Check for updates

OPEN ACCESS

EDITED BY Urszula Wojda, Polish Academy of Sciences, Poland

REVIEWED BY Enrica Caterina Pietronigro, University of Verona, Italy Aashutosh Shetti, Northwestern University, United States

*CORRESPONDENCE Tian Wang Is bluewt2000@163.com Guirong Zhang guirong_zhang@126.com

[†]These authors share first authorship

RECEIVED 01 February 2024 ACCEPTED 27 March 2024 PUBLISHED 10 April 2024

CITATION

Wang S, Xie S, Zheng Q, Zhang Z, Wang T and Zhang G (2024) Biofluid biomarkers for Alzheimer's disease. *Front. Aging Neurosci.* 16:1380237. doi: 10.3389/fnagi.2024.1380237

COPYRIGHT

© 2024 Wang, Xie, Zheng, Zhang, Wang and Zhang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Biofluid biomarkers for Alzheimer's disease

Sensen Wang^{1,2†}, Sitan Xie^{1†}, Qinpin Zheng^{1,2}, Zhihui Zhang¹, Tian Wang^{2*} and Guirong Zhang^{1,2*}

¹Shandong Yinfeng Academy of Life Science, Jinan, Shandong, China, ²School of Pharmacy, Key Laboratory of Molecular Pharmacology and Drug Evaluation, Ministry of Education, Collaborative Innovation Center of Advanced Drug Delivery System and Biotech Drugs in Universities of Shandong, Yantai University, Yantai, Shandong, China

Alzheimer's disease (AD) is a multifactorial neurodegenerative disease, with a complex pathogenesis and an irreversible course. Therefore, the early diagnosis of AD is particularly important for the intervention, prevention, and treatment of the disease. Based on the different pathophysiological mechanisms of AD, the research progress of biofluid biomarkers are classified and reviewed. In the end, the challenges and perspectives of future research are proposed.

KEYWORDS

Alzheimer's disease, diagnosis, biofluid, biomarkers, neurodegenerative diseases

1 Introduction

Alzheimer's disease (AD) is the most common type of dementia, which accounts for 60~80% of all cases (Gauthier et al., 2022). According to the "World Alzheimer' Report in 2019," there were about 55 million dementia patients in the world, and this number was expected to reach 152 million by 2055 (Alzheimer's Disease International, 2019). In 2019, the global societal cost of dementia was around 1.3 trillion US dollars, of which 50% was from the economic value of unpaid care. The 2017 World Health Assembly (WHA) recognized dementia as a public health priority (World Health Organization, 2017). Nowadays, dementia has become one of the biggest public health challenges in the world.

AD can be divided into early-onset AD (EOAD) and late-onset AD (LOAD) at the age boundary of 60 or 65. It could also be classified into familial AD (FAD) and sporadic AD (SAD) based on the family history (Xie et al., 2022). LOAD is mainly SAD, accounting for around 95% of all cases. EOAD is relatively rare, which accounts for less than 5% of AD (Alzheimer's Disease International, 2019). Jia et al. (2022) reported a 19-year-old AD patient with the memory impairment occurring at the age of 17, which was the youngest probable AD case in the world.

The amyloid plaques and the neurofibrillary tangles (NFT) are the two main pathological characteristics of AD (Blennow et al., 2006; Kang et al., 2022; Mahaman et al., 2022). The amyloid plaques are formed by deposition of extracellular β -amyloid protein (A β), and the NFT is induced by intracellular tau hyperphosphorylation. The etiology of AD is still unclear. However, it is generally believed that AD is induced by multiple factors, e.g., genetics, biology, environment, and social psychology (Vermunt et al., 2019; Kulichikhin et al., 2021). The potential mechanism of A β -induced neurodegeneration is always the research focus. Human soluble A β dimers and trimers induce progressive loss of hippocampal synapses. When exposes to picomolar level of soluble A β oligomers, pyramidal neurons in rat brain slice significantly reduce the density of dendritic spines and the number of electrophysiologically active synapses (Shankar et al., 2007). A β could directly incorporate into neuronal membranes

of hypothalamic neurons, and participate in the formation of calciumpermeable pores, leading to an increase in intracellular calcium concentration of GT1-7 cells. Therefore, the disruption of calcium homeostasis by "A β -channels" is recognized as the molecular basis for Aβ neurotoxicity. Previous studies indicated that the lipid composition of cell membrane played an important role in the formation of this channel (Kawahara and Kuroda, 2000). Normally, tau proteins bind to microtubules for maintaining the stability of cytoskeleton. The hyperphosphorylated tau aggregates to form paired helical filaments, which have fewer binding sites and unable to attach to microtubules, thus forming NFT (Liu et al., 2019). NFT disintegrates the microtubule network of nerve cells, resulting in the inhibition of cell biochemical communication, the destruction of the cytoskeleton, and ultimately the production of neurotoxicity (Srivastava et al., 2021). Research data showed that $A\beta$ induced the spread of tau pathology in an unknown way leading to neuronal death (Long and Holtzman, 2019; Karran and De Strooper, 2022). Once Aß accumulation exceeding a specific threshold, the spread of tau pathology was significantly accelerated (Karran and De Strooper, 2022). However, the hypothesis that the interaction between A β and tau leads to cytopathology is still required further investigation.

Since the conceptual framework of preclinical AD was officially proposed by the National Institute of Aging and the Alzheimer's Association (NIA-AA) in 2011, accumulating data suggested that cognitive decline occurred continuously and progressively over a long period (Jack et al., 2018). For example, the whole course of AD for a 70-year-old person could take approximately 15–25 years, including ~10-year asymptomatic stage (preclinical stage), 4-year mild cognitive impairment, and 6-year for ultimately developing into dementia (Scheltens et al., 2021). Therefore, the measurement of AD biomarker should be a continuous process that begins before symptom appearance (Fleisher et al., 2015; Jack et al., 2018).

Cerebrospinal fluid (CSF) directly reflects pathological changes in brain. Core CSF biomarkers of AD include $A\beta_{42}$, total tau protein (T-tau) and P-tau. $A\beta_{42}$ reflects cortical amyloid deposition. T-tau indicates the density of neurodegeneration, and P-tau links to the pathological changes in NFT. The increased concentration of CSF T-tau and P-tau was found in AD patients (Reitz and Mayeux, 2014), and their diagnostic accuracy was around 85–90% (Visser et al., 2009). However, the CSF or imaging analysis of AD biomarkers is either invasive or expensive (e.g., 10,000 RMB/per time for positron emission tomography (PET), 1,500 RMB/per time for magnetic resonance imaging (MRI), 300 RMB/per time for computed tomography (CT), 1,000–3,000 RMB/per time for CSF examination), or both (Teunissen et al., 2022).

Based on the characteristics of accessibility, sampling technology, repeatability and cost-effectiveness, blood biomarkers have more advantages than CSF and imaging analysis (Blennow and Zetterberg, 2018). However, the detection of AD biomarkers in blood is much more complex than in CSF (Kulichikhin et al., 2021). First, there is a blood-brain barrier (BBB) in the human body. The capillary endothelium in BBB lacks pores (Ueno et al., 2016), and therefore ions and polar molecules could only cross the BBB in the presence of some transport proteins (Haas, 2018). However, the transporters that are responsible for transporting tau through BBB have not yet been identified (Ueno et al., 2016). When the axon is damaged, proteins are released from the extracellular space of the brain and only a small fraction could enter the bloodstream. These brain proteins are cleaved,

modified, and degraded before or after passing through BBB (Kulichikhin et al., 2021). Second, the blood-cerebrospinal fluid barrier (BCB) is another important barrier. BCB is porous, so small peptides and hydrophilic molecules could pass through it. Due to the death of neuronal cells and intracellular high concentration, tau is firstly released to CSF (Tarasoff-Conway et al., 2015), and then enters the blood through the barrier (Haas, 2018). Because of the existence of BBB and BCB, there is a difference on the concentration between CSF and blood biomarkers. Third, plasma contains multiple background proteins. Some of them are at high levels. Therefore, the analytical techniques of blood samples should have a high sensitivity and specificity in order to detect a small quantity of biomarkers in the complex backgrounds (Kulichikhin et al., 2021).

10.3389/fnagi.2024.1380237

In this review, the traditional and emerging AD biomarkers are summarized and categorized according to main AD pathologies such as amyloidosis, NFT, neurodegeneration, synaptic dysfunction, neuroinflammation, and BBB breakdown (Figure 1). The physiological function and the biofluid level of these markers are described. In addition, the research of some biomarkers on the other neurodegenerative diseases are also reviewed in this article.

2 Biomarkers of amyloidosis

$\textbf{2.1 A}\beta_{\textbf{42}}$

 $A\beta_{40}$ and $A\beta_{42}$ are the most common subtypes in human. Due to the expanded C-terminal, $A\beta_{42}$ is highly hydrophobic and easier to aggregate than $A\beta_{40}$ (Long and Holtzman, 2019). The level of CSF $A\beta_{42}$ in preclinical stage, mild cognitive impairment (MCI), and AD with dementia symptoms was lower than those in control groups (Bateman et al., 2012). It could decrease to around 50% of the healthy individuals (Table 1; Olsson et al., 2016). A neuropathological examination showed that a decrease in CSF $A\beta_{42}$ was associated with an increase in brain amyloid plaques (Strozyk et al., 2003). Elderly persons with a decrease of CSF A β_{42} were also A β -PET positive, and vice versa (Fagan et al., 2006). The studies indicated that a reduction in CSF A β_{42} preceded the formation of plaques (Palmqvist et al., 2016; Kulichikhin et al., 2021). Thus, CSF A β_{42} is considered as a robust biomarker of early AD diagnosis. A decrease in CSF A β_{42} , coupled with an increase in T-tau and P-tau could help to identify the symptomatic AD (Blennow and Hampel, 2003). Low level of CSF A β_{42} , rather than high level of T-tau, could predict cognitive decline (Stomrud et al., 2007). In addition, a significant decrease in CSF $A\beta_{42}$ was observed in Creutzfeldt-Jakob disease (CJD), multiple system atrophy (MSA), and amyotrophic lateral sclerosis (ALS) (Holmberg et al., 2003), which indicated that CSF A β_{42} could be influenced by other factors besides plaque formation (Mahaman et al., 2022).

Plasma A β analysis is one of the most widely applied peripheral biomarker tests of AD (Mahaman et al., 2022). Plasma and CSF A β_{42} had a weak positive correlation, while plasma A β_{42} levels and brain A β deposition had a negative correlation (Janelidze et al., 2016b). The research results regarding the relationship between plasma A β_{42} and cognitive impairment were not consistent. Some studies showed decreased plasma A β_{42} in MCI and AD (Nakamura et al., 2018), while others reported increased A β_{42} level and following decreased trend before or at the beginning of cognitive decline (Mayeux et al., 2003). The inconsistent results are probably caused by differences in sample



FIGURE 1

Classification of AD biofluid biomarkers described in this review based on different pathophysiological mechanisms. The inner ring indicates different potential mechanisms. The outer ring lists the biomarkers. Biomarkers with different colors present the corresponding research in different biological matrices. Black, CSF and blood; Blue, CSF; Red, blood. $A\beta_{42}$, β -amyloid 42; $A\beta_{40}$, β -amyloid 40; BACE1, β -secretase enzyme; BIN1, Bridging integrator 1; NfL, Neurofilament Light Chain; NFT, Neurofibrillary tangles; VILIP-1, Visinin-like Protein 1; SNAP-25, Synaptosome-associated protein 25; NPTX-2, Neuronal pentraxin 2; GAP-43, Growth-associated protein 43; NG, Neurogranin; TREM2, Triggering receptor 2; sTREM2, Soluble Triggering receptor 2; GFAP, Glial fibrillary acidic protein; NRG-1, Neuregulin-1; YKL-40, Human cartilage glycoprotein-39; TRAIL, TNF-related apoptosis-inducing ligand; PDGFR β , Platelet-derived growth factor receptor- β ; BBB, Blood–brain barrier.

TABLE 1 Summarized information of AD amyloidosis biofluid biomarkers presented in this review.

Pathophysiological mechanism	Biomarker	Biological matrices	Trend	Purpose
Amyloidosis	Αβ ₄₂	CSF	Decrease (Olsson et al., 2016)	Diagnosis (Jack et al., 2018)
		Blood	Controversial (Mayeux et al., 2003; Nakamura et al., 2018)	Research (Wang J. et al., 2018)
	$A\beta_{40}$	CSF	No significant change (Kulichikhin et al., 2021)	Research (Janelidze et al., 2016b;
		Blood	Decrease (Janelidze et al., 2016b)	Kulichikhin et al., 2021)
	$A\beta_{42}/A\beta_{40}$	CSF	Decrease (Schindler et al., 2019)	Diagnosis (Jack et al., 2018)
		Blood	Decrease (Janelidze et al., 2016b)	Research (Schindler et al., 2019)
	T -tau/A β_{42}	CSF	Increase (Kaplow et al., 2020)	Research (Kaplow et al., 2020)
	P-tau181/Aβ ₄₂	CSF		
	ΑΡΡ669-711/Αβ ₄₂	Blood	Increase (Nakamura et al., 2018)	Research (Nakamura et al., 2018)
	BACE1	CSF	Increase (Cervellati et al., 2020)	Research (Decourt and Sabbagh, 2011)
		Blood		
	BIN1	Blood	Increase (Sun et al., 2013)	Research (Sun et al., 2013)

inclusion and exclusion criteria, as well as the analytical methods (Wang J. et al., 2018). Elevated levels of plasma $A\beta_{42}$ and $A\beta_{40}$ were also associated with other diseases, such as hypertension, diabetes and ischemic heart disease, indicating significant differences in metabolic process of amyloid proteins between blood and brain (Janelidze et al., 2016b). Despite these differences, all studies confirmed the changes of blood amyloid proteins at the early stages of AD. In the future, the association between AD pathology and blood amyloid protein is required further exploration to promote its clinical application.

2.2 Aβ₄₀

 $A\beta_{40}$ is the most abundant protein fragment hydrolyzed from APP. The concentration of CSF $A\beta_{40}$ had no significant difference during the development of AD (Kulichikhin et al., 2021). Similar to $A\beta_{42}$, there was a weak positive correlation between plasma and CSF $A\beta_{40}$ (Janelidze et al., 2016b). Although $A\beta_{40}$ does not have the same strong cytotoxicity as $A\beta_{42}$, $A\beta_{40}$ aggregates could be detected in cerebral amyloid vasculopathy (Attems et al., 2011). The decreased plasma concentration of $A\beta_{40}$ was observed in AD patients compared with controls (Janelidze et al., 2016b). Biochemical and molecular simulation showed that $A\beta_{40}$ inhibited the aggregation of $A\beta_{42}$ (Jan et al., 2008).

2.3 Αβ₄₂/Αβ₄₀

The impact of total A β variation could be neutralized through normalization of A β_{42} by A β_{40} (Lewczuk et al., 2004). Several studies showed that CSF A $\beta_{42}/A\beta_{40}$ could better diagnose, differentiate, and monitor AD than CSF A β_{42} (Hansson et al., 2019). CSF A $\beta_{42}/A\beta_{40}$ and A β -PET positive had a good correlation (Kulichikhin et al., 2021). Compared to negative A β -PET population, the CSF A $\beta_{42}/A\beta_{40}$ level of positive A β -PET individuals decreased (Schindler et al., 2019). PD and dementia with Lewy body (DLB) had higher level of CSF A β_{40} . Therefore, A $\beta_{42}/A\beta_{40}$ could help to distinguish AD from these diseases (Nutu et al., 2013; Mahaman et al., 2022). The combined application of CSF A $\beta_{42}/A\beta_{40}$ and other CSF biomarkers, e.g., T-tau or P-tau, gave a better prediction of AD (Baldeiras et al., 2018) and the conversion from MCI to AD (Baldeiras et al., 2018; Mahaman et al., 2022). Thus, CSF A $\beta_{42}/A\beta_{40}$ is proposed as a promising biomarker for preclinical AD diagnosis (Hansson et al., 2019; Kulichikhin et al., 2021).

