



Links Between Adiponectin and Dementia: From Risk Factors to Pathophysiology

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With the aging population, dementia is becoming one of the most serious and troublesome global public health issues. Numerous studies have been seeking for effective strategies to delay or block its progression, but with little success. In recent years, adiponectin (APN) as one of the most abundant and multifunctional adipocytokines related to anti-inflammation, regulating glycogen metabolism and inhibiting insulin resistance (IR) and anti-atherosclerosis, has attracted widespread attention. In this article, we summarize recent studies that have contributed to a better understanding of the extent to which APN influences the risks of developing dementia as well as its pathophysiological progression. In addition, some controversial results interlinked with its effects on cognitive dysfunction diseases will be critically discussed. Ultimately, we aim to gain a novel insight into the pleiotropic effects of APN levels in circulation and suggest potential therapeutic target and future research strategies.

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INTRODUCTION

With the aging population, over 43.8 million people worldwide are suffering from dementia (GBD 2016 Dementia Collaborators, 2019). According to epidemiological statistics and estimates, by 2050, the population with dementia is predicted to triple worldwide (GBD 2016 Disease and Injury Incidence and Prevalence Collaborators, 2017). Furthermore among all chronic diseases, dementia is one of major causes leading to disability and dependance. Dementia will be one of the most serious and troublesome global public health issues (Alzheimer's Association, 2013; GBD 2016 Disease and Injury Incidence and Prevalence Collaborators, 2017). The two most common forms of dementia are Alzheimer's disease (AD) and vascular dementia (VD), accounting for 60% and 30% of dementia, respectively (Kalaria et al., 2008; Crous-Bou et al., 2017). Now, due to the rapid aging of the global population and the prolonged life expectancy, dementia has become a focus issue attracting global attention. This disease imposes a considerable burden on individuals, their families, and society.

AD and VD are distinct diseases with potential overlapping metabolic dysfunction. They share some common risk factors, pathogenesis, and clinical features (Haan and Wallace, 2004). For instance, aging, sex, genetic factors, rate, and vascular factors, including hypertension, diabetes, dietary fat intake, metabolic syndrome (MetS), high cholesterol, stroke, and exercise are their common risk factors (**Figure 1**); peripheral and neuroinflammatory, IR, energy metabolism

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disorder, and oxidative stress are their common pathogenesis; cognitive and behavioral dysfunctions are their common clinical features.

APN is one of the most abundant adipocytokines. APN plays an important role in regulating vasodilation, resisting inflammation and anti-arterial atherosclerosis, increasing insulin sensitivity, and regulating glycogen and lipid metabolism. In this review, we summarize and discuss the latest advances in the association between APN and dementia, from the risk factors to potential pathophysiological mechanisms of dementia, hoping to explore a new way to address the pathophysiological mechanism associated with dementia.

ADIPONECTIN

Adiponectin (APN) is a fat-derived hormone that was first isolated from rat adipocytes by Scherer et al. (1995). Extensive attention was attracted as a key messenger for a connection between adipose tissue and other metabolically related organs. It is a 30-kDa protein, also known as ACRP30, apM1, adipoQ, or GBP28, containing 244 amino acids with an N-terminal collagen-like domain and a C-terminal complement factor C1q-like globular domain (Turer and Scherer, 2012). There are three different complexes of APN in the blood circulation: hexamers, trimers, and high molecular form (HMW; Aso et al., 2006; Wang and Scherer, 2016). Different forms own different biological activities. In related diseases, HMW APN or HMW APN/total APN may be a more effective indicator of insulin sensitivity (Aso et al., 2006). It works by binding to three receptors: AdipoR1, AdipoR2, and T-Cadherin. The three receptors are commonly expressed throughout the body, including adipose tissue, skeletal muscle, liver, pancreas, heart, blood vessels, and endothelial cells (Kiliaan et al., 2014). APN receptors have also been found in hippocampus, hypothalamus, and brainstem, and AdipoR1 expression is more pronounced in the brain (Letra et al., 2019b).

APN participates in regulating fatty acid and glucose catabolism and sensitizing cellular to insulin (Gustafson, 2010; van Himbergen et al., 2012; Letra et al., 2019b). APN has been repeatedly reported that it is negatively correlated with IR, body mass index (BMI), type 2 diabetes mellitus (T2DM), and cardiovascular disease. All these factors can elevate dementia risk (Wang and Scherer, 2016). In addition, APN exerts anti-inflammatory effects by decreasing the production of pro-inflammatory cytokines, including interleukin-6 (IL-6), interferon γ (INF γ) and tumor necrosis factor α (TNF- α), meanwhile increasing interleukin-1 (IL-1) and interleukin-10 (IL-10) expression (Brochu-Gaudreau et al., 2010). Furthermore, it reduces IR in the brain, but also in peripheral organs. Moreover, the role of APN in dementia with neurodegenerative symptoms has been confirmed in many studies.

DEMENTIA

Dementia delineates a clinical syndrome that is characterized by a series of memory difficulties, language disorders, psychological and mental changes, and impaired activities of daily living (Burns and Iliffe, 2009). AD is an irreversible and disabling progressive neurodegenerative disease owing to neuronal and synaptic loss. It is sporadic or shows modest familial clustering. AD tends to be more insidious than VD. It is similar to the symptoms of "normal aging" memory loss or communication disorders, and personality or emotional changes in the early states (Robinson et al., 2015). The diagnostic criteria for AD have been recognized extensively. Hallmarks of neuropathology in AD include deposits of amyloid- β (A β) peptide extracellularly and accumulation of hyperphosphorylated *tau* (p-*tau*) in neurofibrillary tangles (NFTs; Dubois et al., 2016; Crous-Bou et al., 2017). Therefore, this provides detailed information on the molecular pathogenesis of AD. Then, recently, a great deal of effort have been undertaken targeting these pathological mechanisms in an attempt to find potential therapeutic point for AD, especially focusing on the metabolism of the A β peptide (Citron, 2010; De Strooper et al., 2010).

Although second only to AD, VD remains an important form of dementia plaguing the elderly population. It is a progressive disease caused by a decreased blood flow into the brain that affects cognitive ability as well as executive function. It is the most severe form of vascular cognitive dysfunction (VCI; Gorelick et al., 2011), which is defined as cognitive function alterations caused by various vascular factors (Hachinski and Bowler, 1993). According to the contribution of vascular pathology to dementia, VD is classified as different subtypes, including cortical VD, subcortical VD, strategic infarct dementia, hypoperfusion dementia, hemorrhagic dementia, hereditary VD (e.g., CADASIL), and AD with cardiovascular disease (O'Brien and Thomas, 2015). However, the most common causes of vascular brain injury or infarction are atherosclerosis and cardiogenic embolism, while the cognitive impairment caused by microinfarcts in the cortex and subcortex is mostly attributed to cerebral small vessel disease characterized by arteriosclerosis and lacunar infarction (O'Brien and Thomas, 2015). Accurate diagnosis of VD relies on assessment of clinical symptoms, neuropsychological measurements, and neuroimaging and final pathological confirmation (Kalaria, 2016). Numerous cerebrovascular disease (CVD) causes stroke injury and other tissue perfusion changes, along with neurocognitive disorders, behavioral symptoms, motor abnormalities, and autonomic dysfunction (Kalaria, 2018).

AD, VD, or CVD has common risk factors including obesity, hypertension, diabetes, IR, hyperlipidemia, and hyperhomocystinemia (Fillit et al., 2008; Craft, 2009; Purnell et al., 2009). Therefore, learning more about the risk factors and the mechanism factors contributing to dementia could promote new ways to prevent, improve, or delay the progression of dementia.

APN AND MULTIPLE RISK FACTORS OF DEMENTIA

Due to a tremendous increase in dementia and the lack of effective treatment, it is urgent to identify all the risk factors that accelerate or inhibit the cognitive impairment process. In many previous studies, several important risk factors were analyzed, including hypertension, diabetes mellitus, dyslipidemia and obesity at midlife, atherosclerosis, stroke, genes, and other factors (aging, low levels of education, smoking, depression and physical inactivity, etc.; Baumgart et al., 2015). AD and VD share the common risk factors especially vascular risk factors (Gardener et al., 2015). These common vascular risk factors indicate a correlation and potential interaction between

AD and vascular pathology; however, the deep mechanism is unclear.

Some studies have suggested that the clinical manifestations of dementia are serious when vascular diseases co-exist (Snowdon et al., 1997; Jellinger and Attems, 2015). Identifying and reducing risk factors are major strategies for primary prevention. Here, we review the current research progresses on the dementia risk factors, as summarized in **Figure 1** (Kalaria et al., 2008; Song et al., 2014; O'Brien and Thomas, 2015; Crous-Bou et al., 2017).

Aging

Aging is interrelated with progressive declines in physiological function, which leads to multiple chronic diseases and frailty (Kirkland, 2013). Undoubtedly, dementia is an age-related disease. Some epidemiological studies have shown that between 65 and 90 years old, even in people aged 90 years and older, the incidence of all-cause dementia increases exponentially with age and doubles every 5 years (Jorm and Jolley, 1998; Corrada et al., 2010), Systemic decline with aging is characterized by various alterations, including mitochondrial dysfunction, cellular senescence, metabolic declines, adipose tissue dysfunction, IR, chronic sterile inflammation, and dysregulated nutrient sensing (Stout et al., 2017). Notably, IR and chronic low-grade inflammation are two major features of aging that occur nearly in most age-related diseases, such as dementia, diabetes, arthritis, CVD, and cancer. Besides, along with aging, adipose tissues undergo significant changes in distribution, abundance, endocrine signaling, and cellular composition. They play a central role in the development of IR, inflammation, metabolic dysfunction, and regenerative capacity (Palmer and Kirkland, 2016). As one of the most abundant adipocytokines secreted by adipose tissue, APN has versatile properties to positively influence these fundamental aging mechanisms. It is well known as a modulator in improvement of insulin sensitivity, inhibition of systemic inflammation, regulation of lipid and glycogen metabolism, and reduction of atherosclerotic processes (Brochu-Gaudreau et al., 2010). Numerous epidemiologic researches have indicated that low-circulating levels of APN are associated with many age-related metabolic disorders including T2DM, obesity, and cardiovascular disease (Arita et al., 1999; Lindsay et al., 2002, 2003). Besides, macrophage is crucial to regulate the aging process and prominently contribute to inflammatory and immune responses, but also helps maintain metabolic homeostasis (Biswas and Mantovani, 2012). It is reported that APN expressed in macrophages improves insulin sensitivity and protects against inflammation and atherosclerosis (Luo et al., 2010). Reduced IR and inflammation appear to be closely associated with life span (Franceschi et al., 2005; Ewald et al., 2018). In addition, mitochondria impairment increases with age, leading to age-associated disease phenotypes and senescense (Kauppila et al., 2017). Iwabu et al. (2010) dedicated that decreased levels of APN and AdipoR1 in obesity may play causal roles in IR and mitochondrial dysfunction seen in diabetes. Several recent studies suggest that mitochondrial dysfunction is linked to

impairment of insulin sensitivity and decreased APN secretion in adipocytes (Koh et al., 2007; Wang C. H. et al., 2013). Considering these evidences, APN might be a key factor in aging pathway.

Hypertension

Emerging as a leading cause of age-related cognitive dysfunction, hypertension is known to be linked to VD, and is also associated with AD and other adverse cognitive outcomes (Crous-Bou et al., 2017; Iadecola and Gottesman, 2019). These evidences primarily come from many epidemiological studies, which strongly support the idea that hypertension is clearly correlated with steeper cognitive decline. But this adverse effect mainly occurs in middle age, early stage in dementia (Kennelly et al., 2009), while some studies show that later-life hypertension may help to prevent cognitive decline (Kennelly et al., 2009; Corrada et al., 2017). Deficit of nitric oxide (NO) derived from endothelial NO synthase (eNOS) and endothelial dysfunction is one of the vital mechanisms for hypertension to affect cerebral blood flow and lead to cognitive decline (Delles et al., 2004). Endothelial cells secrete a variety of vasodilators (e.g., NO, prostaglandins) and vasoconstrictors (e.g., endothelin ET, thromboxane A2), thereby regulating vasoconstriction and altering cerebral perfusion (Vanhoutte et al., 2017). Meanwhile, the endothelium is also essential to maintain the blood-brain barrier (BBB) for bidirectional molecular transmission between brain and other organs of the body (Sweeney et al., 2019) and exerts crucial trophic effects on brain cells (Marie et al., 2018). Previous studies demonstrated that hypertension causes severe damage to vascular endothelial cells, which in turn leads to BBB permeability changes, dysfunction of cerebral vascular and perivascular, and brain structural failure, including subclinical brain infarcts, white matter hyperintensities, and cerebral microbleeds (Iadecola and Gottesman, 2019). Furthermore, vascular injury and perivascular dysfunction caused by hypertension may impair the $A\beta$ disposal and accumulation. Conversely, loss of NO bioavailability associated with vascular dysfunction may also contribute to the promotion of increased AB and amyloid formation, which is a marker in AD. APN decreases the risk of hypertension and improves cognitive impairment by stimulating NO production. It promotes NO release via AdipoR1 and AdipoR2 and inhibits cerebral inflammatory response through adenosine monophosphateactivated protein kinase (AMPK)/eNOS signaling pathway activation (Shibata et al., 2004). Increased NO reduces platelet aggregation and elevates vasodilation regulating cerebrovascular microcirculation. In addition, APN could suppress amyloid- β in mice (Jian et al., 2019). Waragai et al. (2016) have suggested a possible positive effect of APN by finding that higher cerebrospinal fluid (CSF) level of APN is related with lower amyloid and tau burden. Consequentially, APN could decrease the risk of hypertension and improve vascular cognitive impairment. Despite this, the interaction between APN and hypertension, one of the risk factors in dementia, still has many unsolved mysteries that require further reveals.

