs. J. Endocrinol. 198, 219–230. doi: 10.1677/JOE-07-0294
- Terwel, D., Steffensen, K. R., Verghese, P. B., Kummer, M. P., Gustafsson, J. A., Holtzman, D. M., et al. (2011). Critical role of astroglial apolipoprotein E and liver X receptor-alpha expression for microglial Abeta phagocytosis. *J. Neurosci.* 31, 7049–7059. doi: 10.1523/JNEUROSCI.6546-10.2011
- Thal, L. J., Ferris, S. H., Kirby, L., Block, G. A., Lines, C. R., Yuen, E., et al. (2005). A randomized, double-blind, study of rofecoxib in patients with mild cognitive impairment. *Neuropsychopharmacology* 30, 1204–1215. doi: 10.1038/ sj.npp.1300690
- Toms, T. E., Smith, J. P., Panoulas, V. F., Blackmore, H., Douglas, K. M., and Kitas, G. D. (2012). Apolipoprotein E gene polymorphisms are strong predictors of inflammation and dyslipidemia in rheumatoid arthritis. *J. Rheumatol.* 39, 218–225. doi: 10.3899/jrheum.110683
- Tutuncu, M., Tang, J., Zeid, N. A., Kale, N., Crusan, D. J., Atkinson, E. J., et al. (2013). Onset of progressive phase is an age dependent clinical milestone in multiple sclerosis. *Mult. Scler.* 19, 188–198. doi: 10.1177/1352458512451510
- Ukkola, O., Kunnari, A., Jokela, M., Paivansalo, M., and Kesaniemi, Y. A. (2009). ApoE phenotype is associated with inflammatory markers in middle-aged subjects. *Inflamm. Res.* 58, 54–59. doi: 10.1007/s00011-008-8215-2
- Ungar, L., Altmann, A., and Greicius, M. D. (2014). Apolipoprotein E, gender, and Alzheimer's disease: an overlooked, but potent and promising interaction. *Brain Imaging Behav.* 8, 262–273. doi: 10.1007/s11682-013-9272-x
- Uri-Belapolsky, S., Shaish, A., Eliyahu, E., Grossman, H., Levi, M., Chuderland, D., et al. (2014). Interleukin-1 deficiency prolongs ovarian lifespan in mice. *Proc. Natl. Acad. Sci. U.S.A.* 111, 12492–12497. doi: 10.1073/pnas.1323955111
- Vadasz, Z., Haj, T., Kessel, A., and Toubi, E. (2013). Age-related autoimmunity. BMC Med. 11:94. doi: 10.1186/1741-7015-11-94
- Van Den Heuvel, C., Thornton, E., and Vink, R. (2007). Traumatic brain injury and Alzheimer's disease: a review. *Prog. Brain Res.* 161, 303–316. doi: 10.1016/ S0079-6123(06)61021-2
- Vandanmagsar, B., Youm, Y.-H., Ravussin, A., Galgani, J. E., Stadler, K., Mynatt, R. L., et al. (2011). The NALP3/NLRP3 inflammasome instigates obesityinduced autoinflammation and insulin resistance. *Nat. Med.* 17, 179–188. doi: 10.1038/nm.2279
- VanGuilder, H. D., Bixler, G. V., Brucklacher, R. M., Farley, J. A., Yan, H., Warrington, J. P., et al. (2011). Concurrent hippocampal induction of MHC II pathway components and glial activation with advanced aging is not correlated with cognitive impairment. *J. Neuroinflammation* 8:138. doi: 10.1186/1742-2094-8-138
- Vegeto, E., Benedusi, V., and Maggi, A. (2008). Estrogen anti-inflammatory activity in brain: a therapeutic opportunity for menopause and neurodegenerative diseases. *Front. Neuroendocrinol.* 29, 507–519. doi: 10.1016/j.yfrne.2008.04.001
- Vitek, M. P., Brown, C. M., and Colton, C. A. (2009). APOE genotype-specific differences in the innate immune response. *Neurobiol. Aging* 30, 1350–1360. doi: 10.1016/j.neurobiolaging.2007.11.014
- Vlad, S. C., Miller, D. R., Kowall, N. W., and Felson, D. T. (2008). Protective effects of NSAIDs on the development of Alzheimer disease. *Neurology* 70, 1672–1677. doi: 10.1212/01.wnl.0000311269.57716.63
- von Bernhardi, R., Eugenin-Von Bernhardi, L., and Eugenin, J. (2015). Microglial cell dysregulation in brain aging and neurodegeneration. *Front. Aging Neurosci.* 7:124. doi: 10.3389/fnagi.2015.00124
- Walsh, J. G., Muruve, D. A., and Power, C. (2014). Inflammasomes in the CNS. *Nat. Rev. Neurosci.* 15, 84–97. doi: 10.1038/nrn3638
- Wilson, R. S., Leurgans, S. E., Boyle, P. A., and Bennett, D. A. (2011). Cognitive decline in prodromal Alzheimer disease and mild cognitive impairment. *Arch. Neurol.* 68, 351–356. doi: 10.1001/archneurol.2011.31
- Wirz, K. T., Keitel, S., Swaab, D. F., Verhaagen, J., and Bossers, K. (2014). Early molecular changes in Alzheimer disease: can we catch the disease in its presymptomatic phase? *J. Alzheimers Dis.* 38, 719–740. doi: 10.3233/JAD-130920
- Wotton, C. J., and Goldacre, M. J. (2017). Associations between specific autoimmune diseases and subsequent dementia: retrospective record-linkage

cohort study, UK. J. Epidemiol. Commun. Health 71, 576-583. doi: 10.1136/ jech-2016-207809