In a 719-person cohort study, the plasma concentration of $A\beta_{42}/$ Aβ40 significantly decreased in MCI and AD patients (Janelidze et al., 2016b). Ovod et al. (2017) reported that plasma $A\beta_{42}/A\beta_{40}$ was decreased in patients with cerebral amyloidosis. Cognitive decline and the risk of AD progression were associated with low plasma $A\beta_{42}/A\beta_{40}$ levels (Verberk et al., 2020). Plasma $A\beta_{42}$ alone was not an accurate biomarker for AD brain pathology, while $A\beta_{42}/A\beta_{40}$ could give a better prediction (Ovod et al., 2017; Schindler et al., 2019). Moreover, plasma $A\beta_{42}/A\beta_{40}$ was associated with A β -PET (Schindler et al., 2019). Cognitively normal individuals with declined $A\beta_{42}/A\beta_{40}$ were observed within Aβ-PET-negative groups, indicating the early stage of AD before plaque formation (Schindler et al., 2019). A mathematical simulation study suggested that the risk of positive Aβ-PET for the above population was 15 times higher than those with normal plasma $A\beta_{42}/A\beta_{40}$ in the next 6 years (Schindler et al., 2019). Additionally, plasma $A\beta_{42}/A\beta_{40}$ and $A\beta_{42}/A\beta_{38}$ had higher accuracy than $A\beta_{42}$ in distinguishing AD from DLB, PD, or subcortical vascular dementia (VaD) (Janelidze et al., 2016c).

2.4 Other Aβ ratios

Elevated CSF T-tau/A β_{42} or P-tau181/A β_{42} indicated an obvious AD brain pathology (Kaplow et al., 2020). CSF A β_{42} /P-tau181 showed high accuracy in predicting the progression from MCI to AD (Buchhave et al., 2012; Mahaman et al., 2022). A study based on mass spectrometry suggested that the plasma levels of APP669-711/A β_{42} and A β_{40} /A β_{42} in A β -PET-positive individuals were higher than those with negative A β -PET scan (Nakamura et al., 2018). These two ratios were associated with the levels of CSF A β_{42} (Kaneko et al., 2014; Nakamura et al., 2018), and they could be used to predict the brain A β burden (Nakamura et al., 2018).

2.5 BACE1

BACE1, also known as β -secretase or β -site APP-lyase 1, is encoded by BACE1 gene, which is primarily expressed in neurons of brain and responsible for A β production (Vassar et al., 1999). It was reported that the increase of BACE1 gene expression or the abnormal function of β -secretase was one of the earliest processes in AD (Yang et al., 2003). Knocking out BACE1 in mice resulted in a significant decrease in A β and CTF β concentration (a 99-amino acid fragment starting with the N-terminal aspartic acid residue of $A\beta$) in the brain (McConlogue et al., 2007). In the SAD brain, the expression of BACE1 significantly increased (Yang et al., 2003). The CSF level of BACE1 protein of AD patients were significantly elevated (Cervellati et al., 2020). The activity of CSF BACE1 was higher in the individuals with the conversion from MCI to AD than those without (Zetterberg et al., 2008). A recent research (Hampel et al., 2021) and two large-scale cohort studies (Zuliani et al., 2020) presented similar trend of CSF BACE1 in peripheral blood, with a 30% increase in serum of AD compared to the control group. In APP/PS-1 transgenic mice, abnormal BACE1 expression in the retina predated behavioral defects. Therefore, BACE1 could be used as a sensitive biomarker for the early diagnosis of AD (Decourt and Sabbagh, 2011; Haas, 2018). However, the application of CSF BACE1 as a clinical AD biomarker is limited by its inter-subject variability and technical difference of assay in the lab (Hampel et al., 2020). The research focus is mainly on the BACE1 gene rather than the level of BACE1 protein in the other neurodegenerative diseases such as PD (Li et al., 2020).

2.6 BIN1

Bridging integrator 1 (BIN1) is a member of amphiphysin proteins family. It is associated with the cytoskeleton and cell membrane. BIN1 plays an important role in the nervous system (Sudwarts et al., 2022). BIN1 is widely expressed in mice and human brain (Miyagawa et al., 2016). It participates in the regulation of endocytose (Wechsler-Reya et al., 1998), and it is important in intracellular vesicles sorting (Pant et al., 2009).

Previous study showed that BIN1 affected the AD molecular pathobiology through the regulation membrane trafficking of AD-related proteins (Miyagawa et al., 2016). BIN1 dependent pathophysiological process is possibly associated with tau pathology through various mechanisms (Thomas et al., 2019). However, there is no further evidence to clarify whether this effect is regulated through the modulation of tau phosphorylation (Thomas et al., 2019), the influence of BIN1-tau interactions (Malki et al., 2017; Sartori et al., 2019), or directly facilitating the propagation of pathogenic tau (Crotti et al., 2019). Genome-wide association studies demonstrated that BIN1 was a genetic risk factor of LOAD (Tan et al., 2013). The depletion of BIN1 enhanced the level of cellular BACE1 by impairing endosomal trafficking and decreased BACE1 lysosomal degradation, leading to the overproduction of AB (Miyagawa et al., 2016). A cohort study including 112 AD and 200 control subjects showed significantly elevated levels of BIN1 mRNA and protein in the plasma of ADs. However, this trend is required further investigation in the larger-scale cohort (Sun et al., 2013).

3 Biomarkers of NFT formation

3.1 P-tau

AD-related NFT are composed of P-tau (Grundke-Iqbal et al., 1986). CSF P-tau was elevated at the preclinical stage of AD (Sato et al., 2018), and it continued to rise during the early clinical stage (Palmqvist et al., 2019). Higher CSF T-tau and P-tau indicated faster disease progression (Wallin et al., 2010). The elevated CSF T-tau reflected CNS degeneration, while increased P-tau was specific to AD (Blennow and Hampel, 2003; Skillbäck et al., 2014). Therefore, P-tau could help to distinguish AD from other neurodegenerative diseases (Hampel et al., 2004). The correlation between CSF P-tau and A β was stronger than that between CSF P-tau and tau-PET (La Joie et al., 2018). This is probably because A β pathology is the reason for the increased CSF P-tau, while tau-PET measures NFT (Sato et al., 2018; Smirnov et al., 2022).

Several CSF P-tau proteins were reported to increase at the preclinical stage of AD. Both P-tau181 and P-tau217 began to increase at the early stage of AD, which were around 20 years before the detection of tau aggregation in the brain (Barthélemy et al., 2020b). Compared to the control groups, the CSF P-tau217 increased by 5 times in AD patients, while P-tau181 only increased by 1.3 times (Barthélemy et al., 2019). CSF or plasma P-tau217 could identify patients with Aβ-PET positive but tau-PET negative, which indicated the changes of CSF P-tau before the detection of tau aggregates (Barthélemy et al., 2020a; Kulichikhin et al., 2021). CSF P-tau205 increased in the advanced AD, and it was related to the increase of T-tau and brain atrophy (Barthélemy et al., 2020b). Studies reported that P-tau231 was the earliest increasing biomarker in AD (Ashton et al., 2021; Smirnov et al., 2022). There are few studies about comparison of different CSF P-tau biomarkers. One study showed that the levels of CSF P-tau181, P-tau199 and P-tau231 were strongly associated with each other (Hampel et al., 2004). By combining the application of two or more biomarkers, higher diagnostic accuracy could be obtained (Hansson et al., 2006). The fragments of the microtubule binding region (MTBR) where tau was much easier to aggregate (Blennow et al., 2020) were more reliable indicators for tau aggregation (Simrén et al., 2023). A mass spectrometry study investigating the role of MTBR showed that CSF MTBR-tau, such as MTBR-tau243, MTBR-tau299 or MTBR-tau354, was a promising biomarker for monitoring target participation in the clinical trials (Horie et al., 2021).

Plasma P-tau was suggested to be the most promising analyte as a screening tool in clinical application. The increase of plasma P-tau was closely related to amyloid plaques and tau tangles (Mattsson-Carlgren et al., 2021). The validity of plasma P-tau as AD biomarker has been verified in neuropathologically confirmed cases (Lantero Rodriguez et al., 2020; Palmqvist et al., 2020), which was not affected by common co-pathologies (such as TDP-43 or α -synuclein pathology) (Smirnov et al., 2022). Studies proved that plasma P-tau could predict the progression from cognitively unimpaired individuals to MCI and eventually to AD (Janelidze et al., 2020). Plasma P-tau had excellent accuracy in distinguishing AD from non-AD tau proteinosis (e.g., progressive supranuclear paralysis or cortical basal degeneration) or non-AD neurodegenerative diseases [e.g., DLB or frontotemporal dementia (FTD)] (Palmqvist et al., 2020; Ashton et al., 2021; Thijssen et al., 2021). Several P-tau proteins such as P-tau181, P-tau217 and P-tau231 have been proposed as AD blood biomarkers (Palmqvist et al., 2020; Ashton et al., 2021; Smirnov et al., 2022). With the progression of the disease, plasma P-tau181 increased steadily (Suárez-Calvet et al., 2020; Thijssen et al., 2020), and it could accurately distinguish AD from non-AD neurodegenerative diseases (Janelidze et al., 2020). Plasma P-tau 217 was at a high level before detection of tau pathology by PET.

The concentrations of P-tau181, T-tau and $A\beta_{42}$ in exosomes of AD were associated with the corresponding CSF biomarkers. They were significantly higher than those in MCI and healthy controls (Jia et al., 2019). Compared to the control groups, P-tau, $A\beta_{42}$ and phosphorylated insulin receptor substrate 1 (IRS-1) of neuronal derived extracellular vesicles (NDEVs) in ADs showed high accuracy in predicting and distinguishing AD (Kapogiannis et al., 2019).

4 Biomarkers of neurodegeneration

In 2018, NIA-AA guideline proposed neurodegeneration as the third biomarker of AD (Table 2; Jack et al., 2018). Although neurodegeneration alone was not enough as a diagnostic marker, its change rate could accurately predict the progression of the disease (Frisoni et al., 2010).

4.1 T-tau

In AD patients, the level of CSF T-tau increased (Shui et al., 2018). Similar to CSF P-tau181, CSF T-tau began to increase after the formation of amyloid plaques (Schindler, 2022). This happened $10 \sim 15$ years earlier before NFT (Bateman et al., 2012; Fleisher et al., 2015). Therefore, CSF T-tau and P-tau181 probably reflected neuron dysfunction caused by A β plaques, rather than tau tangles. The level of CSF T-tau also increased in other diseases, such as stroke, acute neuronal injury, and CJD (Skillbäck et al., 2014). It increased within a few days after acute brain injury, and continued to elevate for several weeks until it finally fell back to the normal range (Zetterberg et al., 2006). Previous studies reported that the highest level of CSF T-tau was observed in the most serious neurodegenerative diseases. For example, CSF T-tau level in CJD patients was 10 to 20 times higher than that in AD (Skillbäck et al., 2014).

Pathophysiological mechanism	Biomarker	Biological matrices	Trend	Purpose
Neurodegeneration	T-tau	CSF	Increase (Shui et al., 2018)	Diagnosis (Jack et al., 2018)
		Blood	Increase (Chen et al., 2019)	Research (Mahaman et al., 2022)
	NfL	CSF	Increase (Zetterberg et al., 2016)	Research (Mattsson et al., 2017; Bridel et al., 2019)
		Blood	Increase (Mattsson et al., 2017)	
	VILIP-1	CSF	Increase (Braunewell, 2012; Halbgebauer et al.,	Research (Mroczko et al., 2015;
		Blood	2022b)	Halbgebauer et al., 2022b)

TABLE 2 Summarized information of AD neurodegeneration biofluid biomarkers presented in this review.

A two-year longitudinal study involving 152 people showed that plasma T-tau and T-tau/A β_{42} could predict the deposition of abnormally folded proteins in the brain (Sutphen et al., 2018). There was a correlation between high plasma T-tau and AD (Chen et al., 2019). Although the plasma T-tau concentration in MCI or AD increased, the research data showed that the range of plasma T-tau in AD overlapped with that in normal population. This hinders the application of plasma T-tau as a diagnostic biomarker (Mahaman et al., 2022).

serum NfL increased in asymptomatic FAD, and it was associated with the stage and severity of the disease (Weston et al., 2017). Due to the concomitant pathologies of axons and peripheral axons, big difference in plasma NfL levels among healthy elderly population limited its specificity (Simrén et al., 2023). Nevertheless, the plasma NfL was more specific in autosomal dominant AD (ADAD) and Down syndrome because of the younger onset age of these disease process (Simrén et al., 2023).

metabolism (Mattsson et al., 2017). Weston et al. pointed out that

4.2 Neurofilament light chain (NfL)

Neurofilament plays an important role in maintaining the neuronal integrity and regulating the transport of intracellular components (Liu et al., 2004; Wagner et al., 2007; Yan et al., 2007; Bruno et al., 2012). Because of the imbalance of tau kinase and hyperphosphorylation activity, phosphatase abnormal of neurofilaments occurred leading to the loss of their stability and axonal injury in the white matter and brain (Gong et al., 2000). Axonal injury released neurofilament proteins into interstitial fluid, and they could be detected in CSF (Bruno et al., 2012). The concentration of NfL in CSF rose at the early clinical stages of AD, correlating with cognitive decline and the changes of brain structure (Zetterberg et al., 2016). A longitudinal study involving 389 subjects showed the elevated CSF NfL level in ADs than healthy controls (Mattsson et al., 2016). The lack of correlation between increased CSF NfL and Aß pathology indicated that NfL was not specific to AD. Higher concentration of CSF NfL indicated the existence of axonal injury (Bridel et al., 2019). Although CSF NfL was not a specific biomarker for a disease, it could help clinicians to investigate whether cognitively impaired patients had neurological causes (Schindler, 2022). Studies showed that combined application of NfL with other biomarkers could greatly improve the accuracy of dementia diagnosis (de Jong et al., 2007; Mattsson et al., 2016). For example, the combination of CSF $A\beta_{42}$ and P-tau181 with CSF NfL improved the diagnosis accuracy of early AD and FTD (de Jong et al., 2007).

Blood NfL was closely related to CSF NfL (Hansson et al., 2017), and it has become an easy-to-obtain biomarker for reflecting the intensity of neuronal damage and neurodegeneration (Simrén et al., 2023). Some studies proposed that blood NfL was an effective tool for the early diagnosis of AD (Fortea et al., 2018). Elevated plasma NfL concentrations were observed in SAD (Ashton et al., 2021). Compared to the healthy controls, the plasma NfL level of AD increased by nearly 150% (Mattsson et al., 2017). The increased NFL was related to the cognitive decline, brain atrophy and low

4.3 VILIP-1

Visinin-like Protein 1 (VILIP-1) is a new biomarker for reflecting the pathological changes of AD. It is a calcium-binding protein of neuronal calcium sensor (NCS), which plays an important role in neuronal signaling cascades (Burgoyne and Weiss, 2001). In AD and other neurodegenerative diseases, neurons showed abnormal Ca²⁺ homeostasis, which resulted in the aberrant regulation of Ca²⁺ ion channels and reduction of Ca²⁺ buffering capacity (Marambaud et al., 2009). VILIP-1 participated in calciummediated neuronal injury through regulation of Ca²⁺ ions. The expression of VILIP-1 was closely related to Aß plaque and NFT in ADs (Braunewell et al., 2001; Braunewell, 2012). CSF VILIP-1 could predict the change rate of global and focal brain atrophy which was similar as T-tau and P-tau181 (Luo et al., 2013). The CSF VILIP-1 level of ApoE £4 carriers was significantly increased, and it was positively correlated with tau and P-tau levels (Wang et al., 2020). A meta-analysis indicated enhanced levels of CSF VILIP-1 in AD compared to the control group. In addition, the CSF VILIP-1 level was higher in MCI patients who progressed to AD than stable MCIs (Mavroudis I. A. et al., 2021). Similarly, several longitudinal studies pointed out that the concentration of VILIP-1 and/or VILIP-1/A β_{42} in CSF could be applied to diagnose diseases at an early stage, and predicted the future cognitive impairment of normal individuals (Luo et al., 2013). For example, the VILIP-1/A β_{42} and VILIP-1 in the CSF could help to differentiate AD from CJD (Halbgebauer et al., 2022b). No significant difference of VILIP-1 was observed between AD and DLB (Mavroudis I. A. et al., 2021). Serum VILIP-1 could not discriminate AD from PD, ALS, and behavioral variant frontotemporal dementia (bvFTD) (Halbgebauer et al., 2022b). It was reported that serum VILIP-1 was correlated with acute ischemic stroke (Stejskal et al., 2011; Liu D. et al., 2020) and the neuronal injury caused by epileptic seizures (Tan et al., 2020). To date, there is no convincing data supporting the correlation between plasma VILIP-1 and AD or other neurodegenerative diseases.