Obesity

Obesity is another independent risk factor of dementia in the diverse adult urban population. Traditionally, obesity refers to an increase in whole body mass, which cannot analyze body composition or distinguish between subcutaneous and visceral fat, while excessive accumulation of visceral fat can better illustrate metabolic abnormalities (Duvnjak and Duvnjak, 2009).

Overweight and obesity are the cornerstones of vascular risk contributed to various diseases. BMI, waist circumference, and waist-to-hip ratio are widely quantitative measurement methods to assess obesity, and the BMI is the most effectively popularized in clinic (over 30 kg/m²). Significant increase in abdominal circumference means central adiposity or abdominal obesity, which is more associated with visceral fat deposits and has a stronger relationship with adverse metabolic outcomes (Luchsinger, 2008). Abdominal obesity, hypertension, dyslipidemia, and IR are collectively defined as MetS. MetS is a systemic inflammatory response, which is a cluster of risk factors interrelated with cardiovascular disease and T2DM (Grundy et al., 2005; O'Neill et al., 2016). Epidemiological evidence suggests that MetS may also be linked to cognitive dysfunction, involving VD and AD (Martins et al., 2006; Cooper et al., 2015). Obesity and MetS represent an increase in adipose tissue and lead to an adipocyte endocrine dysfunction, secreting excess or inadequate adipose tissue hormones and adipokines, which may be a clue to the mechanisms associated with dementia. APN regulates cerebral inflammatory responses, central food intake, energy expenditure glucose, and fatty acid catabolism. It is also a potent insulin sensitizer with a negative correlation with obesity, T2DM, MetS, and cardiovascular diseases (Chandran et al., 2003). Thus, it exerts beneficial effects on neuroprotection, neurotrophic actions, and neurogenesis. This may be attributed to modulation of insulin receptor signaling, sensitizing the insulin receptor signaling pathway and suppression of neuroinflammation. Though much data support obesity associated with multiple types of dementia, the links between adipokines and dementia risk remain to be further explored.

Diabetes

T2DM is also linked with dementia. It is characterized by hyperglycemia, IR, and pancreatic β cell dysfunction. Epidemiology studies have shown that the probability of cognitive decline in elder patients with T2DM is 1.5 times higher than that of non-diabetics (Biessels et al., 2006; Cheng et al., 2012). The latest cross-sectional and longitudinal researches have strongly demonstrated their association (Cukierman et al., 2005; Biessels et al., 2006). In addition, recent studies suggest that AD is a brain-specific diabetes and define it as "type 3 diabetes" on an account of the common risk factors between diabetes and AD, such as IR, BBB disruption, and altered glucose homeostasis (de la Monte and Wands, 2008; Kroner, 2009; de la Monte, 2014). But the exact mechanism between T2DM-related dementia especially in AD is unclear. The ultimate underlying mechanism is possibly related to IR, and systemic inflammatory response is associated with diabetes, vascular abnormalities,

neurodegenerative changes, and other multifactorial effects. Combined mechanisms may lead to mixed pathology. First, CVD pathology is likely to be an important determinant of the risks of all-cause dementia in individuals with diabetes. In several different studies, T2DM was consistently associated with an increased risk of pathologically verified infarcts at autopsy (Arvanitakis et al., 2006; Pruzin et al., 2017). These macroscopic brain infarcts may contribute to insidious ischemia of the brainrelated impaired cognitive function (Pruzin et al., 2018). Another potential pathology is neurodegenerative change including deposits of neurotic NP and NFTs, which may be attributed to brain IR and altered insulin signaling (Verdile et al., 2015). In addition, disruption of the BBB, dysregulation of lymphatics, and activation of the hypothalamic-pituitary-adrenal (HPA) axis are also proposed in many published articles (Ng and Chan, 2017; Pruzin et al., 2018). In conclusion, present data indicate that the relationship between T2DM and dementia is probably multifactorial in etiology. APN plays an important anti-inflammatory and anti-oxidant role, enhancing insulin sensitization and maintaining BBB and anti-atherosclerotic properties, which are crucial potential mechanisms in diabetesinduced dementia as described above Ng and Chan (2017). Some studies have reported that subjects with T2DM have lower plasma APN concentrations than matched non-diabetic controls (Hotta et al., 2000), and low APN levels can be used as predictors of the incidence of T2DM (Lindsay et al., 2002; Spranger et al., 2003).

Atherosclerosis

Atherosclerosis is a progressive vessel disease of large- or medium-sized arteries that eventually leads to cardiovascular diseases and stroke. Chronic inflammation, abnormal immune responses, and lipid depository are involved in the development of atherosclerosis (Solanki et al., 2018). Then, intracranial atherosclerosis (ICAS) causes degenerative vessel stenosis and cerebral hypoperfusion associated with an increased risk for ischemic stroke and dementia (Gorelick et al., 2008; Yarchoan et al., 2012). Atherosclerotic plaque development and rupture adding to subsequent rupture of thrombosis contribute to partial or total occlusion of the affected artery. These constitute main events of ICA (Wang et al., 2019). Based on valuable studies, APN may inhibit the release of pro-inflammatory cytokines (e.g., TGF- α), enhance the release of anti-inflammatory cytokines (e.g., IL-10), and promote the shift of macrophages toward the anti-inflammatory phenotype M2. Furthermore, it increases macrophages cholesterol efflux and prevent generation of foam cells (Lovren et al., 2010; Jenke et al., 2013; Wang M. et al., 2013). Another critical factor leading to atherosclerotic plaque is dyslipidemia, which is characterized by high triglycerides (TG), low high-density lipoprotein cholesterol (HDL-C), and small-dense low-density lipoprotein (sd-LDL) particles (Lusis, 2000). APN is well-known as a mediator in lipid metabolism. It is positively correlated with HDL-C and negatively associated with LDL cholesterol (LDL-C) and triglyceride concentrations (Katsiki et al., 2017). All evidences indicate that APN exerts a multifaceted effect in improving atherosclerosis in brain by regulating atherogenic factors.

Stroke

Stroke is defined as a cerebral disease that the blood supply to the brain is interrupted and subsequently the brain lacks vital oxygen and nutrients, leading to focal or global disruption of neurological function, without other obvious cause apart from vascular disorders (WHO MONICA project principal investigators, 1988). Accumulating evidences suggest that vascular factors and stroke injury increase risk for dementia including AD, not only VD (Tosto et al., 2016; Nucera and Hachinski, 2018). Mechanisms might involve hypoxia, hypoperfusion, and neuroinflammation. Furthermore, dementia and stroke share the corresponding risks and protective factors (Nucera and Hachinski, 2018). Stroke increases the risk of dementia in two ways. One is a sharp decline in cognition that occurs after a stroke (post-stroke dementia), while the other is to accelerate the development of cognitive decline several years after stroke (Levine et al., 2015). Dementia becomes a severe problem in stroke survivors. Several prospective cohort studies have demonstrated cognitive deficits after stroke (Lusis, 2000; Katsiki et al., 2017). In addition, meta-analysis studies have shown that some form of cognitive impairment affects approximately 30% of stroke patients (Ben Assayag et al., 2012; Wollenweber et al., 2014). The risks for stroke are shared with dementia, including non-modifiable (e.g., age, sex, and genes) and modifiable (e.g., hypertension, diabetes, obesity, MetS, and smoking) factors (Vijayan and Reddy, 2016). Association between APN and stroke have been evaluated in a number of studies, but the results are contradictory. Some retrospective case-control studies suggest that low APN level is related to greater stroke risk (Stott et al., 2009; Savopoulos et al., 2011; Prugger et al., 2012). Furthermore, several studies suggest that APN knockout (APN-KO) mice are more prone to serious ischemia-reperfusion injury in the brain (Shibata et al., 2005). However, several cases report that there are no independent links between APN and stroke risk (Stott et al., 2009). In addition, some other studies have shown that elevated APN levels in the elderly may contribute to an increased risk of ischemic stroke (Hao et al., 2013).

Considering all above results, whether APN is positively correlated with the risk factors of stroke still requires a lot of research data to identify, though numerous studies have confirmed its affirmative effects on vascular risk factors such as anti-atherosclerosis, anti-inflammation, and IR.

Other Risk Factors

Genetics, ethnicity, smoking, physical activity, social engagement, cognitive training, diet, traumatic brain injury, depression, sleep, and level of education, etc. These are also crucial factors influencing the progression of dementia. For instance, genetic diagnosis is becoming a hot spot in dementia research. But most of them have been on AD while investigations on VD have mainly been on rare familial syndromes. According to a system review involved of meta-analysis research, there are six polymorphisms strongly associated with vascular cognitive impairment (APOE, ACT, ACE, MTHFR, PON1, and PSEN-1 genes; Dwyer et al., 2013). APOE e4 has been reported as a significant related gene of AD in another independent meta-analysis (Farrer et al., 1997; Yin et al., 2012). Moreover, exercise is another important factor that has been reported to possibly maintain and improve cognitive function (Brown et al., 2013; Pedersen, 2019). A randomized controlled trial has shown that physical or mental activity may enhance cognitive function in older adults and that the amount of activity is more important than the type in the subject population (Barnes et al., 2013). Hence, distinguishing and utilizing these modifiable factors could help us find effective interventions to prevent dementia at the preclinical stage. Understanding well the role of APN in the progress is a key to explore novel strategies.

REGULATION MECHANISM OF APN ON DEMENTIA

APN has pleiotropic effects that may benefit AD and VD, involving anti-inflammatory and insulin-sensitizing effects, regulating sugar and lipid metabolism, regulating glucose and lipid metabolism, improving mitochondrial dysfunction, decreasing A β amyloid deposition, and inhibition of *tau* phosphorylation (**Figure 2**).

Anti-neuroinflammation

Epidemiological and clinical studies have repeatedly demonstrated that APN plays a chronic anti-inflammatory



FIGURE 2 | The potential effect of adiponectin to target diseases and the relationship between them. As demonstrated here, adiponectin has pleiotropic effects on numerous organs and tissues, and there is a close correlation between various diseases. For example, hypertension, atherosclerosis, diabetes, and obesity are risk factors in both dementia and coronary heart disease, while hypertension, atherosclerosis, and obesity are associated with the onset of diabetes. Adiponectin has anti-atherogenic, glucose metabolism-regulating, anti-inflammatory, insulin-sensitizing, and cardioprotective effects in multiple diseases.

role in dementia and other related diseases, such as T2DM, cardiovascular disease, and cancer. For example, Chabry et al. (2015) showed that APN could reduce neuroinflammation and depressive-like behaviors in mice by regulating microglia and macrophage phenotype and activation state. Lecompte et al. (2017) found that APN retained its anti-inflammatory feature in dystrophic muscle by activating the AdipoR1-AMPK-SIRT1-PGC-1apathway in mice. Besides, Jian et al. (2019) demonstrated that APN suppressed inflammatory response of microglia to amyloid-β oligomer (AβO) and APN deficiency may aggravate microglia-mediated neuroinflammation in AD mice. In patients with severe carotid stenosis, APN's negative association with glucose, insulin, and intraplaque inflammatory markers was observed (Liberale et al., 2018). Macrophages are the main targets for APN to exert anti-inflammatory effects. It acts by suppressing macrophage differentiation, modulating macrophage function or shifting phenotype from proinflammatory M1 state to an anti-inflammatory M2 state, decreasing expression of Toll-like receptor 4 (TLR4), and regulating inflammation responses (Yokota et al., 2000; Gordon and Martinez, 2010). It inhibits the transformation of human monocyte-derived macrophages into foam cells and the production of inflammatory chemokines and upregulates the production of the anti-inflammatory cytokine interleukin 10 (IL-10) in macrophages binding AdipoR1 primarily. Additionally, it may modulate the suppression of M1 macrophage activation; in contrast, it regulates promotion of M2 macrophage proliferation (Fang and Judd, 2018). It binds three receptor subtypes, AdipoR1, AdipoR2, and T-cadherin receptor, and mediates its effects primarily through AMPK signaling pathway (Thundyil et al., 2012). APN is also reported to act protective effects against inflammation on a variety of cell types (e.g., cardiomyocytes, endothelial cells, and vascular smooth muscle cells or microglia, astrocytes, and neurons in brain), and it mediates the phenotypes of these cells to exert anti-inflammatory effects (Ohashi et al., 2014; Sargolzaei et al., 2018). APN gene expression is widely expressed in the cortex and the hippocampus. It is suggested that APN inhibits pro-inflammatory signal release, IL-6 and TNF-a, from BBB endothelial cells, indirectly modulating the inflammatory signaling across the BBB (Spranger et al., 2006). In some studies, it is demonstrated that it down-regulates neuroinflammation by decreasing AB amyloid deposition in AD, while improving dementia (Chan et al., 2012; Song et al., 2017). Therefore APN's anti-inflammation properties in target organs is multiplex and complicated, and worth discussing further.