- Xiang, Z., Haroutunian, V., Ho, L., Purohit, D., and Pasinetti, G. M. (2006). Microglia activation in the brain as inflammatory biomarker of Alzheimer's disease neuropathology and clinical dementia. *Dis. Markers* 22, 95–102. doi: 10.1155/2006/276239
- Xie, F., Zhang, J. C., Fu, H., and Chen, J. (2013). Age-related decline of myelin proteins is highly correlated with activation of astrocytes and microglia in the rat CNS. *Int. J. Mol. Med.* 32, 1021–1028. doi: 10.3892/ijmm.2013.1486
- Xu, J., and Ikezu, T. (2009). The comorbidity of HIV-associated neurocognitive disorders and Alzheimer's disease: a foreseeable medical challenge in post-HAART Era. J. Neuroimmune Pharmacol. 4, 200–212. doi: 10.1007/s11481-008-9136-0
- Yao, J., Hamilton, R. T., Cadenas, E., and Brinton, R. D. (2010). Decline in mitochondrial bioenergetics and shift to ketogenic profile in brain during reproductive senescence. *Biochim. Biophys. Acta* 1800, 1121–1126. doi: 10.1016/ j.bbagen.2010.06.002
- Yasui, T., Maegawa, M., Tomita, J., Miyatani, Y., Yamada, M., Uemura, H., et al. (2007). Changes in serum cytokine concentrations during the menopausal transition. *Maturitas* 56, 396–403. doi: 10.1016/j.maturitas.2006. 11.002
- Yin, F., Yao, J., Sancheti, H., Feng, T., Melcangi, R. C., Morgan, T. E., et al. (2015). The perimenopausal aging transition in the female rat brain: decline in bioenergetic systems and synaptic plasticity. *Neurobiol. Aging* 36, 2282–2295. doi: 10.1016/j.neurobiolaging.2015.03.013
- Youm, Y. H., Grant, R. W., Mccabe, L. R., Albarado, D. C., Nguyen, K. Y., Ravussin, A., et al. (2013). Canonical Nlrp3 inflammasome links systemic lowgrade inflammation to functional decline in aging. *Cell Metab.* 18, 519–532. doi: 10.1016/j.cmet.2013.09.010
- Youm, Y. H., Kanneganti, T. D., Vandanmagsar, B., Zhu, X., Ravussin, A., Adijiang, A., et al. (2012). The Nlrp3 inflammasome promotes age-related thymic demise and immunosenescence. *Cell Rep.* 1, 56–68. doi: 10.1016/j.celrep. 2011.11.005
- Youm, Y. H., Nguyen, K. Y., Grant, R. W., Goldberg, E. L., Bodogai, M., Kim, D., et al. (2015). The ketone metabolite beta-hydroxybutyrate blocks NLRP3 inflammasome-mediated inflammatory disease. *Nat. Med.* 21, 263–269. doi: 10.1038/nm.3804
- Yun, Y. W., Kweon, S. S., Choi, J. S., Rhee, J. A., Lee, Y. H., Nam, H. S., et al. (2015). APOE polymorphism is associated with c-reactive protein levels but not with white blood cell count: dong-gu study and namwon study. *J. Korean Med. Sci.* 30, 860–865. doi: 10.3346/jkms.2015.30.7.860
- Zandi, P. P., Carlson, M. C., Plassman, B. L., Welsh-Bohmer, K. A., Mayer, L. S., Steffens, D. C., et al. (2002). Hormone replacement therapy and incidence of Alzheimer disease in older women: the Cache County Study. *JAMA* 288, 2123–2129. doi: 10.1001/jama.288.17.2123
- Zhang, G., Li, J., Purkayastha, S., Tang, Y., Zhang, H., Yin, Y., et al. (2013). Hypothalamic programming of systemic ageing involving IKKbeta, NF-kappaB and GnRH. *Nature* 497, 211–216. doi: 10.1038/nature 12143
- Zhao, J., Jiang, C. Q., Lam, T. H., Liu, B., Cheng, K. K., Kavikondala, S., et al. (2014). Genetically predicted 17beta-estradiol and systemic inflammation in women: a separate-sample Mendelian randomisation analysis in the Guangzhou Biobank Cohort Study. J. Epidemiol. Commun. Health 68, 780–785. doi: 10.1136/jech-2013-203451

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer AG and handling Editor declared their shared affiliation at the time of the review.

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