5 Biomarkers of synaptic dysfunction

Synaptic dysfunction is closely correlated with the loss of synaptic integrity and neuropathological changes in the brain, which is another important pathophysiological mechanism of cognitive decline in AD. Synaptic dysfunction is largely driven by A β and tau pathology, and/or by the indirect consequence of neuroinflammatory response (Jackson et al., 2019). Therefore, synaptic alterations are considered as an early pathological event of AD (Scheff et al., 2007). Biomarkers indicating synaptic integrity and plasticity are helpful for early diagnosis and prognosis monitoring of AD (Table 3; Marambaud et al., 2009).

5.1 Presynaptic protein

Synaptosome-associated protein 25 (SNAP-25) exists in synaptic vesicles (Geppert et al., 1994). CSF SNAP-25 is a key participant in synaptic degeneration (Zhang et al., 2018). An increasing trend of CSF SNAP-25 was observed in the AD patients (Öhrfelt et al., 2016). The significantly reduced SNAP-25 in the cerebral cortex indicated the synaptic dysfunction (Davidsson and Blennow, 1998). It was reported that CSF SNAP-25 could differentiate AD from PD, FTD and ALS, and high concentration of CSF SNAP-25 could be applied as a biomarker for both AD and CJD (Halbgebauer et al., 2022a). CSF SNAP-25 level was correlated with APOE £4 (Wang S. et al., 2018). MCI patients carrying APOE £4 showed higher CSF SNAP-25 than non-carriers (Wang S. et al., 2018). There is few research on the correlation between blood SNAP-25 and AD. A study presented the reduced SNAP-25 in the neuron-derived exosomes (NDEs) isolated from serum, and it was associated with the cognition evaluated by Mini-Mental State Examination (MMSE) (Agliardi et al., 2019).

The increase of growth-associated protein 43 (GAP-43) was considered as another biomarker reflecting AD-related synaptic dysfunction. CSF GAP-43 increased with the cognitive decline, and it was correlated with A β plaque and NFT in hippocampus, amygdala, and cortex (Sandelius et al., 2019). CSF GAP-43 could predict the progression from MCI to AD, and this correlation was suspected to be related with *APOE* ϵ 4 (Zhu et al., 2023). Elevated level of CSF GAP-43 was specific to AD compared to the other neurodegenerative diseases, e.g., MCI, ALS, behavioral variant FTD (bvFTD), PD, DLB, primary progressive aphasia (PPA), progressive supranuclear palsy, corticobasal syndrome, and posterior cortical atrophy (PCA). Therefore, it was considered as a promise AD biomarker for the clinical research. However, another study reported a temporary

increase of CSF GAP-43 after ischemic stroke (Sandelius et al., 2018). A blood neuro-exosomal study presented that exosomal GAP-43 could predict MCI 5–7 years in advance (Jia et al., 2021).

Neuronal pentraxin 2 (NPTX-2) is a protein correlated with the inhibitory circuit dysfunction. NPTX-2 showed different trends from the above biomarkers. The level of NPTX-2 in AD cerebral cortex and CSF decreased, and it had a strong correlation with cognitive ability and the volume of hippocampus (Xiao et al., 2017; Nilsson et al., 2021). Reduced CSF NPTX-2 level can predict early AD in adults with Down syndrome (Belbin et al., 2020). A longitudinal CSF proteomics study found that the change rate of NPTX-2 concentration was significantly associated with cognitive decline. CSF NPTX-2 was suggested as a strong biomarker for accelerated cognitive decline (Libiger et al., 2021). NPTX-2 regulates the complement activity and the loss of microglial synapses in the brain. Reduced NPTX-2 level may exacerbate complement-mediated neurodegeneration in FTD (Zhou J. et al., 2023). Studies reported significantly reduced CSF NPTX-2 level in FTD (Das et al., 2023) and DLB (Boiten et al., 2020), indicating that CSF NPTX-2 was not a specific AD biomarker. NPTX-2/tau was closely related to the cognition of AD and MCI, and it had the best discrimination on AD (Galasko et al., 2019). The correlation between serum NPTX-2 and AD still remains unclear. There was an independent correlation between the serum NPTX-2 and cognition of VaD patients (Shao et al., 2020). Reduced NPTX-2 level was observed in the plasma NDEs of AD patients, but this change could not be detected 6-11 years before dementia (Goetzl et al., 2018).

 α -syn induced hyperphosphorylation and aggregation of tau protein (Duka et al., 2006). There was a correlation between CSF α -syn and MMSE score (Korff et al., 2013). Compared to the cognitively normal population, the level of CSF α -syn in AD increased (Hall et al., 2012). CSF α -syn in AD was significantly higher than those in PD and other neurodegenerative diseases (Wang et al., 2015). More research found that α -syn was a protein associated with familial PD, and it has identified as a major protein of the neuropathological hallmark for idiopathic PD (Fayyad et al., 2019).

5.2 Postsynaptic protein

Neurogranin (NG) is the most well-studied postsynaptic protein. It is a protein composed of 78 amino acids, which is involved in synaptic dysfunction or neuronal injury (Represa et al., 1990). NG participates in the induction of synaptic plasticity by accelerating the dissociation of calmodulin, e.g., long-term potentiation (LTP) and long-term depression (LTD) (Kubota et al., 2007). High levels of CSF NG reflected the loss of

TABLE 3 Summarized information of AD synaptic dysfunction biofluid biomarkers presented in this review.

Pathophysiological mechanism	Biomarker	Biological matrices	Trend	Purpose
	SNAP-25	CSF	Increase (Öhrfelt et al., 2016)	Research (Öhrfelt et al., 2016;
		Blood NDEs	Decrease (Agliardi et al., 2019)	Agliardi et al., 2019)
	GAP-43	CSF	Increase (Sandelius et al., 2019)	Research (Sandelius et al., 2019)
Synaptic dysfunction	NPTX-2	CSF	Decrease (Goetzl et al., 2018; Nilsson et al., 2021)	Research (Goetzl et al., 2018; Nilsson
		Blood NDEs		et al., 2021)
	NG	CSF	Increase (Portelius et al., 2018)	Research (Liu W. et al., 2020)
		Blood exosome	Decrease (Liu W. et al., 2020)	

10.3389/fnagi.2024.1380237

brain neurogranin protein (Saunders et al., 2023), and it was positively correlated with A β plaque and tau pathology (Pereira et al., 2017; Portelius et al., 2018). The content of CSF NG48-76 increased significantly indicating the neurodegenerative process in the brain (Kvartsberg et al., 2015). CSF NG in MCI and AD was higher than cognitive normal population, and it could help to predict the progression from MCI to AD (Kester et al., 2015). CSF NG can predict MCI 5-7 years in advance (Jia et al., 2021), but it could not differentiate MCI from FTD or DLB (Mavroudis et al., 2020). The specificity of CSF NG as a biomarker for AD diagnosis is controversial. Studies indicated that the elevation of CSF NG was highly specific to AD (Portelius et al., 2018), while others presented opposite conclusion (Willemse et al., 2021). Blood NG did not show a good trend. Plasma NG had no significant change or in a similar range between AD and healthy controls (De Vos et al., 2015; Kvartsberg et al., 2015). NG level in NDEVs of MCI and AD decreased, and it was correlated with the cognitive decline (Goetzl et al., 2016; Winston et al., 2016; Liu W. et al., 2020). A meta-analysis reported decreased NG level in plasma exosomes, and plasma NG was closely correlated with the cognitive decline (Liu W. et al., 2020). The interpretation of NDEVs biomarkers should be taken with caution because of their controversial origin (Mahaman et al., 2022).

6 Biomarkers of neuroinflammation

Neuropathologist Alois Alzheimer firstly observed lipid accumulation and fibrous structures in glial cells of dementia population's brain (Simrén et al., 2023). Glial cells in CNS include astrocytes and microglia, and they were reported to be involved in AD pathophysiology (Table 4; Simrén et al., 2023).

6.1 TREM2 and sTREM2

Triggering receptor 2 (TREM2) is a cell surface receptor mainly expressed in CNS microglia, and it plays an important role in regulating energy metabolism and phenotypic transformation of microglia. TREM2 acts as a key regulator, allowing microglia to switch between steady state and activated state (disease-associated microglia) (Keren-Shaul et al., 2017). Heterozygous loss-of-function variants in TREM2 were associated with the increasing AD risk (Guerreiro et al., 2013), and another study presented that they also increased the risk of the other neurodegenerative diseases, such as FTD, PD, and ALS (Carmona et al., 2018). Studies reported that TREM2 tripled the risk of AD (Guerreiro et al., 2013), probably through its influence on tau pathology (Lill et al., 2015) and $A\beta$ clearance pathway (Kleinberger et al., 2014). CSF TREM2 level was deranged at the early stage of AD, and it was closely related to neurodegenerative biomarkers such as T-tau and/or P-tau (Heslegrave et al., 2016; Suárez-Calvet et al., 2016b). Compared to the healthy controls, the levels of TREM2 mRNA and protein in peripheral blood of AD were higher (Hu et al., 2014). TREM2 level in MCI-AD patients was significantly higher than that in AD or healthy controls (Casati et al., 2018).

After hydrolysis of the TREM2 protein, the soluble fragments of sTREM2 are produced outside the cell (Colonna and Wang, 2016). Transgelin-2 (TG2) is expressed on neurons. The sTREM2-TG2 interaction mediates the crosstalk between neurons and microglia (Zhang et al., 2023). The concentration of CSF sTREM2 in AD was higher than the healthy controls (Piccio et al., 2016; Suárez-Calvet et al., 2016b). Before the appearance of expected symptoms and after the increasing of A β and T-tau, the level of CSF sTREM2 in ADAD mutation carriers was higher than non-carriers (Suárez-Calvet et al., 2016a). It was suggested that sTREM2 was potential to predict the progression from MCI to AD. The MCI patients with low CSF or high plasma sTREM2 levels had a higher risk of progression to AD. In addition, the concentration of CSF sTREM2 was closely correlated with sTREM2 in plasma (Zhao et al., 2022). The dynamically changes of CSF sTREM2 was observed in preclinical AD. In the absence of neurodegeneration and tau deposition, AB pathology was correlated with the decreased CSF sTREM2. However, tau pathology and neurodegeneration are associated with the increased CSF sTREM2 (Ma et al., 2020). A meta-analysis reported that CSF sTREM2 was significantly elevated in the entire continuum of AD compared to controls. However, an increasing trend of CSF sTREM2 was also observed in PD, multiple sclerosis (MS), FTD and DLB (Zhou W. et al., 2023).

TABLE 4 Summarized information of AD neuroinflammation biofluid biomarkers presented in this review.

Pathophysiological mechanism	Biomarker	Biological matrices	Trend	Purpose
Neuroinflammation	TREM2 & sTREM2	CSF	Controversial (Ma et al., 2020)	Research (Hu et al., 2014; Suárez-Calvet et al., 2016b)
		Blood	Increase (Hu et al., 2014)	
	TAM receptor	CSF	Increase (Brosseron et al., 2022, 2023)	Research (Brosseron et al., 2022, 2023)
		Blood		
	YKL-40	CSF	Increase (Janelidze et al., 2016a)	Research (Janelidze et al., 2016a; Villar-Piqué et al., 2019)
		Blood	Increase (Craig-Schapiro et al., 2010)	
	GFAP	CSF	Increase (Benedet et al., 2021)	Research (Benedet et al., 2021; Cicognola et al., 2021; Katsipis et al., 2021)
		Blood	Increase (Benedet et al., 2021)	
		Saliva	Decrease (Katsipis et al., 2021)	
	NRG-1	CSF	Increase (Chang et al., 2016; Mouton-Liger	Research (Chang et al., 2016; Mouton-Liger et al.,
		Blood	et al., 2020)	2020)
	TRAIL	Blood	Decrease (Wu et al., 2015)	Research (Wu et al., 2015)

6.2 TAM receptor

TAM receptors are widely expressed in various types of cells and tissues in the immune, neurological, vascular, and reproductive systems (Zhou S. et al., 2023). Axl and Mertk are expressed in microglia, but Tyro3 is not (Fourgeaud et al., 2016). The TAM system is an important regulator for microglia to recognize and engulfment of amyloid plaques. Axl and Mertk proteins were correlated with AD (Huang et al., 2021). Axl reduces inflammatory responses, while Mertk plays a role in the phagocytosis of aggregated proteins and cellular debris (Zhou S. et al., 2023). Research reported that Aβ plaque-associated microglia exhibited hyperreactivity upon systemic inflammation and upregulated phagocytic genes in the transgenic AD mouse model, including Axl (Yin et al., 2017). Microglial phagocytosis driven by TAM does not inhibit, but promotes the formation of amyloid plaques. Mertk is the major participant in this process (Huang et al., 2021). Phosphatidylserine (PtdSer) serves as the ligand decorating the plaques for TAM receptors. Mertk, expressed by activated microglia, recognized, and phagocytosed neurons exposed to PtdSer, thereby promoting tau-induced neuronal loss (Pampuscenko et al., 2020; Zhou S. et al., 2023). The increasing of soluble Axl level in CSF reflected the pathophysiology of AD and was positively correlated with P-tau181 (Brosseron et al., 2022). However, extensive clinical data showed that CSF Axl was correlated with larger brain volume and slower cognitive decline, which indicated that Axl had a protective effect on related processes (Brosseron et al., 2022). The serum Axl was significantly elevated in AD, and it was negatively correlated with cognition and structural imaging (Brosseron et al., 2023). However, the specificity of Axl as an AD biomarker is still required further investigation.

6.3 YKL-40

Human cartilage glycoprotein-39 (YKL-40) is a biomarker of inflammation, as well as the activated astrocytes (Olsson et al., 2016). It is primarily produced by astrocytes in the brain and it can predict the neurotoxicity induced by inflammation and other stress signals (Connolly et al., 2023). The research focus on the role of YKL-40 in AD is limited. It was presented that YKL-40 may promote AD progression via altering amyloid burden and neuroinflammatory processes (Connolly et al., 2023). Another study indicated a correlation between YKL-40 and tau pathology (Baldacci et al., 2017a). YKL-40 showed good performance in distinguishing tau-positive patients from controls (Baldacci et al., 2017b). A metaanalysis found that CSF YKL-40 could serve as a biomarker to predict the progression from MCI to AD, as well as for the prognosis of MCI. CSF YKL-40 was significantly elevated in AD, MCI, MCI-AD, and stable MCI compared to the controls, and it was higher in MCI-AD than in the stable MCI (Mavroudis I. et al., 2021). The level of CSF YKL-40 increased in AD and FTD, and it was relatively low in DLB (Craig-Schapiro et al., 2010; Janelidze et al., 2016a). Another study showed that the level of CSF YKL-40 in AD increased compared to non-dementia control groups, DLB and PD (Wennström et al., 2015). The above results suggested that CSF YKL-40 could be used as a non-specific neuroinflammatory biomarker to distinguish AD from PD and DLB (Wennström et al., 2015). The application of plasma YKL-40 in diagnosis of AD is controversial. It was reported that higher plasma YKL-40 in Clinical Dementia Rating (CDR) 1 and 0.5 than CDR 0 (Craig-Schapiro et al., 2010). Another study indicated the possibility of YKL-40 for predicting the progression from MCI to mild AD (Choi et al., 2011). Some studies presented no statistical difference of plasma YKL-40 between AD and controls. However, the significantly increased plasma YKL-40 was observed in CJD and LBD (Craig-Schapiro et al., 2010; Villar-Piqué et al., 2019).