Inhibiting Insulin Resistance

Definition of IR in T2DM means "reduced sensitivity in body tissues to the action of insulin," ordinarily observed in T2DM and obesity, characterized as hyperinsulinemia and dyslipidemia (Goldstein, 2002). Correspondingly, brain IR is equivalent to a weakened response of the brain cells to insulin (Mielke et al., 2005). Insulin is involved in neuromodulation, including regulation of neurotransmitter (e.g., acetylcholine) concentrations, neuronal differentiation, repair, proliferation, regeneration, and suppression of neuronal apoptosis, subsequently mediating memory and learning processes (Kang et al., 2017). Thus, brain IR may be attributed to a decrease in insulin receptors, a loss of insulin receptor-binding insulin function, or defective insulin signaling cascade in the central nervous system. At the cellular level, this lack of response may appear as alteration of neuroplasticity, receptor regulation or neurotransmitter release dysfunction, or insulin metabolism disorders. Functionally, dysregulation of brain or peripheral metabolism, as well as cognitive and emotional impairment might be the main manifestations. In addition, numerous epidemiological data propose that obesity, T2DM, and other MetS of IR are risk factors for AD and other associated dysfunctions (Ahtiluoto et al., 2010; Bosco et al., 2012; Gao et al., 2013; Chatterjee et al., 2016).

Besides, it is delineated that in IR, target tissues do not respond insufficiently to insulin stimulation, eventually leading to hyperglycemia, hyperlipidemia, inflammation, and reduced plasma APN concentrations (Shibata et al., 2005). Upregulating plasma APN can reverse the sequelae of IR in various target organs, such as skeletal muscle, liver, pancreatic, and adipose tissue, established by a mass of basic and clinical outcome (Combs et al., 2004; Otabe et al., 2007; Wang et al., 2014; Ye et al., 2014). In contrast, some other documents show that APN deficiency is closely related to IR-exacerbating metabolic disorders such as T2DM and obesity (Maeda et al., 2002; Spranger et al., 2003; Bajaj et al., 2004). In these diseases, infiltration of inflammatory cells, particularly activated macrophages, aggregating into adipose tissue induces a highly inflammatory status. Meanwhile, in these conditions, a significant reduction exists in serum APN (Zhang et al., 2009). In addition, studies have shown that continuing elevated levels of inflammatory cytokines may directly deteriorate IR and lead to disruption of insulin sensitivity (Weisberg et al., 2003; Xu et al., 2003). APN decreases oxidative stress and inflammatory cytokines, which contribute to an improvement of IR. Animals and human studies have also identified that serum APN is transported into the CSF, regulating various central physiological functions of the brain (Qi et al., 2004). Intraventricular injection of APN can improve hypothalamic insulin signaling activity in diabetic rats and adjust glucose homeostasis (Park et al., 2011). Therefore, APN defined as an insulin-sensitizing hormone is a crucial mediator participating in the protection of brain health. Exploiting and better understanding its function in the crucial pathological mechanism in nerve provide powerful opportunities to novel therapeutic interventions that may improve dementia.

Regulating Glucose and Lipid Metabolism

As mentioned above, the molecular alterations in diabetes, obesity, and dementia are accompanied by impaired glucose uptake, lipid and fatty acid metabolism dysfunction, and energy metabolism disorder (Hotamisligil, 2006; Lourenco et al., 2015). Furthermore, increasing evidence has clearly demonstrated that cellular IR exists in the brains of AD patients and even those of non-diabetic patients; therefore, AD is also referred to as "type 3 diabetes" (de la Monte and Wands, 2008).

This means that dementia especially AD can absolutely be considered a kind of metabolic disease (De Felice et al., 2014; Ferreira et al., 2014). However, APN directly or indirectly exerts anti-insulin resistance, regulation of glycogen, lipid and fatty acid metabolism, and reduction of oxidative stress at the hub or periphery. In terms of glycogen metabolism, it is involved in mediating hepatic glucose production, glycogen uptake, decomposition, utilization, storage, transport, energy expenditure, and protection of pancreatic β cell as well as maintaining glycogen homeostasis in the brain. While in lipid metabolism, it is demonstrated that circulating APN levels positively associate with HDL-C (Matsubara et al., 2002; Yamamoto et al., 2002; Ezenwaka et al., 2004; Kazumi et al., 2004; Shetty et al., 2004; Kangas-Kontio et al., 2010; Christou et al., 2012) and show an inverse correlation with TG (Matsubara et al., 2002; Yamamoto et al., 2002; Ezenwaka et al., 2004; Siebel et al., 2015; Yanai and Yoshida, 2019). The possible mechanisms underlying up-regulation of HDL-C due to APN involve increasing production of apolipoprotein apo-AI and ATP-binding cassette transporter A1 (ABCA1; Matsuura et al., 2007; Oku et al., 2007; Qiao et al., 2008; Kitajima et al., 2011), down-regulation of hepatic lipase (HL) activity (Schneider et al., 2005; Clarenbach et al., 2007), and activation of lipoprotein lipase (LPL; Yanai and Yoshida, 2019). On the other hand, the plausible mechanism of TG reduction can be attributed to the regulation of LPL activity by APN (von Eynatten et al., 2004; Kobayashi et al., 2005) and the decrease of APN-induced serum APO-CIII, a well-known LPL inhibitor (Chan et al., 2005; Tsubakio-Yamamoto et al., 2012). With regard to LDL-C, the majority of studies have indicated no association with circulating APN (Kazumi et al., 2004; Shetty et al., 2004; Tomono et al., 2018). Nevertheless, high sd-LDL levels have been demonstrated to be correlated with elevated TG levels and decreased HDL-C levels, which constitute a common feature of diabetes and MetS (Eckel et al., 2005; Rizzo et al., 2009). In addition, APN-mediated improvement of HDL and TG may decrease the atherogenic lipoprotein sd-LDL and remnant lipoproteins, derived from very low density lipoprotein (VLDL) and chylomicrons (Yanai and Yoshida, 2019). Summarily, increase of serum APN levels may protect against atherosclerosis and other related diseases via mediating lipid metabolism.

Special Effects in AD Pathogenesis

AD is referred to a degenerative brain disease characterized by extracellular A β plaques and intraneuronal accumulation of NFTs. Amyloid plaques contain A β , while NFTs are composed of hyperphosphorylated *tau* proteins (Takahashi et al., 2017). APN has been repeatedly reported to play special effects in AD pathogenesis.

Reducing Aβ-Amyloid Deposition

Extracellular $A\beta$ -amyloid deposition into oligomers, fibrils, and plaques is a major hallmark of AD pathological mechanism. It may cause the dysfunction of several crucial processes, such as synaptogenesis, neurotrophy, and apoptosis, showing neurotoxin

in the disruption of learning and memory (Rad et al., 2018). Then, the amyloid cascade hypothesis is the most prevailing hypothesis and propose that $A\beta$ accumulation is the initiating mechanistic event. In this case, various stages of aggregates, involving protofibrils of AB, fibrillar forms of AB, as well as different soluble and insoluble AB oligomers, are neurotoxic as they could damage synapses and, in turn, cause neuron loss, ultimately leading to chronic neurodegeneration and dementia (Hardy, 2009; Blennow et al., 2015). Currently, targeting this molecular mechanism of AB neurotoxicity, numerous research therapies emerge aiming to reduce further $A\beta$ aggregation and plaque formation in brain, but remain invalid (Doody et al., 2013, 2014; Salloway et al., 2014). It could be that A β accumulation is just a bystander, rather than the cause, of neurodegeneration in AD. Studies have posed that insulin modulates various steps in the amyloid cascade, affecting Aß aggregation in the brain. The disturbance of insulin signaling may inhibit AB clearance and accelerate the formation of neurotoxic Aß plaque (Kim and Feldman, 2015; Rad et al., 2018). In addition, there is no doubt about the relevance of neuroinflammation in AD. Astroglia and microglia are believed to be the major sources of pro-inflammatory cytokines in the brain and can be stimulated by AB aggregation (Letra et al., 2019b). However, it has been reported that APN inhibits the inflammatory response and improve IR to indirectly reduce the production of amyloid plaques (Kamat et al., 2016). Furthermore, a study has shown that APN suppresses inflammatory response of microglia by inhibiting ABO and that APN deficiency promotes aggravation of microglia activation and deteriorates neuroinflammation in AD mice (Jian et al., 2019). Thus, it appears to be valuable to probe and perceive crosstalk of APN to AD for effective therapies.

Inhibiting Hyperphosphorylation of Tau in NFTs

NFTs are referred to as the twisted fibers involving abnormal phosphorylated tau proteins (also named phospho-tau, or p-tau), which exist as oligomers primarily in neurons suppressing microtubule assembly (Grundke-Iqbal et al., 1986; Iqbal et al., 1986). The lack of successful clinical trials targeting $A\beta$ plaques provides a novel opportunity to seek out potential therapy targeting pathological tau in AD progression. The crucial role of tau has been emphasized by several clinical studies in regard to the close correlation between *tau*-positive NFTs and AD development in the brain. NFT-positive cell density correlates with disease stages, which are measured by clinical parameters for disease severity or cognitive decline (Giannakopoulos et al., 2003). Contrarily, senile plaque density is not associated with stages (Delaère et al., 1990). Hyperphosphorylated tau is neurotoxic, suppressing microtubule assembly and inducing prion-like template activity. Therefore, the most promising treatment is to inhibit hyperphosphorylation and clear pathological tau; in addition, nerve regeneration can save tau pathology and cognitive decline (Iqbal et al., 2016). Xu et al. (2018) found that in the ICV-STZ rat model experiment, APN supplements inhibit hyperphosphorylation of tau protein at multiple AD-related sites, improve cognitive deficits, and have neuroprotective effects. But intriguingly, it is also reported that adaptation of APN to IR may play a dual role in the formation of two markers of AD: A β plaques and NFTs, and perhaps its fluctuation acts as a driving force in the disease pathogenesis (Sekiyama et al., 2014; Waragai et al., 2016, 2017). Therefore, according to this unique biological mechanism of APN in AD, a selective therapeutic strategy that is distinct from previous concepts may be required.

CONTROVERSY

Although a large number of studies have shown that APN has beneficial effects such as anti-inflammation, attenuating IR, regulation of sugar and lipid metabolism, and antiatherosclerosis, its neuroprotective effects are still controversial. Some researches suggest that APN is of no significance, and some even think it is harmful (Table 1; Kamogawa et al., 2010; Une et al., 2011; Teixeira et al., 2013; Dukic et al., 2016; Gorska-Ciebiada et al., 2016; Kitagawa et al., 2016; Bednarska-Makaruk et al., 2017; Bossolasco et al., 2017; Fujita et al., 2018; Gilbert et al., 2018; Benavente et al., 2019; Letra et al., 2019a). Furthermore, high serum APN levels have been reported to be positively associated with cardiovascular mortality (Ortega Moreno et al., 2016). These conflicting results about association between the APN serum levels and the outcomes of different stages of disease (MCI or dementia) suggest that we still know very little about the complex involvement of APN in AD. What contributes to the incongruences described above? Here, are five possible underlying causes.

First of all, the most important one is that individual diagnostic accuracy varies in different study teams. Classification of patients in cohorts depends on the accurate diagnostic process. Using diagnostic tools [such as Mini Mental State Examination (MMSE) and Montreal Cognitive Assessment (MoCA)] or exclusively relying on them, ignoring the limitation of examinations may influence the result (Chapman et al., 2016). Second, lack of detailed treatment data for patients enrolled, such as drugs and other treatments. Certain special drugs such as acetylcholinesterase inhibitors (AChEIs), statins, and thiazolidinediones all might affect the circulating levels of APN (Montecucco and Mach, 2009). Third, the nonuniformity in basic characteristics of study design and studied populations or lack of adjustments to confounding factors including vascular risk factors may also explain the inconsistency of different experimental results. Fourth, few studies have distinguished different proportions of circulating APN isomers, which are the decisive isomers, hexamers, trimers, or HMW. As mentioned above, different forms determine their different biological activities. In related diseases, HMW APN or HMW APN/total APN may be a more effective indicator of insulin sensitivity (Aso et al., 2006). Lastly, high circulating APN levels may result in subsequent resistance to APN in a manner similar to IR. Studies have demonstrated that high circulating APN levels are related to inverse outcome, defined as APN resistance (Van Berendoncks et al., 2010; Sente et al., 2016). Thus, APN may show dual effects in the pathological process of dementia.

TABLE 1 | Population-based studies aiming to assess the association between adiponectin and cognitive dysfunction diseases.

References	Study objects	Title	Disease	Sample, method	Results
Gorska-Ciebiada et al. (2016) and Xu et al. (2018)	62 seniors with type 2 diabetes (T2DM) and MCI, and 132 seniors with T2DM but without MCI (controls)	Adiponectin, leptin, and IL-1β in elderly diabetic patients with mild cognitive impairment.	Mild cognitive impairment.	Serum, ELISA	Serum leptin and IL-1 β levels were higher and adiponectin concentration was lower in MCI patients than controls. In MCI subjects, adiponectin level was negatively correlated with leptin, IL-1 β levels, and BMI. Leptin concentration was correlated with IL-1 β level. Univariate logistic regression models revealed that the factors that increased the likelihood of diagnosis of MCI in elderly patients with T2DM were higher levels of HbA1c, leptin, IL-1 β , and triglycerides, as well as lower levels of adiponectin and HDL cholesterol. Similarly, previous CVD, hypertension, hyperlipidemia, retinopathy, nephropathy, hypoglycemia, longer duration of diabetes, increased number of comorbidities, older age, and fewer years of formal education were found to be associated with MCI. The multivariable model indicated fewer years of formal education, previous CVD, hypertension, increased number of comorbidities, higher HbA1c and IL-1 β levels, and lower adiponectin level. Elderly diabetic patients with MCI have higher levels of leptin and IL-1 β and lower levels of adiponectin.
Kamogawa et al. (2010) nd Waragai et al. (2017)	517middle-aged-to-elderly community-dwelling persons.	Abdominal fat, adipose-derived hormones and mild cognitive impairment: the J-SHIPP study.	Mild cognitive impairment (MCI)	Serum, ELISA	In men, the abdominal subcutaneous fat area was significantly lower in participants with MCI than in those with normal cognitive function [median (interquartile range): 107.4 (85.9, 133.1) cm ² vs. 136.4 (93.1, 161.4) cm ² ; $p = 0.002$]. Logistic regression analyses with confounding factors including age and abdominal subcutaneous fat area showed that a 10 mg/L increase in plasma adiponectin had a protective effect against the development of MCI in men (odds ratio: 0.46; 95% CI: 0.20–0.97; $p = 0.041$).
Teixeira et al. (2013) and Sekiyama et al. (2014)	54 subjects with MCI and 43 controls	Decreased levels of circulating adiponectin in mild cognitive impairment and Alzheimer's disease.	Mild cognitive impairment (MCI) and Alzheimer's disease (AD)	Serum, ELISA	Serum levels of adiponectin were significantly lower in MCI and AD as compared to controls ($p < 0.001$). After controlling for age, educational level, and APOE genotype, adiponectin levels remained significantly reduced in these groups ($p < 0.001$). Circulating adiponectin levels did not predict cognitive decline in the elderly controls (i.e., progression from normal cognition to MCI) or progression to Alzheimer's disease in subjects with MCI.
Letra et al. (2019a) and Gorska-Ciebiada et al. (2016)	Human, amnestic mild cognitive impairment (MCI, n = 71) and Alzheimer's dementia (AD, $n = 53$)	Association between adipokines and biomarkers of Alzheimer's disease: A cross-sectional study.	Alzheimer's disease	serum and CSF, ELISA	Serum adiponectin was 33% higher in AD when compared to MCI patients. Adiponectin CSF levels, similar in both groups, were positively correlated with A β 42 and cognitive function, though only in women. The area under the ROC curve was 0.673 (95% CI: 0.57–0.78) for serum adiponectin as predictor of dementia stage and the cutoff 10.85 μ g/ml maximized the sum of specificity (87%) and sensitivity (44%).
Fujita et al. (2018) and Kamogawa et al. (2010)	20 male and 32 female, aged 60–93 years, mean 80.0	Increased adiponectin is associated with cerebral white matter lesions in the elderly with cognitive impairment	Alzheimer's disease	Serum, ELISA	High serum adiponectin levels correlated with more severe WML ($\rho = 0.013$). Low BMI ($\rho < 0.001$), female sex ($\rho = 0.025$), and high WML scores ($\rho = 0.039$) were significant determinants of high serum adiponectin. HT ($\rho = 0.032$) and high adiponectin levels ($\rho = 0.021$) were independent risk factors for WML. Overall, we observed an association between serum adiponectin levels and WML severity in elderly people with cognitive decline.

Adiponectin and Dementia

References	Study objects	Title	Disease	Sample, method	Results
Bednarska- Makaruk et al. (2017) and Teixeira et al. (2013)	205 patients with dementia [89 with Alzheimer's disease (AD), 47 with vascular dementia (VaD), 69 with mixed dementia (MD)], 113 persons with mild cognitive impairment and in 107 controls	Association of adiponectin, leptin, and resistin with inflammatory markers and obesity in dementia.	Dementia.	Serum and CSF, ELISA	In all-cause dementia, adiponectin and resistin levels were significantly higher as compared to the controls; leptin levels did not show differences. Higher adiponectinlevels concerned AD and MD, whereas higher resistin concerned VaD and MD. After stratification by abdominal obesity, the differences in adiponectin levels remained significant in subjects without obesity. In all-cause dementia, negative correlation of adiponectin with obesity, glucose metabolism parameters, IL-6, and hsCRP and positive correlation with HDL-cholesterol were found. Positive correlation of resistin with age, IL-6, hsCRP, and chitotriosidase and negative correlation with HDL-cholesterol and paraoxonase 1 were stated.
Une et al. (2011) and Letra et al. (2019a)	Normal controls ($n = 28$), MCI ($n = 18$), and AD ($n = 27$) subjects	Adiponectin in plasma and cerebrospinal fluid in MCI and Alzheimer's disease.	MCI and Alzheimer's disease.	Serum and CSF, ELISA,	The levels of adiponectin in plasma and in CSF showed a positive correlation. Plasma adiponectin was significantly higher in MCI and AD compared to NC, whereas CSF adiponectin was significantly higher in MCI compared to NC.
Gilbert et al. (2018) and Fujita et al. (2018)	205 patients over 65 years of age	Association between peripheral leptin and adiponectin levels and cognitive decline in patients with neurocognitive disorders \geq 65 years.	Neurocognitive disorders	Serum, ELISA	The mean BMI was significantly lower (by 2 kg/m ² , $\rho = 0.01$) in patients with AD than in patients with either mild-NCD or vascular/mixed dementia. Leptin levels were significantly higher ($\rho = 0.043$) and adiponectin levels were significantly lower ($\rho = 0.045$) in patients with mild-NCD than in patients with major-NCD (AD or vascular/mixed dementia). However, the mixed model suggested no influence of the baseline levels of these two biomarkers on the course of cognitive decline.
Benavente et al. (2019) and Bednarska- Makaruk et al. (2017)	50% splits of TARCC's data (Group 1 n = 1,691; Group 2 n = 1,690)	Serum adiponectin is related to dementia.	Alzheimer's disease	MIMIC models	Serum APN was significantly related to δ scores ($r = 0.10$, $p = 0.015$). APN hadno significant effect on g' ($r =25$, $p = 0.66$), nor did it have any independent direct effects on cognitive performance. These results were replicated across random subsets (Δ CHISQ = 2.8 (Scherer et al., 1995), $p > 0.90$).
Bossolasco et al. (2017) and Une et al. (2011)	88 samples in the whole cohort	Adiponectin levels in the serum and cerebrospinal fluid of amyotrophic lateral sclerosis patients: possible influence on neuroinflammation?	Cerebrovascular and neurodegenerative diseases	Serum and CSF, ELISA	In the whole ALS group, serum APN levels were not different when compared to the age- and sex-matched control group (CTR), but a gender-specific analysis enlightened a significant opposite APN trend between ALS males, characterized by lower values (ALS 9.8 ± 5.2 vs. CTR 15 ± 9.7 µg/ml), and ALS females, showing higher amounts (ALS 26.5 ± 11.6 vs. CTR 14.6 ± 5.2 µg/ml). This sex-linked difference was significantly enhanced in familial ALS cases ($p \leq 0.01$). The APN levels in ALS cerebrospinal fluids wereunrelated toserum values and not linked to sex and/or familiarity of the disease. Finally, the screening of serum APN levels in patients affected by other neurological disorders revealed the highest serum values in FTD patients.
Kitagawa et al. (2016) and Gilbert et al. (2018)	466 patients (mean age 67.8 years, male 57%)	Serum high-molecular-weight adiponectin level and incident dementia in patients with vascular risk factors.	Dementia	Serum, ELISA	Serum HMW adiponectin level was $4.33 \pm 2.95 \mu$ g/ml; the levels were lower in men than in women and negatively correlated with body mass index. During the follow-up period (median 6.9 years), 47 patients had incident dementia including Alzheimer's disease dementia (Fillit et al., 2008), vascular dementia (Gustafson, 2010), mixed dementia (four), other dementia (three). Risks of dementia in patients with high vs. low HMW adiponectin levels were almost identical ($\rho = 0.689$). No association was found between adiponectin levels and Alzheimer's disease dementia or vascular dementia in the whole group or among men and women separately.

(Continued)

Adiponectin and Dementia

CONCLUSION AND FUTURE PERSPECTIVE

Researches about dementia are rapidly increasing. Advances in basic science and clinic studies in molecular and pathological mechanism have provided unprecedented possibilities for novel therapeutic strategy. The association between APN and AD or VD, whether direct or indirect, positive or negative, in risk factors or in pathological mechanism, central or peripheral, has been deeply evaluated and elaborated in numerous studies. Overall, it is significantly associated with risk factors of dementia such as obesity, T2DM, hyperlipidemia, atherosclerosis, and other vascular factors. APN has multiple effects on the pathological process of dementia. Positive effects are involved in anti-inflammation, reduced IR, anti-atherosclerosis, and regulation of energy metabolism, such as glycogen and lipids. Insulin sensitivity and neuroinflammatory responses are key cellular mechanisms involved in age-related cardiovascular disease, metabolic disease, cerebrovascular dysfunction, and cognitive decline. APN is highly correlated with insulin sensitivity and inflammatory response. Many studies have shown that elevated levels of APN can improve damaged insulin signaling; inhibit neuroinflammation, oxidative stress, nitrosative stress, etc.; and affect cerebral blood vessels. However, its potential mechanism is still not well explored and even controversial. Further studies are required to illustrate the exact functions and receptor-dependent or -independent downstream pathways of each isoform activation, and how APN's peripheral concentration can modulate its central effect. This will help us enhance congruency of the results and further facilitate the search for the possible role and pathophysiological mechanism of APN in the onset and development of dementia, especially its actions on the hippocampus and cerebral cortex. Finally, identifying and using these potential relationships, and thoroughly understanding APN's physiology, may help us seek out a multi-target cocktail therapy for individuals with cognitive impairment disease, or find a novel way to delay or block the process of dementia in the early stage. Is there an individual treatment that can take into account of the different physiological characteristics, genotypes, comorbidities, or even individual biomarker levels of dementia patients to achieve a cure? Maybe it's no longer a dream in the coming future.

AUTHOR CONTRIBUTIONS

YZ determined the structure of the review. RC selected the references and contributed to the writing. YS contributed to the revision and finalization of the article.

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TABLE 1 Continued					
References	Study objects	Title	Disease	Sample, method	Results
Dukic et al. (2016) and Benavente et al. (2019)	235 participants	The role of human kallikrein 6, clusterin and adiponectin as potential blood biomarkers of dementia.	Dementia.	Serum, ELISA	Serum concentrations of KLK6 ($p = 0.137$), CLU ($p = 0.178$), and ADPN ($p = 0.268$) did not differ between AD, VAD, MCl and cognitively healthy control group of participants, whereas IL-6 was significantly higher in VAD patients than in AD, MCl, and CHP individuals ($p = 0.014$). There was no association between investigated biomarkers and clinical patient parameters.