6.4 GFAP

Glial fibrillary acidic protein (GFAP) is the main intermediate filament of the glial cytoskeleton (Pekny and Nilsson, 2005). GFAP mediates insulin-like growth factor 1 signaling pathway that involved in AD pathology (Wang et al., 2023). CSF GFAP increased with plaque deposition (Benedet et al., 2021), and it could be used to predict the progression from MCI to AD (Cicognola et al., 2021). In addition, studies found that CSF GFAP can effectively differentiate healthy population from preclinical ADs (Wojdała et al., 2023). Plasma GFAP is considered as an early biomarker for Aβ pathology, but it is not related with tau pathology (Pereira et al., 2021). Plasma GFAP was sensitive to AD pathology in LBD, especially to Aβ plaque accumulation (Cousins et al., 2023). Studies showed that plasma GFAP level continued to increase as the disease progression (AD dementia>MCI> cognitively normal elderly with Aß positive), but CSF GFAP did not reflect the consistent trend (Verberk et al., 2020; Chatterjee et al., 2021). The reason why plasma GFAP performs better than CSF GFAP as AD biomarker is still unclear. This is probably correlated with the direct release of GFAP into the blood by astrocytic end-feet, and/or different biological degradation between these two matrices (Simrén et al., 2022). Additionally, plasma GFAP can serve as a biomarker for adult AD in Down syndrome, and it could be applied in clinical practice and trials (Montoliu-Gaya et al., 2023). Notably, significantly decreasing GFAP concentration was observed in saliva of MCI and AD compared to controls. Salivary GFAP is suggested as an excellent biomarker for distinguishing MCI or AD from the controls (Katsipis et al., 2021).

6.5 NRG-1

Neuregulin (NRG) plays an important role in the development of nervous system (Esper et al., 2006). NRG-1 is the first and best characterized NRG gene (Pankonin et al., 2009). Soluble NRG (sNRG) was preferentially accumulated on the surface of white matter astrocytes (Pankonin et al., 2009). NRG-1 signaling exerts neuroprotection through the activation of the phosphatidylinositol 3-kinase/Akt (PI3K/Akt) pathway to prevent the neurotoxicity induced by $A\beta_{42}$ (Baik et al., 2016). Another study presented prevention of AD via blocking NRG-1 signaling on microglia (Liu et al., 2023). Both the above studies are based on the animal models. Research found that ADs and MCI-ADs had higher CSF NRG-1 than controls and non-AD dementias, and the CSF NRG-1 was correlated with cognitive evolution (Mouton-Liger et al., 2020). Plasma sNRG-1 level in AD increased and there was a significant correlation between plasma sNRG-1 and MMSE score. The lower MMSE score was correlated with the higher plasma sNRG-1 concentration (Chang et al., 2016). In addition, plasma NRG-1 was found to be correlated with CSF GAP-43, SNAP-25, and NG (Vrillon et al., 2022). Therefore,

6.6 TRAIL

Previous study presented the involvement of TNF-related apoptosis-inducing ligand (TRAIL) in $A\beta$ induced neurotoxicity in a human neuronal cell line (Cantarella et al., 2003). Blockade of the TRAIL-death receptor DR5 prevented $A\beta$ -neurotoxicity (Uberti et al., 2007). It was reported that the TRAIL was specifically expressed in the brains of ADs (Uberti et al., 2004). The reduced plasma TRAIL level was observed in AD group compared to controls (Wu et al., 2015), and it was significantly associated with its level in CSF (Wu et al., 2015). No significant difference was observed in the serum TRAIL between ADs and controls. However, a negative correlation was indicated between serum TRAIL and MMSE scores of AD patients (Genc et al., 2009).

7 Biomarkers of BBB breakdown

ApoE ϵ 4 could accelerate the breakdown of BBB and damage the pericapillary cells of the brain (Halliday et al., 2016). High level of platelet-derived growth factor receptor- β (PDGFR- β) in CSF could predict the cognitive decline of *ApoE* ϵ 4 carriers (Montagne et al., 2020). Brain capillary injury and BBB breakdown occurred in the hippocampus of patients with early cognitive impairment (Nation et al., 2019). However, the BBB disruptions were not correlated with alterations in A β and/or tau levels (Nation et al., 2019; Montagne et al., 2020). BBB breakdown was proposed as an early biomarker for human cognitive impairment that was uncorrelated to A β and tau pathology (Mahaman et al., 2022).

The ratio of CSF to serum albumin was applied as a standard method to measure BBB function (Tibbling et al., 1977; Skillbäck et al., 2017; Menendez-Gonzalez and Gasparovic, 2019). The mean value of this ratio was slightly higher in AD patients with vascular factors when compared to the healthy controls, but no significant difference was observed in AD group without vascular factors (Blennow et al., 1990). It is suggested that the BBB damage in AD was associated with clinical vascular factors but not a result of the disease itself (Blennow et al., 1990). This ratio might help to exclude some of the cerebrovascular diseases and indirectly assist the diagnosis of AD (Reitz and Mayeux, 2014).

8 Outlook

The Ronald and Nancy Reagan Research Institute of the Alzheimer's Association, and the National Institute on Aging Working Group proposed that "ideal" biomarkers for AD diagnosis should meet the following criteria (Khan and Alkon, 2015): (1) the basic characteristics of AD neuropathology could be detected; (2) the application of biomarkers should be validated in neuropathologically confirmed AD cases; (3) biomarkers could diagnose AD at the early stage with high sensitivity and specificity; and (4) the analysis should be repeatable, reliable, non-invasive, simple and cost-effective (Kulichikhin et al., 2021). Several challenges still exist. First, the diagnosis of AD mainly depends on the clinical features. Some clinical features are overlapped between AD and non-AD dementia patients, resulting in a high rate of misdiagnosis at the early stage of AD, especially in non-specialist clinical

centers (Engelborghs et al., 2008). Second, other dementias, e.g., VaD and DLB, have the pathological characteristics of AD (Schneider et al., 2009), which hinders the application of some AD biomarkers. Third, A β plaques and NFT are also presented in the elderly population without cognitive impairment (Price and Morris, 1999). Fourth, the detection of most AD biomarkers has not been standardized. Most of the results are preliminary and retrospective, and lacking comparison between patients (Sunderland et al., 2003; Mahaman et al., 2022). Finally, more meta-analysis or the correlation analysis studies among different biomarkers are insufficient. Further investigation to rank and confirm their importance is required.

In the future, the research could focus on demonstrating the accuracy of AD biomarkers, especially for the blood-based biomarkers. The relationship between AD biomarkers and the pathogenesis of AD needs further exploration. Large-scale head-to-head studies are required to determine the most appropriate application scenario of biomarkers at different stages of AD, involving diagnosis, prediction, prognosis, and clinical trial design (Ossenkoppele et al., 2019).

Author contributions

SW: Writing – review & editing, Writing – original draft. SX: Writing – review & editing, Writing – original draft. QZ: Writing – original draft. ZZ: Writing – original draft. TW: Writing – review & editing, Supervision, Conceptualization. GZ: Writing – review & editing, Supervision, Conceptualization.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by Shandong Yinfeng Academy of Life Science.

Acknowledgments

We thank the Shandong Yinfeng Academy of Life Science for financial support. Special thanks to Professor Guirong Zhang and Professor Tian Wang for their guidance in this work, as well as to Shandong Yinfeng Academy of Life Science and School of Pharmacy of Yantai University for their academic resources and support.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

Agliardi, C., Guerini, F. R., Zanzottera, M., Bianchi, A., Nemni, R., and Clerici, M. (2019). SNAP-25 in serum is carried by exosomes of neuronal origin and is a potential biomarker of Alzheimer's disease. *Mol. Neurobiol.* 56, 5792–5798. doi: 10.1007/s12035-019-1501-x

Alzheimer's Disease International (2019). Alzheimer's disease international: world Alzheimer report 2019. London: Alzheimer's Disease International.

Ashton, N. J., Pascoal, T. A., Karikari, T. K., Benedet, A. L., Lantero-Rodriguez, J., Brinkmalm, G., et al. (2021). Plasma p-tau231: a new biomarker for incipient Alzheimer's disease pathology. *Acta Neuropathol.* 141, 709–724. doi: 10.1007/s00401-021-02275-6

Attems, J., Jellinger, K., Thal, D. R., and Van Nostrand, W. (2011). Review: sporadic cerebral amyloid angiopathy. *Neuropathol. Appl. Neurobiol.* 37, 75–93. doi: 10.1111/j.1365-2990.2010.01137.x

Baik, T. K., Kim, Y. J., Kang, S. M., Song, D. Y., Min, S. S., and Woo, R. S. (2016). Blocking the phosphatidylinositol 3-kinase pathway inhibits neuregulin-1-mediated rescue of neurotoxicity induced by A β 1-42. *J. Pharm. Pharmacol.* 68, 1021–1029. doi: 10.1111/jphp.12563

Baldacci, F., Lista, S., Cavedo, E., Bonuccelli, U., and Hampel, H. (2017a). Diagnostic function of the neuroinflammatory biomarker YKL-40 in Alzheimer's disease and other neurodegenerative diseases. *Expert Rev. Proteomics* 14, 285–299. doi: 10.1080/14789450.2017.1304217

Baldacci, F., Toschi, N., Lista, S., Zetterberg, H., Blennow, K., Kilimann, I., et al. (2017b). Two-level diagnostic classification using cerebrospinal fluid YKL-40 in Alzheimer's disease. *Alzheimers Dement.* 13, 993–1003. doi: 10.1016/j.jalz.2017.01.021

Baldeiras, I., Santana, I., Leitão, M. J., Gens, H., Pascoal, R., Tábuas-Pereira, M., et al. (2018). Addition of the $A\beta 42/40$ ratio to the cerebrospinal fluid biomarker profile increases the predictive value for underlying Alzheimer's disease dementia in mild cognitive impairment. *Alzheimers Res. Ther.* 10:33. doi: 10.1186/s13195-018-0362-2

Barthélemy, N. R., Horie, K., Sato, C., and Bateman, R. J. (2020a). Blood plasma phosphorylated-tau isoforms track CNS change in Alzheimer's disease. *J. Exp. Med.* 217:e20200861. doi: 10.1084/jem.20200861

Barthélemy, N. R., Li, Y., Joseph-Mathurin, N., Gordon, B. A., Hassenstab, J., Benzinger, T. L. S., et al. (2020b). A soluble phosphorylated tau signature links tau, amyloid and the evolution of stages of dominantly inherited Alzheimer's disease. *Nat. Med.* 26, 398–407. doi: 10.1038/s41591-020-0781-z

Barthélemy, N. R., Mallipeddi, N., Moiseyev, P., Sato, C., and Bateman, R. J. (2019). Tau Phosphorylation Rates Measured by Mass Spectrometry Differ in the Intracellular Brain vs. Extracellular Cerebrospinal Fluid Compartments and Are Differentially Affected by Alzheimer's Disease. *Front. Aging Neurosci.* 11:121. doi: 10.3389/ fnagi.2019.00121

Bateman, R. J., Xiong, C., Benzinger, T. L., Fagan, A. M., Goate, A., Fox, N. C., et al. (2012). Clinical and biomarker changes in dominantly inherited Alzheimer's disease. *N. Engl. J. Med.* 367, 795–804. doi: 10.1056/NEJMoa1202753

Belbin, O., Xiao, M. F., Xu, D., Carmona-Iragui, M., Pegueroles, J., Benejam, B., et al. (2020). Cerebrospinal fluid profile of NPTX2 supports role of Alzheimer's disease-related inhibitory circuit dysfunction in adults with Down syndrome. *Mol. Neurodegener.* 15:46. doi: 10.1186/s13024-020-00398-0

Benedet, A. L., Milà-Alomà, M., Vrillon, A., Ashton, N. J., Pascoal, T. A., Lussier, F., et al. (2021). Differences Between Plasma and Cerebrospinal Fluid Glial Fibrillary Acidic Protein Levels Across the Alzheimer Disease Continuum. *JAMA Neurol.* 78, 1471–1483. doi: 10.1001/jamaneurol.2021.3671

Blennow, K., Chen, C., Cicognola, C., Wildsmith, K. R., Manser, P. T., Bohorquez, S. M. S., et al. (2020). Cerebrospinal fluid tau fragment correlates with tau PET: a candidate biomarker for tangle pathology. *Brain* 143, 650–660. doi: 10.1093/brain/awz346

Blennow, K., De Leon, M. J., and Zetterberg, H. (2006). Alzheimer's disease. *Lancet* 368, 387-403. doi: 10.1016/s0140-6736(06)69113-7

Blennow, K., and Hampel, H. (2003). CSF markers for incipient Alzheimer's disease. *Lancet Neurol.* 2, 605–613. doi: 10.1016/s1474-4422(03)00530-1

Blennow, K., Wallin, A., Fredman, P., Karlsson, I., Gottfries, C. G., and Svennerholm, L. (1990). Blood-brain barrier disturbance in patients with Alzheimer's disease is related to vascular factors. *Acta Neurol. Scand.* 81, 323–326. doi: 10.1111/j.1600-0404.1990. tb01563.x

Blennow, K., and Zetterberg, H. (2018). Biomarkers for Alzheimer's disease: current status and prospects for the future. *J. Intern. Med.* 284, 643–663. doi: 10.1111/joim.12816

Boiten, W. A., Van Steenoven, I., Xiao, M. F., Worley, P. F., Noli, B., Cocco, C., et al. (2020). Pathologically Decreased CSF Levels of Synaptic Marker NPTX2 in DLB Are Correlated with Levels of Alpha-Synuclein and VGF. *Cells* 10:38. doi: 10.3390/ cells10010038

Braunewell, K. H. (2012). The visinin-like proteins VILIP-1 and VILIP-3 in Alzheimer's disease-old wine in new bottles. *Front. Mol. Neurosci.* 5:20. doi: 10.3389/fnmol.2012.00020

Braunewell, K., Riederer, P., Spilker, C., Gundelfinger, E. D., Bogerts, B., and Bernstein, H. G. (2001). Abnormal localization of two neuronal calcium sensor proteins,

visinin-like proteins (vilips)-1 and -3, in neocortical brain areas of Alzheimer disease patients. *Dement. Geriatr. Cogn. Disord.* 12, 110-116. doi: 10.1159/000051244

Bridel, C., Van Wieringen, W. N., Zetterberg, H., Tijms, B. M., Teunissen, C. E., Alvarez-Cermeño, J. C., et al. (2019). Diagnostic Value of Cerebrospinal Fluid Neurofilament Light Protein in Neurology: A Systematic Review and Meta-analysis. *JAMA Neurol.* 76, 1035–1048. doi: 10.1001/jamaneurol.2019.1534

Brosseron, F., Maass, A., Kleineidam, L., Ravichandran, K. A., González, P. G., Mcmanus, R. M., et al. (2022). Soluble TAM receptors sAXL and sTyro3 predict structural and functional protection in Alzheimer's disease. *Neuron* 110, 1009–1022.e4. doi: 10.1016/j.neuron.2021.12.016

Brosseron, F., Maass, A., Kleineidam, L., Ravichandran, K. A., Kolbe, C. C., Wolfsgruber, S., et al. (2023). Serum IL-6, sAXL, and YKL-40 as systemic correlates of reduced brain structure and function in Alzheimer's disease: results from the DELCODE study. *Alzheimers Res. Ther.* 15:13. doi: 10.1186/s13195-022-01118-0

Bruno, D., Pomara, N., Nierenberg, J., Ritchie, J. C., Lutz, M. W., Zetterberg, H., et al. (2012). Levels of cerebrospinal fluid neurofilament light protein in healthy elderly vary as a function of TOMM40 variants. *Exp. Gerontol.* 47, 347–352. doi: 10.1016/j. exger.2011.09.008

Buchhave, P., Minthon, L., Zetterberg, H., Wallin, A. K., Blennow, K., and Hansson, O. (2012). Cerebrospinal fluid levels of β -amyloid 1-42, but not of tau, are fully changed already 5 to 10 years before the onset of Alzheimer dementia. *Arch. Gen. Psychiatry* 69, 98–106. doi: 10.1001/archgenpsychiatry.2011.155

Burgoyne, R. D., and Weiss, J. L. (2001). The neuronal calcium sensor family of Ca2+binding proteins. *Biochem. J.* 353, 1–12.