REFERENCES

- Ahtiluoto, S., Polvikoski, T., Peltonen, M., Solomon, A., Tuomilehto, J., Winblad, B., et al. (2010). Diabetes, Alzheimer disease, and vascular dementia: a population-based neuropathologic study. *Neurology* 75, 1195–1202. doi: 10.1212/wnl.0b013e3181f4d7f8
- Alzheimer's Association. (2013). Alzheimer's disease facts and figures. Alzheimers Dement. 9, 208–245. doi: 10.1016/j.jalz.2013.02.003
- Arita, Y., Kihara, S., Ouchi, N., Takahashi, M., Maeda, K., Miyagawa, J., et al. (1999). Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. *Biochem. Biophys. Res. Commun.* 257, 79–83. doi: 10.1006/bbrc. 1999.0255
- Arvanitakis, Z., Schneider, J. A., Wilson, R. S., Li, Y., Arnold, S. E., Wang, Z., et al. (2006). Diabetes is related to cerebral infarction but not to AD pathology in older persons. *Neurology* 67, 1960–1965. doi: 10.1212/01.wnl.0000247053. 45483.4e
- Aso, Y., Yamamoto, R., Wakabayashi, S., Uchida, T., Takayanagi, K., Takebayashi, K., et al. (2006). Comparison of serum high-molecular weight (HMW) adiponectin with total adiponectin concentrations in type 2 diabetic patients with coronary artery disease using a novel enzyme-linked immunosorbent assay to detect HMW adiponectin. *Diabetes* 55, 1954–1960. doi: 10.2337/db05-1525
- Bajaj, M., Suraamornkul, S., Piper, P., Hardies, L. J., Glass, L., Cersosimo, E., et al. (2004). Decreased plasma adiponectin concentrations are closely related to hepatic fat content and hepatic insulin resistance in pioglitazone-treated type 2 diabetic patients. *J. Clin. Endocrinol. Metab.* 89, 200–206. doi: 10.1210/jc. 2003-031315
- Barnes, D. E., Santos-Modesitt, W., Poelke, G., Kramer, A. F., Castro, C., Middleton, L. E., et al. (2013). The Mental Activity and eXercise (MAX) trial: a randomized controlled trial to enhance cognitive function in older adults. *JAMA Intern. Med.* 173, 797–804. doi: 10.1001/jamainternmed.2013.189
- Baumgart, M., Snyder, H. M., Carrillo, M. C., Fazio, S., Kim, H., and Johns, H. (2015). Summary of the evidence on modifiable risk factors for cognitive decline and dementia: a population-based perspective. *Alzheimers Dement.* 11, 718–726. doi: 10.1016/j.jalz.2015.05.016
- Bednarska-Makaruk, M., Graban, A., Wisniewska, A., Łojkowska, W., Bochyńska, A., Gugaa-Iwaniuk, M., et al. (2017). Association of adiponectin, leptin and resistin with inflammatory markers and obesity in dementia. *Biogerontology* 18, 561–580. doi: 10.1007/s10522-017-9701-0
- Ben Assayag, E., Korczyn, A. D., Giladi, N., Goldbourt, U., Berliner, A. S., Shenhar-Tsarfaty, S., et al. (2012). Predictors for poststroke outcomes: the Tel Aviv Brain Acute Stroke Cohort (TABASCO) study protocol. *Int. J. Stroke* 7, 341–347. doi: 10.1111/j.1747-4949.2011.00652.x
- Benavente, K. S. K., Palmer, R. F., and Royall, D. R. (2019). Serum adiponectin is related to dementia. J. Gerontol. A Biol. Sci. Med. Sci. doi: 10.1093/gerona/glz102 [Epub ahead of print].
- Biessels, G. J., Staekenborg, S., Brunner, E., Brayne, C., and Scheltens, P. (2006). Risk of dementia in diabetes mellitus: a systematic review. *Lancet Neurol.* 5, 64–74. doi: 10.1016/S1474-4422(05)70284-2
- Biswas, S. K., and Mantovani, A. (2012). Orchestration of metabolism by macrophages. *Cell Metab.* 15, 432–437. doi: 10.1016/j.cmet.2011.11.013
- Blennow, K., Mattsson, N., Schöll, M., Hansson, O., and Zetterberg, H. (2015). Amyloid biomarkers in Alzheimer's disease. *Trends Pharmacol. Sci.* 36, 297–309. doi: 10.1016/j.tips.2015.03.002
- Bosco, D., Plastino, M., Cristiano, D., Colica, C., Ermio, C., De Bartolo, M., et al. (2012). Dementia is associated with insulin resistance in patients with Parkinson's disease. J. Neurol. Sci. 315, 39–43. doi: 10.1016/j.jns.2011.12.008
- Bossolasco, P., Cancello, R., Doretti, A., Morelli, C., Silani, V., and Cova, L. (2017). Adiponectin levels in the serum and cerebrospinal fluid of amyotrophic lateral sclerosis patients: possible influence on neuroinflammation? *J. Neuroinflammation* 14:85. doi: 10.1186/s12974-017-0861-2
- Brochu-Gaudreau, K., Rehfeldt, C., Blouin, R., Bordignon, V., Murphy, B. D., and Palin, M. F. (2010). Adiponectin action from head to toe. *Endocrine* 37, 11–32. doi: 10.1007/s12020-009-9278-8
- Brown, B. M., Peiffer, J. J., and Martins, R. N. (2013). Multiple effects of physical activity on molecular and cognitive signs of brain aging: can exercise slow neurodegeneration and delay Alzheimer's disease? *Mol. Psychiatry* 18, 864–874. doi: 10.1038/mp.2012.162

Burns, A., and Iliffe, S. (2009). Dementia. BMJ 338:b75. doi: 10.1136/bmj.b75

- Chabry, J., Nicolas, S., Cazareth, J., Murris, E., Guyon, A., Glaichenhaus, N., et al. (2015). Enriched environment decreases microglia and brain macrophages inflammatory phenotypes through adiponectin-dependent mechanisms: relevance to depressive-like behavior. *Brain Behav. Immun.* 50, 275–287. doi: 10.1016/j.bbi.2015.07.018
- Chan, K. H., Lam, K. S., Cheng, O. Y., Kwan, J. S., Ho, P. W., Cheng, K. K., et al. (2012). Adiponectin is protective against oxidative stress induced cytotoxicity in amyloid-β neurotoxicity. *PLoS One* 7:e52354. doi: 10.1371/journal.pone. 0052354
- Chan, D. C., Watts, G. F., Ng, T. W., Uchida, Y., Sakai, N., Yamashita, S., et al. (2005). Adiponectin and other adipocytokines as predictors of markers of triglyceride-rich lipoprotein metabolism. *Clin. Chem.* 51, 578–585. doi: 10.1373/clinchem.2004.045120
- Chandran, M., Phillips, S. A., Ciaraldi, T., and Henry, R. R. (2003). Adiponectin: more than just another fat cell hormone? *Diabetes Care* 26, 2442–2450. doi: 10.2337/diacare.26.8.2442
- Chapman, K. R., Bing-Canar, H., Alosco, M. L., Steinberg, E. G., Martin, B., Chaisson, C., et al. (2016). Mini Mental State Examination and Logical Memory scores for entry into Alzheimer's disease trials. *Alzheimers Res. Ther.* 8:9. doi: 10.1186/s13195-016-0176-z
- Chatterjee, S., Peters, S. A., Woodward, M., Mejia Arango, S., Batty, G. D., Beckett, N., et al. (2016). Type 2 diabetes as a risk factor for dementia in women compared with men: a pooled analysis of 2.3 million people comprising more than 100,000 cases of dementia. *Diabetes Care* 39, 300–307. doi: 10.2337/ dc15-1588
- Cheng, G., Huang, C., Deng, H., and Wang, H. (2012). Diabetes as a risk factor for dementia and mild cognitive impairment: a meta-analysis of longitudinal studies. *Intern. Med. J.* 42, 484–491. doi: 10.1111/j.1445-5994.2012.02758.x
- Christou, G. A., Tellis, K. C., Elisaf, M. C., Tselepis, A. D., and Kiortsis, D. N. (2012). High density lipoprotein is positively correlated with the changes in circulating total adiponectin and high molecular weight adiponectin during dietary and fenofibrate treatment. *Hormones* 11, 178–188. doi: 10.14310/horm. 2002.1345
- Citron, M. (2010). Alzheimer's disease: strategies for disease modification. *Nat. Rev. Drug Discov.* 9, 387–398. doi: 10.1038/nrd2896
- Clarenbach, J. J., Vega, G. L., Adams-Huet, B., Considine, R. V., Ricks, M., and Sumner, A. E. (2007). Variability in postheparin hepatic lipase activity is associated with plasma adiponectin levels in African Americans. J. Investig. Med. 55, 187–194. doi: 10.2310/6650.2007.07001
- Combs, T. P., Pajvani, U. B., Berg, A. H., Lin, Y., Jelicks, L. A., Laplante, M., et al. (2004). A transgenic mouse with a deletion in the collagenous domain of adiponectin displays elevated circulating adiponectin and improved insulin sensitivity. *Endocrinology* 145, 367–383. doi: 10.1210/en.2003-1068
- Cooper, C., Sommerlad, A., Lyketsos, C. G., and Livingston, G. (2015). Modifiable predictors of dementia in mild cognitive impairment: a systematic review and meta-analysis. *Am. J. Psychiatry* 172, 323–334. doi: 10.1176/appi.ajp.2014. 14070878
- Corrada, M. M., Brookmeyer, R., Paganini-Hill, A., Berlau, D., and Kawas, C. H. (2010). Dementia incidence continues to increase with age in the oldest old: the 90+ study. *Ann. Neurol.* 67, 114–121. doi: 10.1002/ana.21915
- Corrada, M. M., Hayden, K. M., Paganini-Hill, A., Bullain, S. S., DeMoss, J., Aguirre, C., et al. (2017). Age of onset of hypertension and risk of dementia in the oldest-old: the 90+ study. *Alzheimers Dement*. 13, 103–110. doi: 10.1016/j. jalz.2016.09.007
- Craft, S. (2009). The role of metabolic disorders in Alzheimer disease and vascular dementia: two roads converged. *Arch. Neurol.* 66, 300–305. doi:10.1001/archneurol.2009.27
- Crous-Bou, M., Minguillon, C., Gramunt, N., and Molinuevo, J. L. (2017). Alzheimer's disease prevention: from risk factors to early intervention. *Alzheimers Res. Ther.* 9:71. doi: 10.1186/s13195-017-0297-z
- Cukierman, T., Gerstein, H. C., and Williamson, J. D. (2005). Cognitive decline and dementia in diabetes—systematic overview of prospective observational studies. *Diabetologia* 48, 2460–2469. doi: 10.1007/s00125-005-0023-4
- De Felice, F. G., Lourenco, M. V., and Ferreira, S. T. (2014). How does brain insulin resistance develop in Alzheimer's disease? *Alzheimers Dement.* 10, S26–S32. doi: 10.1016/j.jalz.2013.12.004

- de la Monte, S. M. (2014). Type 3 diabetes is sporadic Alzheimers disease: minireview. *Eur. Neuropsychopharmacol.* 24, 1954–1960. doi: 10.1016/j.euroneuro. 2014.06.008
- de la Monte, S. M., and Wands, J. R. (2008). Alzheimer's disease is type 3 diabetes-evidence reviewed. J. Diabetes Sci. Technol. 2, 1101–1113. doi: 10.1177/193229680800200619
- De Strooper, B., Vassar, R., and Golde, T. (2010). The secretases: enzymes with therapeutic potential in Alzheimer disease. *Nat. Rev. Neurol.* 6, 99–107. doi: 10.1038/nrneurol.2009.218
- Delaère, P., Duyckaerts, C., Masters, C., Beyreuther, K., Piette, F., and Hauw, J. J. (1990). Large amounts of neocortical β A4 deposits without neuritic plaques nor tangles in a psychometrically assessed, non-demented person. *Neurosci. Lett.* 116, 87–93. doi: 10.1016/0304-3940(90)90391-1
- Delles, C., Michelson, G., Harazny, J., Oehmer, S., Hilgers, K. F., and Schmieder, R. E. (2004). Impaired endothelial function of the retinal vasculature in hypertensive patients. *Stroke* 35, 1289–1293. doi: 10.1161/01.str. 0000126597.11534.3b
- Doody, R. S., Raman, R., Farlow, M., Iwatsubo, T., Vellas, B., Joffe, S., et al. (2013). A phase 3 trial of semagacestat for treatment of Alzheimer's disease. N. Engl. J. Med. 369, 341–350. doi: 10.1056/NEJMoa1210951
- Doody, R. S., Thomas, R. G., Farlow, M., Iwatsubo, T., Vellas, B., Joffe, S., et al. (2014). Phase 3 trials of solanezumab for mild-to-moderate Alzheimer's disease. N. Engl. J. Med. 370, 311–321. doi: 10.1056/NEJMoa1312889
- Dubois, B., Hampel, H., Feldman, H. H., Scheltens, P., Aisen, P., Andrieu, S., et al. (2016). Preclinical Alzheimer's disease: definition, natural history, and diagnostic criteria. *Alzheimers Dement.* 12, 292–323. doi: 10.1016/j.jalz.2016. 02.002
- Dukic, L., Simundic, A. M., Martinic-Popovic, I., Kackov, S., Diamandis, A., Begcevic, I., et al. (2016). The role of human kallikrein 6, clusterin and adiponectin as potential blood biomarkers of dementia. *Clin. Biochem.* 49, 213–218. doi: 10.1016/j.clinbiochem.2015.10.014
- Duvnjak, L., and Duvnjak, M. (2009). The metabolic syndrome—an ongoing story. *J. Physiol. Pharmacol.* 60, 19–24. doi: 10.1007/BF03185938
- Dwyer, R., Skrobot, O. A., Dwyer, J., Munafo, M., and Kehoe, P. G. (2013). Using Alzgene-like approaches to investigate susceptibility genes for vascular cognitive impairment. *J. Alzheimers Dis.* 34, 145–154. doi: 10.3233/JAD-121069
- Eckel, R. H., Grundy, S. M., and Zimmet, P. Z. (2005). The metabolic syndrome. Lancet 365, 1415–1428. doi: 10.1016/S0140-6736(05)66378-7
- Ewald, C. Y., Castillo-Quan, J. I., and Blackwell, T. K. (2018). Untangling longevity, dauer, and healthspan in caenorhabditis elegans insulin/IGF-1-signalling. *Gerontology* 64, 96–104. doi: 10.1159/000480504
- Ezenwaka, C. E., Kalloo, R., Uhlig, M., and Eckel, J. (2004). Relationship between adiponectin and metabolic variables in Caribbean offspring of patients with type 2 diabetes mellitus. *Horm. Metab. Res.* 36, 238–242. doi: 10.1055/s-2004-814454
- Fang, H., and Judd, R. L. (2018). Adiponectin regulation and function. Compr. Physiol. 8, 1031–1063. doi: 10.1002/cphy.c170046
- Farrer, L. A., Cupples, L. A., Haines, J. L., Hyman, B., Kukull, W. A., Mayeux, R., et al. (1997). Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease. *JAMA* 278, 1349–1356. doi: 10.1001/jama.1997.03550160069041
- Ferreira, S. T., Clarke, J. R., Bomfim, T. R., and De Felice, F. G. (2014). Inflammation, defective insulin signaling, and neuronal dysfunction in Alzheimer's disease. *Alzheimers Dement.* 10, S76–S83. doi: 10.1016/j.jalz.2013. 12.010
- Fillit, H., Nash, D. T., Rundek, T., and Zuckerman, A. (2008). Cardiovascular risk factors and dementia. Am. J. Geriatr. Pharmacother. 6, 100–118. doi: 10.1016/j. amjopharm.2008.06.004
- Franceschi, C., Olivieri, F., Marchegiani, F., Cardelli, M., Cavallone, L., Capri, M., et al. (2005). Genes involved in immune response/inflammation, IGF1/insulin pathway and response to oxidative stress play a major role in the genetics of human longevity: the lesson of centenarians. *Mech. Ageing Dev.* 126, 351–361. doi: 10.1016/j.mad.2004.08.028
- Fujita, Y., Toyomoto, T., Sakoh-Goshima, T., Kohno, Y., Okada, M., Hamano, T., et al. (2018). Increased adiponectin is associated with cerebral white matter lesions in the elderly with cognitive impairment. *Metab. Brain Dis.* 33, 1385–1388. doi: 10.1007/s11011-018-0247-x

- Gao, C., Liu, Y., Li, L., and Holscher, C. (2013). New animal models of Alzheimer's disease that display insulin desensitization in the brain. *Rev. Neurosci.* 24, 607–615. doi: 10.1515/revneuro-2013-0034
- Gardener, H., Wright, C. B., Rundek, T., and Sacco, R. L. (2015). Brain health and shared risk factors for dementia and stroke. *Nat. Rev. Neurol.* 11, 651–657. doi: 10.1038/nrneurol.2015.195
- GBD 2016 Dementia Collaborators. (2019). Global regional, and national burden of Alzheimer's disease and other dementias, 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet Neurol.* 18, 88–106. doi: 10.1016/S1474-4422(18)30403-4
- GBD 2016 Disease and Injury Incidence and Prevalence Collaborators. (2017). Global, regional, and national incidence, prevalence, and years lived with disability for 328 diseases and injuries for 195 countries 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet* 390, 1211–1259. doi: 10.1016/S0140-6736(17)32154-2
- Giannakopoulos, P., Herrmann, F. R., Bussiere, T., Bouras, C., Kovari, E., Perl, D. P., et al. (2003). Tangle and neuron numbers, but not amyloid load, predict cognitive status in Alzheimer's disease. *Neurology* 60, 1495–1500. doi: 10.1212/01.wnl.0000063311.58879.01
- Gilbert, T., Roche, S., Blond, E., Bar, J. Y., Drai, J., Cuerq, C., et al. (2018). Association between peripheral leptin and adiponectin levels and cognitive decline in patients with neurocognitive disorders ≥65 years. J. Alzheimers Dis. 66, 1255–1264. doi: 10.3233/jad-180533
- Goldstein, B. J. (2002). Insulin resistance as the core defect in type 2 diabetes mellitus. Am. J. Cardiol. 90, 3G-10G. doi: 10.1016/s0002-9149(02)02553-5
- Gordon, S., and Martinez, F. O. (2010). Alternative activation of macrophages: mechanism and functions. *Immunity* 32, 593–604. doi: 10.1016/j.immuni.2010. 05.007
- Gorelick, P. B., Scuteri, A., Black, S. E., Decarli, C., Greenberg, S. M., Iadecola, C., et al. (2011). Vascular contributions to cognitive impairment and dementia: a statement for healthcare professionals from the american heart association/american stroke association. *Stroke* 42, 2672–2713. doi: 10.1161/STR.0b013e3182299496
- Gorelick, P. B., Wong, K. S., Bae, H. J., and Pandey, D. K. (2008). Large artery intracranial occlusive disease: a large worldwide burden but a relatively neglected frontier. *Stroke* 39, 2396–2399. doi: 10.1161/strokeaha.107. 505776
- Gorska-Ciebiada, M., Saryusz-Wolska, M., Borkowska, A., Ciebiada, M., and Loba, J. (2016). Adiponectin, leptin and IL-1 β in elderly diabetic patients with mild cognitive impairment. *Metab. Brain Dis.* 31, 257–266. doi: 10.1007/s11011-015-9739-0
- Grundke-Iqbal, I., Iqbal, K., Tung, Y. C., Quinlan, M., Wisniewski, H. M., and Binder, L. I. (1986). Abnormal phosphorylation of the microtubule-associated protein tau (tau) in Alzheimer cytoskeletal pathology. *Proc. Natl. Acad. Sci. U S* A 83, 4913–4917. doi: 10.1073/pnas.83.13.4913
- Grundy, S. M., Cleeman, J. I., Daniels, S. R., Donato, K. A., Eckel, R. H., Franklin, B. A., et al. (2005). Diagnosis and management of the metabolic syndrome: an american heart association/national heart, lung, and blood institute scientific statement. *Circulation* 112, 2735–2752. doi: 10.1161/CIRCULATIONAHA.105.169404
- Gustafson, D. R. (2010). Adiposity hormones and dementia. J. Neurol. Sci. 299, 30–34. doi: 10.1016/j.jns.2010.08.036
- Haan, M. N., and Wallace, R. (2004). Can dementia be prevented? Brain aging in a population-based context. *Annu. Rev. Public Health* 25, 1–24. doi: 10.1146/annurev.publhealth.25.101802.122951
- Hachinski, V. C., and Bowler, J. V. (1993). Vascular dementia. *Neurology* 43, 2159–2160; author reply 2160–2161. doi: 10.1212/wnl.43.10.2159-a
- Hao, G., Li, W., Guo, R., Yang, J. G., Wang, Y., Tian, Y., et al. (2013). Serum total adiponectin level and the risk of cardiovascular disease in general population: a meta-analysis of 17 prospective studies. *Atherosclerosis* 228, 29–35. doi: 10.1016/j.atherosclerosis.2013.02.018
- Hardy, J. (2009). The amyloid hypothesis for Alzheimer's disease: a critical reappraisal. *J. Neurochem.* 110, 1129–1134. doi: 10.1111/j.1471-4159.2009. 06181.x
- Hotamisligil, G. S. (2006). Inflammation and metabolic disorders. *Nature* 444, 860-867. doi: 10.1038/nature05485
- Hotta, K., Funahashi, T., Arita, Y., Takahashi, M., Matsuda, M., Okamoto, Y., et al. (2000). Plasma concentrations of a novel, adipose-specific protein, adiponectin,

in type 2 diabetic patients. Arterioscler. Thromb. Vasc. Biol. 20, 1595–1599. doi: 10.1161/01.atv.20.6.1595

- Iadecola, C., and Gottesman, R. F. (2019). Neurovascular and cognitive dysfunction in hypertension. *Circ. Res.* 124, 1025–1044. doi: 10.1161/CIRCRESAHA.118.313260
- Iqbal, K., Grundke-Iqbal, I., Zaidi, T., Merz, P. A., Wen, G. Y., Shaikh, S. S., et al. (1986). Defective brain microtubule assembly in Alzheimer's disease. *Lancet* 2, 421–426. doi: 10.1016/s0140-6736(86)92134-3
- Iqbal, K., Liu, F., and Gong, C. X. (2016). Tau and neurodegenerative disease: the story so far. Nat. Rev. Neurol. 12, 15–27. doi: 10.1038/nrneurol.2015.225
- Iwabu, M., Yamauchi, T., Okada-Iwabu, M., Sato, K., Nakagawa, T., Funata, M., et al. (2010). Adiponectin and AdipoR1 regulate PGC-1 α and mitochondria by Ca²⁺ and AMPK/SIRT1. *Nature* 464, 1313–1319. doi: 10.1038/nature08991
- Jellinger, K. A., and Attems, J. (2015). Challenges of multimorbidity of the aging brain: a critical update. J. Neural Transm. 122, 505–521. doi: 10.1007/s00702-014-1288-x
- Jenke, A., Wilk, S., Poller, W., Eriksson, U., Valaperti, A., Rauch, B. H., et al. (2013). Adiponectin protects against Toll-like receptor 4-mediated cardiac inflammation and injury. *Cardiovasc. Res.* 99, 422–431. doi: 10.1093/cvr/ cvt118
- Jian, M., Kwan, J. S., Bunting, M., Ng, R. C., and Chan, K. H. (2019). Adiponectin suppresses amyloid-β oligomer (AβO)-induced inflammatory response of microglia via AdipoR1-AMPK-NF-κB signaling pathway. J. Neuroinflammation 16:110. doi: 10.1186/s12974-019-1492-6
- Jorm, A. F., and Jolley, D. (1998). The incidence of dementia: a meta-analysis. *Neurology* 51, 728–733. doi: 10.1212/wnl.51.3.728
- Kalaria, R. N. (2016). Neuropathological diagnosis of vascular cognitive impairment and vascular dementia with implications for Alzheimer's disease. *Acta Neuropathol.* 131, 659–685. doi: 10.1007/s00401-016-1571-z
- Kalaria, R. N. (2018). The pathology and pathophysiology of vascular dementia. Neuropharmacology 134, 226–239. doi: 10.1016/j.neuropharm.2017.12.030
- Kalaria, R. N., Maestre, G. E., Arizaga, R., Friedland, R. P., Galasko, D., Hall, K., et al. (2008). Alzheimer's disease and vascular dementia in developing countries: prevalence, management and risk factors. *Lancet Neurol.* 7, 812–826. doi: 10.1016/S1474-4422(08)70169-8
- Kamat, P. K., Kalani, A., Rai, S., Tota, S. K., Kumar, A., and Ahmad, A. S. (2016). Streptozotocin intracerebroventricular-induced neurotoxicity and brain insulin resistance: a therapeutic intervention for treatment of sporadic Alzheimer's disease (sAD)-like pathology. *Mol. Neurobiol.* 53, 4548–4562. doi: 10.1007/s12035-015-9384-y
- Kamogawa, K., Kohara, K., Tabara, Y., Uetani, E., Nagai, T., Yamamoto, M., et al. (2010). Abdominal fat, adipose-derived hormones and mild cognitive impairment: the J-SHIPP study. *Dement. Geriatr. Cogn. Disord.* 30, 432–439. doi: 10.1159/000321985
- Kang, S., Lee, Y. H., and Lee, J. E. (2017). Metabolism-centric overview of the pathogenesis of Alzheimer's disease. *Yonsei Med. J.* 58, 479–488. doi: 10.3349/ymj.2017.58.3.479
- Kangas-Kontio, T., Huotari, A., Ruotsalainen, H., Herzig, K. H., Tamminen, M., Ala-Korpela, M., et al. (2010). Genetic and environmental determinants of total and high-molecular weight adiponectin in families with low HDL-cholesterol and early onset coronary heart disease. *Atherosclerosis* 210, 479–485. doi: 10.1016/j.atherosclerosis.2009.12.022
- Katsiki, N., Mantzoros, C., and Mikhailidis, D. P. (2017). Adiponectin, lipids and atherosclerosis. *Curr. Opin. Lipidol.* 28, 347–354. doi: 10.1097/mol. 000000000000431
- Kauppila, T. E. S., Kauppila, J. H. K., and Larsson, N. G. (2017). Mammalian mitochondria and aging: an update. *Cell Metab.* 25, 57–71. doi: 10.1016/j.cmet. 2016.09.017
- Kazumi, T., Kawaguchi, A., Hirano, T., and Yoshino, G. (2004). Serum adiponectin is associated with high-density lipoprotein cholesterol, triglycerides and low-density lipoprotein particle size in young healthy men. *Metabolism* 53, 589–593. doi: 10.1016/j.metabol.2003.12.008
- Kennelly, S. P., Lawlor, B. A., and Kenny, R. A. (2009). Blood pressure and the risk for dementia: a double edged sword. *Ageing Res. Rev.* 8, 61–70. doi: 10.1016/j. arr.2008.11.001
- Kiliaan, A. J., Arnoldussen, I. A., and Gustafson, D. R. (2014). Adipokines: a link between obesity and dementia? *Lancet Neurol.* 13, 913–923. doi: 10.1016/s1474-4422(14)70085-7