Cantarella, G., Uberti, D., Carsana, T., Lombardo, G., Bernardini, R., and Memo, M. (2003). Neutralization of TRAIL death pathway protects human neuronal cell line from beta-amyloid toxicity. *Cell Death Differ.* 10, 134–141. doi: 10.1038/sj.cdd.4401143

Carmona, S., Zahs, K., Wu, E., Dakin, K., Bras, J., and Guerreiro, R. (2018). The role of TREM2 in Alzheimer's disease and other neurodegenerative disorders. *Lancet Neurol.* 17, 721–730. doi: 10.1016/s1474-4422(18)30232-1

Casati, M., Ferri, E., Gussago, C., Mazzola, P., Abbate, C., Bellelli, G., et al. (2018). Increased expression of TREM2 in peripheral cells from mild cognitive impairment patients who progress into Alzheimer's disease. *Eur. J. Neurol.* 25, 805–810. doi: 10.1111/ ene.13583

Cervellati, C., Trentini, A., Rosta, V., Passaro, A., Bosi, C., Sanz, J. M., et al. (2020). Serum beta-secretase 1 (BACE1) activity as candidate biomarker for late-onset Alzheimer's disease. *Geroscience* 42, 159–167. doi: 10.1007/s11357-019-00127-6

Chang, K. A., Shin, K. Y., Nam, E., Lee, Y. B., Moon, C., Suh, Y. H., et al. (2016). Plasma soluble neuregulin-1 as a diagnostic biomarker for Alzheimer's disease. *Neurochem. Int.* 97, 1–7. doi: 10.1016/j.neuint.2016.04.012

Chatterjee, P., Pedrini, S., Stoops, E., Goozee, K., Villemagne, V. L., Asih, P. R., et al. (2021). Plasma glial fibrillary acidic protein is elevated in cognitively normal older adults at risk of Alzheimer's disease. *Transl. Psychiatry* 11:27. doi: 10.1038/s41398-020-01137-1

Chen, T. B., Lee, Y. J., Lin, S. Y., Chen, J. P., Hu, C. J., Wang, P. N., et al. (2019). Plasma A β 42 and Total Tau Predict Cognitive Decline in Amnestic Mild Cognitive Impairment. *Sci. Rep.* 9:13984. doi: 10.1038/s41598-019-50315-9

Choi, J., Lee, H. W., and Suk, K. (2011). Plasma level of chitinase 3-like 1 protein increases in patients with early Alzheimer's disease. *J. Neurol.* 258, 2181–2185. doi: 10.1007/s00415-011-6087-9

Cicognola, C., Janelidze, S., Hertze, J., Zetterberg, H., Blennow, K., Mattsson-Carlgren, N., et al. (2021). Plasma glial fibrillary acidic protein detects Alzheimer pathology and predicts future conversion to Alzheimer dementia in patients with mild cognitive impairment. *Alzheimers Res. Ther.* 13:68. doi: 10.1186/ s13195-021-00804-9

Colonna, M., and Wang, Y. (2016). TREM2 variants: new keys to decipher Alzheimer disease pathogenesis. *Nat. Rev. Neurosci.* 17, 201–207. doi: 10.1038/nrn.2016.7

Connolly, K., Lehoux, M., O'rourke, R., Assetta, B., Erdemir, G. A., Elias, J. A., et al. (2023). Potential role of chitinase-3-like protein 1 (CHI3L1/YKL-40) in neurodegeneration and Alzheimer's disease. *Alzheimers Dement*. 19, 9–24. doi: 10.1002/alz.12612

Cousins, K., Irwin, D. J., Chen-Plotkin, A., Shaw, L. M., Arezoumandan, S., Lee, E. B., et al. (2023). Plasma GFAP associates with secondary Alzheimer's pathology in Lewy body disease. *Ann. Clin. Transl. Neurol.* 10, 802–813. doi: 10.1002/acn3.51768

Craig-Schapiro, R., Perrin, R. J., Roe, C. M., Xiong, C., Carter, D., Cairns, N. J., et al. (2010). YKL-40: a novel prognostic fluid biomarker for preclinical Alzheimer's disease. *Biol. Psychiatry* 68, 903–912. doi: 10.1016/j.biopsych.2010.08.025

Crotti, A., Sait, H. R., Mcavoy, K. M., Estrada, K., Ergun, A., Szak, S., et al. (2019). BIN1 favors the spreading of Tau via extracellular vesicles. *Sci. Rep.* 9:9477. doi: 10.1038/ s41598-019-45676-0

Das, S., Goossens, J., Jacobs, D., Dewit, N., Pijnenburg, Y., In 't Veld, S. G. J. G., et al. (2023). Synaptic biomarkers in the cerebrospinal fluid associate differentially with classical neuronal biomarkers in patients with Alzheimer's disease and frontotemporal dementia. *Alzheimers Res. Ther.* 15:62. doi: 10.1186/s13195-023-01212-x

Davidsson, P., and Blennow, K. (1998). Neurochemical dissection of synaptic pathology in Alzheimer's disease. *Int. Psychogeriatr.* 10, 11–23. doi: 10.1017/s1041610298005110

De Jong, D., Jansen, R. W., Pijnenburg, Y. A., Van Geel, W. J., Borm, G. F., Kremer, H. P., et al. (2007). CSF neurofilament proteins in the differential diagnosis of dementia. *J. Neurol. Neurosurg. Psychiatry* 78, 936–938. doi: 10.1136/jnnp.2006.107326

De Vos, A., Jacobs, D., Struyfs, H., Fransen, E., Andersson, K., Portelius, E., et al. (2015). C-terminal neurogranin is increased in cerebrospinal fluid but unchanged in plasma in Alzheimer's disease. *Alzheimers Dement.* 11, 1461–1469. doi: 10.1016/j. jalz.2015.05.012

Decourt, B., and Sabbagh, M. N. (2011). BACE1 as a potential biomarker for Alzheimer's disease. J. Alzheimers Dis. 24, 53-59. doi: 10.3233/jad-2011-110017

Duka, T., Rusnak, M., Drolet, R. E., Duka, V., Wersinger, C., Goudreau, J. L., et al. (2006). Alpha-synuclein induces hyperphosphorylation of Tau in the MPTP model of parkinsonism. *FASEB J.* 20, 2302–2312. doi: 10.1096/fj.06-6092com

Engelborghs, S., De Vreese, K., Van De Casteele, T., Vanderstichele, H., Van Everbroeck, B., Cras, P., et al. (2008). Diagnostic performance of a CSF-biomarker panel in autopsy-confirmed dementia. *Neurobiol. Aging* 29, 1143–1159. doi: 10.1016/j. neurobiolaging.2007.02.016

Esper, R. M., Pankonin, M. S., and Loeb, J. A. (2006). Neuregulins: versatile growth and differentiation factors in nervous system development and human disease. *Brain Res. Rev.* 51, 161–175. doi: 10.1016/j.brainresrev.2005.11.006

Fagan, A. M., Mintun, M. A., Mach, R. H., Lee, S. Y., Dence, C. S., Shah, A. R., et al. (2006). Inverse relation between in vivo amyloid imaging load and cerebrospinal fluid Abeta42 in humans. *Ann. Neurol.* 59, 512–519. doi: 10.1002/ana.20730

Fayyad, M., Salim, S., Majbour, N., Erskine, D., Stoops, E., Mollenhauer, B., et al. (2019). Parkinson's disease biomarkers based on α -synuclein. J. Neurochem. 150, 626–636. doi: 10.1111/jnc.14809

Fleisher, A. S., Chen, K., Quiroz, Y. T., Jakimovich, L. J., Gutierrez Gomez, M., Langois, C. M., et al. (2015). Associations between biomarkers and age in the presenilin 1 E280A autosomal dominant Alzheimer disease kindred: a cross-sectional study. *JAMA Neurol.* 72, 316–324. doi: 10.1001/jamaneurol.2014.3314

Fortea, J., Carmona-Iragui, M., Benejam, B., Fernández, S., Videla, L., Barroeta, I., et al. (2018). Plasma and CSF biomarkers for the diagnosis of Alzheimer's disease in adults with Down syndrome: a cross-sectional study. *Lancet Neurol.* 17, 860–869. doi: 10.1016/s1474-4422(18)30285-0

Fourgeaud, L., Través, P. G., Tufail, Y., Leal-Bailey, H., Lew, E. D., Burrola, P. G., et al. (2016). TAM receptors regulate multiple features of microglial physiology. *Nature* 532, 240–244. doi: 10.1038/nature17630

Frisoni, G. B., Fox, N. C., Jack, C. R. Jr., Scheltens, P., and Thompson, P. M. (2010). The clinical use of structural MRI in Alzheimer disease. *Nat. Rev. Neurol.* 6, 67–77. doi: 10.1038/nrneurol.2009.215

Galasko, D., Xiao, M., Xu, D., Smirnov, D., Salmon, D. P., Dewit, N., et al. (2019). Synaptic biomarkers in CSF aid in diagnosis, correlate with cognition and predict progression in MCI and Alzheimer's disease. *Alzheimers Dement* 5, 871–882. doi: 10.1016/j.trci.2019.11.002

Gauthier, S., Wevster, C., Servaes, S., Morais, J. A., and Rosa-Neto, P. (2022). World Alzheimer report 2022 life after diagnosis: navigating treatment, care and support. (London, England: Alzheimer's Disease International).

Genc, S., Egrilmez, M. Y., Yaka, E., Cavdar, Z., Iyilikci, L., Yener, G., et al. (2009). TNFrelated apoptosis-inducing ligand level in Alzheimer's disease. *Neurol. Sci.* 30, 263–267. doi: 10.1007/s10072-009-0047-5

Geppert, M., Goda, Y., Hammer, R. E., Li, C., Rosahl, T. W., Stevens, C. F., et al. (1994). Synaptotagmin I: a major Ca2+ sensor for transmitter release at a central synapse. *Cell* 79, 717–727. doi: 10.1016/0092-8674(94)90556-8

Goetzl, E. J., Abner, E. L., Jicha, G. A., Kapogiannis, D., and Schwartz, J. B. (2018). Declining levels of functionally specialized synaptic proteins in plasma neuronal exosomes with progression of Alzheimer's disease. *FASEB J.* 32, 888–893. doi: 10.1096/ fi.201700731R

Goetzl, E. J., Kapogiannis, D., Schwartz, J. B., Lobach, I. V., Goetzl, L., Abner, E. L., et al. (2016). Decreased synaptic proteins in neuronal exosomes of frontotemporal dementia and Alzheimer's disease. *FASEB J.* 30, 4141–4148. doi: 10.1096/fj.201600816R

Gong, C. X., Lidsky, T., Wegiel, J., Zuck, L., Grundke-Iqbal, I., and Iqbal, K. (2000). Phosphorylation of microtubule-associated protein tau is regulated by protein phosphatase 2A in mammalian brain. Implications for neurofibrillary degeneration in Alzheimer's disease. *J. Biol. Chem.* 275, 5535–5544. doi: 10.1074/ jbc.275.8.5535

Grundke-Iqbal, I., Iqbal, K., Tung, Y. C., Quinlan, M., Wisniewski, H. M., and Binder, L. I. (1986). Abnormal phosphorylation of the microtubule-associated protein tau (tau) in Alzheimer cytoskeletal pathology. *Proc. Natl. Acad. Sci. U. S. A.* 83, 4913–4917. doi: 10.1073/pnas.83.13.4913

Guerreiro, R., Wojtas, A., Bras, J., Carrasquillo, M., Rogaeva, E., Majounie, E., et al. (2013). TREM2 variants in Alzheimer's disease. *N. Engl. J. Med.* 368, 117–127. doi: 10.1056/NEJMoa1211851

Haas, D. (2018). "Chapter 15- Biomarker for Alzheimer's disease" in *Precision medicine*. eds. H.-P. Deigner and M. Kohl (London: Academic Press)

Halbgebauer, S., Steinacker, P., Hengge, S., Oeckl, P., Abu Rumeileh, S., Anderl-Straub, S., et al. (2022a). CSF levels of SNAP-25 are increased early in Creutzfeldt-Jakob and Alzheimer's disease. *J. Neurol. Neurosurg. Psychiatry*. doi: 10.1136/jnnp-2021-328646

Halbgebauer, S., Steinacker, P., Riedel, D., Oeckl, P., Anderl-Straub, S., Lombardi, J., et al. (2022b). Visinin-like protein 1 levels in blood and CSF as emerging markers for Alzheimer's and other neurodegenerative diseases. *Alzheimers Res. Ther.* 14:175. doi: 10.1186/s13195-022-01122-4

Hall, S., Öhrfelt, A., Constantinescu, R., Andreasson, U., Surova, Y., Bostrom, F., et al. (2012). Accuracy of a panel of 5 cerebrospinal fluid biomarkers in the differential diagnosis of patients with dementia and/or parkinsonian disorders. *Arch. Neurol.* 69, 1445–1452. doi: 10.1001/archneurol.2012.1654

Halliday, M. R., Rege, S. V., Ma, Q., Zhao, Z., Miller, C. A., Winkler, E. A., et al. (2016). Accelerated pericyte degeneration and blood-brain barrier breakdown in apolipoprotein E4 carriers with Alzheimer's disease. *J. Cereb. Blood Flow Metab.* 36, 216–227. doi: 10.1038/icbfm.2015.44

Hampel, H., Buerger, K., Zinkowski, R., Teipel, S. J., Goernitz, A., Andreasen, N., et al. (2004). Measurement of Phosphorylated Tau Epitopes in the Differential Diagnosisof Alzheimer Disease: A Comparative Cerebrospinal Fluid Study. *Arch. Gen. Psychiatry* 61, 95–102. doi: 10.1001/archpsyc.61.1.95

 $\begin{array}{l} Hampel, H., Lista, S., Vanmechelen, E., Zetterberg, H., Giorgi, F. S., Galgani, A., et al. \\ (2020). \\ \beta-Secretase1 biological markers for Alzheimer's disease: state-of-art of validation and qualification. \\ Alzheimers Res. Ther. 12:130. \\ doi: 10.1186/s13195-020-00686-3 \end{array}$

Hampel, H., Vassar, R., de Strooper, B., Hardy, J., Willem, M., Singh, N., et al. (2021). The β -Secretase BACE1 in Alzheimer's Disease. *Biol. Psychiatry* 89, 745–756. doi: 10.1016/j.biopsych.2020.02.001

Hansson, O., Janelidze, S., Hall, S., Magdalinou, N., Lees, A. J., Andreasson, U., et al. (2017). Blood-based NfL: A biomarker for differential diagnosis of parkinsonian disorder. *Neurology* 88, 930–937. doi: 10.1212/wnl.00000000003680

Hansson, O., Lehmann, S., Otto, M., Zetterberg, H., and Lewczuk, P. (2019). Advantages and disadvantages of the use of the CSF Amyloid β (A β) 42/40 ratio in the diagnosis of Alzheimer's Disease. *Alzheimers Res. Ther.* 11:34. doi: 10.1186/s13195-019-0485-0

Hansson, O., Zetterberg, H., Buchhave, P., Londos, E., Blennow, K., and Minthon, L. (2006). Association between CSF biomarkers and incipient Alzheimer's disease in patients with mild cognitive impairment: a follow-up study. *Lancet Neurol.* 5, 228–234. doi: 10.1016/s1474-4422(06)70355-6

Heslegrave, A., Heywood, W., Paterson, R., Magdalinou, N., Svensson, J., Johansson, P., et al. (2016). Increased cerebrospinal fluid soluble TREM2 concentration in Alzheimer's disease. *Mol. Neurodegener.* 11:3. doi: 10.1186/s13024-016-0071-x

Holmberg, B., Johnels, B., Blennow, K., and Rosengren, L. (2003). Cerebrospinal fluid Abeta42 is reduced in multiple system atrophy but normal in Parkinson's disease and progressive supranuclear palsy. *Mov. Disord.* 18, 186–190. doi: 10.1002/mds.10321

Horie, K., Barthélemy, N. R., Sato, C., and Bateman, R. J. (2021). CSF tau microtubule binding region identifies tau tangle and clinical stages of Alzheimer's disease. *Brain* 144, 515–527. doi: 10.1093/brain/awaa373

Hu, N., Tan, M. S., Yu, J. T., Sun, L., Tan, L., Wang, Y. L., et al. (2014). Increased expression of TREM2 in peripheral blood of Alzheimer's disease patients. *J. Alzheimers Dis.* 38, 497–501. doi: 10.3233/jad-130854

Huang, Y., Happonen, K. E., Burrola, P. G., O'connor, C., Hah, N., Huang, L., et al. (2021). Microglia use TAM receptors to detect and engulf amyloid β plaques. *Nat. Immunol.* 22, 586–594. doi: 10.1038/s41590-021-00913-5