- Kim, B., and Feldman, E. L. (2015). Insulin resistance as a key link for the increased risk of cognitive impairment in the metabolic syndrome. *Exp. Mol. Med.* 47:e149. doi: 10.1038/emm.2015.3
- Kirkland, J. L. (2013). Translating advances from the basic biology of aging into clinical application. *Exp. Gerontol.* 48, 1–5. doi: 10.1016/j.exger.2012.11.014
- Kitagawa, K., Miwa, K., Okazaki, S., Sakaguchi, M., and Mochizuki, H. (2016). Serum high-molecular-weight adiponectin level and incident dementia in patients with vascular risk factors. *Eur. J. Neurol.* 23, 641–647. doi: 10.1111/ene. 12915
- Kitajima, K., Miura, S., Yamauchi, T., Uehara, Y., Kiya, Y., Rye, K. A., et al. (2011). Possibility of increasing cholesterol efflux by adiponectin and its receptors through the ATP binding cassette transporter A1 in HEK293T cells. *Biochem. Biophys. Res. Commun.* 411, 305–311. doi: 10.1016/j.bbrc.2011. 06.131
- Kobayashi, J., Kusunoki, M., Murase, Y., Kawashiri, M., Higashikata, T., Miwa, K., et al. (2005). Relationship of lipoprotein lipase and hepatic triacylglycerol lipase activity to serum adiponectin levels in Japanese hyperlipidemic men. *Horm. Metab. Res.* 37, 505–509. doi: 10.1055/s-2005-870318
- Koh, E. H., Park, J. Y., Park, H. S., Jeon, M. J., Ryu, J. W., Kim, M., et al. (2007). Essential role of mitochondrial function in adiponectin synthesis in adipocytes. *Diabetes* 56, 2973–2981. doi: 10.2337/db07-0510
- Kroner, Z. (2009). The relationship between Alzheimer's disease and diabetes: type 3 diabetes? *Altern. Med. Rev.* 14, 373–379.
- Lecompte, S., Abou-Samra, M., Boursereau, R., Noel, L., and Brichard, S. M. (2017). Skeletal muscle secretome in Duchenne muscular dystrophy: a pivotal anti-inflammatory role of adiponectin. *Cell Mol. Life Sci.* 74, 2487–2501. doi: 10.1007/s00018-017-2465-5
- Letra, L., Matafome, P., Rodrigues, T., Duro, D., Lemos, R., Baldeiras, I., et al. (2019a). Association between adipokines and biomarkers of Alzheimer's disease: a cross-sectional study. J. Alzheimers Dis. 67, 725–735. doi: 10.3233/JAD-180669
- Letra, L., Rodrigues, T., Matafome, P., Santana, I., and Seica, R. (2019b). Adiponectin and sporadic Alzheimer's disease: clinical and molecular links. *Front. Neuroendocrinol.* 52, 1–11. doi: 10.1016/j.yfrne.2017.10.002
- Levine, D. A., Galecki, A. T., Langa, K. M., Unverzagt, F. W., Kabeto, M. U., Giordani, B., et al. (2015). Trajectory of cognitive decline after incident stroke. *JAMA* 314, 41–51. doi: 10.1001/jama.2015.6968
- Liberale, L., Carbone, F., Bertolotto, M., Bonaventura, A., Vecchie, A., Mach, F., et al. (2018). Serum adiponectin levels predict acute coronary syndrome (ACS) in patients with severe carotid stenosis. *Vascul. Pharmacol.* 102, 37–43. doi: 10.1016/j.vph.2017.12.066
- Lindsay, R. S., Funahashi, T., Hanson, R. L., Matsuzawa, Y., Tanaka, S., Tataranni, P. A., et al. (2002). Adiponectin and development of type 2 diabetes in the Pima Indian population. *Lancet* 360, 57–58. doi: 10.1016/S0140-6736(02)09335-2
- Lindsay, R. S., Funahashi, T., Krakoff, J., Matsuzawa, Y., Tanaka, S., Kobes, S., et al. (2003). Genome-wide linkage analysis of serum adiponectin in the Pima Indian population. *Diabetes* 52, 2419–2425. doi: 10.2337/diabetes.52.9.2419
- Lourenco, M. V., Ferreira, S. T., and De Felice, F. G. (2015). Neuronal stress signaling and eIF2α phosphorylation as molecular links between Alzheimer's disease and diabetes. *Prog. Neurobiol.* 129, 37–57. doi: 10.1016/j.pneurobio. 2015.03.003
- Lovren, F., Pan, Y., Quan, A., Szmitko, P. E., Singh, K. K., Shukla, P. C., et al. (2010). Adiponectin primes human monocytes into alternative anti-inflammatory M2 macrophages. *Am. J. Physiol. Heart Circ. Physiol.* 299, H656–H663. doi: 10.1152/ajpheart.00115.2010
- Luchsinger, J. A. (2008). Adiposity, hyperinsulinemia, diabetes and Alzheimer's disease: an epidemiological perspective. *Eur. J. Pharmacol.* 585, 119–129. doi: 10.1016/j.ejphar.2008.02.048
- Luo, N., Liu, J., Chung, B. H., Yang, Q., Klein, R. L., Garvey, W. T., et al. (2010). Macrophage adiponectin expression improves insulin sensitivity and protects against inflammation and atherosclerosis. *Diabetes* 59, 791–799. doi: 10.2337/db09-1338
- Lusis, A. J. (2000). Atherosclerosis. Nature 407, 233-241. doi: 10.1038/350 25203
- Maeda, N., Shimomura, I., Kishida, K., Nishizawa, H., Matsuda, M., Nagaretani, H., et al. (2002). Diet-induced insulin resistance in mice lacking adiponectin/ACRP30. *Nat. Med.* 8, 731–737. doi: 10.1038/nm724

- Marie, C., Pedard, M., Quirie, A., Tessier, A., Garnier, P., Totoson, P., et al. (2018). Brain-derived neurotrophic factor secreted by the cerebral endothelium: a new actor of brain function? *J. Cereb. Blood Flow Metab.* 38, 935–949. doi: 10.1177/0271678x18766772
- Martins, I. J., Hone, E., Foster, J. K., Sunram-Lea, S. I., Gnjec, A., Fuller, S. J., et al. (2006). Apolipoprotein E, cholesterol metabolism, diabetes, and the convergence of risk factors for Alzheimer's disease and cardiovascular disease. *Mol. Psychiatry* 11, 721–736. doi: 10.1038/sj.mp.4001854
- Matsubara, M., Maruoka, S., and Katayose, S. (2002). Decreased plasma adiponectin concentrations in women with dyslipidemia. J. Clin. Endocrinol. Metab. 87, 2764–2769. doi: 10.1210/jcem.87.6.8550
- Matsuura, F., Oku, H., Koseki, M., Sandoval, J. C., Yuasa-Kawase, M., Tsubakio-Yamamoto, K., et al. (2007). Adiponectin accelerates reverse cholesterol transport by increasing high density lipoprotein assembly in the liver. *Biochem. Biophys. Res. Commun.* 358, 1091–1095. doi: 10.1016/j.bbrc.2007. 05.040
- Mielke, J. G., Taghibiglou, C., Liu, L., Zhang, Y., Jia, Z., Adeli, K., et al. (2005). A biochemical and functional characterization of diet-induced brain insulin resistance. *J. Neurochem.* 93, 1568–1578. doi: 10.1111/j.1471-4159.2005. 03155.x
- Montecucco, F., and Mach, F. (2009). Update on therapeutic strategies to increase adiponectin function and secretion in metabolic syndrome. *Diabetes Obes. Metab.* 11, 445–454. doi: 10.1111/j.1463-1326.2008. 00986.x
- Ng, R. C., and Chan, K. H. (2017). Potential neuroprotective effects of adiponectin in Alzheimer-s disease. *Int. J. Mol. Sci.* 18:E592. doi: 10.3390/ijms180 30592
- Nucera, A., and Hachinski, V. (2018). Cerebrovascular and Alzheimer disease: fellow travelers or partners in crime? J. Neurochem. 144, 513–516. doi: 10.1111/jnc.14283
- O'Brien, J. T., and Thomas, A. (2015). Vascular dementia. *Lancet* 386, 1698–1706. doi: 10.1016/S0140-6736(15)00463-8
- Ohashi, K., Shibata, R., Murohara, T., and Ouchi, N. (2014). Role of anti-inflammatory adipokines in obesity-related diseases. *Trends Endocrinol. Metab.* 25, 348–355. doi: 10.1016/j.tem.2014.03.009
- Oku, H., Matsuura, F., Koseki, M., Sandoval, J. C., Yuasa-Kawase, M., Tsubakio-Yamamoto, K., et al. (2007). Adiponectin deficiency suppresses ABCA1 expression and ApoA-I synthesis in the liver. *FEBS Lett.* 581, 5029–5033. doi: 10.1016/j.febslet.2007.09.038
- O'Neill, S., Bohl, M., Gregersen, S., Hermansen, K., and O'Driscoll, L. (2016). Blood-based biomarkers for metabolic syndrome. *Trends Endocrinol. Metab.* 27, 363–374. doi: 10.1016/j.tem.2016.03.012
- Ortega Moreno, L., Copetti, M., Fontana, A., De Bonis, C., Salvemini, L., Trischitta, V., et al. (2016). Evidence of a causal relationship between high serum adiponectin levels and increased cardiovascular mortality rate in patients with type 2 diabetes. *Cardiovasc. Diabetol.* 15:17. doi: 10.1186/s12933-016-0339-z
- Otabe, S., Yuan, X., Fukutani, T., Wada, N., Hashinaga, T., Nakayama, H., et al. (2007). Overexpression of human adiponectin in transgenic mice results in suppression of fat accumulation and prevention of premature death by high-calorie diet. Am. J. Physiol. Endocrinol. Metab. 293, E210–E218. doi: 10.1152/ajpendo.00645.2006
- Palmer, A. K., and Kirkland, J. L. (2016). Aging and adipose tissue: potential interventions for diabetes and regenerative medicine. *Exp. Gerontol.* 86, 97–105. doi: 10.1016/j.exger.2016.02.013
- Park, S., Kim, D. S., Kwon, D. Y., and Yang, H. J. (2011). Long-term central infusion of adiponectin improves energy and glucose homeostasis by decreasing fat storage and suppressing hepatic gluconeogenesis without changing food intake. J. Neuroendocrinol. 23, 687–698. doi: 10.1111/j.1365-2826.2011.02165.x
- Pedersen, B. K. (2019). Physical activity and muscle-brain crosstalk. Nat. Rev. Endocrinol. 15, 383–392. doi: 10.1038/s41574-019-0174-x
- Prugger, C., Luc, G., Haas, B., Arveiler, D., Machez, E., Ferrieres, J., et al. (2012). Adipocytokines and the risk of ischemic stroke: the PRIME Study. Ann. Neurol. 71, 478–486. doi: 10.1002/ana.22669
- Pruzin, J. J., Nelson, P. T., Abner, E. L., and Arvanitakis, Z. (2018). Review: relationship of type 2 diabetes to human brain pathology. *Neuropathol. Appl. Neurobiol.* 44, 347–362. doi: 10.1111/nan.12476

- Pruzin, J. J., Schneider, J. A., Capuano, A. W., Leurgans, S. E., Barnes, L. L., Ahima, R. S., et al. (2017). Diabetes, hemoglobin A1C, and regional alzheimer disease and infarct pathology. *Alzheimer Dis. Assoc. Disord.* 31, 41–47. doi: 10.1097/wad.00000000000172
- Purnell, C., Gao, S., Callahan, C. M., and Hendrie, H. C. (2009). Cardiovascular risk factors and incident Alzheimer disease: a systematic review of the literature. *Alzheimer Dis. Assoc. Disord.* 23, 1–10. doi: 10.1097/wad.0b013e3181 87541c
- Qi, Y., Takahashi, N., Hileman, S. M., Patel, H. R., Berg, A. H., Pajvani, U. B., et al. (2004). Adiponectin acts in the brain to decrease body weight. *Nat. Med.* 10, 524–529. doi: 10.1038/nm1029
- Qiao, L., Zou, C., van der Westhuyzen, D. R., and Shao, J. (2008). Adiponectin reduces plasma triglyceride by increasing VLDL triglyceride catabolism. *Diabetes* 57, 1824–1833. doi: 10.2337/db07-0435
- Rad, S. K., Arya, A., Karimian, H., Madhavan, P., Rizwan, F., Koshy, S., et al. (2018). Mechanism involved in insulin resistance *via* accumulation of β -amyloid and neurofibrillary tangles: link between type 2 diabetes and Alzheimer's disease. *Drug Des. Devel. Ther.* 12, 3999–4021. doi: 10.2147/dddt. s173970
- Rizzo, M., Kotur-Stevuljevic, J., Berneis, K., Spinas, G., Rini, G. B., Jelic-Ivanovic, Z., et al. (2009). Atherogenic dyslipidemia and oxidative stress: a new look. *Transl. Res.* 153, 217–223. doi: 10.1016/j.trsl.2009.01.008
- Robinson, L., Tang, E., and Taylor, J. P. (2015). Dementia: timely diagnosis and early intervention. BMJ 350:h3029. doi: 10.1136/bmj.h3029
- Salloway, S., Sperling, R., Fox, N. C., Blennow, K., Klunk, W., Raskind, M., et al. (2014). Two phase 3 trials of bapineuzumab in mild-to-moderate Alzheimer's disease. N. Engl. J. Med. 370, 322–333. doi: 10.1056/NEJMoa13 04839
- Sargolzaei, J., Chamani, E., Kazemi, T., Fallah, S., and Soori, H. (2018). The role of adiponectin and adipolin as anti-inflammatory adipokines in the formation of macrophage foam cells and their association with cardiovascular diseases. *Clin. Biochem.* 54, 1–10. doi: 10.1016/j.clinbiochem.2018.02.008
- Savopoulos, C., Michalakis, K., Apostolopoulou, M., Miras, A., and Hatzitolios, A. (2011). Adipokines and stroke: a review of the literature. *Maturitas* 70, 322–327. doi: 10.1016/j.maturitas.2011.09.002
- Scherer, P. E., Williams, S., Fogliano, M., Baldini, G., and Lodish, H. F. (1995). A novel serum protein similar to C1q, produced exclusively in adipocytes. J. Biol. Chem. 270, 26746–26749. doi: 10.1074/jbc.270.45.26746
- Schneider, J. G., von Eynatten, M., Schiekofer, S., Nawroth, P. P., and Dugi, K. A. (2005). Low plasma adiponectin levels are associated with increased hepatic lipase activity *in vivo*. *Diabetes Care* 28, 2181–2186. doi: 10.2337/diacare.28. 9.2181
- Sekiyama, K., Waragai, M., Akatsu, H., Sugama, S., Takenouchi, T., Takamatsu, Y., et al. (2014). Disease-modifying effect of adiponectin in model of α-synucleinopathies. Ann. Clin. Transl. Neurol. 1, 479–489. doi: 10.1002/ acn3.77
- Sente, T., Van Berendoncks, A. M., Hoymans, V. Y., and Vrints, C. J. (2016). Adiponectin resistance in skeletal muscle: pathophysiological implications in chronic heart failure. *J. Cachexia Sarcopenia Muscle* 7, 261–274. doi: 10.1002/jcsm.12086
- Shetty, G. K., Economides, P. A., Horton, E. S., Mantzoros, C. S., and Veves, A. (2004). Circulating adiponectin and resistin levels in relation to metabolic factors, inflammatory markers and vascular reactivity in diabetic patients and subjects at risk for diabetes. *Diabetes Care* 27, 2450–2457. doi: 10.2337/diacare. 27.10.2450
- Shibata, R., Ouchi, N., Kihara, S., Sato, K., Funahashi, T., and Walsh, K. (2004). Adiponectin stimulates angiogenesis in response to tissue ischemia through stimulation of amp-activated protein kinase signaling. *J. Biol. Chem.* 279, 28670–28674. doi: 10.1074/jbc.m402558200
- Shibata, R., Sato, K., Pimentel, D. R., Takemura, Y., Kihara, S., Ohashi, K., et al. (2005). Adiponectin protects against myocardial ischemia-reperfusion injury through AMPK- and COX-2-dependent mechanisms. *Nat. Med.* 11, 1096–1103. doi: 10.1038/nm1295
- Siebel, A. L., Heywood, S. E., and Kingwell, B. A. (2015). HDL and glucose metabolism: current evidence and therapeutic potential. *Front. Pharmacol.* 6:258. doi: 10.3389/fphar.2015.00258
- Snowdon, D. A., Greiner, L. H., Mortimer, J. A., Riley, K. P., Greiner, P. A., and Markesbery, W. R. (1997). Brain infarction and the clinical expression of

Alzheimer disease. The nun study. JAMA 277, 813–817. doi: 10.1001/jama.277. 10.813

- Solanki, A., Bhatt, L. K., and Johnston, T. P. (2018). Evolving targets for the treatment of atherosclerosis. *Pharmacol. Ther.* 187, 1–12. doi: 10.1016/j. pharmthera.2018.02.002
- Song, J., Choi, S. M., Whitcomb, D. J., and Kim, B. C. (2017). Adiponectin controls the apoptosis and the expression of tight junction proteins in brain endothelial cells through AdipoR1 under β amyloid toxicity. *Cell Death Dis.* 8:e3102. doi: 10.1038/cddis.2017.491
- Song, J., Lee, W. T., Park, K. A., and Lee, J. E. (2014). Association between risk factors for vascular dementia and adiponectin. *Biomed. Res. Int.* 2014:261672. doi: 10.1155/2014/261672
- Spranger, J., Kroke, A., Mohlig, M., Bergmann, M. M., Ristow, M., Boeing, H., et al. (2003). Adiponectin and protection against type 2 diabetes mellitus. *Lancet* 361, 226–228. doi: 10.1016/s0140-6736(03)12255-6
- Spranger, J., Verma, S., Gohring, I., Bobbert, T., Seifert, J., Sindler, A. L., et al. (2006). Adiponectin does not cross the blood-brain barrier but modifies cytokine expression of brain endothelial cells. *Diabetes* 55, 141–147. doi: 10.2337/diabetes.55.1.141
- Stott, D. J., Welsh, P., Rumley, A., Robertson, M., Ford, I., Sattar, N., et al. (2009). Adipocytokines and risk of stroke in older people: a nested case-control study. *Int. J. Epidemiol.* 38, 253–261. doi: 10.1093/ije/dyn215
- Stout, M. B., Justice, J. N., Nicklas, B. J., and Kirkland, J. L. (2017). Physiological aging: links among adipose tissue dysfunction, diabetes and frailty. *Physiology* 32, 9–19. doi: 10.1152/physiol.00012.2016
- Sweeney, M. D., Zhao, Z., Montagne, A., Nelson, A. R., and Zlokovic, B. V. (2019). Blood-brain barrier: from physiology to disease and back. *Physiol. Rev.* 99, 21–78. doi: 10.1152/physrev.00050.2017
- Takahashi, R. H., Nagao, T., and Gouras, G. K. (2017). Plaque formation and the intraneuronal accumulation of β-amyloid in Alzheimer's disease. *Pathol. Int.* 67, 185–193. doi: 10.1111/pin.12520
- Teixeira, A. L., Diniz, B. S., Campos, A. C., Miranda, A. S., Rocha, N. P., Talib, L. L., et al. (2013). Decreased levels of circulating adiponectin in mild cognitive impairment and Alzheimer's disease. *Neuromolecular Med.* 15, 115–121. doi: 10.1007/s12017-012-8201-2
- Thundyil, J., Pavlovski, D., Sobey, C. G., and Arumugam, T. V. (2012). Adiponectin receptor signalling in the brain. Br. J. Pharmacol. 165, 313–327. doi: 10.1111/j.1476-5381.2011.01560.x
- Tomono, Y., Hiraishi, C., and Yoshida, H. (2018). Age and sex differences in serum adiponectin and its association with lipoprotein fractions. Ann. Clin. Biochem. 55, 165–171. doi: 10.1177/0004563217699233
- Tosto, G., Bird, T. D., Bennett, D. A., Boeve, B. F., Brickman, A. M., Cruchaga, C., et al. (2016). The role of cardiovascular risk factors and stroke in familial Alzheimer disease. *JAMA Neurol.* 73, 1231–1237. doi: 10.1001/jamaneurol. 2016.2539
- Tsubakio-Yamamoto, K., Sugimoto, T., Nishida, M., Okano, R., Monden, Y., Kitazume-Taneike, R., et al. (2012). Serum adiponectin level is correlated with the size of HDL and LDL particles determined by high performance liquid chromatography. *Metabolism* 61, 1763–1770. doi: 10.1016/j.metabol.2012. 05.011
- Turer, A. T., and Scherer, P. E. (2012). Adiponectin: mechanistic insights and clinical implications. *Diabetologia* 55, 2319–2326. doi: 10.1007/s00125-012-2598-x
- Une, K., Takei, Y. A., Tomita, N., Asamura, T., Ohrui, T., Furukawa, K., et al. (2011). Adiponectin in plasma and cerebrospinal fluid in MCI and Alzheimer's disease. *Eur. J. Neurol.* 18, 1006–1009. doi: 10.1111/j.1468-1331.2010.03194.x
- Van Berendoncks, A. M., Garnier, A., Beckers, P., Hoymans, V. Y., Possemiers, N., Fortin, D., et al. (2010). Functional adiponectin resistance at the level of the skeletal muscle in mild to moderate chronic heart failure. *Circ. Heart Fail.* 3, 185–194. doi: 10.1161/circheartfailure.109.885525
- van Himbergen, T. M., Beiser, A. S., Ai, M., Seshadri, S., Otokozawa, S., Au, R., et al. (2012). Biomarkers for insulin resistance and inflammation and the risk for all-cause dementia and Alzheimer disease: results from the Framingham Heart Study. *Arch. Neurol.* 69, 594–600. doi: 10.1001/archneurol. 2011.670
- Vanhoutte, P. M., Shimokawa, H., Feletou, M., and Tang, E. H. (2017). Endothelial dysfunction and vascular disease—a 30th anniversary update. *Acta Physiol*. 219, 22–96. doi: 10.1111/apha.12646

- Verdile, G., Fuller, S. J., and Martins, R. N. (2015). The role of type 2 diabetes in neurodegeneration. *Neurobiol. Dis.* 84, 22–38. doi: 10.1016/j.nbd.2015. 04.008
- Vijayan, M., and Reddy, P. H. (2016). Stroke, vascular dementia and Alzheimer's disease: molecular links. J. Alzheimers Dis. 54, 427–443. doi: 10.3233/jad-160527
- von Eynatten, M., Schneider, J. G., Humpert, P. M., Rudofsky, G., Schmidt, N., Barosch, P., et al. (2004). Decreased plasma lipoprotein lipase in hypoadiponectinemia: an association independent of systemic inflammation and insulin resistance. *Diabetes Care* 27, 2925–2929. doi: 10.2337/diacare.27. 12.2925
- Wang, Y., Meng, R., Liu, G., Cao, C., Chen, F., Jin, K., et al. (2019). Intracranial atherosclerotic disease. *Neurobiol. Dis.* 124, 118–132. doi: 10.1016/j.nbd.2018. 11.008
- Wang, X., Pu, H., Ma, C., Jiang, T., Wei, Q., Zhang, C., et al. (2014). Adiponectin abates atherosclerosis by reducing oxidative stress. *Med. Sci. Monit.* 20, 1792–1800. doi: 10.12659/msm.892299
- Wang, Z. V., and Scherer, P. E. (2016). Adiponectin, the past two decades. J. Mol. Cell Biol. 8, 93–100. doi: 10.1093/jmcb/mjw011
- Wang, C. H., Wang, C. C., Huang, H. C., and Wei, Y. H. (2013). Mitochondrial dysfunction leads to impairment of insulin sensitivity and adiponectin secretion in adipocytes. *FEBS J.* 280, 1039–1050. doi: 10.1111/febs. 12096
- Wang, M., Wang, D., Zhang, Y., Wang, X., Liu, Y., and Xia, M. (2013). Adiponectin increases macrophages cholesterol efflux and suppresses foam cell formation in patients with type 2 diabetes mellitus. *Atherosclerosis* 229, 62–70. doi: 10.1016/j. atherosclerosis.2013.01.017
- Waragai, M., Adame, A., Trinh, I., Sekiyama, K., Takamatsu, Y., Une, K., et al. (2016). Possible involvement of adiponectin, the anti-diabetes molecule, in the pathogenesis of Alzheimer's disease. J. Alzheimers Dis. 52, 1453–1459. doi: 10.3233/jad-151116
- Waragai, M., Ho, G., Takamatsu, Y., Sekiyama, K., Sugama, S., Takenouchi, T., et al. (2017). Importance of adiponectin activity in the pathogenesis of Alzheimer's disease. Ann. Clin. Transl. Neurol. 4, 591–600. doi: 10.1002/ acn3.436
- Weisberg, S. P., McCann, D., Desai, M., Rosenbaum, M., Leibel, R. L., Ferrante, A. W., et al. (2003). Obesity is associated with macrophage accumulation in adipose tissue. *J. Clin. Invest.* 112, 1796–1808. doi: 10.1172/JCI19246
- WHO MONICA project principal investigators. (1988). The World Health Organization MONICA Project (monitoring trends and determinants in cardiovascular disease): a major international collaboration. J. Clin. Epidemiol. 41, 105–114. doi: 10.1016/0895-4356(88)90084-4
- Wollenweber, F. A., Zietemann, V., Rominger, A., Opherk, C., Bayer-Karpinska, A., Gschwendtner, A., et al. (2014). The determinants of dementia after stroke (DEDEMAS) study: protocol and pilot data. *Int. J. Stroke* 9, 387–392. doi: 10.1111/ijs.12092
- Xu, H., Barnes, G. T., Yang, Q., Tan, G., Yang, D., Chou, C. J., et al. (2003). Chronic inflammation in fat plays a crucial role in the development of obesityrelated insulin resistance. *J. Clin. Invest.* 112, 1821–1830. doi: 10.1172/JCI 19451
- Xu, Z. P., Gan, G. S., Liu, Y. M., Xiao, J. S., Liu, H. X., Mei, B., et al. (2018). Adiponectin attenuates streptozotocin-induced tau hyperphosphorylation and cognitive deficits by rescuing PI3K/Akt/GSK-3β pathway. *Neurochem. Res.* 43, 316–323. doi: 10.1007/s11064-017-2426-2
- Yamamoto, Y., Hirose, H., Saito, I., Tomita, M., Taniyama, M., Matsubara, K., et al. (2002). Correlation of the adipocyte-derived protein adiponectin with insulin resistance index and serum high-density lipoprotein-cholesterol, independent of body mass index, in the Japanese population. *Clin. Sci.* 103, 137–142. doi: 10.1042/cs1030137
- Yanai, H., and Yoshida, H. (2019). Beneficial effects of adiponectin on glucose and lipid metabolism and atherosclerotic progression: mechanisms and perspectives. *Int. J. Mol. Sci.* 20:E1190. doi: 10.3390/ijms200 51190
- Yarchoan, M., Xie, S. X., Kling, M. A., Toledo, J. B., Wolk, D. A., Lee, E. B., et al. (2012). Cerebrovascular atherosclerosis correlates with Alzheimer pathology in neurodegenerative dementias. *Brain* 135, 3749–3756. doi: 10.1093/brain/aws271

- Ye, R., Holland, W. L., Gordillo, R., Wang, M., Wang, Q. A., Shao, M., et al. (2014). Adiponectin is essential for lipid homeostasis and survival under insulin deficiency and promotes β-cell regeneration. *Elife* 3:3. doi: 10.7554/eLife.03851
- Yin, Y. W., Li, J. C., Wang, J. Z., Li, B. H., Pi, Y., Yang, Q. W., et al. (2012). Association between apolipoprotein E gene polymorphism and the risk of vascular dementia: a meta-analysis. *Neurosci. Lett.* 514, 6–11. doi: 10.1016/j. neulet.2012.02.031
- Yokota, T., Oritani, K., Takahashi, I., Ishikawa, J., Matsuyama, A., Ouchi, N., et al. (2000). Adiponectin, a new member of the family of soluble defense collagens, negatively regulates the growth of myelomonocytic progenitors and the functions of macrophages. *Blood* 96, 1723–1732. doi: 10.1182/blood. v96.5.1723
- Zhang, P., Wang, Y., Fan, Y., Tang, Z., and Wang, N. (2009). Overexpression of adiponectin receptors potentiates the antiinflammatory action of subeffective

dose of globular adiponectin in vascular endothelial cells. *Arterioscler. Thromb.* Vasc. Biol. 29, 67–74. doi: 10.1161/atvbaha.108.178061

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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