Jack, C. R. Jr., Bennett, D. A., Blennow, K., Carrillo, M. C., Dunn, B., Haeberlein, S. B., et al. (2018). NIA-AA Research Framework: Toward a biological definition of Alzheimer's disease. *Alzheimers Dement*. 14, 535–562. doi: 10.1016/j.jalz.2018.02.018

Jackson, J., Jambrina, E., Li, J., Marston, H., Menzies, F., Phillips, K., et al. (2019). Targeting the Synapse in Alzheimer's Disease. *Front. Neurosci.* 13:735. doi: 10.3389/ fnins.2019.00735

Jan, A., Gokce, O., Luthi-Carter, R., and Lashuel, H. A. (2008). The ratio of monomeric to aggregated forms of Abeta40 and Abeta42 is an important determinant of amyloidbeta aggregation, fibrillogenesis, and toxicity. *J. Biol. Chem.* 283, 28176–28189. doi: 10.1074/jbc.M803159200

Janelidze, S., Hertze, J., Zetterberg, H., Landqvist Waldö, M., Santillo, A., Blennow, K., et al. (2016a). Cerebrospinal fluid neurogranin and YKL-40 as biomarkers of Alzheimer's disease. *Ann. Clin. Transl. Neurol.* 3, 12–20. doi: 10.1002/acn3.266

Janelidze, S., Mattsson, N., Palmqvist, S., Smith, R., Beach, T. G., Serrano, G. E., et al. (2020). Plasma P-tau181 in Alzheimer's disease: relationship to other biomarkers, differential diagnosis, neuropathology and longitudinal progression to Alzheimer's dementia. *Nat. Med.* 26, 379–386. doi: 10.1038/s41591-020-0755-1

Janelidze, S., Stomrud, E., Palmqvist, S., Zetterberg, H., Van Westen, D., Jeromin, A., et al. (2016b). Plasma β -amyloid in Alzheimer's disease and vascular disease. *Sci. Rep.* 6:26801. doi: 10.1038/srep26801

Janelidze, S., Zetterberg, H., Mattsson, N., Palmqvist, S., Vanderstichele, H., Lindberg, O., et al. (2016c). CSF A β 42/A β 40 and A β 42/A β 38 ratios: better diagnostic markers of Alzheimer disease. *Ann. Clin. Transl. Neurol.* 3, 154–165. doi: 10.1002/acn3.274

Jia, L., Qiu, Q., Zhang, H., Chu, L., Du, Y., Zhang, J., et al. (2019). Concordance between the assessment of A β 42, T-tau, and P-T181-tau in peripheral blood neuronal-derived exosomes and cerebrospinal fluid. *Alzheimers Dement.* 15, 1071–1080. doi: 10.1016/j.jalz.2019.05.002

Jia, J., Zhang, Y., Shi, Y., Yin, X., Wang, S., Li, Y., et al. (2022). A 19-Year-Old Adolescent with Probable Alzheimer's Disease 1. *J. Alzheimers Dis.* 91, 915–922. doi: 10.3233/JAD-221065

Jia, L., Zhu, M., Kong, C., Pang, Y., Zhang, H., Qiu, Q., et al. (2021). Blood neuroexosomal synaptic proteins predict Alzheimer's disease at the asymptomatic stage. *Alzheimers Dement.* 17, 49–60. doi: 10.1002/alz.12166

Kaneko, N., Nakamura, A., Washimi, Y., Kato, T., Sakurai, T., Arahata, Y., et al. (2014). Novel plasma biomarker surrogating cerebral amyloid deposition. *Proc. Jpn. Acad. Ser. B Phys. Biol. Sci.* 90, 353–364. doi: 10.2183/pjab.90.353

Kang, J., Tian, Z., Wei, J., Mu, Z., Liang, J., and Li, M. (2022). Association between obstructive sleep apnea and Alzheimer's disease-related blood and cerebrospinal fluid biomarkers: a meta-analysis. *J. Clin. Neurosci.* 102, 87–94. doi: 10.1016/j.jocn.2022.06.004

Kaplow, J., Vandijck, M., Gray, J., Kanekiyo, M., Huyck, E., Traynham, C. J., et al. (2020). Concordance of Lumipulse cerebrospinal fluid t-tau/Aβ42 ratio with amyloid PET status. *Alzheimers Dement.* 16, 144–152. doi: 10.1002/alz.12000

Kapogiannis, D., Mustapic, M., Shardell, M. D., Berkowitz, S. T., Diehl, T. C., Spangler, R. D., et al. (2019). Association of Extracellular Vesicle Biomarkers With Alzheimer Disease in the Baltimore Longitudinal Study of Aging. *JAMA Neurol.* 76, 1340–1351. doi: 10.1001/jamaneurol.2019.2462

Karran, E., and De Strooper, B. (2022). The amyloid hypothesis in Alzheimer disease: new insights from new therapeutics. *Nat. Rev. Drug Discov.* 21, 306–318. doi: 10.1038/ s41573-022-00391-w

Katsipis, G., Tzekaki, E. E., Tsolaki, M., and Pantazaki, A. A. (2021). Salivary GFAP as a potential biomarker for diagnosis of mild cognitive impairment and Alzheimer's disease and its correlation with neuroinflammation and apoptosis. *J. Neuroimmunol.* 361:577744. doi: 10.1016/j.jneuroim.2021.577744

Kawahara, M., and Kuroda, Y. (2000). Molecular mechanism of neurodegeneration induced by Alzheimer's β -amyloid protein: channel formation and disruption of calcium homeostasis. *Brain Res. Bull.* 53, 389–397. doi: 10.1016/S0361-9230(00)00370-1

Keren-Shaul, H., Spinrad, A., Weiner, A., Matcovitch-Natan, O., Dvir-Szternfeld, R., Ulland, T. K., et al. (2017). A Unique Microglia Type Associated with Restricting Development of Alzheimer's Disease. *Cell* 169, 1276–1290.e17. doi: 10.1016/j. cell.2017.05.018

Kester, M. I., Teunissen, C. E., Crimmins, D. L., Herries, E. M., Ladenson, J. H., Scheltens, P., et al. (2015). Neurogranin as a Cerebrospinal Fluid Biomarker for Synaptic Loss in Symptomatic Alzheimer Disease. *JAMA Neurol.* 72, 1275–1280. doi: 10.1001/ jamaneurol.2015.1867

Khan, T. K., and Alkon, D. L. (2015). Peripheral biomarkers of Alzheimer's disease. J. Alzheimers Dis. 44, 729–744. doi: 10.3233/jad-142262

Kleinberger, G., Yamanishi, Y., Suárez-Calvet, M., Czirr, E., Lohmann, E., Cuyvers, E., et al. (2014). TREM2 mutations implicated in neurodegeneration impair cell surface transport and phagocytosis. *Sci. Transl. Med.* 6:243ra86. doi: 10.1126/ scitranslmed.3009093

Korff, A., Liu, C., Ginghina, C., Shi, M., and Zhang, J. (2013). α-Synuclein in cerebrospinal fluid of Alzheimer's disease and mild cognitive impairment. *J. Alzheimers Dis.* 36, 679–688. doi: 10.3233/jad-130458

Kubota, Y., Putkey, J. A., and Waxham, M. N. (2007). Neurogranin controls the spatiotemporal pattern of postsynaptic Ca2+/CaM signaling. *Biophys. J.* 93, 3848–3859. doi: 10.1529/biophysj.107.106849

Kulichikhin, K. Y., Fedotov, S. A., Rubel, M. S., Zalutskaya, N. M., Zobnina, A. E., Malikova, O. A., et al. (2021). Development of molecular tools for diagnosis of Alzheimer's disease that are based on detection of amyloidogenic proteins. *Prion* 15, 56–69. doi: 10.1080/19336896.2021.1917289

Kvartsberg, H., Portelius, E., Andreasson, U., Brinkmalm, G., Hellwig, K., Lelental, N., et al. (2015). Characterization of the postsynaptic protein neurogranin in paired cerebrospinal fluid and plasma samples from Alzheimer's disease patients and healthy controls. *Alzheimers Res. Ther.* 7:40. doi: 10.1186/s13195-015-0124-3

La Joie, R., Bejanin, A., Fagan, A. M., Ayakta, N., Baker, S. L., Bourakova, V., et al. (2018). Associations between [(18)F]AV1451 tau PET and CSF measures of tau pathology in a clinical sample. *Neurology* 90, e282–e290. doi: 10.1212/wnl.000000000004860

Lantero Rodriguez, J., Karikari, T. K., Suárez-Calvet, M., Troakes, C., King, A., Emersic, A., et al. (2020). Plasma p-tau181 accurately predicts Alzheimer's disease pathology at least 8 years prior to post-mortem and improves the clinical characterisation of cognitive decline. *Acta Neuropathol.* 140, 267–278. doi: 10.1007/ s00401-020-02195-x

Lewczuk, P., Esselmann, H., Otto, M., Maler, J. M., Henkel, A. W., Henkel, M. K., et al. (2004). Neurochemical diagnosis of Alzheimer's dementia by CSF Abeta42, Abeta42/

Abeta40 ratio and total tau. *Neurobiol. Aging* 25, 273–281. doi: 10.1016/ s0197-4580(03)00086-1

Li, Y., Fang, J., Zhou, Z., Zhou, Q., Sun, S., Jin, Z., et al. (2020). Downregulation of lncRNA BACE1-AS improves dopamine-dependent oxidative stress in rats with Parkinson's disease by upregulating microRNA-34b-5p and downregulating BACE1. *Cell Cycle* 19, 1158–1171. doi: 10.1080/15384101.2020.1749447

Libiger, O., Shaw, L. M., Watson, M. H., Nairn, A. C., Umaña, K. L., Biarnes, M. C., et al. (2021). Longitudinal CSF proteomics identifies NPTX2 as a prognostic biomarker of Alzheimer's disease. *Alzheimers Dement.* 17, 1976–1987. doi: 10.1002/alz.12353

Lill, C. M., Rengmark, A., Pihlstrøm, L., Fogh, I., Shatunov, A., Sleiman, P. M., et al. (2015). The role of TREM2 R47H as a risk factor for Alzheimer's disease, frontotemporal lobar degeneration, amyotrophic lateral sclerosis, and Parkinson's disease. *Alzheimers Dement.* 11, 1407–1416. doi: 10.1016/j.jalz.2014.12.009

Liu, D., Dong, X., Yang, R., Guo, H., Wang, T., and Xu, G. (2020). Visinin-like protein-1 level is associated with short-term functional outcome of acute ischemic stroke: A prospective cohort study. *Medicine* 99:e19252. doi: 10.1097/md.000000000019252

Liu, J., Geraghty, J. R., Schram, S., Cropper, H. C., Lei, J., Loeb, J. A., et al. (2023). Prevention of Alzheimer pathology by blocking neuregulin signaling on microglia. *eNe uro*:ENEURO.0422-23.2023:10. doi: 10.1523/eneuro.0422-23.2023

Liu, W., Lin, H., He, X., Chen, L., Dai, Y., Jia, W., et al. (2020). Neurogranin as a cognitive biomarker in cerebrospinal fluid and blood exosomes for Alzheimer's disease and mild cognitive impairment. *Transl. Psychiatry* 10:125. doi: 10.1038/ s41398-020-0801-2

Liu, P. P., Xie, Y., Meng, X. Y., and Kang, J. S. (2019). History and progress of hypotheses and clinical trials for Alzheimer's disease. *Signal Transduct. Target. Ther.* 4:29. doi: 10.1038/s41392-019-0063-8

Liu, Q., Xie, F., Siedlak, S. L., Nunomura, A., Honda, K., Moreira, P. I., et al. (2004). Neurofilament proteins in neurodegenerative diseases. *Cell. Mol. Life Sci.* 61, 3057–3075. doi: 10.1007/s00018-004-4268-8

Long, J. M., and Holtzman, D. M. (2019). Alzheimer disease: an update on pathobiology and treatment strategies. *Cell* 179, 312–339. doi: 10.1016/j.cell.2019.09.001

Luo, X., Hou, L., Shi, H., Zhong, X., Zhang, Y., Zheng, D., et al. (2013). CSF levels of the neuronal injury biomarker visinin-like protein-1 in Alzheimer's disease and dementia with Lewy bodies. *J. Neurochem.* 127, 681–690. doi: 10.1111/jnc.12331

Ma, L. Z., Tan, L., Bi, Y. L., Shen, X. N., Xu, W., Ma, Y. H., et al. (2020). Dynamic changes of CSF sTREM2 in preclinical Alzheimer's disease: the CABLE study. *Mol. Neurodegener.* 15:25. doi: 10.1186/s13024-020-00374-8

Mahaman, Y., Embaye, K. S., Huang, F., Li, L., Zhu, F., Wang, J. Z., et al. (2022). Biomarkers used in Alzheimer's disease diagnosis, treatment, and prevention. *Ageing Res. Rev.* 74:101544. doi: 10.1016/j.arr.2021.101544

Malki, I., Cantrelle, F. X., Sottejeau, Y., Lippens, G., Lambert, J. C., and Landrieu, I. (2017). Regulation of the interaction between the neuronal BIN1 isoform 1 and Tau proteins - role of the SH3 domain. *FEBS J.* 284, 3218–3229. doi: 10.1111/febs.14185

Marambaud, P., Dreses-Werringloer, U., and Vingtdeux, V. (2009). Calcium signaling in neurodegeneration. *Mol. Neurodegener.* 4:20. doi: 10.1186/1750-1326-4-20

Mattsson, N., Andreasson, U., Zetterberg, H., and Blennow, K. (2017). Association of Plasma Neurofilament Light With Neurodegeneration in Patients With Alzheimer Disease. JAMA Neurol. 74, 557–566. doi: 10.1001/jamaneurol.2016.6117

Mattsson, N., Insel, P. S., Palmqvist, S., Portelius, E., Zetterberg, H., Weiner, M., et al. (2016). Cerebrospinal fluid tau, neurogranin, and neurofilament light in Alzheimer's disease. *EMBO Mol. Med.* 8, 1184–1196. doi: 10.15252/emmm.201606540

Mattsson-Carlgren, N., Janelidze, S., Bateman, R. J., Smith, R., Stomrud, E., Serrano, G. E., et al. (2021). Soluble P-tau217 reflects amyloid and tau pathology and mediates the association of amyloid with tau. *EMBO Mol. Med.* 13:e14022. doi: 10.15252/emmm.202114022

Mavroudis, I., Chowdhury, R., Petridis, F., Karantali, E., Chatzikonstantinou, S., Balmus, I. M., et al. (2021). YKL-40 as a Potential Biomarker for the Differential Diagnosis of Alzheimer's Disease. *Medicina* 58:60. doi: 10.3390/medicina 58010060

Mavroudis, I. A., Petridis, F., Chatzikonstantinou, S., Karantali, E., and Kazis, D. (2021). A meta-analysis on the levels of VILIP-1 in the CSF of Alzheimer's disease compared to normal controls and other neurodegenerative conditions. *Aging Clin. Exp. Res.* 33, 265–272. doi: 10.1007/s40520-019-01458-2

Mavroudis, I. A., Petridis, F., Chatzikonstantinou, S., and Kazis, D. (2020). A metaanalysis on CSF neurogranin levels for the diagnosis of Alzheimer's disease and mild cognitive impairment. *Aging Clin. Exp. Res.* 32, 1639–1646. doi: 10.1007/ s40520-019-01326-z

Mayeux, R., Honig, L. S., Tang, M. X., Manly, J., Stern, Y., Schupf, N., et al. (2003). Plasma A[beta]40 and A[beta]42 and Alzheimer's disease: relation to age, mortality, and risk. *Neurology* 61, 1185–1190. doi: 10.1212/01.wnl.0000091890.32140.8f

McConlogue, L., Buttini, M., Anderson, J. P., Brigham, E. F., Chen, K. S., Freedman, S. B., et al. (2007). Partial reduction of BACE1 has dramatic effects on Alzheimer plaque and synaptic pathology in APP Transgenic Mice. *J. Biol. Chem.* 282, 26326–26334. doi: 10.1074/jbc.M611687200

Menendez-Gonzalez, M., and Gasparovic, C. (2019). Albumin Exchange in Alzheimer's Disease: Might CSF Be an Alternative Route to Plasma? *Front. Neurol.* 10:1036. doi: 10.3389/fneur.2019.01036

Miyagawa, T., Ebinuma, I., Morohashi, Y., Hori, Y., Young Chang, M., Hattori, H., et al. (2016). BIN1 regulates BACE1 intracellular trafficking and amyloid- β production. *Hum. Mol. Genet.* 25, 2948–2958. doi: 10.1093/hmg/ddw146

Montagne, A., Nation, D. A., Sagare, A. P., Barisano, G., Sweeney, M. D., Chakhoyan, A., et al. (2020). APOE4 leads to blood-brain barrier dysfunction predicting cognitive decline. *Nature* 581, 71–76. doi: 10.1038/s41586-020-2247-3

Montoliu-Gaya, L., Alcolea, D., Ashton, N. J., Pegueroles, J., Levin, J., Bosch, B., et al. (2023). Plasma and cerebrospinal fluid glial fibrillary acidic protein levels in adults with Down syndrome: a longitudinal cohort study. *EBioMedicine* 90:104547. doi: 10.1016/j. ebiom.2023.104547

Mouton-Liger, F., Dumurgier, J., Cognat, E., Hourregue, C., Zetterberg, H., Vanderstichele, H., et al. (2020). CSF levels of the BACE1 substrate NRG1 correlate with cognition in Alzheimer's disease. *Alzheimers Res. Ther.* 12:88. doi: 10.1186/s13195-020-00655-w

Mroczko, B., Groblewska, M., Zboch, M., Muszyński, P., Zajkowska, A., Borawska, R., et al. (2015). Evaluation of visinin-like protein 1 concentrations in the cerebrospinal fluid of patients with mild cognitive impairment as a dynamic biomarker of Alzheimer's disease. *J. Alzheimers Dis.* 43, 1031–1037. doi: 10.3233/jad-141050

Nakamura, A., Kaneko, N., Villemagne, V. L., Kato, T., Doecke, J., Doré, V., et al. (2018). High performance plasma amyloid- β biomarkers for Alzheimer's disease. Nature 554, 249–254. doi: 10.1038/nature25456

Nation, D. A., Sweeney, M. D., Montagne, A., Sagare, A. P., D'orazio, L. M., Pachicano, M., et al. (2019). Blood-brain barrier breakdown is an early biomarker of human cognitive dysfunction. *Nat. Med.* 25, 270–276. doi: 10.1038/s41591-018-0297-y

Nilsson, J., Gobom, J., Sjödin, S., Brinkmalm, G., Ashton, N. J., Svensson, J., et al. (2021). Cerebrospinal fluid biomarker panel for synaptic dysfunction in Alzheimer's disease. *Alzheimers Dement* 13:e12179. doi: 10.1002/dad2.12179

Nutu, M., Zetterberg, H., Londos, E., Minthon, L., Nägga, K., Blennow, K., et al. (2013). Evaluation of the cerebrospinal fluid amyloid- β 1-42/amyloid- β 1-40 ratio measured by alpha-LISA to distinguish Alzheimer's disease from other dementia disorders. *Dement. Geriatr. Cogn. Disord.* 36, 99–110. doi: 10.1159/000353442

Öhrfelt, A., Brinkmalm, A., Dumurgier, J., Brinkmalm, G., Hansson, O., Zetterberg, H., et al. (2016). The pre-synaptic vesicle protein synaptotagmin is a novel biomarker for Alzheimer's disease. *Alzheimers Res. Ther.* 8:41. doi: 10.1186/s13195-016-0208-8

Olsson, B., Lautner, R., Andreasson, U., Öhrfelt, A., Portelius, E., Bjerke, M., et al. (2016). CSF and blood biomarkers for the diagnosis of Alzheimer's disease: a systematic review and meta-analysis. *Lancet Neurol.* 15, 673–684. doi: 10.1016/s1474-4422(16)00070-3

Ossenkoppele, R., Smith, R., Ohlsson, T., Strandberg, O., Mattsson, N., Insel, P. S., et al. (2019). Associations between tau, $A\beta$, and cortical thickness with cognition in Alzheimer disease. *Neurology* 92, e601–e612. doi: 10.1212/wnl.000000000006875

Ovod, V., Ramsey, K. N., Mawuenyega, K. G., Bollinger, J. G., Hicks, T., Schneider, T., et al. (2017). Amyloid β concentrations and stable isotope labeling kinetics of human plasma specific to central nervous system amyloidosis. *Alzheimers Dement.* 13, 841–849. doi: 10.1016/j.jalz.2017.06.2266

Palmqvist, S., Insel, P. S., Stomrud, E., Janelidze, S., Zetterberg, H., Brix, B., et al. (2019). Cerebrospinal fluid and plasma biomarker trajectories with increasing amyloid deposition in Alzheimer's disease. *EMBO Mol. Med.* 11:e11170. doi: 10.15252/emmm.201911170

Palmqvist, S., Janelidze, S., Quiroz, Y. T., Zetterberg, H., Lopera, F., Stomrud, E., et al. (2020). Discriminative Accuracy of Plasma Phospho-tau217 for Alzheimer Disease vs Other Neurodegenerative Disorders. *JAMA* 324, 772–781. doi: 10.1001/jama.2020.12134

Palmqvist, S., Mattsson, N., and Hansson, O.Alzheimer's Disease Neuroimaging Initiative (2016). Cerebrospinal fluid analysis detects cerebral amyloid-β accumulation earlier than positron emission tomography. *Brain* 139, 1226–1236. doi: 10.1093/brain/ aww015

Pampuscenko, K., Morkuniene, R., Sneideris, T., Smirnovas, V., Budvytyte, R., Valincius, G., et al. (2020). Extracellular tau induces microglial phagocytosis of living neurons in cell cultures. *J. Neurochem.* 154, 316–329. doi: 10.1111/jnc.14940

Pankonin, M. S., Sohi, J., Kamholz, J., and Loeb, J. A. (2009). Differential distribution of neuregulin in human brain and spinal fluid. *Brain Res.* 1258, 1–11. doi: 10.1016/j. brainres.2008.12.047

Pant, S., Sharma, M., Patel, K., Caplan, S., Carr, C. M., and Grant, B. D. (2009). AMPH-1/Amphiphysin/Bin1 functions with RME-1/Ehd1 in endocytic recycling. *Nat. Cell Biol.* 11, 1399–1410. doi: 10.1038/ncb1986

Pekny, M., and Nilsson, M. (2005). Astrocyte activation and reactive gliosis. *Glia* 50, 427–434. doi: 10.1002/glia.20207

Pereira, J. B., Janelidze, S., Smith, R., Mattsson-Carlgren, N., Palmqvist, S., Teunissen, C. E., et al. (2021). Plasma GFAP is an early marker of amyloid- β but not tau pathology in Alzheimer's disease. *Brain* 144, 3505–3516. doi: 10.1093/brain/awab223

Pereira, J. B., Westman, E., and Hansson, O. (2017). Association between cerebrospinal fluid and plasma neurodegeneration biomarkers with brain atrophy in Alzheimer's disease. *Neurobiol. Aging* 58, 14–29. doi: 10.1016/j.neurobiolaging.2017.06.002

Piccio, L., Deming, Y., Del-Águila, J. L., Ghezzi, L., Holtzman, D. M., Fagan, A. M., et al. (2016). Cerebrospinal fluid soluble TREM2 is higher in Alzheimer disease and associated with mutation status. *Acta Neuropathol.* 131, 925–933. doi: 10.1007/s00401-016-1533-5

Portelius, E., Olsson, B., Höglund, K., Cullen, N. C., Kvartsberg, H., Andreasson, U., et al. (2018). Cerebrospinal fluid neurogranin concentration in neurodegeneration: relation to clinical phenotypes and neuropathology. *Acta Neuropathol.* 136, 363–376. doi: 10.1007/s00401-018-1851-x

Price, J. L., and Morris, J. C. (1999). Tangles and plaques in nondemented aging and "preclinical" Alzheimer's disease. *Ann. Neurol.* 45, 358–368. doi: 10.1002/1531-8249(199903)45:3<358::aid-ana12>3.0.co;2-x

Reitz, C., and Mayeux, R. (2014). Alzheimer disease: epidemiology, diagnostic criteria, risk factors and biomarkers. *Biochem. Pharmacol.* 88, 640–651. doi: 10.1016/j. bcp.2013.12.024

Represa, A., Deloulme, J. C., Sensenbrenner, M., Ben-Ari, Y., and Baudier, J. (1990). Neurogranin: immunocytochemical localization of a brain-specific protein kinase C substrate. *J. Neurosci.* 10, 3782–3792. doi: 10.1523/jneurosci.10-12-03782.1990

Sandelius, Å., Cullen, N. C., Källén, Å., Rosengren, L., Jensen, C., Kostanjevecki, V., et al. (2018). Transient increase in CSF GAP-43 concentration after ischemic stroke. *BMC Neurol.* 18:202. doi: 10.1186/s12883-018-1210-5

Sandelius, Å., Portelius, E., Källén, Å., Zetterberg, H., Rot, U., Olsson, B., et al. (2019). Elevated CSF GAP-43 is Alzheimer's disease specific and associated with tau and amyloid pathology. *Alzheimers Dement.* 15, 55–64. doi: 10.1016/j.jalz.2018.08.006

Sartori, M., Mendes, T., Desai, S., Lasorsa, A., Herledan, A., Malmanche, N., et al. (2019). BIN1 recovers tauopathy-induced long-term memory deficits in mice and interacts with Tau through Thr(348) phosphorylation. *Acta Neuropathol.* 138, 631–652. doi: 10.1007/s00401-019-02017-9

Sato, C., Barthélemy, N. R., Mawuenyega, K. G., Patterson, B. W., Gordon, B. A., Jockel-Balsarotti, J., et al. (2018). Tau Kinetics in Neurons and the Human Central Nervous System. *Neuron* 97, 1284–1298.e7. doi: 10.1016/j.neuron.2018.02.015

Saunders, T., Gunn, C., Blennow, K., Kvartsberg, H., Zetterberg, H., Shenkin, S. D., et al. (2023). Neurogranin in Alzheimer's disease and ageing: A human post-mortem study. *Neurobiol. Dis.* 177:105991. doi: 10.1016/j.nbd.2023.105991

Scheff, S. W., Price, D. A., Schmitt, F. A., Dekosky, S. T., and Mufson, E. J. (2007). Synaptic alterations in CA1 in mild Alzheimer disease and mild cognitive impairment. *Neurology* 68, 1501–1508. doi: 10.1212/01.wnl.0000260698.46517.8f

Scheltens, P., De Strooper, B., Kivipelto, M., Holstege, H., Chételat, G., Teunissen, C. E., et al. (2021). Alzheimer's disease. *Lancet* 397, 1577–1590. doi: 10.1016/S0140-6736(20)32205-4

Schindler, S. E. (2022). Fluid Biomarkers in Dementia Diagnosis. Continuum 28, 822–833. doi: 10.1212/con.00000000001083

Schindler, S. E., Bollinger, J. G., Ovod, V., Mawuenyega, K. G., Li, Y., Gordon, B. A., et al. (2019). High-precision plasma β -amyloid 42/40 predicts current and future brain amyloidosis. *Neurology* 93, e1647–e1659. doi: 10.1212/wnl.00000000008081

Schneider, J. A., Arvanitakis, Z., Leurgans, S. E., and Bennett, D. A. (2009). The neuropathology of probable Alzheimer disease and mild cognitive impairment. *Ann. Neurol.* 66, 200–208. doi: 10.1002/ana.21706

Shankar, G. M., Bloodgood, B. L., Townsend, M., Walsh, D. M., Selkoe, D. J., and Sabatini, B. L. (2007). Natural oligomers of the Alzheimer amyloid-beta protein induce reversible synapse loss by modulating an NMDA-type glutamate receptor-dependent signaling pathway. J. Neurosci. 27, 2866–2875. doi: 10.1523/jneurosci.4970-06.2007

Shao, K., Shan, S., Ru, W., and Ma, C. (2020). Association between serum NPTX2 and cognitive function in patients with vascular dementia. *Brain Behav.* 10:e01779. doi: 10.1002/brb3.1779

Shui, B., Tao, D., Florea, A., Cheng, J., Zhao, Q., Gu, Y., et al. (2018). Biosensors for Alzheimer's disease biomarker detection: a review. *Biochimie* 147, 13–24. doi: 10.1016/j. biochi.2017.12.015

Simrén, J., Elmgren, A., Blennow, K., and Zetterberg, H. (2023). Fluid biomarkers in Alzheimer's disease. Adv. Clin. Chem. 112, 249–281. doi: 10.1016/bs.acc.2022.09.006

Simrén, J., Weninger, H., Brum, W. S., Khalil, S., Benedet, A. L., Blennow, K., et al. (2022). Differences between blood and cerebrospinal fluid glial fibrillary Acidic protein levels: The effect of sample stability. *Alzheimers Dement.* 18, 1988–1992. doi: 10.1002/alz.12806

Skillbäck, T., Delsing, L., Synnergren, J., Mattsson, N., Janelidze, S., Nägga, K., et al. (2017). CSF/serum albumin ratio in dementias: a cross-sectional study on 1861 patients. *Neurobiol. Aging* 59, 1–9. doi: 10.1016/j.neurobiolaging.2017.06.028

Skillbäck, T., Rosén, C., Asztely, F., Mattsson, N., Blennow, K., and Zetterberg, H. (2014). Diagnostic performance of cerebrospinal fluid total tau and phosphorylated tau in Creutzfeldt-Jakob disease: results from the Swedish Mortality Registry. *JAMA Neurol.* 71, 476–483. doi: 10.1001/jamaneurol.2013.6455

Smirnov, D. S., Ashton, N. J., Blennow, K., Zetterberg, H., Simrén, J., Lantero-Rodriguez, J., et al. (2022). Plasma biomarkers for Alzheimer's Disease in

relation to neuropathology and cognitive change. Acta Neuropathol. 143, 487–503. doi: 10.1007/s00401-022-02408-5

Srivastava, S., Ahmad, R., and Khare, S. K. (2021). Alzheimer's disease and its treatment by different approaches: a review. *Eur. J. Med. Chem.* 216:113320. doi: 10.1016/j.ejmech.2021.113320

Stejskal, D., Sporova, L., Svestak, M., and Karpisek, M. (2011). Determination of serum visinin like protein-1 and its potential for the diagnosis of brain injury due to the stroke: a pilot study. *Biomed. Pap. Med. Fac. Univ. Palacky Olomouc Czech Repub.* 155, 263–268. doi: 10.5507/bp.2011.049

Stomrud, E., Hansson, O., Blennow, K., Minthon, L., and Londos, E. (2007). Cerebrospinal fluid biomarkers predict decline in subjective cognitive function over 3 years in healthy elderly. *Dement. Geriatr. Cogn. Disord.* 24, 118–124. doi: 10.1159/000105017

Strozyk, D., Blennow, K., White, L. R., and Launer, L. J. (2003). CSF Abeta 42 levels correlate with amyloid-neuropathology in a population-based autopsy study. *Neurology* 60, 652–656. doi: 10.1212/01.wnl.0000046581.81650.d0

Suárez-Calvet, M., Araque Caballero, M., Kleinberger, G., Bateman, R. J., Fagan, A. M., Morris, J. C., et al. (2016a). Early changes in CSF sTREM2 in dominantly inherited Alzheimer's disease occur after amyloid deposition and neuronal injury. *Sci. Transl. Med.* 8:369ra178. doi: 10.1126/scitranslmed.aag1767

Suárez-Calvet, M., Karikari, T. K., Ashton, N. J., Lantero Rodríguez, J., Milà-Alomà, M., Gispert, J. D., et al. (2020). Novel tau biomarkers phosphorylated at T181, T217 or T231 rise in the initial stages of the preclinical Alzheimer's continuum when only subtle changes in A β pathology are detected. *EMBO Mol. Med.* 12:e12921. doi: 10.15252/emmm.202012921

Suárez-Calvet, M., Kleinberger, G., Araque Caballero, M., Brendel, M., Rominger, A., Alcolea, D., et al. (2016b). sTREM2 cerebrospinal fluid levels are a potential biomarker for microglia activity in early-stage Alzheimer's disease and associate with neuronal injury markers. *EMBO Mol. Med.* 8, 466–476. doi: 10.15252/emmm.201506123

Sudwarts, A., Ramesha, S., Gao, T., Ponnusamy, M., Wang, S., Hansen, M., et al. (2022). BIN1 is a key regulator of proinflammatory and neurodegeneration-related activation in microglia. *Mol. Neurodegener.* 17:33. doi: 10.1186/s13024-022-00535-x

Sun, L., Tan, M. S., Hu, N., Yu, J. T., and Tan, L. (2013). Exploring the value of plasma BIN1 as a potential biomarker for alzheimer's disease. *J. Alzheimers Dis.* 37, 291–295. doi: 10.3233/jad-130392

Sunderland, T., Linker, G., Mirza, N., Putnam, K. T., Friedman, D. L., Kimmel, L. H., et al. (2003). Decreased beta-amyloid1-42 and increased tau levels in cerebrospinal fluid of patients with Alzheimer disease. *JAMA* 289, 2094–2103. doi: 10.1001/jama.289.16.2094

Sutphen, C. L., Mccue, L., Herries, E. M., Xiong, C., Ladenson, J. H., Holtzman, D. M., et al. (2018). Longitudinal decreases in multiple cerebrospinal fluid biomarkers of neuronal injury in symptomatic late onset Alzheimer's disease. *Alzheimers Dement.* 14, 869–879. doi: 10.1016/j.jalz.2018.01.012

Tan, Z., Jiang, J., Tian, F., Peng, J., Yang, Z., Li, S., et al. (2020). Serum Visinin-Like Protein 1 Is a Better Biomarker Than Neuron-Specific Enolase for Seizure-Induced Neuronal Injury: A Prospective and Observational Study. *Front. Neurol.* 11:567587. doi: 10.3389/fneur.2020.567587

Tan, M. S., Yu, J. T., and Tan, L. (2013). Bridging integrator 1 (BIN1): form, function, and Alzheimer's disease. *Trends Mol. Med.* 19, 594–603. doi: 10.1016/j. molmed.2013.06.004

Tarasoff-Conway, J. M., Carare, R. O., Osorio, R. S., Glodzik, L., Butler, T., Fieremans, E., et al. (2015). Clearance systems in the brain-implications for Alzheimer disease. *Nat. Rev. Neurol.* 11, 457–470. doi: 10.1038/nrneurol.2015.119

Teunissen, C. E., Verberk, I. M. W., Thijssen, E. H., Vermunt, L., Hansson, O., Zetterberg, H., et al. (2022). Blood-based biomarkers for Alzheimer's disease: towards clinical implementation. *Lancet Neurol.* 21, 66–77. doi: 10.1016/s1474-4422(21)00361-6

Thijssen, E. H., La Joie, R., Strom, A., Fonseca, C., Iaccarino, L., Wolf, A., et al. (2021). Plasma phosphorylated tau 217 and phosphorylated tau 181 as biomarkers in Alzheimer's disease and frontotemporal lobar degeneration: a retrospective diagnostic performance study. *Lancet Neurol.* 20, 739–752. doi: 10.1016/s1474-4422(21)00214-3

Thijssen, E. H., La Joie, R., Wolf, A., Strom, A., Wang, P., Iaccarino, L., et al. (2020). Diagnostic value of plasma phosphorylated tau181 in Alzheimer's disease and frontotemporal lobar degeneration. *Nat. Med.* 26, 387–397. doi: 10.1038/s41591-020-0762-2

Thomas, S., Hoxha, K., Tran, A., and Prendergast, G. C. (2019). Bin1 antibody lowers the expression of phosphorylated Tau in Alzheimer's disease. *J. Cell. Biochem.* 120, 18320–18331. doi: 10.1002/jcb.29142

Tibbling, G., Link, H., and Ohman, S. (1977). Principles of albumin and IgG analyses in neurological disorders. I. Establishment of reference values. *Scand. J. Clin. Lab. Invest.* 37, 385–390. doi: 10.1080/00365517709091496

Uberti, D., Cantarella, G., Facchetti, F., Cafici, A., Grasso, G., Bernardini, R., et al. (2004). TRAIL is expressed in the brain cells of Alzheimer's disease patients. *Neuroreport* 15, 579–581. doi: 10.1097/00001756-200403220-00002

Uberti, D., Ferrari-Toninelli, G., Bonini, S. A., Sarnico, I., Benarese, M., Pizzi, M., et al. (2007). Blockade of the tumor necrosis factor-related apoptosis inducing ligand death

receptor DR5 prevents beta-amyloid neurotoxicity. *Neuropsychopharmacology* 32, 872–880. doi: 10.1038/sj.npp.1301185

Ueno, M., Chiba, Y., Murakami, R., Matsumoto, K., Kawauchi, M., and Fujihara, R. (2016). Blood-brain barrier and blood-cerebrospinal fluid barrier in normal and pathological conditions. *Brain Tumor Pathol.* 33, 89–96. doi: 10.1007/s10014-016-0255-7

Vassar, R., Bennett, B. D., Babu-Khan, S., Kahn, S., Mendiaz, E. A., Denis, P., et al. (1999). Beta-secretase cleavage of Alzheimer's amyloid precursor protein by the transmembrane aspartic protease BACE. *Science* 286, 735–741. doi: 10.1126/science.286.5440.735

Verberk, I. M. W., Thijssen, E., Koelewijn, J., Mauroo, K., Vanbrabant, J., De Wilde, A., et al. (2020). Combination of plasma amyloid beta((1-42/1-40)) and glial fibrillary acidic protein strongly associates with cerebral amyloid pathology. *Alzheimers Res. Ther.* 12:118. doi: 10.1186/s13195-020-00682-7

Vermunt, L., Sikkes, S., van den Hout, A., Handels, R., Bos, I., van der Flier, W. M., et al. (2019). Duration of preclinical, prodromal, and dementia stages of Alzheimer's disease in relation to age, sex, and APOE genotype. *Alzheimers Dement.* 15, 888–898. doi: 10.1016/j.jalz.2019.04.001

Villar-Piqué, A., Schmitz, M., Hermann, P., Goebel, S., Bunck, T., Varges, D., et al. (2019). Plasma YKL-40 in the spectrum of neurodegenerative dementia. *J. Neuroinflammation* 16:145. doi: 10.1186/s12974-019-1531-3

Visser, P. J., Verhey, F., Knol, D. L., Scheltens, P., Wahlund, L. O., Freund-Levi, Y., et al. (2009). Prevalence and prognostic value of CSF markers of Alzheimer's disease pathology in patients with subjective cognitive impairment or mild cognitive impairment in the DESCRIPA study: a prospective cohort study. *Lancet Neurol.* 8, 619–627. doi: 10.1016/s1474-4422(09)70139-5

Vrillon, A., Mouton-Liger, F., Martinet, M., Cognat, E., Hourregue, C., Dumurgier, J., et al. (2022). Plasma neuregulin 1 as a synaptic biomarker in Alzheimer's disease: a discovery cohort study. *Alzheimers Res. Ther.* 14:71. doi: 10.1186/s13195-022-01014-7

Wagner, O. I., Rammensee, S., Korde, N., Wen, Q., Leterrier, J. F., and Janmey, P. A. (2007). Softness, strength and self-repair in intermediate filament networks. *Exp. Cell Res.* 313, 2228–2235. doi: 10.1016/j.yexcr.2007.04.025

Wallin, Å. K., Blennow, K., Zetterberg, H., Londos, E., Minthon, L., and Hansson, O. (2010). CSF biomarkers predict a more malignant outcome in Alzheimer disease. *Neurology* 74, 1531–1537. doi: 10.1212/WNL.0b013e3181dd4dd8

Wang, Z. Y., Han, Z. M., Liu, Q. F., Tang, W., Ye, K., and Yao, Y. Y. (2015). Use of CSF α-synuclein in the differential diagnosis between Alzheimer's disease and other neurodegenerative disorders. *Int. Psychogeriatr.* 27, 1429–1438. doi: 10.1017/s1041610215000447

Wang, J., Qiao, F., Shang, S., Li, P., Chen, C., Dang, L., et al. (2018). Elevation of Plasma Amyloid- β Level is More Significant in Early Stage of Cognitive Impairment: A Population-Based Cross-Sectional Study. *J. Alzheimers Dis.* 64, 61–69. doi: 10.3233/ jad-180140

Wang, Z., Tan, L., Zong, Y., Ma, Y. H., Wang, Z. B., Wang, H. F., et al. (2023). sTREM2 and GFAP Mediated the Association of IGF-1 Signaling Biomarkers with Alzheimer's Disease Pathology. J. Alzheimers Dis. 92, 791–797. doi: 10.3233/jad-220725

Wang, S., Zhang, J., and Pan, T. (2018). APOE £4 is associated with higher levels of CSF SNAP-25 in prodromal Alzheimer's disease. *Neurosci. Lett.* 685, 109–113. doi: 10.1016/j.neulet.2018.08.029

Wang, L., Zhang, M., Wang, Q., Jiang, X., Li, K., and Liu, J. (2020). APOE & Allele Is Associated with Elevated Levels of CSF VILIP-1 in Preclinical Alzheimer's Disease. *Neuropsychiatr. Dis. Treat.* 16, 923–931. doi: 10.2147/ndt.S235395

Wechsler-Reya, R., Elliott, K., and Prendergast, G. (1998). A Role for the Putative Tumor Suppressor Bin1 in Muscle Cell Differentiation. *Mol. Cell. Biol.* 18, 566–575. doi: 10.1128/MCB.18.1.566

Wennström, M., Surova, Y., Hall, S., Nilsson, C., Minthon, L., Hansson, O., et al. (2015). The Inflammatory Marker YKL-40 Is Elevated in Cerebrospinal Fluid from Patients with Alzheimer's but Not Parkinson's Disease or Dementia with Lewy Bodies. *PLoS One* 10:e0135458. doi: 10.1371/journal.pone.0135458

Weston, P. S. J., Poole, T., Ryan, N. S., Nair, A., Liang, Y., Macpherson, K., et al. (2017). Serum neurofilament light in familial Alzheimer disease: A marker of early neurodegeneration. *Neurology* 89, 2167–2175. doi: 10.1212/wnl.000000000004667

Willemse, E., Sieben, A., Somers, C., Vermeiren, Y., De Roeck, N., Timmers, M., et al. (2021). Neurogranin as biomarker in CSF is non-specific to Alzheimer's disease dementia. *Neurobiol. Aging* 108, 99–109. doi: 10.1016/j. neurobiolaging.2021.08.002

Winston, C. N., Goetzl, E. J., Akers, J. C., Carter, B. S., Rockenstein, E. M., Galasko, D., et al. (2016). Prediction of conversion from mild cognitive impairment to dementia with neuronally derived blood exosome protein profile. *Alzheimers Dement* 3, 63–72. doi: 10.1016/j.dadm.2016.04.001

Wojdała, A. L., Bellomo, G., Gaetani, L., Toja, A., Chipi, E., Shan, D., et al. (2023). Trajectories of CSF and plasma biomarkers across Alzheimer's disease continuum: disease staging by NF-L, p-tau181, and GFAP. *Neurobiol. Dis.* 189:106356. doi: 10.1016/j. nbd.2023.106356

World Health Organization (2017). Global action plan on the public health response to dementia 2017–2025. (Geneva: World Health Organization).

Wu, Y. Y., Hsu, J. L., Wang, H. C., Wu, S. J., Hong, C. J., and Cheng, I. H. (2015). Alterations of the neuroinflammatory markers IL-6 and TRAIL in Alzheimer's disease. *Dement. Geriatr. Cogn. Dis. Extra* 5, 424–434. doi: 10.1159/000439214

Xiao, M. F., Xu, D., Craig, M. T., Pelkey, K. A., Chien, C. C., Shi, Y., et al. (2017). NPTX2 and cognitive dysfunction in Alzheimer's Disease. *Elife* 6:e23798. doi: 10.7554/ eLife.23798

Xie, X.-Y., Zhao, Q.-H., Huang, Q., Dammer, E., Chen, S.-D., Ren, R.-J., et al. (2022). Genetic profiles of familial late-onset Alzheimer's disease in China: the Shanghai FLOAD study. *Genes Diseases* 9, 1639–1649. doi: 10.1016/j.gendis.2021.05.001

Yan, Y., Jensen, K., and Brown, A. (2007). The polypeptide composition of moving and stationary neurofilaments in cultured sympathetic neurons. *Cell Motil. Cytoskeleton* 64, 299–309. doi: 10.1002/cm.20184

Yang, L. B., Lindholm, K., Yan, R., Citron, M., Xia, W., Yang, X. L., et al. (2003). Elevated beta-secretase expression and enzymatic activity detected in sporadic Alzheimer disease. *Nat. Med.* 9, 3–4. doi: 10.1038/nm0103-3

Yin, Z., Raj, D., Saiepour, N., Van Dam, D., Brouwer, N., Holtman, I. R., et al. (2017). Immune hyperreactivity of A β plaque-associated microglia in Alzheimer's disease. *Neurobiol. Aging* 55, 115–122. doi: 10.1016/j.neurobiolaging.2017.03.021

Zetterberg, H., Andreasson, U., Hansson, O., Wu, G., Sankaranarayanan, S., Andersson, M. E., et al. (2008). Elevated cerebrospinal fluid BACE1 activity in incipient Alzheimer disease. *Arch. Neurol.* 65, 1102–1107. doi: 10.1001/archneur.65.8.1102

Zetterberg, H., Hietala, M. A., Jonsson, M., Andreasen, N., Styrud, E., Karlsson, I., et al. (2006). Neurochemical aftermath of amateur boxing. *Arch. Neurol.* 63, 1277–1280. doi: 10.1001/archneur.63.9.1277

Zetterberg, H., Skillbäck, T., Mattsson, N., Trojanowski, J. Q., Portelius, E., Shaw, L. M., et al. (2016). Association of cerebrospinal fluid neurofilament light concentration with Alzheimer disease progression. *JAMA Neurol.* 73, 60–67. doi: 10.1001/jamaneurol.2015.3037

Zhang, X., Tang, L., Yang, J., Meng, L., Chen, J., Zhou, L., et al. (2023). Soluble TREM2 ameliorates tau phosphorylation and cognitive deficits through activating transgelin-2 in Alzheimer's disease. *Nat. Commun.* 14:6670. doi: 10.1038/s41467-023-42505-x

Zhang, H., Therriault, J., Kang, M. S., Ng, K. P., Pascoal, T. A., Rosa-Neto, P., et al. (2018). Cerebrospinal fluid synaptosomal-associated protein 25 is a key player in synaptic degeneration in mild cognitive impairment and Alzheimer's disease. *Alzheimers Res. Ther.* 10:80. doi: 10.1186/s13195-018-0407-6

Zhao, A., Jiao, Y., Ye, G., Kang, W., Tan, L., Li, Y., et al. (2022). Soluble TREM2 levels associate with conversion from mild cognitive impairment to Alzheimer's disease. *J. Clin. Invest.* 132:e158708. doi: 10.1172/jci158708

Zhou, S., Li, Y., Zhang, Z., and Yuan, Y. (2023). An insight into the TAM system in Alzheimer's disease. *Int. Immunopharmacol.* 116:109791. doi: 10.1016/j. intimp.2023.109791

Zhou, J., Wade, S. D., Graykowski, D., Xiao, M. F., Zhao, B., Giannini, L. A. A., et al. (2023). The neuronal pentraxin Nptx2 regulates complement activity and restrains microglia-mediated synapse loss in neurodegeneration. *Sci. Transl. Med.* 15:eadf0141. doi: 10.1126/scitranslmed.adf0141

Zhou, W., Zhou, Y., and Li, J. (2023). Association between cerebrospinal fluid soluble TREM2, Alzheimer's disease and other neurodegenerative diseases. J. Clin. Med. 12:3589. doi: 10.3390/jcm12103589

Zhu, Y., Guo, X., Zhu, F., Zhang, Q., and Yang, Y.For The Alzheimer's Disease Neuroimaging Initiative (2023). Association of CSF GAP-43 and APOE ε 4 with Cognition in Mild Cognitive Impairment and Alzheimer's Disease. *Cells* 12:13. doi: 10.3390/cells12010013

Zuliani, G., Trentini, A., Rosta, V., Guerrini, R., Pacifico, S., Bonazzi, S., et al. (2020). Increased blood BACE1 activity as a potential common pathogenic factor of vascular dementia and late onset Alzheimer's disease. *Sci. Rep.* 10:14980. doi: 10.1038/ s41598-020-72168-3